



Application of the Quality-by-Design (QbD) Approach to Improve the Nose-to-Brain Delivery of Diazepam-Loaded Nanostructured Lipid Carriers (NLCs) ⁺

Cláudia Pina Costa ^{1,*}, Sara Cunha ¹, Andreia F. Peixoto ², João Nuno Moreira ^{3,4}, José Manuel Sousa Lobo ¹ and Ana Catarina Silva ^{1,5}

- ¹ UCIBIO/REQUIMTE, MEDTECH, Laboratory of Pharmaceutical Technology, Department of Drug Sciences, Faculty of Pharmacy, University of Porto, 4099-002 Porto, Portugal; up201510339@ff.up.pt (S.C.); slobo@ff.up.pt (J.M.S.L.); ana.silva@ff.up.pt (A.C.S.)
- ² LAQV/REQUIMTE, Department of Chemistry and Biochemistry, Faculty of Sciences, University of Porto, 4169-007 Porto, Portugal; andreia.peixoto@fc.up.pt
- ³ CNC-Center for Neuroscience and Cell Biology, Center for Innovative Biomedicine and Biotechnology (CIBB), Faculty of Medicine (Pólo I), University of Coimbra, 3004-531 Coimbra, Portugal; jmoreira@ff.uc.pt
- ⁴ UC—University of Coimbra, CIBB, Faculty of Pharmacy, Pólo das Ciências da Saúde, Azinhaga de Santa Comba, 3000-548 Coimbra, Portugal
- ⁵ FP-ENAS (UFP Energy, Environment and Health Research Unit), CEBIMED (Biomedical Research Centre), Faculty of Health Sciences, University Fernando Pessoa, 4249-004, Porto, Portugal
- * Correspondence: claudiaspinacosta@gmail.com or up201610743@ff.up.pt
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Abstract: The intranasal administration of nanostructured lipid carriers (NLCs) has been suggested as a promising strategy to improve the fast treatment of epilepsy. This route allows for drug passage directly from the nose to the brain, avoiding the need of bypassing the blood–brain barrier. In addition, the quality-by-design (QbD) approach is a useful tool for the optimization of manufacturing variables, resulting in effective and safe pharmaceutical formulations. The aim of this work was to use the QbD approach to optimize a NLCs formulation for the nose-to-brain delivery of diazepam. The studies began with the screening of excipients and the assessment of the lipid-drug compatibility. The central composite design was used to evaluate the effects of critical material attributes (CMAs) (ratio of solid and liquid lipids and the amount of drug and emulsifiers) on the CQAs of the diazepam-loaded NLCs formulation (particle size, polydispersity index (PDI), zeta potential (ZP) and encapsulation efficiency (EE)). The results showed that the most adequate ratios of lipids and emulsifiers were 6.65:2.85 and 4.2:0.3 (%, *w/w*), with values of 84.92 nm, 0.18, -18.20 mV and 95.48% for particle size, PDI, ZP and EE, respectively. This formulation was selected for further studies related to the optimization of critical process parameters (CPPs).

Keywords: epilepsy; nose-to-brain delivery; intranasal delivery; nanostructured lipid carriers; quality-by-design

1. Introduction

Neurological disorders, including epilepsy, require a rapid and effective treatment targeting the brain. In this area, the intranasal administration of lipid nanosystems, such as nanostructured lipid carriers (NLCs) has been suggested as a promising strategy. This route allows for drug passage directly from the nose to the brain, avoiding the need of bypassing the blood–brain barrier [1]. The quality-by-design (QbD) approach has been applied to optimize NLCs formulations, improving the manufacturing processes and ensuring the quality and safety of the final products. Herein, the quality target product

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Copyright: © 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses /by/4.0/). profile (QTPP) and critical quality attributes (CQAs) are identified, and a risk assessment analysis is conducted to evaluate the critical material attributes (CMAs) and critical process parameters (CPPs) [2]. The aim of this work was to use the QbD approach to optimize a NLCs formulation for the nose-to-brain delivery of diazepam, improving epilepsy emergency treatment. Studies started with the screening of excipients and evaluation of the lipid-drug compatibility. Subsequently, the QbD approach was applied to evaluate the effects of CMAs on CQAs in the diazepam-loaded NLCs formulation.

2. Experiments

2.1. Materials

Diazepam was purchased from Acofarma (Barcelona, Spain). Precirol® 5 ATO (glyceryl palmitostearate), Compritol® 888 ATO (glyceryl behenate), Gelucire® 43/01 (hard fat compounds), Gelucire[®] 44/14 (lauroyl polyoxyl-32 glycerides), Gelucire[®] 50/13 (stearoyl polyoxyl-32 glycerides), cetyl palmitate, Apifil® (PEG-8 beeswax), Labrafac® W1349 (medium chain triglycerides) and Capryol® 90 (propylene glycol monocaprylate) were kindly provided by Gattefossé (Lyon, France). Imwitor® 900K (glyceryl stearate), Softisan® 100 (hydrogenated coco-glycerides), Softisan® 154 (hydrogenated palm oil), Dynasan® 118 (glyceryl tristearate) and Witepsol® E85 (hard fat compounds) were from Oxi-med (Barcelona, Spain). Cetiol[®] V (decyl oleate) and glyceryl monostearate were from Guinama (Valencia, Spain). Miglyol® 812 (medium-chain triglycerides of caprylic and capric acids), Tween 80[®] (polysorbate 80), sodium deoxycholate, oleic acid, isopropyl myristate, vitamin E, chloride benzalkonium, stearic acid, sodium chloride and sodium phosphate were purchased from Acofarma (Barcelona, Spain). Lutrol® F68 (poloxamer 188) and Lutrol® F127 (poloxamer 407) were acquired from BASF (Gordon, USA), Phospolipon® 90 G and Phospolipon[®] 90 H were obtained from Lipoid (Ludwigshafen am Rhein, Germany) and acetonitrile was from Thermo Fisher Scientific (Loughborough, UK). The purified water used for the NLCs production was obtained from a Direct-Q[®] Ultrapure Water Systems, Merck Millipore (Darmstadt, Germany).

2.2. Methods

2.2.1. Screening of Excipients

The excipients chosen to develop the NLCs were based on previous research, where the suitability of lipids and emulsifiers for nasal administration was confirmed [3,4]. Tested solid lipids were Precirol® 5 ATO, Imwitor® 900K, Compritol® 888 ATO, Gelucire® 43/01, Gelucire® 44/14, Gelucire® 50/13, glyceryl monostearate, stearic acid, cetyl palmitate, Softisan® 100, Softisan® 154, Dynasan® 118, Apifil® and Witepsol® E85. Tested liquid lipids were Miglyol® 812, oleic acid, isopropyl myristate, Cetiol® V, vitamin E, Labrafac® W1349, Capryol® 90 and Microcare®. The tested emulsifiers were non-ionic emulsifiers, such as Tween 80®, Lutrol® F68, Lutrol® F127, and anionic emulsifiers, such as sodium deoxycholate and phospholipids (Phospolipon® 90 G and Phospolipon® 90 H).

2.2.2. Compatibility between Solid and Liquid Lipids

Compatibility between lipids was evaluated by screening different ratios of solid and liquid lipids, i.e., 60:40, 70:30, 80:20 and 90:10, heated 5–10 °C above the melting point of the solid lipid under stirring for 1 h, and cooled at room temperature (25 ± 0.5 °C). The mixture was examined for any phase separation and color change. Afterwards, the mixture was placed in a hydrophilic filter paper, followed by visual observation to determine the presence/absence of liquid oil droplets on the filter to detect the existence/absence of immiscibility.

2.2.3. Drug-Lipid Solubility

To evaluate which solid lipid solubilizes the highest diazepam amount, an excess of the drug (5–10%, w/w) was added to the lipid and heated to 5–10 °C above the melting point under continuous stirring for 1 h. After solidification by cooling to room temperature, the presence/absence of insoluble drug crystals was observed. The same procedure was conducted with the liquid lipid.

2.2.4. Preparation of the Diazepam-Loaded NLCs Formulation

Diazepam-loaded NLCs were prepared from the method previously employed by Silva et al. [5]. Briefly, the aqueous and lipid phases were heated to 5–10 °C above the melting point of the solid lipid. Afterwards, the aqueous phase was added to the lipid phase and the mixture was emulsified under high-speed stirring using an Ultra-Turrax[®] T25 (Janke and Kunkel GmbH (Staufen, Germany) at 13,400 rpm for 5 min. The formed emulsion was sonicated, with an amplitude of 75% for 15 min, using a VCX 130 ultrasonic processor (Sonics, Switzerland). The obtained O/W nano-emulsion was transferred to glass vials and drastically cooled down to room temperature (20 ± 0.5 °C) to generate NLCs.

2.2.5. Characterization of the Diazepam-Loaded NLCs Formulation

Particle size was evaluated by laser diffractometry (Mastersizer 3000, Malvern, UK) and dynamic light scattering (DLS), using a Malvern Nano-Zetasizer (Malvern, UK). The polydispersity index (PDI) and zeta potential (ZP) were evaluated using the same Malvern Nano-Zetasizer.

The encapsulation efficiency (EE) of diazepam in the NLCs was calculated according to the following equation: EE (%) = [(total amount of drug – free drug)/total amount of drug] \times 100.

Diazepam was quantified by high-pressure liquid homogenization (HPLC), at 254 nm.

2.2.6. QbD Approach

The QbD approach was applied to optimize the diazepam-loaded NLCs formulation, improving the manufacturing process and ensuring the quality and safety of the final product. The effect of the CMAs and CPPs on the CQAs is shown in Figure 1.



Figure 1. Ishikawa diagram showing the effects of critical material attributes (CMAs) and critical process parameters (CPPs) on the critical quality attributes (CQAs), for the optimization of a nanostructured lipid carrier (NLCs) formulation.

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3. Results

3.1. Preparation and Characterization of the Diazepam-Loaded NLCs Formulation

Precirol[®] 5 ATO and Cetiol[®] V were selected as the solid lipid (SL) and the liquid lipid (LL), respectively. Tween 80[®] and sodium deoxycholate were selected as emulsifiers and a drug concentration of 0.50% was selected to prepare the diazepam-loaded NLCs formulation (Table 1).

Table 1. Composition of diazepam-loaded nanostructured lipid carriers (NLCs) formulation.

Composition	(<i>w</i> / <i>w</i>)%
Precirol [®] 5 ATO	6.65
Cetiol® V	2.85
Diazepam	0.50
Tween 80®	4.20
Sodium deoxycholate	0.30
Benzalkonium chloride	0.02
Ultrapure water	q.s. 100.00

3.2. *QbD* Approach

Central Composite Design (CCD)

CCD was used to evaluate the effects of CMAs on the CQAs of the NLCs formulation (Tables 2 and 3 and Figure 2).

Table 2. Selection of the central composite design (CCD) variables and respective levels.

CMAs	_7	_1	0	1	2
CIVIAS	-2	-1	0	1	2
Ratio SL:LL	5:5	6:4	7:3	8:2	9:1
Ratio E1:E2	2.4:0.1	2.9:0.1	3.3:0.2	3.7:0.3	4.2:0.3

CMAs: critical material attributes; SL: Precirol[®] 5 ATO; LL: Cetiol[®] V; E1: Tween 80[®]; E2: sodium deoxycholate.

Table 3. Effect of the critical material attributes (CMAs) on the critical quality attributes (CQAs) of the diazepam-loaded NLCs.

CMAs	D(50) nm	D(90) nm	Z-Ave (nm)	PDI	ZP (mV)	EE (%)
A1	57.100 ± 0.001	167.000 ± 0.003	129.300 ± 46.960	0.179 ± 0.000	-16.100 ± 7.240	93.960 ± 0.001
A2	60.600 ± 0.000	146.000 ± 0.004	107.000 ± 44.690	0.188 ± 0.000	-20.200 ± 7.710	94.770 ± 0.001
A3	53.700 ± 0.002	141.000 ± 0.002	140.300 ± 48.400	0.205 ± 0.000	-18.000 ± 6.440	92.720 ± 0.003
A4	55.300 ± 0.000	145.000 ± 0.003	93.980 ± 47.430	0.180 ± 0.000	-19.200 ± 9.440	94.470 ± 0.001
A5	77.200 ± 0.000	126.000 ± 0.007	109.600 ± 40.260	0.157 ± 0.000	-20.600 ± 8.500	94.600 ± 0.002
A6	68.700 ± 0.001	192.000 ± 0.005	113.900 ± 47.660	0.185 ± 0.000	-16.200 ± 8.660	94.430 ± 0.000
A7	55.700 ± 0.003	155.000 ± 0.009	158.400 ± 45.660	0.164 ± 0.000	-14.100 ± 9.500	92.000 ± 0.003
A8	53.300 ± 0.000	137.000 ± 0.003	84.920 ± 45.750	0.178 ± 0.000	-18.200 ± 7.220	95.480 ± 0.001
A9	75.500 ± 0.005	134.000 ± 0.007	110.900 ± 40.080	0.153 ± 0.000	-18.400 ± 8.340	94.750 ± 0.002
A10	75.200 ± 0.001	133.000 ± 0.008	110.500 ± 42.760	0.151 ± 0.000	-18.100 ± 8.890	94.790 ± 0.002

CMAs: critical material attributes; D(50): 50% of particles with size equal to or lower than the given value; D(90): 90% of particles with size equal to or lower than the given value; EE: encapsulation efficiency; PDI: polydispersity index; ZP: zeta potential; Z-Ave: mean particle size.



Figure 2. The 3D surface plots portraying the effect of the ratio between the solid and liquid lipids (SL:LL) and the two emulsifiers (E1:E2) on the size (D(50): 50% of particles with size equal to or lower than the given value, D(90): 90% of particles with size equal to or lower than the given value and Z-Ave: mean particle size), polydispersity index (PDI), zeta potential (ZP) and encapsulation efficiency (EE).

4. Discussion

The studies for screening excipients, compatibility between lipids and the drug-lipid solubility allowed for the preparation of a diazepam-loaded NLCs formulation with the typical characteristics of these systems, such as low viscosity and milky appearance. In addition, the use of the QbD approach was effective for the optimization of the diazepam-loaded NLCs formulation. Other studies have also reported the effectiveness of QbD optimizing NLCs formulations. For example, Cunha et al. used the QbD and two different experiment designs (CCD and Box-Behnken design) to optimize a rivastigmine-loaded NLCs formulation prepared through two different production methods (sonication and high-pressure homogenization). In this study, the variations in the CMAs and CPPs originated important outcomes in the CQAs of the final formulation [6]. From Table 3 and Figure 2, it can be observed that the most adequate ratios of lipids and emulsifiers were 6.65:2.85 and 4.2:0.3 (%, w/w). The results of particle size, PDI, ZP and EE were, respectively, 84.92 nm, 0.178, -18.20 mV and 95.48%. These values are in accordance with the requisites of intranasal delivery of NLCs formulations, which are a particle size of less than 200 nm, a PDI less than 0.3, a ZP close to -20 mV and an EE higher than 80% [6–8].

5. Conclusions

Optimizing NLCs formulations is critical to achieve a reproducible quality of the final pharmaceutical products, in terms of both efficacy and safety. The QbD approach is a useful tool for the development of these systems, being observed that manufacturing variables related to the materials and process parameters are important for the optimization of the diazepam-loaded NLCs formulation.

The diazepam-loaded NLCs formulation with the best CQAs was selected for further optimization, related to the selection of the best CPPs and using the same design of experiment, which will be tested in vitro and in vivo in the future.

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Abbreviations

CCD	central composite design
CMAs	critical material attributes
CPPs	critical process parameters
CQAs	critical quality attributes
DLS	dynamic light scattering
E1	Tween [®] 80
E2	sodium deoxycholate
EE	encapsulation efficiency
HPLC	high-pressure liquid homogenization
LL	liquid lipid
NLCs	nanostructured lipid carriers
PDI	polydispersity index
QbD	quality-by-design

QTPP	quality target profile product
SL	solid lipid
ZP	zeta potential

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