

Review

Phototherapy-Based Treatment for Sexually Transmitted Infections—Shining Light into Unexplored Territory

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Abstract: New therapeutic strategies are urgently needed to overcome drawbacks in the treatment of some infections, particularly sexually transmitted infections (STI). STIs are easily spread by the transmission of various bacteria, viruses, and parasites with some of the infections being incurable or even lethal, leading to a serious impact on reproductive health worldwide. Phototherapy (PT) is a major therapeutic approach based on the controlled administration of light in the visible, near infrared, or UV spectrum, with or without the application of an external photosensitizer. Despite the fact that PT has not been explored to its full potential in the control of STIs, it has already demonstrated good clinical response rates and lower recurrence rates in genital infections. For instance, increasing evidence has demonstrated that 5-aminolevulinic acid photodynamic therapy (5-ALA-PDT) is effective in the treatment of condyloma acuminatum (CA), by eliminating the causative latent human papillomavirus (HPV) infection, and also in the antiviral treatment of recurrent genital herpes simplex virus (HSV) infections. The clinical application of PDT is a new treatment for oral fungal infection caused by *Candida albicans* in adult acquired immune deficiency syndrome (AIDS) patients, with human immunodeficiency virus (HIV), and could also be used for genital fungal infections. Another antimicrobial PT strategy, water-filtered infrared A combined with visible light irradiation, has been shown to be effective against genital *Chlamydia trachomatis* bacterial infection, and an optical nano-genosensor has been designed for the diagnosis of trichomoniasis, a parasitic *Trichomonas vaginalis* infection. This review aims to summarize the published evidence for the effectiveness of PT in the treatment of STIs, and for the suppression of STI-related pathogens of various types.

Keywords: sexually transmitted infections; phototherapy; photodynamic therapy



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1. Introduction

Sexually transmitted infections (STIs) are a major public health problem throughout the world [1]. They can be either ancient or emerging infections, with an increasing likelihood of being resistant to treatment [2]. Three types of viruses are often responsible for STIs [3]. First and foremost are human papillomavirus (HPV) infections, which have recently been brought under control by the development of highly effective vaccines [4]. Herpes virus infections (HSV) are characterized by high prevalence and morbidity [5]. No effective vaccine against viruses of the herpes group is currently available. This is also the case with HIV (human immunodeficiency virus) infections [6]. STIs can also be caused by non-viral pathogens such as bacteria, parasites, or fungi. Examples of these are *Chlamydia trachomatis*, *Neisseria gonorrhoeae*, *Treponema pallidum*, *Mycoplasma genitalium* and *Trichomonas vaginalis* [7]. While the site of infection is mainly urogenital, it can also be pharyngeal and/or anorectal [2]. Although pharyngeal infections with *C. trachomatis* and *N. gonorrhoeae*

often resolve spontaneously, they can be transmitted by oral sex, suggesting a large oral reservoir of germs as a source of contamination [8,9]. Genital and extra-genital co-infection is far from rare and could explain the occurrence of re-infection and the sustainability of an epidemic [8]. The clinical presentation could be an erosive or ulcerative anorectal or genital lesion, more or less painful or raised [10]. The context is important to confirm the diagnosis, and possible co-infections should be systematically sought out because they are frequent, and this is essential in HIV-positive patients [11].

Moreover, the emergence of antimicrobial resistance in sexually acquired infection pathogens is an important global public health threat. There is an urgent need for novel STI treatment and prevention strategies to tackle the rising incidence of STIs in high-income settings, and the static incidence in low- and middle-income settings over the past decade [12]. Both the treatment and the prevention of these infections are often complex. Recent clinical studies have made it possible to use new light-based therapeutic approaches to treat skin lesions caused by sexually transmitted viral infections [13].

The primary care therapeutic approaches to treat benign skin lesions associated with HPV infection are quite diverse, such as topical therapy with agents such as podophyllin, trichloroacetic acid, salicylic acid, or 5-fluorouracil. However, 5-fluorouracil has the disadvantage of inducing a strong local inflammatory response, which makes its use on mucous membranes difficult [14]. Similar treatments have been used in cases of vaginal intraepithelial neoplasia with high-risk HPV infection, and other sites of this infection, including excision by surgery and intravaginal radiotherapy or chemotherapy [15]. Some physical methods constitute an alternative approach employed in current practice [16,17]. Several different approaches could be considered: cryotherapy [18]; photodynamic therapy (PDT) [19]; thermocoagulation [20]; CO₂ laser ablation [21]. However, chemotherapy has certain disadvantages and side effects which can vary according to the individual patient, the drugs used, the doses, and the combination treatments. In some cases chemotherapy is performed during concomitant radiochemotherapy [22]. This therapeutic approach does not have the same side effects as chemotherapy alone, and it may be more effective than chemotherapy alone, because it targets the tissue with malignant cells in a spatially confined manner [23,24]. The undesirable effects often include digestive disorders, a drop in immune cells, red blood cells, and platelets, oral mucositis, and the appearance of skin disorders [25,26]. Locally applied laser and radiation therapies could improve the cure rate while avoiding the damage caused by surgery [27], but they could also cause damage to the vaginal mucosa resulting in scarring or vaginal stenosis, and other adverse effects such as burning sensations, pain, and dyspareunia [28,29]. Moreover, the development of therapeutic HPV vaccines could be used for the treatment of persistent infections, and to prevent the progression of HPV-associated cancers [6]. There are three vaccines called Cervarix (GlaxoSmithKline, London, United Kingdom), Gardasil and Gardasil9 (Merck Sharp & Dohme Corp, Whitehouse Station, Township of Readington, New Jersey, United States). Cervarix can protect against oncogenic HPV types HPV16 and HPV18. Gardasil provides protection against oncogenic types HPV16 and 18, as well as low-risk HPV types 6 and 11. Gardasil9 provides protection against a broad panel of HPV strains, protecting against the seven most common oncogenic HPV types (HPV16, 18, 31, 33, 45, 52 and 58) and the two low risk types (HPV6 and 11) [30,31]. For HSV lesions, the primary therapeutic arsenal consists of anti-herpesviridae antiviral drugs such as aciclovir, valaciclovir, or famciclovir [32]. The use of topical aciclovir still has a strong following. However, new approaches are in the evaluation phase, but do not yet present an established management plan in current practice [33].

For infectious diseases caused by *C. trachomatis*, the first-line treatment is doxycycline in the European and American guidelines [34]. However, in the event of an allergy to doxycycline or in pregnant women, a single dose of oral azithromycin or erythromycin (500 mg) has been used [35]. Exceptional treatment failure has been reported with this treatment, which could then be controlled with moxifloxacin. Due to the frequent co-infection with gonococcus bacteria, treatment with ceftriaxone is often added to this regimen. Neverthe-

less, the notable limitations of these antibiotics, highlight the need for further, updated research in this area, particularly for low- and middle-income settings [36]. Furthermore, azithromycin (500 mg) and doxycycline (100 mg) are also recommended for the treatment of *Mycoplasma genitalium*. The persistence of this infection requires treatment with pristinamycin (1 g) [37]. To date, all findings suggest that doxycycline is inefficient for the eradication of *M. genitalium*. Although azithromycin was not significantly less efficient than extended dosage doxycycline, it was associated with the selection of macrolide-resistant *M. genitalium* strains. The monitoring of *M. genitalium* macrolide resistance should be encouraged.

In the case of gonococcal infections, the WHO Global Gonococcal Antimicrobial Surveillance Program (WHO GASP) reported that between 2009 and 2014, there emerged a persistent and widespread resistance to penicillin, tetracycline, ciprofloxacin, and azithromycin. This was accompanied by a decrease in sensitivity to broad-spectrum cephalosporins, especially cefixime [38]. To date, three strains of gonococci with a high resistance to ceftriaxone have been reported in France, Japan and Spain [39,40]. There is a need to develop and implement a national sentinel program for gonococcal antimicrobial susceptibility and to develop new therapeutic strategies to combat this scourge.

STIs can also be caused by a parasitic infection, particularly *Trichomonas vaginalis*. The standard treatment for trichomoniasis is metronidazole. This molecule is an antibiotic and an antiparasitic drug. Treatment adherence should be followed closely to avoid re-infection [41]. Therapeutic failure of this protocol sometimes occurs, and then the treatment dose is increased. Because metronidazole can cause leukopenia, an antabuse effect (similar to disulfiram) or a candida secondary infection, there is a relative contraindication in early pregnancy [42].

Drug resistance is increasingly observed in pathogens causing STIs. Clinical trials have used some physical therapy approaches that do not cause resistance. In this review, we are particularly interested in the treatment of STIs by phototherapy (PT). PT is a major therapeutic approach based on the controlled administration of light in the visible, near infrared, or UV spectrum, with or without the application of an external photosensitizer. When the light is combined with a photosensitizer, it is called photodynamic therapy (PDT). PDT is a physicochemical method initially developed to treat cancer and tumors [19]. It was first approved in the 1980s, as a new approach intended for cancer patients who could not be treated by surgery or radiotherapy [43]. During phototherapy, the non-toxic photosensitizer agents can be activated by light irradiation to induce cell death without causing much damage to normal tissues [44]. A successful clinical PDT involves complex procedures, but can lead to the eradication of tumor or infected tissue with lower toxicity due to more limited light penetration compared to PT [44]. The three key components of the PDT are photosensitizer, oxygen and light, with light dosimetry being a key factor. The photosensitizer should have some selective affinity for the target cells. Since the 1980s, three generations of photosensitizers have emerged. The first generation derivatives were the first photosensitizers to have been used clinically, hematoporphyrin and its derivatives (HpD) [45]. HpD is difficult to obtain pure and contains oligomeric compounds responsible for photocytotoxicity [44]. Photofrin (first generation) is a purified form of HpD. Photofrin can persist in normal skin for long periods of time [46,47], meaning that patients should be protected from strong light sources for several weeks after Photofrin administration. This problem was minimized when new photosensitizing agents were identified with shorter persistence [46]. The second generation photosensitizers (e.g., Visudyne, Foscan, 5-aminolevulinic acid (5-ALA) ...) have been developed to overcome some of these drawbacks [48]. They are pure compounds; mostly absorbing strongly in the red part of the spectrum, and have a high quantum yield of singlet oxygen formation. Rapid elimination of the photosensitizer from the body is also desirable in order to limit residual phototoxicity after treatment. The third generation photosensitizers have been modified by conjugation or by encapsulation (e.g., liposomes and nanoparticles) of the second generation photosensitizers in order to allow passive or active targeting of neoplastic cells, and thus improve their selectivity for the lesion to be treated [49]. Basic studies made it

possible to establish the mechanistic principles of PDT, which relies on the absorption of light by a photosensitizer molecule or dye, leading to a photochemical reaction to produce reactive oxygen species that can kill cancer cells or microorganisms [46]. Furthermore, it is only relatively recently that PDT has been studied as a treatment for various types of localized infections. This resurgence of interest has been partly motivated by the alarming increase in drug resistance amongst bacteria and other pathogens. The clinical application of antimicrobial PDT to localized viral infections caused by herpes or papilloma viruses, or non-viral dermatological infections such as acne, yeast, fungal, and bacterial skin infections has been validated. PDT has been used to treat bacterial infections in brain abscesses and non-healing ulcers [50]. Figure 1 schematically illustrates the applications of PT and PDT to treat a variety of STIs.

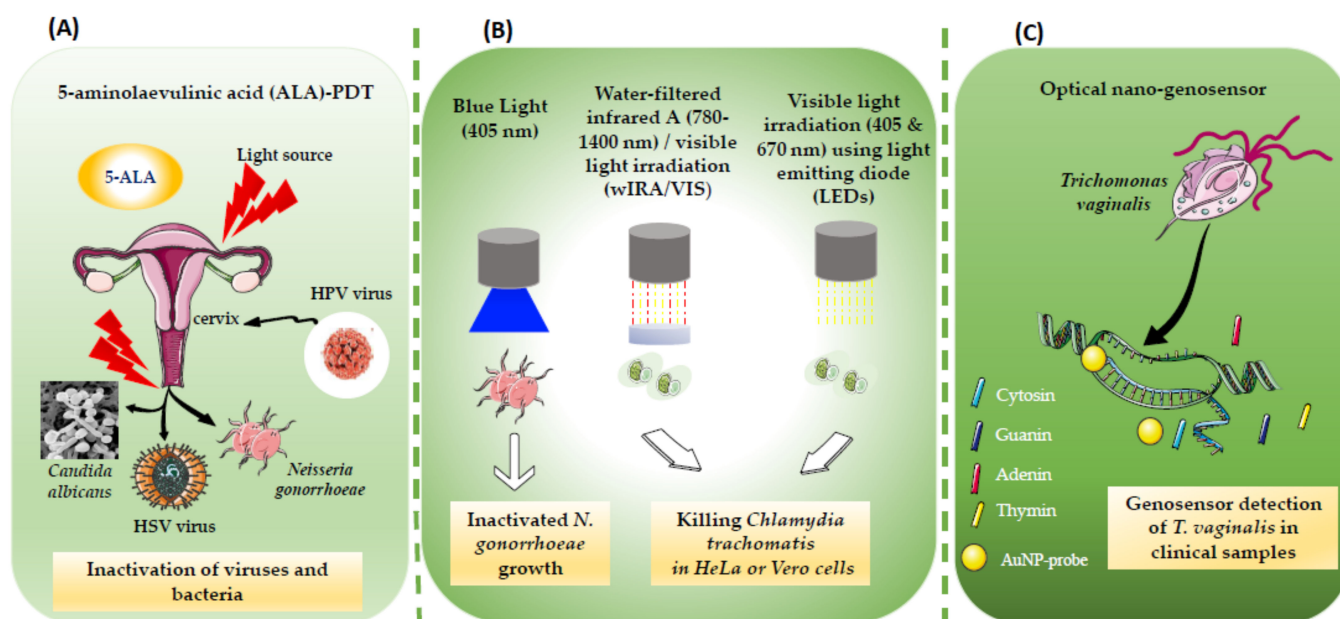


Figure 1. (A) Treatment of multifocal disease based on 5-aminolaevulinic acid (ALA)-PDT, effective in treating HPV lesions. The PDT can be also used for selective and specific destruction of subclinical bacteria and fungi-infected areas, as in the case of *N. gonorrhoeae* and *C. albicans*. (B) The combination of water-filtered infrared A (wIRA) with visible light VIS (wIRA/VIS) is used for killing intracellular and extracellular *Chlamydia trachomatis* strains in Vero and HeLa cell lines. Visible light irradiation (405 and 670 nm) also leads to dose-dependent inhibitory effect on *Chlamydia*. (C) Diagnostic approach based on an optical nano-genosensor and gold nanoparticles conjugated to a specific oligonucleotide for *T. vaginalis* PCR detection.

2. PT for Viral STIs

Human papilloma virus (HPV) infection, the most common sexually transmitted disease in the world and the main cause of genital warts, infects millions of people worldwide every year [51], with an estimated 291 million HPV-positive women worldwide in 2007 [52]. Among diseases caused by HPV, condyloma acuminata refers to an anogenital infection caused by HPV strains 6 and 11 [53,54]. Successful treatment can still be challenging. Traditional forms of treatment for condyloma acuminata that are effective at removing wart tissue including topical approaches, carbon dioxide laser (CO₂ laser), cryotherapy, or electrosurgery [55], or medical treatment with topical agents (imiquimod, podofilox, podophyllin, trichloroacetic acid, sinecatechins ointment) [56,57].

However, these therapies are often ineffective with a high recurrence rate, as they cannot eliminate subclinical latent HPV infection. Multiple reports have shown that 5-aminolaevulinic acid (ALA)-PDT can be effective in treating HPV lesions [16,58]. A real benefit of PDT lies in its ability to treat multifocal disease without tissue loss [59]. Treatment with 5-ALA-PDT can destroy the warts and cause selective and specific destruction of

subclinical virus infected areas [58]. The cure rate and viral clearance rate are significantly higher after ALA-PDT therapy compared to pharmacotherapy or physically destructive therapies. However, clinical trials have demonstrated the feasibility of applying topical ALA cream for photodiagnosis (PD) and PDT of condylomas caused by HPV. Women receiving PDT in this clinical trial had often failed with conventional treatment [56]. Conventional treatment can often be painful and sometimes disfiguring, and often results in high recurrence rates, which reinforces the need for new approaches [56,60]. One study evaluated the persistence or clearance of HPV infection after ALA-PDT in patients suffering from genital warts. The data was analyzed between January 2019 and December 2020 at Nanfang Hospital and Dermatology Department, Hospital of Southern Medical University in China, and showed that different variables such as multiple sexual partners, a history of recurrent infection, and severe pain during PDT affected the overall outcomes of PDT treatment. The authors suggested that the patients may need additional PDT sessions. Interestingly, PDT appears from this study to be effective against single strain HPV infections [61].

In one patient with condyloma acuminata covering the glans penis, a case study reported a patient who had a single large lesion. ALA-PDT was used as a therapeutic approach to reduce the risk of recurrence and minimize the trauma caused by traditional methods such as CO₂ laser therapy. The choice of therapy by ALA-PDT is dictated by the fact that ALA accumulates the warts and reduce the rate of recurrence in the surrounding tissue with subclinical infection [55]. ALA-PDT is an effective, safe and curative alternative to the conventional treatment of genital warts [55]. ALA-PDT-based treatment was also effective against urethral condyloma acuminata; in this case, the authors suggested that the dynamic monitoring of the HPV viral load could objectively demonstrate the effectiveness and guide the treatment of PDT [62]. These results were reinforced by a randomized controlled clinical trial carried out recently. The objective was to compare the use of PDT with the application of trichloroacetic acid (TAA) in the treatment of HPV condyloma in the perianal and vulval regions. A total of 16 patients was treated with PDT using the prodrug methyl aminolevulinate incubated for 3 h and irradiation at 630 nm (100 J/cm²). Fifteen patients were treated with TAA, received acid using a cotton swab. The results of these experimentations revealed that the PDT-based treatment appeared to be effective in the treatment of lesions due to the physical destruction of condyloma and subclinical lesions. A complete response rate was evaluated at 63% for PDT versus 60% (10 had a complete response and 6 had a partial response) for TAA (9 patients had a complete response in the elimination of lesions, 3 had a partial response), and a recurrence rate of 0% for PDT versus 33% for TAA. In addition, treatment with PDT led to complete clearance in an area with many warts; the authors suggested that PDT may be more beneficial for patients with recurrent HPV warts [63].

Another therapeutic strategy based on a combination of CO₂ laser and ALA-PDT was tested to treat condyloma acuminata in 98 adult patients (male and female). Firstly, patients were treated by CO₂ laser to remove the visible warts. Secondly, the patients had the ALA-PDT treatment immediately after the laser exposition. The ALA surface application was performed for 3 h at light irradiation of 100–150 J/cm². The ALA-PDT was continued once a week for three weeks. A combination of CO₂ laser and ALA-PDT has been shown to be feasible and effective in the treatment of condyloma acuminata. The cure rate was high at 93.8% (92/98) [64]. Shi et al., (2013) tested a different treatment strategy on 361 patients diagnosed with condyloma acuminata. Patients were divided into three groups according to the maximum diameter of their lesion (A < 0.5, B = 0.5–2.0, and C > 2.0–4.0 cm). Five treatments were compared in each group (cryotherapy, CO₂ laser, ALA-PDT alone, ALA-PDT plus CO₂ laser, ALA-PDT plus cryotherapy). The clinical outcomes evaluated during follow up after each treatment showed that the ALA-PDT was best if the maximum lesion diameter was <0.5 cm, while ALA-PDT plus cryotherapy was better for lesions 0.5–2.0 cm. ALA-PDT treatment, after either cryotherapy or CO₂ laser was effective for lesions >2.0–4.0 cm, which should be the first choice. They suggested that

all treatments could be effective, but the choice depended on the size of the condyloma lesion [65].

However, a study by Szeimies et al. (2009) reported a different result. CO₂ laser ablation followed by ALA-PDT was investigated in a phase III prospective randomized bicenter double-blind clinical trial to prevent recurrence of condyloma acuminata. One hundred seventy-five patients with condyloma acuminata received CO₂ laser vaporization plus adjuvant ALA-PDT or adjuvant placebo-PDT. Results showed no statistically significant difference between the groups with regard to recurrence rates up to 12 months after treatment. No major complications were observed [66].

Persistent HPV infection can lead to the development of malignant lesions in the vaginal and cervical epithelium. Indeed, HPV infection represents the main cause of cervical intraepithelial neoplasia [15]. The clinical treatments recommended for cervical intraepithelial neoplasia have already been mentioned, and mainly include the topical application of certain drugs or surgical excision, as well as irradiation by intravaginal radiotherapy or ablation by a laser [67]. PDT is a new therapeutic tool which has been used mainly in HPV infections causing condyloma acuminata, or in the case of non-melanoma skin tumors [58,68]. PDT has become a promising therapeutic method used in the treatment of various tumors including cervical intraepithelial neoplasia, cervical HPV infection, and vaginal intraepithelial neoplasia [69]. One meta-analysis revealed that out of 77 patients with cervical HPV infection included in four randomized controlled trials who received PDT, 48 of them showed complete remission. The complete remission rate ranged from 53.5 to 94.4% [69]. The authors of this meta-analysis concluded that PDT was effective for HPV clearance, particularly high-risk HPV genotypes. In addition, the study showed that out of 120 patients with cervical intraepithelial neoplasia treated by PDT, 77 patients achieved full primary remission by the end of the 3-month follow-up. Interestingly, the complete remission rate ranged from 31.3 to 100% [69]. Clinically, PDT is becoming increasingly employed to treat malignant HPV viral infection, including cervical intraepithelial neoplasia and cervical HPV infection, and has shown complete remission and local eradication of the virus. Another systematic review also demonstrated the efficacy of PDT for the treatment of cervical intraepithelial neoplasia. Analysis revealed that the complete remission rate of PDT for cervical intraepithelial neoplasia ranged from 0 to 100%, with an HPV eradication rate varying from 53.4 to 80% [70]. These results were based on analysis of the data published in several studies [71,72]. The effectiveness of PDT for the treatment of cervical intraepithelial neoplasia and for the eradication of HPV has also been confirmed in several studies by Li et al. [73], Cang et al. [74], Wu et al. [75], Su et al. [76] and Zang et al. [15]. Furthermore, the efficacy of PDT for the treatment of cervical cancer has also recently been demonstrated. At three months after PDT, complete elimination of HPV was detected in more than 90% of patients with early stage cervical cancer [19].

Herpes simplex viruses, HSV-1 and HSV-2, are the most common cause of mucocutaneous herpes lesions with a chronic or recurrent course. Approximately 20% of people infected with HSV have clinical manifestations that recur, especially during periods of weakened immunity [1]. HSV-1 is mainly transmitted by oral-to-oral contact causing oral herpes, but can also cause genital herpes. The World Health Organization has estimated that 3.7 billion people under age 50 (67%) have HSV-1 infection globally. HSV-2 is a sexually transmitted infection that causes genital herpes and has been estimated to affect 491 million people aged 15–49 (13%) worldwide [77]. HSV infections cause discomfort (itching, pain) associated with the eruption of vesicles and development of erosions. Lesion recurrence and aesthetic discomfort significantly reduce patient quality of life [78]. To improve the quality of life of women affected by genital HSV, several therapeutic approaches are being developed. Conventional therapy with nucleotide analogue antiviral drugs inhibits viral replication and shortens the duration of symptoms, but does not prevent recurrence. PDT is selective, non-invasive, not harmful to the patient, and can be used in parallel with other therapies, even in immunocompromised subjects (e.g., transplantation or oncology patients). Recent studies have shown the effectiveness of PDT in the inactivation of dif-

ferent types of virus in vitro and in vivo [79]. The HSV strains VR-3 and MS were used to infect Vero cell cultures and the antiherpetic effect was evaluated after PDT and laser irradiation treatments (Figure 1A). The results showed a significant reduction in virus load (100–1000 times) [79]. Nevertheless, a study determining the effect of ALA-PDT on the recurrence of herpes simplex showed that one patient irradiated with a higher dose of light (630 ± 20 nm; 120 J/cm^2) developed an acute inflammation, accompanied by the appearance of prolonged scabs [80].

Published case reports regarding the treatment of recurrent herpes with phototherapy (including lasers) sometimes combined with synthetic dyes have focused mainly on reducing the duration of HSV infection symptoms and its accompanying pain, reducing viral titers, and the acceleration of wound healing, in cases of herpes labialis and genital herpes [80]. Donnarumma et al. demonstrated that laser phototherapy applied in HSV infection could also act on the immune response, restoring the expression of proinflammatory cytokines, tumor necrosis factor α (TNF α), interleukin (IL)-1 β , and IL-6 suppressed by the virus, and limiting the viral spread from cell to cell [81]. A case report by Ferreira et al., (2011) used low-level laser therapy on a patient (50-year-old heterosexual female), with recurrent episodes of labial herpes over the preceding 5 years. Follow up showed that the patient remained symptom free for 17 months. Low-level laser therapy showed promising clinical results as a long-lasting suppression therapy in a single patient, but its wider efficacy for long-term suppression has yet to be established [82]. HSV infection can cause hyperpigmentation of the skin which may be present even when the infection is latent. Hyperpigmentation is linked to an inflammatory process in the tissues. Staining can cause people aesthetic inconvenience with a sense of shame. Hyperpigmentation can be treated topically with alpha-hydroxy acids (AHAs), hydroquinone, azelaic acid, retinoic acid, ascorbic acid, kojic acid can be effective alone or in combination with others therapies such as phototherapy or laser therapy [83]. A multitude of different lasers are currently available for the treatment of cutaneous hyperpigmentation. Five major classes of dermatological lasers are currently used: ablative and non-ablative lasers in their fractionated and non-fractionated forms as well as radio frequency technologies. Non-ablative lasers are gentler on the skin and allow faster healing, while harsher ablative lasers tend to be more effective. Fractionating increases the number of treatments but minimizes downtime and complications [84]. In addition, given the absorption spectrum of melanin (250–1200 nm), visible and near-infrared lasers can be used successfully to target excess melanin in the skin in the case of postinflammatory hyperpigmentation [85]. The Q-switched Nd-YAG laser, the fractional thulium laser (via tranexamic acid) and the picosecond alexandrite laser are the most used in the treatment of hyperpigmentation of the skin [86,87].

3. PT for Bacterial STIs

Neisseria gonorrhoeae (also known as gonococcus) is a Gram-negative bacterial species that can be sexually transmitted. *N. gonorrhoeae* infects the urogenital tract, causing dysuria with a penile discharge in men, and a vaginal mucopurulent discharge, along with severe pelvic pain in women [88]. Antimicrobial resistance is emerging, which limits antibiotic treatment for *N. gonorrhoeae*. Novel treatment strategies are urgently needed, according to Klausner et al. (2021) [89].

To combat antibiotic resistance in sexually transmitted bacteria, PDT has been tested against *N. gonorrhoeae*, including antibiotic-resistant strains cocultured with human vaginal epithelial cells in vitro. No PDT-induced genotoxicity to vaginal epithelial cells was observed upon delivering a sufficient radiant exposure to inactivate *N. gonorrhoeae*. PDT effectively inactivated *N. gonorrhoeae*, which had attached and invaded vaginal epithelial cells in their co-culture model [90]. In addition, studies have demonstrated that blue light (405 nm) used alone has an intrinsic antimicrobial activity against *N. gonorrhoeae*. This wavelength of light inactivated *N. gonorrhoeae* cells, including antibiotic-resistant strains, without causing epithelial cell cytotoxicity. Blue light was still effective against these bacteria after 15 successive cycles of exposure [90,91].

The use of water-filtered infrared A (wIRA), a short wavelength band of infrared radiation with a spectrum from 780–1400 nm, which is combined with visible light VIS (wIRA/VIS) showed efficacy in killing intracellular and extracellular *Chlamydia* strains (*C. pecorum*, *C. trachomatis* serovar E) in two different cell lines (Vero and HeLa). Irradiation of the infected cells (HeLa and Vero) neither affected cell viability nor induced any molecular markers of cytotoxicity. However, a single exposure to wIRA/VIS at 40 h post infection (hpi) led to a significant reduction in the frequency of *C. pecorum* incorporation in Vero cells, and *C. trachomatis* in HeLa cells (Figure 1B). Three sessions of irradiation (24, 36, 40 hpi) during the course of *C. trachomatis* infection further reduced the chlamydial incorporation frequency in HeLa cells [92].

In another study carried out in vitro, Wasson et al. (2021) investigated the effects of visible light irradiation (405 and 670 nm) using light emitting diodes (LEDs) on *Chlamydial* growth in HeLa cells (Figure 1B). The results demonstrated a significant dose-dependent inhibitory effect and a diminished bacterial load during both active and persistent infection, following irradiation [93].

4. PT for Parasitic STIs

Trichomoniasis is a condition caused by *Trichomonas vaginalis*, and is a very common vaginal infection that, in some women, manifests as a vaginal discharge, with a disagreeable smell and a yellowish or greenish color. *T. vaginalis* is usually transmitted sexually [94]. Recently, a new diagnostic approach based on an optical nano-genosensor was designed by conjugation of gold nanoparticles to a specific oligonucleotide that recognized a *T. vaginalis* gene sequence (AuNP-probe) for specific and sensitive PCR diagnosis. An investigation was performed using the AuNP-probe with different concentrations of a synthetic complementary sequence as a standard for *T. vaginalis*. Complete hybridization was detected by adding acid to the medium and observing the changes in the color and the spectroscopic absorption spectrum. The results confirmed the accurate function of the genosensor for the detection of *T. vaginalis* in clinical samples (Figure 1C). This new diagnosis strategy used a photo-genosensor for *T. vaginalis* detection [95]. PDT has also been used for *T. vaginalis* inactivation. The trophozoites (JT and CDC 085 strains) were exposed to PDT using methylene blue as a photosensitizer. The degree of parasite inhibition was significant, with $80.21\% \pm 7.11$ for the JT strain and $91.13\% \pm 2.31$ for the CDC 085 strain. This study confirmed that PDT using methylene blue could inhibit parasite multiplication and therefore could possibly reduce infection [96].

5. PDT for Fungal STIs

There was a pilot clinical trial carried out by Du et al. (2021), in which they investigated the effectiveness of a PDT-based treatment on 21 adult AIDS patients with *C. albicans* oral candidiasis. After two consecutive PDT treatments the clinical symptoms of oral candidiasis in adult AIDS patients were improved [97]. This success suggests that PDT might also be effective in genital thrush caused by *C. albicans*. In addition, one case report has shown that treatment with methylene blue (660 nm red laser) for PDT of vulvovaginal candidiasis significantly decreased fungal infection in the vaginal canal. This report claimed that this therapy improved the quality of life of patients who reported a reduction in symptoms [98]. Moreover, antifungal blue light therapy (400–470 nm) has been reported to be effective in decreasing vaginal candidiasis. A viability study using both *C. albicans* and human vaginal epithelial cells was performed. These two models were irradiated at different wavelength ranges of 405, 415 and 450 nm. The experimental data showed that an inhibition rate of approximately 80% of *C. albicans* was achieved, while the epithelial cells had a survival rate which varied according to the wavelength: 0.6700, 0.7748, and 0.6027, respectively, for treatment with light of wavelength 405, 415 and 450 nm. Additionally, 415 nm light showed a more effective antifungal effect with less damage to epithelial cells compared to 405 nm or 450 nm light [99].

The efficacy of PDT was studied against yeast cells in a mouse model of vaginal infection. PDT was carried out using two photosensitizers, methylene blue and protoporphyrin IX. The study was carried out on mice with persistent vaginitis evoked by an intravaginal inoculation of *C. albicans*. PDT was performed 5 days after fungal inoculation using both photosensitizers. The irradiation was carried out using two custom-made LED devices at 660 and 630 nm. The results showed that PDT reduced fungal colony-forming units. After a follow-up time, the results did not change and the colonies did not increase from the initial value immediately after PDT. The use of PDT as a therapy to reduce fungal infection in a model of vaginal candidiasis produced a significant reduction in *C. albicans* while causing no damage to the vaginal mucosa [100]. Another mouse model study testing the effects of PDT on vaginal candidiasis using methylene blue and red (660 nm) laser irradiation, demonstrated that PDT significantly reduced *C. albicans* colonies. In addition, this study also demonstrated that the percentage of inflammatory cells per unit surface area was significantly reduced after only two sessions of PDT [101].

Table 1 summarizes the published reports of PT or PDT to treat a variety of STIs, either in vitro, in animal models, or in humans.

Table 1. Selected studies of PT or PDT treatment for sexually transmitted infections.

Type of Disease	Type of Photosensitizer	Light Source	Light Parameters	References
Condyloma acuminata (penis)	5-ALA	Helium-neon laser	Wavelength 635 nm Power density 100 mW/cm ²	[55]
Genital warts and Subclinical Human papilloma virus (HPV)	5-aminolevulinic acid (ALA)	Carbon diode (CO ₂) laser	Wavelength 635 nm Fluence 100 J/cm ² Power density 100 mW/cm ²	[61]
Condyloma acuminata (urethral)	5-ALA	Semiconductor laser	Wavelength 635 nm Power density 100 mW/cm ² Fluence 100 J/cm ²	[62]
HPV Condyloma (perianal and vulval regions)	Methyl aminolevulinate	LEDs	Wavelength 630 nm Power density 80 mW/cm ² Fluence 100 J/cm ²	[63]
Condyloma acuminata	5-ALA	CO ₂ laser light	Fluence 100–150 J/cm ² Power density 60–100 mW/cm ²	[64]
Condyloma acuminata	5-ALA	Cylindrical laser fiber High energy narrow-band red light therapy equipment	Urethral meatus with a spot size <3 cm: Wavelength 630 ± 5 nm Fluence 100–150 J/cm ² Power density 150–300 mW/cm ² Size ≥ 3 cm Wavelength 633 ± 5 nm Fluence 105 J/cm ²	[65]

Table 1. Cont.

Type of Disease	Type of Photosensitizer	Light Source	Light Parameters	References
Condyloma acuminata	5-ALA	CO ₂ laser red light	Wavelength 600–740 nm Fluence 100 J/cm ² Power density 100 mW/cm ²	[66]
Cervical intraepithelial neoplasia	Polyhematoporphyrin ether/ester	YAG-OPO laser (laser pumped optical parametric oscillator)	Wavelength 630 nm Fluence 100 J/cm ²	[102]
Cervical intraepithelial neoplasia	5-ALA	Thermal light source emitting a broadband red light Illuminate the cervical canal	150 W halogen lamp Power density 90 mW/cm ² Fluence 100 J/cm ² Power density 300 mW/cm ² Fluence 50 J/cm ²	[103]
Cervical intraepithelial neoplasia	Hexaminolevulinate (HAL)	Red coherent laser and a special light catheter	Wavelength 633 nm	[104]
Cervical intraepithelial neoplasia	Photofrin	Excimer dye laser Or YAG-OPO laser	Wavelength 630 nm Fluence 100 J/cm ²	[105]
Cervical intraepithelial neoplasia	Photolon (a combination of chlorin e6 potassium salt and low-weight polyvinylpyrrolidone)	Therapeutic laser device “LD680-2000”	Wavelength 670 nm Power density 200 mW/cm ² Fluence 100 J/cm ²	[106]
Cervical intraepithelial neoplasia	5-ALA	Semiconductor laser	Wavelength 635 nm Power density 100–150 mW/cm ² Fluence 100 J/cm ²	[72]
Cervical intraepithelial neoplasia combined with high-risk HPV	5-ALA	Red laser	Wavelength 633 nm Fluence 80 J/cm ²	[73]
High-risk HPV without cervical lesions	5-ALA	Cylindrical semiconductor laser fiber	Wavelength 635 nm Power density 100 mW/cm ² Fluence 100 J/cm ²	[74]
Cervical intraepithelial neoplasia combined with high-risk HPV	5-ALA	LED	Wavelength 635 nm Fluence 100 J/cm ²	[75]
Cervical intraepithelial neoplasia with vaginal intraepithelial neoplasia	5-ALA	LED optical fibers Semiconductor laser	Wavelength 635 nm Fluence 80–120 J/cm ²	[76]

Table 1. Cont.

Type of Disease	Type of Photosensitizer	Light Source	Light Parameters	References
Early stage cervical cancer	Photoran E6 Fotoditazin	LED Flexible cylindrical diffuser	Wavelength 400 nm Power 1–1.2 W Fluence 400 J/cm ²	[19]
HSV (HSV-1 (VR-3 strain) and HSV-2 (MS strain)) infected Vero cell culture	Fotoditazin preparation (chlorin E6 derivative)	NI	NI	[79]
Genital and oral herpes	5-ALA	Red light from a halogen lamp	Wavelength 630 ± 20 nm Power density 100 mW/cm ² Fluence 120 J/cm ²	[80]
HSV-I strain infected human epithelial cell	NI	Diode laser	Wavelength 830 nm	[81]
Hyperemic lesions labial herpes	NI	Low intensity red laser Direct machining control (DMC) Photon Laser II	Wavelength 660 nm Power density 100 mW/cm ² Fluence 30 J/cm ²	[82]
<i>Neisseria gonorrhoeae</i> (<i>N.</i> <i>gonorrhoeae</i>) (ATCC 700825) 4 clinical <i>N. gonorrhoeae</i> isolates	NI	LED	Wavelength 405 nm Power density 60 mW/cm ²	[90]
<i>N. gonorrhoeae</i> (ATCC 700825) one multidrug-resistant clinical strain of <i>N. gonorrhoeae</i>	NI	Blue light	Wavelength 405 nm Fluence 54 J/cm ²	[91]
<i>N. gonorrhoeae</i> (ATCC 700825)	NI	LED	Wavelength 405 nm, 470 nm Power density 60 mW/cm ²	[107]
<i>Chlamydia</i> (<i>C.</i>) <i>pecorum</i> 1710S <i>C. trachomatis</i> serovar E	NI	Water-filtered infrared A combined with visible light (wIRA/VIS)	Wavelengths 380 nm up to 1400 nm Power density 3700 W/m ²	[92]
<i>C. trachomatis</i> serovar E	NI	Warfighter Accelerated Recovery by Photobiomodulation (WARP) 10 LED	Wavelengths 405 nm, 670 nm Power density 60 mW/cm ² Fluence 5 J/cm ²	[93]
<i>Trichomonas vaginalis</i> CDC 085 strain (ATCC 50143)	Methylene blue	LED monochromatic light source	Wavelength 630 nm Power 300 mW	[96]
HIV / AIDS, co-infected with <i>Candida</i> (<i>C.</i>) <i>albicans</i> in the oral cavity	Methylene blue	LED	Wavelength 633 nm Power density 20.72 mW/cm ² Fluence 37.29 J/cm ²	[97]

Table 1. Cont.

Type of Disease	Type of Photosensitizer	Light Source	Light Parameters	References
Vulvovaginal candidiasis (<i>C. albicans</i>)	Methylene blue	PDT using the MAC Scar Acceleration Method Red laser	Wavelength 660 nm Power 100 mW	[98]
<i>C. albicans</i>	NI	Blue LED light sources	Wavelength 415 nm Power density 50 mW/cm ²	[99]
Vaginal candidiasis <i>C. albicans</i> (mouse model)	Methylene blue and protoporphyrin IX	LEDs	Wavelengths 660 nm, 630 nm Power 800 mW	[100]
<i>C. albicans</i> (mouse model)	Methylene blue	Red laser	Wavelength 660 nm Power density 100 mW/cm ² Fluence 18 J/cm ² , 36 J/cm ²	[101]

6. Conclusions

So far, despite the range of treatment options available, no light-based approach has been able to achieve satisfactory results against all pathogens that cause STIs. In addition, to date and to our knowledge, there are no randomized controlled clinical trials evaluating the efficacy of phototherapy-based treatment against each pathogen causing a specific infection and comparing this efficacy to a reference treatment. Nevertheless, controlled trials comparing treatments are currently being optimized. In the case of HPV infection, no current PDT-based treatment completely eradicates the HPV virus. However, the variables that could be considered include (but are not limited to) the morphology of the lesions such as thickness and size, quantity, anatomic location, and HPV strain. The viral replication cycle and stage of infection appear to play a role in the response of the infection. It would be difficult at present for this type of treatment to be implicated as a specific mode of treatment (e.g., phototherapy) and could cover a wide variety of biologically unrelated pathogens that belonged only to STIs. PDT approaches should be further developed against genital HSV infection, and against STIs with bacterial and parasitic causes (*C. trachomatis*, *N. gonorrhoeae*, *Treponema pallidum*, *Mycoplasma genitalium* and *T. vaginalis*). The development of nanotechnology over the past decades has led to the incorporation of nanomedicine into PT for cancer treatment. The merging of nanomedicine into PT has allowed the continuous refinement of PDT approaches. With the careful design of phototherapeutic agents and good control of light illumination at the site of the lesions, effective PDT can be achieved, with reductions in the systemic toxicity associated with traditional chemotherapy and radiotherapy. However, we hope that this localized therapy could also be used in the case of STIs, especially in view of the rise in drug resistance observed in most types of pathogens.

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Abbreviations

HPV: human papilloma virus; 5-ALA: 5-aminolevulinic acid; CO₂ laser: carbon diode laser; LED: light-emitting diode; YAG-OPO laser: laser pumped optical parametric oscillator; HAL: hexaminolevulinate; PDT: photodynamic therapy; HSV: virus herpes simplex; DMC: direct machining control; *N. gonorrhoeae*: *Neisseria gonorrhoeae*; Chlamydia pecorum: *C. pecorum*; *Chlamydia trachomatis*: *C. trachomatis*; wIRA/VIS: water-filtered infrared A combined with visible light; WARP: Warfighter Accelerated Recovery by Photobiomodulation; HIV/AIDS: human immunodeficiency virus/acquired immunodeficiency syndrome; *C. albicans*: *Candida albicans*; MAC: scar acceleration method. NI: not indicated.

References

1. Sexually Transmitted Infections (STIs). Available online: <https://www.who.int/westernpacific/health-topics/sexually-transmitted-infections> (accessed on 19 November 2021).
2. Williamson, D.A.; Chen, M.Y. Emerging and Reemerging Sexually Transmitted Infections. *N. Engl. J. Med.* **2020**, *382*, 2023–2032. [CrossRef] [PubMed]
3. Smith, L.; Angarone, M.P. Sexually Transmitted Infections. *Urol. Clin. N. Am.* **2015**, *42*, 507–518. [CrossRef] [PubMed]
4. Ciccicarese, G.; Herzum, A.; Pastorino, A.; Dezzana, M.; Casazza, S.; Mavilia, M.G.; Copello, F.; Parodi, A.; Drago, F. Prevalence of Genital HPV Infection in STI and Healthy Populations and Risk Factors for Viral Persistence. *Eur. J. Clin. Microbiol. Infect. Dis.* **2021**, *40*, 885–888. [CrossRef] [PubMed]
5. Cole, S. Herpes Simplex Virus: Epidemiology, Diagnosis, and Treatment. *Nurs. Clin. N. Am.* **2020**, *55*, 337–345. [CrossRef]
6. Kardani, K.; Basimi, P.; Fekri, M.; Bolhassani, A. Antiviral Therapy for the Sexually Transmitted Viruses: Recent Updates on Vaccine Development. *Expert Rev. Clin. Pharmacol.* **2020**, *13*, 1001–1046. [CrossRef]
7. Juliana, N.C.A.; Deb, S.; Ouburg, S.; Chauhan, A.; Pleijster, J.; Ali, S.M.; Morré, S.A.; Sazawal, S.; Ambrosino, E. The Prevalence of Chlamydia Trachomatis and Three Other Non-Viral Sexually Transmitted Infections among Pregnant Women in Pemba Island Tanzania. *Pathogens* **2020**, *9*, 625. [CrossRef]
8. Gannon-Loew, K.E.; Holland-Hall, C. A Review of Current Guidelines and Research on the Management of Sexually Transmitted Infections in Adolescents and Young Adults. *Ther. Adv. Infect. Dis.* **2020**, *7*, 2049936120960664. [CrossRef]
9. Doernberg, S.B.; Komarow, L.; Tran, T.T.T.; Sund, Z.; Pandori, M.W.; Jensen, D.; Tsalik, E.L.; Deal, C.D.; Chambers, H.F.; Fowler, V.G.; et al. Simultaneous Evaluation of Diagnostic Assays for Pharyngeal and Rectal *Neisseria Gonorrhoeae* and Chlamydia Trachomatis Using a Master Protocol. *Clin. Infect. Dis.* **2020**, *71*, 2314–2322. [CrossRef]
10. Yarbrough, M.L.; Burnham, C.-A.D. The ABCs of STIs: An Update on Sexually Transmitted Infections. *Clin. Chem.* **2016**, *62*, 811–823. [CrossRef]
11. Lin, K.-Y.; Sun, H.-Y.; Lee, T.-F.; Chuang, Y.-C.; Wu, U.-I.; Liu, W.-C.; Chang, S.-Y.; Chen, Y.-J.; Hung, C.-C.; Chang, S.-C. High Prevalence of Sexually Transmitted Coinfections among At-Risk People Living with HIV. *J. Formos. Med. Assoc.* **2021**, *120*, 1876–1883. [CrossRef]
12. Tien, V.; Punjabi, C.; Holubar, M.K. Antimicrobial Resistance in Sexually Transmitted Infections. *J. Travel. Med.* **2020**, *27*, taz101. [CrossRef]
13. Williams, E.; Fairley, C.K.; Williamson, D. Novel Strategies for Prevention and Treatment of Antimicrobial Resistance in Sexually-Transmitted Infections. *Curr. Opin. Infect. Dis.* **2021**, *34*, 591–598. [CrossRef]

14. Li, Y.; Yu, T.; Yan, H.; Li, D.; Yu, T.; Yuan, T.; Rahaman, A.; Ali, S.; Abbas, F.; Dian, Z.; et al. Vaginal Microbiota and HPV Infection: Novel Mechanistic Insights and Therapeutic Strategies. *Infect. Drug Resist.* **2020**, *13*, 1213–1220. [\[CrossRef\]](#)
15. Zhang, T.; Hu, R.; Tang, Y.; Zhang, Y.; Qin, L.; Shen, Y.; Wang, B.; Zhang, L.; Cao, L.; Zhou, Y.; et al. The Effect of Local Photodynamic Therapy with 5-Aminolevulinic Acid in the Treatment of Vaginal Intraepithelial Lesions with High-Risk HPV Infection. *Photodiagn. Photodyn. Ther.* **2022**, *37*, 102728. [\[CrossRef\]](#)
16. Fathi, R.; Tsoukas, M.M. Genital Warts and Other HPV Infections: Established and Novel Therapies. *Clin. Dermatol.* **2014**, *32*, 299–306. [\[CrossRef\]](#)
17. Schnürch, H.-G.; Ackermann, S.; Alt-Radtke, C.D.; Angleitner, L.; Barinoff, J.; Beckmann, M.W.; Böing, C.; Dannecker, C.; Fehm, T.; Gaase, R.; et al. Diagnosis, Therapy and Follow-up of Vaginal Cancer and Its Precursors. Guideline of the DGGG and the DKG (S2k-Level, AWMF Registry No. 032/042, October 2018). *Geburtshilfe Und Frauenheilkd.* **2019**, *79*, 1060–1078. [\[CrossRef\]](#)
18. Chumworathayi, B.; Thinkhamrop, J.; Blumenthal, P.D.; Thinkhamrop, B.; Pientong, C.; Ekalaksananan, T. Cryotherapy for HPV Clearance in Women with Biopsy-Confirmed Cervical Low-Grade Squamous Intraepithelial Lesions. *Int. J. Gynaecol. Obstet.* **2010**, *108*, 119–122. [\[CrossRef\]](#)
19. Afanasiev, M.D.D.S.M.S.; Dushkin, M.D.A.D.; Grishacheva, D.S.T.G.; Afanasiev, M.D.D.S.S.S.; Karaulov Academician Ras, M.D.D.S.A.V. Photodynamic Therapy for Early-Stage Cervical Cancer Treatment. *Photodiagn. Photodyn. Ther.* **2021**, *37*, 102620. [\[CrossRef\]](#)
20. Viviano, M.; Kenfack, B.; Catarino, R.; Tincho, E.; Temogne, L.; Benski, A.-C.; Tebeu, P.-M.; Meyer-Hamme, U.; Vassilakos, P.; Petignat, P. Feasibility of Thermocoagulation in a Screen-and-Treat Approach for the Treatment of Cervical Precancerous Lesions in Sub-Saharan Africa. *BMC Womens Health* **2017**, *17*, 2. [\[CrossRef\]](#)
21. Gutierrez, P.; Garza, J.; Gandhi, K.; Voice, A.; Stout, E.; Ventolini, G. Carbon Dioxide (CO₂) Laser Ablation Treatment of a Peri-Urethral Genital Wart: A Case Report. *Case Rep. Womens Health* **2020**, *27*, e00226. [\[CrossRef\]](#)
22. Cospes, P.F.; McNair, C.; González, I.; Wong, N.; Knudsen, K.E.; Chen, J.J.; Markovina, S.; Schwarz, J.K.; Grigsby, P.W.; Wang, X. Decreased Local Immune Response and Retained HPV Gene Expression during Chemoradiotherapy Are Associated with Treatment Resistance and Death from Cervical Cancer. *Int. J. Cancer* **2020**, *146*, 2047–2058. [\[CrossRef\]](#)
23. Foster, C.C.; Lee, A.Y.; Furtado, L.V.; Hart, J.; Alpert, L.; Xiao, S.-Y.; Hyman, N.H.; Sharma, M.R.; Liauw, S.L. Treatment Outcomes and HPV Characteristics for an Institutional Cohort of Patients with Anal Cancer Receiving Concurrent Chemotherapy and Intensity-Modulated Radiation Therapy. *PLoS ONE* **2018**, *13*, e0194234. [\[CrossRef\]](#)
24. Chera, B.S.; Amdur, R.J.; Green, R.; Shen, C.; Gupta, G.; Tan, X.; Knowles, M.; Fried, D.; Hayes, N.; Weiss, J.; et al. Phase II Trial of De-Intensified Chemoradiotherapy for Human Papillomavirus-Associated Oropharyngeal Squamous Cell Carcinoma. *J. Clin. Oncol.* **2019**, *37*, 2661–2669. [\[CrossRef\]](#)
25. Sindhu, S.K.; Bauman, J.E. Current Concepts in Chemotherapy for Head and Neck Cancer. *Oral. Maxillofac. Surg. Clin. N. Am.* **2019**, *31*, 145–154. [\[CrossRef\]](#)
26. Bonomi, M.; Ahmed, T.; Warner, D.; Waltonen, J.; Sullivan, C.; Porosnicu, M.; Batt, K.; Ruiz, J.; Cappellari, J. Human Papillomavirus-Related Small Cell Carcinoma of the Oropharynx: A Case Report and Literature Review. *Cancers Head Neck* **2017**, *2*, 3. [\[CrossRef\]](#) [\[PubMed\]](#)
27. Perrone, A.M.; Tesei, M.; Ferioli, M.; De Terlizzi, F.; Della Gatta, A.N.; Boussedra, S.; Dondi, G.; Galuppi, A.; Morganti, A.G.; De Iaco, P. Results of a Phase I-II Study on Laser Therapy for Vaginal Side Effects after Radiotherapy for Cancer of Uterine Cervix or Endometrium. *Cancers* **2020**, *12*, 1639. [\[CrossRef\]](#) [\[PubMed\]](#)
28. Bogani, G.; Ditto, A.; Martinelli, F.; Mosca, L.; Chiappa, V.; Rossetti, D.; Leone Roberti Maggiore, U.; Sabatucci, I.; Lorusso, D.; Raspagliesi, F. LASER Treatment for Women with High-Grade Vaginal Intraepithelial Neoplasia: A Propensity-Matched Analysis on the Efficacy of Ablative versus Excisional Procedures. *Lasers Surg. Med.* **2018**, *50*, 933–939. [\[CrossRef\]](#) [\[PubMed\]](#)
29. Boonlikit, S. Recurrence of High-Grade Vaginal Intraepithelial Neoplasia after Various Treatments. *Curr. Probl. Cancer* **2022**, *46*, 100792. [\[CrossRef\]](#) [\[PubMed\]](#)
30. Farmer, E.; Cheng, M.A.; Hung, C.-F.; Wu, T.-C. Vaccination Strategies for the Control and Treatment of HPV Infection and HPV-Associated Cancer. *Recent Results Cancer Res.* **2021**, *217*, 157–195. [\[CrossRef\]](#)
31. Dadar, M.; Chakraborty, S.; Dhama, K.; Prasad, M.; Khandia, R.; Hassan, S.; Munjal, A.; Tiwari, R.; Karthik, K.; Kumar, D.; et al. Advances in Designing and Developing Vaccines, Drugs and Therapeutic Approaches to Counter Human Papilloma Virus. *Front. Immunol.* **2018**, *9*, 2478. [\[CrossRef\]](#)
32. Taylor, M.; Gerriets, V. Acyclovir. In *StatPearls*; StatPearls Publishing: Treasure Island, FL, USA, 2021.
33. Feng, E.; Balint, E.; Vahedi, F.; Ashkar, A.A. Immunoregulatory Functions of Interferons during Genital HSV-2 Infection. *Front. Immunol.* **2021**, *12*, 724618. [\[CrossRef\]](#)
34. Lau, A.; Kong, F.Y.S.; Fairley, C.K.; Templeton, D.J.; Amin, J.; Phillips, S.; Law, M.; Chen, M.Y.; Bradshaw, C.S.; Donovan, B.; et al. Azithromycin or Doxycycline for Asymptomatic Rectal Chlamydia Trachomatis. *N. Engl. J. Med.* **2021**, *384*, 2418–2427. [\[CrossRef\]](#)
35. Zofkie, A.C.; Fomina, Y.Y.; Roberts, S.W.; McIntire, D.D.; Nelson, D.B.; Adhikari, E.H. Effectiveness of Chlamydia Trachomatis Expedited Partner Therapy in Pregnancy. *Am. J. Obstet. Gynecol.* **2021**, *225*, 325.e1–325.e7. [\[CrossRef\]](#)
36. Adachi, K.N.; Nielsen-Saines, K.; Klausner, J.D. Chlamydia Trachomatis Screening and Treatment in Pregnancy to Reduce Adverse Pregnancy and Neonatal Outcomes: A Review. *Front. Public Health* **2021**, *9*, 531073. [\[CrossRef\]](#)
37. Gnanadurai, R.; Fifer, H. Mycoplasma Genitalium: A Review. *Microbiology* **2020**, *166*, 21–29. [\[CrossRef\]](#)

38. Unemo, M.; Lahra, M.M.; Cole, M.; Galarza, P.; Ndowa, F.; Martin, I.; Dillon, J.-A.R.; Ramon-Pardo, P.; Bolan, G.; Wi, T. World Health Organization Global Gonococcal Antimicrobial Surveillance Program (WHO GASP): Review of New Data and Evidence to Inform International Collaborative Actions and Research Efforts. *Sex. Health* **2019**, *16*, 412–425. [\[CrossRef\]](#)
39. Poncin, T.; Fouere, S.; Braille, A.; Camelena, F.; Agsous, M.; Bebear, C.; Kumanski, S.; Lot, F.; Mercier-Delarue, S.; Ngangro, N.N.; et al. Multidrug-Resistant *Neisseria Gonorrhoeae* Failing Treatment with Ceftriaxone and Doxycycline in France, November 2017. *Eurosurveillance* **2018**, *23*, 1800264. [\[CrossRef\]](#)
40. Salmerón, P.; Viñado, B.; Arando, M.; Alcoceba, E.; Romero, B.; Menéndez, B.; Bernal, S.; Idigoras, P.; Colomina, J.; Martin-Saco, G.; et al. *Neisseria Gonorrhoeae* Antimicrobial Resistance in Spain: A Prospective Multicentre Study. *J. Antimicrob. Chemother.* **2021**, *76*, 1523–1531. [\[CrossRef\]](#)
41. Bouchemal, K.; Bories, C.; Loiseau, P.M. Strategies for Prevention and Treatment of *Trichomonas Vaginalis* Infections. *Clin. Microbiol. Rev.* **2017**, *30*, 811–825. [\[CrossRef\]](#)
42. Graves, K.J.; Novak, J.; Secor, W.E.; Kissinger, P.J.; Schwebke, J.R.; Muzny, C.A. A Systematic Review of the Literature on Mechanisms of 5-Nitroimidazole Resistance in *Trichomonas Vaginalis*. *Parasitology* **2020**, *147*, 1383–1391. [\[CrossRef\]](#)
43. Lam, S. Photodynamic Therapy of Lung Cancer. *Semin. Oncol.* **1994**, *21*, 15–19. [\[CrossRef\]](#)
44. Kessel, D. Photodynamic Therapy: Critical PDT Theory. *Photochem. Photobiol.* **2022**. [\[CrossRef\]](#)
45. Dougherty, T.J. Hematoporphyrin Derivative for Detection and Treatment of Cancer. *J. Surg. Oncol.* **1980**, *15*, 209–210. [\[CrossRef\]](#)
46. Kou, J.; Dou, D.; Yang, L. Porphyrin Photosensitizers in Photodynamic Therapy and Its Applications. *Oncotarget* **2017**, *8*, 81591–81603. [\[CrossRef\]](#)
47. Zhang, J.; Jiang, C.; Figueiró Longo, J.P.; Azevedo, R.B.; Zhang, H.; Muehlmann, L.A. An Updated Overview on the Development of New Photosensitizers for Anticancer Photodynamic Therapy. *Acta. Pharm. Sin. B* **2018**, *8*, 137–146. [\[CrossRef\]](#)
48. Bellnier, D.A.; Greco, W.R.; Loewen, G.M.; Nava, H.; Oseroff, A.R.; Dougherty, T.J. Clinical Pharmacokinetics of the PDT Photosensitizers Porfimer Sodium (Photofrin), 2-[1-Hexyloxyethyl]-2-Deviny Pyropheophorbide-a (Photochlor) and 5-ALA-Induced Protoporphyrin IX. *Lasers Surg. Med.* **2006**, *38*, 439–444. [\[CrossRef\]](#)
49. Mfouo-Tynga, I.S.; Dias, L.D.; Inada, N.M.; Kurachi, C. Features of Third Generation Photosensitizers Used in Anticancer Photodynamic Therapy: Review. *Photodiagn. Photodyn. Ther.* **2021**, *34*, 102091. [\[CrossRef\]](#)
50. Kharkwal, G.B.; Sharma, S.K.; Huang, Y.-Y.; Dai, T.; Hamblin, M.R. Photodynamic Therapy for Infections: Clinical Applications. *Lasers Surg. Med.* **2011**, *43*, 755–767. [\[CrossRef\]](#)
51. Kombe Kombe, A.J.; Li, B.; Zahid, A.; Mengist, H.M.; Bounda, G.-A.; Zhou, Y.; Jin, T. Epidemiology and Burden of Human Papillomavirus and Related Diseases, Molecular Pathogenesis, and Vaccine Evaluation. *Front. Public Health* **2020**, *8*, 552028. [\[CrossRef\]](#)
52. De Sanjosé, S.; Diaz, M.; Castellsagué, X.; Clifford, G.; Bruni, L.; Muñoz, N.; Bosch, F.X. Worldwide Prevalence and Genotype Distribution of Cervical Human Papillomavirus DNA in Women with Normal Cytology: A Meta-Analysis. *Lancet Infect. Dis.* **2007**, *7*, 453–459. [\[CrossRef\]](#)
53. Pennycook, K.B.; McCreedy, T.A. Condyloma Acuminata. In *StatPearls*; StatPearls Publishing: Treasure Island, FL, USA, 2021.
54. Hu, Z.; Zheng, H.; Zeng, K. Patterns of Multiple Human Papillomavirus Clearance during 5-Aminolevulinic Acid-Based Photodynamic Therapy in Patients with Genital Warts. *Photodiagn. Photodyn. Ther.* **2021**, *35*, 102454. [\[CrossRef\]](#) [\[PubMed\]](#)
55. Yin, G.; Zhang, Y.; Geng, M.; Cai, B.; Zheng, Y. Cure of Condyloma Acuminata Covering the Glans Penis Using Aminolevulinic Acid/Photodynamic Therapy. *Photodiagn. Photodyn. Ther.* **2020**, *30*, 101658. [\[CrossRef\]](#) [\[PubMed\]](#)
56. Inada, N.M.; Buzza, H.H.; Carbinatto, F.M.; Blanco, K.C.; de Andrade, C.T.; Vollet-Filho, J.D.; Bagnato, V.S.; Allison, R.R. Optical Techniques for the Diagnosis and Treatment of Lesions Induced by the Human Papillomavirus—A Resource Letter. *Photodiagn. Photodyn. Ther.* **2018**, *23*, 106–110. [\[CrossRef\]](#) [\[PubMed\]](#)
57. Owczarek, W.; Slowinska, M.; Walecka, I.; Ciazynska, M.; Nowicka, D.; Walczak, L.; Paluchowska, E. Correlation of the ALA-PDT Treatment Efficacy and the HPV Genotype Profile of Genital Warts after Cryotherapy Failure and Podophyllotoxin Therapy in Male Patients. *Life* **2021**, *11*, 146. [\[CrossRef\]](#)
58. Kechichian, E.; Helou, E.; Sarkis, J.; Hayek, C.; Labaki, C.; Nemr, E.; Tomb, R. The Place of 5-Aminolaevulinic Acid-Photodynamic Therapy in the Treatment Landscape of Urethral Warts: A Systematic Review. *Photodiagn. Photodyn. Ther.* **2021**, *33*, 102204. [\[CrossRef\]](#)
59. Stern, P.L.; van der Burg, S.H.; Hampson, I.N.; Broker, T.R.; Fiander, A.; Lacey, C.J.; Kitchener, H.C.; Einstein, M.H. Therapy of Human Papillomavirus-Related Disease. *Vaccine* **2012**, *30* (Suppl. S5), F71–F82. [\[CrossRef\]](#)
60. Mistrangelo, M.; Dal Conte, I.; Volpato, S.; Di Benedetto, G.; Testa, V.; Currado, F.; Morino, M. Current Treatments for Anal Condylomata Acuminata. *Minerva Chir.* **2018**, *73*, 100–106. [\[CrossRef\]](#)
61. Hu, Z.; Zheng, H.; Zeng, K. Predictors of Human Papillomavirus Persistence or Clearance after 5-Aminolevulinic Acid-Based Photodynamic Therapy in Patients with Genital Warts. *Photodiagn. Photodyn. Ther.* **2021**, *35*, 102431. [\[CrossRef\]](#)
62. Xie, J.; Ao, C.; Li, J.; Jiang, L.; Liu, H.; Zeng, K. 5-Aminolevulinic Acid Photodynamic Therapy for Condyloma Acuminatum of Urethral Meatus. *J. Dermatolog. Treat.* **2019**, *30*, 714–717. [\[CrossRef\]](#)
63. Buzzá, H.H.; Stringasci, M.D.; de Arruda, S.S.; Crestana, R.H.S.; de Castro, C.A.; Bagnato, V.S.; Inada, N.M. HPV-Induced Condylomata Acuminata Treated by Photodynamic Therapy in Comparison with Trichloroacetic Acid: A Randomized Clinical Trial. *Photodiagn. Photodyn. Ther.* **2021**, *35*, 102465. [\[CrossRef\]](#)

64. Hu, S.; Yang, Y.; Jiang, B.; Su, D.; Zhang, L.; Huang, Z.; Zhang, F. Treatment of Condyloma Acuminatum Using the Combination of Laser Ablation and ALA-PDT. *Photodiagn. Photodyn. Ther.* **2019**, *25*, 193–196. [CrossRef]
65. Shi, H.; Zhang, X.; Ma, C.; Yu, N.; Wang, J.; Xia, L.; Ge, X.; Liu, M.; Duan, A. Clinical Analysis of Five Methods Used to Treat Condylomata Acuminata. *Dermatology* **2013**, *227*, 338–345. [CrossRef]
66. Szeimies, R.-M.; Schleyer, V.; Moll, I.; Stocker, M.; Landthaler, M.; Karrer, S. Adjuvant Photodynamic Therapy Does Not Prevent Recurrence of Condylomata Acuminata after Carbon Dioxide Laser Ablation-A Phase III, Prospective, Randomized, Bicentric, Double-Blind Study. *Dermatol. Surg.* **2009**, *35*, 757–764. [CrossRef]
67. Kim, J.-H.; Kim, J.; Kim, K.; No, J.H.; Kim, Y.B.; Suh, D.H. Risk Factor and Treatment of Vaginal Intraepithelial Neoplasia After Hysterectomy for Cervical Intraepithelial Neoplasia. *J. Low Genit. Tract. Dis.* **2022**, *26*, 147–151. [CrossRef]
68. Hampson, L.; Martin-Hirsch, P.; Hampson, I.N. An Overview of Early Investigational Drugs for the Treatment of Human Papilloma Virus Infection and Associated Dysplasia. *Expert. Opin. Investig. Drugs* **2015**, *24*, 1529–1537. [CrossRef]
69. Zhang, W.; Zhang, A.; Sun, W.; Yue, Y.; Li, H. Efficacy and Safety of Photodynamic Therapy for Cervical Intraepithelial Neoplasia and Human Papilloma Virus Infection. *Medicine* **2018**, *97*, e10864. [CrossRef]
70. Tao, X.H.; Guan, Y.; Shao, D.; Xue, W.; Ye, F.S.; Wang, M.; He, M.H. Efficacy and Safety of Photodynamic Therapy for Cervical Intraepithelial Neoplasia: A Systemic Review. *Photodiagn. Photodyn. Ther.* **2014**, *11*, 104–112. [CrossRef]
71. Bodner, K.; Bodner-Adler, B.; Wierrani, F.; Kubin, A.; Szölts-Szölts, J.; Spängler, B.; Grünberger, W. Cold-Knife Conization versus Photodynamic Therapy with Topical 5-Aminolevulinic Acid (5-ALA) in Cervical Intraepithelial Neoplasia (CIN) II with Associated Human Papillomavirus Infection: A Comparison of Preliminary Results. *Anticancer. Res.* **2003**, *23*, 1785–1788.
72. Wang, J.; Xu, J.; Chen, J.; He, Q.; Xiang, L.; Huang, X.; Ding, G.; Xu, S. Successful Photodynamic Therapy with Topical 5-Aminolevulinic Acid for Five Cases of Cervical Intraepithelial Neoplasia. *Arch. Gynecol. Obstet.* **2010**, *282*, 307–312. [CrossRef]
73. Li, D.; Zhang, F.; Shi, L.; Lin, L.; Cai, Q.; Xu, Y. Treatment of HPV Infection-Associated Low Grade Cervical Intraepithelial Neoplasia with 5-Aminolevulinic Acid-Mediated Photodynamic Therapy. *Photodiagn. Photodyn. Ther.* **2020**, *32*, 101974. [CrossRef]
74. Cang, W.; Gu, L.; Hong, Z.; Wu, A.; Di, W.; Qiu, L. Effectiveness of Photodynamic Therapy with 5-Aminolevulinic Acid on HPV Clearance in Women without Cervical Lesions. *Photodiagn. Photodyn. Ther.* **2021**, *34*, 102293. [CrossRef] [PubMed]
75. Wu, A.; Li, Q.; Ling, J.; Gu, L.; Hong, Z.; Di, W.; Qiu, L. Effectiveness of Photodynamic Therapy in Women of Reproductive Age with Cervical High-Grade Squamous Intraepithelial Lesions (HSIL/CIN2). *Photodiagn. Photodyn. Ther.* **2021**, *36*, 102517. [CrossRef] [PubMed]
76. Su, Y.; Zhang, Y.; Tong, Y.; Zhang, L.; Li, P.; Zhang, H.; Zhang, X.; Tang, Y.; Qin, L.; Shen, Y.; et al. Effect and Rational Application of Topical Photodynamic Therapy (PDT) with 5-Aminolevulinic Acid (5-ALA) for Treatment of Cervical Intraepithelial Neoplasia with Vaginal Intraepithelial Neoplasia. *Photodiagn. Photodyn. Ther.* **2022**, *37*, 102634. [CrossRef] [PubMed]
77. Herpes Simplex Virus. Available online: <https://www.who.int/news-room/fact-sheets/detail/herpes-simplex-virus> (accessed on 26 November 2021).
78. Fisman, D.N. Health Related Quality of Life in Genital Herpes: A Pilot Comparison of Measures. *Sex. Transm. Infect.* **2005**, *81*, 267–270. [CrossRef]
79. Makarov, O.V.; Khashukaeva, A.Z.; Svitich, O.A.; Markova, È.A.; Khlynova, S.A.; Labzhinov, P.A.; Zverev, V.V. Anti-herpetic effect of photodynamic action in an in vitro experiment. *Zhurnal Mikrobiol. Epidemiol. Immunobiol.* **2014**, *1*, 48–55.
80. Osiecka, B.J.; Nockowski, P.; Kwiatkowski, S.; Szepietowski, J.C. Photodynamic Therapy with Red Light and 5-Aminolaevulinic Acid for Herpes Simplex Recurrence: Preliminary Results. *Acta. Derm. Venereol.* **2017**, *97*, 1239–1240. [CrossRef]
81. Donnarumma, G.; De Gregorio, V.; Fusco, A.; Farina, E.; Baroni, A.; Esposito, V.; Contaldo, M.; Petrucci, M.; Pannone, G.; Serpico, R. Inhibition of HSV-1 Replication by Laser Diode-Irradiation: Possible Mechanism of Action. *Int. J. Immunopathol. Pharmacol.* **2010**, *23*, 1167–1176. [CrossRef]
82. Ferreira, D.C.; Reis, H.L.B.; Cavalcante, F.S.; Santos, K.R.N.D.; Passos, M.R.L. Recurrent Herpes Simplex Infections: Laser Therapy as a Potential Tool for Long-Term Successful Treatment. *Rev. Soc. Bras. Med. Trop.* **2011**, *44*, 397–399. [CrossRef]
83. Davis, E.C.; Callender, V.D. Postinflammatory Hyperpigmentation. *J. Clin. Aesthet. Dermatol.* **2010**, *3*, 20–31.
84. Preissig, J.; Hamilton, K.; Markus, R. Current Laser Resurfacing Technologies: A Review That Delves Beneath the Surface. *Semin. Plast. Surg.* **2012**, *26*, 109–116. [CrossRef]
85. Kaufman, B.P.; Aman, T.; Alexis, A.F. Postinflammatory Hyperpigmentation: Epidemiology, Clinical Presentation, Pathogenesis and Treatment. *Am. J. Clin. Dermatol.* **2018**, *19*, 489–503. [CrossRef]
86. Barrett, T.; de Zwaan, S. Picosecond Alexandrite Laser Is Superior to Q-Switched Nd:YAG Laser in Treatment of Minocycline-Induced Hyperpigmentation: A Case Study and Review of the Literature. *J. Cosmet. Laser Ther.* **2018**, *20*, 387–390. [CrossRef]
87. Wang, J.V.; Christman, M.P.; Feng, H.; Ferzli, G.; Jeon, H.; Geronemus, R.G. Laser-Assisted Delivery of Tranexamic Acid for Melasma: Pilot Study Using a Novel 1927 Nm Fractional Thulium Fiber Laser. *J. Cosmet. Dermatol.* **2021**, *20*, 105–109. [CrossRef]
88. Unemo, M.; Ross, J.; Serwin, A.B.; Gomberg, M.; Cusini, M.; Jensen, J.S. Background Review for the “2020 European Guideline for the Diagnosis and Treatment of Gonorrhoea in Adults”. *Int. J. STD AIDS* **2021**, *32*, 108–126. [CrossRef]
89. Klausner, J.D.; Bristow, C.C.; Soge, O.O.; Shahkolahi, A.; Waymer, T.; Bolan, R.K.; Philip, S.S.; Asbel, L.E.; Taylor, S.N.; Mena, L.A.; et al. Resistance-Guided Treatment of Gonorrhea: A Prospective Clinical Study. *Clin. Infect. Dis.* **2021**, *73*, 298–303. [CrossRef]
90. Wang, Y.; Ferrer-Espada, R.; Baglo, Y.; Goh, X.S.; Held, K.D.; Grad, Y.H.; Gu, Y.; Gelfand, J.A.; Dai, T. Photoinactivation of Neisseria Gonorrhoeae: A Paradigm-Changing Approach for Combating Antibiotic-Resistant Gonococcal Infection. *J. Infect. Dis.* **2019**, *220*, 873–881. [CrossRef]

91. Wang, Y.; Ferrer-Espada, R.; Gu, Y.; Dai, T. Antimicrobial Blue Light: An Alternative Therapeutic for Multidrug-Resistant Gonococcal Infections? *MOJ Sol. Photoenergy Syst.* **2017**, *1*, 00009. [\[CrossRef\]](#)
92. Marti, H.; Koschwanetz, M.; Pesch, T.; Blenn, C.; Borel, N. Water-Filtered Infrared a Irradiation in Combination with Visible Light Inhibits Acute Chlamydial Infection. *PLoS ONE* **2014**, *9*, e102239. [\[CrossRef\]](#)
93. Wasson, C.J.; Zourelis, J.L.; Aardsma, N.A.; Eells, J.T.; Ganger, M.T.; Schober, J.M.; Skwor, T.A. Inhibitory Effects of 405 Nm Irradiation on Chlamydia Trachomatis Growth and Characterization of the Ensuing Inflammatory Response in HeLa Cells. *BMC Microbiol.* **2012**, *12*, 176. [\[CrossRef\]](#)
94. Kissinger, P. Trichomonas Vaginalis: A Review of Epidemiologic, Clinical and Treatment Issues. *BMC Infect. Dis.* **2015**, *15*, 307. [\[CrossRef\]](#)
95. Ilbeigi, S.; Dehdari Vais, R.; Sattarahmady, N. Photo-Genosensor for Trichomonas Vaginalis Based on Gold Nanoparticles-Genomic DNA. *Photodiagn. Photodyn. Ther.* **2021**, *34*, 102290. [\[CrossRef\]](#)
96. Silva Fonseca, T.H.; Alacoque, M.; Silva Oliveira, F.M.; Soares, B.M.; Leite, H.V.; Caliar, M.V.; Gomes, M.A.; Busatti, H. Photodynamic Therapy as a New Approach to Trichomonas Vaginalis Inactivation. *Photodiagn. Photodyn. Ther.* **2018**, *22*, 91–95. [\[CrossRef\]](#)
97. Du, M.; Xuan, W.; Zhen, X.; He, L.; Lan, L.; Yang, S.; Wu, N.; Qin, J.; Zhao, R.; Qin, J.; et al. Antimicrobial Photodynamic Therapy for Oral Candida Infection in Adult AIDS Patients: A Pilot Clinical Trial. *Photodiagn. Photodyn. Ther.* **2021**, *34*, 102310. [\[CrossRef\]](#)
98. Pinto, M.V.M.; Baron, M.; Corrêa, M.; Berton, J.; de Mattos, C.O.C.; Pieczaki, M.; Ronis, A.; Fortuny, E.; Padilha, M.R. Use of the Scar Acceleration Method—Mac@in the Treatment of Vulvovaginal Candidiasis: A Proposal for Treatment in Public Health in Sus, Brazil. *Open J. Appl. Sci.* **2020**, *10*, 758–765. [\[CrossRef\]](#)
99. Wang, T.; Dong, J.; Yin, H.; Zhang, G. Blue Light Therapy to Treat Candida Vaginitis with Comparisons of Three Wavelengths: An in Vitro Study. *Lasers Med. Sci.* **2020**, *35*, 1329–1339. [\[CrossRef\]](#)
100. De Santi, M.E.S.O.; Prates, R.A.; França, C.M.; Lopes, R.G.; Sousa, A.S.; Ferreira, L.R.; Bussadori, S.K.; Fernandes, A.U.; Deana, A.M. Antimicrobial Photodynamic Therapy as a New Approach for the Treatment of Vulvovaginal Candidiasis: Preliminary Results. *Lasers Med. Sci.* **2018**, *33*, 1925–1931. [\[CrossRef\]](#)
101. Machado-de-Sena, R.M.; Corrêa, L.; Kato, I.T.; Prates, R.A.; Senna, A.M.; Santos, C.C.; Picanço, D.A.; Ribeiro, M.S. Photodynamic Therapy Has Antifungal Effect and Reduces Inflammatory Signals in Candida Albicans-Induced Murine Vaginitis. *Photodiagn. Photodyn. Ther.* **2014**, *11*, 275–282. [\[CrossRef\]](#)
102. Ichimura, H.; Yamaguchi, S.; Kojima, A.; Tanaka, T.; Niiya, K.; Takemori, M.; Hasegawa, K.; Nishimura, R. Eradication and Reinfection of Human Papillomavirus after Photodynamic Therapy for Cervical Intraepithelial Neoplasia. *Int. J. Clin. Oncol.* **2003**, *8*, 322–325. [\[CrossRef\]](#)
103. Wierrani, F.; Kubin, A.; Jindra, R.; Henry, M.; Gharehbaghi, K.; Grin, W.; Söltz-Szötz, J.; Alth, G.; Grünberger, W. 5-Aminolevulinic Acid-Mediated Photodynamic Therapy of Intraepithelial Neoplasia and Human Papillomavirus of the Uterine Cervix—A New Experimental Approach. *Cancer Detect. Prev.* **1999**, *23*, 351–355. [\[CrossRef\]](#)
104. Soergel, P.; Wang, X.; Stepp, H.; Hertel, H.; Hillemanns, P. Photodynamic Therapy of Cervical Intraepithelial Neoplasia with Hexaminolevulinic. *Lasers Surg. Med.* **2008**, *40*, 611–615. [\[CrossRef\]](#)
105. Yamaguchi, S.; Tsuda, H.; Takemori, M.; Nakata, S.; Nishimura, S.; Kawamura, N.; Hanioka, K.; Inoue, T.; Nishimura, R. Photodynamic Therapy for Cervical Intraepithelial Neoplasia. *Oncology* **2005**, *69*, 110–116. [\[CrossRef\]](#) [\[PubMed\]](#)
106. Istomin, Y.P.; Lapzevich, T.P.; Chalau, V.N.; Shliakhtsin, S.V.; Trukhachova, T.V. Photodynamic Therapy of Cervical Intraepithelial Neoplasia Grades II and III with Photolon. *Photodiagn. Photodyn. Ther.* **2010**, *7*, 144–151. [\[CrossRef\]](#) [\[PubMed\]](#)
107. Wang, Y.; Ferrer-Espada, R.; Baglo, Y.; Gu, Y.; Dai, T. Antimicrobial Blue Light Inactivation of Neisseria Gonorrhoeae: Roles of Wavelength, Endogenous Photosensitizer, Oxygen, and Reactive Oxygen Species. *Lasers Surg. Med.* **2019**, *51*, 815–823. [\[CrossRef\]](#) [\[PubMed\]](#)