

Entry

Lichen as Multipartner Symbiotic Relationships

Lourdes Morillas ^{1,2,*} , Javier Roales ^{1,3} , Cristina Cruz ¹  and Silvana Munzi ^{1,4} 

- ¹ Center for Ecology, Evolution and Environmental Changes & CHANGE—Global Change and Sustainability Institute, Faculdade de Ciências, Universidade de Lisboa, Campo Grande, Bloco C2, 1749-016 Lisbon, Portugal; jroabat@upo.es (J.R.); ccruz@fc.ul.pt (C.C.); ssmunzi@fc.ul.pt (S.M.)
- ² Departamento de Biología Vegetal y Ecología, Universidad de Sevilla, Av. Reina Mercedes 10, 41080 Seville, Spain
- ³ Departamento de Sistemas Físicos, Químicos y Naturales, Universidad Pablo de Olavide, Ctra. Utrera Km 1, 41013 Seville, Spain
- ⁴ Centro Interuniversitário de História das Ciências e da Tecnologia, Faculdade de Ciências, Universidade de Lisboa, Campo Grande, 1749-016 Lisbon, Portugal
- * Correspondence: lmorillas@fc.ul.pt

Definition: Lichens have long been considered as composite organisms composed of algae and/or cyanobacteria hosted by a fungus in a mutualistic relationship. Other organisms have been gradually discovered within the lichen thalli, such as multiple algal species, yeasts, or even viruses. Of pivotal relevance is the existence of the lichen microbiome, which is a community of microorganisms that can be found living together on the lichen surface. This community performs a growing number of functions. In this entry, we explore the journey of lichens being considered from a dual partnership to a multi-species symbiotic relationship.

Keywords: symbiosis; microbiome; partnership; mycobiont; photobiont; holobiont; bacterial layer



Citation: Morillas, L.; Roales, J.; Cruz, C.; Munzi, S. Lichen as Multipartner Symbiotic Relationships. *Encyclopedia* **2022**, *2*, 1421–1431. <https://doi.org/10.3390/encyclopedia2030096>

Academic Editors: Milva Pepi and Raffaele Barretta

Received: 5 May 2022

Accepted: 25 July 2022

Published: 3 August 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. General Context of Lichens

Lichens are just one of many symbiotic relationships that can be established between heterotrophic fungi and photoautotrophic partners, such as plants, mosses, cyanobacteria, and algae. Partnerships between fungi and vascular plants are highly diverse and ecologically relevant. Some of these partnerships, such as those of ectomycorrhizas [1], endomycorrhizas, or the unique orchid mycorrhizal associations [2], are well known. The relationships between fungi and cyanobacteria or algae are also well known and very diverse, including the relation between algicolous fungi and bacteria or algae [3] and lichens [4]. Algicolous fungi can parasitize algae or cyanobacteria [5,6] or alternatively can establish a mutualistic relationship, as in the case of mycophycobioses [7]. Lichen-forming fungi may establish symbiotic relationships with algae or cyanobacteria and form a unique identity, i.e., the lichen. However, although unique, lichens are just one example of the highly diverse partnerships between fungi and photosynthetic organisms.

More than 18,000 fungal species, comprising a highly diverse group and representing around 20% of those currently identified, participate in lichen partnerships. They occur in all terrestrial ecosystems, ranging from polar to tropical areas and from coastal to high mountain ecosystems. Lichens form vegetative structures called thalli and can grow on a large variety of substrates such as minerals, rocks, bare soil, and the wood or leaves of plants, even in streams and marine zones [8], as well as on synthetic material surfaces.

The symbiotic condition of lichens remained unknown for a long time and, until 1869 [9], they were thought to be individual organisms. The German mycologist Anton de Bary introduced the term “symbiosis” to describe the condition of dissimilar organisms living together [10] supporting Schwendener’s theory that lichen is formed by two separate organisms, a fungus and an alga. From then on, lichens were considered to be an obligate partnership between a fungus (mycobiont) and either cyanobacteria and/or green algae,

that acted as a photoautotrophic partner (photobiont) [8,11]. Figure 1 shows the location of the mycobiont and the photobiont within heteromerous lichens, in which the algae and fungal components are arranged in definite layers. The stability of this symbiotic association depends on the mutualistic–antagonistic relationships of a multitude of interlinked organisms, also known as the “holobiont” [12,13]. This complex relationship is determined by the symbionts’ interactive intimacy, stability of environmental conditions, and partner availability [14,15].

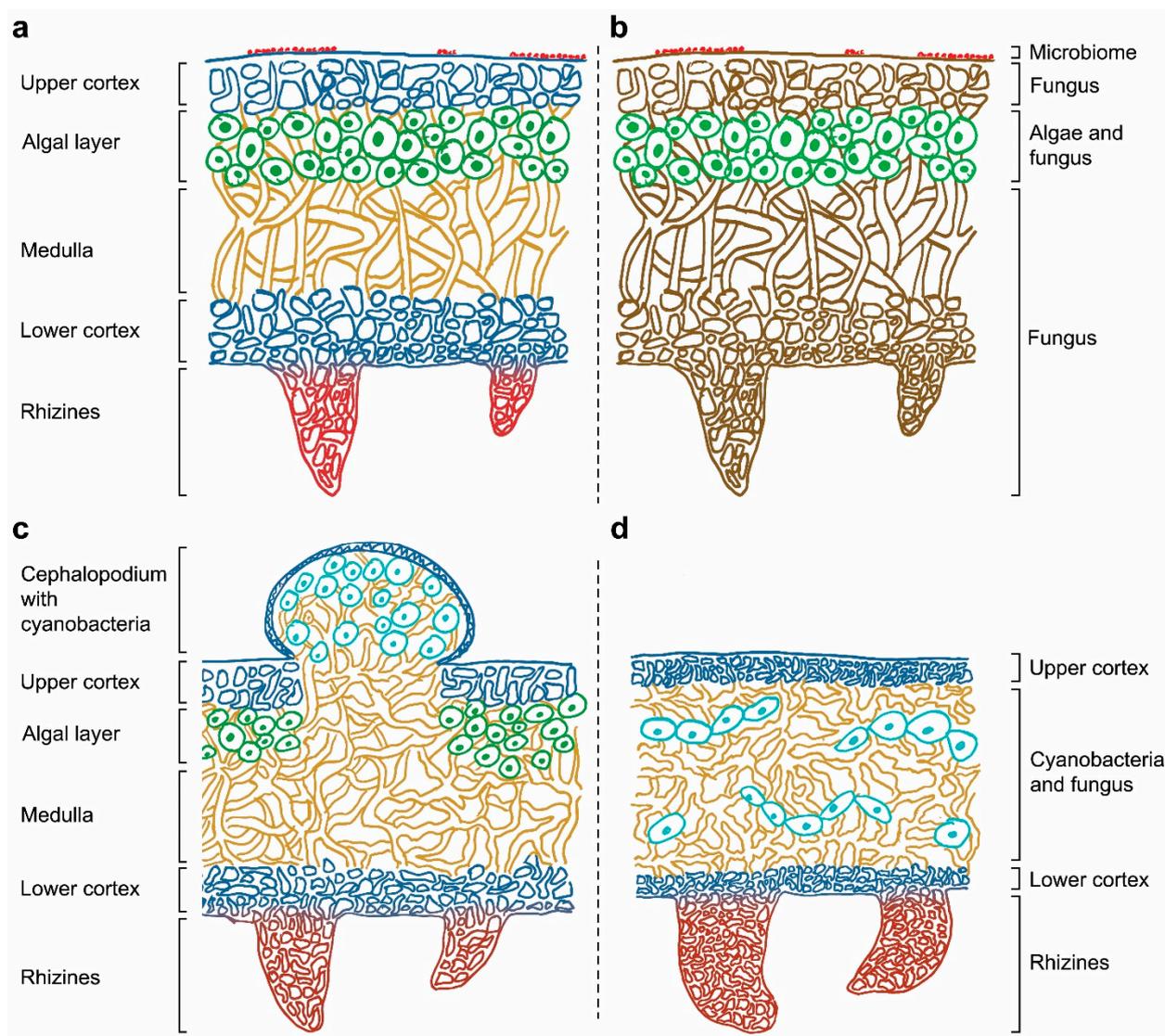


Figure 1. (a) Correspondence between the lichen structure and (b) their individual components in a heteromerous thallus. Depiction of tripartite lichen (c) and homoioimerous lichen (d).

Thalli are specialized structures unique to lichen-forming fungi and are not observed when the mycobiont grows in isolation. The thalli hold the photosynthetic partners and are one of the most complex structures in the entire fungal kingdom. Thallus structures are mainly determined by the fungal partner and can be grouped into three main growth forms: crustose, foliose, or fruticose. Crustose lichens, lacking the lower cortex, are completely attached to the substrate, while foliose and fruticose lichens are only partially attached through anchor like structures such as rhizines and hapteres [16]. The internal space of the thallus can be made up of either an internal stratification with a fungal upper layer and underlying algal layer (heteromerous, Figure 1a), or an even distribution of the mycobiont and photobiont (homoioimerous, Figure 1d). Lichens do not have a waxy cuticle to isolate

the thalli from their surroundings and therefore everything in the lichen's environment is absorbed into its structure, including water and nutrients coming from air and rain. They also lack vascular tissues (such as xylem and phloem in plants) to distribute nutrients and water around their thalli.

The general structure of a heteromorous lichen is shown in Figure 1 and it is mainly composed of layers of fungus and alga. The upper cortex is the outer layer of the lichen thallus, and it is formed by the mycobiont. The cells in this layer are tightly packed to provide certain physical and chemical protection from the environment. The algal layer contains the photobiont, which frequently is a green alga. Cyanobacteria, if present, can be located in small vacuoles called cephalopodiums, which exist on top of the upper cortex or within the tissues of the lichen when there is a green algal layer already present (secondary photobiont, Figure 1c), or in a layer under the upper cortex (primary photobiont). Part of the lichen thallus is composed of filamentous fungal cells that form the medulla. This layer is loosely packed and has a threadlike structure. The lower cortex, with a similar structure to the upper cortex, protects the medulla and give support to the rhizines or other basal attachments that allow lichens to get linked to their substrate. Rhizines are fungal multicellular structures originated on the lower surface with no vascular purposes: no water or nutrients can be absorbed by them, and their unique function is to support the lichen attachment. Although this is the most frequent structural organization found in a thallus, some lichens present no distinguishable layers of mycobiont and photobiont. In these cases, the components are distributed in one big uniform layer, resulting in a gelatinous growth form (Figure 1d).

Most lichen-forming fungi belong to the phylum Ascomycota, while only 0.3% of lichenized fungi are known to be derived from Basidiomycota [8,17]. The majority of lichen-forming algae belong to the green algae (85%), and 10% have a cyanobacterium as primary photosynthetic partner, but also brown or yellow-green algae have been identified in these relationships [18,19]. Sexual reproduction of the fungal partner involves the growth of fruiting bodies from the thallus which produce ascospores to be dispersed. In addition to this sexual mechanism, lichens have also developed other processes of asexual reproduction to disperse both partners together in varied and specific joint propagules [16].

2. From a Dual Partnership to a Multi-Species Symbiotic Relationship

In this intimate and long-term partnership, the fungal partner provides water, mineral nutrients, and sheltering structures for the photobiont, which contributes photosynthetically fixed carbon as the energy source for the system. Although the fungus can be very specific when selecting its photobiont [20,21], generalist fungi have also been frequently described [22–25]. Yahr et al. [26] established three categories according to the range of photobionts with which they are able to lichenize: photobiont specialists, intermediates and generalists. Following this scheme, photobiont specialists partner with single algal lineages, while photobiont generalists accept a high variety of algal partners and can establish associations with a number of strains according to environmental conditions. Intermediates form symbiotic relationships with a reduced number of algal partners. Photobiont generalists are frequently associated to lichens with wide both geographical range and ecological niches, which can associate with locally adapted photobionts in different climatic regions [25,27,28]. The exact factors that determine the photobiont selection are well known and appear to be related to phylogenetic specialization, fungus reproductive strategy, photobiont cell availability and ecological factors such as climate or substrate [29,30]. Surprisingly, multiple algal species have been observed in association with the same thallus in a large number of studies [24,31,32]. For example, Backor et al. [33] confirmed the presence of multiple algal genotypes in a single lichen thallus, Casano et al. [34] found that *Ramalina farinacea* thalli represent a specific and selective form of symbiotic association involving the same two *Trebouxia* phycobionts, and Del Campo et al. [35] concluded that ecological diversification and speciation of lichen symbionts in different habitats could include a transient phase consisting of associations with more than one photobiont in individual thalli. This

pattern of algal coexistence is probably promoted by their different and complementary ecophysiological responses which facilitate the proliferation of the lichen in a wide range of habitats and geographic areas [34].

In the last decade, it has been shown that lichens are far from being a simple association between two unrelated organismal groups, and instead involve a bacterial (and other fungi including yeast) component which is a key contributor to the biology of the holobiont. The inclusion of bacteria within the lichen partnership was first observed around the 1930's [36–38] (Figure 2), and it has been described as a discontinuous monolayer on the thallus surface (Figure 1). At that time, rudimentary methods only facilitated associating these bacteria with a possible and unspecific role in nitrogen fixation. At the beginning of the 21st century, the first molecular analyses started using bacterial isolates (e.g., Gonzalez et al. [39]). However, as culture-dependent methods can only account for 0.001–15% of the bacterial diversity [40], most microorganisms had remained unrevealed [41]. More recent studies allowed the observation that the structure of the nitrogen-fixing bacteria present in the cyanolichens is different from that of chlorolichens and that chlorolichens have a higher diversity of nitrogen-fixing bacteria than cyanolichens [42]. Also, in addition to the main cyanobiont, other cyanobacteria have been found within the microbiota of lichen thalli and substrates [43,44], which can also contribute with part of the nitrogen input to the symbiosis.

Once new methods were developed to complement the previously applied techniques, the limitations of the bacterial isolation were overcome, which allowed researchers to holistically explore the bacterial community. Fingerprinting techniques [45] and molecular cloning methods (e.g., Hodkinson and Lutzoni, [46]) allowed the production of microbial community profiles of lichen-associated microbiota. Bates et al. [47] revealed the microbial community associated with lichens based on next generation pyrosequencing for the first time. Thus, multi-omics approaches, that allow the integration of multiple omics research and datasets explaining the mechanisms underlying biological processes and molecular functions [48], along with bioinformatic tools have put the spotlight on the host-specific bacterial microbiome [49–52]. Recent findings about bacterial associations with lichens support their relationship as a multi-species symbiosis in which different roles are played by an increasingly recognized diversity of organisms associated with the thalli (see [49] and references therein; Figure 2).

3. Potential Roles of Recently Discovered Partners

Bacterial and secondary fungal communities inhabiting lichens have been found through next-generation sequencing techniques (e.g., Grube et al. [45], Tuovinen et al. [53]). Key roles as functional components in structuring the thalli and modulating the response to environmental factors could be performed by these overlooked organisms [49,50,54]. Since the first half of the 20th century, individual strains of bacteria have been isolated from lichens, with Alpha proteobacteria making up the dominant group, first observed by Cardinale et al. [55]. Studies of the diversity of lichen-associated bacteria suggest that different parts of the thallus, providing different chemical and physiological micro-niches, can influence microbial colonization ([45,46] and references therein). Another main factor driving the bacterial composition is the fungal partner. Accordingly, Aschenbrenner et al. [56] demonstrated that the lichen *Lobaria pulmonaria* presented a core and shared fraction of its bacterial biome, as well as a transient fraction. They also demonstrated that bacteria were present in the lichen's vegetative propagules which allowed them to vertically transmit through asexual reproduction. Traditionally, it has been accepted that the mycobiont builds up the thallus as a result of the specific interaction with a suitable algal partner, and then numerous associated and potentially interacting bacterial and other partners colonize the lichen more or less specifically. Recently, it has been hypothesized that the microbiome contributes to the lichenization process [57].

Similar questions to those posed regarding the role and specificity of bacteria within lichens have also been raised in relation to secondary fungal communities inhabiting lichens.

Spribille et al. [58] found that a specific group of basidiomycetous yeasts played a part in the microbiome of two epiphytic lichens (Figure 2), and that the abundance of the yeast within the lichen was correlated with concentrations of vulpinic acid, which is a secondary metabolite associated with lichen defenses. More recently, Cernajova and Skaloud [59] discovered previously unknown cystobasidiomycete symbionts in a number of *Cladonia* species in the northern hemisphere.

The consistent presence of basidiomycetous yeast within lichens in these studies suggested these were the third partner within the lichen complex. However, Millanes et al. [60] considered *Cyphobasidium* spp. to be a lichen-related fungi that can form galls on the thalli instead of as a previously unseen third mutualistic partner. The relevance of these lichen-related yeasts was also discussed by Oberwinkler [61], who pointed out that “it is obvious that basidiomycetous yeasts in lichen thalli are not a third component of symbiosis, but rather the vegetative propagules of mycoparasites”. Supporting this view, Lendemmer et al. [51] failed to detect basidiomycete yeasts in over 97% out of 339 lichen species from the Appalachian Mountains in North America in a metagenomic study. Although their metagenomic approach is likely less sensitive than PCR assays with specific primers, these findings raise questions about the ubiquity and specificity of yeasts in lichens.

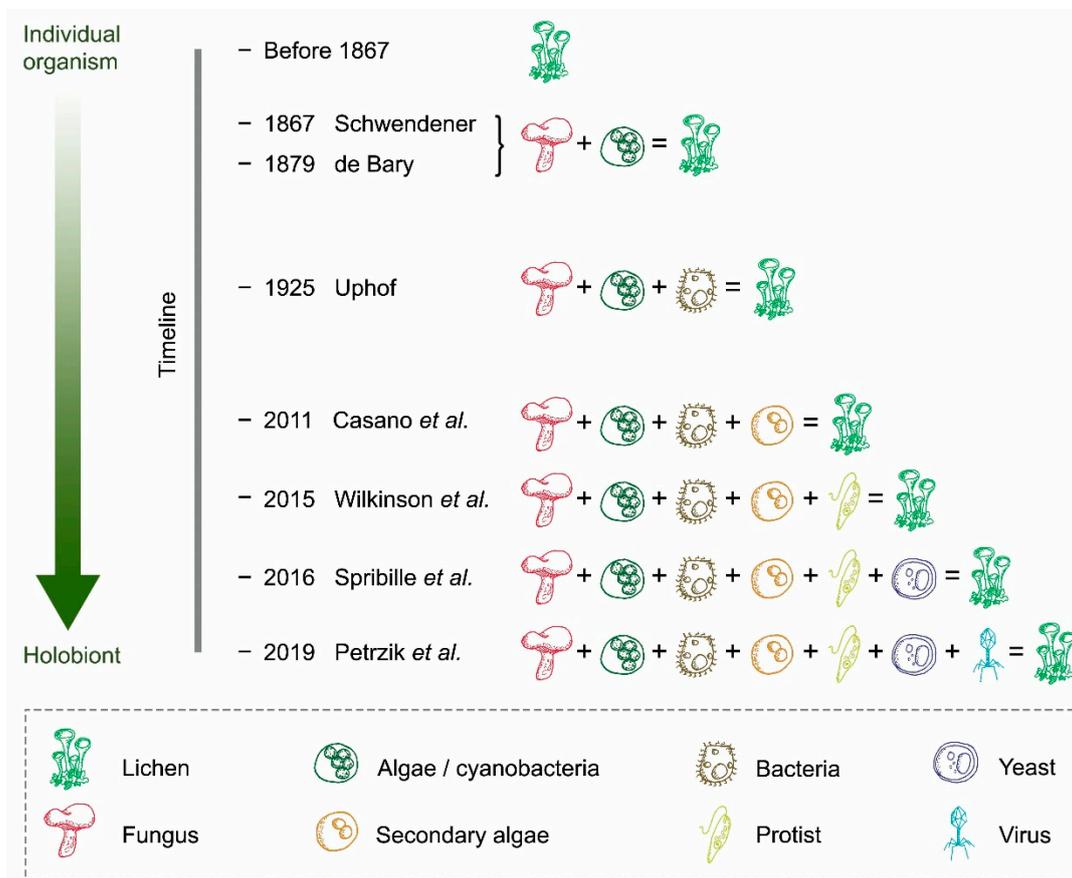


Figure 2. Temporal line indicating the timing at which the different components of the lichen partnership were discovered. This figure reflects the journey from considering lichens as a single organism to the complex multipartner symbiotic relationship known nowadays [9,10,34,36,58,62,63].

4. The Increasing Complexity Surrounding the Concept of Lichen

The discovery of potential new partners increased the complexity of species interactions within these supraorganisms. However, the lack of experimental evidence regarding the lichen microbiome hindered our ability to reveal the network of interactions within this holobiont. As opposed to what was traditionally believed, the photobiont should not be

limited to a single strain of algae [34] and protists and even viruses can form symbiotic associations with lichens [62,63] (Figure 2).

Conceptualizing a lichen means accounting for a vast array of related microorganisms, providing the ideal example of a holobiont, composed of a dominant mycobiont and diverse microbiome [64]. This evolved network of biotic connections whose morphology is shaped by the mycobiont, is at the service of the fitness of the entire superorganism. Hawksworth and Grube [65] re-defined the lichen symbiosis as: ‘a self-sustaining ecosystem formed by the interaction of an exhabitant fungus, an extracellular arrangement of one or more photosynthetic partners and an indeterminate number of other microscopic organisms’ [66].

While a detailed analysis of the scientific scrutiny focused on lichens is beyond the purpose of this text, a rough indication of the increasing interest in the lichen microbiome can be obtained by looking at the publications of the last 20 years (Figure 3). Although a linear relationship is missing, since 2014, the concept of a lichen symbiotic association, including more than just the mycobiont and the photobiont was well established. The development of “omic” technologies is a key element that has allowed the proliferation of studies in this research field [67]. The number of citations of a specific paper can increase with the time from its publication, which can explain the low number of citations for the most recent works; however, since 2015, the annual number of citations has exceeded 50, testifying to an increasing interest in this specialized topic.

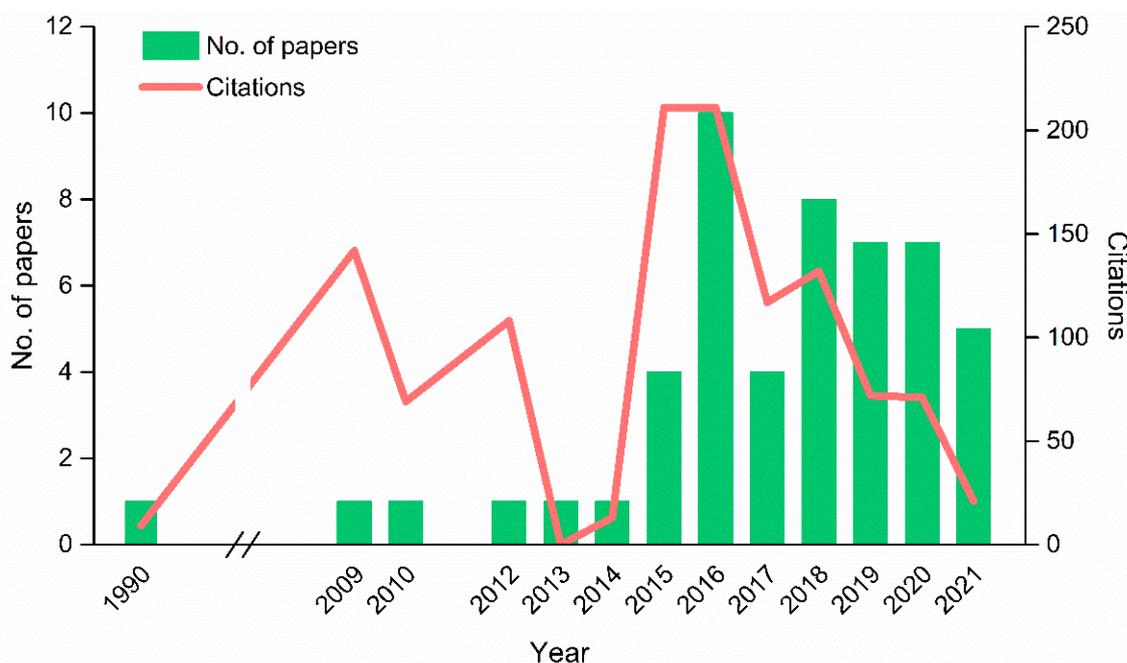


Figure 3. Number of papers related to the lichen microbiome and their citations. Data obtained using the search terms “lichen microbiome” or “lichen microbiota” or “lichen bacterial community” or “lichen microbial” or “lichen bacteria” or “lichen-associated bacteria” in the Web of Science (<https://www.webofscience.com/wos/woscc/basic-search>) accessed on 13 February 2022. All results were checked to ensure that they were referred to the presence of microorganisms in the lichen symbiosis.

5. Mutualism or Parasitism?

Although lichens are usually considered mutualistic symbioses, many lichen characteristics identify them as controlled parasitic interactions [68,69]. Initial studies in this field deemed lichens as algae parasitized by fungi because they found algal cells in a lichen thallus which were dead or penetrated by fungal haustoria. However, other authors considered lichens as mutualists based on the seemingly healthy and long-lasting

relationship among its partners. Two different experimental approaches have been developed to study selectivity among the lichen partners. The first one is based on in vitro resynthesis of the independently cultured mycobiont with different photobiont species. An alternative approach involves the assessment of the specimens collected in different geographical areas and the identification of the partners present in the lichen thalli [70]. In vitro studies give researchers the opportunity to observe initial stages of the lichenization process. Several developmental stages of lichenization have been described depending on the interacting alga, and timing of these events is variable depending on the species, media, and incubation conditions. The first developmental stage which has been described is the “pre-contact” stage, crucial for the establishment of symbiont recognition mechanisms and biont specificity [71,72], where the partners are in close proximity to share extracellular secretions but not physical contact. In a second phase, the “contact” stage, the two bionts start making physical contact by fungal appressoria (which are flattened hyphal tips that bind to the host cell surface and start a penetration peg), whereas, in a third phase termed the “growth together” stage, the two partners grow together in a network to form cellular masses containing both bionts [73–76].

Attempts of resynthesis in the laboratory starting from the isolated partners have been made with various and inconsistent outcomes [52,68,69,77–86], making the study of lichenization mechanisms hard due to the lack of consistently repeatable results. In vitro re-synthesis experiments showed that the interaction of a mycobiont with its compatible algal partner triggers specific morphological differentiation that is not seen when in contact with incompatible algal partners [68,71,87]. Conversely, parasitic behaviors of the fungus can be observed in interaction with nonlichenized algae or lichenized algae from different lichen species [68,71,87,88]. Switching to parasitism and saprotrophic nutrition is also known in some lichens, like *Ochrolechia frigida* when growing without its alga partner. In algal-free stages, this species seems to be capable of saprotrophic nutrition on mosses, phanerogams and other lichens [89].

In agreement with an optional parasitic or saprotrophic lifestyle of the mycobiont, Munzi et al. recorded for the first time in lichens a high activity of extracellular enzymes able to digest organic matter of different types and that are usually found in mycorrhizal fungi, which also alternate between different lifestyles (unpublished). A common characteristic of axenic reconstitutions is either that they do not progress beyond the soredia or squamule stages or, if they do, the resulting thalli do not resemble closely the corresponding natural lichens in shape, size, and full differentiation. Gene expression studies [72] and specific exudation patterns of lichen photobionts [90] also point to extracellular communication between lichen symbionts without cellular contact [57]. For example, ribitol was capable of overcoming fungal growth arrest [90], fungal lectins induced chemotropism of compatible *Nostoc* cells in cyanolichens [91], and chitinase, a defense enzyme in plants against pathogens, was downregulated in the photobiont during resynthesis stages [87]. Interestingly, the results of proteomic analyses in the thalli of the lichen *Xanthoria parietina* [92] included expression of proteins linked to the signaling compound pathways mentioned above.

Our knowledge about the cytological and biochemical interactions between the symbionts in lichens is still scarce [88]. However, increasing evidence indicates conservation of signaling pathways involved in the establishment of other major symbioses between plants and mutualistic microbes. Both common effectors and genes have been found to be essential for the establishment of rhizobial, Frankia, mycorrhizal and fungal endophytic symbioses, including plant-produced strigolactones, microbial partner-produced chitoooligosaccharides (COs) and lipo-chitoooligosaccharides (LCOs) [93–99], genes encoding Vapyrin [93,100,101] and several transcription factors [102,103]. Three of these transcription factors (CYCLOPS, NSP1, and NSP2) are well conserved between actinorhizal, legume, non-legume, and mycorrhizal symbioses [96]. It is therefore possible that some or all these factors are also present and play essential roles in lichenization, and their absence or differential presence may relate to re-synthesis failures or establishment of associations with parasitic outcomes.

6. Conclusions and Prospects

A large scientific effort is still needed to achieve the goal of revealing the physiological mechanisms operating in convoluted lichen symbioses and the roles of the various organisms involved in this complex holobiont. As the latest scientific evidence has shown, this research challenge must be addressed by considering lichens as self-sustained and adaptable systems of partnerships, just like us!

Author Contributions: Conceptualization, S.M. and L.M.; methodology, S.M. and L.M.; software, J.R.; validation, S.M., L.M., J.R. and C.C.; formal analysis, J.R.; investigation, L.M.; resources, C.C., L.M. and S.M.; data curation, L.M. and J.R.; writing—original draft preparation, L.M., J.R. and S.M.; writing—review and editing, S.M., L.M., J.R. and C.C.; visualization, L.M. and J.R.; supervision, C.C. and S.M.; project administration, L.M.; funding acquisition, L.M. All authors have read and agreed to the published version of the manuscript.

Funding: This project has received funding from the European Union’s Horizon 2020 Research and Innovation programme under the Marie Skłodowska–Curie grant agreement #793965 (Med-N-Change).

Acknowledgments: We are grateful to Lucy Sheppard for language revision and useful insights.

Conflicts of Interest: The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

References

1. Agerer, R. Characterization of Ectomycorrhiza. *Methods Microbiol.* **1991**, *23*, 25–73.
2. Rasmussen, H.N. Recent developments in the study of orchid mycorrhiza. *Plant Soil* **2002**, *244*, 149–163. [[CrossRef](#)]
3. Hawksworth, D.L. Observations on three algicolous microfungi. *Notes R. Bot. Gard. Edinburgh* **1987**, *44*, 549–560.ill.
4. Hawksworth, D.L. The variety of fungal-algal symbioses, their evolutionary significance, and the nature of lichens. *Bot. J. Linn. Soc.* **1988**, *96*, 3–20. [[CrossRef](#)]
5. Kohlmeyer, J.; Demoulin, D. Parasitic and Symbiotic Fungi on Marine Algae. *Bot. Mar.* **1981**, *24*, 9–18. [[CrossRef](#)]
6. Sønstebo, J.H.; Rohrlack, T. Possible implications of Chytrid parasitism for population subdivision in freshwater cyanobacteria of the genus *Planktothrix*. *Appl. Environ. Microbiol.* **2011**, *77*, 1344–1351. [[CrossRef](#)]
7. Kohlmeyer, J.; Kohlmeyer, E. Is Ascophyllum nodosum Lichenized? *Bot. Mar.* **1972**, *15*, 109–112. [[CrossRef](#)]
8. Nash, T.H.I. *Lichen Biology*, 2nd ed.; Cambridge University Press: Cambridge, UK, 2008; ISBN 9780521871624.
9. Schwendener, S. *Die Algentyphen der Flechtengonidien*; Universitätsbuchdruckerei von C. Schultze: Basel, Switzerland, 1869.
10. Oulhen, N.; Schulz, B.J.; Carrier, T.J. English translation of Heinrich Anton de Bary’s 1878 speech, ‘Die Erscheinung der Symbiose’ (‘De la symbiose’). *Symbiosis* **2016**, *69*, 131–139. [[CrossRef](#)]
11. Rikkinen, J. Molecular studies on cyanobacterial diversity in lichen symbioses. *MycKeys* **2013**, *6*, 3–32. [[CrossRef](#)]
12. Margulis, L.; Fester, R. *Symbiosis as a Source of Evolutionary Innovation: Speciation and Morphogenesis*; Margulis, L., Fester, R., Eds.; MIT Press: Cambridge, MA, USA, 1991; ISBN 0262132699.
13. Douglas, A.E.; Werren, J.H. Holes in the hologenome: Why host-microbe symbioses are not holobionts. *mBio* **2016**, *7*, e02099-15. [[CrossRef](#)]
14. Rafferty, N.E.; Caradonna, P.J.; Bronstein, J.L. Phenological shifts and the fate of mutualisms. *Oikos* **2015**, *124*, 14–21. [[CrossRef](#)]
15. Chomicki, G.; Renner, S.S. Partner abundance controls mutualism stability and the pace of morphological change over geologic time. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, 3951–3956. [[CrossRef](#)]
16. Büdel, B.; Scheidegger, C. Thallus morphology and anatomy. In *Lichen Biology*; Nash, T.H.I., Ed.; Cambridge University Press: Cambridge, UK, 2008; pp. 40–68, ISBN 9780521871624.
17. Lücking, R.; Dal-Forno, M.; Sikaroodi, M.; Gillevet, P.M.; Bungartz, F.; Moncada, B.; Yáñez-Ayabaca, A.; Chaves, J.L.; Coca, L.F.; Lawrey, J.D. A single macrolichen constitutes hundreds of unrecognized species. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, 11091–11096. [[CrossRef](#)] [[PubMed](#)]
18. Honegger, R. Lichen-Forming Fungi and Their Photobionts. In *Plant Relationships*; Deising, H.B., Ed.; Springer: Berlin/Heidelberg, Germany, 2009; pp. 307–333, ISBN 978-3-540-87407-2.
19. Henskens, F.L.; Green, T.G.A.; Wilkins, A. Cyanolichens can have both cyanobacteria and green algae in a common layer as major contributors to photosynthesis. *Ann. Bot.* **2012**, *110*, 555–563. [[CrossRef](#)]
20. Piercey-Normore, M.D.; DePriest, P.T. Algal switching among lichen symbioses. *Am. J. Bot.* **2001**, *88*, 1490–1498. [[CrossRef](#)]
21. Magain, N.; Miadlikowska, J.; Goffinet, B.; Serusiaux, E.; Lutzoni, F. Macroevolution of specificity in cyanolichens of the genus *Peltigera* section *Polydactylon* (Lecanoromycetes, Ascomycota). *Syst. Biol.* **2017**, *66*, 74–99. [[CrossRef](#)]

22. Wirtz, N.; Lumbsch, H.T.; Green, T.G.A.; Türk, R.; Pintado, A.; Sancho, L.; Schroeter, B. Lichen fungi have low cyanobiont selectivity in maritime Antarctica. *New Phytol.* **2003**, *160*, 177–183. [[CrossRef](#)] [[PubMed](#)]
23. Guzow-Krzemińska, B. Photobiont flexibility in the lichen *Protoparmeliopsis muralis* as revealed by ITS rDNA analyses. *Lichenol.* **2006**, *38*, 469–476. [[CrossRef](#)]
24. Muggia, L.; Vancurova, L.; Škaloud, P.; Peksa, O.; Wedin, M.; Grube, M. The symbiotic playground of lichen thalli—A highly flexible photobiont association in rock-inhabiting lichens. *FEMS Microbiol. Ecol.* **2013**, *85*, 313–323. [[CrossRef](#)]
25. Blaha, J.; Baloch, E.; Grube, M. High photobiont diversity associated with the euryoecious lichen-forming ascomycete *Lecanora rupicola* (Lecanoraceae, Ascomycota). *Biol. J. Linn. Soc.* **2006**, *88*, 283–293. [[CrossRef](#)]
26. Yahr, R.; Vilgalys, R.; DePriest, P.T. Geographic variation in algal partners of *Cladonia subtenuis* (Cladoniaceae) highlights the dynamic nature of a lichen symbiosis. *New Phytol.* **2006**, *171*, 847–860. [[CrossRef](#)]
27. Fernández-Mendoza, F.; Domaschke, S.; García, M.A.; Jordan, P.; Martín, M.P.; Printzen, C. Population structure of mycobionts and photobionts of the widespread lichen *Cetraria aculeata*. *Mol. Ecol.* **2011**, *20*, 1208–1232. [[CrossRef](#)]
28. Vargas Castillo, R.; Beck, A. Photobiont selectivity and specificity in *Caloplaca* species in a fog-induced community in the Atacama Desert, northern Chile. *Fungal Biol.* **2012**, *116*, 665–676. [[CrossRef](#)]
29. Scheidegger, C. Systematische Studien zur Krustenflechte *Anzina carneonivea* (Trapeliaceae, Lecanorales). *Nov. Hedwigia* **1985**, *41*, 191–218.
30. Hoz, C.J.P.-D.L.; Magain, N.; Lutzoni, F.; Goward, T.; Restrepo, S.; Miadlikowska, J. Contrasting Symbiotic Patterns in Two Closely Related Lineages of Trimembered Lichens of the Genus *Peltigera*. *Front. Microbiol.* **2018**, *9*, 2770. [[CrossRef](#)]
31. del Campo, E.M.; Casano, L.M.; Gasulla, F.; Barreno, E. Suitability of chloroplast LSU rDNA and its diverse group I introns for species recognition and phylogenetic analyses of lichen-forming *Trebouxia* algae. *Mol. Phylogenet. Evol.* **2010**, *54*, 437–444. [[CrossRef](#)] [[PubMed](#)]
32. Moya, P.; Molins, A.; Chiva, S.; Bastida, J.; Barreno, E. Symbiotic microalgal diversity within lichenicolous lichens and crustose hosts on Iberian Peninsula gypsum biocrusts. *Sci. Rep.* **2020**, *10*, 14060. [[CrossRef](#)]
33. Bačkor, M.; Peksa, O.; Škaloud, P.; Bačkorová, M. Photobiont diversity in lichens from metal-rich substrata based on ITS rDNA sequences. *Ecotoxicol. Environ. Saf.* **2010**, *73*, 603–612. [[CrossRef](#)]
34. Casano, L.M.; Del Campo, E.M.; García-Breijó, F.J.; Reig-Armiñana, J.; Gasulla, F.; Del Hoyo, A.; Guéra, A.; Barreno, E. Two *Trebouxia* algae with different physiological performances are ever-present in lichen thalli of *Ramalina farinacea*. Coexistence versus Competition? *Environ. Microbiol.* **2011**, *13*, 806–818. [[CrossRef](#)]
35. del Campo, E.M.; Catalá, S.; Gimeno, J.; del Hoyo, A.; Martínez-Alberola, F.; Casano, L.M.; Grube, M.; Barreno, E. The genetic structure of the cosmopolitan three-partner lichen *Ramalina farinacea* evidences the concerted diversification of symbionts. *FEMS Microbiol. Ecol.* **2013**, *83*, 310–323. [[CrossRef](#)]
36. Uphof, J.C.T. Purple bacteria as symbionts of a lichen. *Science* **1925**, *61*, 67. [[CrossRef](#)] [[PubMed](#)]
37. Henkel, P.A.; Yuzhakova, L.A. Nitrogen-fixing bacteria in lichens. *Izv. Biol. Inst. Permsk. Gos. Univ.* **1936**, *10*, 9–10.
38. Iskina, R.Y. On nitrogen fixing bacteria in lichens. *Isv. Biol. Inst. Permsk.* **1938**, *11*, 133–139.
39. González, I.; Ayuso-Sacido, A.; Anderson, A.; Genilloud, O. Actinomycetes isolated from lichens: Evaluation of their diversity and detection of biosynthetic gene sequences. *FEMS Microbiol. Ecol.* **2005**, *54*, 401–415. [[CrossRef](#)]
40. Amann, R.I.; Ludwig, W.; Schleifer, K.-H. Phylogenetic identification and in situ detection of individual microbial cells without cultivation. *Microbiol. Rev.* **1995**, *59*, 143–169. [[CrossRef](#)]
41. Rappé, M.S.; Giovannoni, S.J. The Uncultured Microbial Majority. *Annu. Rev. Microbiol.* **2003**, *57*, 369–394. [[CrossRef](#)]
42. Almendras, K.; García, J.; Carú, M.; Orlando, J. Nitrogen-fixing bacteria associated with *Peltigera* cyanolichens and *Cladonia* chlorolichens. *Molecules* **2018**, *23*, 3077. [[CrossRef](#)]
43. Zúñiga, C.; Leiva, D.; Carú, M.; Orlando, J. Substrates of *Peltigera* Lichens as a Potential Source of Cyanobionts. *Microb. Ecol.* **2017**, *74*, 561–569. [[CrossRef](#)]
44. Graham, L.E.; Trest, M.T.; Will-Wolf, S.; Mücke, N.S.; Atonio, L.M.; Piotrowski, M.J.; Knack, J.J. Microscopic and metagenomic analyses of *peltigera ponojensis* (Peltigerales, ascomycota). *Int. J. Plant Sci.* **2018**, *179*, 241–255. [[CrossRef](#)]
45. Grube, M.; Cardinale, M.; De Castro, J.V., Jr.; Müller, H.; Berg, G. Species-specific structural and functional diversity of bacterial communities in lichen symbioses. *ISME J.* **2009**, *3*, 1105–1115. [[CrossRef](#)] [[PubMed](#)]
46. Hodkinson, B.P.; Lutzoni, F. A microbiotic survey of lichen-associated bacteria reveals a new lineage from the Rhizobiales. *Symbiosis* **2009**, *49*, 163–180. [[CrossRef](#)]
47. Bates, S.T.; Cropsey, G.W.G.; Caporaso, J.G.; Knight, R.; Fierer, N. Bacterial communities associated with the lichen symbiosis. *Appl. Environ. Microbiol.* **2011**, *77*, 1309–1314. [[CrossRef](#)] [[PubMed](#)]
48. Krassowski, M.; Das, V.; Sahu, S.K.; Misra, B.B. State of the Field in Multi-Omics Research: From Computational Needs to Data Mining and Sharing. *Front. Genet.* **2020**, *11*, 610798. [[CrossRef](#)] [[PubMed](#)]
49. Grube, M.; Cernava, T.; Soh, J.; Fuchs, S.; Aschenbrenner, I.; Lassek, C.; Wegner, U.; Becher, D.; Riedel, K.; Sensen, C.W.; et al. Exploring functional contexts of symbiotic sustain within lichen-associated bacteria by comparative omics. *ISME J.* **2015**, *9*, 412–424. [[CrossRef](#)] [[PubMed](#)]
50. Cernava, T.; Erlacher, A.; Aschenbrenner, I.A.; Krug, L.; Lassek, C.; Riedel, K.; Grube, M.; Berg, G. Deciphering functional diversification within the lichen microbiota by meta-omics. *Microbiome* **2017**, *5*, 82. [[CrossRef](#)]

51. Lendemer, J.C.; Keepers, K.G.; Tripp, E.A.; Pogoda, C.S.; McCain, C.M.; Kane, N.C. A taxonomically broad metagenomic survey of 339 species spanning 57 families suggests cystobasidiomycete yeasts are not ubiquitous across all lichens. *Am. J. Bot.* **2019**, *106*, 1090–1095. [[CrossRef](#)]
52. Kono, M.; Kon, Y.; Ohmura, Y.; Satta, Y.; Terai, Y. In vitro resynthesis of lichenization reveals the genetic background of symbiosis-specific fungal-algal interaction in *Usnea hakonensis*. *BMC Genom.* **2020**, *21*, 671. [[CrossRef](#)]
53. Tuovinen, V.; Ekman, S.; Thor, G.; Vanderpool, D.; Spribille, T.; Johannesson, H. Two Basidiomycete Fungi in the Cortex of Wolf Lichens. *Curr. Biol.* **2019**, *29*, 476–483.e5. [[CrossRef](#)]
54. Noh, H.-J.; Park, Y.; Hong, S.G.; Lee, Y.M. Diversity and Physiological Characteristics of Antarctic Lichens-Associated Bacteria. *Microorganisms* **2021**, *9*, 607. [[CrossRef](#)] [[PubMed](#)]
55. Cardinale, M.; Vieira De Castro Jr., J.; Müller, H.; Berg, G.; Grube, M. In situ analysis of the bacterial community associated with the reindeer lichen *Cladonia arbuscula* reveals predominance of Alphaproteobacteria. *FEMS Microbiol. Ecol.* **2008**, *66*, 63–71. [[CrossRef](#)]
56. Aschenbrenner, I.A.; Cernava, T.; Berg, G.; Grube, M. Understanding microbial multi-species symbioses. *Front. Microbiol.* **2016**, *7*, 180. [[CrossRef](#)]
57. Spribille, T.; Tagirdzhanova, G.; Goyette, S.; Tuovinen, V.; Case, R.; Zandberg, W.F. 3D biofilms: In search of the polysaccharides holding together lichen symbioses. *FEMS Microbiol. Lett.* **2020**, *367*, fnaa023. [[CrossRef](#)]
58. Spribille, T.; Tuovinen, V.; Resl, P.; Vanderpool, D.; Wolinski, H.; Aime, M.C.; Schneider, K.; Stabenheimer, E.; Toome-Heller, M.; Thor, G.; et al. Basidiomycete yeasts in the cortex of ascomycete macrolichens. *Science* **2016**, *353*, 488–492. [[CrossRef](#)]
59. Černajová, I.; Škaloud, P. The first survey of Cystobasidiomycete yeasts in the lichen genus *Cladonia*; with the description of *Lichenozyma pisutiana* gen. nov., sp. nov. *Fungal Biol.* **2019**, *123*, 625–637. [[CrossRef](#)]
60. Millanes, A.M.; Diederich, P.; Wedin, M. *Cyphobasidium* gen. nov., a new lichen-inhabiting lineage in the Cystobasidiomycetes (Pucciniomycotina, Basidiomycota, Fungi). *Fungal Biol.* **2016**, *120*, 1468–1477. [[CrossRef](#)]
61. Oberwinkler, F. Yeasts in Pucciniomycotina. *Mycol. Prog.* **2017**, *16*, 831–856. [[CrossRef](#)]
62. Wilkinson, D.M.; Creevy, A.L.; Kalu, C.L.; Schwartzman, D.W. Are heterotrophic and silica-rich eukaryotic microbes an important part of the lichen symbiosis? *Mycology* **2015**, *6*, 4–7. [[CrossRef](#)]
63. Petrzik, K.; Koloniuk, I.; Sehadová, H.; Sarkisova, T. Chrysovirus inhabited symbiotic fungi of lichens. *Viruses* **2019**, *11*, 1120. [[CrossRef](#)]
64. Simon, J.-C.; Marchesi, J.R.; Mougél, C.; Selosse, M.-A. Host-microbiota interactions: From holobiont theory to analysis. *Microbiome* **2019**, *7*, 5. [[CrossRef](#)] [[PubMed](#)]
65. Hawksworth, D.L.; Grube, M. Lichens redefined as complex ecosystems. *New Phytol.* **2020**, *227*, 1281–1283. [[CrossRef](#)] [[PubMed](#)]
66. Honegger, R. The symbiotic phenotype of lichen-forming ascomycetes and their endo- and epibionts. In *Fungal Associations*, 2nd ed.; Springer Berlin Heidelberg; Institute of Plant Biology, University of Zürich: Zürich, Switzerland, 2012; Volume 9, pp. 287–339, ISBN 9783642308260.
67. Grimm, M.; Grube, M.; Schiefelbein, U.; Zühlke, D.; Bernhardt, J.; Riedel, K. The Lichens' Microbiota, Still a Mystery? *Front. Microbiol.* **2021**, *12*, 714. [[CrossRef](#)] [[PubMed](#)]
68. Ahmadjian, V.; Jacobs, J.B. Relationship between fungus and alga in the lichen *Cladonia cristatella* Tuck. *Nature* **1981**, *289*, 169–172. [[CrossRef](#)]
69. Athukorala, S.N.P.; Huebner, E.; Piercey-Normore, M.D. Identification and comparison of the 3 early stages of resynthesis for the lichen *Cladonia rangiferina*. *Can. J. Microbiol.* **2014**, *60*, 41–52. [[CrossRef](#)] [[PubMed](#)]
70. Honegger, R. Morphogenesis. In *Lichen Biology*; Nash, T.H., III, Ed.; Cambridge University Press: Cambridge, UK, 2008; pp. 69–93, ISBN 9780521871624.
71. Meeßen, J.; Ott, S. Recognition mechanisms during the pre-contact state of lichens: I. Mycobiont-photobiont interactions of the mycobiont of *Fulgensia bracteata*. *Symbiosis* **2013**, *59*, 121–130. [[CrossRef](#)]
72. Joneson, S.; Armaleo, D.; Lutzoni, F. Fungal and algal gene expression in early developmental stages of lichen-symbiosis. *Mycologia* **2011**, *103*, 291–306. [[CrossRef](#)] [[PubMed](#)]
73. Ahmadjian, V.; Jacobs, J.B.; Russell, L.A. Scanning Electron Microscope Study of Early Lichen Synthesis. *Science* **1978**, *200*, 1062–1064. [[CrossRef](#)] [[PubMed](#)]
74. Galun, M. Lichenization. In *CRC Handbook of Lichenology*; Galun, M., Ed.; CRC Press: Boca Raton, FL, USA, 1988; Volume II, pp. 153–169.
75. Armaleo, D. Experimental Microbiology of Lichens—Mycelia Fragmentation, a Novel Growth Chamber, and the Origins of Thallus Differentiation. *Symbiosis* **1991**, *11*, 163–177.
76. Joneson, S.; Lutzoni, F. Compatibility and thigmotropism in the lichen symbiosis: A reappraisal. *Symbiosis* **2009**, *47*, 109–115. [[CrossRef](#)]
77. Yoshimura, I.; Kurokawa, T.; Yamamoto, Y.; Kinoshita, Y. Development of Lichen Thalli in Vitro. *Bryologist* **1993**, *96*, 412–421. [[CrossRef](#)]
78. Kon, Y.; Kashiwadani, H.; Masada, M.; Tamura, G. Artificial Syntheses of Mycobionts of *Usnea confusa* ssp. *kitamiensis* and *Usnea orientalis* with Their Natural and Non-Natural Phycobiont. *J. Jpn. Bot.* **1993**, *68*, 129–137.
79. Stocker-Wörgötter, E. Experimental studies of the lichen symbiosis: DNA-analyses, differentiation and secondary chemistry of selected mycobionts, artificial resynthesis of two- and tripartite symbioses. *Symbiosis* **2001**, *30*, 207–227.

80. Trembley, M.L.; Ringli, C.; Honegger, R. Morphological and molecular analysis of early stages in the resynthesis of the lichen *Baeomyces rufus*. *Mycol. Res.* **2002**, *106*, 768–776. [[CrossRef](#)]
81. Guzow-Krzemińska, B.; Stocker-Wörgötter, E. In vitro culturing and resynthesis of the mycobiont *Prototermeliopsis muralis* with algal bionts. *Lichenologist* **2013**, *45*, 65–76. [[CrossRef](#)]
82. Ahmadjian, V.; Russell, L.A.; Hildreth, K.C. Artificial Reestablishment of Lichens. I. Morphological Interactions Between the Phycobionts of Different Lichens and the Mycobionts *Cladonia cristatella* and *Lecanora Chrysoleuca*. *Mycologia* **1980**, *72*, 73–89. [[CrossRef](#)]
83. Bubrick, P.; Galun, M. Spore to spore resynthesis of *Xanthoria Parietina*. *Lichenologist* **1986**, *18*, 47–49. [[CrossRef](#)]
84. Culberson, C.F.; Culberson, W.L.; Johnson, A. Genetic and environmental effects of growth and production of secondary compounds in *Cladonia cristatella*. *Biochem. Syst. Ecol.* **1983**, *11*, 77–84. [[CrossRef](#)]
85. Stocker-Wörgötter, E.; Türk, R. Artificial cultures of the cyanobacterial lichen *Peltigera didactyla* (Peltigeraceae) in the natural environment. *Plant Syst. Evol.* **1989**, *165*, 39–48. [[CrossRef](#)]
86. Stocker-Wörgötter, E.; Türk, R. The resynthesis of thalli of *Dermatocarpon miniatum* under laboratory conditions. *Symbiosis* **1989**, *7*, 37–50.
87. Athukorala, S.N.P.; Piercey-Normore, M.D. Recognition-and defense-related gene expression at 3 resynthesis stages in lichen symbionts. *Can. J. Microbiol.* **2014**, *61*, 1–12. [[CrossRef](#)]
88. Insarova, I.D.; Blagoveshchenskaya, E.Y. Lichen symbiosis: Search and recognition of partners. *Biol. Bull.* **2016**, *43*, 408–418. [[CrossRef](#)]
89. Gaßmann, A.; Ott, S. Growth strategy and the gradual symbiotic interactions of the lichen *Ochrolechia frigida*. *Plant Biol.* **2000**, *2*, 368–378. [[CrossRef](#)]
90. Meeßen, J.; Eppenstein, S.; Ott, S. Recognition mechanisms during the pre-contact state of lichens: II. Influence of algal exudates and ribitol on the response of the mycobiont of *Fulgensia bracteata*. *Symbiosis* **2013**, *59*, 131–143. [[CrossRef](#)]
91. Díaz, E.M.; Vicente-Manzanares, M.; Sacristan, M.; Vicente, C.; Legaz, M.-E. Fungal lectin of *Peltigera canina* induces chemotropism of compatible *Nostoc* cells by constriction-relaxation pulses of cyanobiont cytoskeleton. *Plant Signal. Behav.* **2011**, *6*, 1525–1536. [[CrossRef](#)] [[PubMed](#)]
92. Munzi, S.; Gouveia, C.; Cruz, C.; Branquinho, C.; Coelho, A.V. Proteomic analysis contributes to unveil the mechanisms of nitrogen tolerance in the lichen *Xanthoria parietina*. *Not. Della Soc. Lichenol. Ital.* **2018**, *31*, 23.
93. Crosino, A.; Moscato, E.; Blangetti, M.; Carotenuto, G.; Spina, F.; Bordignon, S.; Puech-Pagès, V.; Anfossi, L.; Volpe, V.; Prandi, C.; et al. Extraction of short chain chitoooligosaccharides from fungal biomass and their use as promoters of arbuscular mycorrhizal symbiosis. *Sci. Rep.* **2021**, *11*, 3798. [[CrossRef](#)]
94. Volpe, V.; Carotenuto, G.; Berzero, C.; Cagnina, L.; Puech-Pagès, V.; Genre, A. Short chain chito-oligosaccharides promote arbuscular mycorrhizal colonization in *Medicago truncatula*. *Carbohydr. Polym.* **2020**, *229*, 115505. [[CrossRef](#)]
95. Feng, F.; Sun, J.; Radhakrishnan, G.V.; Lee, T.; Bozsóki, Z.; Fort, S.; Gavrin, A.; Gysel, K.; Thygesen, M.B.; Andersen, K.R.; et al. A combination of chitoooligosaccharide and lipochitoooligosaccharide recognition promotes arbuscular mycorrhizal associations in *Medicago truncatula*. *Nat. Commun.* **2019**, *10*, 5047. [[CrossRef](#)]
96. Diédhiou, I.; Diouf, D. Transcription factors network in root endosymbiosis establishment and development. *World J. Microbiol. Biotechnol.* **2018**, *34*, 37. [[CrossRef](#)]
97. Omoarelojie, L.O.; Van Staden, J. Plant-endophytic fungi interactions: A strigolactone perspective. *S. Afr. J. Bot.* **2020**, *134*, 280–284. [[CrossRef](#)]
98. Peláez-Vico, M.A.; Bernabéu-Roda, L.; Kohlen, W.; Soto, M.J.; López-Ráez, J.A. Strigolactones in the *Rhizobium*-legume symbiosis: Stimulatory effect on bacterial surface motility and down-regulation of their levels in nodulated plants. *Plant Sci.* **2016**, *245*, 119–127. [[CrossRef](#)]
99. Maclean, A.M.; Bravo, A.; Harrison, M.J. Plant signaling and metabolic pathways enabling arbuscular mycorrhizal symbiosis. *Plant Cell* **2017**, *29*, 2319–2335. [[CrossRef](#)] [[PubMed](#)]
100. Kuga, Y.; Schläppi, K.; Reinhardt, D. From Imaging to Functional Traits in Interactions between Roots and Microbes. In *Methods in Rhizosphere Biology Research*; Reinhardt, D., Sharma, A.K., Eds.; Springer: Singapore, 2019; pp. 227–239, ISBN 978-981-13-5767-1.
101. Liu, C.-W.; Breakspear, A.; Stacey, N.; Findlay, K.; Nakashima, J.; Ramakrishnan, K.; Liu, M.; Xie, F.; Endre, G.; de Carvalho-Niebel, F.; et al. A protein complex required for polar growth of rhizobial infection threads. *Nat. Commun.* **2019**, *10*, 2848. [[CrossRef](#)] [[PubMed](#)]
102. Radhakrishnan, G.V.; Keller, J.; Rich, M.K.; Vernié, T.; Mbadanga Mbadanga, D.L.; Vigneron, N.; Cottret, L.; Clemente, H.S.; Libourel, C.; Cheema, J.; et al. An ancestral signalling pathway is conserved in intracellular symbioses-forming plant lineages. *Nat. Plants* **2020**, *6*, 280–289. [[CrossRef](#)] [[PubMed](#)]
103. Boschiero, C.; Dai, X.; Lundquist, P.K.; Roy, S.; de Bang, T.C.; Zhang, S.; Zhuang, Z.; Torres-Jerez, I.; Udvardi, M.K.; Scheible, W.-R.; et al. MtSSPDB: The *Medicago truncatula* Small Secreted Peptide Database. *Plant Physiol.* **2020**, *183*, 399–413. [[CrossRef](#)] [[PubMed](#)]