

## Review

# Biophysical Dissection of Isolated GPCRs: The Adenosine A<sub>2A</sub> Receptor under the Bistouries

Jean-Louis Banères <sup>1</sup>, Thomas Botzanowski <sup>2,†</sup>, Jean A. Boutin <sup>3,\*</sup>, Barbara Calamini <sup>4,‡</sup>, Jérôme Castel <sup>2,5</sup>, Laurent J. Catoire <sup>6</sup>, Sarah Cianférani <sup>2</sup>, Claire Demesmay <sup>7</sup>, Gavin Ferguson <sup>8</sup>, Gilles Ferry <sup>9</sup>, Julie Kniazeff <sup>1</sup>, Isabelle Krimm <sup>10</sup>, Thierry Langer <sup>11</sup>, Guillaume Lebon <sup>8</sup>, Marie Ley <sup>2</sup>, Miklos Nyerges <sup>12</sup>, Magali Schwob <sup>5,13</sup>, Catherine Venien-Bryan <sup>14</sup>, Renaud Wagner <sup>13</sup>, Gabrielle Zeder-Lutz <sup>13</sup> and Claudia Zilian-Stohrer <sup>6</sup>

- <sup>1</sup> Institut des Biomolécules Max Mousseron (IBMM), UMR 5247, CNRS-Université de Montpellier-ENSCM, 34093 Montpellier, France
  - <sup>2</sup> Laboratoire de Spectrométrie de Masse BioOrganique (LSMBO), Institut Pluridisciplinaire Hubert Curien, IPHC, CNRS UMR 7178, Infrastructure Nationale de Protéomique ProFI, FR 2048 CNRS CEA, Université de Strasbourg, 67084 Strasbourg, France
  - <sup>3</sup> Laboratory of Neuroendocrine Endocrine and Germinal Differentiation and Communication (NorDiC), Université de Rouen Normandie, Inserm, UMR 1239, 76000 Rouen, France
  - <sup>4</sup> Center for Drug Discovery, Department of Neurobiology, Duke University Medical Center, Durham, NC 27710, USA
  - <sup>5</sup> Department of Structural Biology, NovAliX, 67080 Strasbourg, France
  - <sup>6</sup> Laboratoire de Biologie Physico-Chimique des Protéines Membranaires, UMR 7099, IBPC, 75005 Paris, France
  - <sup>7</sup> Equipe Techsep, Institut des Sciences Analytiques, 69100 Villeurbanne, France
  - <sup>8</sup> Institut de Génomique Fonctionnelle, Université de Montpellier, CNRS, INSERM, 34094 Montpellier, France
  - <sup>9</sup> Institut de Recherches Servier, 78290 Croissy-sur-Seine, France
  - <sup>10</sup> Small Molecules for Biological Targets Team, Université Claude Bernard Lyon 1, INSERM 1052, CNRS 5286, Centre Léon Bérard, Centre de Recherche en Cancérologie de Lyon, 69373 Lyon, France
  - <sup>11</sup> Department of Pharmaceutical Sciences, University of Vienna, 1090 Vienna, Austria
  - <sup>12</sup> Servier Research Institute of Medicinal Chemistry, 1031 Budapest, Hungary
  - <sup>13</sup> Plateforme IMPReSs, UMR 7242, Biotechnologie et Signalisation Cellulaire, Ecole Supérieure de Biotechnologie de Strasbourg, 67412 Illkirch, France
  - <sup>14</sup> Institut de Minéralogie, de Physique des Matériaux et de Cosmochimie, UMR 7590 CNRS, UPMC, IRD, MNHN, 75005 Paris, France
- \* Correspondence: ja.boutin.pro@gmail.com  
† Present Address: Department of Structural Biology, NovAliX, 67080 Strasbourg, France.  
‡ Present Address: Eli Lilly and Company, Boston, MA 02142, USA.



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**Abstract:** In an effort to provide an overview of the biophysical approaches used to study G-protein-coupled receptors, we chose to consider the adenosine A<sub>2A</sub> receptor as a model, as it is widely reported in the literature to explore the way GPCRs are studied nowadays. After a brief introduction of the receptor, we gathered descriptions of the various tools used to investigate the pharmacology and structure of the A<sub>2A</sub> receptor. We began by describing the key developments which have led to successful studies of GPCRs including the cloning, expression and purification of A<sub>2A</sub>, and the subsequent characterizations including quality control, binding and functional studies that have been necessary for the further understanding of the receptor. Then, we reviewed the reconstitution of A<sub>2A</sub> into nanodiscs as well as the use of this biological material in structural mass spectrometry, NMR, calorimetry and various other approaches to gain not only information about the structure and function of A<sub>2A</sub>, but also the dynamics of the receptor and the tools necessary to pursue such investigations. The body of techniques presented herein are applicable to all GPCRs amenable to purification.

**Keywords:** adenosine receptor A<sub>2A</sub>; biophysical approach; NMR; mass spectrometry; molecular pharmacology; expression; purification; reconstitution; ligands; techniques; review

## 1. Foreword: Biophysical Approaches to Answer Which Questions?

During the last two to three decades, as researchers in the GPCR area, we saw a profound evolution in the study of these membrane-bound proteins. It started with the description of whole genomes and the systematic comparison of sequences that revealed the remarkable conserved topology of this exceptionally large family of proteins. This in turn shed light on the necessity to explore the subtle molecular mechanisms and determinants that were responsible for recognizing such a variety of ligands, to understand how these receptors were selectively activated and how they are involved in so many cellular processes. Initially, the aim was to better understand the pharmacology of these proteins, and thus it became key to be able to study these entities in solution, or in reconstituted and controlled environments. The endpoint is to identify and develop directly interacting molecules that could become novel or “better” drugs. This includes molecules which provide safer pharmacological profiles, improved efficacy, in addition to greater precision, efficiency, and potency. Our scientific community therefore invested intensive efforts to produce and isolate these GPCRs and, hence, to assay and develop integrated analytical techniques to enable the characterization of GPCR structures, functions and dynamics, and further understand the way these receptors are modulated. To make such a characterization, a large diversity of biophysical principles have been exploited, including light, fluorescence, mass and NMR spectroscopies, surface plasmon resonance (SPR), X-ray diffraction and electron microscopy, among others. All of these techniques have been successfully applied to the biophysical dissection of the molecular functioning of GPCRs.

Like many other protein families in similar studies, a small number of prototypical receptors served the development of these GPCR-tailored methodologies, among which is the adenosine A<sub>2A</sub> receptor (A<sub>2A</sub>) which we chose to focus on in the present review. This receptor from the Class A GPCR family is indeed of continuing interest as a major therapeutic target in a panel of inflammatory, neurodegenerative and cancer disease types. More importantly, unlike a large proportion of GPCRs that are still recalcitrant to molecular investigations, this receptor rapidly appeared favorably throughout the literature to be produced and isolated in various experimental formats that prove compatible with a remarkable diversity.

The present review summarizes the main biophysical techniques that brought important information on GPCRs through the data obtained on the adenosine A<sub>2A</sub> receptor. In the first section, we will remind the reader of the main facts known about this receptor. The next sections extensively review the preparative conditions leading to the isolation of the receptor and the many biophysical approaches that contributed to our understanding of the structure and function of GPCRs in general.

## 2. Background and Introduction

Adenosine is a natural compound formed by an adenine attached to a ribose via a  $\beta$ -N9-glycosidic bond. It occurs widely in nature in the form of many diverse derivatives from multiple sources. All those molecules in which adenosine is a central chemical play important roles in living organisms. It is a part of one of the four building blocks of which the nucleic acids are made and its phosphorylated derivatives are key elements of the energetic system in all living systems. Finally, it exists also under a cyclic form, cAMP, which is a clear element of signaling pathways via the regulation of its key synthesis enzyme, adenylate cyclase. Adenosine is recognized essentially by four GPCRs: A<sub>1</sub>, A<sub>2A</sub>, A<sub>2B</sub> and A<sub>3</sub>. A full survey of these receptor characteristics can be found in Fredholm et al. [1]. The human A<sub>2A</sub> receptor is a 412-amino-acid-long protein encoded by the gene located at chromosome position 22q11.23. The A<sub>2A</sub> receptor was initially isolated and purified as early as 1974 [2]. Although membrane protein purification under an active form is not trivial, this receptor was widely used because its cloning and expression from human sources were reported early [3]. It was possible to express it in *E. coli* [4] and its characterization began from there.

A complete review of A<sub>2A</sub> pharmacology exists and is a reference for the matters concerning this receptor [5]. In brief, the A<sub>2A</sub> expression pattern concerns mainly the brain [6], peripheral T cells [7] and platelets as well as various organs such as the colon, heart and kidneys [5].

The main coupling routes of the receptors are in human platelets, cAMP [8], while coupling to ERK<sub>1/2</sub> has also been reported on the human cloned receptor [9]. Early works clearly indicated that A<sub>2A</sub> is not linked to intracellular calcium modifications, at least upon reference to its agonist effects [10]. In 1995, interplays were suspected with other GPCRs, leading to the A<sub>2A</sub>-dependent reduction in dopamine receptor activation [10]. A very complex picture of the co-regulations of ion channels was drawn later (see, for example, Ikeuchi et al. [11] and Wang and Zhou [12]).

The key step for receptor studies is the binding assay on membranes, whether derived from native tissues or from hosts in which the receptor has been cloned. This is also dependent on the availability of specific radioligands, such as the agonists [<sup>3</sup>H]-CGS21680 [13] or [<sup>3</sup>H]-NECA [14], generally preferred over tritiated adenosine [15], and the antagonist [<sup>3</sup>H]-XAC (xanthine amine congener) [16], with pK<sub>d</sub> in the range of 8. New ligands were added, with pK<sub>d</sub> in the range of 9, such as [<sup>3</sup>H]-ZM241385 [17].

A complete survey of the clinical status of A<sub>2A</sub> receptor ligands as therapeutic drugs can be found in Ijzermann et al. [5]. More than 50 different drug trials were still active in 2022 for targeting A<sub>2A</sub> as treatments for cancer (Imaradenant), pain (Spongiosine), Parkinson's disease (BIIB014, Taminadenant) and sickle cell disease (Regadenoson), for which the infusion of the A<sub>2A</sub> agonist reduces the production of IFN-γ and enhances the production of IL-13 and CD39 [18].

### 3. Mass Production of A<sub>2A</sub> Receptors

#### 3.1. Expression Systems

Owing to their finely regulated functions, GPCRs are naturally scarce in native biological sources. Accordingly, robust recombinant expression systems are needed to produce the milligram amounts required for their biophysical analyses. These systems should not only fit yield criteria but should also offer the most adapted environment for producing properly folded and functional GPCRs. Several systems have been developed with the aim of producing large amounts of the receptor properly folded with various degrees of sophistication and success rates. These include cell-free approaches [19,20], bacterial [21] or yeast [22–24] microorganisms, insect and animal cells using viral-infection or DNA-transfection strategies [25], photoreceptor cells of transgenic animals [26,27] and even whole organisms such as silkworm (*Bombyx mori*) larvae [28].

Among the many GPCRs investigated in these studies, A<sub>2A</sub> is remarkably one of the very few that has been successfully expressed at relatively high levels in nearly all recombinant systems. It was indeed produced in milligram amounts in *E. coli*, either unfolded in inclusion bodies [29], or as ligand-binding active receptors in the bacterial membranes [30–32]. A<sub>2A</sub> was also efficiently produced in the membranes of yeast cells, mainly *Saccharomyces cerevisiae* [33–35] and *Pichia pastoris* [22,36–39]. These hosts combine the advantages of handling the simplicity of microorganisms with the complex cellular machineries of eukaryotes. *S. cerevisiae* notably offers a large panel of molecular tools and engineering possibilities that were exploited for functional studies of many GPCRs [40] as well as for extensive screenings of stabilizing mutations and preparative conditions, including for A<sub>2A</sub> [41–46]. Concerning *P. pastoris*, its peculiar methylotrophic metabolism, the tightly regulatable expression conditions and the exceptional cell densities it can afford, make this yeast well suited for expressing membrane proteins for structural studies. Accordingly, it has not only allowed two of the high-resolution structures of A<sub>2A</sub> to be solved by X-ray crystallography [38,47], but it has also delivered various isotopically labelled samples for most of the NMR-based studies conducted on A<sub>2A</sub> [47–50].

Though technically more demanding and more time-consuming than microbes, animal cell systems offer a closer native-like environment and have therefore been extensively

used as recombinant hosts for A<sub>2A</sub> production. Most notably, recombinant baculovirus infecting either *Trichoplusia ni* (Tni) or *Spodoptera frugiperda* (Sf9) insect cells became the reference systems for generating crystallography-grade GPCR samples over the years. For A<sub>2A</sub> in particular, the majority of high resolution structures referenced to date were obtained from various forms of the receptor produced with this insect cell expression system [51–55]. Concerning mammalian cells, low expression levels and cost issues initially precluded the direct use of the high-quality receptors they produce for further biophysical analyses. Until recently, CHO or HEK 293 cell lines were thus mainly used as orthogonal recombinant systems for evaluating and validating the pharmacological properties of engineered or mutated A<sub>2A</sub> receptors that were designed for structural studies with other systems [39,51,56–59]. With the development of HEK 293-derived cell lines adapted to suspension and high cell density culturing in optimized media, combined with their transient transfection with tetracycline-inducible plasmids, GPCRs produced in these quasi-homologous host cells, including A<sub>2A</sub>, have now become accessible to thorough biophysical and structural studies [60,61].

### 3.2. Molecular Constructs

The design of A<sub>2A</sub> sequences that were expressed in these different systems are representative of the strategies classically employed for the large-scale production and purification of membrane proteins. The A<sub>2A</sub> coding sequence, sometimes optimized to fit the codon preference of the host cell [38], has been N- or C-terminally fused to various tags (most commonly His6, His8, His10 and FLAG) to enable detection and purification schemes, or to reporter proteins (GFP, RFP) [33,41,62,63] to monitor expression and purification optimization. These supplementary sequences were often separated from the A<sub>2A</sub> coding sequence by a cleavage sequence (TEV or 3C protease sequence) [22,62,64,65]. Finally, signal peptide sequences adapted to the expression host (from influenza hemagglutinin HA in animal cells, or from  $\alpha$ -Factor in yeast systems) were almost always added upstream of the fusion constructs to improve protein translocation in the ER and target the plasma membrane [22,33,51,66,67].

More specific modifications were further implemented in the A<sub>2A</sub> sequence when homogeneity and stability issues had to be addressed, notably for crystallography or cryo-electron microscopy (cryo-EM) structural studies. As further described in Section 8 below, besides a systematic truncation of the last C-terminal amino acids 317 to 412 [38,51,52], two complementary or combined sequence engineering strategies were employed to stabilize A<sub>2A</sub>. The first one consists of replacing flexible domains of the receptor with more structured and stable soluble proteins, the most common being the phage T4 lysozyme (T4L) [51,58,68] and the apo-cytochrome b562RIL (BRIL) [54,56,61,65,69,70]; however, other less-used proteins such as flavodoxin, xylanase and rubredoxin [71], as well as custom-made de-novo-designed  $\alpha$ -helical proteins have been successfully employed [43]. The second approach relied on the introduction of thermostabilizing point mutations, identified via directed-evolution [31,45] or structure-guided [55] approaches, allowing crystallographic studies [52,72] and other biochemical analyses [42,59,62]. An additional point mutation aiming at suppressing a glycosylation site at position N154 to improve receptor homogeneity was often introduced [38,47,52,72]. However, similar constructs bearing no such mutation also led to homogenous and crystallography grade samples [51,56,68]. Finally, for some specific NMR studies, several amino acids exposed at the intracellular surface were replaced by cysteine residues for targeted labeling with <sup>19</sup>F chemical probes [48,73–75].

## 4. Extraction and Purification

When the expression conditions are met, the next challenge before biophysical investigations of isolated membrane proteins consists of extracting and purifying the expected amounts of pure, homogenous, and correctly folded receptors. As for a large majority of GPCRs and other membrane proteins studied in solution [25,76], A<sub>2A</sub> was mainly solubilized using the mild detergent n-dodecyl- $\beta$ -D-maltopyranoside (DDM), independently

of the expression system used [30,38,62,63,67,77], or with the more recent lauryl maltose neopentyl glycol (LMNG) [47,48,50,61]. The physicochemical properties of these two surfactants are particularly suited for destabilizing biological membranes, while providing the receptors a reasonably stable hydrophobic environment to maintain their folding and function throughout their preparation and the planned analyses [78].

In the case of thermostabilized A<sub>2A</sub> sequences, the higher stability conferred by the selected point mutations allowed the solubilization of the receptors with n-decyl-β-D-maltopyranoside (DM), a harsher maltoside-based detergent. Compared to DDM, DM forms smaller micelles and has a higher critical micelle concentration, which was better adapted to further detergent exchanges and crystallographic studies using vapor diffusion crystallization conditions [52,69,72,79,80].

For the non-thermostabilized A<sub>2A</sub> constructs, the stability issues during solubilization and purification were addressed via the addition of specific ligands, locking the receptors into a limited number of conformations. Theophylline was the most frequently used [30,38,47–51,55,56,58,71,79–82], even if other ligands such as caffeine [69], DPCPX [77] and ZM241385 [61,63] appeared to also be useful. The stabilizing effect of these ligands [56,83–85] was very often combined with the supplementation of cholesteryl hemisuccinate (CHS), a cholesterol derivative that modulates the shape of detergent micelles in a pseudo-bicelle architecture. The addition of CHS into the purification buffers has been known to be beneficial for improving the stability of numerous solubilized GPCRs [86], including A<sub>2A</sub> [30,56,63,87]. Finally, other agents commonly applied to minimize the aggregation of membrane proteins in solution were also often added, such as glycerol which likely acts as an amphiphilic interface between hydrophobic domains and the polar solvent [88], and NaCl which contributes to maintaining proteins soluble in aqueous solutions through a “salting in” effect. Incidentally, sodium ions are also considered an allosteric modulator of A<sub>2A</sub> [89]. Iodoacetamide can also be used to prevent protein aggregation by intermolecular disulfide-bond formation.

The strategies used for purifying detergent-solubilized A<sub>2A</sub> were essentially based on affinity chromatography (AC) approaches. Since nearly all recombinant receptors were fused to a poly-histidine tag, an immobilized-metal affinity chromatography was most often performed as a first step, either on Co<sup>2+</sup> [50,51,63,67,90,91] or Ni<sup>2+</sup>-grafted resins [30,34,52,77]. Some procedures also involved agarose resins covalently bound to XAC, a non-specific, high A<sub>2A</sub> affinity antagonist at adenosine receptors [92], which were implemented as ligand-affinity columns in a first [30] or second purification step [49,50,93,94]. Recently, AC procedures using anti-tag antibodies coupled to agarose resins, such as M1 anti-FLAG [61], or anti-Rho-1D4 [59] antibodies, were also reported.

Purification of A<sub>2A</sub> has often been finalized by a polishing step on size exclusion chromatography (SEC) Superdex 200 columns, allowing the recovery of homogenous populations of monomeric receptors, and the elimination of additional impurities and specific compounds eluted from the previous AC step(s).

## 5. Isolation in Lipid Nanoparticles

As detailed in the previous section, a variety of conditions has allowed the isolation of A<sub>2A</sub> in detergent, in a sufficiently stable and convenient environment for a panel of biochemical and biophysical studies. However, the hydrophobic environment brought by detergents is chemically and physically distinct from the complexity of biological membranes, which poses certain limitations and issues for a number of investigations. Examples of such issues include the interference of detergents with certain analytical methods, insufficient stability for prolonged periods of analyses, absence of critical lipid and/or protein interactants for a receptor’s structure and function [95]. To overcome these difficulties, several methods have been developed over the years to reinsert detergent-isolated GPCRs in a more stable and more native lipid environment, including lipid-detergent bicelles, lipid bilayer nanoparticles or unilamellar lipid vesicles (ULVs) [95–97].



Bicelles are discoidal particles made of a planar phospholipid bilayer surrounded by a belt of detergent. While the reconstitution of several purified GPCRs in bicelles proved useful for their structural and functional investigations [98–100], none of these approaches have been reported so far for the characterization of A<sub>2A</sub>.

Concerning the implementation of ULVs, or liposomes, the situation is just slightly better documented. In an NMR study on A<sub>2A</sub> purified in detergent, liposome reconstitution was used as a quality control material to evaluate the functionality of the highly deuterated receptor for its capacity to activate its heterotrimeric G protein [67]. In another study, an original liposome synthesis method called INSYRT (in situ lipid synthesis for protein reconstitution technology) was recently developed and validated with A<sub>2A</sub> [101]. The receptor was solubilized and purified with DDM thioester analogs that served as precursors for the synthesis of phospholipids via a native chemical ligation (NCL) reaction. The resulting proteoliposomes were further characterized to contain A<sub>2A</sub>, showing ligand-binding pharmacological properties similar to those described in cellular membranes, and to the receptor reconstituted in the so-called nanodiscs (NDs).

Nanodiscs are nanometric discoidal lipid bilayers stabilized by two copies of a membrane scaffold proteins (MSP) deriving from the Apo-A1 lipoprotein [102]. Since it has been demonstrated that likely every detergent-solubilized membrane protein can self-assemble in such particles and maintain its structure and activity in a stable environment [103], NDs have been the prevailing source of material for the biophysical investigation of numerous GPCRs reconstituted in a lipid bilayer in recent times [104]. This is particularly the case for A<sub>2A</sub> for which the use of ND particles generated led to the discovery of a number of findings on receptor–ligand interactions and the structural dynamics of the receptor. The different approaches used included surface plasmon resonance (SPR) [62,105], mass spectrometry (MS) [106], miniaturized weak affinity chromatography (nanoWAC) [77], single molecule fluorescence microscopy [75] and ligand- or receptor-oriented NMR techniques [49,50,74,75,107]. In these studies, detergent-purified A<sub>2A</sub> was successfully assembled with different versions of the engineered MSP, either MSP1ΔH5 [50,74], MSP1D1 [62,80,106,107], MSP1E3D1 [49,77] or the less widespread Apolipoprotein A1 from zebrafish, Zap1 [105], which essentially differ in their molecular size and in the disk size of the generated NDs (from approximately 9 to 12 nm in diameter [103]). Some of these MSP were genetically [62] or chemically [77] modified to allow the grafting of the resulting NDs via their MSP on different supports for SPR and WAC analyses. The lipid composition essentially consisted of POPC:POPG mixtures incubated with MSP and A<sub>2A</sub> at various molecular ratios, in some cases replaced by or supplemented with other components used to evaluate their effect on the ND assembly efficiency [80] and/or on the receptor function [49,50,74].

Amphiphilic polymers, mainly amphipols, have also been largely used for isolating GPCRs in a membrane-mimicking environment [108,109]. While these particles have proven to be highly stable and allowed many investigations in detergent-free solutions, they are essentially generated after the reconstitution of proteins already purified in detergents. Thus, they may lack some lipids important for the conformational dynamic and function of the receptors. To date, no publication has reported on A<sub>2A</sub> reconstitution in amphipols.

Alternative amphiphilic co-polymers made of styrene and maleic acid (SMAs) are being increasingly used for the characterization of isolated membrane proteins, notably GPCRs, in lipid bilayers [97,110]. Unlike the standard amphipols, which are acrylate-based polymers grafted with octylamine, SMA compounds hold the ability, under defined conditions, to destabilize biological membranes and spontaneously form SMA lipid disc particles (SMALPs). Those containing the protein of interest can then be directly purified. A<sub>2A</sub> has been successfully isolated in SMALPs generated from the membranes of *Pichia pastoris* and HEK recombinant cells, and was characterized for its ligand-binding capacity and its increased stability compared to the receptor isolated in detergent [39]. Further studies reported by the same group investigated A<sub>2A</sub> SMALPs from *Pichia pastoris* membranes to

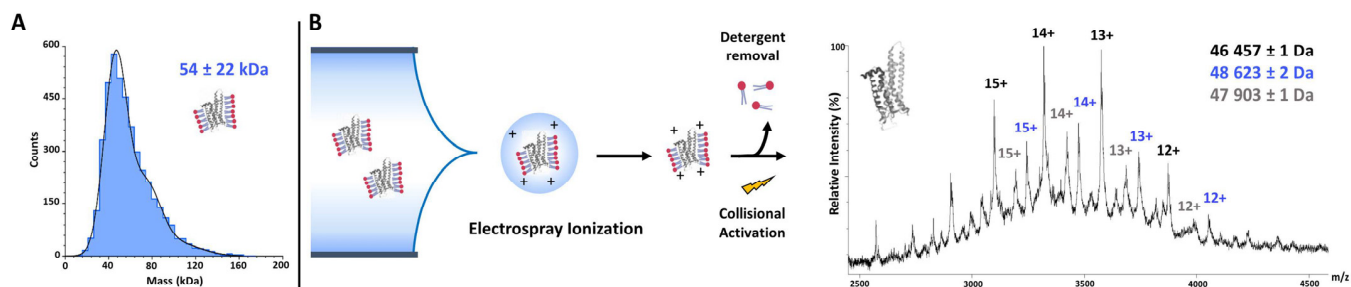
evaluate ligand-binding events via fluorescence correlation spectroscopy [111], as well as some ligand-induced conformational changes in A<sub>2A</sub> via fluorescence spectroscopy [112].

## 6. Quality Controls on Isolated Receptors

Once the protein material is isolated and purified, a panel of biochemical and/or biophysical tests are generally performed to assess the integrity of the receptor including the purity, homogeneity, stability and the activity of the isolated receptors. These are all important or even critical criteria to be met before initiating the functional and structural characterizations that are described in sections below.

Receptor integrity and purity are almost always assessed via classical SDS-PAGE analyses, sometimes complemented with A<sub>2A</sub>-specific immunodetection. The evaluation of sample homogeneity is often necessary to identify the presence of potential residual contaminant, but also to appreciate the oligomerization states of the receptor. This is essentially assessed through standard size-exclusion chromatography (SEC) analyses [35,51,71,94,106], or by discriminating the particle size either in the final sample via dynamic light scattering (DLS) [80] or during its purification via SEC-MALS [35,43,91,94]. Negative-staining EM imaging may also provide a useful qualitative complementary approach [61,77,91].

Native mass spectrometry (native MS) is often used as a first-line technique after the protein purification of soluble proteins to address oligomerization issues. This technique aims at transferring and detecting intact non-covalent interactions in the gas phase of a mass spectrometer. While native MS has been established for the analysis of soluble proteins and protein complexes, it is however challenging for membrane protein analyses [113,114], but has been successfully applied to A<sub>2A</sub>. Figure 1, panel B, represents a native mass spectrum of A<sub>2A</sub> solubilized in DDM, highlighting several species corresponding to different populations of receptors, including full-length, degraded or modified forms.



**Figure 1.** A<sub>2A</sub> mass measurement using mass photometry and native MS. Mass Photometry (A) is an emerging technique for fast mass and oligomeric state determination of proteins. Analysis of A<sub>2A</sub> showed a good homogeneity and enabled the determination of a mass of  $54 \pm 22$  kDa (molecular mass distribution histogram of A<sub>2A</sub>—the solid blue line represents major species fit with a Gaussian function). The quite large mass distribution obtained can be explained by the accuracy of the technique itself but also by the presence of potentially remaining micelles around the protein which may average out the overall mass obtained and thus increase the mass deviation. Native MS (B) is a powerful analytical tool for the study of membrane proteins embedded in detergent. The technique, preserving non-covalent interactions in the gas phase for mass analysis, enabled the visualization of the A<sub>2A</sub> protein with nano electrospray. Several populations (48.6 kDa, the expected mass of A<sub>2A</sub>; 46.4 and 47.9 kDa, truncated forms of the protein) have been identified after optimization of the instrumental parameters. These finely tuned and harsh conditions are essential for efficient detergent removal.

The implementation of mass photometry, a very recent label-free technique that quantifies molecular weights of biomolecules at the single-molecule level by interferometric detection of scattered light [115], provides an additional way for the characterization of membrane protein sample homogeneity [116]. In this context, we evaluated the technique for the analyses of A<sub>2A</sub> samples (Figure 1, panel A). The obtained results highlight the

complementarity of mass photometry and native MS for the characterization of membrane proteins. Requiring low amounts of starting material (approximately 100 ng protein) and no additional buffer exchange steps during the preparation, mass photometry can be used to obtain information on the sample studied with less optimization than native MS, providing heterogeneity and oligomerization information very quickly but with considerably less accuracy.

As mentioned earlier, the stability of the isolated protein is often critical for a number of analyses, in particular for crystallography studies or when the analytical methods require extended periods of measurement. For A<sub>2A</sub>, this issue has been mostly addressed via methods assessing the thermal stability of the isolated protein. Those were mainly fluorescence-based techniques, either involving the thiol-reactive chemical probe N-[4-(7-diethylamino-4-methyl-3-coumarinyl)phenyl]maleimide that covalently binds to internal cysteines and fluoresces upon thermal denaturation [55,56,71,83], consisting of a fluorescence size-exclusion chromatography (FSEC) evaluation that requires the presence of a fluorescent protein (i.e., GFP) fused to the receptor [62], or label-free tryptophan fluorescence monitoring upon unfolding [117]. A<sub>2A</sub> thermal stability was also assessed via the analyses of migration profiles of the protein on acrylamide gel after denaturation, either revealed via Western blotting [84] or via clear-native PAGE [63]. As an alternative to thermal denaturation, an isothermal chemical denaturation method using guanidium chloride has been successfully applied to assess the stability of A<sub>2A</sub> when isolated in the presence of various ligands [90].

Perhaps the most important indication of the quality of a protein sample relies on its capacity to fulfill its function. A variety of techniques have been applied to assess the activity of A<sub>2A</sub> via its ability either to bind specific ligands or to activate its associated heterotrimeric G-protein partner upon agonist stimulation. These methods include radioligand binding assays in filtration [36,77,91] or scintillation proximity assay (SPA) formats [62,105], GTPγS activity assays [67], fluorescence analyses [111], and a panel of SPR-, MS- or NMR-based approaches, which are described in more detail in the following sections.

## 7. Functionality of the Isolated A<sub>2A</sub>

### 7.1. Ligand Binding

#### 7.1.1. Radioactive Ligand Binding

As far as specific radiolabeled compounds are available, for decades, this direct evaluation of receptor–ligand interactions has been the preferred technique to study and define the pharmacology of GPCRs. It is a highly sensitive approach based on the counting of the radiation emitted by the radioactive compounds bound to their receptors, from which are deduced precise affinity parameters either in saturation or competition modes. In the most classical methods, receptor-bound and free radioligands are separated via filter-based techniques that are particularly well suited to evaluating GPCRs in membrane fraction samples. When GPCRs are solubilized and isolated, standard filter-based assays are generally not compatible with the small size of the particles to be analyzed, so alternative separation methods were needed. For A<sub>2A</sub>, these mainly relied on the use of mini-spin gel filtration columns [30–32,36,39,49,118,119], on the filter-based separation of the receptor–ligand complexes either precipitated with polyethylene glycol (PEG) [77,91] or immobilized on agarose beads [34,83], or by immobilization on functionalized scintillating bead supports designed for SPA [47,62,77,79,105]. These studies were essentially conducted to characterize and validate the pharmacology of A<sub>2A</sub> isolated either in detergent micelles [30,31,36,39,49,91], in NDs [49,62,77,105], in SMALPs [39] or in liposomes [101]. Interestingly, one study conducted on different A<sub>2A</sub> samples generated from *P. pastoris* or HEK cells, either in membrane fractions or isolated in detergent or in SMA particles [39], determined comparable affinity values for several agonist and antagonist ligands. In addition, this work demonstrated the increased stability overtime of SMALP or ND samples over detergent micelles on the ligand-binding activity of the isolated receptor [39].



Radioactive ligand binding has also been the key method for evaluating the thermal stability of solubilized A<sub>2A</sub>, either for screening thermostabilizing mutations [31,52] or for evaluating the stabilizing effect of various detergents and additives [119].

#### 7.1.2. Surface Plasmon Resonance

Several alternative biophysical methods, such as SPR, have emerged to determine binding affinities and kinetics, as they are essential for drug discovery and development [120]. Indeed, the kinetics of drug–receptor complex formation, and in particular the residence time of the ligand [121], is a crucial element that affects the functional properties of a ligand and its pharmacological profile [122–124].

SPR is a label-free technique that allows the quantitative analysis of molecular binding. One binding partner is immobilized on a biosensor surface and the other is introduced into a microfluidic system and flows in solution over the surface. The binding causes a small change in the refractive index at the biosensor surface due to the accumulated adsorbed mass. Continuous monitoring of this signal allows the determination of the kinetic parameters such as the association rate constant ( $k_{on}$ ), the dissociation rate constant ( $k_{off}$ ) and the affinity ( $K_d$ ) of the interaction [125].

SPR has become the method of choice for studying ligand binding to various GPCRs [126–129]. This technique requires small amounts of membrane proteins in a native or similar environment. The challenges are to maintain high levels of binding activity of the immobilized receptor and to detect the binding of low-molecular-weight ligands, as this technology is based on mass changes [130–132]. This has been successfully applied to assess the ligand-binding affinities of a number of GPCRs. A variety of A<sub>2A</sub>/ligand interactions have been studied using SPR over the past decades. Findings from these studies illustrates how SPR technology supports structure-based ligand discovery and fragment screening and contributes to the better understanding of the molecular aspects of signaling. Detergent-solubilized A<sub>2A</sub>, either non modified or thermostabilized (A<sub>2A</sub>-StaR), has been widely investigated and generally extracted from insect cells membranes. Several A<sub>2A</sub>-StaR (purified or as crude solubilized extracts) were captured through their C-terminal 10His tag to a Ni-loaded NTA sensor surface (carboxymethylated dextran functionalized with nitrilotriacetic acid) [90,131,133–135]. Small antagonists (molecular weight ranging from 285 to 345 Da) were shown to interact with a  $K_d$  ranging from 0.3 to 160 nM in accordance with the classical competition radioligand binding assay [133]. Importantly, from the screening point of view, SPR is able to detect even weaker GPCR/compound complexes ( $K_d > 1 \mu\text{M}$ ) [131]. The groundwork was laid for the cross-screening of a panel of antagonists with mutated A<sub>2A</sub>-StaR. The resulting matrix of binding kinetics data was then combined with molecular modelling and docking data from a refined map of the A<sub>2A</sub> binding site, thereby facilitating the structure-based drug design [135,136]. Affinity and kinetic rate constants measured at different temperatures allow the calculation of enthalpic and entropic energy components as well as the analysis of kinetics of the transition state, constituting the thermodynamic signature of a target-ligand complex [137–139]. Calculating the kinetic transition state components for six ligand–A<sub>2A</sub>–StaR complexes and combining them with supervised molecular dynamics (MD) and metadynamics approaches clarified the crucial role of receptor and antagonist solvation/desolvation in the transition states of entropic nature. Thermodynamics combined with kinetic analyses allows a better understanding of the details of the molecular interactions. They could also help rationalize the drug design since kinetics better correlates with EC<sub>50</sub> than thermodynamic equilibrium affinity constant values for some GPCR–ligand complexes [138].

Solubilized GPCR micelles are unstable and tend to denature and/or aggregate over time. As detailed previously in Section 5, several strategies have been developed to reconstitute GPCRs in a lipid bilayer, such as proteoliposomes or high-density lipoproteins known as NDs, to provide more stable conditions for the receptor compared to detergents. The kinetic characterization, using SPR, of the binding of nine adenosine antagonists to a

A<sub>2A</sub>-StaR reconstituted in NDs correlates well with those of the same receptor in a membrane environment obtained from the conventional radioligand binding assay. NDs likely correspond to a more native environment than detergent and may avoid the problems of ligand partitioning in micelles [105]. Bocquet et al. conducted different binding assays (SPA and SPR) with the A<sub>2A</sub>-StaR in membranes, in NDs and in detergent micelles [62]. Different set-ups were evaluated in the SPR assay, with NDs being immobilized via different tags fused either on the receptor or on the MSP of the NDs, without impacting the binding kinetics of small antagonists (337 to 428 Da). In addition, the receptor was shown to be more stable in NDs than in detergent micelles.

The SPR approach has also been validated for fragment screening investigations on A<sub>2A</sub> (see Section 9). The success of such experiments requires a high density of immobilized GPCRs with high binding capacity, as well as careful experimental design, to reduce false positive results since high concentrations of fragments are tested and tend to non-specifically bind to many targets (reviewed in Shepherd et al. [132] and Coyle and Walser [140]). This approach was first developed for a solubilized A<sub>2A</sub>-StaR with a panel of 70 fragments derived from xanthine, with molecular weights ranging from 136 to 194 Da. [134]. SPR was also used to validate putative ligands of a A<sub>2A</sub>-StaR issued from a mass spectrometry fragment screening [141]. In another study, the native, non-stabilized A<sub>2A</sub> receptor, as well as three other receptor subtypes from the same family (A<sub>1</sub>, A<sub>2B</sub> and A<sub>3</sub>), were investigated via SPR. [142].

As GPCRs interact with different intracellular partners of the downstream signaling cascade, SPR was also used to study some of these molecular interactions. It was notably shown that the binding of solubilized A<sub>2A</sub> to immobilized G $\alpha$  subunit proteins was affected by GDP analogues and that the C-terminus of A<sub>2A</sub> was necessary for its high-affinity association to this G-protein subunit [143]. In another study on A<sub>2A</sub> in NDs bound to a full or a partial agonist, the binding characteristics for the heterotrimeric G $\alpha\beta\gamma$  proteins were determined using SPR [50]. No differences in affinity and kinetics were observed, but complementary structural analyses using NMR suggested a model based on conformational changes in the receptor to explain their different functional output. Other aspects of A<sub>2A</sub> functioning have been explored via SPR, such as the impact of mutations in a cholesterol consensus motif on G-protein binding [59].

### 7.1.3. Calorimetry

Isothermal titration calorimetry (ITC) is a label-free biophysical technique that can measure kinetics and the thermodynamic parameters of binding, such as affinity, stoichiometry, enthalpy and entropy between macromolecules (e.g., between two proteins) or between a macromolecule and its small molecule ligands [144,145]. Applied to GPCRs, ITC allows for a deeper understanding of the binding interactions. Indeed, it reflects the subtle structural and dynamic changes between bound and unbound forms, identifying the most important regions of the binding interface [146,147] as well as receptor–ligand specificity, cell surface expression, endocytosis and recycling [148]. A<sub>2A</sub> is of particular interest because it possesses a long and flexible C-terminal region which favors interactions with several proteins, including G-protein-coupled receptor kinases,  $\beta$ -arrestins,  $\alpha$ -actinins, calmodulin (CaM), ubiquitin-specific protease 4 (USP4), and neuronal calcium sensor protein 1 (NCS-1) [149–156]. The affinity and stoichiometry of A<sub>2A</sub> binding to two of its partners, CaM and  $\alpha$ -actinin 1, have been recently studied together with their regulation by calcium using ITC [157,158]. In this study, human CaM and two A<sub>2A</sub> C-terminal regions of different lengths (A<sub>2A</sub>-ctL comprising 293–412 amino acids and A<sub>2A</sub>-ctS comprising 321–412 amino acids). CaM was titrated into the protein in the presence of calcium with or without the addition of EDTA. In the absence of EDTA, the binding between A<sub>2A</sub> and CaM displayed strong affinity ( $K_d = 97.9$  nM) with a 1:1 stoichiometry. In the presence of calcium, the binding of CaM to A<sub>2A</sub>-ctL led to an exothermic reaction with a favorable enthalpy, but unfavorable entropy, possibly indicating conformational changes. No binding occurs in the presence of the calcium chelator, EDTA. The binding site of CaM on the A<sub>2A</sub>-ctL

region was further identified using the A<sub>2A</sub>-ctS construct lacking the amino acids 293 to 320. CaM binding in the presence of calcium led to small exothermic peaks signalling a loss of affinity. These results, confirmed by NMR spectroscopy, identified the residues 293–310 as the binding epitope of CaM onto A<sub>2A</sub> [158].

Similarly, mutating arginine residues in this region abolished CaM binding. These data are in agreement with previously published results [151].  $\alpha$ -Actinin binding to A<sub>2A</sub> was suspected to be implicated in the internalization of the receptor after agonist activation. To characterize the domain responsible for the interaction between the C-terminal domains of A<sub>2A</sub> and  $\alpha$ -actinin 1,  $\alpha$ -actinin1 wild-type constructs limited to the calcium binding domain (CABD), the rod domain or the CABD-rod domains were used. In the absence of calcium, the titration of the rod-CABD domains led to exothermic peaks and to a saturable binding curve. The affinity of rod-CABD domains to  $\alpha$ -actinin was found to be in the micromolar range ( $K_d = 39 \mu\text{M}$ ), which was weaker than for A<sub>2A</sub>-ctL with CaM. Together, the results of the ITC experiments suggest that the A<sub>2A</sub> C-terminus binds to the distal part of  $\alpha$ -actinin 1 C-terminal domain, either in the CABD or at the subsequent C-terminus, a region shown to interact with CaM, and in fact it has been demonstrated that these proteins compete when binding to A<sub>2A</sub>. It is worth highlighting that ITC is a classical approach to identify GPCR partners and should be systematically used to validate and characterize receptor partners at the molecular level.

#### 7.1.4. Receptor Fluorescent Labeling

Many different techniques have been applied to illuminate the molecular bases of trans-membrane signaling through GPCRs in general, and through A<sub>2A</sub>, in particular. Among them, fluorescence-based studies appear to be particularly adapted to investigate the functioning of these receptors, as they can be used in many different environments ranging from native and recombinant cell lines to isolated receptors in membrane-mimicking systems [159]. These techniques nevertheless require GPCRs to be specifically labelled with adapted fluorophores. For the last two decades, several approaches have been developed to label receptors for fluorescence-based applications. Besides the fluorescent ligands described later (Section 9), these include protein and peptide tags compatible with measurements in live cells (e.g., GFP, YFP, SnapTag and FAsH-tetracysteine tag) [160]. Several of these labelling strategies have been applied to the A<sub>2A</sub>. For instance, the concomitant insertion of a Cyan Fluorescent Protein and the FAsH-tetracysteine tag into human A<sub>2A</sub> was combined with fluorescence resonance energy transfer (FRET) measurements to provide a description of the activation process of this receptor in cellular model systems [161]. More recently, a covalent ligand was used to couple a fluorophore to A<sub>2A</sub> [162]. Although this latter approach was used in a cell-based system, it could be extended to isolated receptors as well. Besides cell-based studies, strategies to label purified receptors in detergent micelles or nanodiscs have also been developed. In many cases, the insertion of the fluorescent probe primarily involves the labelling of unique reactive cysteines with a fluorescent probe through a maleimide-based reaction. This strategy was applied to recombinant A<sub>2A</sub> to successfully introduce probes for NMR- and fluorescence-based applications [75]. For the latter, cyanine 3 was attached to the purified receptor expressed in *P. pastoris* and assembled into nanodiscs [163]. This opened the way to single-molecule fluorescence experiments aimed at describing the conformational transitions of A<sub>2A</sub>. In the same way, a purified mutant receptor containing single reactive cysteines was labeled with an environment-sensitive dye, BODIPY-FL, for fluorescence quenching experiments aimed at delineating the dynamics of the TM6 domain of A<sub>2A</sub> [164]. In addition, orthogonal labeling strategies were also described in this same study [164]. Specifically, a double-cysteine mutant with reactive Cys residues on TM4 and TM6 was used for double-labeling with a fluorescence donor (Alexa Fluor 488) and acceptor (Alexa Fluor 647), paving the way for single molecule-FRET analyses of receptor conformational fluctuations. In addition to these classical protein engineering approaches, alternative labeling strategies have been described for A<sub>2A</sub>. In particular, a modified aminoacyl-tRNA-synthetase and a suppressor tRNA were used in

a cell-free system to incorporate an unnatural amino acid that was further coupled to a fluorescent dye [165]. Together, these studies provide evidence that the isolated adenosine A<sub>2A</sub> receptor in different membrane-mimicking environments can be efficiently labeled with fluorophores, providing a convenient system to describe its energy ensemble and the modulations by the environment using fluorescence-based techniques.

#### 7.1.5. Ligand-Based NMR

Ligand-observed NMR experiments are widely used to measure and analyze small molecule binding to soluble and membrane proteins such as GPCRs [166]. No protein labeling is required, and experiments can be performed in micelles, membranes and nanodiscs. Ligand-observed NMR experiments are suitable for moderate- to low-affinity ligands (10 nM to mM affinities). This is of particular interest as far as screening approaches are concerned, as the net results of most of the screening campaigns are poor affinity binders. Nevertheless, the binding of higher-affinity ligands can be monitored through competition experiments, using the NMR binding signal of a moderate-affinity compound as a reference signal. Saturation transfer difference (STD), nuclear Overhauser effect spectroscopy (NOESY) experiments or <sup>19</sup>F NMR on labeled ligands have been reported on A<sub>2A</sub> [74,91,107]). Experiments are typically compared to a control experiment where the receptor is either missing or blocked by a high-affinity ligand. The NMR experiments allow the identification of novel ligands and of their binding pockets when a reference compound at a known binding site is available. Furthermore, they may reveal other binding pockets on A<sub>2A</sub>. For example, the binding of adenosine in an allosteric pocket was observed through STD experiments recorded with A<sub>2A</sub> bound to ZM241358 [91].

Three-dimensional structures of A<sub>2A</sub>–ligand complexes can be calculated from NOESY experiments called INPHARMA (Interligand Noes for PHarmacophore Mapping) [107,167]. For example, the structure of the complex between A<sub>2A</sub> and 3-pyrrolidin-1-ylquinoxalin-2-amine (PQA) was obtained from the interligand NOESY observed between PQA and the antagonist compound ZM241358 [107].

Finally, ligand-observed NMR can be used to investigate allosteric mechanisms, as positive and negative allosteric modulators (PAMs and NAMs), respectively, increasing or decreasing the binding of a ligand and thus its NMR binding signal. Recently, Huang et al. [74] investigated the binding of cholesterol on A<sub>2A</sub> using <sup>19</sup>F-labeled cholesterol analogs. They observed the broadening of the NMR resonances as well as a 0.5 ppm up-field shift for the <sup>19</sup>F-labeled cholesterol bound to A<sub>2A</sub> in nanodiscs compared to empty nanodiscs. However, the NMR spectrum was not modified upon the addition of an inverse agonist, full agonist, or a G-protein mimetic, suggesting that the cholesterol interactions with A<sub>2A</sub> are transient and unspecific, and that the effect of cholesterol is due to changes in the membrane bilayer properties [74]. The investigation of the allosteric modulation of A<sub>2A</sub> by small molecules using ligand-observed NMR represents a rapid and robust approach that should be further used to explore the biology of A<sub>2A</sub>.

#### 7.1.6. Native Mass Spectrometry

As described above (see Section 6), native MS is often used to check the quality of a purified protein and to address oligomerization issues. In addition, native MS is also a valuable method to evaluate molecular interactions between the protein and various partners. Yen et al. [168] used native MS to reveal that A<sub>2A</sub> in its apo state still binds to endogenous phosphatidylserines (PS) and phosphatidylinositol (PI). Native MS was next applied for the affinity ranking of such compounds, demonstrating that PI(4,5)P<sub>2</sub> is a better binder compared to other phospholipids during binding assays [168]. Even more challenging is the native MS detection of a heterotrimeric G-protein complex involving the monomeric A<sub>2A</sub>, the Gα subunit and a nanobody to stabilize the entire assembly [168]. The lipids bound to this receptor complex were measured after quadrupole selection and collisional activation of the whole complex. To obtain such high-quality, high-resolution native MS data of GPCRs, all steps of the native MS workflow must be carefully optimized,

from sample injection using nano-emitters, to MS methods. Furthermore, an optimization of collisional conditions is required to allow efficient desolvation and membrane-mimic removal while maintaining an intact membrane for protein analysis. Comparisons between ligand-binding native MS results obtained from classical  $\text{NH}_4\text{OAc}$  buffer and  $\text{NaCl}$ -based buffers on  $\text{A}_{2\text{A}}$  and glucagon receptors in the presence of different ligands demonstrated that ligand-binding interactions are better conserved on GPCRs using  $\text{NaCl}$ -based buffers than  $\text{NH}_4\text{OAc}$  buffers [169]. For  $\text{A}_{2\text{A}}$ , the authors could differentiate agonist from antagonist compounds based on the capacity of the receptor to retain sodium adducts from the storage buffer, thus paving the way for the use of native MS screenings on GPCRs for the detection and characterization of non-covalent interactions under more relevant physiological conditions.

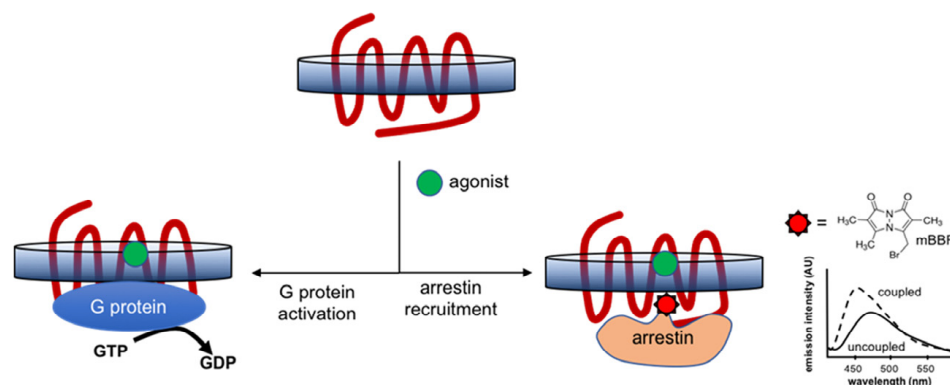
## 7.2. Activity

### 7.2.1. G-Protein Coupling and Arrestin Recruitment

Advanced Resonance Energy Transfer (BRET/FRET) strategies have been developed during the last decade to monitor receptor–effector coupling and selectivity in signaling. These approaches have been extensively reviewed [170,171] and have been mostly used with recombinant cell systems expressing receptor and effectors labeled with fluorophores and/or fused to fluorescent/bioluminescent partners or fragments. In the case of isolated receptors, apart from the purification of stable signaling complexes for subsequent structural studies, *in vitro* assays monitoring the coupling to signaling partners are also currently used to assess the functional properties of the isolated GPCRs, in parallel to ligand-binding assays. Initially, these assays focused on the allosteric stabilization of the active state of the receptor and its impact on an agonist's affinity and/or receptor stability. Such assays were used, for instance, to reveal the coupling of the isolated adenosine receptor to engineered mini-G proteins [53]. Specifically, mini-Gs were shown to increase the affinity of the purified  $\text{A}_{2\text{A}}$  in detergents for NECA, and significantly increased the stability of the receptor in the presence of the agonist. Besides these assays, experimental designs have been implemented to monitor receptor-catalyzed G-protein activation through GTP turnover (Figure 2). This process can be visualized by monitoring the binding of a radiolabeled [ $^{35}\text{S}$ ]GTP $\gamma\text{S}$  [172] or the fluorescent analog of GTP (Bodipy-GTP $\gamma\text{S}$ ) [173] to the  $\text{G}\alpha$ -subunit of the purified  $\text{G}\alpha\beta\gamma$  heterotrimer. [ $^{35}\text{S}$ ]GTP $\gamma\text{S}$  binding was used to decipher the G-protein coupling to  $\text{A}_{2\text{A}}$  in Sf9 membranes [174]. Besides membrane fractions, labeled GTP $\gamma\text{S}$  binding assays can also be applied to GPCRs isolated in detergent micelles [175,176] or nanodiscs [177,178]. More recently, a GTP turnover assay has been devised that monitors the amount of free GTP after receptor-catalyzed G-protein activation. Initially described for other GPCRs such as the  $\alpha 2$ -adrenergic [179] or ghrelin [180] receptors, this assay has been recently used to reveal the allosteric modulation of the adenosine  $\text{A}_{2\text{A}}$  receptor in nanodiscs by cholesterol [74].

Regarding arrestin coupling, although some alternative assays have been reported that are based, for instance, on the use of radiolabeled arrestin [181], a popular *in vitro* assay with isolated GPCRs relies on the use of recombinant arrestin labeled on its finger loop with a specific fluorophore, monobromobimane (mBBR) (Figure 2). This fluorophore is attached to recombinant arrestin through a unique reactive cysteine [182]. The emission properties (intensity, maximum wavelength) of mBBR vary upon coupling to the receptor, and as such, allow for visualization of the GPCR–arrestin interaction. Initially developed with rhodopsin and visual arrestins [183], this assay has since been extended to other GPCRs such as the vasopressin,  $\beta 2$ -adrenergic or ghrelin receptors in detergents, amphipols and nanodiscs [178,184–186]. To the best of our knowledge, this assay has not been used with purified  $\text{A}_{2\text{A}}$  so far but could be directly applied to this receptor isolated in membrane-mimicking systems, allowing, for instance, screening for particular ligands that would selectively trigger arrestin recruitment.





**Figure 2.** Schematic representation of the in vitro assays for G-protein activation and arrestin recruitment. These assays monitor receptor-catalyzed G-protein activation through GDP-to-GTP exchange by measuring either the binding of a labeled GTP analog or the depletion of GTP in the solution. To study arrestin recruitment, the most popular assay relies on monitoring the changes in the emission profile of a fluorophore, monobromobimane (mBBR), attached to the finger loop of recombinant arrestin.

It is clear from the last years in GPCR biology that our understanding of receptor signaling pathways has been completely changed by two concepts: (i) the description of the signaling bias of receptors [187–189] and (ii) the numerous proteins that have been found associated with receptors, such as the MT<sub>1</sub> receptor [190]. For the first point, isolated systems are necessary to better understand how the activation of receptors are controlled. For the second point, an understanding of the activation routes of a receptor will be clearly defined from isolated systems for which potential coupling between purified receptors and their protein binding partners will be studied in the defined environment of nanodiscs.

#### 7.2.2. Functionality of Isolated Receptors: Activity of Receptor Dimers and Oligomers

Even though a monomeric A<sub>2A</sub> receptor is sufficient to promote G-protein activation or  $\beta$ -arrestin recruitment, this receptor, like many other GPCRs, has been shown to form dimers and larger oligomers [191–194]. GPCR oligomers offer additional inter-receptor communications to potentially regulate their own functional properties. A<sub>2A</sub> receptor was shown to associate with itself [195], forming A<sub>2A</sub> homodimers, but also with other receptors to form heterodimers or larger oligomers. In particular, A<sub>2A</sub> could form heterodimers with the other adenosine receptors A<sub>1</sub>, A<sub>2B</sub> and A<sub>3</sub> [196,197], as well as with the cannabinoid CB<sub>1</sub> receptor [198], the dopamine D<sub>2</sub> and D<sub>3</sub> receptors [199–202] or the glutamate mGlu5 receptor [203]. Most of these studies were performed in heterologous systems, mainly using BRET or FRET approaches. Among the heterodimers, A<sub>2A</sub>/D<sub>2</sub> has gained a lot of attention. First, its existence is well documented in vivo both in animal models [204] and in the post-mortem human brain [205]. Second, A<sub>2A</sub>/D<sub>2</sub> heterodimers are located in the GABAergic striatopallidal neurons, a region critical in locomotor activity and central in the physiopathology of Parkinson's disease [206]. Interestingly, A<sub>2A</sub>/D<sub>2</sub> heteromers were shown to operate a reciprocal negative functional crosstalk and the interaction between the two receptors is reduced along the development of the pathology [206,207]. Therefore, A<sub>2A</sub>/D<sub>2</sub> constitutes a major therapeutic target for the treatment of Parkinson's disease.

While cellular and in vivo studies provide a general picture of the functional crosstalk within GPCR oligomers [208] (whether homodimers, heterodimers or heteromers) including A<sub>2A</sub>/D<sub>2</sub>, a precise characterization of the pharmacological, conformational or signaling specificity of each oligomer relative to the monomers or other oligomers is very difficult to reach in these complex systems where several oligomeric species are likely to co-exist. Therefore, isolated systems where a single receptor combination is present, either monomer or oligomer, bring invaluable information to elucidate the molecular, pharmacological and signaling signatures. Additionally, in isolated systems, specific mutations (e.g., non-binding

or non-coupling mutations) can easily be introduced to a given protomer. They can analyze the overall effect of the absence of the oligomer function. To date, a single publication reported A<sub>2A</sub> oligomers in isolated systems [94]. However, other GPCR oligomers have been analyzed in such systems illustrating the power of the method but also highlighting the technical challenges to overcome [197,209].

As indicated above, purified receptors can be obtained either in detergent micelles, or reconstituted in proteoliposomes or in lipid nanodiscs. Depending on the protein concentration, isolation procedures and purification steps, samples containing the desired receptor composition are obtained. For example, the neurotensin NTS1 receptor is mostly monomeric in detergent at low protein concentrations and becomes dimeric at higher concentrations [210]. Leukotriene BLT2 receptor has been shown to be present in a mixture of monomers and dimers with each species successfully isolated after HPLC fractionation [176]. For nanodiscs, the incorporation of monomeric or dimeric GPCRs can be achieved by acting on the receptor-to-scaffold protein ratio as reported for rhodopsin [211]. Alternatively, the reconstitution of the purified glutamate mGlu2 receptor into nanodiscs has been shown to lead to a mixture of discs containing monomers or dimers that can be isolated by HPLC fractionation [212]. Upon reconstitution, the dimer can assemble in parallel or anti-parallel orientations, with only the former having a physiological relevance. A general assessment of the relative orientation can be conducted using fluorescently labeled protomers either at the N- or C-terminus. Indeed, FRET would occur only at a short distance, i.e., when both fluorophores are located at the same side of the receptor relative to the lipids. Thus, if FRET is detected between probes inserted at the N- and C-terminus, anti-parallel dimers are present [176]. To overcome this phenomenon and to purify only the parallel dimers from a mixture, an original strategy has been developed using the stable CaM binding simultaneously to two similar sequences derived from the CaM binding domain from *Petunia* glutamate decarboxylase (PGD). Accordingly, after the fusion of the 26-residue PGD-tag to the N-terminus of the receptor of interest, only parallel dimers can be purified using a CaM binding step followed by SEC. This method was efficiently used on lipid nanodiscs containing LTB1 receptor dimers or ghrelin GHSR receptor dimers [213]. When considering heterodimers, their formation in detergents or in nanodiscs gives a mixture of several species such as monomers, homodimers and heterodimers. In order to select the heterodimers from this mixture, double tag purification strategies have been developed. Indeed, the fusion of a different tag to each receptor of interest, followed by two successive purification steps, each using one of the tags, lead to the purification of only dimers presenting one of each tag [214]. This method has been used to prepare BLT1 receptor dimers in detergent where one of the subunits only is mutated in its binding site for coupling with a G protein [214]. It is worth noting that some receptors assemble in complexes larger than dimers. Therefore, nanodisc size is not compatible with the insertions of these large complexes. To overcome this limitation, proteoliposomes are an alternative that have been used to reconstitute a heterotetramer composed of two GHSR and two D2 receptors [215]. The heterotetramers were further isolated from the proteoliposomes using SMALPs.

Following the isolation of receptor complexes, functional and conformational experiments can be performed similarly to those carried out on isolated monomeric receptors, highlighting the specific properties of the different complexes. For example, in a rhodopsin dimer, only one of the protomers is in a high-affinity state upon recruitment of G protein while the dimers have a lower affinity for  $\beta$ -arrestin 1 than the monomer [211,216]. The analysis of the GHSR/D2 heteromer highlights that heteromerization directly modulated D2-mediated G $\alpha$ i protein activation. More specifically, such heterocomplexes act directly on the conformation of the G $\alpha$ i subunit [215]. Applying similar strategies to the analysis of the A<sub>2A</sub> homodimer or A<sub>2A</sub>/D<sub>2</sub> heteromer analysis would bring important information on the precise mechanism underlying the function of the complexes and would lead to new hypotheses of how to more precisely regulate these pathophysiologically relevant heteromers.

## 8. Structure of A<sub>2A</sub>

The early evidence for a necessary transfer of information from structure to function has been reviewed by Piirainen et al. [217] and clearly showed an early path for the rationalization of GPCR structure determination to gain an understanding of the function of the A<sub>2A</sub> receptor and GPCRs in general. The central role of biophysical techniques in the determination of the structure and function of GPCRs has been nicely reviewed by Langelaan et al. [218]. Furthermore, artificial intelligence approaches, such as with AlphaFold, are powerful tools to predict three-dimensional protein structures and protein–protein interactions. This technology can be accessed for the prediction of the human A<sub>2A</sub> three-dimensional structure, in addition to almost all other GPCRs [219,220].

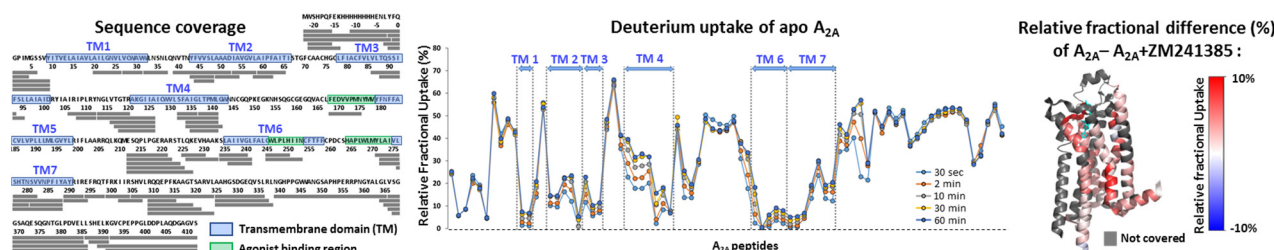
### 8.1. Structural Mass Spectrometry Approaches

Among the large panel of biophysical methods available, structural MS techniques have become a valuable tool for either ligand screening or for more detailed structural characterization of purified GPCRs, including the A<sub>2A</sub> receptor. The structural MS toolbox encompasses a series of techniques, such as native MS for stoichiometry and oligomerization determination, ion mobility MS for the assessment of global conformation, and labeling methods such as hydrogen/deuterium exchange MS (HDX-MS) and chemical cross-linking to investigate interacting regions and/or conformational changes [221,222].

While native MS and mass photometry have great capacity for the characterization of oligomeric state or binding events (see Sections 6 and 7 above), both techniques are not adapted to provide either amino acid resolution information on binding regions, or the dynamics of assembly or disassembly [113,114]. In recent years, HDX-MS has become a very powerful labeling method for membrane protein analysis, adapted to tackle conformational issues related to ligand binding and even allostery [223]. A typical HDX-MS experiment includes the incubation of a protein of interest in two or more states (apo- or holo-states for example) in deuterated buffer at different time points to allow hydrogens from backbone amides to exchange with deuterium. The exchange reaction is then quenched by switching to acidic pH and low temperature followed by online digestion with acid-functional proteases. The generated peptides are then separated on a reversed phase-HPLC system and detected using MS. Dynamics and binding-site mapping are then extracted by comparing apo- and holo-states. While HDX-MS is becoming a well-established technique for soluble proteins with standardized workflows and protocols [224], the study of membrane proteins remains technically challenging. Thus, extensive work on the optimization of the experimental conditions (composition of buffers, choice of digestion protease, etc.) must be carried out systematically. For instance, nepenthesin protease seems to be particularly suited for membrane protein studies compared to the standard pepsin one. Similarly, the addition of urea as denaturant in quench buffers seems to be beneficial when working with membrane proteins [225]. Other dedicated workflows and methodological developments have been set up to prevent the early deterioration of the instruments caused by the large amounts of detergents and phospholipids in the samples, including an additional off-line cleaning of the LC columns, or the use of zirconium beads for lipid depletion, etc. [226,227].

Figure 3 presents a typical HDX-MS workflow used to monitor the binding of the antagonist ZM241385 on A<sub>2A</sub> and the induced conformational changes. Online digestion of the protein allowed to cover 70.8% of the sequence with 79 identified peptides and a redundancy of 2.7, due to the use of nepenthesin II protease, was reported as more efficient on GPCRs. Experiments carried out on apo A<sub>2A</sub> showed a low deuterium incorporation on transmembrane domains, reflecting their known low accessibility to the solvent. Among these domains, some peptides (20–31; 58–62; 86–93; 134–141; 234–241; 245–253; 271–280) exhibited a dramatically low deuterium incorporation typically less than 10%, which is consistent with the known structure of the protein. Upon ZM241385 binding, a significant part of the protein was protected meaning that A<sub>2A</sub> incorporates less deuterium in the presence of the ligand. Several peptides exhibited statistically significant differences in deuterium incorporation (*p*-value of 0.01). Peptides 168–175, 245–250 and 245–253 in the known ligand

binding region [38] (F168, N253 and I274 residues) presented a H/D protection higher than 5% confirming the ZM241385 binding site. The significant protection of additional peptides 59–65, 276–288, 277–282 and 277–288 supports the known ligand fixation, as involved in the following residues (A63, H278), forming a cleft to accommodate the phenol group of ZM241385, as well as for the end of the transmembrane domain 7 (TM7) represented by residues 281 to 298 [72]. Other regions showing differences in incorporation (TM3, TM4 and the loop in between) may reflect the induced-structural rearrangement of the receptor upon ligand binding. This example highlights the ability of HDX-MS experiments to measure membrane protein conformational dynamics in their native state.



**Figure 3.** Structural characterization of  $A_{2A}$  upon ligand ZM241385 binding with HDX-MS. HDX-MS identified peptides shown on the  $A_{2A}$  sequence; transmembrane domains are highlighted in blue and the agonist ligand-binding region in green (left). The relative deuterium uptake of  $A_{2A}$  of each identified peptide at each deuteration times are plotted, with peptides in the transmembrane regions framed in blue (middle). Relative fractional uptake differences between  $A_{2A}$  and  $A_{2A}$  with the ligand ZM241385 are represented on the protein structure (PDB code: 4E1Y [56]) at a 30 min deuteration time; regions not covered are colored in grey (right).

In addition to continuous labeling studies (deuteration time points from a few seconds to hours), HDX-MS can also be used in a time-resolved manner to monitor sequential conformational changes in the protein of interest over time, using very short “pulsed” deuteration time [223] either in the apo state to report on folding and aggregation [228] or in an holo state to report on protein–protein interaction. To our knowledge, this pulsed HDX-MS approach has been reported only once on  $A_{2A}$  by Du et al. [229], in order to better understand why GPCR/G-protein complexes are stabilized in a GDP free-state, how this nucleotide is released and to lay the structural basis of coupling specificity between G-protein subtypes and GPCRs. Interestingly, the authors used hydroxyl radical protein footprinting with mass spectrometry [230,231], which aims at looking at protein conformational changes after the irreversible labeling of amino acid side chains by hydroxyl radicals (generated by radiolysis or photolysis reactions), as an orthogonal technique to detect very early events (from a few milliseconds to seconds).

Among the emerging structural MS methods, chemical cross-linking followed by MS analysis is still rarely used for in vitro membrane protein analysis. The first step consists of the covalent binding of a cross-linker agent to the side chain of amino acids, followed by the quenching of the reaction and the reduction, alkylation and digestion step. Then, peptides including cross-linked dipeptides are separated and analyzed using LC-MS/MS and finally identified via different software due to the mass increment due to the cross-linking reagent [232,233]. The  $A_{2A}$  receptor was used as a model to assess which chemical cross-linkers are suitable for the characterization of GPCRs by cross-linking mass spectrometry (XL-MS) [234]. Similar to HDX-MS, all steps of the XL-MS workflow have to be adapted to membrane protein analyses, especially the choice of the cross-linker reagent and the cross-linking conditions, enzymatic digestion and sample clean up, along with adapted LC-MS methods and data treatment. By comparing the MS-identified cross-links to the structurally compatible cross-links from available GPCR structures, Jones et al. concluded arginine-arginine cross-linker and lysine-arginine are best adapted for XL-MS analyses of GPCRs and are highly complementary to the existing classical cross-linkers

present in the toolbox of specialists [235]. Altogether, structural MS methods have great potential to complement more classical high-resolution structural biology techniques, and are becoming more and more popular.

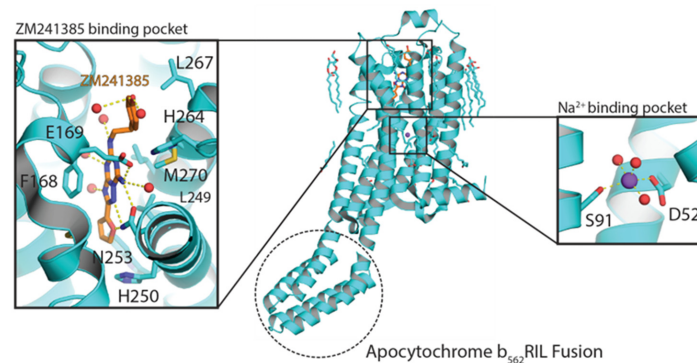
### *8.2. X-ray Crystallography of GPCRs: Uncovering Conserved Activation Mechanisms Using A<sub>2A</sub> as a Model*

GPCRs are highly dynamic and become particularly unstable during the purification process once extracted from the cell membrane. As a consequence, crystallization of GPCRs remains a challenging task. To overcome such difficulties, protein engineering strategies have been developed to reduce the flexibility and to increase the stability of these membrane receptors. A<sub>2A</sub> was one of the first receptors for which the high-resolution structure was solved using X-ray crystallography. The T4 Lysozyme (T4L) fusion partner, initially established for resolving the high-resolution crystal structure of the beta-2 adrenergic receptor ( $\beta_{2A}$ R) [236], was later applied to solve the first structure of A<sub>2A</sub> [51]. The T4L increases the hydrophilic surface of detergent-solubilized GPCRs to promote crystal contacts, and reduces the flexibility associated with the TM6 helix [236]. Subsequent developments led to the application of the BRIL fusion partner into intracellular loop 3 to solve the A<sub>2A</sub> crystal structure at 1.8 Å [56]. Another key engineering strategy to overcome the instability of GPCRs in detergents is by introducing point mutations in the 7TM domain to select thermostabilized ligand-bound conformations. The development of conformational thermostabilization of the A<sub>2A</sub> [31,237] has been successfully applied for a range of co-crystallized agonist- and antagonist-bound structures [52,72].

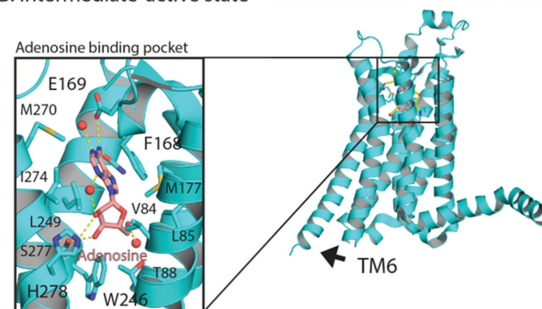
Ciancetta pointed out in 2019 that [238] “A<sub>2A</sub> represents to date (2019) one of the rare Class A GPCR that has been solved in the inactive (R) [79], active-intermediate (R\*) [68], and fully active state coupled to an engineered Gs protein  $\alpha$  subunit (R \* G) [53,64]”. Most of these structures have been obtained using X-ray crystallography. During the last 3 years, more structures were solved in similar configurations for other Class A GPCRs.

The structural determination of multiple A<sub>2A</sub> conformational states has contributed greatly to the understanding of Class A GPCR activation mechanisms [38,51–53,68,72,239]. The antagonist-binding pocket has been thoroughly characterized by the co-crystallization of A<sub>2A</sub> bound to ZM241385 (Figure 4A) and to various xanthine-containing compounds including caffeine and XAC, using either thermostabilization or fusion protein strategies [72]. Later developments combining the antagonist-thermostabilized point mutations with the BRIL fusion partner provided the highly utilized construct known as A<sub>2A</sub>-StaR2-bRIL [69,79,240]. Multiple structures of antagonist-bound states also revealed the sodium allosteric modulation of A<sub>2A</sub> [56,58]. The allosteric sodium ion-binding site is nested in a set of highly conserved residues, located below the orthosteric binding pocket, and bound to the highly conserved Asp<sup>2.50</sup> [56,58].

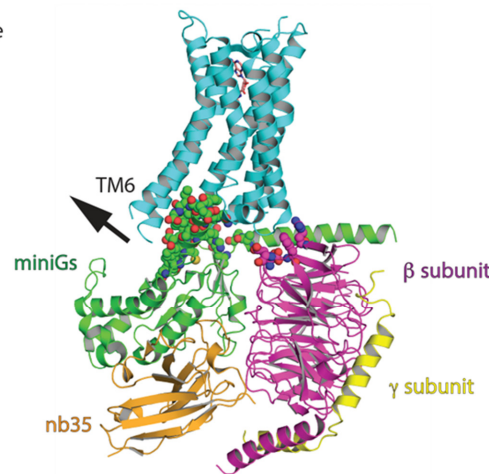


A. A<sub>2A</sub> inactive state

## B. Intermediate-active state



## C. Fully active state



**Figure 4.** Three-dimensional structures of the human adenosine A<sub>2A</sub> receptor. The A<sub>2A</sub> receptor is bound to the antagonist ZM241385 (A; PDB: 4EIY [56]), agonist adenosine (B; PDB: 2YDO [52]), NECA and mini-Gs protein (C; PDB: 6GDG [241]). These figures represent the inactive, intermediate-active, and fully active states, respectively. Polar contacts are represented as yellow dashed lines, with the sodium ion depicted as a purple sphere and water molecules as red spheres.

Agonist-binding modes of UK-432097 [68], NECA and adenosine (Figure 4B) to the thermostabilized A<sub>2A</sub> highlight the molecular interaction of the ribose moiety of all agonists with Ser277 and His278 and contraction of the receptor binding site [52]. Agonist binding also initiates a slight outward movement of TM6 that defines an intermediate-active conformation [52,57,58,68,242]. The crystal structure of the A<sub>2A</sub> bound to an engineered Gs protein (known as mini-Gs) illustrated the large outward movement of TM6 that is required for the mini-Gs to bind, in addition to slight movements of TM5 and TM7 and rotamer shifts of several amino acids near the G-protein binding site (Figure 4C) [53].

This was further validated by solving the cryo-EM structure of A<sub>2A</sub> bound to the heterotrimeric mini-Gs (Figure 4C) [241]. However, comparisons between the NECA

binding pocket in the intermediate [52] and fully activated conformation [53] show identical binding modes. Together, these studies provided an overview of the conserved activation mechanisms of Class A GPCRs [243].

High-resolution crystal structures of A<sub>2A</sub> have also led to the advancement of structural-based drug design (SBDD) [70,135]. Recent reports have outlined chemical modifications of an adenosine scaffold, commonly attributed to A<sub>2A</sub> agonist activity, which led to the alteration of the compound from an agonist to an antagonist [244]. Although the ribose moiety is accepted to be important for A<sub>2A</sub> agonist activity, recent structural and functional data have demonstrated the binding mode of a non-riboside partial agonist [242]. Although allosteric regulation of the human A<sub>2A</sub> remains relatively unexplored, differences between antagonist-bound conformations have suggested the possibility of an allosteric binding site within the 7TM binding pocket [245]. Taken together, these studies highlight future avenues for the design of selective orthosteric A<sub>2A</sub> modulators and the possibility of development of allosteric ligands.

Technical advances have been made in macromolecular crystallography for speeding up the workflow for screening, data collection and processing, proving to be essential for the progress of GPCR structural biology. The infrastructure developments of synchrotron microfocus beamlines have also been key for solving A<sub>2A</sub> structures and GPCRs in general. Moreover, the use X-ray free-electron lasers (XFELs) and lipid cubic phase (LCP) injectors for sample delivery has allowed the structural determination of A<sub>2A</sub> at room temperature, opening new possibilities for investigating the highly dynamic nature of GPCRs [78,81,246–251]. Currently, X-ray crystallography has become one of the more accessible approaches for the structural determination of GPCRs. Understanding the dynamic activation mechanism of A<sub>2A</sub> remains of strong interest. Accordingly, the use of integrated approaches, such as cryo-EM, will uncover more details of A<sub>2A</sub> in complex with its signaling partners in the future.

### 8.3. Cryo-Electron Microscopy

Cryo-EM is a powerful technique for solving structures of challenging and dynamic proteins that are difficult to obtain using X-ray crystallography. While there is a debate on the respective virtues of cryo-EM versus X-ray crystallography for the determination of protein structures, particularly those of membrane-bound and integral membrane proteins [252–254], it is clear to us that the complementarity between those approaches opens avenues for a better description of membrane-bound proteins in the future, as reviewed by others [255,256].

A key advantage of cryo-EM is that it does not require the preparation of protein crystals, rather it allows for the imaging of single isolated particles in isomorphous ice prepared directly from purified samples. Conversely, the application of the cryo-EM technology for proteins or complexes smaller than 60 kDa remains challenging due to the inherent low signal-to-noise ratio (or low contrast) that limits the accurate alignment of projections during image analyses. Accordingly, the relatively small size of a ligand-bound Class A GPCR such as A<sub>2A</sub> (approximately 35–45 kDa) is typically not adapted to cryo-EM studies but is suitable for X-ray crystallography which remains the preferred technique for the determination of GPCR structure in the inactive state [257].

This size limitation has been recently circumvented by increasing the molecular weight of GPCRs isolated in protein complexes. Most remarkably, advances of the near-atomic resolution of activated GPCRs in complex with G-protein binding partners have been achievable with the use of cryo-EM [258]. Cryo-EM has since been successfully employed to solve the structure of an engineered A<sub>2A</sub> thioredoxin fusion protein (TrxA-A<sub>2A</sub>) in complex with the mini-Gs protein [241]. A recent study made use of an anti-BRIL Fab in complex with the A<sub>2A</sub>-BRIL to increase the molecular weight of A<sub>2A</sub> and to facilitate cryo-EM data processing [61].

One method recently developed, known as microcrystal electron diffraction (MicroED), combines crystallization in LCP with the acquisition of cryo-EM for structural determina-

tion. Here, a beam of electrons is transmitted through protein microcrystals grown in LCP, rather than X-rays by conventional crystallography. A<sub>2A</sub> is currently the only GPCR for which this method has been successfully employed [259]. Future work ought to apply the use of cryo-EM to investigate the structures of the purified A<sub>2A</sub> in complex with signaling partners such as the full-length Gs heterotrimer and other intracellular signaling partners.

#### 8.4. NMR Approaches

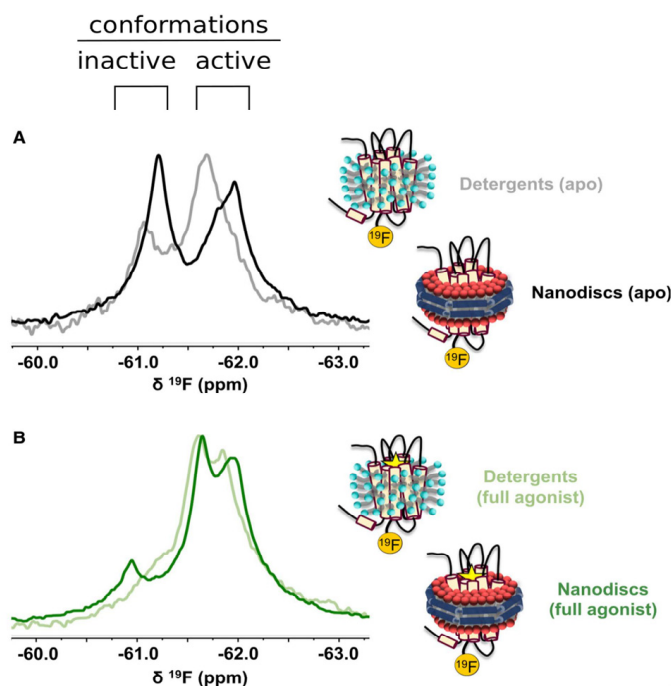
At a molecular level, GPCR signaling can be described through the concept of free energy landscape [260] where orthosteric ligands, intracellular effectors and allosteric modulators influence both the population and lifetime of various inactive, intermediate and active states. High-resolution NMR spectroscopy is a powerful tool to investigate these complex energy landscapes. While NMR can be used to obtain de novo structures of proteins, in the case of large complexes such as GPCRs associated with various surfactants or lipids, most of the NMR studies that are conducted in solution concern conformational landscapes or interactions, with considerations sometimes relating to chemical exchanges. They rely advantageously on the plethora of atomic scale structures obtained from X-ray crystallography and cryo-EM to highlight the coexistence of several inactive and active sub-states, including lowly populated states, but also, to a much lesser extent, the detection of some chemical exchanges. Along with the  $\beta_{2A}R$ , A<sub>2A</sub> is one of the GPCRs most studied with NMR. These studies are based on different isotopic labeling strategies and/or expression systems.

An appreciable amount of NMR studies of A<sub>2A</sub> is based on fluorine (<sup>19</sup>F) NMR [48,50,73,93]. This nucleus can be easily incorporated in proteins via biosynthetic pathways or chemical reactions and has the advantage of displaying a large chemical shift dispersion compared to other nuclei that are traditionally detected to study biomolecules [261,262]. This partly counterbalances the fast <sup>19</sup>F transversal relaxation due to the large chemical shift anisotropy, which becomes the major source of linewidth in high magnetic fields. As GPCRs are usually expressed in eukaryotic expression systems, <sup>19</sup>F labeling via chemical reactions is generally preferred. However, in this case, only solvent-exposed residues can be labeled due to the accessibility for the chemical reaction to occur and, consequently, all these studies are based on single extra-membrane <sup>19</sup>F probes. Following such a strategy, the conformational landscape of the receptor was observed using one-dimensional (1D) <sup>19</sup>F NMR at the three following positions that are all located at the extracellular tip of TM helices: V229C<sup>6.31</sup> (TM helix VI, TM6) [48,93], L225C<sup>6.27</sup> (TM6) and A289C<sup>7.54</sup> (TM7) [73]. 1D <sup>19</sup>F spectra in these locations display a pattern characteristic of a complex conformational ensemble, composed of several inactive and active sub-states in equilibrium. Their population strongly depends on the presence or absence of ligands and their associated pharmacological properties, as well as the presence or absence of effectors. This illustrates that signal transduction by A<sub>2A</sub> relies on a dynamic equilibrium of several conformations that can be detected through a fluorine probe. Interestingly, several coexisting active states, whose populations are related to specific ligands were observed, fitting with the concept of partial agonism, i.e., when a GPCR is activated to a sub-maximal level.

By investigating these three positions of the <sup>19</sup>F probe, different distributions of populations in the conformational ensemble could be identified in the apo or holo (e.g., in the presence of the full agonist NECA) states. Between V229C<sup>6.31</sup> [48,93], or L225C<sup>6.27</sup> [73], and A289C<sup>7.54</sup> [73], this may reflect a different response to drug efficacy at the intracellular ends of TM6 and TM7 [73]. In contrast, it is most puzzling that the distribution of populations is not the same between V229C<sup>6.31</sup> and L225C<sup>6.27</sup>, as these two residues are localized very closely with each other at the extracellular tip of TM6 [48,73]. Moreover, these two studies were performed in the same detergent solution. This discrepancy could be associated with different possible effects of aromatic ring current fields in the vicinity of these two <sup>19</sup>F NMR probes that could partially respond differently to conformational changes [263].

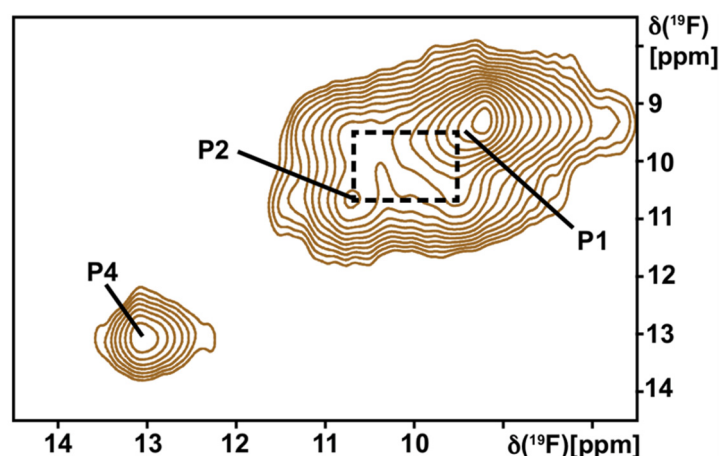
Interestingly, looking at the conformational ensemble of V229C<sup>6.31</sup> in the apo state or in the presence of NECA of A<sub>2A</sub> embedded in a lipid nanodiscs, the 1D <sup>19</sup>F NMR spectra

are very different from what has been previously observed in detergent [50] (Figure 5), highlighting the importance of studying the energy landscape of membrane proteins in a native-like lipid bilayer environment, even if using very small nanodiscs where the receptor is surrounded by just one layer of lipid (lipoprotein  $\Delta H5$  [264]), such as those used in Huang et al. [50]. Furthermore, this study is also remarkable as they could additionally depict the conformational ensemble of V229C<sup>6.31</sup> in the presence of a heterotrimeric G protein, revealing a G-protein-bound intermediate and distinct nucleotide-free state in the presence of either a partial or a full agonist. This study further suggests a scaffolding role for the G $\beta\gamma$ -associated subunits, facilitating the allosteric transmission through the transmembrane part of the receptor from the orthosteric ligand site to the intra-cellular side in a ligand-dependent manner [50].



**Figure 5.**  $A_{2A}$  exhibits a different distribution of states and slower exchange dynamics in a lipid bilayer environment than in detergent micelles (A,B). Comparison of the  $^{19}\text{F}$  NMR spectra of apo (A) or agonist-bound (B)  $A_{2A}$  reconstituted in either LMNG micelles or phospholipid nanodiscs (legend and figure adapted from Huang et al. [50] with automatic permission from Elsevier).

A common feature identified in all of these studies is that most of the sub-states sampled by the  $^{19}\text{F}$  probe suggest a slow chemical exchange at the  $^{19}\text{F}$  chemical shift timescale (millisecond timescale), under the assumption that some chemical exchanges are present between the conformations revealed by the fluorine probe. A strong indication that such chemical exchanges are indeed present is given by the observed variations in either chemical shifts and/or signal linewidths in association with measurements of the transversal relaxation time ( $T_2$ ) upon the addition of various ligands and/or a heterotrimeric G protein or a G $\alpha$  mimetic peptide. 1D  $^{19}\text{F}$  NMR can also be expanded with a second  $^{19}\text{F}$  frequency axis to directly visualize conformational exchanges by 2D [ $^{19}\text{F}$ - $^{19}\text{F}$ ] exchange spectroscopy (EXSY) spectra [73] (Figure 6).



**Figure 6.** Conformational exchange in the  $A_{2A}$  complex with the full agonist NECA observed using 2D exchange spectroscopy. A contour plot is shown of a 2D  $[^{19}\text{F}, ^{19}\text{F}]$ -EXSY spectrum collected at 280 °K with a mixing time of 100 ms. The diagonal peak positions of the active-like sub-states P1, P2, and P4 are labeled, and a dashed box indicates cross-peaks observed between conformations P1 and P2. This spectrum indicates the existence of a low-energy barrier for interconversion between sub-states P1 and P2 (legend and figure adapted from Susac et al. [73] with automatic permission from PNAS).

Among the various allosteric modulators that have been identified in the activation of GPCRs, lipids play an important role by either binding to specific sites, e.g., cholesterol or phosphatidylinositol molecules acting as specific cofactors [180], or by modulating the physical properties of the lipid bilayer [265], thereby impacting the activation of these receptors. Docosahexaenoic acid (DHA) phospholipid chains are well represented in the membrane of cells where  $A_{2A}$  is highly expressed. With the help of the introduction of a  $^{13}\text{C}$ -labeled and protonated methyl group in methionine residues, Mizumura et al. investigated the effect of these acyl chains on the activation of  $A_{2A}$  [49]. Thanks to deuteration, this study could be performed on quite large nanodiscs stabilized by the lipoprotein MSP1E3 [103]. Based on 2D  $[^1\text{H}, ^{13}\text{C}]$  NMR spectra, they deduced that in the presence of a partial agonist,  $A_{2A}$  is in an equilibrium between inactive and active states, while a full agonist shifts the equilibrium towards the active state only. Interestingly, the addition of DHA enhances the G-protein activation by  $A_{2A}$ , a positive allosteric effect that has also been observed, for instance, with phosphatidylinositol (4,5)-biphosphate and an analog of cholesterol in the activation of ghrelin [180] and leukotriene receptors [266], respectively. Based on these observations, they make the assumption that DHA acyl chains modify the conformational equilibrium centered at the NPxxY motif that would induce a large clockwise rotation of TM6, which is based on the lipid-dependent chemical shift variations of methionine residues located at the cytoplasmic ends of TM3 and TM6 (M106 and M232, respectively).

Another important class of allosteric modulators of GPCRs are cations. Deciphering  $A_{2A}$  conformation landscape in various situations, Ye et al. confirmed the negative allosteric modulation of  $A_{2A}$  activity by sodium ions, as already observed in pharmacological and structural studies and confirmed as being common to many Class A GPCRs [56]. Interestingly, they observed an opposite trend upon the addition of divalent cations, e.g.,  $\text{Ca}^{2+}$  and  $\text{Mg}^{2+}$ , which had positive allosteric effects that were boosted by the presence of an agonist and a G-protein-derived peptide [93]. With the aid of molecular dynamic (MD) simulations, they propose that divalent cations would establish bridges between extra-cellular acidic residues belonging to TM5 and TM6, which would allosterically contribute to the opening of the G-protein binding cavity on the intracellular side of the receptor.

$^{19}\text{F}$  NMR can be useful to study GPCR conformational ensembles thanks to a large chemical shift dispersion, a high sensitivity (a gyromagnetic ratio close to the proton) and the fact that there are no background signals. However, if using just a single incorporated



probe, in particular, localized in an extra-membrane region, it is very difficult to obtain a global and coherent view of the molecular mechanism governing activation. To overcome these limitations, Eddy et al. used uniform-labeling of the receptor with  $^{15}\text{N}$  and  $^2\text{H}$  isotopes to investigate the allosteric coupling between the drug binding event and intracellular signaling [47]. The authors took advantage of the  $^1\text{H}$ ,  $^{15}\text{N}$  correlation peak chemical shifts of exchangeable Gly amide ( $^1\text{HN}$ ) and Trp indole protons, as their  $^1\text{H}/^{15}\text{N}$  cross-peaks resonate in well-isolated parts in typical 2D [ $^1\text{H}$ ,  $^{15}\text{N}$ ] correlation spectra. Thanks to an additional partial deuteration (~70%), this strategy enabled them to identify well resolved NMR cross-peaks that could be assigned to six individual Trp indole protons and eight Gly amide protons by single mutagenesis. As discussed in this study, multiplying the number of sampling points in the receptor is only interesting if most of the assigned residues are located in the vicinity of “ring current fields” to increase the sensitivity of these nuclei to subtle changes in the conformational polymorphism of the receptor. Hence, significant  $^1\text{H}$  and/or  $^{15}\text{N}$  chemical shift variations for 11 of the 14 assigned residues were observed between  $\text{A}_{2\text{A}}$  bound to an agonist and to an antagonist. Incidentally, no data were reported on the apo state in this work. The synthesis of all their observations leads to a model where there is a strong interplay between the very-well-conserved residue of A52<sup>2,50</sup> and W256<sup>6,48</sup>; the latter, which is located just below the ligand orthosteric pocket, has previously been hypothesized to change its rotameric state upon activation (“toggle switch”) [267]. The authors also extended the use of indole protons of tryptophan to introduce extrinsic tryptophan residues in  $\text{A}_{2\text{A}}$  to sample the energy landscape of the receptor at additional locations, confirming that the receptor responds differently to ligands with variable efficacy and that the intracellular end of TM6 samples wide conformational variations upon activation [268]. More recently, they applied this same strategy to compare the conformational ensembles of  $\text{A}_{2\text{A}}$  associated with two different partial agonists (LUF5834 and Regadenoson) and the full agonist NECA [269]. Their study suggests a different signal propagation pathway based on variations in populations of simultaneously coexisting conformations at equilibrium [269].

Solution-state NMR undoubtedly represents a key ally to classical structural approaches using crystallography or electron microscopy to further explore the energy landscape of very dynamic proteins, such as GPCRs. Numerous studies performed at or nearly at physiological temperatures with no or just a few chemical modifications depict a complex conformational landscape often associated with high kinetic barriers.  $\text{A}_{2\text{A}}$  is no exception, and beyond representing an important target for several human diseases, it also portrays an interesting model to study the fundamental properties of GPCRs, such as biased or partial agonism and the impact of allosteric modulators in the activation process. From the studies performed so far, it appears to be better to sample several locations at the same time within the receptor, whenever possible. In order to connect different parts of the receptor in the molecular activation mechanism(s), it will also be important to sample novel positions beyond the water-exposed regions. This should also limit the discrepancies between different approaches relying on nuclei with different physical properties and/or sample conditions. Moreover, as a membrane protein in solution is highly sensitive to its swimming belt, it seems to be essential to try to study the receptor in a membrane-like environment. Furthermore, in consideration of the importance of the constitutive activity in GPCR signaling, it will be crucial to include the *apo* state in all subsequent studies, as this has not been conducted systematically in the studies mentioned here. One major objective for NMR remains the determination of the kinetic barriers separating the different sub-states observed, which, in addition to the determination of populations, will finally provide fundamental kinetic and thermodynamic data to better understand the signaling process through these integral membrane receptors such as  $\text{A}_{2\text{A}}$ . One possible path to help to increase the amount of information gained from an NMR experiment would be to improve the deuteration level of the receptor in conjunction with the use of appropriate isotope-labeling schemes, high magnetic fields and other suitable methodologies [270]. This would allow not only base NMR-derived activation models on chemical shift variations or

intensities of NMR signals, but would also be of particular advantage to investigate the dynamics in GPCRs using the relaxation phenomena of nuclear spins [271].

### 8.5. Future Trends for GPCR Structure Studies

#### 8.5.1. Cryo-Electron Tomography

Cryo-electron tomography (cryo-ET) data can be collected with the same transmission electron microscopes as single-particle analysis data. Cryo-ET is label-free and provides three-dimensional snapshots of organelles and proteins at nanometer resolutions within their functional cellular environments, allowing users to visualize and understand how they, and other molecules, work together to carry out major processes in a cell.

The majority of Class A GPCRs are relatively small and thus very difficult to identify *in vivo* in a cellular environment. However, one can cite the very interesting example on visual rhodopsin, a member of the GPCR family [272]. In this work, the authors identified the highly ordered arrangement of rhodopsin molecules in the disk membrane of intact photoreceptors with molecular resolution. Due to the small size of rhodopsin (40 kDa), the resulting molecular map was not amenable to fitting with the X-ray structure of rhodopsin. However, the resolution was sufficient to identify that rhodopsin was forming dimers as building blocks of a hierarchical supramolecular architecture.

This structure is characterized by aligned rows of dimers, and pairs of rows. The authors proposed that GPCR dimerization and oligomerization offer possibilities for the allosteric regulation of GPCR activity. These results support previously published work, which show for the first time that rhodopsin molecules assemble in dimers and form paracrystals in native rod outer segment membranes using atomic force microscopy (AFM) images [273].

#### 8.5.2. Structural Flexibility of GPCR Investigated Using Single Molecule Fluorescence and Atomic Force Microscopy

Evidence has emerged that, rather than being activated by ligands in an on/off manner switching from an inactive to an active state, GPCRs exhibit high structural flexibility in the absence and even in the presence of ligands. The physiological as well as pharmacological impact of this structural flexibility remains puzzling. Single molecule fluorescence offers the advantage of monitoring the structural dynamics of biomolecules in real-time, with minimal structural invasiveness and in the context of complex biological environments; in particular, single-molecule Förster resonance energy transfer is a key method to measure actual distance changes between two probes, and highlight conformational changes occurring at timescales relevant for protein conformational movements [159].

AFM-based high-resolution imaging and force spectroscopy have been also successfully used to better understand the molecular mechanism details of how GPCR conformations are modulated by ligand-binding or GPCR-transducer complex formation [274].

## 9. Pharmacology of the Isolated Protein A<sub>2A</sub>

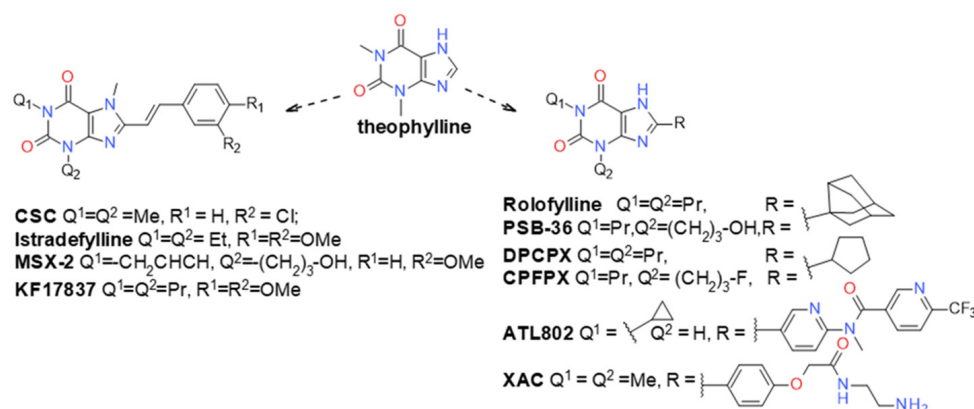
### 9.1. Background

While finding inhibitors or ligands for soluble proteins is relatively easy, and basically concerns the number of molecules one can screen, for membrane-associated proteins, such as GPCRs, it is far more complicated. A key to understand GPCR function still resides in the availability of molecules with chemically diverse structures—in other words, structurally diverse ligands (agonists as well as antagonists) are needed. Although this is more important for some GPCRs (melatonin, dopamine) than for others (histamine, serotonin), it is crucial that efforts are made in screening large libraries of molecules with the hope of finding new ligands. The availability of new approaches, particularly *in silico*, will assist in the discovery of new molecules, although many obstacles remain on these avenues.

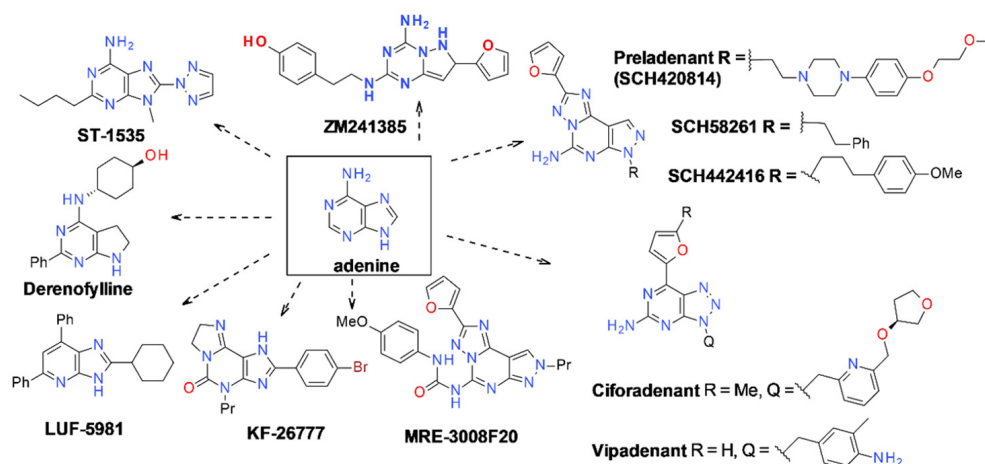


number of adenine derivatives without the ribose moiety have been reported as adenosine receptor antagonists, starting with the A<sub>1</sub>-selective N-0840 first reported in 1987 [280]. It was followed by many alternative but closely related heterocycles, including fused five- and six-membered rings. One of the tricyclic series has progressed to Preladenant [281], which reached the clinical phase (Figure 9). It is worth noting that the most powerful antagonist is SCH442416 [282,283], although the actual antagonistic property seemed not to be experimentally determined, but was based on the fact that this compound displaced the potent antagonist, SCH58261 [284]. This compound and its reported analogues were also structurally related to adenine. The development of this molecule led to a positron emission tomography (PET) imaging compound applied to visualize the A<sub>2A</sub> expression in vivo [283,285].

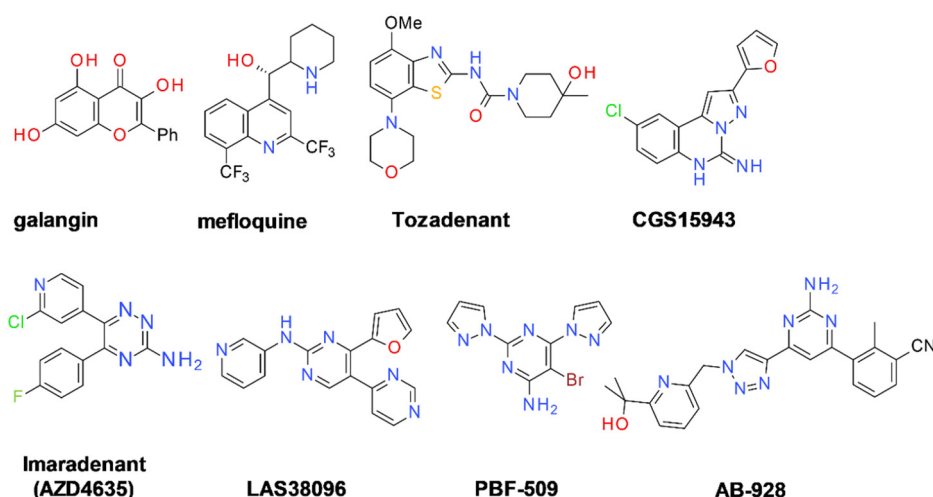
Molecular pharmacology shows that the A<sub>2A</sub> agonist and antagonist families are well-populated. They form a large ensemble of molecules with a large panel of properties, although their chemical diversity is somewhat restricted, as is often the case for small molecule GPCRs, such as melatonin [286] or serotonin [287] receptors.



**Figure 8.** Structures of the main A<sub>2A</sub> receptors agonists derived from theophylline.



**Figure 9.** Structures of the main A<sub>2A</sub> receptors agonists derived from adenine.



**Figure 10.** Structures of the main A<sub>2A</sub> receptors antagonists.

### 9.3. Allosteric Modulators

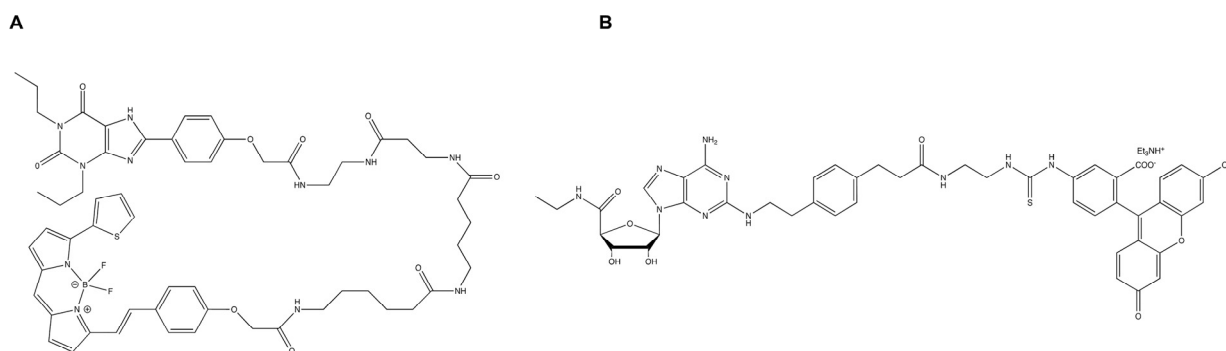
Among the new descriptions of GPCR ligands, besides the classical antagonists, agonists and inverse agonists, are also the allosteric modulators of GPCR signaling which have previously been reviewed in depth [288–290]. In brief, such compounds bind to the receptor at a site different to the orthosteric site, leading to subtle modifications of the area, and hence to changes in either the affinity for the agonist or of the activity of orthosteric ligands. Two major classes of allosteric ligands have been described, i.e., negative allosteric modulators (NAMs) or positive allosteric modulators (PAMs). NAMs inhibit the activation of the receptor and the downstream signal transduction. On the other hand, PAMs increase the activation of the receptor. Furthermore, ions are also considered as allosteric modulators for multiple GPCRs, such as Na<sup>+</sup> for the adenosine A<sub>2A</sub> receptor [89,291] or Zn<sup>2+</sup> for the β<sub>2</sub>-adrenergic receptor [292] for which they have been shown to bind at allosteric sites which differ from the orthosteric site as reviewed by van der Westhuizen et al. [290]. While they offer new ways to modulate and fine-tune GPCR activity, they are quite difficult to screen for, as they might be dependent on the nature of the agonist they work in conjunction with. This poses a great challenge for the screening format, especially when functional assays are performed on living cells. Furthermore, to characterize this novel, more peculiar type of binding sites of NAMs and PAMs, a large panel of biophysical techniques is required, including molecular dynamics (MD) simulations, ligand structure–activity relationship studies, and in-solution NMR analyses, to name a few [293].

The main example of an A<sub>2A</sub> PAM is perhaps DFIABA (3,4-difluoro-2-((2-fluoro-4-iodophenyl)amino)benzoic acid), a compound that enhances adenosine signaling at the A<sub>2A</sub> receptor, and thereby induces slow wave sleep without affecting body temperature in mice [294]. This remarkable feature exemplifies the role of these types of molecules, promising precise modulations of signaling processes with reduced side effects. Other molecules with similar functions include, AEA061 [295], or inosine. Inosine is an endogenous molecule, that could open avenues in the concept of natural PAMs or NAMs, adding to the arsenal of molecules modulating GPCRs functions [296]. The interplay between several receptors, in pathological models illustrates the complexity of various pharmacological ligands, as soon as they interfere with several profoundly different GPCRs, e.g., M4 and A<sub>2A</sub> [297], mGlu<sub>4</sub> and A<sub>2A</sub> [298] or (TLR)4 and A<sub>2A</sub> [299]. Adenosine receptors have also been screened for NAMs, as adenosine receptor subtype-specific negative allosteric modulators would offer access to new potential treatments for analgesia, possibly with minimum negative off-target effects, an area where immense treatment progress remains to be done [300].



#### 9.4. Fluorescent Ligands

Adenosine receptors are of therapeutic interest and, as such, selective ligands attract particular attention. Indeed, a plethora of ligands with well-defined pharmacological profiles have been developed during the last years for  $A_{2A}$ . Due to the accumulation of the three-dimensional structures of  $A_{2A}$  in different activation states, the design of ligands is now largely structure-based. Different compounds with particular pharmacological profiles have been identified and reviewed exhaustively [5], as well as their modified counterparts such as radiolabeled, covalent or bivalent ligands [301]. We will focus here on a particular class of labeled compounds, fluorescent ligands. Indeed, such compounds are of major interest when developing ligand-binding assays with isolated receptors [302]. A full series of fluorescent  $A_{2A}$ -selective ligands have been devised to be used in native membranes or cell-based systems [303]. These include agonists such as the initial compound FITC-APEC that was derived from the conjugation of the  $A_{2A}$  agonist APEC to fluorescein [304]. An alternative version of FITC-APEC, with Alexa Fluor 488 as the fluorophore instead of fluorescein, has been more recently used to study the kinetics of  $A_{2A}$  redistribution in living cells [305]. Besides agonists, fluorescent antagonists have also been designed, for instance by coupling the selective  $A_{2A}$  antagonist preladenant to Alexa Fluor 647 [306], or using either of the two pharmacophores, 1,3-dipropyl-8-phenylxanthine and triazolo [1,5-c]quinazolin-5-yl)amine [307]. Regarding isolated receptors, the number of studies with fluorescent ligands is much more limited, and these compounds have been mostly used in the context of ligand-binding experiments. A fluorescent antagonist, CA600245, was recently used to assess the ligand-binding ability of  $A_{2A}$  encapsulated into SMALPs generated by solubilizing membranes of *P. pastoris* expressing  $A_{2A}$  with the SMA polymer. In this case, the fluorescent ligand comprised the antagonist xanthine amine moiety linked to a red BODIPY630/650 fluorophore (Figure 11A). The resulting study demonstrated that  $A_{2A}$  in SMALPS is a powerful model system to characterize ligand-receptor complexes when used in combination with fluorescence spectroscopy [111]. More recently, the fluorescent compound FITC-APEC described above (Figure 11B) was used to provide an experimental and theoretical framework for monitoring ligand-binding kinetics to the  $A_{2A}$  in detergent micelles under ligand depletion conditions using fluorescence anisotropy measurements [308]. Taken together, the availability of different fluorescent probes from distinct pharmacological classes now provides a toolbox of labeled ligands to develop radioactive-free ligand-binding assays with purified receptors, as well as to probe the pharmacological behavior of  $A_{2A}$  isolated in detergent micelles and nanodiscs.



**Figure 11.** Chemical structures of the fluorescent ligands used for in vitro assays with purified  $A_{2A}$ . (A) CA600245 was used to assess the functional properties of the isolated receptor in SMALPS [111]. (B) FITC-APEC was used to monitor the ligand-binding kinetics to the  $A_{2A}$  in detergent micelles [308].

#### 9.5. Structure-Based Ligand Discovery for $A_{2A}$

Structure-based methods in drug discovery comprise a variety of different computer-assisted strategies, including either ligand- or structure-based virtual screening approaches, as well as pharmacophore-based, fragment-based and comparative molecular field anal-

ysis [309–312]. These methods are widely used to identify novel ligands exerting either an agonistic or an antagonistic effect on a specific protein target and they also have had considerable impact on the drug-discovery-related pharmacology of A<sub>2A</sub>. There are several examples in the literature that illustrate the practical application of structural information for the discovery of A<sub>2A</sub> antagonists which have been reviewed by Jazayeri et al. [313]. Furthermore, molecular docking has also been employed for the design of either A<sub>2A</sub> agonists or PDE10A inhibitors, and has been a recent focus in the search of novel antiproliferative agents [314]. In addition, MD simulations have been used extensively to investigate the conformational dynamics at the A<sub>2A</sub> or PDE10A [315–321]. In a recent paper, Kalash et al. studied a novel structure-based method for identifying ligands that activate A<sub>2A</sub>, while simultaneously inhibiting the PDE10A [322]. The focus of this approach was on the key interacting residues that are reported in the literature to discriminate between agonist and antagonist activity of A<sub>2A</sub> ligands [52,57,64,68]. It has been postulated that the motion of the residue Val84 in the TM3, upon A<sub>2A</sub> ligand binding, might discriminate between agonist and antagonist activity, which had not been studied previously by any of the MD simulations of A<sub>2A</sub> [64,323]. Kalash et al. show that the motion of this residue can be used as a conformational descriptor for the characterization of receptor activation by A<sub>2A</sub> ligands [322]. Furthermore, it is suggested that a selective interaction of ligands with His<sup>250</sup> accounts for A<sub>2A</sub> sub-type selectivity. This interaction has only been detected in co-crystal structures with selective A<sub>2A</sub> agonists, such as CGS21680 [57] and UK432097 [58]. Moreover, MD simulation analysis demonstrated that this residue undergoes conformational changes only when selective A<sub>2A</sub> agonists are bound and not when non-selective agonists bind to A<sub>2A</sub> [322]. Additionally, the analysis of the MD trajectories highlighted the motion of Val84 in TM3 as an essential requirement for A<sub>2A</sub> activation, which is a strong indicator for the potential application of structure-based methods in the discovery of novel A<sub>2A</sub> ligands.

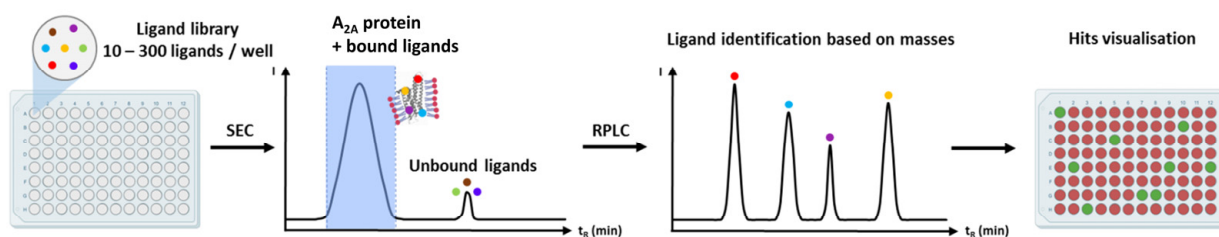
#### 9.6. Fragment-Based Ligand Discovery for A<sub>2A</sub>

Fragment-based ligand design is a well-established approach for the generation of novel protein inhibitors or modulators [324]. In a first step, fragment hits are identified from the screening of a fragment library against the protein target. In a second step, selected fragments are iteratively optimized using three-dimensional structures of the protein–ligand complex until a high-affinity compound is obtained. Fragment libraries typically regroup thousands of highly soluble small molecules with low molecular weight (<300 g/mol). Fragment screening using biophysical methods has been achieved for A<sub>2A</sub> using target immobilized NMR screening [325], SPR [134], STD-NMR [91] and Affinity Mass Spectrometry [293] with A<sub>2A</sub> samples prepared in detergents, or weak-affinity chromatography with A<sub>2A</sub> prepared in nanodiscs [77]. Whereas main studies are conducted with the wild-type receptor, a thermostabilized receptor was used in the study reported by Congreve et al. [134]. All techniques, except for STD-NMR, require the immobilization of A<sub>2A</sub> onto a solid support. To avoid the identification of false positive hits, the binding signal observed in the presence of A<sub>2A</sub> is compared to the one observed in the absence of A<sub>2A</sub> or in the presence of the receptor blocked with a reference ligand. In addition to experimental biophysical screening, fragment-based computer-aided design was reported by several groups [293,311,326–329]. To probe the conformational changes in the receptor upon fragment-based ligand binding, serial crystallography could play a key role in the future as a method for obtaining high-resolution conformational dynamics of protein–inhibitor complexes.

#### 9.7. Mass Spectrometry-Based Ligand Screening

Among the available modern screening methods described in this Section, affinity selection-mass spectrometry (AS-MS) is emerging as a technique of choice for the identification and confirmation of binders from large compound libraries, due to its high throughput capacity. AS-MS has been applied by the group of Shui to find new A<sub>2A</sub> receptor binders [106]. As a proof of concept, they first developed AS-MS using a small model

library consisting of a fifteen-compound mixture including eight known binders (such as adenosine, NECA, caffeine or ZM241385) and  $A_{2A}$  reconstituted in nanodiscs. All the known compounds were identified as positive binders from the AS-MS screen, validating the set-up for hit discovery on GPCRs. They next highlighted the potential of AS-MS to screen large compound libraries (approximately 20,000 compounds) on  $A_{2A}$  in one pool, leading to the discovery of three new antagonist compounds [141]. Interestingly, affinity MS experiments were successfully performed on a purified  $A_{2A}$  immobilized on nickel agarose beads as well as on the receptor embedded in cell-membranes. More recently [293], the same authors adapted their methodology to screen a library of 1100 fragments with the purpose of finding new NAMs of  $A_{2A}$ . Among the 28 initial hits, 17 have been confirmed to be specific binders at  $A_{2A}$ , and 9 as strong binders by SPR. These nine binders were then analyzed using affinity mass spectrometry in the presence of  $A_{2A}$ , free or bound to ZM241385. Thanks to an extensive optimization of immobilization and incubation conditions (see scheme in Figure 12), their method was initially validated as it allowed the detection of three known weak antagonists with binding affinity above 20  $\mu$ M (theophylline, NG-52 and LH-846). The six fragments that retained 50% binding in the presence of ZM241385 were further tested in competition with HMA (5-(N,N-hexamethylene)amiloride), one of the most potent NAMs of  $A_{2A}$ . Two new HMA-competitive binders were discovered using this approach and confirmed as NAMs using a competitive radioligand binding assay, molecular docking and  $^{19}\text{F}$ -labeled protein NMR experiments [293].



**Figure 12.** Schematic workflow of modern affinity selection mass spectrometry (AS-MS) used for  $A_{2A}$  hit identification. Additionally, called automated ligand identification system (ALIS), this technique is performed to identify small molecules that engage a specific target, such as  $A_{2A}$ . The protein of interest is incubated with a mixture of compounds, each with unique molar mass. The protein–ligand complexes are separated from unbound compounds using SEC. The protein peak (depicted in blue on the figure) is then directed to a reverse-phase liquid chromatography (RPLC) column for dissociation, desalting, and elution of any ligands into an ESI-MS system for identification. Alternatively to this 2D-LC set-up, an ultracentrifugation or an offline gel filtration step can be performed prior to LC-MS. This high-throughput screening technique enables hit identification for large collections of ligands.

## 10. Concluding Remarks and Perspectives

As the isolation of  $A_{2A}$  and its reconstitution in an active form were achieved relatively early compared to most other GPCRs, its handling became more streamlined compared to other GPCRs. Accordingly,  $A_{2A}$  has become a highly amenable model for the application and development of novel innovative approaches for the biophysical characterization of GPCRs. Moreover,  $A_{2A}$  is also a very important drug target in several human disease areas, and it goes without saying that the accumulation of knowledge of the structure and function of  $A_{2A}$  was also driven by the desire to discover novel and more specific pharmacological compounds. Indeed, the progress made in many of the cellular biology and cellular chemistry domains, together with the *in silico* approach of modelling the receptor structures and the three-dimensional structure of ligands, lead to the discovery of a series of novel ligands including compounds containing novel chemical scaffolds. Nonetheless, it remains to be seen if this knowledge will be transferable to the studies of other, more challenging to obtain GPCRs, especially from other families.

Many other areas could be further explored for A<sub>2A</sub> that might be more advanced in the context of other receptors. For example, the structural effects of the variety of agonists existing for this receptor should be further investigated by cryo-EM, possibly also in complex with different G proteins, to investigate the possibility of biased agonism at this receptor and how it can be explained at the structural level. Similarly, the structural effects on signal transduction of allosteric modulators are still largely unknown for the A<sub>2A</sub> receptor. To further understand these mechanisms, as well as the effects of subtle variations of the agonists on the structure of the G-protein binding site, dynamics kinetics need to further be studied. To this end, single molecule studies, molecular dynamics and also ensemble NMR studies will be invaluable.

While studies of the isolated receptor are crucial for understanding the mechanisms of signal transduction, it is of the utmost importance to also study receptors in their cellular context. Indeed, the progress recently made in the chemical alkylation of proteins in situ (for instance by click chemistry) might eventually allow high resolution studies of the receptor in a “native” milieu, surrounded by its “natural” neighbors in A<sub>2A</sub>-richly expressing membranes isolated from human tissues. However, the prerequisite of such experimental endeavors will be to introduce a handle at the receptor that can be recognized specifically by the chemical reaction. Combining the use of stem cells with the expression of such handles in a neutral genomic area of the transgene could provide a closer model of the physiological situation.

Finally, A<sub>2A</sub> has been proven to be an excellent model in many cases of biophysical, structural and even pharmacological characterizations of a GPCR. This is probably due to the relative easiness with which the receptor has been cloned, expressed and isolated, together with a natural stability that helped keep it active once purified.

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**Conflicts of Interest:** The authors declare no conflict of interest.

## Abbreviations

A<sub>2A</sub>, adenosine A<sub>2A</sub> receptor; AC, affinity chromatography; AFM, atomic force microscopy; AS-MS, affinity selection-mass spectrometry; BRIL, apo-cytochrome b562RIL; CABD, calcium binding domain; CaM, calmodulin; cryo-EM, electron cryo-microscopy; cryo-ET, cryo-electron tomography; DDM, n-dodecyl-β-D-maltopyranoside; DHA, docosahexaenoic acid; EXSY, exchange spectroscopy; FRET, fluorescence resonance energy transfer; GFP, green fluorescent protein; HDX, H/D exchange mass spectrometry; ITC, isothermal titration calorimetry; LCP, lipid cubic phase; LMNG, lauryl maltose neopentyl glycol; mBBR, monobromobimane; MS, mass spectrometry; MSP, membrane scaffold proteins; NAM, negative allosteric modulators; NDs, nanodiscs; NOESY, nuclear Overhauser effect spectroscopy; PAM, positive allosteric modulators; PGD, *petunia* glutamate decarboxylase; PI, Phosphatidylinositol; PQA, 3-pyrrolidin-1-ylquinoxalin-2-amine; PS, Phosphatidylserines; SEC, size-exclusion chromatography; Sf9, *Spodoptera frugiperda*; SMALPs, SMA lipid disc particles; SPA, Scintillation Proximity Assay; SPR, surface plasmon resonance; StaR, thermostabilized; STD, saturation transfer difference; T4L, T4 Lysozyme; TM, GPCR transmembrane domain; ULVs, unilamellar lipid vesicles; XAC, xanthine amine congener; XFEL, X-ray free-electron lasers; XL-MS, Cross-linking mass spectrometry.

## References

1. Fredholm, B.B.; Frenguelli, B.G.; Hills, R.; IJzerman, A.P.; Jacobson, K.A.; Klotz, K.-N.; Linden, J.; Müller, C.E.; Schwabe, U.; Stiles, G.L. Adenosine receptors in GtoPdb v.2021.2. *IUPHAR/BPS Guide Pharmacol. CITE* **2021**, 2021. [\[CrossRef\]](#)
2. Blackmore, R.V.; Linn, S. Partial purification and characterization of four endodeoxyribonuclease activities from *Escherichia coli* K-12. *Nucleic Acids Res.* **1974**, *1*, 1–17. [\[CrossRef\]](#)
3. Pierce, K.D.; Furlong, T.J.; Selbie, L.A.; Shine, J. Molecular cloning and expression of an adenosine A2b receptor from human brain. *Biochem. Biophys. Res. Commun.* **1992**, *187*, 86–93. [\[CrossRef\]](#) [\[PubMed\]](#)
4. Michalke, K.; Huyghe, C.; Lichère, J.; Gravière, M.-E.; Siponen, M.; Sciara, G.; Lepaul, I.; Wagner, R.; Magg, C.; Rudolph, R.; et al. Mammalian G protein-coupled receptor expression in *Escherichia coli*: II. Refolding and biophysical characterization of mouse cannabinoid receptor 1 and human parathyroid hormone receptor 1. *Anal. Biochem.* **2010**, *401*, 74–80. [\[CrossRef\]](#) [\[PubMed\]](#)
5. IJzerman, A.P.; Jacobson, K.A.; Müller, C.E.; Cronstein, B.N.; Cunha, R.A. International Union of Basic and Clinical Pharmacology. CXII: Adenosine Receptors: A Further Update. *Pharmacol. Rev.* **2022**, *74*, 340–372. [\[CrossRef\]](#)
6. Yu, L.; Frith, M.C.; Suzuki, Y.; Peterfreund, R.A.; Gearan, T.; Sugano, S.; Schwarzschild, M.A.; Weng, Z.; Fink, J.S.; Chen, J.-F. Characterization of genomic organization of the adenosine A<sub>2A</sub> receptor gene by molecular and bioinformatics analyses. *Brain Res.* **2004**, *1000*, 156–173. [\[CrossRef\]](#) [\[PubMed\]](#)
7. Koshiba, M.; Rosin, D.L.; Hayashi, N.; Linden, J.; Sitkovsky, M.V. Patterns of A<sub>2A</sub> extracellular adenosine receptor expression in different functional subsets of human peripheral T cells. Flow cytometry studies with anti-A<sub>2A</sub> receptor monoclonal antibodies. *Mol. Pharmacol.* **1999**, *55*, 614–624.
8. Cooper, J.A.; Hill, S.J.; Alexander, S.P.; Rubin, P.C.; Horn, E.H. Adenosine receptor-induced cyclic AMP generation and inhibition of 5-hydroxytryptamine release in human platelets. *Br. J. Clin. Pharmacol.* **1995**, *40*, 43–50. [\[CrossRef\]](#)
9. Schulte, G.; Fredholm, B.B. Human adenosine A<sub>1</sub>, A<sub>2A</sub>, A<sub>2B</sub>, and A<sub>3</sub> receptors expressed in Chinese hamster ovary cells all mediate the phosphorylation of extracellular-regulated kinase  $\frac{1}{2}$ . *Mol. Pharmacol.* **2000**, *58*, 477–482. [\[CrossRef\]](#)
10. Yang, S.N.; Dasgupta, S.; Lledo, P.M.; Vincent, J.D.; Fuxe, K. Reduction of dopamine D<sub>2</sub> receptor transduction by activation of adenosine A<sub>2a</sub> receptors in stably A<sub>2a</sub>/D<sub>2</sub> (long-form) receptor co-transfected mouse fibroblast cell lines: Studies on intracellular calcium levels. *Neuroscience* **1995**, *68*, 729–736. [\[CrossRef\]](#)
11. Ikeuchi, Y.; Nishizaki, T.; Mori, M.; Okada, Y. Adenosine activates the K<sup>+</sup> channel and enhances cytosolic Ca<sup>2+</sup> release via a P<sub>2Y</sub> purinoceptor in hippocampal neurons. *Eur. J. Pharmacol.* **1996**, *304*, 191–199. [\[CrossRef\]](#) [\[PubMed\]](#)
12. Wang, Q.; Zhou, F.M. cAMP-producing chemogenetic and adenosine A<sub>2a</sub> receptor activation inhibits the inwardly rectifying potassium current in striatal projection neurons. *Neuropharmacology* **2019**, *148*, 229–243. [\[CrossRef\]](#)
13. Varani, K.; Gessi, S.; Dalpiaz, A.; Borea, P.A. Pharmacological and biochemical characterization of purified A<sub>2a</sub> adenosine receptors in human platelet membranes by [<sup>3</sup>H]-CGS 21680 binding. *Br. J. Pharmacol.* **1996**, *117*, 1693–1701. [\[CrossRef\]](#) [\[PubMed\]](#)
14. Buckley, N.; Burnstock, G. Autoradiographic demonstration of peripheral adenosine binding sites using [<sup>3</sup>H]NECA. *Brain Res.* **1983**, *269*, 374–377. [\[CrossRef\]](#) [\[PubMed\]](#)
15. Schütz, W.; Brugger, G. Characterization of [<sup>3</sup>H]-adenosine binding to media membranes of hog carotid arteries. *Pharmacology* **1982**, *24*, 26–34. [\[CrossRef\]](#)
16. Ji, X.D.; Stiles, G.L.; van Galen, P.J.; Jacobson, K.A. Characterization of human striatal A<sub>2</sub>-adenosine receptors using radioligand binding and photoaffinity labeling. *J. Recept. Res.* **1992**, *12*, 149–169. [\[CrossRef\]](#)
17. Alexander, S.P.; Millns, P.J. [<sup>3</sup>H]ZM241385—An antagonist radioligand for adenosine A<sub>2A</sub> receptors in rat brain. *Eur. J. Pharmacol.* **2001**, *411*, 205–210. [\[CrossRef\]](#)
18. Yu, J.C.; Lin, G.; Field, J.J.; Linden, J. Induction of antiinflammatory purinergic signaling in activated human iNKT cells. *JCI Insight* **2018**, *3*, e91954. [\[CrossRef\]](#)
19. Klammt, C.; Schwarz, D.; Eifler, N.; Engel, A.; Piehler, J.; Haase, W.; Hahn, S.; Dötsch, V.; Bernhard, F. Cell-free production of G protein-coupled receptors for functional and structural studies. *J. Struct. Biol.* **2007**, *158*, 482–493. [\[CrossRef\]](#)
20. Köglér, L.M.; Stichel, J.; Beck-Sickinger, A.G. Structural investigations of cell-free expressed G protein-coupled receptors. *Biol. Chem.* **2019**, *401*, 97–116. [\[CrossRef\]](#)
21. Banères, J.-L.; Popot, J.-L.; Mouillac, B. New advances in production and functional folding of G-protein-coupled receptors. *Trends Biotechnol.* **2011**, *29*, 314–322. [\[CrossRef\]](#)
22. André, N.; Cherouati, N.; Prual, C.; Steffan, T.; Zeder-Lutz, G.; Magnin, T.; Pattus, F.; Michel, H.; Wagner, R.; Reinhart, C. Enhancing functional production of G protein-coupled receptors in *Pichia pastoris* to levels required for structural studies via a single expression screen. *Protein Sci.* **2006**, *15*, 1115–1126. [\[CrossRef\]](#)
23. Krettlér, C.; Reinhart, C.; Bevans, C.G. Expression of GPCRs in *Pichia pastoris* for structural studies. *Methods Enzymol.* **2013**, *520*, 1–29. [\[CrossRef\]](#)
24. Blocker, K.M.; Britton, Z.T.; Naranjo, A.N.; McNeely, P.M.; Young, C.L.; Robinson, A.S. Recombinant G protein-coupled receptor expression in *Saccharomyces cerevisiae* for protein characterization. *Methods Enzymol.* **2015**, *556*, 165–183. [\[CrossRef\]](#)
25. Errey, J.C.; Fiez-Vandal, C. Production of membrane proteins in industry: The example of GPCRs. *Protein Expr. Purif.* **2020**, *169*, 105569. [\[CrossRef\]](#) [\[PubMed\]](#)
26. Panneels, V.; Kock, I.; Krijnse-Locker, J.; Rezgaoui, M.; Sinning, I. Drosophila photoreceptor cells exploited for the production of eukaryotic membrane proteins: Receptors, transporters and channels. *PLoS ONE* **2011**, *6*, e18478. [\[CrossRef\]](#)



27. Zhang, L.; Salom, D.; He, J.; Okun, A.; Ballesteros, J.; Palczewski, K.; Li, N. Expression of functional G protein-coupled receptors in photoreceptors of transgenic *Xenopus laevis*. *Biochemistry* **2005**, *44*, 14509–14518. [[CrossRef](#)] [[PubMed](#)]
28. Kajikawa, M.; Sasaki, K.; Wakimoto, Y.; Toyooka, M.; Motohashi, T.; Shimojima, T.; Takeda, S.; Park, E.Y.; Maenaka, K. Efficient silkworm expression of human GPCR (nociceptin receptor) by a *Bombyx mori* bacmid DNA system. *Biochem. Biophys. Res. Commun.* **2009**, *385*, 375–379. [[CrossRef](#)]
29. Michalke, K.; Gravière, M.-E.; Huyghe, C.; Vincentelli, R.; Wagner, R.; Pattus, F.; Schroeder, K.; Oschmann, J.; Rudolph, R.; Cambillau, C.; et al. Mammalian G-protein-coupled receptor expression in *Escherichia coli*: I. High-throughput large-scale production as inclusion bodies. *Anal. Biochem.* **2009**, *386*, 147–155. [[CrossRef](#)] [[PubMed](#)]
30. Weiss, H.M.; Grisshammer, R. Purification and characterization of the human adenosine A<sub>2a</sub> receptor functionally expressed in *Escherichia coli*. *Eur. J. Biochem.* **2002**, *269*, 82–92. [[CrossRef](#)] [[PubMed](#)]
31. Magnani, F.; Shibata, Y.; Serrano-Vega, M.J.; Tate, C.G. Co-evolving stability and conformational homogeneity of the human adenosine A<sub>2a</sub> receptor. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 10744–10749. [[CrossRef](#)]
32. Lebon, G.; Bennett, K.; Jazayeri, A.; Tate, C.G. Thermostabilisation of an Agonist-Bound Conformation of the Human Adenosine A<sub>2A</sub> Receptor. *J. Mol. Biol.* **2011**, *409*, 298–310. [[CrossRef](#)]
33. Niebauer, R.T.; Robinson, A.S. Exceptional total and functional yields of the human adenosine (A<sub>2a</sub>) receptor expressed in the yeast *Saccharomyces cerevisiae*. *Protein Expr. Purif.* **2006**, *46*, 204–211. [[CrossRef](#)]
34. O'Malley, M.A.; Lazarova, T.; Britton, Z.T.; Robinson, A.S. High-level expression in *Saccharomyces cerevisiae* enables isolation and spectroscopic characterization of functional human adenosine A<sub>2a</sub> receptor. *J. Struct. Biol.* **2007**, *159*, 166–178. [[CrossRef](#)] [[PubMed](#)]
35. Schonenbach, N.S.; Rieth, M.D.; Han, S.; O'Malley, M.A. Adenosine A<sub>2a</sub> receptors form distinct oligomers in protein detergent complexes. *FEBS Lett.* **2016**, *590*, 3295–3306. [[CrossRef](#)] [[PubMed](#)]
36. Fraser, N.J. Expression and functional purification of a glycosylation deficient version of the human adenosine 2a receptor for structural studies. *Protein Expr. Purif.* **2006**, *49*, 129–137. [[CrossRef](#)] [[PubMed](#)]
37. Singh, S.; Gras, A.; Fiez-Vandal, C.; Ruprecht, J.; Rana, R.; Martinez, M.; Strange, P.G.; Wagner, R.; Byrne, B. Large-scale functional expression of WT and truncated human adenosine A<sub>2A</sub> receptor in *Pichia pastoris* bioreactor cultures. *Microb. Cell Fact.* **2008**, *7*, 28. [[CrossRef](#)]
38. Hino, T.; Arakawa, T.; Iwanari, H.; Yurugi-Kobayashi, T.; Ikeda-Suno, C.; Nakada-Nakura, Y.; Kusano-Arai, O.; Weyand, S.; Shimamura, T.; Nomura, N.; et al. G-protein-coupled receptor inactivation by an allosteric inverse-agonist antibody. *Nature* **2012**, *482*, 237–240. [[CrossRef](#)]
39. Jamshad, M.; Charlton, J.; Lin, Y.-P.; Routledge, S.J.; Bawa, Z.; Knowles, T.J.; Overduin, M.; Dekker, N.; Dafforn, T.R.; Bill, R.M.; et al. G-protein coupled receptor solubilization and purification for biophysical analysis and functional studies, in the total absence of detergent. *Biosci. Rep.* **2015**, *35*, e00188. [[CrossRef](#)]
40. Lengger, B.; Jensen, M.K. Engineering G protein-coupled receptor signalling in yeast for biotechnological and medical purposes. *FEMS Yeast Res.* **2020**, *20*, foz087. [[CrossRef](#)]
41. Shiroishi, M.; Tsujimoto, H.; Makyio, H.; Asada, H.; Yurugi-Kobayashi, T.; Shimamura, T.; Murata, T.; Nomura, N.; Haga, T.; Iwata, S.; et al. Platform for the rapid construction and evaluation of GPCRs for crystallography in *Saccharomyces cerevisiae*. *Microb. Cell Fact.* **2012**, *11*, 78. [[CrossRef](#)]
42. Bertheleme, N.; Strege, A.; Bunting, S.E.; Dowell, S.J.; Byrne, B. Arginine 199 and leucine 208 have key roles in the control of adenosine A<sub>2A</sub> receptor signalling function. *PLoS ONE* **2014**, *9*, e89613. [[CrossRef](#)]
43. Mitsumoto, M.; Sugaya, K.; Kazama, K.; Nakano, R.; Kosugi, T.; Murata, T.; Koga, N. State-Targeting Stabilization of Adenosine A<sub>2A</sub> Receptor by Fusing a Custom-Made de Novo Designed  $\alpha$ -Helical Protein. *Int. J. Mol. Sci.* **2021**, *22*, 12906. [[CrossRef](#)]
44. Wang, X.; van Westen, G.J.P.; Heitman, L.H.; IJzerman, A.P. G protein-coupled receptors expressed and studied in yeast. The adenosine receptor as a prime example. *Biochem. Pharmacol.* **2021**, *187*, 114370. [[CrossRef](#)]
45. Meltzer, M.; Zvagelsky, T.; Hadad, U.; Papo, N.; Engel, S. Yeast-based directed-evolution for high-throughput structural stabilization of G protein-coupled receptors (GPCRs). *Sci. Rep.* **2022**, *12*, 8657. [[CrossRef](#)]
46. Yamamoto, T.; Yasuda, S.; Kasai, R.S.; Nakano, R.; Hikiri, S.; Sugaya, K.; Hayashi, T.; Ogasawara, S.; Shiroishi, M.; Fujiwara, T.K.; et al. A methodology for creating mutants of G-protein coupled receptors stabilized in active state by combining statistical thermodynamics and evolutionary molecular engineering. *Protein Sci.* **2022**, *31*, e4425. [[CrossRef](#)] [[PubMed](#)]
47. Eddy, M.T.; Lee, M.-Y.; Gao, Z.-G.; White, K.L.; Didenko, T.; Horst, R.; Audet, M.; Stanczak, P.; McClary, K.M.; Han, G.W.; et al. Allosteric Coupling of Drug Binding and Intracellular Signaling in the A<sub>2A</sub> Adenosine Receptor. *Cell* **2018**, *172*, 68–80.e12. [[CrossRef](#)]
48. Ye, L.; van Eps, N.; Zimmer, M.; Ernst, O.P.; Prosser, R.S. Activation of the A<sub>2A</sub> adenosine G-protein-coupled receptor by conformational selection. *Nature* **2016**, *533*, 265–268. [[CrossRef](#)] [[PubMed](#)]
49. Mizumura, T.; Kondo, K.; Kurita, M.; Kofuku, Y.; Natsume, M.; Imai, S.; Shiraishi, Y.; Ueda, T.; Shimada, I. Activation of adenosine A<sub>2A</sub> receptor by lipids from docosahexaenoic acid revealed by NMR. *Sci. Adv.* **2020**, *6*, eaay8544. [[CrossRef](#)] [[PubMed](#)]
50. Huang, S.K.; Pandey, A.; Tran, D.P.; Villanueva, N.L.; Kitao, A.; Sunahara, R.K.; Sljoka, A.; Prosser, R.S. Delineating the conformational landscape of the adenosine A<sub>2A</sub> receptor during G protein coupling. *Cell* **2021**, *184*, 1884–1894.e14. [[CrossRef](#)] [[PubMed](#)]

51. Jaakola, V.-P.; Griffith, M.T.; Hanson, M.A.; Cherezov, V.; Chien, E.Y.T.; Lane, J.R.; IJzerman, A.P.; Stevens, R.C. The 2.6 angstrom crystal structure of a human A<sub>2A</sub> adenosine receptor bound to an antagonist. *Science* **2008**, *322*, 1211–1217. [[CrossRef](#)] [[PubMed](#)]
52. Lebon, G.; Warne, T.; Edwards, P.C.; Bennett, K.; Langmead, C.J.; Leslie, A.G.W.; Tate, C.G. Agonist-bound adenosine A<sub>2A</sub> receptor structures reveal common features of GPCR activation. *Nature* **2011**, *474*, 521–525. [[CrossRef](#)]
53. Carpenter, B.; Nehmé, R.; Warne, T.; Leslie, A.G.W.; Tate, C.G. Structure of the adenosine A<sub>2A</sub> receptor bound to an engineered G protein. *Nature* **2016**, *536*, 104–107. [[CrossRef](#)]
54. Miyagi, H.; Asada, H.; Suzuki, M.; Takahashi, Y.; Yasunaga, M.; Suno, C.; Iwata, S.; Saito, J.-I. The discovery of a new antibody for BRIL-fused GPCR structure determination. *Sci. Rep.* **2020**, *10*, 11669. [[CrossRef](#)]
55. Claff, T.; Klapschinski, T.A.; Tiruttani Subhramanyam, U.K.; Vaaßen, V.J.; Schlegel, J.G.; Vielmuth, C.; Voß, J.H.; Labahn, J.; Müller, C.E. Single Stabilizing Point Mutation Enables High-Resolution Co-Crystal Structures of the Adenosine A<sub>2A</sub> Receptor with Preladenant Conjugates. *Angew. Chem. Int. Ed Engl.* **2022**, *61*, e202115545. [[CrossRef](#)]
56. Liu, W.; Chun, E.; Thompson, A.A.; Chubukov, P.; Xu, F.; Katritch, V.; Han, G.W.; Roth, C.B.; Heitman, L.H.; IJzerman, A.P.; et al. Structural basis for allosteric regulation of GPCRs by sodium ions. *Science* **2012**, *337*, 232–236. [[CrossRef](#)] [[PubMed](#)]
57. Lebon, G.; Edwards, P.C.; Leslie, A.G.W.; Tate, C.G. Molecular Determinants of CGS21680 Binding to the Human Adenosine A<sub>2A</sub> Receptor. *Mol. Pharmacol.* **2015**, *87*, 907–915. [[CrossRef](#)] [[PubMed](#)]
58. White, K.L.; Eddy, M.T.; Gao, Z.-G.; Han, G.W.; Lian, T.; Deary, A.; Patel, N.; Jacobson, K.A.; Katritch, V.; Stevens, R.C. Structural Connection between Activation Microswitch and Allosteric Sodium Site in GPCR Signaling. *Structure* **2018**, *26*, 259–269.e5. [[CrossRef](#)]
59. McGraw, C.; Koretz, K.S.; Oseid, D.; Lyman, E.; Robinson, A.S. Cholesterol Dependent Activity of the Adenosine A<sub>2A</sub> Receptor Is Modulated via the Cholesterol Consensus Motif. *Molecules* **2022**, *27*, 3529. [[CrossRef](#)]
60. Yeliseev, A.; van den Berg, A.; Zoubak, L.; Hines, K.; Stepnowski, S.; Williston, K.; Yan, W.; Gawrisch, K.; Zmuda, J. Thermostability of a recombinant G protein-coupled receptor expressed at high level in mammalian cell culture. *Sci. Rep.* **2020**, *10*, 16805. [[CrossRef](#)]
61. Zhang, K.; Wu, H.; Hoppe, N.; Manglik, A.; Cheng, Y. Fusion protein strategies for cryo-EM study of G protein-coupled receptors. *Nat. Commun.* **2022**, *13*, 4366. [[CrossRef](#)]
62. Bocquet, N.; Kohler, J.; Hug, M.N.; Kuszniir, E.A.; Rufer, A.C.; Dawson, R.J.; Hennig, M.; Ruf, A.; Huber, W.; Huber, S. Real-time monitoring of binding events on a thermostabilized human A<sub>2A</sub> receptor embedded in a lipid bilayer by surface plasmon resonance. *Biochim. Biophys. Acta* **2015**, *1848*, 1224–1233. [[CrossRef](#)] [[PubMed](#)]
63. Suzuki, N.; Takamuku, Y.; Asakawa, T.; Inai, M.; Hino, T.; Iwata, S.; Kan, T.; Murata, T. An efficient screening method for purifying and crystallizing membrane proteins using modified clear-native PAGE. *Anal. Biochem.* **2018**, *548*, 7–14. [[CrossRef](#)] [[PubMed](#)]
64. Carpenter, B.; Lebon, G. Human Adenosine A<sub>2A</sub> Receptor: Molecular Mechanism of Ligand Binding and Activation. *Front. Pharmacol.* **2017**, *8*, 898. [[CrossRef](#)]
65. Cui, M.; Zhou, Q.; Xu, Y.; Weng, Y.; Yao, D.; Zhao, S.; Song, G. Crystal structure of a constitutive active mutant of adenosine A<sub>2A</sub> receptor. *IUCr* **2022**, *9*, 333–341. [[CrossRef](#)] [[PubMed](#)]
66. Lundstrom, K.; Wagner, R.; Reinhart, C.; Desmyter, A.; Cherouati, N.; Magnin, T.; Zeder-Lutz, G.; Courtot, M.; Prual, C.; André, N.; et al. Structural genomics on membrane proteins: Comparison of more than 100 GPCRs in 3 expression systems. *J. Struct. Funct. Genom.* **2006**, *7*, 77–91. [[CrossRef](#)] [[PubMed](#)]
67. Clark, L.D.; Dikay, I.; Chapman, K.; Rödröm, K.E.; Aramini, J.; LeVine, M.V.; Khelashvili, G.; Rasmussen, S.G.; Gardner, K.H.; Rosenbaum, D.M. Ligand modulation of sidechain dynamics in a wild-type human GPCR. *eLife* **2017**, *6*, e28505. [[CrossRef](#)] [[PubMed](#)]
68. Xu, F.; Wu, H.; Katritch, V.; Han, G.W.; Jacobson, K.A.; Gao, Z.-G.; Cherezov, V.; Stevens, R.C. Structure of an agonist-bound human A<sub>2A</sub> adenosine receptor. *Science* **2011**, *332*, 322–327. [[CrossRef](#)] [[PubMed](#)]
69. Cheng, R.K.Y.; Segala, E.; Robertson, N.; Deflorian, F.; Doré, A.S.; Errey, J.C.; Fiez-Vandal, C.; Marshall, F.H.; Cooke, R.M. Structures of Human A<sub>1</sub> and A<sub>2A</sub> Adenosine Receptors with Xanthines Reveal Determinants of Selectivity. *Structure* **2017**, *25*, 1275–1285.e4. [[CrossRef](#)]
70. Jespers, W.; Verdon, G.; Azuaje, J.; Majellaro, M.; Keränen, H.; García-Mera, X.; Congreve, M.; Deflorian, F.; de Graaf, C.; Zhukov, A.; et al. X-ray Crystallography and Free Energy Calculations Reveal the Binding Mechanism of A<sub>2A</sub> Adenosine Receptor Antagonists. *Angew. Chem. Int. Ed Engl.* **2020**, *59*, 16536–16543. [[CrossRef](#)] [[PubMed](#)]
71. Chun, E.; Thompson, A.A.; Liu, W.; Roth, C.B.; Griffith, M.T.; Katritch, V.; Kunken, J.; Xu, F.; Cherezov, V.; Hanson, M.A.; et al. Fusion partner toolchest for the stabilization and crystallization of G protein-coupled receptors. *Structure* **2012**, *20*, 967–976. [[CrossRef](#)] [[PubMed](#)]
72. Doré, A.S.; Robertson, N.; Errey, J.C.; Ng, I.; Hollenstein, K.; Tehan, B.; Hurrell, E.; Bennett, K.; Congreve, M.; Magnani, F.; et al. Structure of the adenosine A<sub>2A</sub> receptor in complex with ZM241385 and the xanthines XAC and caffeine. *Structure* **2011**, *19*, 1283–1293. [[CrossRef](#)] [[PubMed](#)]
73. Sušac, L.; Eddy, M.T.; Didenko, T.; Stevens, R.C.; Wüthrich, K. A<sub>2A</sub> adenosine receptor functional states characterized by 19F-NMR. *Proc. Natl. Acad. Sci. USA* **2018**, *115*, 12733–12738. [[CrossRef](#)]
74. Huang, S.K.; Almurad, O.; Pejana, R.J.; Morrison, Z.A.; Pandey, A.; Picard, L.-P.; Nitz, M.; Sljoka, A.; Prosser, R.S. Allosteric modulation of the adenosine A<sub>2A</sub> receptor by cholesterol. *eLife* **2022**, *11*, e73901. [[CrossRef](#)]

75. Thakur, N.; Wei, S.; Ray, A.P.; Lamichhane, R.; Eddy, M.T. Production of human A<sub>2A</sub> AR in lipid nanodiscs for 19F-NMR and single-molecule fluorescence spectroscopy. *STAR Protoc.* **2022**, *3*, 101535. [\[CrossRef\]](#) [\[PubMed\]](#)
76. Choy, B.C.; Cater, R.J.; Mancía, F.; Pryor, E.E. A 10-year meta-analysis of membrane protein structural biology: Detergents, membrane mimetics, and structure determination techniques. *Biochim. Biophys. Acta Biomembr.* **2021**, *1863*, 183533. [\[CrossRef\]](#)
77. Lecas, L.; Hartmann, L.; Caro, L.; Mohamed-Bouteben, S.; Raingeval, C.; Krimm, I.; Wagner, R.; Dugas, V.; Demesmay, C. Miniaturized weak affinity chromatography for ligand identification of nanodiscs-embedded G-protein coupled receptors. *Anal. Chim. Acta* **2020**, *1113*, 26–35. [\[CrossRef\]](#)
78. Lee, M.-Y.; Geiger, J.; Ishchenko, A.; Han, G.W.; Barty, A.; White, T.A.; Gati, C.; Batyuk, A.; Hunter, M.S.; Aquila, A.; et al. Harnessing the power of an X-ray laser for serial crystallography of membrane proteins crystallized in lipidic cubic phase. *IUCr* **2020**, *7*, 976–984. [\[CrossRef\]](#)
79. Segala, E.; Guo, D.; Cheng, R.K.Y.; Bortolato, A.; Deflorian, F.; Doré, A.S.; Errey, J.C.; Heitman, L.H.; IJzerman, A.P.; Marshall, F.H.; et al. Controlling the Dissociation of Ligands from the Adenosine A<sub>2A</sub> Receptor through Modulation of Salt Bridge Strength. *J. Med. Chem.* **2016**, *59*, 6470–6479. [\[CrossRef\]](#)
80. Reis, R.I.; Moraes, I. Probing Membrane Protein Assembly into Nanodiscs by In Situ Dynamic Light Scattering: A<sub>2A</sub> Receptor as a Case Study. *Biology* **2020**, *9*, 400. [\[CrossRef\]](#)
81. Batyuk, A.; Galli, L.; Ishchenko, A.; Han, G.W.; Gati, C.; Popov, P.A.; Lee, M.-Y.; Stauch, B.; White, T.A.; Barty, A.; et al. Native phasing of x-ray free-electron laser data for a G protein-coupled receptor. *Sci. Adv.* **2016**, *2*, e1600292. [\[CrossRef\]](#) [\[PubMed\]](#)
82. Dikiy, I.; Clark, L.D.; Gardner, K.H.; Rosenbaum, D.M. Isotopic Labeling of Eukaryotic Membrane Proteins for NMR Studies of Interactions and Dynamics. *Methods Enzymol.* **2019**, *614*, 37–65. [\[CrossRef\]](#) [\[PubMed\]](#)
83. O'Malley, M.A.; Naranjo, A.N.; Lazarova, T.; Robinson, A.S. Analysis of Adenosine A<sub>2A</sub> Receptor Stability: Effects of Ligands and Disulfide Bonds. *Biochemistry* **2010**, *49*, 9181–9189. [\[CrossRef\]](#) [\[PubMed\]](#)
84. Ashok, Y.; Nanekar, R.; Jaakola, V.-P. Defining thermostability of membrane proteins by western blotting. *Protein Eng. Des. Sel.* **2015**, *28*, 539–542. [\[CrossRef\]](#)
85. Zhang, X.; Stevens, R.C.; Xu, F. The importance of ligands for G protein-coupled receptor stability. *Trends Biochem. Sci.* **2015**, *40*, 79–87. [\[CrossRef\]](#)
86. Thompson, A.A.; Liu, J.J.; Chun, E.; Wacker, D.; Wu, H.; Cherezov, V.; Stevens, R.C. GPCR stabilization using the bicelle-like architecture of mixed sterol-detergent micelles. *Methods* **2011**, *55*, 310–317. [\[CrossRef\]](#)
87. O'Malley, M.A.; Helgeson, M.E.; Wagner, N.J.; Robinson, A.S. The morphology and composition of cholesterol-rich micellar nanostructures determine transmembrane protein (GPCR) activity. *Biophys. J.* **2011**, *100*, L11–L13. [\[CrossRef\]](#)
88. Vagenende, V.; Yap, M.G.S.; Trout, B.L. Mechanisms of protein stabilization and prevention of protein aggregation by glycerol. *Biochemistry* **2009**, *48*, 11084–11096. [\[CrossRef\]](#)
89. Wittmann, H.-J.; Seifert, R.; Strasser, A. Mathematical analysis of the sodium sensitivity of the human histamine H<sub>3</sub> receptor. *Silico Pharmacol.* **2014**, *2*, 1. [\[CrossRef\]](#)
90. Ross, P.; Weihofen, W.; Siu, F.; Xie, A.; Katakia, H.; Wright, S.K.; Hunt, I.; Brown, R.K.; Freire, E. Isothermal chemical denaturation to determine binding affinity of small molecules to G-protein coupled receptors. *Anal. Biochem.* **2015**, *473*, 41–45. [\[CrossRef\]](#)
91. Igonet, S.; Raingeval, C.; Cecon, E.; Pučić-Baković, M.; Lauc, G.; Cala, O.; Baranowski, M.; Perez, J.; Jockers, R.; Krimm, I.; et al. Enabling STD-NMR fragment screening using stabilized native GPCR: A case study of adenosine receptor. *Sci. Rep.* **2018**, *8*, 8142. [\[CrossRef\]](#)
92. Ukena, D.; Jacobson, K.A.; Kirk, K.L.; Daly, J.W. A [<sup>3</sup>H]amine congener of 1,3-dipropyl-8-phenylxanthine. *FEBS Lett.* **1986**, *199*, 269–274. [\[CrossRef\]](#)
93. Ye, L.; Neale, C.; Slijo, A.; Lyda, B.; Pichugin, D.; Tsuchimura, N.; Larda, S.T.; Pomès, R.; García, A.E.; Ernst, O.P.; et al. Mechanistic insights into allosteric regulation of the A<sub>2A</sub> adenosine G protein-coupled receptor by physiological cations. *Nat. Commun.* **2018**, *9*, 1372. [\[CrossRef\]](#) [\[PubMed\]](#)
94. Nguyen, K.D.Q.; Vigers, M.; Sefah, E.; Seppälä, S.; Hoover, J.P.; Schonenbach, N.S.; Mertz, B.; O'Malley, M.A.; Han, S. Homooligomerization of the human adenosine A<sub>2A</sub> receptor is driven by the intrinsically disordered C-terminus. *eLife* **2021**, *10*, e66662. [\[CrossRef\]](#)
95. Jones, A.J.Y.; Gabriel, F.; Tandale, A.; Nietlispach, D. Structure and Dynamics of GPCRs in Lipid Membranes: Physical Principles and Experimental Approaches. *Molecules* **2020**, *25*, 4729. [\[CrossRef\]](#)
96. Serebryany, E.; Zhu, G.A.; Yan, E.C.Y. Artificial membrane-like environments for in vitro studies of purified G-protein coupled receptors. *Biochim. Biophys. Acta* **2012**, *1818*, 225–233. [\[CrossRef\]](#) [\[PubMed\]](#)
97. Overduin, M.; Trieber, C.; Prosser, R.S.; Picard, L.-P.; Sheff, J.G. Structures and Dynamics of Native-State Transmembrane Protein Targets and Bound Lipids. *Membranes* **2021**, *11*, 451. [\[CrossRef\]](#)
98. Park, S.H.; Das, B.B.; Casagrande, F.; Tian, Y.; Nothnagel, H.J.; Chu, M.; Kiefer, H.; Maier, K.; de Angelis, A.A.; Marassi, F.M.; et al. Structure of the chemokine receptor CXCR1 in phospholipid bilayers. *Nature* **2012**, *491*, 779–783. [\[CrossRef\]](#) [\[PubMed\]](#)
99. Duc, N.M.; Du, Y.; Zhang, C.; Lee, S.Y.; Thorsen, T.S.; Kobilka, B.K.; Chung, K.Y. Effective application of bicelles for conformational analysis of G protein-coupled receptors by hydrogen/deuterium exchange mass spectrometry. *J. Am. Soc. Mass Spectrom.* **2015**, *26*, 808–817. [\[CrossRef\]](#)
100. Schmidt, P.; Bender, B.J.; Kaiser, A.; Gulati, K.; Scheidt, H.A.; Hamm, H.E.; Meiler, J.; Beck-Sicking, A.G.; Huster, D. Improved in Vitro Folding of the Y2 G Protein-Coupled Receptor into Bicelles. *Front. Mol. Biosci.* **2017**, *4*, 100. [\[CrossRef\]](#)



101. Brea, R.J.; Cole, C.M.; Lyda, B.R.; Ye, L.; Prosser, R.S.; Sunahara, R.K.; Devaraj, N.K. In Situ Reconstitution of the Adenosine A<sub>2A</sub> Receptor in Spontaneously Formed Synthetic Liposomes. *J. Am. Chem. Soc.* **2017**, *139*, 3607–3610. [\[CrossRef\]](#)
102. Bayburt, T.H.; Grinkova, Y.V.; Sligar, S.G. Self-Assembly of Discoidal Phospholipid Bilayer Nanoparticles with Membrane Scaffold Proteins. *Nano Lett.* **2002**, *2*, 853–856. [\[CrossRef\]](#)
103. Ritchie, T.K.; Grinkova, Y.V.; Bayburt, T.H.; Denisov, I.G.; Zolnerchikov, J.K.; Atkins, W.M.; Sligar, S.G. Chapter 11—Reconstitution of membrane proteins in phospholipid bilayer nanodiscs. *Methods Enzymol.* **2009**, *464*, 211–231. [\[CrossRef\]](#)
104. Lavington, S.; Watts, A. Lipid nanoparticle technologies for the study of G protein-coupled receptors in lipid environments. *Biophys. Rev.* **2020**, *12*, 1287–1302. [\[CrossRef\]](#)
105. Segala, E.; Errey, J.C.; Fiez-Vandal, C.; Zhukov, A.; Cooke, R.M. Biosensor-based affinities and binding kinetics of small molecule antagonists to the adenosine A<sub>2A</sub> receptor reconstituted in HDL like particles. *FEBS Lett.* **2015**, *589*, 1399–1405. [\[CrossRef\]](#) [\[PubMed\]](#)
106. Ma, J.; Lu, Y.; Wu, D.; Peng, Y.; Loa-Kum-Cheung, W.; Peng, C.; Quinn, R.J.; Shui, W.; Liu, Z.-J. Ligand identification of the adenosine A<sub>2A</sub> receptor in self-assembled nanodiscs by affinity mass spectrometry. *Anal. Methods* **2017**, *9*, 5851–5858. [\[CrossRef\]](#)
107. Fredriksson, K.; Lottmann, P.; Hinz, S.; Onila, I.; Shymanets, A.; Harteneck, C.; Müller, C.E.; Griesinger, C.; Exner, T.E. Nanodiscs for INPHARMA NMR Characterization of GPCRs: Ligand Binding to the Human A<sub>2A</sub> Adenosine Receptor. *Angew. Chem. Int. Ed Engl.* **2017**, *56*, 5750–5754. [\[CrossRef\]](#)
108. Mary, S.; Damian, M.; Rahmeh, R.; Mouillac, B.; Marie, J.; Granier, S.; Banères, J.-L. Amphipols in G protein-coupled receptor pharmacology: What are they good for? *J. Membr. Biol.* **2014**, *247*, 853–860. [\[CrossRef\]](#)
109. Zoonens, M.; Popot, J.-L. Amphipols for each season. *J. Membr. Biol.* **2014**, *247*, 759–796. [\[CrossRef\]](#) [\[PubMed\]](#)
110. Overduin, M.; Klumperman, B. Advancing membrane biology with poly(styrene-co-maleic acid)-based native nanodiscs. *Eur. Polym. J.* **2019**, *110*, 63–68. [\[CrossRef\]](#)
111. Grime, R.L.; Goulding, J.; Uddin, R.; Stoddart, L.A.; Hill, S.J.; Poyner, D.R.; Briddon, S.J.; Wheatley, M. Single molecule binding of a ligand to a G-protein-coupled receptor in real time using fluorescence correlation spectroscopy, rendered possible by nano-encapsulation in styrene maleic acid lipid particles. *Nanoscale* **2020**, *12*, 11518–11525. [\[CrossRef\]](#)
112. Routledge, S.J.; Jamshad, M.; Little, H.A.; Lin, Y.-P.; Simms, J.; Thakker, A.; Spickett, C.M.; Bill, R.M.; Dafforn, T.R.; Poyner, D.R.; et al. Ligand-induced conformational changes in a SMALP-encapsulated GPCR. *Biochim. Biophys. Acta Biomembr.* **2020**, *1862*, 183235. [\[CrossRef\]](#) [\[PubMed\]](#)
113. Marty, M.T.; Hoi, K.K.; Robinson, C.V. Interfacing Membrane Mimetics with Mass Spectrometry. *Acc. Chem. Res.* **2016**, *49*, 2459–2467. [\[CrossRef\]](#) [\[PubMed\]](#)
114. Keener, J.E.; Zhang, G.; Marty, M.T. Native Mass Spectrometry of Membrane Proteins. *Anal. Chem.* **2021**, *93*, 583–597. [\[CrossRef\]](#)
115. Young, G.; Hundt, N.; Cole, D.; Fineberg, A.; Andrecka, J.; Tyler, A.; Olerinyova, A.; Ansari, A.; Marklund, E.G.; Collier, M.P.; et al. Quantitative mass imaging of single biological macromolecules. *Science* **2018**, *360*, 423–427. [\[CrossRef\]](#)
116. Olerinyova, A.; Sonn-Segev, A.; Gault, J.; Eichmann, C.; Schimpf, J.; Kopf, A.H.; Rudden, L.S.P.; Ashkinadze, D.; Bomba, R.; Frey, L.; et al. Mass Photometry of Membrane Proteins. *Chem* **2021**, *7*, 224–236. [\[CrossRef\]](#)
117. Schlick, K.H.; Lange, C.K.; Gillispie, G.D.; Cloninger, M.J. Characterization of protein aggregation via intrinsic fluorescence lifetime. *J. Am. Chem. Soc.* **2009**, *131*, 16608–16609. [\[CrossRef\]](#) [\[PubMed\]](#)
118. Singh, S.; Zhang, M.; Bertheleme, N.; Kara, E.; Strange, P.G.; Byrne, B. Radioligand binding analysis as a tool for quality control of GPCR production for structural characterization: Adenosine A<sub>2A</sub>R as a template for study. *Curr. Protoc. Protein Sci.* **2012**, *29*, 29.3. [\[CrossRef\]](#) [\[PubMed\]](#)
119. Lee, S.; Ghosh, S.; Jana, S.; Robertson, N.; Tate, C.G.; Vaidehi, N. How Do Branched Detergents Stabilize GPCRs in Micelles? *Biochemistry* **2020**, *59*, 2125–2134. [\[CrossRef\]](#)
120. Holdgate, G.; Embrey, K.; Milbradt, A.; Davies, G. Biophysical methods in early drug discovery. *ADMET DMPK* **2019**, *7*, 222–241. [\[CrossRef\]](#) [\[PubMed\]](#)
121. van der Velden, W.J.C.; Heitman, L.H.; Rosenkilde, M.M. Perspective: Implications of Ligand-Receptor Binding Kinetics for Therapeutic Targeting of G Protein-Coupled Receptors. *ACS Pharmacol. Transl. Sci.* **2020**, *3*, 179–189. [\[CrossRef\]](#)
122. Copeland, R.A. The drug-target residence time model: A 10-year retrospective. *Nat. Rev. Drug Discov.* **2016**, *15*, 87–95. [\[CrossRef\]](#)
123. Guo, D.; Heitman, L.H.; IJzerman, A.P. Kinetic Aspects of the Interaction between Ligand and G Protein-Coupled Receptor: The Case of the Adenosine Receptors. *Chem. Rev.* **2017**, *117*, 38–66. [\[CrossRef\]](#) [\[PubMed\]](#)
124. IJzerman, A.P.; Guo, D. Drug-Target Association Kinetics in Drug Discovery. *Trends Biochem. Sci.* **2019**, *44*, 861–871. [\[CrossRef\]](#)
125. Rich, R.L.; Myszk, D.G. Advances in surface plasmon resonance biosensor analysis. *Curr. Opin. Biotechnol.* **2000**, *11*, 54–61. [\[CrossRef\]](#)
126. Locatelli-Hoops, S.; Yeliseev, A.A.; Gawrisch, K.; Gorshkova, I. Surface plasmon resonance applied to G protein-coupled receptors. *Biomed. Spectrosc. Imaging* **2013**, *2*, 155–181. [\[CrossRef\]](#) [\[PubMed\]](#)
127. Patching, S.G. Surface plasmon resonance spectroscopy for characterisation of membrane protein-ligand interactions and its potential for drug discovery. *Biochim. Biophys. Acta* **2014**, *1838*, 43–55. [\[CrossRef\]](#)
128. Aristotelous, T.; Hopkins, A.L.; Navratilova, I. Surface plasmon resonance analysis of seven-transmembrane receptors. *Methods Enzymol.* **2015**, *556*, 499–525. [\[CrossRef\]](#)
129. Olaru, A.; Bala, C.; Jaffrezic-Renault, N.; Aboul-Enein, H.Y. Surface plasmon resonance (SPR) biosensors in pharmaceutical analysis. *Crit. Rev. Anal. Chem.* **2015**, *45*, 97–105. [\[CrossRef\]](#)

130. Giannetti, A.M. From experimental design to validated hits a comprehensive walk-through of fragment lead identification using surface plasmon resonance. *Methods Enzymol.* **2011**, *493*, 169–218. [[CrossRef](#)] [[PubMed](#)]
131. Rich, R.L.; Errey, J.; Marshall, F.; Myszk, D.G. Biacore analysis with stabilized G-protein-coupled receptors. *Anal. Biochem.* **2011**, *409*, 267–272. [[CrossRef](#)] [[PubMed](#)]
132. Shepherd, C.A.; Hopkins, A.L.; Navratilova, I. Fragment screening by SPR and advanced application to GPCRs. *Prog. Biophys. Mol. Biol.* **2014**, *116*, 113–123. [[CrossRef](#)] [[PubMed](#)]
133. Robertson, N.; Jazayeri, A.; Errey, J.; Baig, A.; Hurrell, E.; Zhukov, A.; Langmead, C.J.; Weir, M.; Marshall, F.H. The properties of thermostabilised G protein-coupled receptors (StaRs) and their use in drug discovery. *Neuropharmacology* **2011**, *60*, 36–44. [[CrossRef](#)] [[PubMed](#)]
134. Congreve, M.; Rich, R.L.; Myszk, D.G.; Figaroa, F.; Siegal, G.; Marshall, F.H. Fragment screening of stabilized G-protein-coupled receptors using biophysical methods. *Methods Enzymol.* **2011**, *493*, 115–136. [[CrossRef](#)]
135. Congreve, M.; Andrews, S.P.; Doré, A.S.; Hollenstein, K.; Hurrell, E.; Langmead, C.J.; Mason, J.S.; Ng, I.W.; Tehan, B.; Zhukov, A.; et al. Discovery of 1,2,4-triazine derivatives as adenosine A<sub>2A</sub> antagonists using structure based drug design. *J. Med. Chem.* **2012**, *55*, 1898–1903. [[CrossRef](#)]
136. Zhukov, A.; Andrews, S.P.; Errey, J.C.; Robertson, N.; Tehan, B.; Mason, J.S.; Marshall, F.H.; Weir, M.; Congreve, M. Biophysical mapping of the adenosine A<sub>2A</sub> receptor. *J. Med. Chem.* **2011**, *54*, 4312–4323. [[CrossRef](#)]
137. Roos, H.; Karlsson, R.; Nilshans, H.; Persson, A. Thermodynamic analysis of protein interactions with biosensor technology. *J. Mol. Recognit.* **1998**, *11*, 204–210. [[CrossRef](#)]
138. Babazada, H.; Alekberli, T.; Hajieva, P.; Farajov, E. Biosensor-based kinetic and thermodynamic characterization of opioids interaction with human  $\mu$ -opioid receptor. *Eur. J. Pharm. Sci.* **2019**, *138*, 105017. [[CrossRef](#)]
139. Deganutti, G.; Zhukov, A.; Deflorian, F.; Federico, S.; Spalluto, G.; Cooke, R.M.; Moro, S.; Mason, J.S.; Bortolato, A. Impact of protein-ligand solvation and desolvation on transition state thermodynamic properties of adenosine A<sub>2A</sub> ligand binding kinetics. *Silico Pharmacol.* **2017**, *5*, 16. [[CrossRef](#)]
140. Coyle, J.; Walser, R. Applied Biophysical Methods in Fragment-Based Drug Discovery. *SLAS Discov.* **2020**, *25*, 471–490. [[CrossRef](#)]
141. Lu, Y.; Qin, S.; Zhang, B.; Dai, A.; Cai, X.; Ma, M.; Gao, Z.-G.; Yang, D.; Stevens, R.C.; Jacobson, K.A.; et al. Accelerating the Throughput of Affinity Mass Spectrometry-Based Ligand Screening toward a G Protein-Coupled Receptor. *Anal. Chem.* **2019**, *91*, 8162–8169. [[CrossRef](#)] [[PubMed](#)]
142. Shepherd, C.; Robinson, S.; Berizzi, A.; Thompson, L.E.J.; Bird, L.; Culurgioni, S.; Varzandeh, S.; Rawlins, P.B.; Olsen, R.H.J.; Navratilova, I.H. Surface Plasmon Resonance Screening to Identify Active and Selective Adenosine Receptor Binding Fragments. *ACS Med. Chem. Lett.* **2022**, *13*, 1172–1181. [[CrossRef](#)]
143. Koretz, K.S.; McGraw, C.E.; Stradley, S.; Elbaradei, A.; Malmstadt, N.; Robinson, A.S. Characterization of binding kinetics of A<sub>2A</sub>R to G $\alpha$ s protein by surface plasmon resonance. *Biophys. J.* **2021**, *120*, 1641–1649. [[CrossRef](#)] [[PubMed](#)]
144. Damian, L. Isothermal titration calorimetry for studying protein-ligand interactions. *Methods Mol. Biol.* **2013**, *1008*, 103–118. [[CrossRef](#)]
145. Pierce, M.M.; Raman, C.S.; Nall, B.T. Isothermal titration calorimetry of protein-protein interactions. *Methods* **1999**, *19*, 213–221. [[CrossRef](#)]
146. Drackowski, P.; Matosiuk, D.; Jozwiak, K. Isothermal titration calorimetry in membrane protein research. *J. Pharm. Biomed. Anal.* **2014**, *87*, 313–325. [[CrossRef](#)]
147. Rajarathnam, K.; Rösger, J. Isothermal titration calorimetry of membrane proteins-progress and challenges. *Biochim. Biophys. Acta* **2014**, *1838*, 69–77. [[CrossRef](#)]
148. Magalhaes, A.C.; Dunn, H.; Ferguson, S.S. Regulation of GPCR activity, trafficking and localization by GPCR-interacting proteins. *Br. J. Pharmacol.* **2012**, *165*, 1717–1736. [[CrossRef](#)] [[PubMed](#)]
149. Klaasse, E.C.; IJzerman, A.P.; de Grip, W.J.; Beukers, M.W. Internalization and desensitization of adenosine receptors. *Purinergic Signal.* **2008**, *4*, 21–37. [[CrossRef](#)]
150. Burgueño, J.; Blake, D.J.; Benson, M.A.; Tinsley, C.L.; Esapa, C.T.; Canela, E.I.; Penela, P.; Mallol, J.; Mayor, F.; Lluís, C.; et al. The adenosine A<sub>2A</sub> receptor interacts with the actin-binding protein alpha-actinin. *J. Biol. Chem.* **2003**, *278*, 37545–37552. [[CrossRef](#)]
151. Woods, A.S.; Marcellino, D.; Jackson, S.N.; Franco, R.; Ferré, S.; Agnati, L.F.; Fuxe, K. How calmodulin interacts with the adenosine A<sub>2A</sub> and the dopamine D<sub>2</sub> receptors. *J. Proteome Res.* **2008**, *7*, 3428–3434. [[CrossRef](#)]
152. Milojevic, T.; Reiterer, V.; Stefan, E.; Korkhov, V.M.; Dorostkar, M.M.; Ducza, E.; Ogris, E.; Boehm, S.; Freissmuth, M.; Nanoff, C. The ubiquitin-specific protease Usp4 regulates the cell surface level of the A<sub>2A</sub> receptor. *Mol. Pharmacol.* **2006**, *69*, 1083–1094. [[CrossRef](#)] [[PubMed](#)]
153. Canela, L.; Luján, R.; Lluís, C.; Burgueño, J.; Mallol, J.; Canela, E.I.; Franco, R.; Ciruela, F. The neuronal Ca<sup>2+</sup>-binding protein 2 (NECAB2) interacts with the adenosine A<sub>2A</sub> receptor and modulates the cell surface expression and function of the receptor. *Mol. Cell. Neurosci.* **2007**, *36*, 1–12. [[CrossRef](#)] [[PubMed](#)]
154. Sun, C.-N.; Cheng, H.-C.; Chou, J.-L.; Lee, S.-Y.; Lin, Y.-W.; Lai, H.-L.; Chen, H.-M.; Chern, Y. Rescue of p53 blockage by the A<sub>2A</sub> adenosine receptor via a novel interacting protein, translin-associated protein X. *Mol. Pharmacol.* **2006**, *70*, 454–466. [[CrossRef](#)]
155. Gsandtner, I.; Charalambous, C.; Stefan, E.; Ogris, E.; Freissmuth, M.; Zetzula, J. Heterotrimeric G protein-independent signaling of a G protein-coupled receptor. Direct binding of ARNO/cytohesin-2 to the carboxyl terminus of the A<sub>2A</sub> adenosine receptor is necessary for sustained activation of the ERK/MAP kinase pathway. *J. Biol. Chem.* **2005**, *280*, 31898–31905. [[CrossRef](#)]



156. Navarro, G.; Hradsky, J.; Lluís, C.; Casadó, V.; McCormick, P.J.; Kreutz, M.R.; Mikhaylova, M. NCS-1 associates with adenosine A<sub>2A</sub> receptors and modulates receptor function. *Front. Mol. Neurosci.* **2012**, *5*, 53. [\[CrossRef\]](#) [\[PubMed\]](#)
157. Piirainen, H.; Hellman, M.; Tossavainen, H.; Permi, P.; Kursula, P.; Jaakola, V.-P. Human adenosine A<sub>2A</sub> receptor binds calmodulin with high affinity in a calcium-dependent manner. *Biophys. J.* **2015**, *108*, 903–917. [\[CrossRef\]](#) [\[PubMed\]](#)
158. Piirainen, H.; Taura, J.; Kursula, P.; Ciruela, F.; Jaakola, V.-P. Calcium modulates calmodulin/ $\alpha$ -actinin 1 interaction with and agonist-dependent internalization of the adenosine A<sub>2A</sub> receptor. *Biochim. Biophys. Acta Mol. Cell Res.* **2017**, *1864*, 674–686. [\[CrossRef\]](#)
159. Quast, R.B.; Margeat, E. Studying GPCR conformational dynamics by single molecule fluorescence. *Mol. Cell. Endocrinol.* **2019**, *493*, 110469. [\[CrossRef\]](#) [\[PubMed\]](#)
160. Tian, H.; Fürstenberg, A.; Huber, T. Labeling and Single-Molecule Methods To Monitor G Protein-Coupled Receptor Dynamics. *Chem. Rev.* **2017**, *117*, 186–245. [\[CrossRef\]](#)
161. Hoffmann, C.; Gaietta, G.; Bünnemann, M.; Adams, S.R.; Oberdorff-Maass, S.; Behr, B.; Vilardaga, J.-P.; Tsien, R.Y.; Ellisman, M.H.; Lohse, M.J. A FIAsh-based FRET approach to determine G protein-coupled receptor activation in living cells. *Nat. Methods* **2005**, *2*, 171–176. [\[CrossRef\]](#) [\[PubMed\]](#)
162. Stoddart, L.A.; Kondon, N.D.; Otun, O.; Harwood, C.R.; Patera, F.; Veprintsev, D.B.; Woolard, J.; Briddon, S.J.; Franks, H.A.; Hill, S.J.; et al. Ligand-directed covalent labelling of a GPCR with a fluorescent tag in live cells. *Commun. Biol.* **2020**, *3*, 722. [\[CrossRef\]](#) [\[PubMed\]](#)
163. Wei, S.; Thakur, N.; Ray, A.P.; Jin, B.; Obeng, S.; McCurdy, C.R.; McMahon, L.R.; Gutiérrez-de-Terán, H.; Eddy, M.T.; Lamichhane, R. Slow conformational dynamics of the human A<sub>2A</sub> adenosine receptor are temporally ordered. *Structure* **2022**, *30*, 329–337.e5. [\[CrossRef\]](#)
164. Fernandes, D.D.; Neale, C.; Gomes, G.-N.W.; Li, Y.; Malik, A.; Pandey, A.; Oraziatti, A.P.; Wang, X.; Ye, L.; Scott Prosser, R.; et al. Ligand modulation of the conformational dynamics of the A<sub>2A</sub> adenosine receptor revealed by single-molecule fluorescence. *Sci. Rep.* **2021**, *11*, 5910. [\[CrossRef\]](#) [\[PubMed\]](#)
165. Zemella, A.; Richter, T.; Thoring, L.; Kubick, S. A Combined Cell-Free Protein Synthesis and Fluorescence-Based Approach to Investigate GPCR Binding Properties. *Methods Mol. Biol.* **2019**, *1947*, 57–77. [\[CrossRef\]](#)
166. Raingeval, C.; Krimm, I. NMR investigation of protein-ligand interactions for G-protein coupled receptors. *Future Med. Chem.* **2019**, *11*, 1811–1825. [\[CrossRef\]](#)
167. Stauch, B.; Orts, J.; Carlomagno, T. The description of protein internal motions aids selection of ligand binding poses by the INPHARMA method. *J. Biomol. NMR* **2012**, *54*, 245–256. [\[CrossRef\]](#)
168. Yen, H.-Y.; Hoi, K.K.; Liko, I.; Hedger, G.; Horrell, M.R.; Song, W.; Wu, D.; Heine, P.; Warne, T.; Lee, Y.; et al. PtdIns(4,5)P<sub>2</sub> stabilizes active states of GPCRs and enhances selectivity of G-protein coupling. *Nature* **2018**, *559*, 423–427. [\[CrossRef\]](#)
169. Agasid, M.T.; Sørensen, L.; Urner, L.H.; Yan, J.; Robinson, C.V. The Effects of Sodium Ions on Ligand Binding and Conformational States of G Protein-Coupled Receptors-Insights from Mass Spectrometry. *J. Am. Chem. Soc.* **2021**, *143*, 4085–4089. [\[CrossRef\]](#) [\[PubMed\]](#)
170. Olsen, R.H.J.; English, J.G. Advancements in G protein-coupled receptor biosensors to study GPCR-G protein coupling. *Br. J. Pharmacol.* **2022**. [\[CrossRef\]](#)
171. Zhou, Y.; Meng, J.; Xu, C.; Liu, J. Multiple GPCR Functional Assays Based on Resonance Energy Transfer Sensors. *Front. Cell Dev. Biol.* **2021**, *9*, 611443. [\[CrossRef\]](#) [\[PubMed\]](#)
172. Milligan, G. Principles: Extending the utility of [35S]GTP $\gamma$ S binding assays. *Trends Pharmacol. Sci.* **2003**, *24*, 87–90. [\[CrossRef\]](#)
173. McEwen, D.P.; Gee, K.R.; Kang, H.C.; Neubig, R.R. Fluorescent BODIPY-GTP analogs: Real-time measurement of nucleotide binding to G proteins. *Anal. Biochem.* **2001**, *291*, 109–117. [\[CrossRef\]](#)
174. Ravyn, V.; Bostwick, J.R. Functional coupling of the Galpha(olf) variant XLGalpha(olf) with the human adenosine A<sub>2A</sub> receptor. *J. Recept. Signal Transduct. Res.* **2006**, *26*, 241–258. [\[CrossRef\]](#) [\[PubMed\]](#)
175. Beckner, R.L.; Zoubak, L.; Hines, K.G.; Gawrisch, K.; Yeliseev, A.A. Probing thermostability of detergent-solubilized CB2 receptor by parallel G protein-activation and ligand-binding assays. *J. Biol. Chem.* **2020**, *295*, 181–190. [\[CrossRef\]](#)
176. Arcemishère, L.; Sen, T.; Boudier, L.; Balestre, M.-N.; Gaibelet, G.; Detouillon, E.; Orcel, H.; Mendre, C.; Rahmeh, R.; Granier, S.; et al. Leukotriene BLT2 receptor monomers activate the G(i2) GTP-binding protein more efficiently than dimers. *J. Biol. Chem.* **2010**, *285*, 6337–6347. [\[CrossRef\]](#) [\[PubMed\]](#)
177. Inagaki, S.; Ghirlando, R.; White, J.F.; Gvozdenovic-Jeremic, J.; Northup, J.K.; Grishammer, R. Modulation of the interaction between neurotensin receptor NTS1 and Gq protein by lipid. *J. Mol. Biol.* **2012**, *417*, 95–111. [\[CrossRef\]](#) [\[PubMed\]](#)
178. Damian, M.; Marie, J.; Leyris, J.-P.; Fehrentz, J.-A.; Verdié, P.; Martinez, J.; Banères, J.-L.; Mary, S. High constitutive activity is an intrinsic feature of ghrelin receptor protein: A study with a functional monomeric GHS-R1a receptor reconstituted in lipid discs. *J. Biol. Chem.* **2012**, *287*, 3630–3641. [\[CrossRef\]](#)
179. Gregorio, G.G.; Masurel, M.; Hilger, D.; Terry, D.S.; Juette, M.; Zhao, H.; Zhou, Z.; Perez-Aguilar, J.M.; Hauge, M.; Mathiasen, S.; et al. Single-molecule analysis of ligand efficacy in  $\beta$ 2AR-G-protein activation. *Nature* **2017**, *547*, 68–73. [\[CrossRef\]](#)
180. Damian, M.; Louet, M.; Gomes, A.A.S.; M'Kadmi, C.; Denoyelle, S.; Cantel, S.; Mary, S.; Bisch, P.M.; Fehrentz, J.-A.; Catoire, L.J.; et al. Allosteric modulation of ghrelin receptor signaling by lipids. *Nat. Commun.* **2021**, *12*, 3938. [\[CrossRef\]](#) [\[PubMed\]](#)

181. Bayburt, T.H.; Vishnivetskiy, S.A.; McLean, M.A.; Morizumi, T.; Huang, C.-C.; Tesmer, J.J.G.; Ernst, O.P.; Sligar, S.G.; Gurevich, V.V. Monomeric rhodopsin is sufficient for normal rhodopsin kinase (GRK1) phosphorylation and arrestin-1 binding. *J. Biol. Chem.* **2011**, *286*, 1420–1428. [[CrossRef](#)] [[PubMed](#)]
182. Srivastava, A.; Baidya, M.; Dwivedi-Agnihotri, H.; Shukla, A.K. Site-directed labeling of  $\beta$ -arrestin with monobromobimane for measuring their interaction with G protein-coupled receptors. *Methods Enzymol.* **2020**, *633*, 271–280. [[CrossRef](#)] [[PubMed](#)]
183. Sommer, M.E.; Farrens, D.L. Arrestin can act as a regulator of rhodopsin photochemistry. *Vis. Res.* **2006**, *46*, 4532–4546. [[CrossRef](#)] [[PubMed](#)]
184. Rahmeh, R.; Damian, M.; Cottet, M.; Orcel, H.; Mendre, C.; Durroux, T.; Sharma, K.S.; Durand, G.; Pucci, B.; Trinquet, E.; et al. Structural insights into biased G protein-coupled receptor signaling revealed by fluorescence spectroscopy. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, 6733–6738. [[CrossRef](#)]
185. Kumari, P.; Srivastava, A.; Banerjee, R.; Ghosh, E.; Gupta, P.; Ranjan, R.; Chen, X.; Gupta, B.; Gupta, C.; Jaiman, D.; et al. Functional competence of a partially engaged GPCR- $\beta$ -arrestin complex. *Nat. Commun.* **2016**, *7*, 13416. [[CrossRef](#)]
186. Staus, D.P.; Wingler, L.M.; Choi, M.; Pani, B.; Manglik, A.; Kruse, A.C.; Lefkowitz, R.J. Sortase ligation enables homogeneous GPCR phosphorylation to reveal diversity in  $\beta$ -arrestin coupling. *Proc. Natl. Acad. Sci. USA* **2018**, *115*, 3834–3839. [[CrossRef](#)]
187. Prosser, R.S.; Ye, L.; Pandey, A.; Oraziotti, A. Activation processes in ligand-activated G protein-coupled receptors: A case study of the adenosine  $A_{2A}$  receptor. *Bioessays* **2017**, *39*, 1700072. [[CrossRef](#)] [[PubMed](#)]
188. Welihinda, A.A.; Kaur, M.; Greene, K.; Zhai, Y.; Amento, E.P. The adenosine metabolite inosine is a functional agonist of the adenosine  $A_{2A}$  receptor with a unique signaling bias. *Cell. Signal.* **2016**, *28*, 552–560. [[CrossRef](#)]
189. Kolb, P.; Kenakin, T.; Alexander, S.P.H.; Bermudez, M.; Bohn, L.M.; Breinholt, C.S.; Bouvier, M.; Hill, S.J.; Kostenis, E.; Martemyanov, K.A.; et al. Community guidelines for GPCR ligand bias: IUPHAR review 32. *Br. J. Pharmacol.* **2022**, *179*, 3651–3674. [[CrossRef](#)]
190. Benleulmi-Chaachoua, A.; Chen, L.; Sokolina, K.; Wong, V.; Jurisica, I.; Emerit, M.B.; Darmon, M.; Espin, A.; Stagljär, I.; Tafelmeyer, P.; et al. Protein interactome mining defines melatonin MT1 receptors as integral component of presynaptic protein complexes of neurons. *J. Pineal Res.* **2016**, *60*, 95–108. [[CrossRef](#)]
191. Overton, M.C.; Blumer, K.J. The extracellular N-terminal domain and transmembrane domains 1 and 2 mediate oligomerization of a yeast G protein-coupled receptor. *J. Biol. Chem.* **2002**, *277*, 41463–41472. [[CrossRef](#)] [[PubMed](#)]
192. Mahesh, G.; Jaiswal, P.; Dey, S.; Sengupta, J.; Mukherjee, S. Cloning, Expression, Purification and Characterization of Oligomeric States of the Native 5HT $_{2A}$  G-Protein-Coupled Receptor. *Protein Pept. Lett.* **2018**, *25*, 390–397. [[CrossRef](#)]
193. Fernández-Dueñas, V.; Bonaventura, J.; Aso, E.; Luján, R.; Ferré, S.; Ciruela, F. Overcoming the Challenges of Detecting GPCR Oligomerization in the Brain. *Curr. Neuropharmacol.* **2022**, *20*, 1035–1045. [[CrossRef](#)]
194. Mansoor, S.; Kayık, G.; Durdagi, S.; Sensoy, O. Mechanistic insight into the impact of a bivalent ligand on the structure and dynamics of a GPCR oligomer. *Comput. Struct. Biotechnol. J.* **2022**, *20*, 925–936. [[CrossRef](#)] [[PubMed](#)]
195. Canals, M.; Burgueño, J.; Marcellino, D.; Cabello, N.; Canela, E.I.; Mallol, J.; Agnati, L.; Ferré, S.; Bouvier, M.; Fuxe, K.; et al. Homodimerization of adenosine  $A_{2A}$  receptors: Qualitative and quantitative assessment by fluorescence and bioluminescence energy transfer. *J. Neurochem.* **2004**, *88*, 726–734. [[CrossRef](#)] [[PubMed](#)]
196. Ciruela, F.; Casadó, V.; Rodrigues, R.J.; Luján, R.; Burgueño, J.; Canals, M.; Borycz, J.; Rebola, N.; Goldberg, S.R.; Mallol, J.; et al. Presynaptic control of striatal glutamatergic neurotransmission by adenosine  $A_1$ - $A_{2A}$  receptor heteromers. *J. Neurosci.* **2006**, *26*, 2080–2087. [[CrossRef](#)]
197. Franco, R.; Cordoní, A.; Del Llinas Torrent, C.; Lillo, A.; Serrano-Marín, J.; Navarro, G.; Pardo, L. Structure and function of adenosine receptor heteromers. *Cell. Mol. Life Sci.* **2021**, *78*, 3957–3968. [[CrossRef](#)] [[PubMed](#)]
198. Carriba, P.; Ortiz, O.; Patkar, K.; Justinova, Z.; Stroik, J.; Themann, A.; Müller, C.; Woods, A.S.; Hope, B.T.; Ciruela, F.; et al. Striatal adenosine  $A_{2A}$  and cannabinoid CB1 receptors form functional heteromeric complexes that mediate the motor effects of cannabinoids. *Neuropsychopharmacology* **2007**, *32*, 2249–2259. [[CrossRef](#)] [[PubMed](#)]
199. Hillion, J.; Canals, M.; Torvinen, M.; Casado, V.; Scott, R.; Terasmaa, A.; Hansson, A.; Watson, S.; Olah, M.E.; Mallol, J.; et al. Coaggregation, cointernalization, and codesensitization of adenosine  $A_{2A}$  receptors and dopamine D $_2$  receptors. *J. Biol. Chem.* **2002**, *277*, 18091–18097. [[CrossRef](#)] [[PubMed](#)]
200. Canals, M.; Marcellino, D.; Fanelli, F.; Ciruela, F.; de Benedetti, P.; Goldberg, S.R.; Neve, K.; Fuxe, K.; Agnati, L.F.; Woods, A.S.; et al. Adenosine  $A_{2A}$ -dopamine D $_2$  receptor-receptor heteromerization: Qualitative and quantitative assessment by fluorescence and bioluminescence energy transfer. *J. Biol. Chem.* **2003**, *278*, 46741–46749. [[CrossRef](#)]
201. Kamiya, T.; Saitoh, O.; Yoshioka, K.; Nakata, H. Oligomerization of adenosine  $A_{2A}$  and dopamine D $_2$  receptors in living cells. *Biochem. Biophys. Res. Commun.* **2003**, *306*, 544–549. [[CrossRef](#)]
202. Torvinen, M.; Marcellino, D.; Canals, M.; Agnati, L.F.; Lluís, C.; Franco, R.; Fuxe, K. Adenosine  $A_{2A}$  receptor and dopamine D $_3$  receptor interactions: Evidence of functional  $A_{2A}$ /D $_3$  heteromeric complexes. *Mol. Pharmacol.* **2005**, *67*, 400–407. [[CrossRef](#)]
203. Cabello, N.; Gandía, J.; Bertarelli, D.C.G.; Watanabe, M.; Lluís, C.; Franco, R.; Ferré, S.; Luján, R.; Ciruela, F. Metabotropic glutamate type 5, dopamine D $_2$  and adenosine  $A_{2A}$  receptors form higher-order oligomers in living cells. *J. Neurochem.* **2009**, *109*, 1497–1507. [[CrossRef](#)] [[PubMed](#)]
204. Trifilieff, P.; Rives, M.-L.; Urizar, E.; Piskorski, R.A.; Vishwasrao, H.D.; Castrillon, J.; Schmauss, C.; Slättman, M.; Gullberg, M.; Javitch, J.A. Detection of antigen interactions ex vivo by proximity ligation assay: Endogenous dopamine D $_2$ -adenosine  $A_{2A}$  receptor complexes in the striatum. *Biotechniques* **2011**, *51*, 111–118. [[CrossRef](#)]

205. Fernández-Dueñas, V.; Gómez-Soler, M.; Valle-León, M.; Watanabe, M.; Ferrer, I.; Ciruela, F. Revealing Adenosine A<sub>2A</sub>-Dopamine D<sub>2</sub> Receptor Heteromers in Parkinson's Disease Post-Mortem Brain through a New AlphaScreen-Based Assay. *Int. J. Mol. Sci.* **2019**, *20*, 3600. [[CrossRef](#)] [[PubMed](#)]
206. Beggiato, S.; Antonelli, T.; Tomasini, M.C.; Borelli, A.C.; Agnati, L.F.; Tanganelli, S.; Fuxe, K.; Ferraro, L. Adenosine A<sub>2A</sub>-D<sub>2</sub> receptor-receptor interactions in putative heteromers in the regulation of the striato-pallidal gaba pathway: Possible relevance for parkinson's disease and its treatment. *Curr. Protein Pept. Sci.* **2014**, *15*, 673–680. [[CrossRef](#)]
207. Ferré, S.; Bonaventura, J.; Tomasi, D.; Navarro, G.; Moreno, E.; Cortés, A.; Lluís, C.; Casadó, V.; Volkow, N.D. Allosteric mechanisms within the adenosine A<sub>2A</sub>-dopamine D<sub>2</sub> receptor heterotetramer. *Neuropharmacology* **2016**, *104*, 154–160. [[CrossRef](#)]
208. Romero-Fernandez, W.; Taura, J.J.; Crans, R.A.J.; Lopez-Cano, M.; Fores-Pons, R.; Narváez, M.; Carlsson, J.; Ciruela, F.; Fuxe, K.; Borroto-Escuela, D.O. The mGlu5 Receptor Protomer-Mediated Dopamine D<sub>2</sub> Receptor Trans-Inhibition Is Dependent on the Adenosine A<sub>2A</sub> Receptor Protomer: Implications for Parkinson's Disease. *Mol. Neurobiol.* **2022**, *59*, 5955–5969. [[CrossRef](#)] [[PubMed](#)]
209. Schuler, M.A.; Denisov, I.G.; Sligar, S.G. Nanodiscs as a new tool to examine lipid-protein interactions. *Methods Mol. Biol.* **2013**, *974*, 415–433. [[CrossRef](#)] [[PubMed](#)]
210. White, J.F.; Grodnitzky, J.; Louis, J.M.; Trinh, L.B.; Shiloach, J.; Gutierrez, J.; Northup, J.K.; Grisshammer, R. Dimerization of the class A G protein-coupled neurotensin receptor NTS1 alters G protein interaction. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 12199–12204. [[CrossRef](#)] [[PubMed](#)]
211. Bayburt, T.H.; Leitz, A.J.; Xie, G.; Oprian, D.D.; Sligar, S.G. Transducin activation by nanoscale lipid bilayers containing one and two rhodopsins. *J. Biol. Chem.* **2007**, *282*, 14875–14881. [[CrossRef](#)]
212. El Moustaine, D.; Granier, S.; Doumazane, E.; Scholler, P.; Rahmeh, R.; Bron, P.; Mouillac, B.; Banères, J.-L.; Rondard, P.; Pin, J.-P. Distinct roles of metabotropic glutamate receptor dimerization in agonist activation and G-protein coupling. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, 16342–16347. [[CrossRef](#)]
213. Mary, S.; Fehrentz, J.-A.; Damian, M.; Gaibelet, G.; Orcel, H.; Verdié, P.; Mouillac, B.; Martinez, J.; Marie, J.; Banères, J.-L. Heterodimerization with Its splice variant blocks the ghrelin receptor 1a in a non-signaling conformation: A study with a purified heterodimer assembled into lipid discs. *J. Biol. Chem.* **2013**, *288*, 24656–24665. [[CrossRef](#)]
214. Mesnier, D.; Banères, J.-L. Cooperative conformational changes in a G-protein-coupled receptor dimer, the leukotriene B(4) receptor BLT1. *J. Biol. Chem.* **2004**, *279*, 49664–49670. [[CrossRef](#)]
215. Damian, M.; Pons, V.; Renault, P.; M'Kadmi, C.; Delort, B.; Hartmann, L.; Kaya, A.I.; Louet, M.; Gagne, D.; Ben Haj Salah, K.; et al. GHSR-D2R heteromerization modulates dopamine signaling through an effect on G protein conformation. *Proc. Natl. Acad. Sci. USA* **2018**, *115*, 4501–4506. [[CrossRef](#)]
216. Tsukamoto, H.; Sinha, A.; DeWitt, M.; Farrens, D.L. Monomeric rhodopsin is the minimal functional unit required for arrestin binding. *J. Mol. Biol.* **2010**, *399*, 501–511. [[CrossRef](#)] [[PubMed](#)]
217. Piirainen, H.; Ashok, Y.; Nanekar, R.T.; Jaakola, V.-P. Structural features of adenosine receptors: From crystal to function. *Biochim. Biophys. Acta* **2011**, *1808*, 1233–1244. [[CrossRef](#)]
218. Langelan, D.N.; Ngweniform, P.; Rainey, J.K. Biophysical characterization of G-protein coupled receptor-peptide ligand binding. *Biochem. Cell Biol.* **2011**, *89*, 98–105. [[CrossRef](#)] [[PubMed](#)]
219. Jumper, J.; Evans, R.; Pritzel, A.; Green, T.; Figurnov, M.; Ronneberger, O.; Tunyasuvunakool, K.; Bates, R.; Židek, A.; Potapenko, A.; et al. Highly accurate protein structure prediction with AlphaFold. *Nature* **2021**, *596*, 583–589. [[CrossRef](#)] [[PubMed](#)]
220. Varadi, M.; Anyango, S.; Deshpande, M.; Nair, S.; Natassia, C.; Yordanova, G.; Yuan, D.; Stroe, O.; Wood, G.; Laydon, A.; et al. AlphaFold Protein Structure Database: Massively expanding the structural coverage of protein-sequence space with high-accuracy models. *Nucleic Acids Res.* **2022**, *50*, D439–D444. [[CrossRef](#)] [[PubMed](#)]
221. Marcoux, J.; Cianférani, S. Towards integrative structural mass spectrometry: Benefits from hybrid approaches. *Methods* **2015**, *89*, 4–12. [[CrossRef](#)]
222. Dafun, A.S.; Marcoux, J. Structural mass spectrometry of membrane proteins. *Biochim. Biophys. Acta Proteins Proteom.* **2022**, *1870*, 140813. [[CrossRef](#)] [[PubMed](#)]
223. Khanal, A.; Pan, Y.; Brown, L.S.; Konermann, L. Pulsed hydrogen/deuterium exchange mass spectrometry for time-resolved membrane protein folding studies. *J. Mass Spectrom.* **2012**, *47*, 1620–1626. [[CrossRef](#)]
224. Masson, G.R.; Burke, J.E.; Ahn, N.G.; Anand, G.S.; Borchers, C.; Brier, S.; Bou-Assaf, G.M.; Engen, J.R.; Englander, S.W.; Faber, J.; et al. Recommendations for performing, interpreting and reporting hydrogen deuterium exchange mass spectrometry (HDX-MS) experiments. *Nat. Methods* **2019**, *16*, 595–602. [[CrossRef](#)]
225. Möller, I.R.; Slivacka, M.; Hausner, J.; Nielsen, A.K.; Pospíšilová, E.; Merkle, P.S.; Lišková, R.; Polák, M.; Loland, C.J.; Kádek, A.; et al. Improving the Sequence Coverage of Integral Membrane Proteins during Hydrogen/Deuterium Exchange Mass Spectrometry Experiments. *Anal. Chem.* **2019**, *91*, 10970–10978. [[CrossRef](#)]
226. Trabjerg, E.; Nazari, Z.E.; Rand, K.D. Conformational analysis of complex protein states by hydrogen/deuterium exchange mass spectrometry (HDX-MS): Challenges and emerging solutions. *TrAC Trends Anal. Chem.* **2018**, *106*, 125–138. [[CrossRef](#)]
227. Martens, C.; Politis, A. A glimpse into the molecular mechanism of integral membrane proteins through hydrogen-deuterium exchange mass spectrometry. *Protein Sci.* **2020**, *29*, 1285–1301. [[CrossRef](#)] [[PubMed](#)]



228. Zhang, Y.; Rempel, D.L.; Zhang, J.; Sharma, A.K.; Mirica, L.M.; Gross, M.L. Pulsed hydrogen-deuterium exchange mass spectrometry probes conformational changes in amyloid beta (A $\beta$ ) peptide aggregation. *Proc. Natl. Acad. Sci. USA* **2013**, *110*, 14604–14609. [[CrossRef](#)]
229. Du, Y.; Duc, N.M.; Rasmussen, S.G.F.; Hilger, D.; Kubiak, X.; Wang, L.; Bohon, J.; Kim, H.R.; Wegrecki, M.; Asuru, A.; et al. Assembly of a GPCR-G Protein Complex. *Cell* **2019**, *177*, 1232–1242.e11. [[CrossRef](#)]
230. Konermann, L.; Tong, X.; Pan, Y. Protein structure and dynamics studied by mass spectrometry: H/D exchange, hydroxyl radical labeling, and related approaches. *J. Mass Spectrom.* **2008**, *43*, 1021–1036. [[CrossRef](#)]
231. Wang, L.; Chance, M.R. Protein Footprinting Comes of Age: Mass Spectrometry for Biophysical Structure Assessment. *Mol. Cell. Proteom.* **2017**, *16*, 706–716. [[CrossRef](#)] [[PubMed](#)]
232. Iacobucci, C.; Piotrowski, C.; Aebersold, R.; Amaral, B.C.; Andrews, P.; Bernfur, K.; Borchers, C.; Brodie, N.I.; Bruce, J.E.; Cao, Y.; et al. First Community-Wide, Comparative Cross-Linking Mass Spectrometry Study. *Anal. Chem.* **2019**, *91*, 6953–6961. [[CrossRef](#)]
233. Steigenberger, B.; Albanese, P.; Heck, A.J.R.; Scheltema, R.A. To Cleave or Not To Cleave in XL-MS? *J. Am. Soc. Mass Spectrom.* **2020**, *31*, 196–206. [[CrossRef](#)] [[PubMed](#)]
234. Xia, L.; Ma, Z.; Tong, J.; Tang, Y.; Li, S.; Qin, S.; Lou, R.; Zhao, S.; Lei, X.; Shui, W. Evaluation of chemical cross-linkers for in-depth structural analysis of G protein-coupled receptors through cross-linking mass spectrometry. *Anal. Chim. Acta* **2020**, *1102*, 53–62. [[CrossRef](#)]
235. Jones, A.X.; Cao, Y.; Tang, Y.-L.; Wang, J.-H.; Ding, Y.-H.; Tan, H.; Chen, Z.-L.; Fang, R.-Q.; Yin, J.; Chen, R.-C.; et al. Improving mass spectrometry analysis of protein structures with arginine-selective chemical cross-linkers. *Nat. Commun.* **2019**, *10*, 3911. [[CrossRef](#)]
236. Rosenbaum, D.M.; Cherezov, V.; Hanson, M.A.; Rasmussen, S.G.F.; Thian, F.S.; Kobilka, T.S.; Choi, H.-J.; Yao, X.-J.; Weis, W.I.; Stevens, R.C.; et al. GPCR engineering yields high-resolution structural insights into beta2-adrenergic receptor function. *Science* **2007**, *318*, 1266–1273. [[CrossRef](#)]
237. Magnani, F.; Serrano-Vega, M.J.; Shibata, Y.; Abdul-Hussein, S.; Lebon, G.; Miller-Gallacher, J.; Singhal, A.; Strege, A.; Thomas, J.A.; Tate, C.G. A mutagenesis and screening strategy to generate optimally thermostabilized membrane proteins for structural studies. *Nat. Protoc.* **2016**, *11*, 1554–1571. [[CrossRef](#)] [[PubMed](#)]
238. Cianchetta, A.; Rubio, P.; Lieberman, D.I.; Jacobson, K.A. A<sub>3</sub> adenosine receptor activation mechanisms: Molecular dynamics analysis of inactive, active, and fully active states. *J. Comput. Aided Mol. Des.* **2019**, *33*, 983–996. [[CrossRef](#)] [[PubMed](#)]
239. Venkatakrisnan, A.J.; Deupi, X.; Lebon, G.; Tate, C.G.; Schertler, G.F.; Babu, M.M. Molecular signatures of G-protein-coupled receptors. *Nature* **2013**, *494*, 185–194. [[CrossRef](#)]
240. Rucktoo, P.; Cheng, R.K.Y.; Segala, E.; Geng, T.; Errey, J.C.; Brown, G.A.; Cooke, R.M.; Marshall, F.H.; Doré, A.S. Towards high throughput GPCR crystallography: In Meso soaking of Adenosine A<sub>2A</sub> Receptor crystals. *Sci. Rep.* **2018**, *8*, 41. [[CrossRef](#)]
241. García-Nafria, J.; Lee, Y.; Bai, X.; Carpenter, B.; Tate, C.G. Cryo-EM structure of the adenosine A<sub>2A</sub> receptor coupled to an engineered heterotrimeric G protein. *eLife* **2018**, *7*, e35946. [[CrossRef](#)]
242. Amelia, T.; van Veldhoven, J.P.D.; Falsini, M.; Liu, R.; Heitman, L.H.; van Westen, G.J.P.; Segala, E.; Verdon, G.; Cheng, R.K.Y.; Cooke, R.M.; et al. Crystal Structure and Subsequent Ligand Design of a Nonriboside Partial Agonist Bound to the Adenosine A<sub>2A</sub> Receptor. *J. Med. Chem.* **2021**, *64*, 3827–3842. [[CrossRef](#)] [[PubMed](#)]
243. Venkatakrisnan, A.J.; Deupi, X.; Lebon, G.; Heydenreich, F.M.; Flock, T.; Miljus, T.; Balaji, S.; Bouvier, M.; Veprintsev, D.B.; Tate, C.G.; et al. Diverse activation pathways in class A GPCRs converge near the G-protein-coupling region. *Nature* **2016**, *536*, 484–487. [[CrossRef](#)]
244. Shiriaeva, A.; Park, D.; Kim, G.; Lee, Y.; Hou, X.; Jarhad, D.B.; Kim, G.; Yu, J.; Hyun, Y.E.; Kim, W.; et al. GPCR Agonist-to-Antagonist Conversion: Enabling the Design of Nucleoside Functional Switches for the A<sub>2A</sub> Adenosine Receptor. *J. Med. Chem.* **2022**, *65*, 11648–11657. [[CrossRef](#)] [[PubMed](#)]
245. Sun, B.; Bachhawat, P.; Chu, M.L.-H.; Wood, M.; Ceska, T.; Sands, Z.A.; Mercier, J.; Lebon, F.; Kobilka, T.S.; Kobilka, B.K. Crystal structure of the adenosine A<sub>2A</sub> receptor bound to an antagonist reveals a potential allosteric pocket. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, 2066–2071. [[CrossRef](#)]
246. Martin-Garcia, J.M.; Conrad, C.E.; Nelson, G.; Stander, N.; Zatspein, N.A.; Zook, J.; Zhu, L.; Geiger, J.; Chun, E.; Kissick, D.; et al. Serial millisecond crystallography of membrane and soluble protein microcrystals using synchrotron radiation. *IUCr* **2017**, *4*, 439–454. [[CrossRef](#)] [[PubMed](#)]
247. Weinert, T.; Olieric, N.; Cheng, R.; Brünle, S.; James, D.; Ozerov, D.; Gashi, D.; Vera, L.; Marsh, M.; Jaeger, K.; et al. Serial millisecond crystallography for routine room-temperature structure determination at synchrotrons. *Nat. Commun.* **2017**, *8*, 542. [[CrossRef](#)]
248. Martin-Garcia, J.M.; Zhu, L.; Mendez, D.; Lee, M.-Y.; Chun, E.; Li, C.; Hu, H.; Subramanian, G.; Kissick, D.; Ogata, C.; et al. High-viscosity injector-based pink-beam serial crystallography of microcrystals at a synchrotron radiation source. *IUCr* **2019**, *6*, 412–425. [[CrossRef](#)]
249. Ishchenko, A.; Stauch, B.; Han, G.W.; Batyuk, A.; Shiriaeva, A.; Li, C.; Zatspein, N.; Weierstall, U.; Liu, W.; Nango, E.; et al. Toward G protein-coupled receptor structure-based drug design using X-ray lasers. *IUCr* **2019**, *6*, 1106–1119. [[CrossRef](#)] [[PubMed](#)]

250. Shimazu, Y.; Tono, K.; Tanaka, T.; Yamanaka, Y.; Nakane, T.; Mori, C.; Terakado Kimura, K.; Fujiwara, T.; Sugahara, M.; Tanaka, R.; et al. High-viscosity sample-injection device for serial femtosecond crystallography at atmospheric pressure. *J. Appl. Crystallogr.* **2019**, *52*, 1280–1288. [[CrossRef](#)]
251. Ihara, K.; Hato, M.; Nakane, T.; Yamashita, K.; Kimura-Someya, T.; Hosaka, T.; Ishizuka-Katsura, Y.; Tanaka, R.; Tanaka, T.; Sugahara, M.; et al. Isoprenoid-chained lipid EROCO17+4: A new matrix for membrane protein crystallization and a crystal delivery medium in serial femtosecond crystallography. *Sci. Rep.* **2020**, *10*, 19305. [[CrossRef](#)]
252. García-Nafria, J.; Tate, C.G. Structure determination of GPCRs: Cryo-EM compared with X-ray crystallography. *Biochem. Soc. Trans.* **2021**, *49*, 2345–2355. [[CrossRef](#)] [[PubMed](#)]
253. Vénien-Bryan, C.; Li, Z.; Vuillard, L.; Boutin, J.A. Cryo-electron microscopy and X-ray crystallography: Complementary approaches to structural biology and drug discovery. *Acta Crystallogr. F Struct. Biol. Commun.* **2017**, *73*, 174–183. [[CrossRef](#)] [[PubMed](#)]
254. Boutin, J.A.; Li, Z.; Vuillard, L.; Vénien-Bryan, C. La cryo-microscopie, une alternative à la cristallographie aux rayons X? *Med. Sci.* **2016**, *32*, 758–767. [[CrossRef](#)] [[PubMed](#)]
255. Rossmann, M.G.; Battisti, A.J.; Plevka, P. Future prospects. *Adv. Protein Chem. Struct. Biol.* **2011**, *82*, 101–121. [[CrossRef](#)]
256. Blundell, T.L.; Chaplin, A.K. The resolution revolution in X-ray diffraction, Cryo-EM and other Technologies. *Prog. Biophys. Mol. Biol.* **2021**, *160*, 2–4. [[CrossRef](#)]
257. Glukhova, A.; Draper-Joyce, C.J.; Sunahara, R.K.; Christopoulos, A.; Wootten, D.; Sexton, P.M. Rules of Engagement: GPCRs and G Proteins. *ACS Pharmacol. Transl. Sci.* **2018**, *1*, 73–83. [[CrossRef](#)]
258. Liang, Y.-L.; Khoshouei, M.; Radjainia, M.; Zhang, Y.; Glukhova, A.; Tarrasch, J.; Thal, D.M.; Furness, S.G.B.; Christopoulos, G.; Coudrat, T.; et al. Phase-plate cryo-EM structure of a class B GPCR-G-protein complex. *Nature* **2017**, *546*, 118–123. [[CrossRef](#)]
259. Martynowycz, M.W.; Shiriaeva, A.; Ge, X.; Hattne, J.; Nannenga, B.L.; Cherezov, V.; Gonen, T. MicroED structure of the human adenosine receptor determined from a single nanocrystal in LCP. *Proc. Natl. Acad. Sci. USA* **2021**, *118*, e2106041118. [[CrossRef](#)]
260. Frauenfelder, H.; Sligar, S.G.; Wolynes, P.G. The energy landscapes and motions of proteins. *Science* **1991**, *254*, 1598–1603. [[CrossRef](#)]
261. Kitevski-LeBlanc, J.L.; Prosser, R.S. Current applications of 19F NMR to studies of protein structure and dynamics. *Prog. Nucl. Magn. Reson. Spectrosc.* **2012**, *62*, 1–33. [[CrossRef](#)]
262. Picard, L.-P.; Prosser, R.S. Advances in the study of GPCRs by 19F NMR. *Curr. Opin. Struct. Biol.* **2021**, *69*, 169–176. [[CrossRef](#)]
263. Liu, D.; Wüthrich, K. Ring current shifts in (19)F-NMR of membrane proteins. *J. Biomol. NMR* **2016**, *65*, 1–5. [[CrossRef](#)] [[PubMed](#)]
264. Hagn, F.; Etzkorn, M.; Raschle, T.; Wagner, G. Optimized phospholipid bilayer nanodiscs facilitate high-resolution structure determination of membrane proteins. *J. Am. Chem. Soc.* **2013**, *135*, 1919–1925. [[CrossRef](#)]
265. Pozza, A.; Giraud, F.; Cece, Q.; Casiraghi, M.; Point, E.; Damian, M.; Le Bon, C.; Moncoq, K.; Banères, J.-L.; Lescop, E.; et al. Exploration of the dynamic interplay between lipids and membrane proteins by hydrostatic pressure. *Nat. Commun.* **2022**, *13*, 1780. [[CrossRef](#)] [[PubMed](#)]
266. Casiraghi, M.; Damian, M.; Lescop, E.; Point, E.; Moncoq, K.; Morellet, N.; Levy, D.; Marie, J.; Guittet, E.; Banères, J.-L.; et al. Functional Modulation of a G Protein-Coupled Receptor Conformational Landscape in a Lipid Bilayer. *J. Am. Chem. Soc.* **2016**, *138*, 11170–11175. [[CrossRef](#)]
267. Nygaard, R.; Frimurer, T.M.; Holst, B.; Rosenkilde, M.M.; Schwartz, T.W. Ligand binding and micro-switches in 7TM receptor structures. *Trends Pharmacol. Sci.* **2009**, *30*, 249–259. [[CrossRef](#)]
268. Eddy, M.T.; Gao, Z.-G.; Mannes, P.; Patel, N.; Jacobson, K.A.; Katritch, V.; Stevens, R.C.; Wüthrich, K. Extrinsic Tryptophans as NMR Probes of Allosteric Coupling in Membrane Proteins: Application to the A<sub>2A</sub> Adenosine Receptor. *J. Am. Chem. Soc.* **2018**, *140*, 8228–8235. [[CrossRef](#)]
269. Eddy, M.T.; Martin, B.T.; Wüthrich, K. A<sub>2A</sub> Adenosine Receptor Partial Agonism Related to Structural Rearrangements in an Activation Microswitch. *Structure* **2021**, *29*, 170–176.e3. [[CrossRef](#)] [[PubMed](#)]
270. Casiraghi, M.; Damian, M.; Lescop, E.; Banères, J.-L.; Catoire, L.J. Illuminating the Energy Landscape of GPCRs: The Key Contribution of Solution-State NMR Associated with *Escherichia coli* as an Expression Host. *Biochemistry* **2018**, *57*, 2297–2307. [[CrossRef](#)]
271. Sekhar, A.; Kay, L.E. NMR paves the way for atomic level descriptions of sparsely populated, transiently formed biomolecular conformers. *Proc. Natl. Acad. Sci. USA* **2013**, *110*, 12867–12874. [[CrossRef](#)]
272. Gunkel, M.; Schöneberg, J.; Alkhalidi, W.; Irsen, S.; Noé, F.; Kaupp, U.B.; Al-Amoudi, A. Higher-order architecture of rhodopsin in intact photoreceptors and its implication for phototransduction kinetics. *Structure* **2015**, *23*, 628–638. [[CrossRef](#)]
273. Fotiadis, D.; Liang, Y.; Filipek, S.; Saperstein, D.A.; Engel, A.; Palczewski, K. Atomic-force microscopy: Rhodopsin dimers in native disc membranes. *Nature* **2003**, *421*, 127–128. [[CrossRef](#)]
274. Sapra, K.T.; Spoerri, P.M.; Engel, A.; Alsteens, D.; Müller, D.J. Seeing and sensing single G protein-coupled receptors by atomic force microscopy. *Curr. Opin. Cell Biol.* **2019**, *57*, 25–32. [[CrossRef](#)] [[PubMed](#)]
275. Jacobson, K.A.; Tosh, D.K.; Jain, S.; Gao, Z.-G. Historical and Current Adenosine Receptor Agonists in Preclinical and Clinical Development. *Front. Cell. Neurosci.* **2019**, *13*, 124. [[CrossRef](#)]
276. Melman, A.; Gao, Z.-G.; Kumar, D.; Wan, T.C.; Gizewski, E.; Auchampach, J.A.; Jacobson, K.A. Design of (N)-methanocarba adenosine 5'-uronamides as species-independent A<sub>3</sub> receptor-selective agonists. *Bioorg. Med. Chem. Lett.* **2008**, *18*, 2813–2819. [[CrossRef](#)]



277. Ivanov, A.A.; Jacobson, K.A. Molecular modeling of a PAMAM-CGS21680 dendrimer bound to an A<sub>2A</sub> adenosine receptor homodimer. *Bioorg. Med. Chem. Lett.* **2008**, *18*, 4312–4315. [\[CrossRef\]](#) [\[PubMed\]](#)
278. Dal Ben, D.; Lambertucci, C.; Buccioni, M.; Martí Navia, A.; Marucci, G.; Spinaci, A.; Volpini, R. Non-Nucleoside Agonists of the Adenosine Receptors: An Overview. *Pharmaceuticals* **2019**, *12*, 150. [\[CrossRef\]](#) [\[PubMed\]](#)
279. Müller, C.E.; Jacobson, K.A. Xanthines as adenosine receptor antagonists. *Handb. Exp. Pharmacol.* **2011**, *200*, 151–199. [\[CrossRef\]](#)
280. Ukena, D.; Padgett, W.L.; Hong, O.; Daly, J.W.; Daly, D.T.; Olsson, R.A. N6-substituted 9-methyladenines: A new class of adenosine receptor antagonists. *FEBS Lett.* **1987**, *215*, 203–208. [\[CrossRef\]](#)
281. Zhou, X.; Khanapur, S.; Huizing, A.P.; Zijlma, R.; Schepers, M.; Dierckx, R.A.J.O.; van Waarde, A.; de Vries, E.F.J.; Elsinga, P.H. Synthesis and preclinical evaluation of 2-(2-furanyl)-7-2-4-4-(2-11Cmethoxyethoxy)phenyl-1-piperazinyethyl-7H-pyrazolo4,3-e1,2,4triazolo1,5-cpyrimidine-5-amine (11CPreladenant) as a PET tracer for the imaging of cerebral adenosine A<sub>2A</sub> receptors. *J. Med. Chem.* **2014**, *57*, 9204–9210. [\[CrossRef\]](#)
282. Shinkre, B.A.; Kumar, T.S.; Gao, Z.-G.; Deflorian, F.; Jacobson, K.A.; Trenkle, W.C. Synthesis and evaluation of 1,2,4-triazolo1,5-cpyrimidine derivatives as A<sub>2A</sub> receptor-selective antagonists. *Bioorg. Med. Chem. Lett.* **2010**, *20*, 5690–5694. [\[CrossRef\]](#) [\[PubMed\]](#)
283. Todde, S.; Moresco, R.M.; Simonelli, P.; Baraldi, P.G.; Cacciari, B.; Spalluto, G.; Varani, K.; Monopoli, A.; Matarrese, M.; Carpinelli, A.; et al. Design, radiosynthesis, and biodistribution of a new potent and selective ligand for in vivo imaging of the adenosine A<sub>2A</sub> receptor system using positron emission tomography. *J. Med. Chem.* **2000**, *43*, 4359–4362. [\[CrossRef\]](#) [\[PubMed\]](#)
284. Zocchi, C.; Ongini, E.; Conti, A.; Monopoli, A.; Negretti, A.; Baraldi, P.G.; Dionisotti, S. The non-xanthine heterocyclic compound SCH 58261 is a new potent and selective A<sub>2A</sub> adenosine receptor antagonist. *J. Pharmacol. Exp. Ther.* **1996**, *276*, 398–404. [\[PubMed\]](#)
285. Marques, T.R.; Natesan, S.; Rabiner, E.A.; Searle, G.E.; Gunn, R.; Howes, O.D.; Kapur, S. Adenosine A<sub>2A</sub> receptor in schizophrenia: An in vivo brain PET imaging study. *Psychopharmacology* **2021**, *239*, 3439–3445. [\[CrossRef\]](#)
286. Boutin, J.A.; Witt-Enderby, P.A.; Sotriffer, C.; Zlotos, D.P. Melatonin receptor ligands: A pharmaco-chemical perspective. *J. Pineal Res.* **2020**, *69*, e12672. [\[CrossRef\]](#)
287. Bojarski, A.J. Pharmacophore models for metabotropic 5-HT receptor ligands. *Curr. Top. Med. Chem.* **2006**, *6*, 2005–2026. [\[CrossRef\]](#)
288. Gao, Z.-G.; Jacobson, K.A. Allosteric modulation and functional selectivity of G protein-coupled receptors. *Drug Discov. Today Technol.* **2013**, *10*, e237–e243. [\[CrossRef\]](#)
289. Lütjens, R.; Rocher, J.-P. Recent advances in drug discovery of GPCR allosteric modulators for neurodegenerative disorders. *Curr. Opin. Pharmacol.* **2017**, *32*, 91–95. [\[CrossRef\]](#)
290. van der Westhuizen, E.T.; Valant, C.; Sexton, P.M.; Christopoulos, A. Endogenous allosteric modulators of G protein-coupled receptors. *J. Pharmacol. Exp. Ther.* **2015**, *353*, 246–260. [\[CrossRef\]](#)
291. Madsen, J.J.; Ye, L.; Frimurer, T.M.; Olsen, O.H. Mechanistic basis of GPCR activation explored by ensemble refinement of crystallographic structures. *Protein Sci.* **2022**, *31*, e4456. [\[CrossRef\]](#)
292. Swaminath, G.; Lee, T.W.; Kobilka, B. Identification of an allosteric binding site for Zn<sup>2+</sup> on the beta2 adrenergic receptor. *J. Biol. Chem.* **2003**, *278*, 352–356. [\[CrossRef\]](#)
293. Lu, Y.; Liu, H.; Yang, D.; Zhong, L.; Xin, Y.; Zhao, S.; Wang, M.-W.; Zhou, Q.; Shui, W. Affinity Mass Spectrometry-Based Fragment Screening Identified a New Negative Allosteric Modulator of the Adenosine A<sub>2A</sub> Receptor Targeting the Sodium Ion Pocket. *ACS Chem. Biol.* **2021**, *16*, 991–1002. [\[CrossRef\]](#) [\[PubMed\]](#)
294. Korkutata, M.; Saitoh, T.; Cherasse, Y.; Ioka, S.; Duo, F.; Qin, R.; Murakoshi, N.; Fujii, S.; Zhou, X.; Sugiyama, F.; et al. Enhancing endogenous adenosine A<sub>2A</sub> receptor signaling induces slow-wave sleep without affecting body temperature and cardiovascular function. *Neuropharmacology* **2019**, *144*, 122–132. [\[CrossRef\]](#) [\[PubMed\]](#)
295. Welihinda, A.A.; Amento, E.P. Positive allosteric modulation of the adenosine A<sub>2A</sub> receptor attenuates inflammation. *J. Inflamm.* **2014**, *11*, 37. [\[CrossRef\]](#) [\[PubMed\]](#)
296. Welihinda, A.A.; Kaur, M.; Raveendran, K.S.; Amento, E.P. Enhancement of inosine-mediated A<sub>2A</sub>R signaling through positive allosteric modulation. *Cell. Signal.* **2018**, *42*, 227–235. [\[CrossRef\]](#)
297. Salovich, J.M.; Sheffler, D.J.; Vinson, P.N.; Lamsal, A.; Utley, T.J.; Blobaum, A.L.; Bridges, T.M.; Le, U.; Jones, C.K.; Wood, M.R.; et al. *Probe Reports from the NIH Molecular Libraries Program: Discovery of a Novel Structural Class of M4 Positive Allosteric Modulators: Characterization of ML293, N-(4-Methoxy-7-Methylbenzothiazol-2-yl)isonicotinamide, with CNS Exposure in Rats*; National Center for Biotechnology Information (US): Bethesda, MD, USA, 2010.
298. Jones, C.K.; Bubser, M.; Thompson, A.D.; Dickerson, J.W.; Turle-Lorenzo, N.; Amalric, M.; Blobaum, A.L.; Bridges, T.M.; Morrison, R.D.; Jadhav, S.; et al. The metabotropic glutamate receptor 4-positive allosteric modulator VU0364770 produces efficacy alone and in combination with L-DOPA or an adenosine 2A antagonist in preclinical rodent models of Parkinson's disease. *J. Pharmacol. Exp. Ther.* **2012**, *340*, 404–421. [\[CrossRef\]](#) [\[PubMed\]](#)
299. Pinhal-Enfield, G.; Ramanathan, M.; Hasko, G.; Vogel, S.N.; Salzman, A.L.; Boons, G.-J.; Leibovich, S.J. An Angiogenic Switch in Macrophages Involving Synergy between Toll-Like Receptors 2, 4, 7, and 9 and Adenosine A<sub>2A</sub> Receptors. *Am. J. Pathol.* **2003**, *163*, 711–721. [\[CrossRef\]](#)
300. Jung, S.-M.; Peyton, L.; Essa, H.; Choi, D.-S. Adenosine receptors: Emerging non-opioids targets for pain medications. *Neurobiol. Pain* **2022**, *11*, 100087. [\[CrossRef\]](#)
301. Federico, S.; Lassiani, L.; Spalluto, G. Chemical Probes for the Adenosine Receptors. *Pharmaceuticals* **2019**, *12*, 168. [\[CrossRef\]](#)

302. Soave, M.; Briddon, S.J.; Hill, S.J.; Stoddart, L.A. Fluorescent ligands: Bringing light to emerging GPCR paradigms. *Br. J. Pharmacol.* **2020**, *177*, 978–991. [\[CrossRef\]](#)
303. Kozma, E.; Jayasekara, P.S.; Squarcialupi, L.; Paoletta, S.; Moro, S.; Federico, S.; Spalluto, G.; Jacobson, K.A. Fluorescent ligands for adenosine receptors. *Bioorg. Med. Chem. Lett.* **2013**, *23*, 26–36. [\[CrossRef\]](#)
304. McCabe, R.T.; Skolnick, P.; Jacobson, K.A. 2-2-4-2-2-1,3-Dihydro- 1,1-bis (4-hydroxyphenyl)-3-oxo-5-isobenzofuranthioureidy lethylaminocarbonylethylphenyl ethylamino-5'-N-ethylcarboxamidoadenosine (FITC-APEC): A Fluorescent Ligand for A<sub>2a</sub>-Adenosine Receptors. *J. Fluoresc.* **1992**, *2*, 217–223. [\[CrossRef\]](#)
305. Brand, F.; Klutz, A.M.; Jacobson, K.A.; Fredholm, B.B.; Schulte, G. Adenosine A<sub>2A</sub> receptor dynamics studied with the novel fluorescent agonist Alexa488-APEC. *Eur. J. Pharmacol.* **2008**, *590*, 36–42. [\[CrossRef\]](#)
306. Comeo, E.; Kindon, N.D.; Soave, M.; Stoddart, L.A.; Kilpatrick, L.E.; Scammells, P.J.; Hill, S.J.; Kellam, B. Subtype-Selective Fluorescent Ligands as Pharmacological Research Tools for the Human Adenosine A<sub>2A</sub> Receptor. *J. Med. Chem.* **2020**, *63*, 2656–2672. [\[CrossRef\]](#)
307. Toti, K.S.; Campbell, R.G.; Lee, H.; Salmaso, V.; Suresh, R.R.; Gao, Z.-G.; Jacobson, K.A. Fluorescent A<sub>2A</sub> and A<sub>3</sub> adenosine receptor antagonists as flow cytometry probes. *Purinergic Signal.* **2022**. [\[CrossRef\]](#) [\[PubMed\]](#)
308. McNeely, P.M.; Naranjo, A.N.; Forsten-Williams, K.; Robinson, A.S. A<sub>2A</sub>R Binding Kinetics in the Ligand Depletion Regime. *SLAS Discov.* **2017**, *22*, 166–175. [\[CrossRef\]](#) [\[PubMed\]](#)
309. Rieger, J.M.; Brown, M.L.; Sullivan, G.W.; Linden, J.; Macdonald, T.L. Design, synthesis, and evaluation of novel A<sub>2A</sub> adenosine receptor agonists. *J. Med. Chem.* **2001**, *44*, 531–539. [\[CrossRef\]](#)
310. Chen, J.-B.; Liu, E.M.; Chern, T.-R.; Yang, C.-W.; Lin, C.-I.; Huang, N.-K.; Lin, Y.-L.; Chern, Y.; Lin, J.-H.; Fang, J.-M. Design and synthesis of novel dual-action compounds targeting the adenosine A<sub>2A</sub> receptor and adenosine transporter for neuroprotection. *ChemMedChem* **2011**, *6*, 1390–1400. [\[CrossRef\]](#) [\[PubMed\]](#)
311. Yuan, G.; Gedeon, N.G.; Jankins, T.C.; Jones, G.B. Novel approaches for targeting the adenosine A<sub>2A</sub> receptor. *Expert Opin. Drug Discov.* **2015**, *10*, 63–80. [\[CrossRef\]](#) [\[PubMed\]](#)
312. Williams, R.; Hosila, F.J. The effect of different extraction sites upon incisor retraction. *Am. J. Orthod.* **1976**, *69*, 388–410. [\[CrossRef\]](#)
313. Jazayeri, A.; Andrews, S.P.; Marshall, F.H. Structurally Enabled Discovery of Adenosine A<sub>2A</sub> Receptor Antagonists. *Chem. Rev.* **2017**, *117*, 21–37. [\[CrossRef\]](#)
314. Hu, E.; Kunz, R.K.; Rumfelt, S.; Andrews, K.L.; Li, C.; Hitchcock, S.A.; Lindstrom, M.; Treanor, J. Use of structure based design to increase selectivity of pyridyl-cinnoline phosphodiesterase 10A (PDE10A) inhibitors against phosphodiesterase 3 (PDE3). *Bioorg. Med. Chem. Lett.* **2012**, *22*, 6938–6942. [\[CrossRef\]](#)
315. Deganutti, G.; Moro, S. Supporting the Identification of Novel Fragment-Based Positive Allosteric Modulators Using a Supervised Molecular Dynamics Approach: A Retrospective Analysis Considering the Human A<sub>2A</sub> Adenosine Receptor as a Key Example. *Molecules* **2017**, *22*, 818. [\[CrossRef\]](#) [\[PubMed\]](#)
316. McGraw, C.; Yang, L.; Levental, I.; Lyman, E.; Robinson, A.S. Membrane cholesterol depletion reduces downstream signaling activity of the adenosine A<sub>2A</sub> receptor. *Biochim. Biophys. Acta Biomembr.* **2019**, *1861*, 760–767. [\[CrossRef\]](#) [\[PubMed\]](#)
317. Yuan, S.; Hu, Z.; Filipek, S.; Vogel, H. W246(6.48) opens a gate for a continuous intrinsic water pathway during activation of the adenosine A<sub>2A</sub> receptor. *Angew. Chem. Int. Ed Engl.* **2015**, *54*, 556–559. [\[CrossRef\]](#) [\[PubMed\]](#)
318. Guo, D.; Pan, A.C.; Dror, R.O.; Mocking, T.; Liu, R.; Heitman, L.H.; Shaw, D.E.; IJzerman, A.P. Molecular Basis of Ligand Dissociation from the Adenosine A<sub>2A</sub> Receptor. *Mol. Pharmacol.* **2016**, *89*, 485–491. [\[CrossRef\]](#) [\[PubMed\]](#)
319. Mondal, C.; Halder, A.K.; Adhikari, N.; Jha, T. Structural findings of cinnolines as anti-schizophrenic PDE10A inhibitors through comparative chemometric modeling. *Mol. Divers.* **2014**, *18*, 655–671. [\[CrossRef\]](#)
320. Rodríguez, D.; Gao, Z.-G.; Moss, S.M.; Jacobson, K.A.; Carlsson, J. Molecular docking screening using agonist-bound GPCR structures: Probing the A<sub>2A</sub> adenosine receptor. *J. Chem. Inf. Model.* **2015**, *55*, 550–563. [\[CrossRef\]](#)
321. Novikov, G.V.; Sivozhelezov, V.S.; Shaïtan, K.V. Investigation of the conformational dynamics of the adenosine A<sub>2A</sub> receptor by means of molecular dynamics simulation. *Biofizika* **2013**, *58*, 618–634.
322. Kalash, L.; Winfield, I.; Safitri, D.; Bermudez, M.; Carvalho, S.; Glen, R.; Ladds, G.; Bender, A. Structure-based identification of dual ligands at the A<sub>2A</sub>R and PDE10A with anti-proliferative effects in lung cancer cell-lines. *J. Cheminform.* **2021**, *13*, 17. [\[CrossRef\]](#) [\[PubMed\]](#)
323. Ng, H.W.; Laughton, C.A.; Doughty, S.W. Molecular dynamics simulations of the adenosine A<sub>2a</sub> receptor: Structural stability, sampling, and convergence. *J. Chem. Inf. Model.* **2013**, *53*, 1168–1178. [\[CrossRef\]](#) [\[PubMed\]](#)
324. Hajduk, P.J.; Greer, J. A decade of fragment-based drug design: Strategic advances and lessons learned. *Nat. Rev. Drug Discov.* **2007**, *6*, 211–219. [\[CrossRef\]](#) [\[PubMed\]](#)
325. Chen, D.; Ranganathan, A.; IJzerman, A.P.; Siegal, G.; Carlsson, J. Complementarity between in silico and biophysical screening approaches in fragment-based lead discovery against the A<sub>2A</sub> adenosine receptor. *J. Chem. Inf. Model.* **2013**, *53*, 2701–2714. [\[CrossRef\]](#)
326. Chen, D.; Errey, J.C.; Heitman, L.H.; Marshall, F.H.; IJzerman, A.P.; Siegal, G. Fragment screening of GPCRs using biophysical methods: Identification of ligands of the adenosine A<sub>2A</sub> receptor with novel biological activity. *ACS Chem. Biol.* **2012**, *7*, 2064–2073. [\[CrossRef\]](#)

327. Matricon, P.; Ranganathan, A.; Warnick, E.; Gao, Z.-G.; Rudling, A.; Lambertucci, C.; Marucci, G.; Ezzati, A.; Jaiteh, M.; Dal Ben, D.; et al. Fragment optimization for GPCRs by molecular dynamics free energy calculations: Probing druggable sub-pockets of the A<sub>2A</sub> adenosine receptor binding site. *Sci. Rep.* **2017**, *7*, 6398. [[CrossRef](#)]
328. Tosh, D.K.; Phan, K.; Gao, Z.-G.; Gakh, A.A.; Xu, F.; Deflorian, F.; Abagyan, R.; Stevens, R.C.; Jacobson, K.A.; Katritch, V. Optimization of adenosine 5'-carboxamide derivatives as adenosine receptor agonists using structure-based ligand design and fragment screening. *J. Med. Chem.* **2012**, *55*, 4297–4308. [[CrossRef](#)]
329. Zhang, M.; Fan, S.; Zhou, X.; Xie, F.; Li, S.; Zhong, W. Design, synthesis and biological evaluation of 2-hydrazinyladenosine derivatives as A<sub>2A</sub> adenosine receptor ligands. *Eur. J. Med. Chem.* **2019**, *179*, 310–324. [[CrossRef](#)]

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