

IECBM
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Modeling and simulation of magnetoliposomes formation by encapsulation of core-shell, magnetite-chitosan nanoparticles in liposomes enabled by a low-cost microfluidic system



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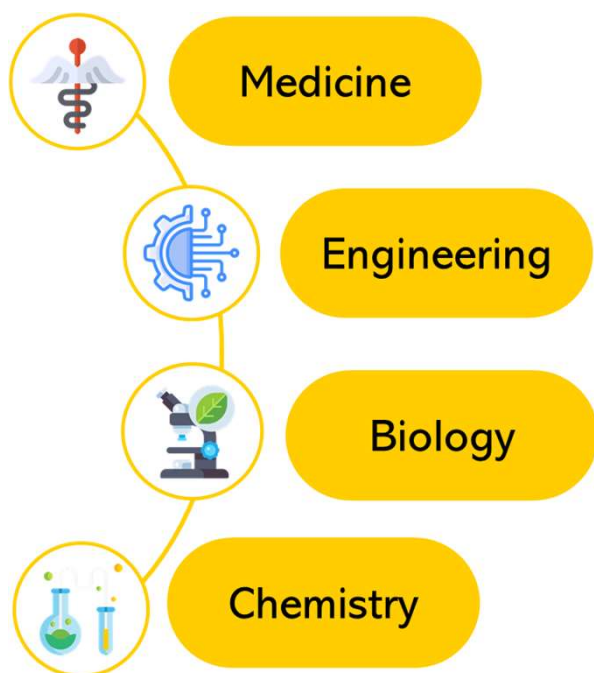
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Nanostructured materials applications

Nanobiotechnology



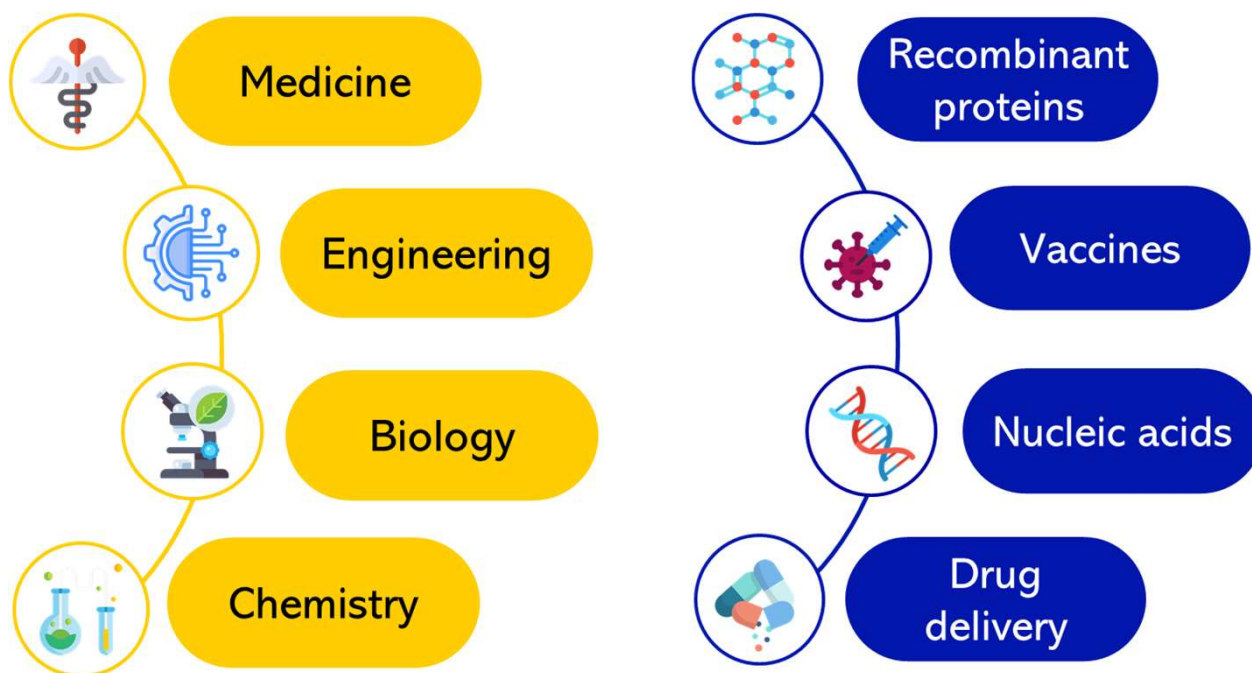
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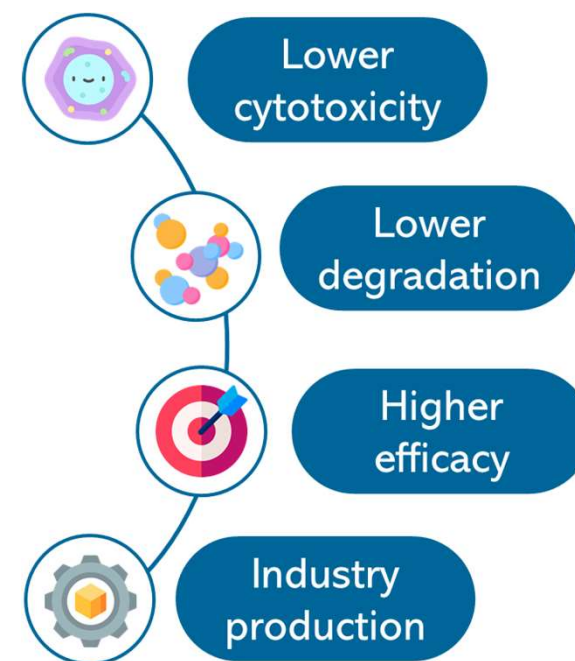
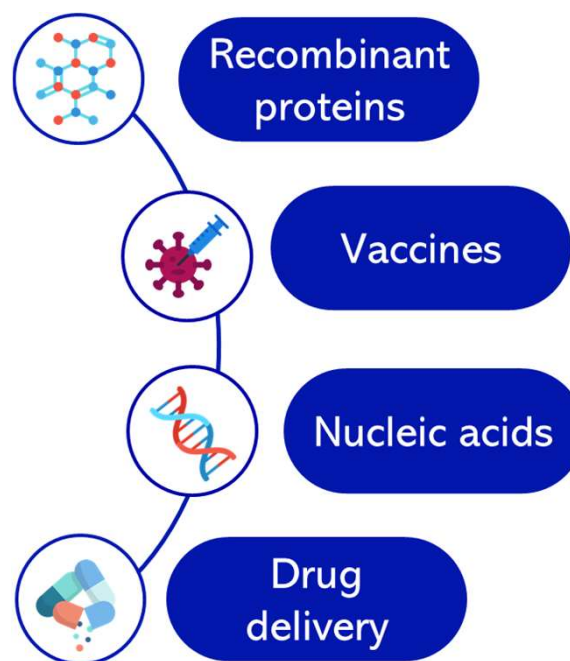
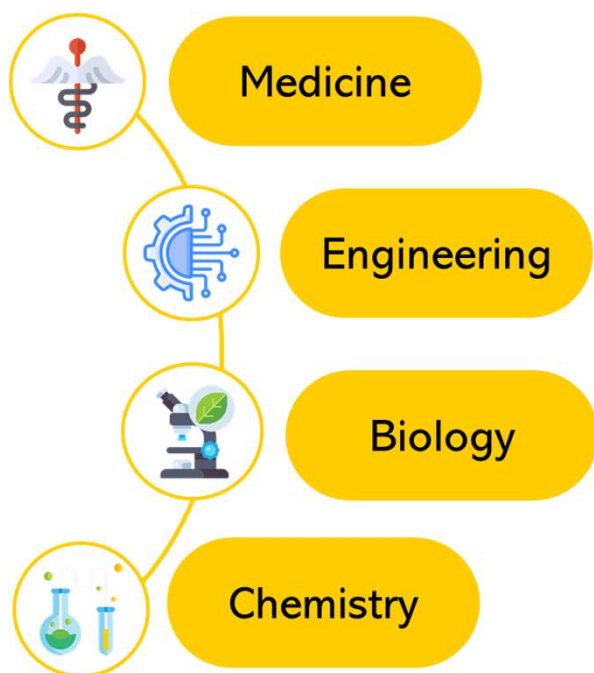
Lubhandwa S Biswaro, Maurício G da Costa Sousa, Taia Rezende, Simoni C Dias, and Octavio L Franco. Antimicrobial peptides and nanotechnology, recent advances and challenges. *Frontiers in microbiology*, 9:855, 2018.

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Nanostructured materials applications

Nanobiotechnology



Nanotransporters advantages like drug delivery systems

Nanobiotechnology



Protection of peptides, vaccines and desired drugs against extracellular degradation



Selectivity of the target therapy



Improvement of the pharmacokinetic profile

Nanotransporters advantages like drug delivery systems

Nanobiotechnology



Protection of peptides, vaccines and desired drugs against extracellular degradation



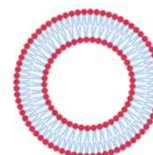
Selectivity of the target therapy



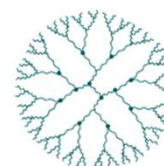
Improvement of the pharmacokinetic profile



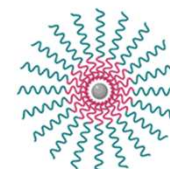
Micelles



Liposomes



Dendrimers

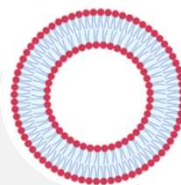


Polymeric nanoparticles

Nano bioconjugates compounds advantages

Liposomes

- Biocompatibility
- Biodegradability
- Encapsulate hydrophilic/hydrophobic compounds
- Similarity to cell's membranes
- Low cytotoxicity / High delivery rates



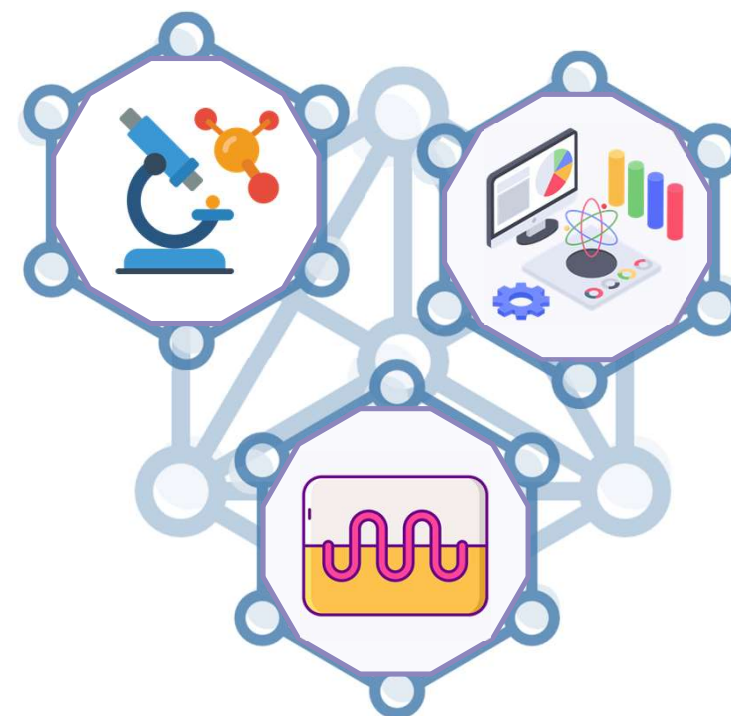
- Nanoparticles as drug delivery vehicles
- Iron oxide nanoparticles (Fe_3O_4)
- Advances in synthesis allows potential applications in diagnostic/therapeutic techniques.



Nanoparticles

Nano bioconjugates synthesis technique

- Manufacture devices to **outperforming** traditional analytical and sensing instruments employed in scientific research.
- **Development of several industries:** medical, cosmetics, pharma and food. Low cost devices
- Explored for the synthesis, manipulation, and separation of microscopic encapsulates.
- Explore in silico and in vitro species dynamics to understand **interaction between different phases in a system**



Objectives

Implement multiphysics simulations to study *in silico* the encapsulation of core-shell, magnetite-chitosan nanoparticles in a microfluidic system by means of the COMSOL Multiphysics® software.



Euler-Euler (EE) Model

Liposome encapsulation of core-shell, magnetite-chitosan nanoparticles in a microfluidic system using a multiphase Euler-Euler computational model.

COMSOL Multiphysics Software

Euler-Euler (EE) model

Parameters

Material	Property	Value
H ₂ O	Density	997 [kg/m ³]
	Dinamic Viscosity	8.9×10^{-4} [Pa * s]
CS-MNP's	Density	1100[kg/m ³]
	Diffusivity	1.0×10^{-9} [m ² /s]
Liposomes	Density	1000[kg/m ³]
	Diffusivity	1.51×10^{-5} [m ² /s]



Euler-Euler (EE) Model

Mass Balance

Continuity relations for continuous and dispersed phases

Continuous phase (liposomes)



$$\frac{\partial}{\partial t}(\rho_c \Phi_c) + \nabla \cdot (\rho_c \Phi_c u_c) = m_{dc}$$

Dispersed phase (CS-MNPs)



$$\frac{\partial}{\partial t}(\rho_d \Phi_d) + \nabla \cdot (\rho_d \Phi_d u_d) = -m_{dc}$$

Where

- Φ = volume fraction (dimensionless)
- ρ = density (Kg/m^3)
- u = velocity (m/s)
- m_{dc} = mass transfer rate



Euler-Euler (EE) Model

Mass Balance

Relation between volume fractions. Continuous and dispersed phases considered incompressible

$$\Phi_c = 1 - \Phi_d$$

Continuous phase (liposomes)



$$\frac{\partial \Phi_c}{\partial t} + \nabla \cdot (\Phi_c u_c) = \frac{m_{dc}}{\rho_c}$$

Dispersed phase (CS-MNPs)



$$\frac{\partial \Phi_d}{\partial t} + \nabla \cdot (\Phi_d u_d) = \frac{-m_{dc}}{\rho_d}$$

Compute volume fraction



Euler-Euler (EE) Model

Mass Balance

Continuity equation continuous phase mass balance and dispersed phase mass balance

$$\nabla \cdot (\Phi_c u_c) + \nabla \cdot (\Phi_d u_d) = \frac{m_{dc}}{\rho_c} - \frac{m_{dc}}{\rho_d} \quad \Phi_c = 1 - \Phi_d$$

$$\nabla \cdot [(1 - \Phi_d)u_c + (\Phi_d u_d)] = m_{dc} \left(\frac{1}{\rho_c} - \frac{1}{\rho_d} \right) \quad \text{Compute mixture pressure}$$



Euler-Euler (EE) Model

Momentum Balance

For fluid-solids mixtures where $\rho_d \gg \rho_c$, momentum balanced dispersed phase is modified in the manner of Enwald

Continuous phase (liposomes)



$$\rho_c \Phi_c \left[\frac{\partial}{\partial t} (u_c) + u_c \nabla \cdot (u_c) \right] = -\Phi_c \nabla p + \underbrace{\Phi_d \nabla \cdot \tau_d}_{\nabla p_s} + \Phi_c p_c \mathbf{g} + \mathbf{F}_{m,c} + \Phi_c \mathbf{F}_c - m_{dc} (u_{int} - u_c)$$

Dispersed phase (CS-MNPs)



$$\rho_d \Phi_d \left[\frac{\partial}{\partial t} (u_d) + u_d \nabla \cdot (u_d) \right] = -\Phi_d \nabla p + \underbrace{\Phi_d \nabla \cdot \tau_d}_{\nabla p_s} + \Phi_d p_d \mathbf{g} + \mathbf{F}_{m,d} + \Phi_d \mathbf{F}_d - m_{dc} (u_{int} - u_d)$$

Where

- p_s = solid pressure (Pa)
- τ = viscous stress tensor (Pa)

Momentum balance was assumed Newtonian fluid



Euler-Euler (EE) Model

Viscous Stress Tensors

Continuous phase (liposomes)



$$\tau_c = \mu_c^m \left(\nabla u_c + (u_c)^T - \frac{2}{3} (\nabla \cdot u_c) I \right)$$

Dispersed phase (CS-MNPs)



$$\tau_d = \mu_d^m \left(\nabla u_d + (u_d)^T - \frac{2}{3} (\nabla \cdot u_d) I \right)$$

Where $\mu_c^m, \mu_d^m = \text{dynamic viscosity (Pa)}$



Euler-Euler (EE) Model

Momentum Balance

Using nonconservative forms

Dispersed phase (CS-MNPs)



$$\rho_c \left[\frac{\partial}{\partial t} (u_d) + u_d \nabla \cdot (u_d) \right] = -\nabla p + \nabla \cdot \tau_d + \rho_d \mathbf{g} + \frac{\mathbf{F}_{m,d}}{d} + \mathbf{F}_d - \frac{m_{dc}(u_{int} - u_d)}{d}$$

Where

- p = mixture pressure (Pa) **equal for two phases**
- τ = viscous stress tensor (Pa)
- \mathbf{g} = vector of gravitational acceleration (m/s^2)
- \mathbf{F}_m = interphase momentum transfer term (N/m^3)
- \mathbf{F} = volume force term (N/m^3)
- u_{int} = interphase velocity (m/s)



Euler-Euler (EE) Model

Viscosity Models

Dispersed phase (CS-MNPs)



Krieger Type

$$\mu_{mix} = \mu_c \left(1 - \frac{\Phi_d}{\Phi_{d,max}} \right)^{-2.5\Phi_{d,max}}$$

Where

Maximum packing limit

$$\Phi_{d,max} = 0.62$$



Euler-Euler (EE) Model

Damping functions - Wall BC

Lam and Bremhorst EE Model (LB)

$$E = 0$$

$$\varepsilon_w = \nu \left(\frac{\partial^2 k}{\partial y^2} \right) \quad \text{Wall Function}$$

Viscous stress tensor related term

Yang et al. EE Model (YS)

$$E = \nu v_t \left(\frac{\partial^2 U^2}{\partial y^2} \right)$$

$$\varepsilon_w = \nu \left(\frac{\partial^2 k}{\partial y^2} \right) \quad \text{Wall Function}$$

Viscous stress tensor related term

Manufacturing Process

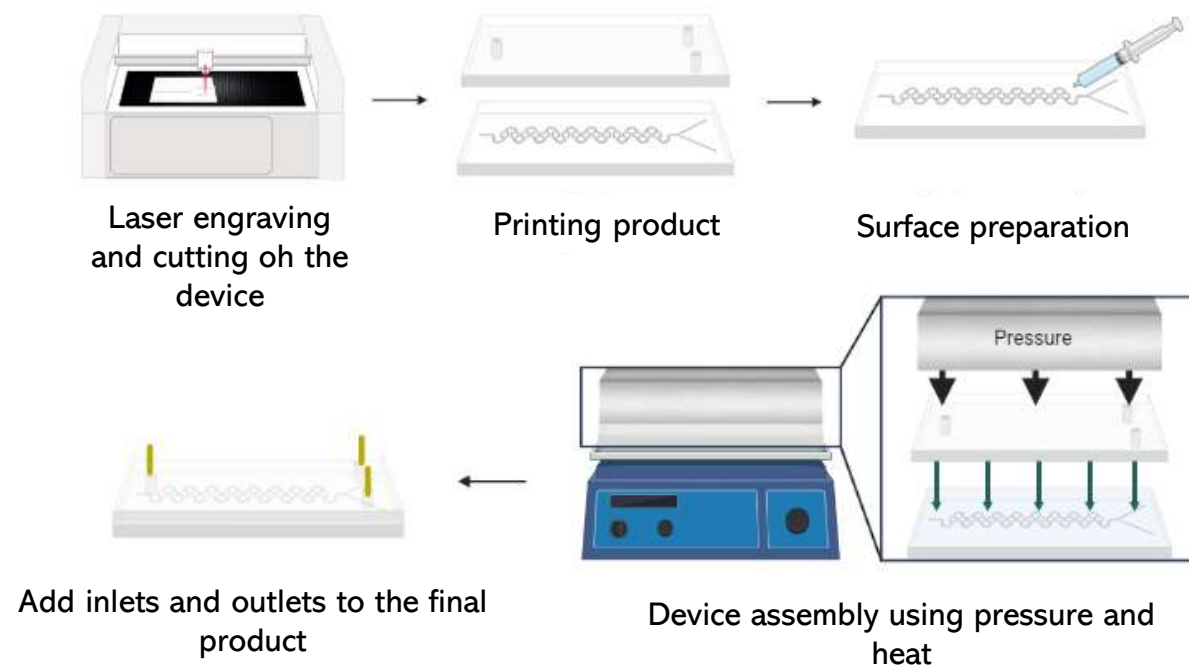


Figure 1. Manufacturing process

Geometries and Boundary conditions

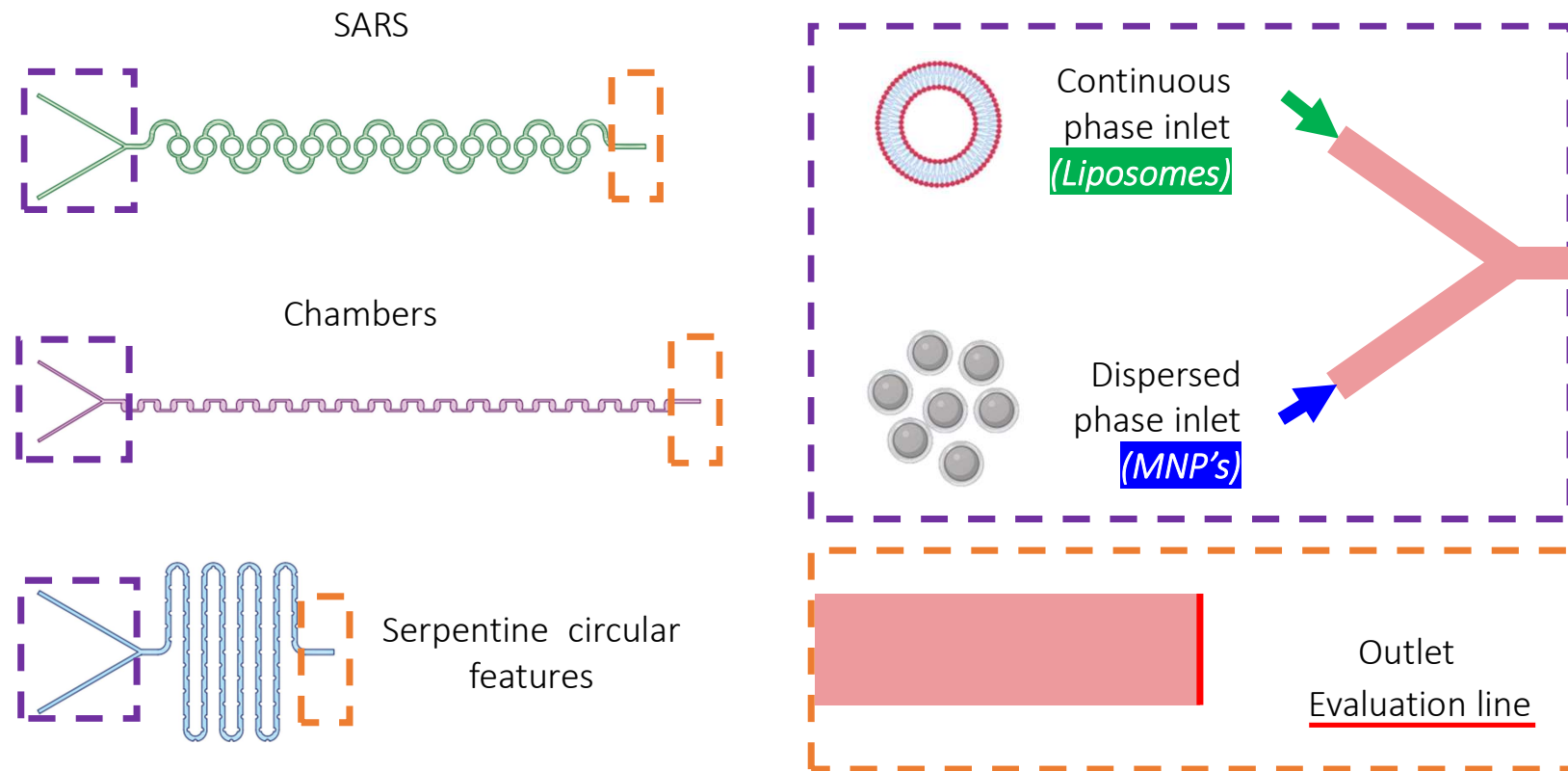


Figure 2. Geometries and boundary conditions, inlets, outlets and evaluation lines

Mesh selection

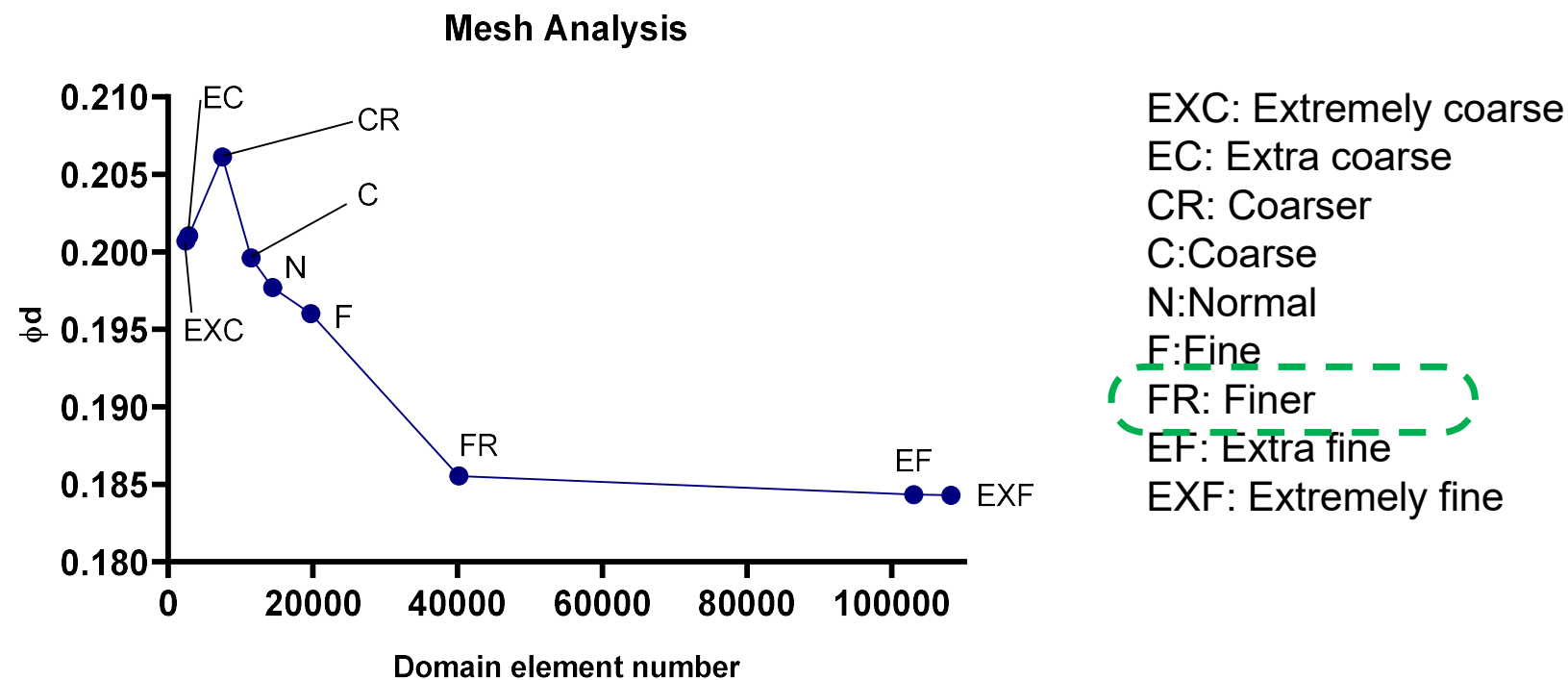


Figure 3. Mesh convergence analysis for SARS geometry. Complete mesh consist of 40175 domain elements

Mesh selection

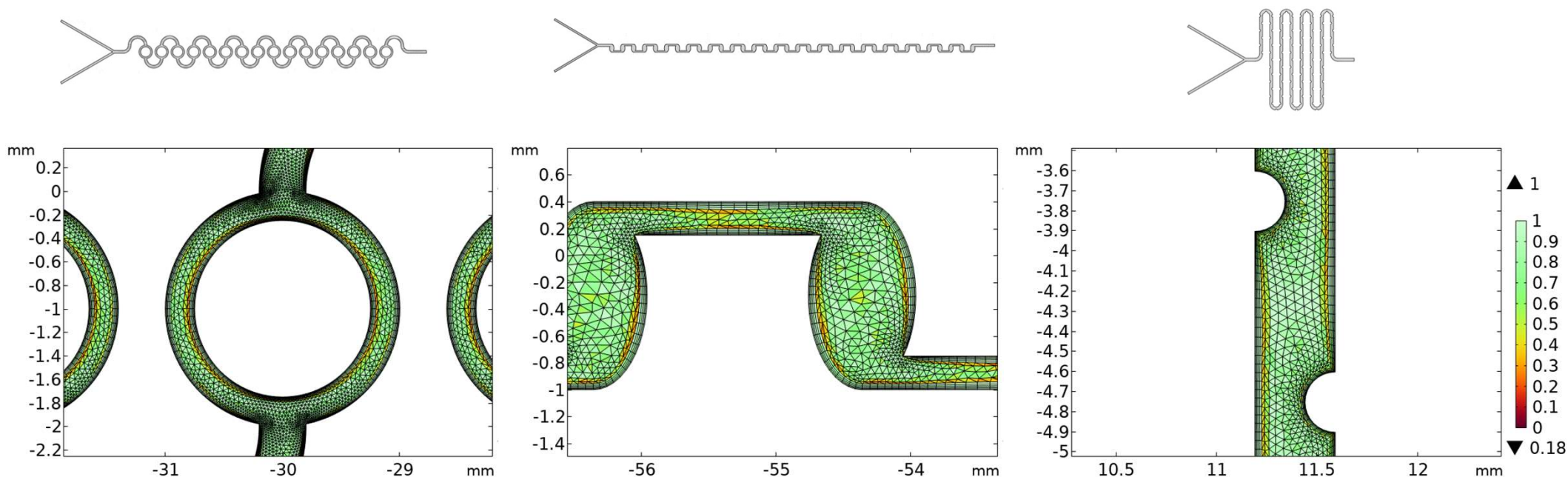


Figure 4. Mesh quality. Left to right: SARS, Chambers and Serpentine circular features geometry

Dispersed phase Volume fraction

Chambers

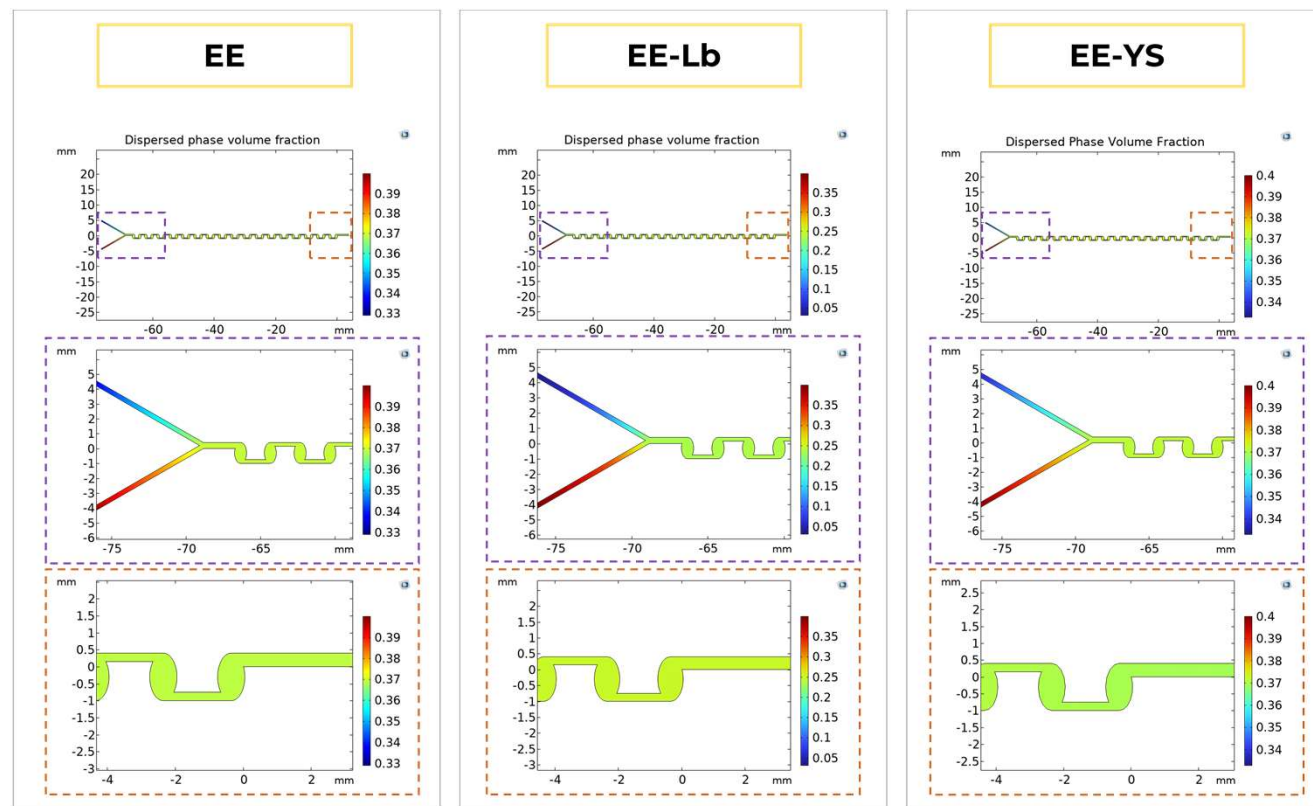


Figure 5. Dispersed phase volume fraction using variations in EE Model in chambers geometry

Dispersed phase Volume fraction

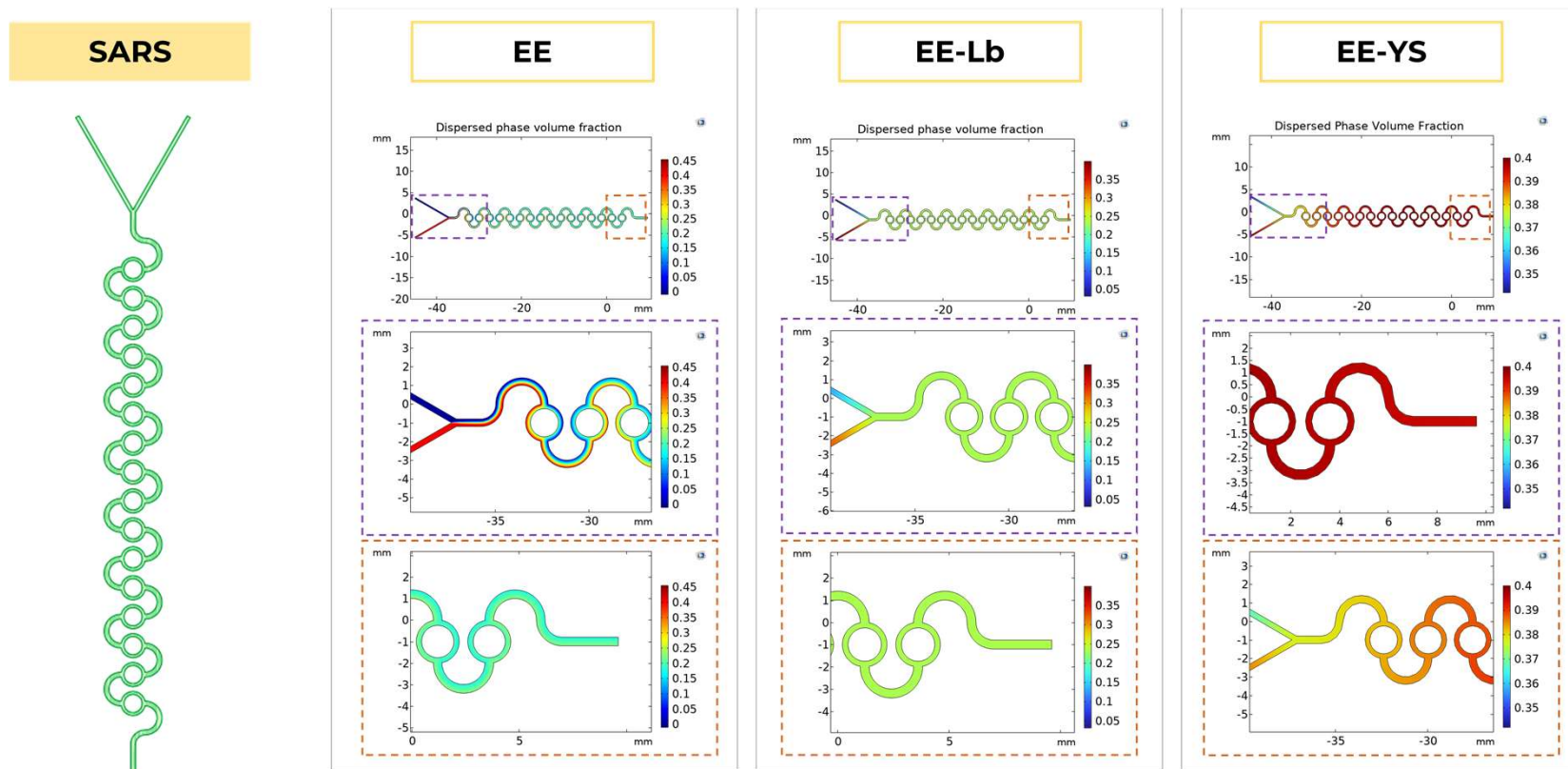


Figure 6. Dispersed phase volume fraction using variations in EE Model in SARS geometry

Dispersed phase Volume fraction

**Serpentine
Circular
Features**

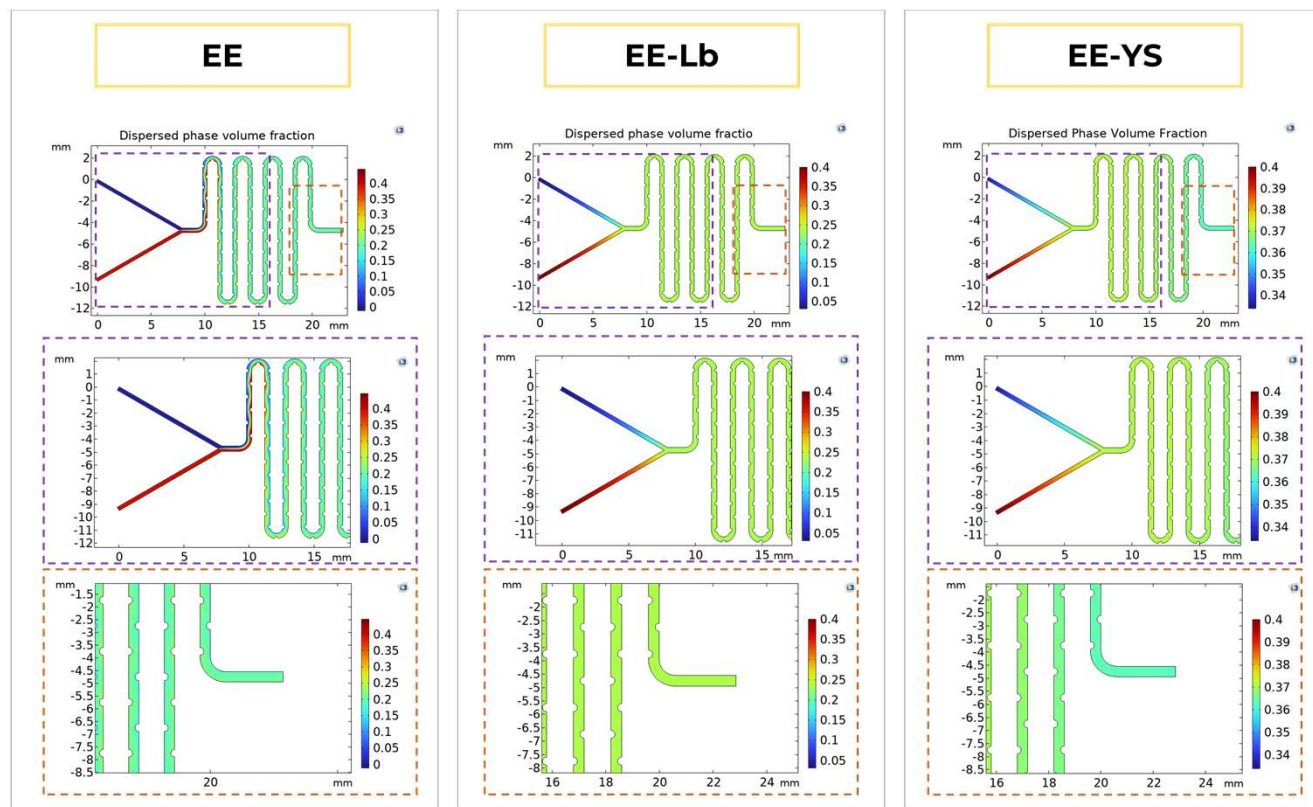
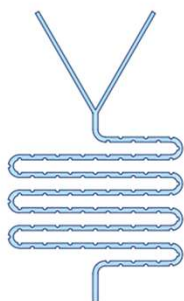
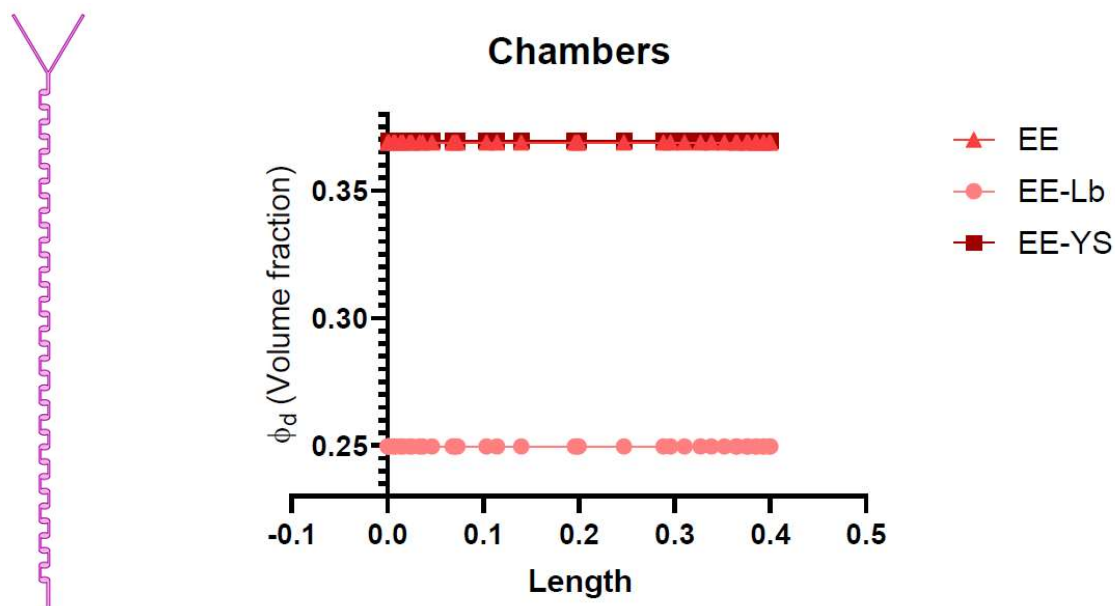


Figure 7. Dispersed phase volume fraction using variations in EE Model in serpentine CF geometry

Encapsulation Efficiency (EE%)



$$EE\% = \left(\frac{\phi_{d,min}}{\phi_d / FRR} \right) \times 100$$

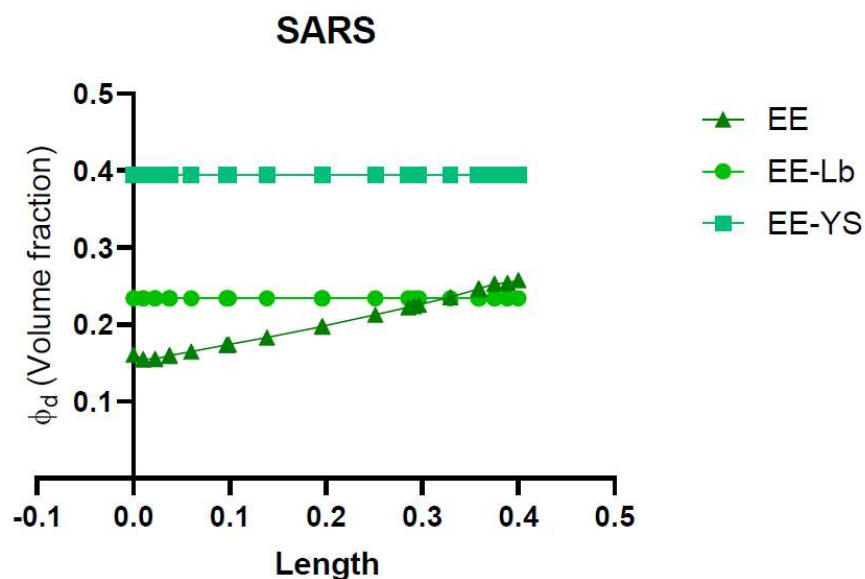
$$EE \ EE\% = 38.69$$

$$EE \ Lb \ EE\% = 58.48$$

$$EE \ Ys \ EE\% = 98.69$$

Figure 8. Encapsulation efficiency Volume fraction variation by length in chambers geometry

Encapsulation Efficiency (EE%)



$$EE\% = \left(\frac{\phi_{d,min}}{\phi_d / FRR} \right) \times 100$$

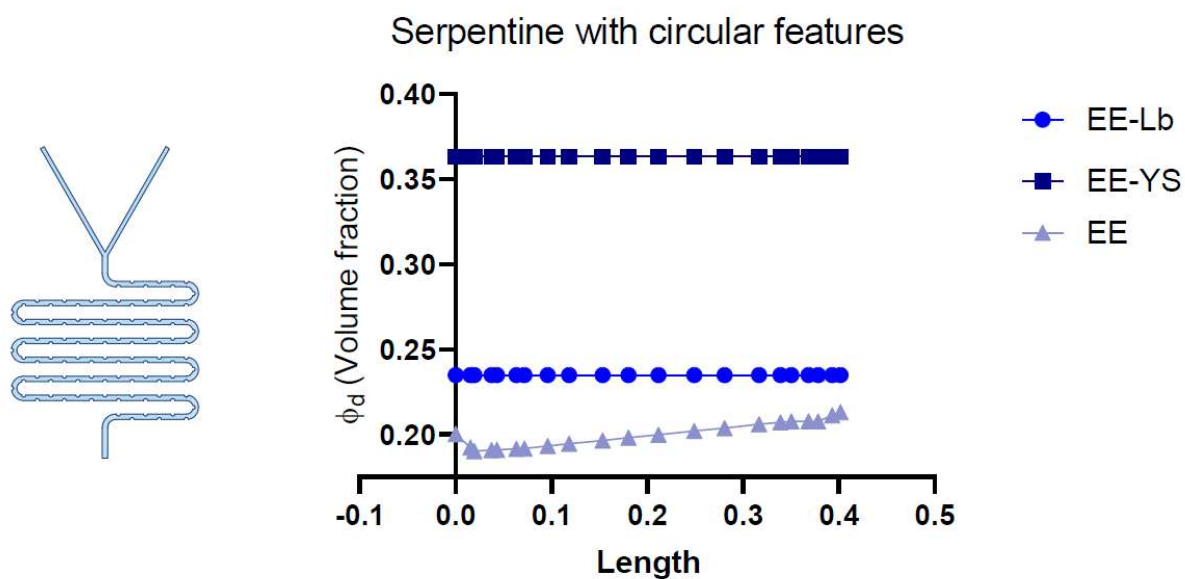
$$EE \ EE\% = 47.58$$

$$EE \ Lb \ EE\% = 58.71$$

$$EE \ Ys \ EE\% = 90.79$$

Figure 9. Encapsulation efficiency Volume fraction variation by length in SARS geometry

Encapsulation Efficiency (EE%)



$$EE\% = \left(\frac{\phi_{d,min}}{\phi_d / FRR} \right) \times 100$$

$$EE \ EE\% = 92.18$$

$$EE \ Lb \ EE\% = 62.38$$

$$EE \ Ys \ EE\% = 92.33$$

Figure 10. Encapsulation efficiency Volume fraction variation by length in serpentine CF geometry

Encapsulation Efficiency (EE%)

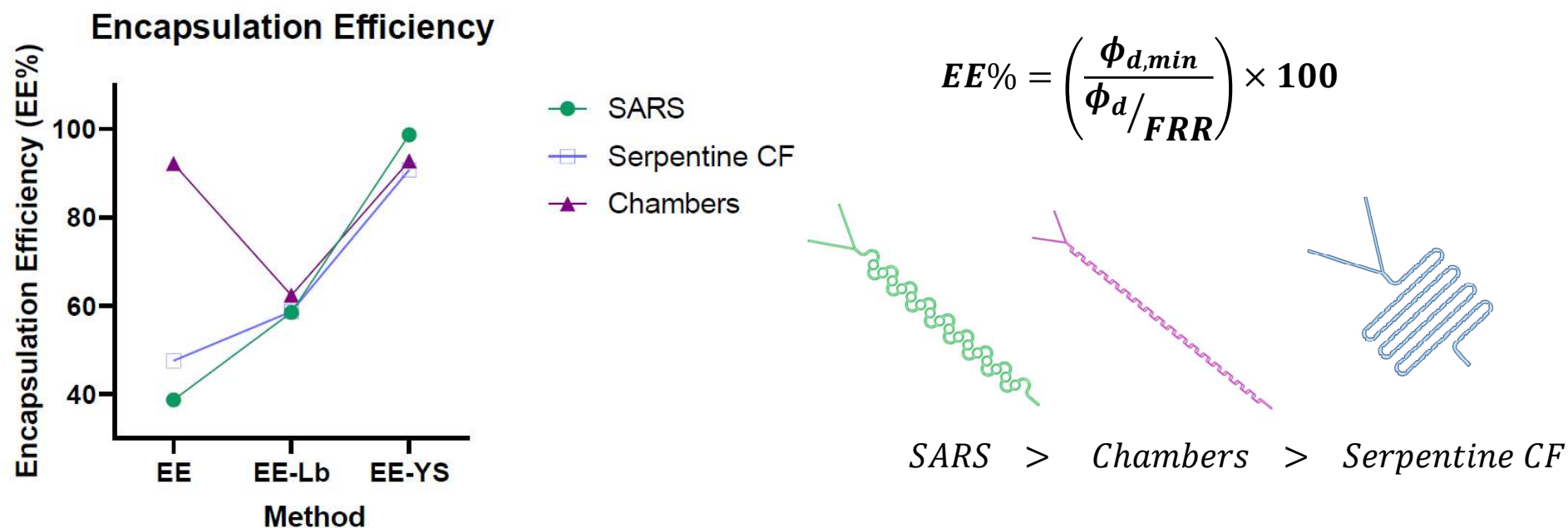


Figure 11. Encapsulation efficiency Volume fraction variation by length. Comparative graph

Encapsulation Efficiency (EE%)

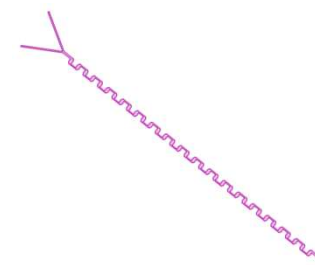
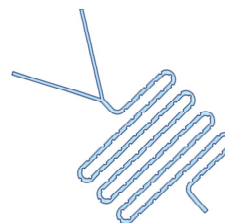
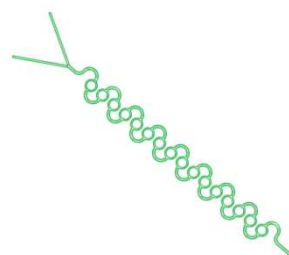


Table 2. Encapsulation efficiency contrast between geometries and math models

Model/Geometry	SARS	Serp CF	Chambers
EE	38.69	47.58	92.18
EE-Lb	58.48	58.71	62.38
EE-YS	98.69	90.79	92.33

Conclusions

- By implementing near-wall damping functions in the Euler-Euler approximation, it is possible to model the mixing process more effectively as they address the **turbulence phenomena at low Reynolds numbers by balancing the rate of of kinetic energy dissipation** and thus ensure a better encapsulation process.
- The LB and YS turbulence models had the best performance, obtaining EE% between **58.48-62.38% and 90.79-98.69%** respectively.
- The Euler-Euler model was sufficient for the analysis; however, **stochastic** processes related to the mixing process will be considered in future work.
- According to the YS model with damping functions, the micromixer with the channel SARS geometry led to the highest EE%, i.e., 98.69%.

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Thank you
for your attention