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Gemellar Competition as a Key Component in Seed–Seedling Transition of *Handroanthus chrysotrichus* (Mart. ex A. DC.) Mattos (Bignoniaceae)

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Abstract: The occurrence of more than one embryo per seed (polyembryony) is common among angiosperms; however, there are gaps in the knowledge of its effects on the early stages of plant development. In this context, we study the effects of polyembryony and intraspecific variability in gemellar competition during the seed-seedling transition in Neotropical Handroanthus chrysotricus (Bignoniaceae). We used seeds from five cultivated trees in an urban environment inserted in a biodiversity hotspot (Cerrado). Embryo mass, seed germination, seedling emergence and seedling morphometry were evaluated. We did not find intraspecific variability in seed germination, seedling emergence or the mean number of embryos and seedlings per seed. On the other hand, intraspecific variability was observed during the transition from embryo to seedling. When only one seedling emerged from a seed, the seed-seedling transition was more asynchronous than when more seedlings emerged from one seed (with higher uncertainty and a longer time to emergence of the last seedling). The mass of embryos and seedlings decreased with the increase in the number of embryos in a seed, reinforcing the occurrence of gemellar competition. However, the total mass of embryos per seed was similar. The increase in seedlings per seed also decreased the morphometric measurements of each one. A positive morphometric aspect of the emergence of two seedlings per seed was that they had the highest total seedling mass, evidencing the positive Allee effect. Polyembryony had both positive and negative effects on seed germination and seedling morphology in the species, which helps to understand how this phenomenon acts on seed biology and plant establishment.

Keywords: apomixis; intraspecific competition; intraspecific variability; polyembryony; seedling emergence; seed germination

1. Introduction

The seed–seedling transition is a central point during the plant's life cycle. Taking this into account, this transition process has been studied from different points of view in areas such as general botany, agronomy and physiology [1]. Nevertheless, there are still many gaps in the knowledge that need to be further studied, and polyembryony is one of the most intriguing areas of research. The presence of more than one embryo per seed, called polyembryony [2], is a common feature in angiosperms and it seems to be present in approximately 30% of the species studied so far. It has been associated with sporophytic apomixis (unreduced embryos originated from maternal tissues) or embryo cleavage (either



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Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). apomictic or, more rarely, sexual embryos) [3]. However, only a few species have been studied in detail regarding these embryos' origins or the ecological consequences of this reproductive phenomenon, with most studies focusing on the effects of polyembryony on the competition for nutrients and space between embryos and seedlings originated from the same seed [4–7]. This is a type of intraspecific competition [8] but with some particularities and important effects on the species' biology.

The competition between embryos and/or seedlings from a polyembryonic seed will be treated in this paper as gemellar competition due to particularities such as: (i) the somatic and possibly clonal origin of the embryos inside a seed; and (ii) the limited space inside a seed, which leads to a reduction in the size of the embryos and seedlings as the number of embryos per seed increases [4–7]. Additional aspects such as morphological anomalies, differences in survival rates in embryos and seedlings, and competition among sexual and asexual embryos [6,7,9]—with a posterior reduction in the competition effects on the seedlings in the long term [4]—have also been reported as particularities of this type of competition.

Some studies have pointed out that polyembryony could promote positive effects. One of them is the Allee effect, described as the mutual collaboration of seedlings while occupying patches [10]. Alternatively, polyembryony would also promote the bet-hedging effect, since the reduction in size and possible decrease in the fitness of polyembryonic seedlings would be compensated by the possibility that at least one of them would survive during germination in stressful conditions [5,10,11]. The positive effect of the increased chance of survival of at least one gemellar seedling per seed would result in greater individual seed survival [7]. However, the effect of gemellar competition is absent from some studies, hindering the creation of models for greater generalizations or future hypotheses on germination and plant development from polyembryonic seeds.

Handroanthus chrysotrichus (Mart. ex DC.) Mattos is a polyploid, self-compatible, apomictic, and polyembryonic tree and is one of the twenty-seven known Handroanthus species of Bignoniaceae [7,12–14]. The species is amply used as ornamental trees—known as "golden trumpet trees"—in urban environments due to their magnificent flowers [7,12–14]. Since it produces a lot of fruits and viable seeds, it seems to be a good model for studies on polyembryony. Like other native species used for ornamentation, its distribution is not clearly known given the low sampling in cities; however, polyembryony screenings are available. All plants studied so far, either natural or cultivated, are polyembryonic [15], which could save time in population selection. Their seeds also show germinability ranging between 25 and 87%, with intraspecific variability [16–20]. This species produces up to seven embryos and five seedlings per seed [7] making it easy to compare seeds with different numbers of embryos and seedlings. Previous data have shown that the higher the number of embryos per seed, the lower the mass of each embryo and seedling and of seedling length, and the higher the mortality of individual seedlings and the frequency of atypical embryos [7]. In addition, the mean seedling number emerged from one seed is also affected by the temperature during the emergence process [21]. Despite this basic knowledge, various previous studies ignored the effect of polyembryony on seedling success during the seed–seedling transition [22].

Taking this into account, it is clear that an increase in the number of gemellar embryos can affect the seed–seedling transition; however, it is not clear how and why this occurs. Therefore, we build the hypothesis that intraspecific gemellar competition affects the seed–seedling transition by affecting the dynamics, uniformity and synchrony of seed germination and of the seedling emergence processes. As a consequence, the morphometry of seedlings may be negatively affected, and early plant establishment hindered. In this context, our expectation is to provide data that will contribute to further the understanding of the impact of polyembryony on the seed biology and early plant development of *Handroanthus chrysotrichus*.

2. Materials and Methods

2.1. Sampling and Study Site

Handroanthus chrysotrichus is a Brazilian tree that occurs naturally in the "Mata Atlântica" and disturbed forest areas, but which is also described in the Cerrado Biome [23], Brazil's Neotropical Savanna [24]. It is also widely cultivated for ornamental use in urban areas [23]. Its origin, natural ecology, invasive and native actual distribution are unknown.

Seeds of five mother plants were collected in 2005, in an urban area of Uberlândia, Minas Gerais State, Brazil (18°53′6.64″ S, 48°15′36.23″ W). The climate region is the Aw Megathermic type according to the updated classification of Köppen–Geiger [25], a tropical wet climate with a dry winter (April to September) and a wet summer (October to March) (see climate diagram in Ranal [26]). The sample was deposited in the Herbarium Uberlandense (HUFU 52589). The origin and cultivation history of these mother plants are unknown. The data related to the embryo mass, number of embryos and seedlings formed per seed were reanalyzed from the original data obtained by Mendes-Rodrigues et al. [7].

2.2. Seed Water Content and Mass

Eight replicates per mother plant were used to evaluate the water content of the seeds. Each replicate had approximately 500 mg of seeds (mean = 490.38, standard deviation = 29.46, n = 40). The number of seeds per replicate and per mother plant was 32 for mother plants four and five, 38 for mother plants one and two and 44 for mother plant three. The water content was obtained after keeping the seeds in a stove at 55 °C for three days, and then at 105 °C for 10 days. Before weighing, the seeds were kept in plastic boxes with dry silica gel until they returned to room temperature. The replicates were weighed individually on an analytical scale (AA250, precision 0.1 mg, Denver Instrument, Bohemia, NY, USA), before and after drying. The water content was calculated on a wet weight basis. These replicas were also used to estimate the fresh and dry mass of the seeds.

2.3. Seed Germination and Seedling Emergence Assays

The germination process was recorded from seeds sown on vermiculite (expansion volume of 0.1 m³) moistened with distilled water at field capacity, and kept in germination plastic boxes in a germination chamber (MPG-2000, Seedburo Equipment Company, Batavia, USA) under continuous fluorescent light and a mean minimum temperature of 24.53 ± 0.24 °C (mean \pm standard deviation; n = 17) and a mean maximum temperature of 25.33 ± 0.20 °C (mean \pm standard deviation; n = 17). We used this temperature as it offered the best results for most Cerrado species [6,7,9]. The seed was considered germinated when the first radicle protruded from the seed, since the species is polyembryonic (Figure 1A,B).

For seedling emergence, seeds were sown in polystyrene multicell boards (cell volume of 34 cm³) with one seed per cell at ca. 0.5 cm depth, in a 1:1 mixture of commercial substrate (Plantmax[®], Eucatex Mineral Ltda, São Paulo, SP, Brazil) and vermiculite (expansion volume of 0.1 m³). The trays were kept in an open area about 100 m away from the mother plants, under natural light conditions, and moistened at field capacity. The appearance of any part of the first seedling above the substrate's surface was used as emergence criterion, since the species is polyembryonic (Figure 1B,C).

Both seed germination and seedling emergence assays were conducted with small samples (n = 32 seeds per replicate). The number of seeds germinated and seedlings emerged was recorded daily, at the same hour. No experiment was censored (break off). Observations continued until all seeds germinated or died. For each replicate, germinability or percentage of emergence, mean germination or emergence time, time to first and last seed germination or seedling emergence, coefficient of variation of germination or emergence time, uncertainty and synchrony were calculated according to Ranal and Santana [27]. Further details of these germination measurements can be found in Ranal and Santana [27], and Ranal et al. [28]. The relative frequency of seed germination and seedling emergence was also calculated, grouping all 20 replicates in one sample based on the absence of intraspecific variability in seed-germination measurements [29].

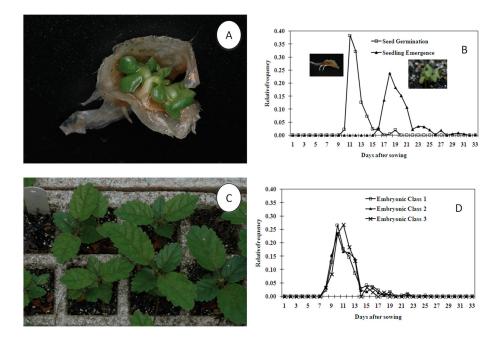


Figure 1. Seed germination and seedling emergence of the polyembryonic *Handroanthus chrysotrichus* (Bignoniaceae) cultivated in an urban area of Uberlândia, Minas Gerais State, Brazil. (**A**) Seed showing the presence of polyembryony, (**B**) relative frequency of seed germination and seedling emergence, (**C**) gemellar seedlings after 54 days of sowing, (**D**) relative frequency of seedling emergence of the embryonic classes one to three.

2.4. Polyembryony Indicators

Each seed germinated was dissected to count the number of embryos, as some could be hidden or did not develop completely inside the seeds. These data were used to calculate the polyembryonic seed percentage (PSP), that is, the percentage of seeds with more than one embryo, and the mean embryo number per seed (MENS). We sampled 124, 118, 120, 125, and 119 seeds for mother plants one to five, respectively. We evaluated only the germinated seeds.

To estimate the total number of seedlings emerged per seed, the observations of seedling emergence were conducted until the number of seedlings effectively emerged per each seed had stabilized. That is needed since seedlings from a polyembryonic seed have different times of emergence. In general, stabilization occurs about seven days after the emergence of the first seedling. During the first seven days, we did not observe any seedling mortality. Seedling mortality was also evaluated during the experiment (see results in Mendes-Rodrigues et al. [7]). We conducted these observations on a daily basis because seedling mortality during the experiment could cause an underestimation of the analysis of the effective number of seedlings produced per seed (see Mendes-Rodrigues et al. [7]). From these data, we calculated the gemellar seed percentage (GSP)—i.e., the percentage of seeds with more than one seedling emerged—and the mean seedling number per seed (MSNS). We sampled 117, 116, 91, 118, and 121 seeds for mother plants one to five, respectively. We evaluated only the seeds with emerged seedlings.

Additionally, we estimated the percentage of embryos that were converted into seedlings, treated here as Embryo Seedling Turnover Percentage (ESTP) [7]. For this, we used the following mathematical expression: ESTP = $[(MSNS \times NSES)/(MENS \times NSS)] \times 100$, where MSNS: mean seedling number per seed, NSES: number of seeds with emerged seedlings, MENS: mean embryo number per seed, and NSS: number of seeds sowed. The ESTP was calculated for each mother plant and species (independently of the mother plant).

We also classified the seeds in seed embryonic classes (SEC) and seedling gemellar classes (SGC). We found from one to seven SEC, containing from one to seven embryos per

seed (SEC1 to SEC7), and found from one to five SGC containing from one to five seedlings per seed (SGC1 to SGC5). The SGCs are named differently in the literature, as singlet, doublet, triplet, quadruplet, and quintuplet, and respectively refer to SGC1 to SGC5.

After the classification of the seeds from the emergence experiment in the SGCs, we calculated the seed-emergence measurements for the seedling gemellar classes singlet, doublet and triplet (except the percentage of emergence). In this analysis, due to the low frequency of each SGC in the original replicates from the seed-emergence experiment, each mother plant was considered to be one replicate for each SGC. In this case, all seeds of each SGC of the four replicates of each mother plant were grouped. SGC4 and SGC5 did not present enough replicates for the calculations of seed-emergence measurements.

2.5. Gemellar Competition

We measured seedling mass just after germination (after the first emission of radicle— Figure 1A) as a direct estimate of embryo mass, because it was practically impossible to isolate the multiple embryos present in a non-imbibed seed without fragmenting them and causing their loss of individuality. Germinated seeds were dissected, and each embryo (now a young seedling) was weighed individually on an analytical scale (AA250, precision 0.1 mg, Denver Instrument, Bohemia, NY, USA). Sample size of each SEC was SEC1 = 50, SEC2 = 106, SEC3 = 183, SEC4 = 132, SEC5 = 65, SEC6 = 12, and SEC7 = 21. The sum of the mass of all embryos of each seed was also calculated, and the sample size of each SEC was SEC1 = 50, SEC2 = 53, SEC3 = 61, SEC4 = 33, SEC5 = 13, SEC6 = 2, and SEC7 = 3. These data were originally presented by Mendes-Rodrigues et al. [7].

After 54 days of sowing, one sample of seedlings of each SGC was exhumed and measured. We adopted 54 days after sowing as a time marker since we did not correct growth with the time of emergence of each seedling. Moreover, after 54 days, emergence does not occur anymore and the seedling stand is homogenous. We sampled only seedlings of SGC with no mortality in a given seed up to 54 days. The survival percentages were presented by Mendes-Rodrigues et al. [7]. At this time (54 days after sowing, Figure 1C) the seedling size permitted hand manipulation without apparent damages. After 54 days of sowing the probability of mortality of the seedlings increased (personal observation). We measured 25 singlets, 20 doublets, 15 triplets, and 16 quadruplets. The gemellar class quintuplet was not included because only one seed from this class appeared in the sample. The analyses of seedling morphometry were not performed separately for each mother plant due to the low frequency of some SGCs at this time. The quadruplets were also not present in all mother plants analyzed.

For each seedling, we recorded the number of leaflets (NL), stem height (SH, from the collar to the apical meristem; the collar is characterized by a separation between the chlorophyllous hypocotyl and the achlorophyllous root) and stem diameter (SD, or diameter of collar). We also evaluated the aerial seedling dry mass (ASDM, or aerial mass part) above the collar, and the subterranean seedling dry mass (SSDM, or subterranean mass part) below the collar. The evaluations were performed using a digital scale (AA250, precision 0.1 mg, Denver Instrument, Bohemia, NY, USA). The seedlings were kept in paper bags at 70 °C until they reached constant dry mass (after 7 days), and 2 to 4 h afterwards in hermetic boxes with dry silica gel until they reached room temperature, before mass evaluations. We calculated the ratio between aerial seedling dry mass and the total seedling dry mass (aerial/seedling dry mass ratio, ASDMR). This ratio was used to evaluate seedling resource allocation. We also calculated the biomass production of each seed individually, treated here as the sum of the biomasses of all seedlings produced per seed, as proposed by Blanchard et al. [5], in order to evaluate the density-dependent facilitation of polyembryonic seedlings, or Allee effects.

2.6. Mass Prediction Model for Seedlings

We applied multiple linear regression models based on the least square method to know if the seedlings' morphometric measurements could predict the aerial and subterranean dry mass of 54-day-old seedlings. This model was proposed to estimate the seedling mass in a seedling competition experiments. We adopted the following traits as independent variables: NL, SH, SD, and density (here considered as the number of seedlings in the SGC) in a model without a constant (or intercept). We sampled these variables in three SGCs (SGC1 = 25, SGC2 = 20, SGC3 = 15) with a total of n = 60 seedlings. SGC 4 and 5 were not included due to their low occurrence in the studied sample. SGC4 also showed high seedling mortality at 54 days. The aerial and subterranean dry mass were considered to be the dependent variables.

2.7. Statistical Analysis

For the germination and emergence experiments, the assumptions of ANOVA were tested. We tested residuals normality using the Kolmogorov–Smirnov Lilliefors test and the homoscedasticity of variances using Levene's test. When both assumptions were accepted, the data were compared using ANOVA and Tukey's test. When at least one of the assumptions was not met, the data were compared using the Kruskall–Wallis and Dunn's test. Data on seed mass and water content were tested with Generalized Linear Models. For seed mass and water content, we adopted the Gaussian and Gamma distribution function, respectively, and the identity link function for both. The Least Significance Difference test was used for multiple comparisons.

Polyembryonic indicators were tested with Generalized Linear Models. For the number of embryos per seed and number of seedlings per seed, we adopted the Poisson distribution function and the logarithmic link function. For the polyembryonic seed percentage, gemellar seed percentage, and embryo seedling turnover percentage, we used the Binomial distribution and identity link functions. Again, the Least Significance Difference test was used for multiple comparisons.

Initially, to test the effect of the mother plant on seedling morphometry, we performed a two-way ANOVA. For the dry-mass analysis of 54-day-old seedlings, the first factor was the mother plant and the second factor was the seedling gemellar class (singlet, doublet and triplet). In this analysis, the mother plants did not show differences ($F_{4,59} = 0.91$, p = 0.4643), the seedling gemellar classes showed differences ($F_{2,59} = 50.87$, p < 0.0001) and the interaction between these two factors was not significant ($F_{8,59} = 1.28$, p = 0.2791). After this, we did not test the effect of the mother plant in the other tested models. In addition, we found low frequency of some gemellar classes in some mother plants, which hindered their use in some of the models.

Embryo-mass data were adjusted to an exponential regression model, and the total mass data of embryos per seed were adjusted to a linear regression model by the ordinary least square method. The model significance was tested using a one-way ANOVA. The effects of initial gemellar competition in seedling morphometry were compared between embryonic classes with a one-way ANOVA. For this, the number of seedlings per seed was used as an independent factor and the seedling traits as a dependent factor. These data were adjusted to linear regression models for all traits, except for the total seedling mass per seed, which was better adjusted to a quadratic regression model. With this, the effect of polyembryony or gemellar competition was inferred by the inclination (or slope) of the linear regression model (see details in Mendes-Rodrigues et al. [6]) whenever a linear regression was used. In these analyses, we included the embryonic classes from one to four seedlings per seed. The number of replicates for each embryonic class were SEC1 = 25, SEC 2 = 20, SEC 3 = 15, and SEC 4 = 16, except for seedling dry mass per seed (SEC1 = 25, SEC2 = 10, SEC3 = 5, and SEC4 = 4).

The density or seed gemellar class was considered a Dummy variable in the mass prediction model (a discrete quantitative variable that represents binary data and can take on only two values, 1 or zero, where 1 represents the presence of a qualitative attribute and zero the absence), since density is an integer. To adjust the mass prediction model, the presence of multicollinearity between the independent variables was tested by the VIF (variation inflation factor), and variables were excluded from the models if VIF > 4.

To evaluate the contribution of each variable to the model, the Partial *F* test was used with a significance of 0.10. The significance of the parameter estimate was tested with Student's *t*-test. The residuals of the regression model were tested for normality by the Shapiro–Wilk test. The homogeneity of variances was tested with Bartlett's test using density as a factor of formation of the groups. The independence of the residuals was tested by the Durbin–Watson test. The tested models were evaluated without the intercept since the presence of the intercept has no biological justification in this case. Multicollinearity was observed among the predictor variables for seedling mass. The VIF value for the stem diameter was higher than four (VIF = 5.54), and the variable was performed again and no VIF value above four was obtained (stem height VIF = 1.44, seedling density VIF = 1.59 and number of leaflets VIF = 1.23).

For the analyses where significance was not declared, 0.05 was adopted. The analyses were performed in the Statistical Package SPSS 20.0 or in the R environment [30].

3. Results

3.1. Seed Water Content and Mass

Mother plants show intraspecific variability in the physical traits of seeds, specifically seed mass and water content. Mother plants one, two and three exhibited lower values for seed mass and water content (Table 1). The seeds' fresh mass varied from 10.4 to 15.66 mg per seed, and the seeds' dry mass varied from 9.63 to 13.19 mg per seed. The seeds showed low water content (mean = 12.1%, SE = 0.89) varying from 7.47 to 20.16%, which indicates an anhydrobiotic pattern (i.e., orthodox seeds; Table 1).

Table 1. Seed water content and mass of *Handroanthus chrysotrichus* (Bignoniaceae). Seeds collected from five polyembryonic mother plants in an urban area of Uberlândia, Minas Gerais State, Brazil, in 2005.

		Trait	
	Seed Wet Mass (mg)	Seed Dry Mass (mg)	Water Content at 105 °C (%)
Plant	Mean \pm SE 1	Mean \pm SE 1	Mean \pm SE ¹
1	$13.14\pm0.06~^{\rm b}$	12.11 ± 0.06 ^b	$7.87\pm0.13~^{\rm d}$
2	13.10 ± 0.14 ^b	$11.88\pm0.11~^{\rm b}$	9.28 ± 0.30 ^c
3	$10.40\pm0.31~^{\rm c}$	9.63 ± 0.32 $^{ m c}$	7.47 ± 0.59 d
4	15.66 ± 0.36 ^a	13.19 ± 0.30 ^a	15.72 ± 0.6 ^b
5	15.50 ± 0.24 $^{\rm a}$	$12.36\pm0.25~^{\mathrm{b}}$	$20.16\pm1.85~^{\rm a}$
All	13.56 ± 0.33	11.83 ± 0.21	12.1 ± 0.89
п	40	40	40
X ²	345.36 **	148.79 **	183.62 **

SE: standard error, X^2 : qui-square statistic based on Generalized Linear Model, ** p < 0.01. ¹ Mean values followed by different letters in the column are different based on the LSD test (p < 0.05).

3.2. Seed Germination and Seedling Emergence Assays

Although mother plants were important for the physical traits of the seed samples, they made no contribution to the intraspecific variability in the functional traits of the seed–seedling transition of Handroanthus chrysotrichus (Table 2). In general, seeds had a high viability (high germinability 97.34% and high percentage of emergence 90.16%) with germination and emergence peaks below 20 days (Table 2). Seed germination was synchronous (Figure 1B) and uniform (coefficient of variation of time less than 15%; Table 2). When emergence measurements were evaluated as a function of the seed's gemellar class (SGC, number of seedlings emerged per seed) regardless of the mother plant sampled, differences were observed in the uncertainty and emergence time of the last seed. The seeds of SGC1 had a longer emergence process (emergence time of the last seed = 29.40 days),

and greater uncertainty (uncertainty = 2.97 bits), when compared to the seeds of SGC3 (emergence time of the last seed = 23 days; uncertainty = 2.27 bits) (Table 3). Despite this, the differences in the relative frequency of the emergence process were slim (Figure 1D).

Table 2. Seed germination and seedling-emergence measurements (mean \pm SE: standard error) of *Handroanthus chrysotrichus* (Bignoniaceae). Seeds collected from five polyembryonic mother plants in an urban area of Uberlândia, Minas Gerais State, Brazil, in 2005.

	Germinati	on (<i>n</i> = 20)	Emergence ($n = 20$)		
Measurement (Unit)	$\mathbf{Mean} \pm \mathbf{SE}$	Statistic	$\mathbf{Mean} \pm \mathbf{SE}$	Statistic	
Percentage (%)	97.34 ± 0.73	F = 1.85 ns	90.16 ± 2.51	F = 2.98 ns	
Mean time (day)	12.04 ± 0.12	F = 1.49 ns	19.68 ± 0.19	F = 0.61 ns	
Uncertainty (bits)	1.85 ± 0.08	F = 1.79 ns	2.69 ± 0.06	F = 0.82 ns	
Synchrony index	0.32 ± 0.02	F = 1.88 ns	0.16 ± 0.01	F = 0.41 ns	
Coefficient of variation of time (%)	9.36 ± 0.62	F = 1.78 ns	13.03 ± 0.69	F = 0.19 ns	
Time of first seed (day)	10.25 ± 0.20	$X^2 = 5.09$ ns	16.75 ± 0.18	$X^2 = 5.59$ ns	
Time of last seed (day)	15.50 ± 0.28	F = 1.83 ns	27.65 ± 0.78	$X^2 = 1.82$ ns	

F: statistic *F* of ANOVA comparing five mother plants, X^2 : qui-square (X^2) statistic based on the Kruskal–Wallis test, ^{ns}: (p > 0.05).

Table 3. Seed-emergence measurements (mean \pm SE: standard error, n = 5) for the first three seed gemellar classes of *Handroanthus chrysotrichus* (Bignoniaceae) in an urban area of Uberlândia, Minas Gerais State, Brazil, in 2005.

	Seed Gemellar Class (Number of Emerged Seedlings)					
Measurement (Unit)	One ¹	Two ¹	Three ¹	Statistic		
Mean emergence time (day)	19.73 ± 0.17	19.29 ± 0.23	19.30 ± 0.06	$X^2 = 3.40$ ns $X^2 = 9.26$ **		
Uncertainty (bits) Synchrony index	2.97 ± 0.03 ^a 0.15 ± 0.00	$2.71 \pm 0.08 \; ^{ m ab} \ 0.16 \pm 0.01$	$2.27 \pm 0.20 \ ^{ m b}$ 0.15 ± 0.05	$X^2 = 9.26^{nx}$ $X^2 = 1.86^{ns}$		
Coefficient of variation of time (%)	13.95 ± 1.09	10.60 ± 1.11	9.36 ± 1.56	F = 3.45 ns		
Time of first seed (day) Time of last seed (day)	$\begin{array}{c} 16.20 \pm 0.20 \\ 29.40 \pm 1.12 \ ^{a} \end{array}$	$\begin{array}{c} 16.20 \pm 0.20 \\ 25.40 \pm 1.29 \ ^{ab} \end{array}$	$\begin{array}{c} 17.00 \pm 0.45 \\ 23.00 \pm 0.95 \ ^{\text{b}} \end{array}$	$X^2 = 3.50 \text{ ns}$ F = 8.21 **		

F: statistic *F* of ANOVA, *X*²: qui-square (*X*²) statistic based on the Kruskal–Wallis test, ^{ns} (p > 0.05), **: p < 0.01. ¹ Mean values followed by different letters in the line are different based on Dunn's or Tukey's test (pairwise comparisons).

3.3. Polyembryony Indicators

Additionally, all mother plants produced polyembryonic seeds and gemellar seedlings. The polyembryonic seed percentage, the gemellar seed percentage and the embryo–seedling turnover percentage showed differences among the five mother plants (Table 4). Mother plants two, four and five performed better in the two former traits while mother plants one and two were more successful in the conversion of embryos into seedlings. On the contrary, the mother plants showed neither variability in the mean embryo number per seed (mean = 2.18 embryos per seed, SE = 0.05), nor in the mean seedling number per seed (mean = 1.66 seedlings per seed, SE = 0.03) (Table 4).

Independently of the mother plant sampled, the frequency of polyembryonic seeds for Handroanthus chrysotrichus was 68.32% (606 seeds). From these 606 seeds, 31.68% had one, 34.82% had two, 23.13% had three, 6.77% had four, 2.31% had five, 0.83% had six, and 0.50% had seven embryos per seed. The frequency of gemellar seeds was 52.04% (563 seeds). From these 563 seeds, 47.96% had one, 39.61% had two, 11.01% had three, 1.24% had four, and 0.18% had five seedlings per seed.

			Trait		
	MENS	PSP	MSNS	GSP	ESTP
Plant	Mean \pm SE 1	$\%\pm { m CIe_{95\%}}\ ^1$	Mean \pm SE 1	$\%\pm {\rm CIe_{95\%}}\ ^1$	$\%\pm {\rm CIe_{95\%}}\ ^1$
1	2.10 ± 0.10	$65.32 \pm 8.38 \ ^{\mathrm{bc}}$	1.59 ± 0.07	$47.01\pm9.04~^{\mathrm{bc}}$	$69.40\pm5.52~^{\mathrm{ab}}$
2	2.11 ± 0.10	$67.80\pm8.43~^{\mathrm{ab}}$	1.78 ± 0.07	$61.21\pm8.87~^{\rm a}$	$76.30\pm5.07~^{\rm a}$
3	1.79 ± 0.08	$55.00\pm8.90~^{\rm c}$	1.47 ± 0.08	$36.26\pm0.99~^{\rm c}$	$58.52\pm6.38~^{\rm c}$
4	2.44 ± 0.10	77.60 ± 7.31 $^{\rm a}$	1.78 ± 0.07	61.86 ± 8.76 $^{\rm a}$	67.31 ± 5.21 ^b
5	2.44 ± 0.11	$75.63\pm7.71~^{\rm ab}$	1.64 ± 0.07	$50.41\pm8.91~^{\rm ab}$	$63.78\pm5.33~^{\rm bc}$
All	2.18 ± 0.05	68.32 ± 3.70	1.66 ± 0.03	52.04 ± 4.13	67.22 ± 2.47
п	606	606	563	563	1391
X ²	5.34 ^{ns}	17.82 **	1.62 ^{ns}	18.44 **	20.06 **

Table 4. Polyembryony indicators of *Handroanthus chrysotrichus* (Bignoniaceae). Seeds collected from five polyembryonic mother plants in an urban area of Uberlândia, Minas Gerais State, Brazil, in 2005.

MENS: mean embryo number per seed, PSP: polyembryonic seed percentage, MSNS: mean seedling number per seed, GSP: gemellar seed percentage, ESTP: embryo seedling turnover percentage, SE: standard error, CIe95%: margin of error of the confidence interval at 95%, X^2 : qui-square statistic based on Generalized Linear Model, ** p < 0.01, ^{ns} p > 0.05. ¹ Mean values followed by different letters in columns are different based on the Least Significance Difference pairwise comparisons.

3.4. Gemellar Competition

The individual mass of embryos decreased with the increase in the number of embryos per seed, with significant adjustment to the exponential model (Figure 2A). The total mass of embryos per seed did not vary according to the SEC and did not fit the linear or other regression models (Figure 2B).

All morphometric variables evaluated in the seedlings 54 days after sowing (Figure 1D) showed significant adjustments to the linear regression model as a function of the number of seedlings per seed (Table 5 and Figure 2C). The estimates of the slope parameter were negative in all cases, showing that the increase in the number of seedlings per seed reduces the individual measurements of each seedling (Table 5, Figure 2C), that is, there is a negative effect of polyembryony on the morphometry of the seedlings. The only exception was the total mass of seedlings per seed that showed a better fit to the second-degree regression model, with doublet seedlings (SGC2) exhibiting the highest total mass of seedlings per seed (Figure 2D). In SGC2, the presence of two seedlings is beneficial, evidenced by the increase in the total mass of seedlings per seed (Figure 2D).

Table 5. Linear regression model applied to seedling morphometry of *Handroanthus chrysotrichus* (Bignoniaceae) as a function of the number of seedlings emerged per seed. The seedlings were cultivated under natural light, collected after 54 days of sowing in Uberlândia, Minas Gerais State, Brazil.

	Estimates				
Trait (Unit)	Intercept Inclination		<i>R</i> ²	F _{3,72}	
Stem height (mm)	39.77	-4.77	0.9085	39.01 **	
Stem diameter (mm)	2.19	-0.35	0.9494	85.77 **	
Number of leaflets	5.03	-0.42	0.9377	93.77 **	
Aerial seedling dry mass (mg)	121.07	-27.77	0.9427	77.38 **	
Subterranean seedling dry mass (mg)	57.29	-11.13	0.9685	25.02 **	
Aerial/seedling dry mass ratio	0.74	-0.05	0.8668	9.52 **	

 R^2 : coefficient of determination, *F*: statistic *F* of ANOVA, **: *p* < 0.001.

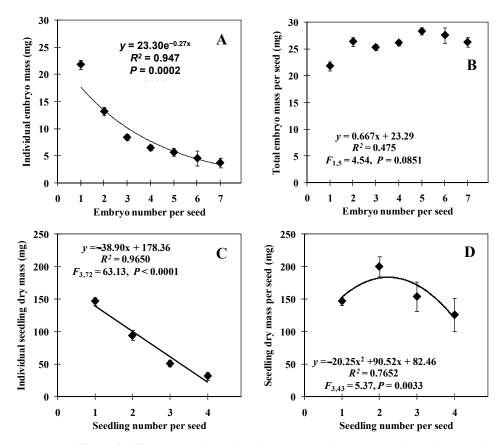


Figure 2. Effects of different numbers of embryos or seedlings per seed on the morphometry of embryos and seedlings of *Handroanthus chrysotrichus* (Bignoniaceae) cultivated under natural light, in Uberlândia, Minas Gerais State, Brazil, for 54 days. (A) Individual Embryo mass, (B) total embryo mass per seed, (C) individual seedling dry mass, (D) seedling dry mass per seed.

3.5. Mass Prediction Model for Seedlings

All the prediction models of the 54-day-old seedlings' dry mass using the morphometric data of the seedlings were adequate (Table 6). For the prediction of the aerial and subterranean dry mass, the model selected the stem height, number of leaflets and density. The stem height and number of leaflets showed positive estimates for the parameters, while the estimates for the parameters were negative for the variable density (Table 6). The dry mass of seedlings decreased as the density increased. Density three had the highest negative impact on the individual mass of seedlings. The models for aerial and subterranean dry mass showed an adequate fit, with coefficients of determination greater than 0.90 (Table 6). All the assumptions of the model were met (normality of residuals, homoscedasticity and independence).

Table 6. Multiple linear regression models for aerial and subterranean seedling dry mass of seedlings of *Handroanthus chrysotrichus* (Bignoniaceae), 54 days after sowing, based on morphometric variables of seedlings.

Aerial Seedling Dry Mass ($R^2 = 0.9508$)								
				$ ext{CI}_{95\%}$ to eta_i				
$\boldsymbol{\beta}_i$: Trait	β_i Estimative	t	LL	UL			
β_1	: Stem height (mm)	2.1066	3.53 **	0.91	3.30			
β_2	: Number of leaflets	7.4042	2.54 *	1.57	13.24			
β3	: Density 1	-4.8822	-0.21 ns	-52.35	42.59			
β_4	: Density 2	-34.9328	-1.19 §	-76.43	6.56			
β_5	: Density 3	-56.1621	-2.84 **	-95.75	-16.58			

1	8	7
1		1

Table 6. Cont.

Subterranean Seedling Dry Mass ($R^2 = 0.9068$)								
				$\text{CI}_{95\%}$ to β_i				
$\boldsymbol{\beta}_i$: Trait	β_i Estimative	t	LL	UL			
β_1	: Stem height (mm)	1.1180	2.62 *	0.26	1.97			
32	: Number of leaflets	3.6875	1.77 [§]	-0.48	7.86			
33	: Density 1	-9.4700	-0.55 ns	-43.39	24.45			
β_4	: Density 2	-10.4101	$-0.70^{\text{ ns}}$	-40.06	19.24			
β ₅	: Density 3	-24.8489	-1.76 §	-53.14	3.44			

 β_{t} : i-th parameter estimative, *t*: statistic based on Student's *t* test, Cl_{95%}: confidence interval to 95%, LL: lower limit of Cl_{95%}, UL: upper limit of Cl_{95%}, *R*²: coefficient of determination. *: *p* < 0.05, **: *p* < 0.01, ns: *p* > 0.05; §: 0.05 < *p* < 0.10. The density variable was considered as Dummy variable and refers to the number of seedlings per seed.

4. Discussion

We showed here that polyembryony affects the seed-seedling transition by altering seedling morphometry and possibly vigor. This occurs more as a consequence of high intraspecific competition (gemellar competition) from the number of embryos in each seed than from intraspecific variability in the mother plants. In general, gemellar competition from two-or-more embryos per seed causes distress in the seed-seedling transition. This behavior is explained by the fact that the size of the embryos and seedlings decreased with the increase in their number per seed, which resulted in a reduction in the individual seedlings' vigor, therefore affecting the emergence process. Only seeds that produced two seedlings per seed showed a higher total mass of seedlings per seed, evidencing a positive Allee effect.

In general, functional and physical traits are correlated in the transition from seed to seedling [31,32]. This is the most studied aspect of the early plant development [33]. Thus, we were surprised when we observed that physical traits had intraspecific variability although the mother plants made no differential contribution to the functional traits. This was the first clue to how polyembryonic seeds (the majority in all samples studied) can alter the early plant development of *H. chrysotrichus*. Taking this into account, we considered polyembryony could be a potential cause of distress for the species. To prove this hypothesis, we observed the way that intraspecific competition from gemellar status affects seed germination, seedling emergence and morphometry.

First, we observed that the high capacity, uniformity, and dynamics of the seedseedling transition follows a species-specific pattern during seed germination and seedling emergence. However, seedling emergence is the key aspect of the seed-seedling transition since it is less efficient and homogeneous, as pointed out by the results. In fact, it is expected that the seed germination process is less variable than the process of seedling emergence, growth, development and establishment [34–37]. However, when more than one embryo is contained in a seed, gemellar competition can alter this expected pattern. Seed germination is an amphibolic event in which enzymatic processes and physical forces overlap, promoting embryo growth until it finally breaks through the teguments, allowing the full embryo's protrusion [32,38-41]. Therefore, more than one embryo in a same seed can provoke a double and conflicting effect in the seed-seedling transition process, increasing physical force to overcome teguments, but reducing reserve material for the optimal development of each embryo. From a physiological point of view, this double effect can play a central role as a trade-off, impacting the time, velocity and uniformity of protrusion and, therefore, the dynamics of seedling emergence. Therefore, the presence of polyembryony can function as a distress agent (sense Kranner et al. [42]) in the early plant development based on the trade-off of seed germination and seedling emergence. Our findings showed this for *H. chrysotrichus*. The increase in the number of embryos in a seed reduces embryo mass and, consequently, stem height and leaf number in seedlings. Those measurements are related to seedling vigor and, thus, with the seedling's ability

to establish itself in a non-optimal environment (see Finch-Savage and Bassel [43]; Reed et al. [44]). However, total seedling mass was optimal when two embryos were contained in a seed. Three embryos per seed led to a reduction in individual seedling mass, making the emergence process less uncertain but rapid. These are characteristics which have been identified as an escape strategy during the seed–seedling transition in order to overcome stress [43,44].

On the other hand, the absence of intraspecific variability in the seed germination and seedling-emergence measurements of *H. chrysotrichus* may be related to the occurrence of mother plants in a restricted and homogeneous urban area. There are records of germinability of the species ranging between 20 and 87% in 17 individuals in an urban area in Jaboticabal city, São Paulo State, Brazil [20]. Another issue that cannot be overlooked is that the apomixis present in the species can generate low genetic variability or clonality, as observed in other Cerrado species [45]. Nevertheless, studies have demonstrated the presence of intraspecific variability in the germination measurements of apomictic species in Cerrado [46–48], and for vegetative and reproductive characters of sexual and apomictic populations of the congeneric *H. ochraceus* [49]. However, the number of individuals needed to characterize the variability in Cerrado Bignoniaceae can be very high, variable and dependent on the reproductive system, with the sample size ranging from 7 to 212 individuals analyzed [49–51]. In addition, even in apomictic populations with a low sampling, it is possible to detect intraspecific variability in some traits, as observed in our case.

Few studies have evaluated the type of germination measurements described here for other Cerrado Bignoniaceae [52,53] (Table 7), or even for other biomes. The seed germination behavior of *H. chrysotrichus* was similar to the one observed in *H. serratifolius*, another polyembryonic species [7], but distinct from that of *J. cuspidifolia*, which has low polyembryony (below 5%), a low germination percentage, and a higher coefficient of variation of germination time (Table 7).

Table 7. Literature data on seed germination and seedling-emergence measurements of Bignoniaceae with known embryony pattern from the Cerrado.

		Seed Germination						
Species	Embryony	G	TFS	MT	TLS	CVT	U	Ζ
Handroanthus chrysotrichus (Mart. ex DC.) Mattos	Polyembryony	99.38	10	12.18	19	13.33	2.25	0.2732
Jacaranda cuspidifolia Mart.	Low Polyembryony	22.25	5	10.79	34	49.70	3.44	0.1004
Handroanthu serratifolius (Vahl) S.O.Grose	Polyembryony	98.00	5	8.30	34	41.94	2.78	0.1780
		Seedling Emergence						
Species	Embryony	Е	TFS	MT	TLS	CVT	U	Ζ
Jacaranda cuspidifolia Mart.	Low Polyembryony	85.75	48	74.31	101	14.49	4.60	0.0572
Handroanthus chrysotrichus (Mart. ex DC.) Mattos	Polyembryony	90.16	16	19.68	33	13.95	3.10	0.1471
Handroanthus impetiginosus (Mart. ex DC.) Mattos	Monoembryony	80.47	10	12.71	22	23.78	2.91	0.1574
Handroanthu serratifolius (Vahl) S.O.Grose	Polyembryony	96.09	18	21.09	35	12.82	2.87	0.1554
Tabebuia aurea (Silva Manso) S.Moor	Monoembryony	97.22	9	13.63	25	24.26	3.21	0.1139
Tabebuia roseoalba (Ridl.) Sandwith	Low Polyembryony	96.00	8	9.49	19	20.09	2.13	0.2816

G: germination percentage (%); E: emergence percentage (%); MT: mean time (day); U: uncertainty (bits); Z: synchrony; CVT: coefficient of variation of time (%); TFS: time of first seed (day); TLS: time of last seed (day). Source: deed-germination measurements adapted from Mendes-Rodrigues et al. [53]; embryonic pattern from Mendes-Rodrigues [54]. Low polyembryony is defined as polyembryony below 5%.

As for seedling emergence, *H. chrysotrichus* showed marked differences from *Tabebuia roseoalba* (Ridl.) Sandwith, which experiences a highly synchronous germination in a short period of time, and from *J. cuspidifolia* Mart, which has a low synchrony, a long germination times and dormancy—when germination takes longer than 30 days [52,53] (Table 7). Both

species have low polyembryony. Despite scarce data, the general emergence and germination pattern of Bignoniaceae seems to be a short germination time, a synchronous process and low variability.

Species with a low polyembryonic rate (lower than 5%) are commonly found in the Cerrado [54] although they probably behave functionally as monoembryonic species. The role of polyembryony in these species and its relationship with seed biology has yet to be elucidated as it also occurs in Bignoniaceae [52], which prevents further discussions. Polyembryonic individuals from the non-Cerrado Bignoniaceae species *Dolichandra unguiscati* (L.) L.G. Lohmann, have a seedling emergence time of more than two months [55], probably showing another case of dormancy in the group. Long and short germination and emergence times also occur in species with low polyembryony frequencies, as in *J. cuspidifolia* (long times) and in *T. roseoalba* (short times). Apparently, polyembryonic species are not associated with variations in the germination and emergence times.

Handroanthus chrysotrichus has commonly high polyembryony [7] and does not present any atypical behavior in germination and or emergence compared to other species of the family. None of the monoembryonic populations of *H. serratifolious* [56], the monoembryonic and polyembryonic populations of *H. ochraceus* [49], or even the other polyembryonic species of the family [57], have been studied for germination measurements. These data could elucidate the role of polyembryony in the group. Some studies have related the seed-germination measurements to polyembryonic expression or the existence of monoembryonic or polyembryonic populations [6,46,49]. In Bignoniaceae, more data are still needed for the group to allow such relationships, probably requiring phylogenetic corrections or the study of populations with differences in polyembryony or its expression.

Changes in the uncertainty and emergence time of the last seed as a function of the seed's gemellar class do not have a definitive explanation. Among the hypotheses, we think that the embryos, even the smaller ones, collaborate with each other or stimulate each other in the rupture of the tegument, which can accelerate and synchronize the germination process. Alternatively, the seeds where only one seedling emerged could have a higher expression of genetic variability because they are more likely to be monoembryonic and sexual seeds (see Mendes-Rodrigues et al. [7]). The emergence time could thus reflect the contribution of the paternal effect as observed in some species [58], leading to a more asynchronous and possibly slower process in seeds with only one seedling. Mendes-Rodrigues et al. [59] showed that for the polyembryonic Malvaceae, *Eriotheca pubescens* (Mart. and Zucc.) Schott and Endl, the type of pollen (auto versus crossed) may affect polyembryony, with the MENS being higher when pollen comes from another individual. This would reinforce the paternal effect on the expression of polyembryony. Further data and other examples are still necessary to test these hypotheses.

The study of germination measurements in the seeds' embryonic or gemellar classes is hampered by some morphological aspects. In most species it is not possible to directly evaluate the number of embryos per seed before germination, except for *Ophiopogon japonicus* (Thunb.) Ker Gawl. var. *umbrosus* Maxim [60], which has a translucent integument that allows the evaluation of the number of embryos per seed, even before the embryo's protrusion. The evaluation of the ESC or GSC does not allow, for example, the estimation of germinability in our case. In these cases, the conversion of embryos into seedlings is an indirect measurement of the viability of extra embryos in the species.

The conversion of 67.22% of embryos into seedlings was high in *H. chrysotrichus*. The decrease in the number of seedlings in relation to the number of embryos produced has been commonly observed [7,59,61,62]; however, the conversion rate in most species has not been measured and its determinants are unknown. It is known that temperature affects *H. chrysotrichus* [21] and temperatures between 20 and 30 °C allow the conversion of more embryos into seedlings. In some genotypes of *Citrus*, about 50% of the embryos were converted into seedlings, with great variability among genotypes [63]. In addition, the seed forehead has a strong relationship with the decrease in the conversion of embryos

into seedlings, as observed in several species in which the removal of the seed tegument increased the conversion (e.g., [61,62]).

Monoembryonic and polyembryonic individuals within the same genus and or species could be an alternative to test these hypotheses. They could show some marked differences in the germination process and its measurements. In *Inga laurina* (Sw.) Willd., a polyembryonic legume species, the process of seedling emergence is much slower than in the monoembryonic species *I. sessilis* (Vell.) Mart. [46]. In *E. pubescens*, the seeds of polyembryonic individuals showed a higher coefficient of variability in the germination time (75.38%) than the seeds of monoembryonic individuals (52.66%) [6]. These effects of monoembryony and polyembryony should be evaluated within each group of species carefully, since they may depend on other factors such as ploidy, phylogenetic and environmental conditions [64,65].

Monoembryonic populations of *H. chrysotrichus* are not known, but if they do exist, they could help to understand the effects of polyembryony more directly. An individual of this species that does not produce fruit by manual self-pollination, indicating that it is self-incompatible, has been observed [66], signaling the possibility that sexual plants with monoembryonic seeds occur. However, the pattern of embryony in this individual has not been described, which prevents its definition as monoembryonic based only on the reproductive system. Population studies are still needed.

Intraspecific variability was also detected in *H. chrysotrichus*'s PSP (14 to 41%) [20], but with values much lower than those observed here. The selection of individuals with no variability in the mean number of embryos and seedlings per seed would be a beneficial factor in experimental competition studies if the individual factor was not of interest to the study. The expression of polyembryony can be very variable among species, individuals, populations, and even in the same individual evaluated at different times [7,67]. On the other hand, the factors that determine the expression of polyembryony are still poorly studied, but there is a record of the effects of pollen type, of the presence of conspecific individuals, the nutritional status, the age of the individual, the orientation of the branch, and of fruit production [54,59,68].

Studies have shown that with the increase in the number of embryos and seedlings per seed, there is a decrease in the individual morphometric measurements of embryos or seedlings [4,6,7,60]. Seedling size is a direct effect of embryo size in polyembryonic species, regardless of whether the embryos are sexual or asexual [6,7,59,69]. The reduction in embryo size and consequently seedling size should be compensated, for example, by the effects of growing together, as observed here in doublet seedlings, or it could be compensated in later stages of development. Apparently, the deleterious effects of polyembryony are more marked and stronger in the early stages of the seed–seedling transition and seedling development and may disappear over time [4] so that an intermediate number of embryos may be more beneficial for the species [60]. This would explain the Allee effect when there was a density of two, as it would decrease the reduction in embryo size. As an example of compensation, polyembryony increases the chances of at least one seedling surviving per seed [7].

The nondestructive model of mass prediction of *H. chrysotrichus* is suitable for estimating the seedling mass at 54 days, a phase in which manipulation can be performed without causing visible damage to the seedling. These results may allow, for example, the proposition of experimental models of nondestructive competition for the species. Models of prediction of dry mass, leaf area and other nondestructive morphometric variables have been proposed for the most diverse groups, using traditional methods of regression and image acquisition, among others [70,71]. These methods are fundamental for the design of experimental studies manipulating seedling growth without destruction, and with registration of the initial point of the experiment. Knowing how seedlings of different sizes respond to gemellar competition is crucial in the study of polyembryonic species; being able to predict their initial mass using nondestructive methods is also essential. The model also clearly demonstrated the impact of increasing the number of seedlings on the reduction of their individual measurements, as discussed previously.

5. Conclusions

We did not observe intraspecific variability in the seed-seedling transition of *H. chrysotrichus* seeds. However, we did observe differences associated with polyembryonic classes. The seed gemellar classes, with different numbers of embryos per seed, interfered with the emergence time, decreasing the time required by the last seedling to emerge in each seed from a higher class. The increase in the number of embryos per seed reduced the size of embryos and seedlings, which reduced individual seedling's vigor. Despite this reduction, the seeds that produced two seedlings per seed (doublet) showed a higher joint growth than the ones that produced one (singlet) or three seedlings per seed (triplet), demonstrating the occurrence of the Allee effect. Both positive and negative effects of polyembryony were observed in the seed biology and early plant development of this species. These data confirm the usefulness of the species as an experimental model.

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References

- 1. Smolikova, G.; Medvedev, S. Seed-to-Seedling Transition: Novel Aspects. Plants 2022, 11, 1988. [CrossRef]
- 2. Strasburger, E. Über Polyembryonie. Z. Nat. 1878, 12, 647–760.
- 3. Carman, J.G. Asynchronous expression of duplicate genes in angiosperms may cause apomixis, bispory, tetraspory, and polyembryony. *Biol. J. Linn. Soc.* **1997**, *61*, 51–94. [CrossRef]
- Hotchkiss, E.E.; DiTommaso, A.; Brainard, D.C.; Mohler, C.L. Survival and performance of the invasive vine *Vincetoxicum rossicum* (Apocynaceae) from seeds of different embryo number under two light environments. *Am. J. Bot.* 2008, 95, 447–453. [CrossRef]
 Blanchard, M.L.; Barney, J.N.; Averill, K.M.; Mohler, C.L.; Ditommaso, A. Does polyembryony confer a competitive advantage to
- the invasive perennial vine *Vincetoxicum rossicum* (Apocynaceae)? *Am. J. Bot.* **2010**, *97*, 251–260. [CrossRef] [PubMed]
- Mendes-Rodrigues, C.; Ranal, M.A.; Oliveira, P.E. Does polyembryony reduce seed germination and seedling development in Eriotheca pubescens (Malvaceae: Bombacoideae)? Am. J. Bot. 2011, 98, 1613–1622. [CrossRef] [PubMed]
- Mendes-Rodrigues, C.; Sampaio, D.S.; Costa, M.E.; de Souza Caetano, A.P.; Ranal, M.A.; Bittencourt, N.S.; Oliveira, P.E. Polyembryony increases embryo and seedling mortality but also enhances seed individual survival in *Handroanthus* species (Bignoniaceae). *Flora* 2012, 207, 264–274. [CrossRef]
- 8. Krebs, C.J. *Ecology: The Experimental Analysis of Distribution and Abundance*, 6th ed.; Benjamin Cummings: San Francisco, CA, USA, 2009; 655p.

- 9. Mendes-Rodrigues, C.; Oliveira, P.E. Polyembryony in Melastomataceae from Brazilian Cerrado: Multiple embryos in a small world. *Plant Biol.* **2012**, *14*, 845–853. [CrossRef]
- Cappuccino, N. Allee effect in an invasive alien plant, pale swallow-wort *Vincetoxicum rossicum* (Asclepiadaceae). *Oikos* 2004, 106, 3–8. [CrossRef]
- 11. Ladd, D.; Cappuccino, N. A field study of seed dispersal and seedling performance in the invasive exotic vine *Vincetoxicum* rossicum. Can. J. Bot. 2005, 83, 1181–1188. [CrossRef]
- 12. Piazzano, M. Nuúmeros cromosoómicos en Bignoniaceae de Argentina. Kurtziana 1998, 26, 179–189.
- 13. Costa, M.E.; Sampaio, D.S.; Paoli, A.A.S.; Leite, S.C.A.L. Polyembryony and aspects of embryogenesis in *Tabebuia ochracea* (Chamisso) Standley (Bignoniaceae). *Braz. J. Bot.* **2004**, 27, 395–406. [CrossRef]
- 14. Bittencourt, N.S.; Moraes, C.I.G. Self-fertility and polyembryony in South American yellow trumpet trees (*Handroanthus chrysotrichus* and *H. ochraceus*, Bignoniaceae): A histological study of postpollination events. *Plant Syst. Evol.* **2010**, *288*, 59–76. [CrossRef]
- Sampaio, D.S. Biologia Reprodutiva de Espeécies de Bignoniaceae Ocorrentes No Cerrado E Variações No Sistema de Autoincompatibilidade. Ph.D. Thesis, Universidade Federal de Uberlândia, Uberlândia, Brazil, 2010. Available online: https: //repositorio.ufu.br/handle/123456789/13259 (accessed on 1 February 2023).
- Fonseca, L.F.; Mengaria, C.; Mori, E.S.; Nakagawa, J. Physiological maturity of ipê amarelo seeds, *Tabebuia chrysotricha* (Mart. Ex DC.) Standl. *Sci. For.* 2005, *69*, 136–141.
- 17. Oliveira, A.K.M.; Scheleder, E.J.D.; Favero, S. Morphological characterization, viability, and vigor of *Tabebuia chrysotricha* (Mart. ex. DC.) Standal. *Rev. Árvore* 2008, 32, 1011–1018. [CrossRef]
- Martins, C.C.; Martinelli-Seneme, A.; Nakagawa, J. Harvest stage and substratum for ipê (*Tabebuia chrysotricha* (Mart. ex DC.) Standl.) seed gemination test. *Rev. Árvore* 2008, 32, 27–32. [CrossRef]
- 19. Martins, L.; Lago, A.A.; Sales, W.R.M. Preservation of *Tabebuia chrysotricha* seeds as a function of seed water content and storage temperature. *Rev. Bras. Sementes* 2009, *31*, 86–95. [CrossRef]
- Dos Santos, F.S.; de Paula, R.C.; Sabonaro, D.Z.; Valadares, J. Biometric and physiological quality of *Tabebuia chrysotricha* (Mart. Ex A. DC.) Standl. seeds from different mother trees. *Sci. For.* 2009, *37*, 163–173.
- Sampaio, D.S.; Costa, M.E.; Mendes-Rodrigues, C. Temperature effect in the number of seedlings per seed in cultivated specimens of *Handroanthus chrysotrichus* (Bignoniaceae). *Iheringia Série Botânica* 2013, 68, 279–283.
- 22. Sarzi, I.; Bôas, R.L.V.; da Silva, M.R. Development of *Tabeluia chrysotricha* seedlings in function of the substrate and fertirrigation solutions. *Cerne* **2008**, *14*, 153–162.
- 23. Gentry, A.H. A synopsis of Bignoniaceae ethnobotany and economic botany. Ann. Mo. Bot. Gard. 1992, 79, 53-64. [CrossRef]
- Oliveira-Filho, A.T.; Ratter, J.A. Vegetation physiognomies and woody flora of the Cerrado Biome. In *The Cerrados of Brazil: Ecology and Natural History of a Neotropical Savanna*; Oliveira, P.S., Marquis, R.J., Eds.; Columbia University Press: New York, NY, USA, 2002; pp. 91–120.
- Kottek, M.; Grieser, J.; Beck, C.; Rudolf, B.; Rubel, F. World map of the Köppen-Geiger climate classification updated. *Meteorol. Z.* 2006, 15, 259–263. [CrossRef] [PubMed]
- Ranal, M.A. Soil spore bank of ferns in a gallery forest of the Ecological Station of Panga, Uberlândia, MG, Brazil. Am. Fern. J. 2003, 93, 97–115. [CrossRef]
- 27. Ranal, M.A.; Santana, D.G. How and why to measure the germination process? Braz. J. Bot. 2006, 29, 1–11. [CrossRef]
- Ranal, M.A.; Santana, D.G.; Ferreira, W.R.; Mendes-Rodrigues, C. Calculating germination measurements and organizing spreadsheets. *Braz. J. Bot.* 2009, 32, 849–855. [CrossRef]
- 29. Labouriau, L.G.; Valadares, M.E.B. On the germination of seeds of *Calotropis procera* (Ait.) Ait. F. An. Acad. Bras. Ciências 1976, 48, 263–284.
- R Core Team. R: A Language and Environment for Statistical Computing; R Foundation for Statistical Computing: Vienna, Austria, 2022. Available online: http://www.R-project.org/ (accessed on 1 February 2023).
- Saatkamp, A.; Cochrane, A.; Commander, L.; Guja, L.K.; Jimenez-Alfaro, B.; Larson, J.; Nicotra, A.; Poschlod, P.; Silveira, F.A.O.; Cross, A.T.; et al. A research agenda for seed-trait functional ecology. *New Phytol.* 2019, 221, 1764–1775. [CrossRef]
- 32. Steinbrecher, T.; Leubner-Metzger, G. The biomechanics of seed germination. J. Exp. Bot. 2016, 68, 765–783. [CrossRef]
- 33. Baskin, J.M.; Baskin, C.C. Seeds: Ecology, Biogeography, and Evolution of Dormancy and Germination; Elsevier: San Diego, CA, USA, 1998. [CrossRef]
- 34. Bassel, G.W. To Grow or not to Grow? Trends Plant Sci. 2016, 21, 498–505. [CrossRef] [PubMed]
- 35. Donohue, K. Completing the cycle: Maternal effects as the missing link in plant life histories. *Philos. Trans. R. Soc.* **2009**, 364, 1059–1074. [CrossRef]
- 36. Lamichhane, J.R.; Debaeke, P.; Steinberg, C.; You, M.P.; Barbetti, M.J.; Aubertot, J.-N. Abiotic and biotic factors affecting crop seed germination and seedling emergence: A conceptual framework. *Plant Soil* **2018**, 432, 1–28. [CrossRef]
- Muscarella, R.; Uriarte, M.; Forero-Montaña, J.; Comita, L.S.; Swenson, N.G.; Thompson, J.; Nytch, C.J.; Jonckheere, I.; Zimmerman, J.K. Life-history trade-offs during the seed-to-seedling transition in a subtropical wet forest community. J. Ecol. 2013, 101, 171–182. [CrossRef]
- Bewley, J.D.; Bradford, K.J.; Hilhorst, H.W.M.; Nonogaki, H. Seeds: Physiology of Development, Germination and Dormancy, 3rd ed.; Springer: New York, NY, USA, 2013; 393p. [CrossRef]

- 39. Carrera-Castaño, G.; Calleja-Cabrera, J.; Pernas, M.; Gómez, L.; Oñate-Sánchez, L. An updated overview on the regulation of seed germination. *Plants* **2020**, *9*, 703. [CrossRef]
- 40. Nonogaki, H.; Bassel, G.W.; Bewley, J.D. Germination-Still a mystery. Plant Sci. 2010, 179, 574-581. [CrossRef]
- 41. Penfield, S. Seed dormancy and germination. *Curr. Biol.* **2017**, *27*, R874–R878. [CrossRef]
- 42. Kranner, I.; Minibayeva, F.V.; Beckett, R.P.; Seal, C.E. What is stress? Concepts, definitions and applications in seed science. *New Phytol.* 2010, *188*, 655–673. [CrossRef]
- Finch-Savage, W.E.; Bassel, G.W. Seed vigour and crop establishment: Extending performance beyond adaptation. J. Exp. Bot. 2016, 67, 567–591. [CrossRef]
- Reed, R.C.; Bradford, K.J.; Khanday, I. Seed germination and vigor: Ensuring crop sustainability in a changing climate. *Heredity* 2022, 128, 450–459. [CrossRef] [PubMed]
- 45. Martins, R.L.; Oliveira, P.E. RAPD Evidence for apomixis and clonal populations in *Eriotheca* (Bombacaceae). *Plant Biol.* **2003**, *5*, 338–340. [CrossRef]
- 46. Mendes-Rodrigues, C.; Ranal, M.; Santana, D.G. Seedling emergence and polyembryony in two species of *Inga* (Mimosaceae). In Proceedings of the II Simpoósio Internacional Savanas Tropicais, IX Simpoósio Nacional Cerrado, Brasiília, Brazil, 12–17 October 2008. Available online: http://www.cpac.embrapa.br/download/429/t (accessed on 16 April 2017).
- Mendes-Rodrigues, C.; Araújo, F.P.; Souza, C.B.; Souza, V.B.; Ranal, M.A.; Santana, D.G.; Oliveira, P.E. Multiple dormancy and maternal effect on *Miconia ferruginata* (Melastomataceae) seed germination, Serra de Caldas Novas, Goiás, Brazil. *Braz. J. Bot.* 2010, 33, 93–105. [CrossRef]
- Ranal, M.A.; Mendes-Rodrigues, C.; Teixeira, W.F.; Oliveira, A.P.; Romero, R. Seed germination of *Microlicia fasciculata*, an apomictic and aluminium accumulator species: Unexpected intraspecific variability in a restricted Neotropical savanna area. *Flora* 2016, 220, 8–16. [CrossRef]
- 49. Mendes, M.G.; de Oliveira, A.P.; Oliveira, P.E.; Bonetti, A.M.; Sampaio, D.S. Sexual, apomictic and mixed populations in *Handroanthus ochraceus* (Bignoniaceae) polyploid complex. *Plant Syst. Evol.* **2018**, *304*, 817–829. [CrossRef]
- 50. Mori, N.T.; de Moraes, M.L.T.; Morita, C.M.; Mori, E.S. Genetic diversity between and within populations of *Handroanthus heptaphyllus* (Vell.) Mattos using microsatellite markers. *Cerne* **2012**, *18*, 9–15. [CrossRef]
- 51. Moreira, P.A.; Fernandes, G.W. Is the São Francisco River a geographic barrier to gene flow in trees of *Handroanthus ochraceus*? J. *Trop. Ecol.* **2013**, *29*, 243–250. [CrossRef]
- 52. Ranal, M.A.; Santana, D.G.; Schiavini, I. Are there germination patterns for cerrado species. In *Encyclopedia of Life Support Systems*. *Tropical Biology and Conservation Management, Savannah Ecosystems*; II Simpoósio Internacional Savanas Tropicais, IX Simpoósio Nacional Cerrado; Del-Claro, K., Oliveira, P.S., Rico-Gray, V., Barbosa, A.A.A., Bonet, A., Scarno, F.R., Garzón, F.J.M., Sampaio, M.V., Morris, M.R., Ramirez, N., et al., Eds.; UNESCO/EOLSS: Oxford, UK, 2010; pp. 106–159.
- 53. Mendes-Rodrigues, C.; Oliveira, P.E.; Marinho, R.C.; Romero, R.; Ranal, M.A. Are the alien species of Melastomataceae and Bombacoideae a potential risk for Brazilian Cerrado? *Open Access Libr. J.* **2019**, *6*, 1–4. [CrossRef]
- Mendes-Rodrigues, C. Ecologia de Espécies Poliembriônicas com ênfase no Bioma Cerrado. Ph.D. Thesis, Universidade Federal de Uberlândia, Uberlândia, Brazil, 2010. Available online: https://repositorio.ufu.br/handle/123456789/13262 (accessed on 1 February 2023).
- Buru, J.C.; Dhileepan, K.; Osunkoya, O.O.; Scharaschkin, T. Germination biology and occurrence of polyembryony in two forms of cats claw creeper vine, *Dolichandra unguis-cati* (Bignoniaceae): Implications for its invasiveness and management. *Am. J. Plant Sci.* 2016, 7, 657–670. [CrossRef]
- 56. Alves, M.F.; Duarte, M.O.; Oliveira, P.E.; Sampaio, D.S. Self-sterility in the hexaploid *Handroanthus serratifolius* (Bignoniaceae), the national flower of Brazil. *Acta Bot. Bras.* 2013, 27, 714–722. [CrossRef]
- 57. Sampaio, D.S.; Bittencourt, N.S., Jr.; Oliveira, P.E. Sporophytic apomixis in polyploid *Anemopaegma* species (Bignoniaceae) from central Brazil. *Bot. J. Linn. Soc.* 2013, 173, 77–91. [CrossRef]
- Baskin, J.M.; Baskin, C.C. How much influence does the paternal parent have on seed germination? Seed Sci. Res. 2019, 29, 1–11. [CrossRef]
- 59. Mendes-Rodrigues, C.; Carmo-Oliveira, R.; Talavera, S.; Arista, M.; Ortiz, P.L.; Oliveira, P.E. Polyembryony and apomixis in *Eriotheca pubescens* (Malvaceae-Bombacoideae). *Plant Biol.* **2005**, *7*, 533–540. [CrossRef]
- 60. Oka, C.; Itagaki, T.; Sakai, S. Effects of the number of embryos in a seed and seed mass on seedling survival and growth in polyembryonic *Ophiopogon japonicus* var. *umbrosus* (Asparagaceae). *Botany* **2016**, *94*, 261–268. [CrossRef]
- 61. Sinnadurai, S. The effect of decortication of seed on germination and the number of nucellar seedlings of some mango cultivars of Ghana. *Acta Hortic.* **1975**, *49*, 95–97. [CrossRef]
- 62. Muralidhara, B.M.; Reddy, Y.T.N.; Srilatha, V.; Akshitha, H.J. Effect of seed coat removal treatments on seed germination and seedling attributes in mango varieties. *Int. J. Fruit Sci.* 2016, *16*, 1–9. [CrossRef]
- Kishore, K.; Rinchen, M.N.; Lepcha, B.; Pandey, B. Polyembryony and seedling emergence traits in apomictic *Citrus. Sci. Hortic.* 2012, 138, 101–107. [CrossRef]
- 64. Astuti, G.; Pratesi, S.; Peruzzi, L.; Carta, A. Two closely related *Tulipa* species with different ploidy levels show distinct germination rates. *Seed Sci. Res.* 2020, *30*, 45–48. [CrossRef]
- 65. Wang, Z.; Wang, L.; Liu, Z.; Li, Y.; Liu, Q.; Liu, B. Phylogeny, seed trait, and ecological correlates of seed germination at the community level in a degraded sandy grassland. *Front. Plant Sci.* **2016**, *7*, 1532. [CrossRef]

- 66. Bittencourt, N.S., Jr.; Semir, J. Late-acting self-incompatibility and other breeding systems in *Tabebuia* (Bignoniaceae). *Int. J. Plant Sci.* 2005, *166*, 493–506. [CrossRef]
- 67. Mendes-Rodrigues, C.; Marinho, R.C.; Balao, F.; Arista, M.; Ortiz, P.; Carmo-Oliveira, R.; Oliveira, P.E. Reproductive diversity, polyploidy, and geographical parthenogenesis in two *Eriotheca* (Malvaceae) species from Brazilian Cerrado. *Perspect. Plant Ecol. Evol. Syst.* **2019**, *36*, 1–12. [CrossRef]
- 68. Bhojwani, S.S.; Bhatnagar, S.P. The Embryology of Angiosperms; Vikas Publishing House Pvt Ltd.: New Delhi, India, 2008.
- 69. Thurlby, K.A.G.; Wilson, P.G.; Sherwin, W.B.; Connelly, C.; Rossetto, M. Reproductive bet-hedging in a rare yet widespread rainforest tree, *Syzygium paniculatum* (Myrtaceae). *Austral. Ecol.* **2012**, *37*, 936–944. [CrossRef]
- Norgren, O.; Elfving, B.; Olsson, O. Non-destructive biomass estimation of tree seedlings using image analysis. *Scand. J. For. Res.* 1995, 10, 347–352. [CrossRef]
- 71. Montagnoli, A.; Terzaghi, M.; Fulgaro, N.; Stoew, B.; Wipenmyr, J.; Ilver, D.; Rusu, C.; Scippa, G.S.; Chiatante, D. Non-destructive phenotypic analysis of early stage tree seedling growth using an automated stereovision imaging method. *Front. Plant Sci.* **2016**, *7*, 1644. [CrossRef] [PubMed]

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