



# Abstract **A Phenylalanine Ammonia Lyase Capacitive Sensor for Phenylalanine Detection**<sup>†</sup>

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**Abstract:** In this paper, an easy-to-use and fast biosensor for phenylalanine quantification in patients affected by phenylketonuria is investigated. The phenylalanine concentration was indirectly estimated through the ammonia released as a by-product of an enzymatic reaction, which was then detected by exploiting an yttria-stabilized zirconia layer deposited over an interdigitated capacitive sensor. The latter was manufactured by rapid-prototyping technologies. A sensor limit of detection higher than 1.25  $\mu$ M was estimated, along with an accuracy better than 18.31  $\mu$ M.

Keywords: phenylalanine; ammonia; biosensor; interdigitated capacitor; rapid prototyping

## 1. Introduction

Phenylketonuria (PKU) is a genetic disorder of phenylalanine (Phe) metabolism that may potentially lead to severe neurological damages. To date, a specific diet and monitoring the Phe levels in the blood is the unique therapy available [1]. Commonly, techniques for Phe quantification are complex, expensive and time-consuming [2]. This research work addresses the development of a low-cost and fast-responsive capacitive sensor for Phe quantification, which offers an accuracy compliant with early warning tasks. The sensing methodology exploits the indirect estimation of Phe by detecting aqueous ammonia  $(NH_3(aq))$  as a by-product of a specific enzymatic reaction with Phenylalanine Ammonia Lyase (PAL). Most  $NH_3$  sensors are aimed at gas detection [3], while few approaches address the quantification of  $NH_3(aq)$  [4], none of which provides solutions for measurements in small volumes. The modeling and design flow of the sensor are discussed in [5], while the ability of YSZ to detect  $NH_3(aq)$  is demonstrated in [6]. In this paper, the behavior of the complete sensor is investigated, including the Phe-PAL enzymatic reaction. The main outcomes of this approach include: (i) a capacitive readout strategy, which is convenient for low analyte concentrations; (ii) high specificity to the target analyte, thanks to the enzymatic reaction; (iii) faster response times compared to those of traditional Phe quantification methods; (iv) low costs, also provided by the adopted rapid-prototyping technologies.



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**Copyright:** © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). The structure and real view of the capacitive sensor are shown in Figure 1a,b. The IDC electrodes were realized by aerosol jet printing a gold-based conductive ink over a 125  $\mu$ m thick Polyether Ether Ketone (PEEK) substrate. The sensor active area was 13.5 × 9.8 mm<sup>2</sup>, and the IDC electrodes were 500  $\mu$ m wide, with a spacing of 300  $\mu$ m. A 50  $\mu$ m thick yttria-stabilized zirconia (YSZ) dielectric functional layer, sensitive to NH<sub>3</sub>(aq), was spray-coated over the IDC electrodes. The YSZ layer was subjected at room temperature, for 45 min, to an O<sub>3</sub>/UV<sub>254nm</sub> process performed by BioForce equipment (Nanosciences) to increase its surface hydrophilicity (126.6° ± 3°). The proposed enzymatic method exploits the deamination of Phe catalyzed by the PAL enzyme to produce trans-cinnamic acid and NH<sub>3</sub>. The yield of the latter is proportional to the Phe concentration [7]. PAL showed optimal sensitivity at temperatures ranging from 32 to 40 °C and pH values ranging from 7 to 8. A rigid supporting frame for the IDC sensor was milled in 1.55 mm thick FR4. The frame also provided the reaction chamber hosting the MUT.



Figure 1. The sensor: (a) top view (layout), (b) cross section and real view, (c) calibration diagram.

### 3. Experimental Results

The sensor behavior was investigated by observing its response to standard test solutions with Phe concentrations in the range of [0, 800]  $\mu$ M. The test solutions were prepared by mixing Phe with 10  $\mu$ L of PAL (containing 0.156 units in 200  $\mu$ L of 10 mM sodium phosphate buffer at pH 8.31) and 1 mL of 2 mM sodium phosphate buffer at pH 7.5. A volume of 500  $\mu$ L of these solutions was dropped on the YSZ sensing layer, and the temperature was set to 37 °C ( $\pm$ 2 °C) for 30 min to realize the enzymatic reaction. In each trial, the sensing chamber was completely filled. Time evolution of the sensor capacitance was acquired by a GW Instek LCR-6300 precision LCR meter. The obtained calibration diagram is shown in Figure 1c. A piecewise linear model was used to interpolate the data. The sensor responsivity was  $5.31 \times 10^{-1}$  pF/ $\mu$ M for Phe concentrations in the [0–200]  $\mu$ M range and  $2.86 \times 10^{-1}$  pF/ $\mu$ M for Phe concentrations in the [200–800]  $\mu$ M range. The accuracy in the 3\sigma limit, for the two considered intervals, resulted to be 18.31  $\mu$ M and 12.34  $\mu$ M, while the estimated limit of detection was 0.98  $\mu$ M and 1.25  $\mu$ M, respectively.

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