

## Article

# Influence of Amitraz-Based Product Characteristics on Varroa Mite Population Control

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**Abstract:** The presence of the *Varroa destructor* mite requires the use of acaricide treatments for honeybee colonies. Amitraz is one of the most common acaricide-active ingredients used by beekeepers. Certain Varroa mite populations have developed resistance to amitraz, thereby leading to a loss in the efficacy of amitraz-based treatments. Two products, Apivar and Supatraz, were applied in the same apiary in France to evaluate their efficacy. Both treatments are amitraz-based but have different galenics. Thanks to field data, a dynamic model was used to simulate the actions of Apivar and Supatraz on the mite population. We considered two parameters to compare the products as follows: the daily mortality rate and the treatment duration. In the field, the percentage of the efficacy of the two products was not significantly different, but Supatraz kills mites faster and decreases 90% of the mite infestation in 28.4 days compared with 50.9 days when using Apivar. Through modeling, we showed the daily impact of the two different products on mite population. Supatraz has a higher daily mortality rate during the first two weeks than Apivar. Supatraz requires a lower efficacy (% of varroa mites killed during all the treatment) to stabilize the varroa mite population due to its faster release of active ingredients than Apivar, thereby needing a shorter period to achieve the same result. Depending on the model, Supatraz conserves effective efficacy when used against moderately resistant mites (with mite mortality being 40–70% at the LC90) but not against highly resistant mites (with mite mortality being <40% at the LC90). These results show that the comparison of the efficacy of the two products with different characteristics (duration of treatment and daily mortality rate) should be analyzed with caution.

**Keywords:** *Apis mellifera*; beekeeping; Apivar; Supatraz; modeling; resistance; efficacy; treatment



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## 1. Introduction

*Varroa destructor* is an ectoparasitic mite of honeybees responsible for varroosis [1]. It is also responsible for the spread of viruses in honeybee colonies [2,3]. Today, beekeepers must integrate varroa management to limit colony collapses. The control methods include the acaricide (chemical) and biotechnical methods [4]. Treatments should be sufficiently effective to significantly reduce the varroa mite population every year. However, a result of the lack of rotations of synthetic acaricides is that of the development of resistance to the acaricide substance by varroa mites [5]. Resistance to tau-fluvalinate, flumethrin, coumaphos and amitraz was detected in the laboratory [6–11] and in the field [11–13]. For the moment, synthetic acaricides are the most prone to resistance. Currently, no resistance to acaricides that are used in organic beekeeping has been demonstrated [5,14,15]. Resistance development can come from different mechanisms as follows: physiological, metabolic or target mutations. In relation to resistance to tau-fluvalinate, protein changes have been demonstrated through increases in detoxifying enzymes [16]. More recently, molecular

resistance, such as target mutation, is also believed to be involved in the development of resistance to tau-fluvalinate [17,18]. The detection of amitraz resistance was demonstrated in the laboratory and in the field [6,8,9,13]. Recently, target mutation resistance has been described in correlation with varroa mite resistance to amitraz [19]. Mutations were different between populations from France (N87S) and United States (Y215H). This discovery demonstrates the ability of the varroa mite to adapt according to its geographical area.

In this context, beekeepers must put in place control strategies to limit the development of resistance and maintain high treatment efficacy [5]. Among these strategies, that of alternating control (alternation of the acaricide substance over several years) has been identified. Unfortunately, the alternation of acaricides is generally not performed in France, and beekeepers mainly use Apivar treatment every year [20]. In addition, because of the loss of efficacy, beekeepers are advised by vets to leave the treatment in the beehives for a longer period [21]. Unfortunately, long-term treatments increase the risk of developing resistance [22].

More generally, for amitraz, several pharmaceutical treatments are available all around the world [4]. Each treatment has different characteristics that influence the varroa population and its resistance to the acaricide substance as follows: duration of treatment, active ingredient concentration and galenic formulation. The release of the active ingredient also varies according to the galenic presentation. This variation in release could explain the differences in efficacy observed for the varroa population between treatments. The choice of products in addition to the choice of the active ingredient may influence the efficacy and the development of resistance.

A recent modeling study shows the influence of resistance on the loss of efficacy with Apivar<sup>®</sup> (Vetopharma, Palaiseau, France) treatment in France [23]. This model explains how the level-resistant mites could be the explanation for the Apivar efficacy observed in the field. However, the model must be adapted to each medicine to fit with the observed field efficacy. A mechanistic model could allow us to understand how treatments kill mites and the impact of the efficacy loss on each type of treatment.

Our goal in this study is to assess the different amitraz-based products that are used for varroa mite management through a mechanistic model. Two points have been studied as follows: (1) the duration of treatment and (2) the rate of falling dead mites. Two products will be compared in the same environment as follows: Apivar<sup>®</sup> and Supatraz<sup>®</sup>. Modeling the action of the treatments may help explain the differences between these two medicines that have been observed for the efficacy parameters, such as those of the optimal treatment duration and residual mites.

## 2. Materials and Methods

### 2.1. Mathematical Modeling

The model of natural mite population dynamics is derived from the model developed by Calis et al. [24]. The action of Apivar<sup>®</sup> treatment was added according to the model developed by Almecija [23]. The input and output variables are presented in Table 1. R software (version 3.6.2) was used for simulations and statistical analysis.

**Table 1.** Input and output variables of the model.

Input Variables for Treatment	Description	Variation
dT	Treatment duration	Variable, from 4 to 12 weeks
pT	Treatment period	Variable, from 190 to 250 calendar days
kT	Daily mortality P(t) during treatment	Variable, from 0.03 to 1
dT	Treatment duration	Variable, from 4 to 12 weeks
Direct Output Variables		
M(t)	Daily number of dead individuals	
Mcum <sub>MODEL</sub> (t)	Cumulative number of dead individuals	

The natural population dynamics of varroa mites is defined by Equation (1) [23]. The number of phoretic mites is represented by  $P(t)$ . The variable  $I(t)$  corresponds to the number of mites infesting the worker brood, and  $E(t)$  corresponds to the number of mites emerging with young, mature females. The number of individual mites that died from natural mortality is given by  $M(t)$ . The treatment's effect on the honeybee population dynamics is represented by an additional daily mortality.  $MT(t)$  is the global number of deaths (dying from natural mortality and from mortality induced through the treatment). The treatment starts on 7 August for all simulations.

$$\begin{aligned}
 P(1) = P_i > 0 \quad P(t) = P(t-1) - I(t) + E(t) - M(t) \text{ for } t \geq 2 \\
 \text{with } M(t) = k \times P(t) \text{ with } k \text{ the natural mortality} \\
 \text{with } MT(t) = kT \times P(t) \text{ with } kT = \text{natural mortality} + \text{mortality during treatment}
 \end{aligned}
 \tag{1}$$

The action of the treatment depends on 3 parameters as follows: duration of treatment,  $dT$ , daily mortality,  $kT$ , and the level of initial infestations in the colony,  $P_i$ . The action of Apivar<sup>®</sup> treatment on the population can be simulated through a daily mortality rate,  $kT$ , and a constant for the duration of treatment,  $kT_{APIVAR}$  [23]. The Apivar<sup>®</sup> treatment duration is 10 weeks (70 days) [25]. The model had to be readjusted for the Supatraz<sup>®</sup> (amitraz) treatment, which has different pharmacokinetic characteristics from those of Apivar<sup>®</sup>. Supatraz<sup>®</sup> is an 8-week treatment (56 days).

## 2.2. Efficacy Determination

### 2.2.1. In the Field

The efficacy monitoring method is that of the standard European method [26]. Efficacy monitoring was carried out in 2019 and 2020 on Apivar<sup>®</sup> and Supatraz<sup>®</sup> in an apiary of 30 beehives located in Nieul-sur-mer, France. Before treatment, the mite infestation level was monitored to homogenize groups (level of phoretic mites by alcohol wash between 4 and 5% and  $3 \pm 0.5$  frames of brood per colony at the beginning of the experiment). In 2020, the efficacies of Apivar<sup>®</sup> and Supatraz<sup>®</sup> were followed, respectively, on 7 and 5 beehives. The colonies used were 6-frame nucs, so only one strip per colony was used. The strips were introduced into the colonies on 26 August and 8 July, respectively, for the years 2019 and 2020. The characteristics of each treatment are presented in Table 2. During treatment, the number of dead mites, which drop to the floor of the beehive, is counted with a sticky board every three days ( $V_{\text{treatment}}$ ).

**Table 2.** Characteristics of Apivar<sup>®</sup> and Supatraz<sup>®</sup>.

	Apivar <sup>®</sup>	Supatraz <sup>®</sup>
Active substance	Amitraz	Amitraz
Amitraz quantity per strip (mg)	500	700
Weight per strip (g)	15 g	12.52 g
Treatment duration	10 weeks (70 days)	6 to 8 weeks (42–56 days)

At the end of the treatment, a control treatment is applied (Apistan<sup>®</sup> (Tau-fluvalinate)) killing the remaining mites ( $V_{\text{remaining}}$ ). From these data, the percentage of field treatment efficacy,  $Eff_{\text{FIELD}}$ , is evaluated according to Equation (2) as follows:

$$Eff_{\text{FIELD}} = \frac{V_{\text{treatment}}}{V_{\text{treatment}} + V_{\text{remaining}}} \times 100
 \tag{2}$$

### 2.2.2. With the Model

Our model replicates the protocol that is applied in the field to determine treatment efficacy (including control treatment). The model estimates the number of cumulative dead mites,  $Mcum_{\text{MODEL}}(t)$ , during treatment and control treatment (Equation (3)). The treatment starts at day  $pT$ . The model simulates the efficacy of the treatment using the same

protocol as that in the field (Equation (4)). The number of remaining mites according to the model,  $V_{remaining_{MODEL}}$ , is evaluated through a simulation of a control treatment (with a daily efficiency of 90%).  $Eff_{field_{MODEL}}$  represents a simulation of the efficacy reproducing the protocol used in the field.

$$M_{cum_{MODEL}}(t) = \sum_{t=pT}^{pT+dT} M(t) \quad (3)$$

$$Eff_{field_{MODEL}} = \frac{M_{cum_{MODEL}} \times 100}{M_{cum_{MODEL}} + V_{remaining_{MODEL}}} \quad (4)$$

### 2.3. Validation of Supatraz<sup>®</sup> Modeling

The Apivar<sup>®</sup> model was validated using previous data from efficacy monitoring in 2020 [23]. The model showed that Apivar<sup>®</sup> presents the constant daily mortality rate,  $kT$ , during the treatment duration. Here, the model had to be adjusted to match the action of the Supatraz<sup>®</sup> treatment. Indeed, Supatraz<sup>®</sup> presents a daily mortality rate,  $kT_{SUPATRAZ}$ , of 0.342 for the first week of treatment and a variable value between 0.03 and 1 after one week of treatment.

The validation of the Supatraz modeling is based on the efficacy monitoring conducted in 2019 and 2020 in Nieul-sur-mer (France). Two parameters were set as follows: start of treatment ( $pT$  = August 7) and duration of treatment ( $dT$  = 8 weeks). The initial infestation,  $P_i$ , is adjusted according to the number of dead mites during the treatment ( $M_{cum_{MODEL}}$ ). The model is validated by comparing the slope of the cumulative number of dead mites over the total duration of treatment between the field and the model. The slopes of cumulative dead mites during treatment are defined in the field as well as through the model. They are evaluated over a duration of treatment of 70 days for Apivar<sup>®</sup> and one of 56 days for Supatraz<sup>®</sup>.

### 2.4. Time to Kill 90% of the Mite Population: $L_{90}$

A lethal time of 90% ( $LT_{90}$ ) represents the time required to kill 90% of the total varroa population. This period is evaluated from data from the field. It is evaluated through the ECOTOX package and the simulation of a GLM model tracing varroa mortality during treatment.  $LT_{90}$ s are evaluated for Apivar<sup>®</sup> and Supatraz<sup>®</sup> in 2019 and 2020.

### 2.5. Influence of the Proportion of Resistant Mites on the Treatment Efficacy

Resistance in the mite population is modeled through the decrease in the daily mortality rate,  $kT$ . The decrease in  $kT$  corresponds to the proportion of resistant mites that are insensitive to treatment. In our model, the proportion of resistant mites hypothetically remains stable and does not increase during the treatment. The percentage of efficacy is evaluated according to whether there are percentages of 0%, 20%, 40%, 60% and 80% of resistant mites in the population observed for both treatments.

Amitraz resistance has different impacts depending on the treatments of either Apivar<sup>®</sup> or Supatraz<sup>®</sup>. Apivar<sup>®</sup> kills mites with a similar  $kT$  during all the treatment. For Supatraz<sup>®</sup>, the resistance simulation is more complicated because of the inconstant  $kT$ . The reduction of  $kT$  to simulate resistance is not enough. The influence of resistance on the efficacy of Supatraz<sup>®</sup> differs depending on the two hypotheses as follows: (1) the resistant mites are killed the first week with stable  $kT$ , 0.394; (2) the resistant mites are not killed in the first week of treatment due to a high resistance ratio. In this second hypothesis, the proportion of  $kT$  decreases in the same proportion as that of resistant mites.

## 3. Results

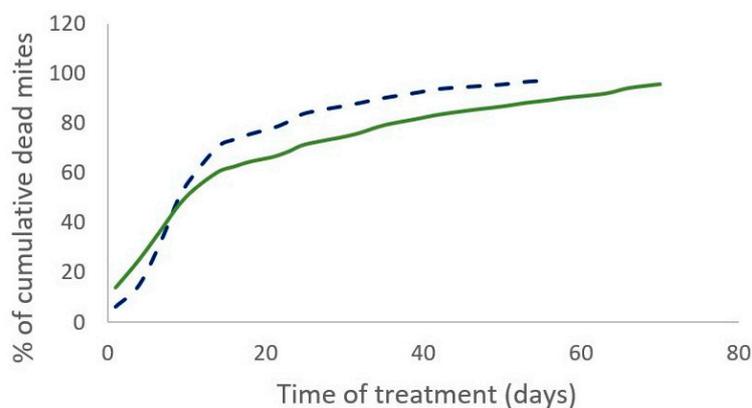
### 3.1. Field Efficacy

The percentage of the field efficacy is not significantly different between both treatments (Mann–Whitney test,  $W = 19$ ,  $p = 0.3$  (2019) and  $W = 12$ ,  $p = 0.43$  (2020)). In 2019, the

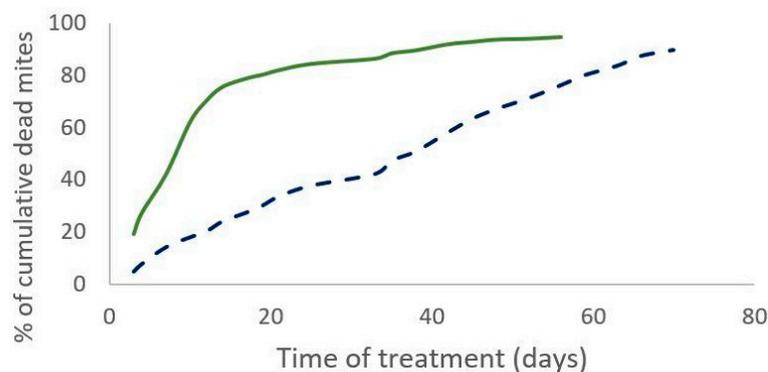
cumulative number of dead mites during treatment was not significantly different between the two treatments (Mann–Whitney test,  $W = 35$ ,  $n = 11$ ,  $p = 0.35$ ). The time required to kill 90% of the varroa population was significantly shorter for the Supatraz<sup>®</sup> treatment than for the Apivar<sup>®</sup> one in 2019 (Wilcoxon test,  $W = 79$ ,  $p = 7 \times 10^{-4}$ ) and in 2020 (Wilcoxon test,  $W = 35$ ,  $p = 2.5 \times 10^{-3}$ ) (Table 3). Supatraz<sup>®</sup> acts faster than Apivar<sup>®</sup> (Figures 1 and 2).

**Table 3.** Comparison of results obtained in the field in 2019 and 2020 between Apivar<sup>®</sup> and Supatraz<sup>®</sup>. Column followed by different letters indicate that they are significantly different ( $p < 0.05$ ).

	2019		2020	
	Apivar <sup>®</sup>	Supatraz <sup>®</sup>	Apivar <sup>®</sup>	Supatraz <sup>®</sup>
Number of beehives	8	12	7	5
Efficacy (%) $\pm$ SE	95.44 $\pm$ 1.19a	97.36 $\pm$ 0.61a	91.35 $\pm$ 2.83a	94.62 $\pm$ 1.45
Average of drop mites $\pm$ SE	2222 $\pm$ 438a	1636 $\pm$ 324a	4324 $\pm$ 1256a	1759 $\pm$ 605b
Average of remaining mites $\pm$ SE	91 $\pm$ 41a	40 $\pm$ 14b	203 $\pm$ 41a	57.4 $\pm$ 8.71b
LT90 (days) $\pm$ SE	50.97 $\pm$ 4.69a	28.49 $\pm$ 2.14b	65.27 $\pm$ 4.39a	37.68 $\pm$ 4.1b



**Figure 1.** Percentage of cumulative dead mites during treatment observed for Apivar<sup>®</sup> (solid line) and Supatraz<sup>®</sup> (dotted line) in 2019.



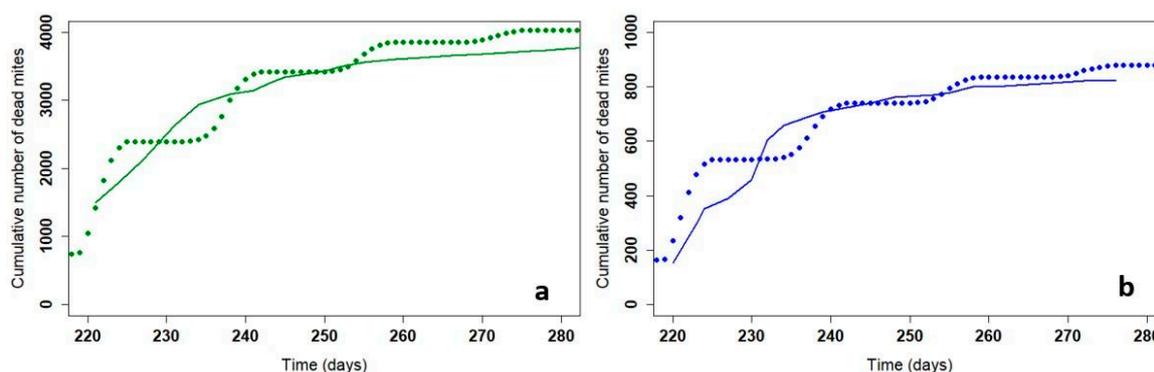
**Figure 2.** Percentage of cumulative dead mites during treatment observed for Apivar<sup>®</sup> (solid line) and Supatraz<sup>®</sup> (dotted line) in 2020.

Residual varroa counts are significantly lower for Supatraz<sup>®</sup>-treated colonies (Mann–Whitney test,  $W = 21$ ,  $n = 11$ ,  $p = 0.04$ ). The same observations are made for the year 2020 (Table 3). The percentage of efficacy alone is not enough to explain the real field efficacy of the treatment.

### 3.2. Modeling Efficacy

#### 3.2.1. Validation of Apivar<sup>®</sup> and Supatraz<sup>®</sup>

The model must represent the kinetics of the mite drop during treatment to match the reality. The action of Apivar<sup>®</sup> treatment was modeled in 2019 and 2020 on beehives followed by the ADA AURA (France) and APINOV in Nieul-sur-Mer (France). Maintaining the same parameter values, we represent the curve of the cumulative dead mites from the apiary in Nieul-sur-Mer (Figure 3).



**Figure 3.** Cumulative dead mites in the field (solid line) and with the model (dotted line) for Apivar (a) and for Supatraz (b). Field data come from one beehive in Nieul-sur-Mer in the 2019 trial.

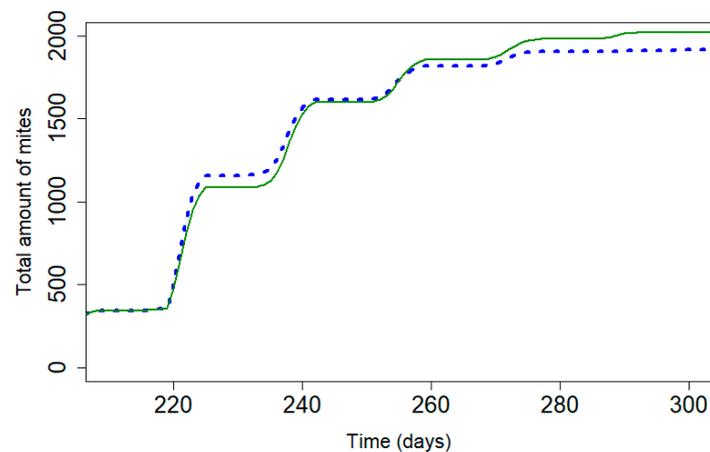
Figure 3 shows the number of mites that fell during treatment with Supatraz<sup>®</sup> in the field and with the model. The cumulative dead mites from the model correspond to the mite count in the field. Moreover, the 56-day slope is not significantly different between the field and model data (Wilcoxon test,  $p = 0.37$ ).

#### 3.2.2. Stabilization of the Mite Population with Supatraz<sup>®</sup> and Apivar<sup>®</sup>

Table 4 presents the different parameters used to stabilize the varroa mite population for both products. The initial daily mortality rate,  $kT$ , is higher for Supatraz<sup>®</sup> than for Apivar<sup>®</sup>. According to the model, a percentage of 97.0% for Supatraz<sup>®</sup> is enough to stabilize the varroa population from year to year, while Apivar<sup>®</sup> must have an efficacy rate of 98.77% [23]. The model shows that the number of dead mites during treatment would be lower for Supatraz<sup>®</sup> and the number of residual varroa mites would be higher. The model, which is free of all environmental factors, shows less of a difference in the cumulative dead mites between those observed for Apivar<sup>®</sup> and Supatraz<sup>®</sup> than the mortalities observed in the field (Figures 1, 2 and 4). In this condition, both treatments present identical results for the mite population as follows: efficacy, LT90 and number of dead mites.

**Table 4.** Modeling values needed to stabilize the varroa population for Apivar<sup>®</sup> (10 weeks of treatment) and Supatraz<sup>®</sup> (8 weeks of treatment) ( $P_i = 50$ ).

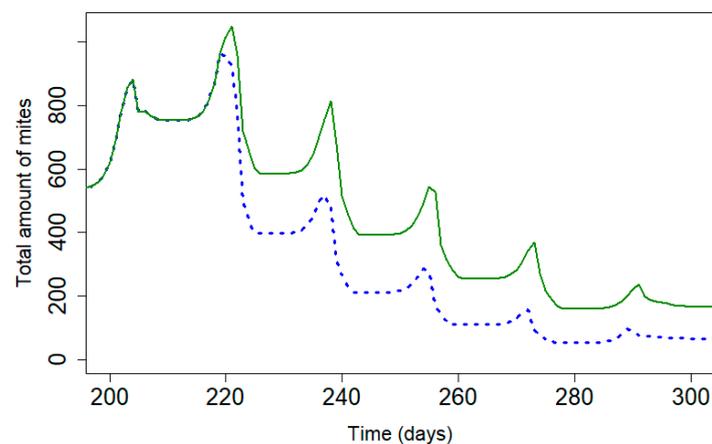
	Apivar <sup>®</sup>	Supatraz <sup>®</sup>
Treatment duration	10	8
Start of treatment	220	220
$P_i$	50	50
$kT_{APIVAR}$ and $kT_{SUPATRAZ}$	0.265	0.3
Percentage of efficacy (to stabilize the population)	98.77	97.17
Slope of the cumulative mite mortality curve (total treatment)	43	25.76
LT90	37	34
Number of cumulative dead mites	1534	1397
Varroa mites remaining	19	39
Maximum number of mites in the year	963	963



**Figure 4.** Cumulative mite mortality with the model for Apivar<sup>®</sup> (solid line) and Supatraz<sup>®</sup> (dotted line) ( $P_i = 50$ ) for stabilized the mite population year to year.

### 3.3. Influence of Decreasing Efficacy on Mite Population

Figure 5 presents the mite population curve during treatment applications of Apivar<sup>®</sup> and Supatraz<sup>®</sup> for a theoretical efficacy that is equal to 95% for both treatments. Supatraz<sup>®</sup> has a faster rate of action on the varroa population than Apivar<sup>®</sup> treatment, thereby allowing for a reduction in the reproduction rate of varroa mites early in treatment. The mite population decreases faster with Supatraz<sup>®</sup> than Apivar<sup>®</sup>, and the number of remaining mites is lower with Supatraz<sup>®</sup>.



**Figure 5.** Mite population during treatment with Apivar<sup>®</sup> (solid line) and Supatraz<sup>®</sup> (dotted line) for 95% efficacy across both products ( $P_i = 50$ ). ( $kT_{APIVAR} = 0.178$  and  $kT_{SUPATRAZ} = 0.24$ ).

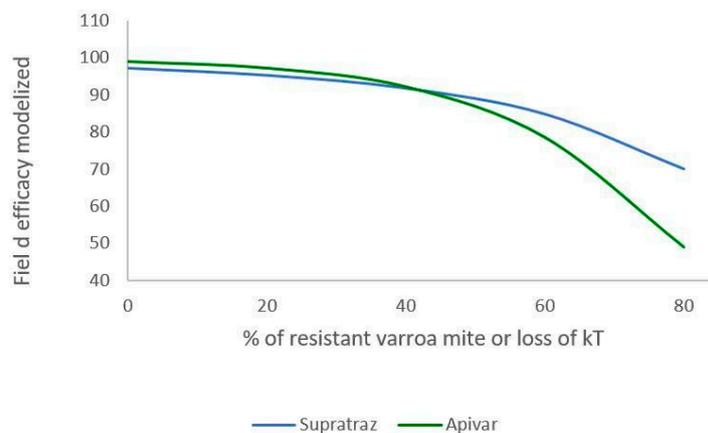
Table 5 presents the influence of both products on mite population depending on the percentage of efficacy. The LT90 is identical to the values observed in the field (Table 2). For the same efficacy, the model shows that Supatraz<sup>®</sup> has fewer remaining mites; moreover, the LT90s are shorter for Supatraz<sup>®</sup>.

**Table 5.** Number of cumulative dead mites and remaining mites with Apivar<sup>®</sup> and Supatraz<sup>®</sup> at different efficacies (with  $P_i = 50$ ).

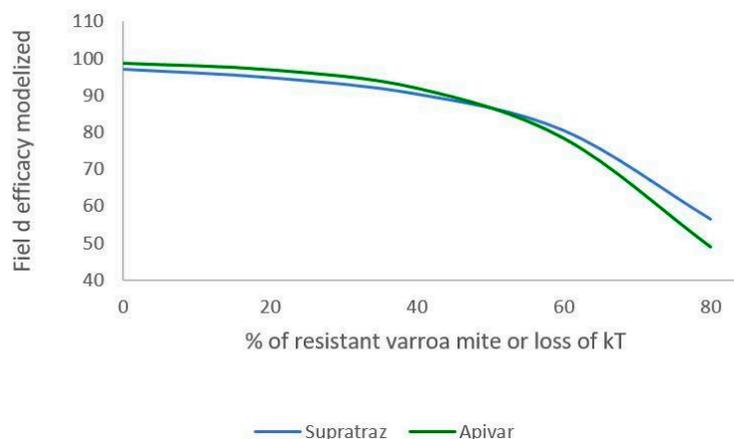
Efficacy	Apivar <sup>®</sup>			Supatraz <sup>®</sup>		
	Dead Mites	Remaining Mites	LT90 (Days)	Dead Mites	Remaining Mites	LT90 (Days)
99%	1505	15	37	1363	13	33
95%	1704	88	57	1529	80	35
90%	1776	196	70	1653	183	38

### 3.4. Influence of Resistant Mites on Treatment Efficacy

Figures 6 and 7 present the decrease in the efficacy when resistance increases in the mite population. With Apivar<sup>®</sup>, efficacy decreases from 98.77% to 48.9% (Figure 6). With Supratraz<sup>®</sup> and hypothesis 1, the efficacy decreases from 97.17 to 70.13% (Figure 7). With hypothesis 2, the efficacy decreases from 97.17% to 56.4%. In both cases, the efficacy is low when the resistance is high (>40%).



**Figure 6.** Modeling of efficacy loss depending on the percentage of resistance (Apivar:  $P_i = 50$ ,  $dT = 10$ ; Supratraz:  $P_i = 50$ ,  $dT = 8$ ). For Supratraz, hypothesis 1 was applied for this simulation. The proportion of  $kT$  decreases in the same proportion as that of resistant mites.



**Figure 7.** Modeling of efficacy loss depending on the percentage of resistance (Apivar:  $P_i = 50$ ,  $dT = 10$ ; Supratraz:  $P_i = 50$ ,  $dT = 8$ ). For Supratraz, hypothesis 2 was applied for this simulation. The varroa mites were sufficiently resistant to be safe during the first week of treatment.

When the proportion of resistant mites is under 40%, the efficacy decreases but is higher than 90% for both Apivar<sup>®</sup> and Supratraz<sup>®</sup>. Thus, the mite population is not stabilized but is sufficiently decreased for beekeepers. However, when the proportion of resistant mites is higher than 40%, the efficacy for all the cases decreases quickly, thereby leading to honeybee colony collapse.

## 4. Discussion

In the field, Supratraz<sup>®</sup> has a higher efficacy than Apivar<sup>®</sup>. The number of cumulative dead mites was lower with Supratraz<sup>®</sup> than Apivar<sup>®</sup>, thereby indicating faster action with Supratraz<sup>®</sup> [23]. Each medicine has different characteristics that affect the impact on the varroa population. The amount of the active ingredient as well as galenic changes the amount or speed of the active ingredient that is released. This could be consistent with the Supratraz<sup>®</sup> galenic due to the support matrix, which contains oil, and the higher amount of

the active ingredient (Figure 5). The product's galenic and formulation play an important role in the action of the treatment. The same observation was evaluated with different thymol-based treatments [27].

A mechanistic model was used to understand the action of both treatments on the varroa population. The action model depends on several parameters. The amitraz release in the beehive is not well understood and we cannot model it; however, the model can show us how each medicine influences the mite population reduction. Without dissociating the origin of this variation in the amitraz released (galenic and amount of the active compound), the modeling of the daily mortality rate of Apivar<sup>®</sup> and Supatraz<sup>®</sup> ( $kT_{APIVAR}$  and  $kT_{SUPATRAZ}$ ) allows for analysis of the performance of the medicines.

The population stabilization point between the treatments shows that the comparison of the efficacy between the treatments is subtle. To stabilize the mite population, the model shows that Supatraz<sup>®</sup> requires lower efficacy (Table 3). Moreover, in the case of a percentage of efficacy that was theoretically the same, Supatraz<sup>®</sup> presents fewer remaining mites and a shorter LT90 due to faster action (Table 4). Supatraz<sup>®</sup> could be useful to quickly reduce infestation, as shown in reference [28], which is probably due to both a higher amitraz concentration and formulation.

Our model also explained how the resistance level influences the efficacy of both treatments, especially for Supatraz<sup>®</sup>, with a higher daily mortality rate ( $kT$ ) in the first week. In both cases, the resistance of varroa mites has a high impact on the varroa population when more than 40% of the mites are considered to be resistant to amitraz. Even if the medicines present a different mode of action on the varroa mite population, the impact of the resistance is quite similar.

Depending on the treatment action, the risk of resistance development may vary, as was shown with other mites [29]. In fact, resistance development in mites and insects depends on many parameters, such as dose, pattern of application and timing and sequence of insecticide, are used [30,31]. Sensitive mites do not need high doses of the active ingredient; however, low doses can rapidly cause resistance. A treatment duration that is too long increases the risk of developing resistance. But, either a too high or too low concentration of the active ingredient can also increase the risk of developing resistance [22]. To prevent resistance, the treatment duration must be as short as possible and the amitraz concentration adapted. At present, no information is available on resistance trigger thresholds for the amitraz concentration and the treatment duration for varroa mites. The amitraz application could influence resistance development. However, more information is needed on the resistance in order to model the impact on the efficacy. It should be noted that our model has the same limitations that are described in the model developed for Apivar<sup>®</sup> [23] as follows: a population dynamic of bees with a total and long winter breakup and no reinfestation. Any changes in bee dynamics would alter these observations as they would affect the varroa population dynamics [28]. In addition, the virus quantity was not included in the model.

## 5. Conclusions

In the field, Supatraz<sup>®</sup> presentation is faster acting than that of Apivar<sup>®</sup> in reducing the mite population. By comparing the action of the Apivar<sup>®</sup> and Supatraz<sup>®</sup> treatments, we have shown that pharmacokinetic characteristics influence the global efficacy of the treatment. The model showed that comparing the efficacy of different products with different treatment times is not reliable. Moreover, low resistance probably affects both products in the same way, even if fast action can lead to an increase in efficacy and reduce the treatment duration. The trade-off between the treatment duration and the quantity of the active ingredient could allow the medications to decrease the risk of resistance development in the mite population.

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