



Use of Algae in Aquaculture: A Review

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Abstract: The utilization of algae in aquaculture is environmentally friendly, safe, and cost-effective and can effectively substitute for fish meal and fish oil in aquatic feeds. Incorporating algae as dietary supplements leads to significant enhancements in aquatic animals' health and also improves the aquatic ecosystem. Algae are rich sources of nutrients and serve as the foundational food source in the aquatic food chain. Currently, 40 different algae species are employed in aquaculture. Furthermore, algae contributes to elevating the overall quality of aquatic feed products. Aquaculture stands as the most vital food production sector globally; however, challenges such as infection outbreaks and aquatic environmental pollution pose significant threats to the sustainable growth of this industry. An alternative strategy for mitigating environmental issues and improving aquatic production involves the utilization of algae. The novelty in the applications of algae in aquaculture stems from their multifaceted roles and benefits, such as their capacity to improve water quality, serve as nutrientrich feed supplements, and enhance the overall health and productivity of aquatic species. These versatile applications of algae represent a fresh and innovative approach to sustainable aquaculture practices. This review furnishes insights into the use of algae, algae extracts, or components derived from algae to enhance water quality. Additionally, it covers the utilization of algae-based feed supplements, boosting of the immune system, enhanced growth performance, and disease resistance in aquatic animals.

Keywords: algae; water treatment; dietary additives; aquatic animal; aquaculture

Key Contribution: Algae are naturally rich nutrient sources; they are the primary food producer in the food chain for aquatic animal life, and the algae cultivation method is eco-friendly, nontoxic, and cost-effective. They have numerous beneficial properties, including immunostimulant, antioxidant, anti-inflammatory, and antimicrobial activity in aquatic animals, and microalgae convert atmospheric carbon into high-nutrient products. Algae improve the circular bioeconomy in the aquaculture industry, and the algae treatment process is a successful method for different types of wastewater from municipal, industrial, agro-industrial, and livestock sources. Both microalgae and macroalgae as well as algae-producing components like carbohydrates, lipids, and proteins are beneficial substances in aquatic feeds and improve the quality of aqua feed, aquatic animal health, and the aquatic environment.



Citation: Vijayaram, S.; Ringø, E.; Ghafarifarsani, H.; Hoseinifar, S.H.; Ahani, S.; Chou, C.-C. Use of Algae in Aquaculture: A Review. *Fishes* **2024**, *9*, 63. https://doi.org/10.3390/ fishes9020063

Academic Editor: Francisco Javier Alarcón López

Received: 8 December 2023 Revised: 26 January 2024 Accepted: 27 January 2024 Published: 1 February 2024



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1. Introduction

The aquaculture sector plays a crucial role in global food production and holds significant economic importance for many countries. Both finfish farming and crustacean production contribute substantially to the global food supply chain, providing essential protein sources and supporting livelihoods. As the global population continues to grow, the aquaculture sector is expected to play an increasingly important role in meeting the demand for nutritious and sustainable protein sources. Sustainable and responsible aquaculture practices are critical for the long-term health of the industry and the environment [1]. Aquatic feeds typically play a crucial role in augmenting aquatic production, with a substantial portion of fish oil (FO) and fish meal (FM) being used as essential feed ingredients to meet the dietary requirements of farmed aquatic species. Consequently, there is a growing imperative to re-evaluate the sustainability of fish farming regarding the consumption of FM and FO [2]. The utilization of algal technology in aquaculture offers a range of benefits and represents an alternative approach to enhancing various biological functions. Algae, including microalgae and macroalgae, can serve multiple purposes in aquaculture systems, contributing to improved health and environmental conditions. The application of algal technology in aquaculture aligns with the goals of sustainability, environmental stewardship, and improved efficiency within the industry. Research and ongoing developments continue to explore innovative ways to integrate algal technologies for the benefit of both aquaculture production and the surrounding ecosystems [3]. Algae, including brown algae (Phaeophyceae), green algae (Chlorophyceae), red algae (Rhodophyceae), and diatoms (Bacillariophyceae), can be broadly categorized into two primary types: microalgae and macroalgae. The distinction between microalgae and macroalgae is primarily based on size, with microalgae being microscopic and macroalgae being visible without the aid of a microscope. Both microalgae and macroalgae play important roles in aquatic ecosystems and have various applications in industries, ranging from food and agriculture to biotechnology and environmental management [4]. The distinctive or combined effects of various algae species contribute to the consistency of protein, lipids, and essential trace minerals in aquaculture. Some of the commonly used algae species include *Tetraselmis* sp., *Pavlova* sp., Chlorella sp., Isochrysis sp., Chaetoceros sp., Phaeodactylum sp., Skeletonema sp., Thalas*siosira* sp., and *Nannochloropsis* sp. [5]. Microalgae, as a diverse group of photosynthetic microorganisms, have played crucial roles (photosynthetic evolution, primary producers, formation of planktonic ecosystems, biodiversity, carbon fixation, nitrogen fixation, symbiotic relationships, biogeochemical cycling, and adaptation to diverse environments) in the evolutionary history of life on Earth. While their exact evolutionary timeline is complex and not fully understood, microalgae have significantly contributed to the development and maintenance of ecosystems. Understanding the evolutionary roles of microalgae is crucial not only for elucidating the history of life on Earth but also for recognizing their ongoing ecological importance in contemporary ecosystems and their potential applications in various industries, such as bioenergy, food, and environmental management. Microalgae are indeed recognized as an essential and indispensable source of nutrition for fish fry and larval shrimp in aquaculture. In the context of aquaculture, particularly in hatcheries where controlled conditions are maintained for the early stages of fish and shrimp development, microalgae are considered a key component in ensuring successful and healthy aquaculture production. [6]. Sivaramakrishnan et al. [7] reported that four green algae, Acetabularia acetabulum, Enteromorpha, Halimeda macroloba, and Halimeda tuna, exhibit significant potential in terms of antioxidant capabilities, immune stimulation, and medicinal applications for aquatic animals. Algae represent vital natural food sources for aquatic animals, and various algae species, such as Leptolyngbya valderiana, L. tenuis, Arthrospira maxima, Navicula minima, Nostoc ellipsosporum, Cytoseira, Ulva, Pavlova, Chaetoceros, Porphyridium, Chlorella, Palmaria, Gracilaria, and Isochrysis, are promising feed components, enhancing the growth performance and immunity of Nile tilapia (Oreochromis niloticus), rainbow trout (Oncorhynchus mykiss), European seabass (Dicentrarchus labrax), golden gourami (Trichopodus trichopterus), cichlid (*Cichlidae*) fish, wag swordtail (*Xiphophorus hellerii*), orange molly pink zebra, tetras

(*Paracheirodon axelrodi*), and prawns (*Dendrobranchiata*). Therefore, algae biomass serves as a cost-effective, safe, and environmentally friendly feed ingredient [8], and administrations of microalgae directly to the rearing water or indirectly by mixing them into the feed has become a crucial practice for enhancing aquatic animal nutrition [9–11]. Both micro- and macroalgae are advantageous components in aqua feeds, as they contribute to improving feed quality, aquatic animal health, and the aquatic environment [12].

Microalgal components offer not only beneficial effects for aquatic animals but also significant economic and ecological advantages in aquaculture [13]. Microalgal biomass contains a variety of biochemical components, such as proteins, lipids, carbohydrates, natural antioxidants, and bioactive products, and these diverse components are harnessed to develop sustainable aquaculture [14]. Microalgae supplements have been shown to enhance the growth performance of aquatic animals, and they synthesize a wide array of beneficial biological elements, including amino acids, vitamins, antioxidants, and pigments, which are advantageous for the health and growth of aquatic animals [15,16]. Compounds generated by macroalgae, particularly polysaccharides, can up-regulate the immune system and enhance protection against Aeromonas hydrophila, Enterobacter sp., Pseudomonas aeruginosa, Streptococcus sp., Escherichia coli, Vibrio parahaemolyticus, Vibrio alginolyticus, Vibrio cholerae, Yersinia enterocolitica, and Proteus sp. [17]. Astaxanthin synthesized by algae enhances antioxidant activity in rainbow trout and offers beneficial effects for human health [18–20]. Furthermore, the inclusion of algae and their various biomolecules, proteins, peptides, amino acids, fatty acids, sterols, polysaccharides, oligosaccharides, phenolic compounds, photosynthetic pigments, vitamins, and minerals collectively contribute significantly to the growth performance, antioxidant activity, and antimicrobial activity, serving as immune-stimulating agents in both aquatic animals and humans [21,22].

Algae are reported to exhibit antibacterial properties against a wide range of bacteria, encompassing both Gram-positive and Gram-negative strains [23,24]. Various algal species synthesize antimicrobial compounds such as short-chain and long-chain fatty acids, which are effective against pathogenic bacteria, including *Vibrio* species, *Aeromonas* species, and *Pseudomonas* species, as well as viruses, including *influenza* B, *mumps* viruses, herpes simplex virus type 1 (HSV-1), and even the human immunodeficiency virus type 1 (HIV-1) [25]. Algae-synthesized polyunsaturated fatty acids (PUFAs), such as eicosapentaenoic acid (EPA) and sterols, have microbicidal effects against several bacteria, including Grampositive strains like *Staphylococcus aureus* and *Streptococcus pyogenes*, as well as Gramnegative genera such as *Aeromonas*, *Pseudomonas*, and *Vibrio* [26–30].

Microalgae cultivation is used in aquaculture wastewater treatment [31,32], as it can improve water quality [33]. Five marine microalgae strains, Porphyridium aerugineum, Nannochloropsis granulate, Tetraselmis chuii, Botryococcus braunii, and Phaeodactylum tricornutum, can produce varying levels of essential minerals, including calcium (ranging from 0.26%) to 2.99%), phosphorus (0.73% to 1.46%), magnesium (0.26% to 0.71%), potassium (0.67% to 2.39%), sodium (0.81% to 2.66%), and sulfur (0.41% to 1.38%). These microalgae strains produce a total of 26 different chemical compounds, and their biomass plays an important role in carbon dioxide (CO_2) conversion, nutrient recycling, land cultivation, and the remediation of wastewater, demonstrating their significance in environmental recycling [34–37]. The use of algae grown on wastewater for aquafeed applications involves several regulatory aspects (wastewater quality standards, nutrient content and composition, microbial safety, toxic substances, harmful algal blooms, feed safety regulations, environmental impact assessment, labeling and documentation, approval and certification, traceability and record keeping, and public health and consumer safety), which need to be considered to ensure both the safety of the aquafeed and the protection of the environment. Regulatory frameworks can vary by country, and specific guidelines may be provided by relevant authorities. Below are some general regulatory considerations. It is essential for individuals or companies engaged in the production of aquafeed using algae grown on wastewater to familiarize themselves with the specific regulations applicable in their region and to work closely with relevant regulatory authorities to ensure compliance. Additionally, seeking

certification from reputable organizations may enhance the credibility and market acceptance of such products [37,38]. The present review aims to provide an overview of recent findings and applications related to the use of both micro- and macroalgae in aquaculture, including the use of macro- and microalgal extracts as functional feed additives as reported by Monteiro et al. [39] and the introduction of innovative approaches that can contribute to the development of sustainable aquaculture.

2. Cultivation Process

The cultivation of algae is the process of growing algae for various purposes, including food production, biofuel, bioremediation, and pharmaceuticals. Algae can be cultivated in open ponds, closed bioreactors, or photobioreactors [40]. The cultivation process involves the following elements: strain selection, growth medium, cultivation system, light and temperature, aeration and mixing, harvesting, and processing. Algae cultivation has gained attention as a sustainable and environmentally friendly way to produce a wide range of products with industrial applications, including biofuel production, food and feed production, wastewater treatment, and pharmaceuticals. Additionally, algae cultivation can contribute to carbon capture and reduce greenhouse gas emissions when used for biofuel production, and as a sustainable and versatile practice, algae cultivation has garnered interest in research and industry for its potential to contribute to a range of sectors. Advances in cultivation technologies and the exploration of novel algal strains continue to shape the field, making algae cultivation a promising solution for addressing global challenges related to food security, environmental sustainability, and renewable energy production [41–44]. In addition, it is worth mentioning that Silkina et al. [45] displayed that far-red light adaptation improved phycocyanin, a pigmented protein complex, and xanthophyll concentrations compared to white-light conditions in the thermophilic cyanobacteria Chlorogloeopsis fritschii.

3. Wastewater Management in Aquaculture

Aquaculture produces substantial volumes of wastewater, which contains nutrients, organic substances, and living organisms. The wastewater contents can vary, influenced by factors such as the species cultivated, the type of feed utilized, and the methods employed for water management [42,43,45,46]. The use of algal cultivation for the enhancement of water quality, especially in reducing Chemical Oxygen Demand (COD), involves considerations of scale application efficiency (system design, nutrient loading rates, algal species selection, harvesting, and recirculation system), commercial exploitation (nutrient recovery, aquaculture and agriculture, research and development, public and private partnerships), and regulatory aspects (water quality standards, environmental permits, biosafety and health regulations, monitoring and reporting, land use regulations, and product safety). By addressing the scale application efficiency, commercial exploitation, and regulatory aspects, the integration of algal cultivation for water quality improvement can be more effectively implemented and sustained over time. Collaboration with regulatory bodies and adherence to standards are crucial for the successful and responsible deployment of algae-based water treatment solutions [32]. Microalgae cultivation has the potential to improve aquaculture wastewater, as it can affect the excess amount of nutrients such as ammonia, nitrate, phosphate, and chemical oxygen demand (COD) in order to enhance water quality. This removal not only benefits water quality but also mitigates the environmental impact of nutrient-rich effluents when discharged. The harvested microalgae biomass can serve as a valuable feed source for aquaculture species, or it can be utilized for various other purposes, making it a cost-effective and environmentally friendly solution for both wastewater treatment and resource utilization in the aquaculture industry [32,47]. Effective management of aquaculture wastewater is imperative to mitigate the detrimental effects on aquatic ecosystems. Employing efficient waste capture, water treatment, and recycling systems can significantly decrease the discharge of pollutants. In addition, regulatory measures and adherence to regulation are crucial for ensuring the long-term environmental

sustainability of aquaculture operations [38,48,49]. Algal biomasses serve a dual purpose in enhancing the nutritional value of aquatic feed supplements, contributing to both environmental sustainability and the efficient utilization of aquaculture resources [50]. The use of algal technology for removing excess nutrients and dissolved carbon from wastewater is not a recent development; it was first suggested in the 1950s and has since evolved and gained prominence as an environmentally friendly and effective method for wastewater treatment and resource re-utilization [51-54]. Microalgae removes biological matter from wastewater through processes like consumption, adsorption, and biodegradation through microalgae's production of extracellular polymers, which can detoxify or modify pollutants [55–58]. The microalgae *Chlorella vulgaris* has shown impressive efficiency in removing total nitrogen (86.1%) and phosphorus (82.7%) from white-leg shrimp (Penaeus vannamei) farming wastewater [59]. Furthermore, C. vulgaris demonstrated superior growth in aquaculture wastewater when bicarbonate was used as the carbon source, underscoring its potential for efficient nutrient removal [60]. The microalgae Scenedesmus sp. is effective in removing pollutants from wet market wastewater, removing 74.8% of total nitrogen (TN) and 92.2% of total phosphorus (TP) [61]. The microalgae Spirulina sp. is also utilized for treating aquaculture wastewater and reducing environmental impact [48].

Microalgae biomass and its components, including astaxanthin, phycocyanin, carbohydrates, proteins, lipids, and PUFAs, are effectively employed to remove eutrophication pollutants from wastewater. The biomass of microalgae species like Chlamydomonas reinhardtii, Monoraphidium griffithii, and Selenastrum sp. can be cultivated and recirculated for use in treating and improving aquaculture wastewater [62]. The use of the microalgae Chlorella sorokiniana in an aquaculture estuary revealed significant effectiveness in nutrient removal, as it showed removal rates of 75.6% for ammonium, 84.5% for nitrates, 73.3% for phosphates, and 71.9% for chemical oxygen demand (COD). [46]. The cultivation of Chlorella vulgaris from Nile tilapia effluents showed impressive results in terms of reducing nutrient levels and bioremediation efficiency. Microalgae biomass produces an average nutrient level of 172.91 mg/day for carbohydrates, 141.57 mg/day for proteins, and 150.19 mg/day for lipids. Furthermore, it achieved exceptional bioremediation efficiency, removing 99.8% of nitrate and 99.7% of phosphate from the tilapia effluents. These findings highlight the potential use of *C. vulgaris* in water quality control [63]. Cultivation of Spirulina platensis in aquaculture wastewater also demonstrated a similar beneficial effect [64,65]. The use of different algal species in combination has also been revealed to be effective in bioremediation and biomass production [66–68]. Cultivating microalgae in an aquaponic system proves beneficial to both fish and plants. Integrated fish and vegetable farming supports sustainable aquaculture and hydroponic plant growth in a mutually beneficial ecosystem [69–71] (Table 1).

	Biochemical Composition					Nutrient Removal Efficiency				
Algae	Wastewater Source	Protein (%)	Lipid (%)	Carbohydrate (%)	COD%	Nitrate (%)	Nitrite (%)	Phosphate (%)	Ammonia (%)	References
Chlorella vulgaris	Aquaculture waste water	47.5	9.1	19.1	46-76	100	100	63.1–92.2	23.4	[58]
Chlorella sp.	Aquaculture wase water	55.28	10	24.77	46–76	62	82	63.1–92.2	84–96	[72]
Chlorella vulgaris and Tetradesmus obliquus	Aquaculture wastewater	57	8	16.8	94	100	-	-	99	[62]
Gracilaria birdiae	Aquaculture wase water	12.94	4.82	58.12	78	~100		93.5	34	[73]
Spirulina platensis	Nile tilapia effluent	56	13	25	87.2	97.5	95	98	98	[63]
Scenedesmus sp.	Tilapia	19.5	30.9	35.15	80	77.7	73.8	~100	88.7	[59]

 Table 1. Biochemical composition and removal nutrient efficiency (%) of algae cultivated in wastewater.

Abbreviation: COD-Chemical Oxygen Demand.

4. Applications of Algae in Aquaculture

4.1. Replacements of Fish Meal and Fish Oil

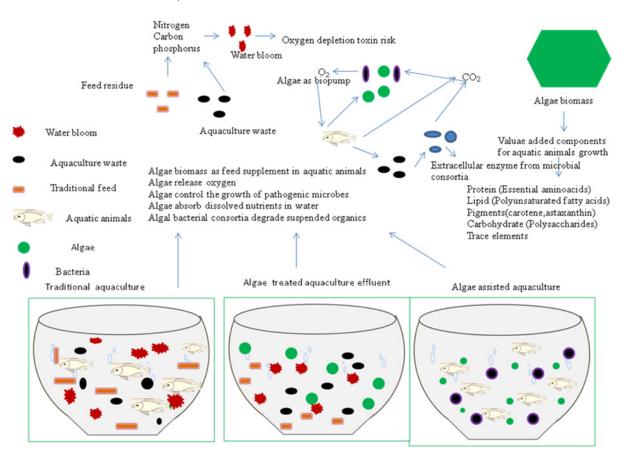
Algae, particularly microalgae, are essential natural nutritional sources in aquatic ecosystems and play an important role in the food chain of aquatic animals. The beneficial effects include improved growth performance, modulation of gut microbiota, and enhanced disease resistance, making them valuable for the aquaculture industry. Furthermore, the substitutions of FM and FO with microalgae contribute to the conservation of marine ecosystems [74–76]. Dietary additives of algae have demonstrated effectiveness as immunomodulators by enhancing survival rates of olive flounder (*Paralichthys olivaceus*) exposed to *Edwardsiella tarda* [77]. Micro- and macroalgae used as a feed supplement have demonstrated positive effects on the green water culture production of various planktivorous species, including Nile tilapia, rohu (*Labeo rohita*), bighead carp (*Hypophthalmichthys nobilis*), catla (*Catla catla*), and shrimp. Microalgae such as Arthrospira (*Spirulina*), *Chlorella*, *Dunaliella*, *Haematococcus*, *Isochrysis*, *Nannochloropsis*, *Pavlova*, and *Schizochytrium* have established themselves as essential nutritional sources, with benefits for aquatic animals [5,18].

4.1.1. Microalgae

In a previous review, Duerr et al. [78] discussed the use of the microalgae Chaetoceros, Thalassiosira, Tetraselmis, Isochrysis, and Nannochloropsis as aquaculture feeds, through Artemia, rotifers, or Daphnia. The dietary inclusion of microalgae supplements represents an innovative and sustainable approach in aquaculture and has garnered increasing attention as an alternative to FM and FO in commercial aquatic feed, enhancing the nutritional value of the feed and promoting environmental sustainability in aquaculture [79–81], which moves the industry closer to achieving sustainable aquaculture [82]. Microalgae species such as Chlorella, Dunaliella, Nannochloropsis, Spirulina, and Scenedesmus are commonly used in aquaculture as feed supplements to improve the health and growth performance of aquatic animals [83]. The combination of algae extracts and organic acids has the potential to partially or completely replace FM and FO in aquafeed [84]. Biomass from species like Dunaliella spp. can serve as a valuable ingredient in aquaculture feeds, particularly for Nile tilapia, and its incorporation can partially replace FM, leading to improved growth and nutrient intake in the cultured species. This practice supports the development of a more sustainable and environmentally friendly aquaculture industry by reducing the dependency on traditional protein sources and promoting resource efficiency [85]. In a subsequent study, it was reported that the incorporation (10%) of Dunaliella sp. in Nile tilapia diets is safe and economically beneficial, as the additive improved fish health and growth performance and reduced the supplementation levels of FO and FM in the diets. This aligns with sustainability goals in aquaculture, emphasizing the responsible use of resources and reducing the reliance on traditional feed ingredients [5].

4.1.2. Macroalgae

The incorporation of macroalgae as a feed additive in an 8-week feeding trial at a dosage of 1000 mg led to significant improvements in growth, antioxidant capacity, and pigmentation in electric yellow cichlid (*Labidochromis caeruleus*) [78]. The combination of dietary supplements containing macroalgae species such as *Ulva* spp., *Gracilaria* spp., and *Fucus* spp. at concentrations ranging from 2.5% to 7.5% yielded several positive effects in European sea bass (*Dicentrarchus labrax*). One of the significant outcomes is the potential to replace traditional feed ingredients FM and FO [86]. The incorporation of algae supplements not only enhances the health and performance of aquaculture species but also contributes to making aquaculture practices more sustainable by reducing the industry's reliance on marine resources. This aligns with the growing emphasis on environmentally responsible aquaculture [84,85,87]. In a previous study with Nile tilapia, the dietary effect of ulvan extract from the green algae *Ulva clathrata* revealed that administration increased hematocrit, phagocytic activity, and white blood cells. However, no significant effect was displayed for growth performance, hemoglobin, red blood cell count, total serum protein,



albumin, and globulin in the dietary levels as a result of the ulvan extract [87,88] (Figure 1, Table 2).

Figure 1. Applications of algae in aquaculture. Algae play diverse and significant roles in aquaculture, contributing to water quality management (nutrient removal, BOD, CO₂ sequestration, and bioremediation) and acting as valuable feed supplements (natural nutrition source of omega-3 fatty acids, pigments, and carotenoids) and providing enhanced growth and reproduction, artemia enrichment, biofloc technology, seedling production, and sustainable practices for aquatic animals. The integration of algae in aquaculture demonstrates its versatility in addressing multiple challenges, from water quality management to providing essential nutrition for aquatic animals. Research and ongoing developments continue to explore ways to optimize the use of algae for sustainable and efficient aquaculture production.

Algae	Algae (Family)	Mode of Algae in Aqua Feed	Aquatic Organisms (Scientific Name)	Aquatic Organisms (Common Name)	Dietary Inclusion Level (g/kg or %)	Replaced Ingredients	Beneficial Effects	References
Arthrospira platensis	Microcoleaceae	Whole	Macrobrachium rosenbergii	Giant river prawn	50%	FM	Increased growth performance and feed conversion efficiency	[89]
Arthrospira platensis	Microcoleaceae	Whole	Oreochromis niloticus	Nile tilapia	10%	FM	Improve growth rate and digestive enzyme activity Enhanced	[90]
Chlorella vulgaris	Chlorellaceae	Whole	Carassius auratus gibelio	Carp	0.4–2.0%	FM	immunoglobulin (Ig) M and D, interleukin-22 (IL)-22, and chemokine (C-C motif) ligand 5 (CCL-5)	[91]
Chlorella vulgaris	Chlorellaceae	Whole	M. rosenbergii	Giant river prawn	4-8%	FM	Enhanced growth rate, immune system, and disease resistance against pathogens	[92]
Dunaliella salina	Dunaliellaceae	Whole	Penaeus monodon	Black tiger shrimp	10%	_	Improved growth rate	[93]
Gracilaria fisheri	Gracilariaceae		P. monodon	Giant tiger prawn	100–200 μg/ml	FM	Enhanced immune system and increased resistance against white spot syndrome virus	[94]
Gracilaria arcuata	Gracilariaceae		O. niloticus	Nile tilapia	20%	FM	Enhanced growth performance	[95]
Nannochloropsis oculata	Monodopsidaceae and Thraustochytriaceae	LEA	O. niloticus	Nile tilapia	33–100%	PR	Enhanced growth rate and nutritional quality of farmed fish	[96]
Pyropia spheroplasts	Bangiaceae	Whole	Apostichopus japonicus	Japanese sea cucumber	50 g/kg	-	Improved growth rate and feed conversion efficiency	[97]
Sargassum wightii	Sargassaceae		Penaeus monodon	Giant tiger prawn	400 mg/L	-	Better growth rate and increased disease resistance	[98,99]
Spirulina sp.	Spirulinaceae	Whole	Cyprinus carpio	Common carp	5 g/kg	-	Higher growth rate and feed conversion ratio	[100]

Table 2. Algae is used as a dietary feed supplement in aquaculture and its effects on aquatic animals.

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Algae	Algae (Family)	Mode of Algae in Aqua Feed	Aquatic Organisms (Scientific Name)	Aquatic Organisms (Common Name)	Dietary Inclusion Level (g/kg or %)	Replaced Ingredients	Beneficial Effects	References
Scenedesmus obliquus	Scenedesmaceae	Whole and LEA	Anarhichas minor	Spotted wolffish	4%	PR	Increased growth performance and skin color Enhanced growth	[101]
<i>Schizochytrium</i> sp.	Thraustochytriacea	e Whole	Salmo salar	Atlantic salmon	100–150 g/kg	FO	performance, fillet quality, nutrient retention efficiency, and blood chemistry	[102]
<i>Schizochytrium</i> sp.	Thraustochytriacea	e Whole	Oncorhynchus mykiss	Rainbow trout	NM	-	Better replacement of FM and FO	[103]
S. platensis	Microcoleaceae	Whole	Pterophyllum scalare	Angel fish	5 g/kg	FM	Better nutrient additives for fish growth	[104]
S. platensis	Microcoleaceae	Whole and LEA	O. niloticus	Nile Tilapia	5–10 g/kg	PR	Better growth and immunity promoter	[105]
S. platensis	Microcoleaceae	Whole	Trachinotus ovatus	Silver fish	5%	FM	Improved growth, body composition and feed utilization	[106]
S. platensis	Microcoleaceae	Whole	C. carpio	Koi carp	5%	FM	Stimulation of the immune system	[100]
Tisochrysis lutea	Isochrysidaceae	-	Sparus aurata	Gilthead seabream	5%	-	Higher growth rate, nutrient utilization, and survival rate	[107]
Ulva rigida	Ulvaceae	-	Mugil cephalus	Common grey mullet	10 mg/kg	-	Enhanced growth response, and antioxidant and immune stimulation	[108]

Abbreviation: Penaeus monodon—P. monodon, Cyprinus carpio—C. carpio, Oreochromis niloticus—O. *niloticus*, Salmo salar—S. salar, Pseudodiaptomus hessei—P. hessei, gram—g, microgram—µg, milligram—mg, kilogram—kg, milliliter—ml, Immunoglobulin—Ig, interleukin—IL, fishmeal—FM, fish oil—FO, lipid-extracted microalgae—LEA, not mention—NM, larval food—LF, protein—PR, digestible protein—DP, apparent digestibility coefficients—ADC.

4.2. Treatment of Aquatic Animal Diseases

The administration of microalgae meal in a 70-day feeding trial with post larval stage of Pacific white shrimp (Litopenaeus vannamei) revealed significant positive impact on disease resistance against V. harveyi infection and various immune parameters, total hemocyte count, phenoloxidase activity, superoxide dismutase activity, and bactericidal activity [109–112]. Polysaccharides derived from the tropical brown algae Turbinaria ornata demonstrated potent antibacterial activity against A. hydrophila, P. aeruginosa, Streptococcus sp., Yersinia enterocolitica, Enterobacter sp., Proteus sp., V. cholerae, V. parahaemolyticus, V. alginolyticus, and E. coli. These findings displayed the potential of algae-derived compounds in combating bacterial infections and promoting the health of aquatic animals [111,113–115]. Various techniques are available for the extraction of polysaccharides from seaweeds, including hot water extraction, enzymaticassisted extraction, microwave-assisted extraction, and ultrasonic-assisted extraction. Each of these methods has advantages and may be chosen based on the specific requirements of the polysaccharide extraction process [112,116]. Scenedesmus quadricauda, a green alga, is not only a good antioxidant but has also demonstrated bactericidal activity against bacteria like Staphylococcus aureus and P. aeruginosa. Additionally, the biomass of S. quadricauda is a rich source of protein, making it suitable for use as a feed supplement in aquatic animal's diets and is a valuable resource for enhancing the nutritional quality of the feeds and promoting health and growth of aquatic species [117]. The dietary incorporation of microalgae such as Sargassum cinereum, Padina gymnospora, and Gracillaria folifera has demonstrated significant antibacterial activity against Pseudomonas spp. in Mozambique tilapia (Oreochromis mossambicus), and these species are important due to their antibacterial and antiviral properties [118,119]. The dietary incorporation of *P. gymnospora* polysaccharide at various concentrations (0.01, 0.1, and 1.0%) showed that inclusion at 0.1% and 1.0% levels increased the production of antibodies and improved the resistance of common carp (Cyprinus carpio) against A. hydrophila and E. tarda. This highlights the potential of *P. gymnospora* polysaccharides as a beneficial dietary supplement by enhancing the immune response and disease resistance in aquatic species [120]. The dietary inclusion of various algae compounds, including polysaccharides, fatty acids, phlorotannins, pigments, lectins, alkaloids, terpenoids, and halogenated compounds, significantly enhanced the antioxidant capacity, immune response, survival rate, and infection resistance against pathogens in Mozambique tilapia [113,121]. Supplements of Azolla pinnata and Nannochloropsis oculata, used in the remediation of environmental pollutants are reported to improve antioxidant capacity, growth performance, immune system function, survival rates, and disease resistance in Nile tilapia [114]. The combination of dietary additives of Chlorella and Spirulina has demonstrated protective effects against A. hydrophila infection in Nile tilapia as these algae-based additives improved the immune response and disease resistance of the fish, contributing to their health and survival [122]. In contrast, dietary supplementation of Dunaliella salina at a dosage of 300 mg/kg for 8 weeks did not lead to changes in phenoloxidase activity and hemocyte count, but administration increased protection against White Spot Syndrome Virus (WSSV) in juvenile black tiger shrimp (Penaeus monodon) compared to control groups. This suggests that D. salina supplementation may not have direct effects on certain immune parameters but can still enhance the shrimp's resistance to specific pathogens like WSSV [9].

4.3. Growth Promoters

The dietary incorporation of *Spirulina* sp. at levels ranging from 5% to 15% in a 60-day feeding trial involving juvenile giant freshwater prawn (*Macrobrachium rosenbergii*) resulted in enhanced growth performance, reduced mortality, and significantly improved feed utilization. This indicates the potential of *Spirulina* sp. as a valuable dietary supplement for improving the health and overall production efficiency of the prawns in aquaculture [96]. *Spirulina* used as a feed supplement is known to be a valuable protein source, and in common carp, its inclusion in the diet has revealed improved growth and increased body weight by more than 25% vs. the control group. [93,123]. The dietary incorporation of *Spirulina* at levels ranging from 5% to 20% significantly enhanced the survival rate, growth,

and feed intake of juvenile Pacific white shrimp and revealed a positive impact on the health and production efficiency of shrimp in aquaculture [124]. In an 11-week study using juvenile Pacific white shrimp, the partial replacement of FM with S. platensis resulted in a significantly improved growth response. This suggests that incorporating S. platensis into the diet can be an effective strategy to enhance the growth performance of shrimp in aquaculture and reduce the dependency of FM [6,100]. The dietary incorporation of the microalgae Scenedesmus almeriensis at various combinations (ranging from 0% to 39%) in a 45-day experiment had a significant positive impact on the growth, body composition, gut microbiota composition, and production of short-chain fatty acids in gilthead sea bream (Sparus aurata) juveniles [125]. The single or combined dietary inclusion of Isochrysis galbana, Pavlova lutheri, and Chaetoceros muelleri significantly enhanced the growth performance and survival of grooved carpet shell (Ruditapes decussatus) larvae well as the biochemical and fatty acid composition during the larval development and underscore the value of algae-based dietary supplements in promoting the health and development of aquatic larvae in aquaculture [10]. The dietary incorporation of the macroalgae Gracilaria arcuata as feed supplement, 10, 20 and 30% to African catfish (*Clarias gariepinus*) revealed significantly lower growth, feed utilization, feed intake, and protein efficient ratio at 20 and 30% inclusion vs. control group and 10% inclusion [126]. In contrast, a later study, using G. arcuata at different levels (20, 40, and 60%) in a 12-week trial with Nile tilapia led to a 20% improvement in growth, enhanced body composition, and increased feed intake compared to control-fed fish [127,128]. In a 60-day feeding trial, the use of flocculated algae-enriched live feed Artemia franciscana resulted in improved growth, survival rates, maintenance of gut microbiota composition, increased villi length, enhanced digestive enzyme activities, improved feed conversion ratio, feed consumption ratio, and better nutrition indices compared to the control group and indicated that the use of algae-enriched live feed can have multiple beneficial effects on the growth, health, and overall performance of catla [129]. Dietary incorporation of different algae or single alga such as A. pinnata, C. vulgaris, S. platensis, and Scenedesmus spp., as well as their extracts, are revealed to be nutrient-rich feed sources for aquatic animals. These feed supplements can partially replace FO and FM, as they significantly improve antioxidant capacity, immune response, nutrient utilization, body weight, and growth performance of aquatic animals and highlight the potential of algae-based feed supplements by enhancing health and production of aquatic species [95,115,130]. Dietary inclusion of the microalgae Pyropia yezoensis extract at different combinations (ranging from 0 to 20 g/kg) in a 9-week feeding trial with olive flounder revealed that a 15% dietary inclusion notably enhanced weight gain, feed efficiency, and immunity and increased the level of PUFAs [92]. Dietary supplementation of red seaweed (Gracilaria pygmaea) at a level of 90 g/kg (GL-90) significantly improved the growth performance of rainbow trout fry [131]. However, the supplementation did not reveal a significant effect on antioxidant responses and digestive activities. Dietary inclusion of 3% Arthrospira platensis significantly improved the growth of Nile tilapia, but administration did not modulate the gut microbiota [132]. Dietary additives of microalgae Spirulina and Schizochytrium, used as replacements for FM, FO or plant oil, significantly improved fish growth and maintained fillet quality and displayed the potential of using these microalgae as sustainable alternatives in aquafeeds to enhance fish growth and the quality of fillets while reducing reliance on traditional protein sources like FM and FO [133]. Dietary supplementation of Sargassum ilicifolium (10%) during an eight-week feeding trial with great sturgeon revealed several positive effects, including enhanced growth performance, hematological parameters (red blood cells (RBCs), white blood cells (WBCs), and hemoglobin levels), immune responses (respiratory bursts, complementary activities, and immunoglobulin M, lysozyme, and serum total protein levels), and defense against Y. ruckeri and highlight the potential of S. ilicifolium as a valuable dietary supplement for improving the health and growth performance of great sturgeon [134]. Dietary inclusion of macroalgae Gracilaria vermiculophylla at a level of 5% in a 91-day feeding trial improved the growth performance of rainbow trout, enhanced the innate immune response, and resulted

in the highest peroxidase, complement, and lysozyme activity [135]. Dietary incorporation of fucoidan-rich algae *Sargassum wightii* extract at various concentrations (1, 2, 3, and 6%) in a 45-day trial experiment using sutchi catfish (*Pangasianodon hypophthalmus*) fingerlings showed that dietary supplementation at 2% notably reduced stress levels, increased the survival rate against *A. hydrophila*, and enhanced immune parameters such as lysozyme activity and respiratory burst activity [136,137]. In a study with Nile tilapia, brown algae *Padina pavonica* was administrated at 2, 4, 6, and 8 g/kg for 45 days, and growth was improved at 8 g/kg inclusion [138]. Furthermore, the study revealed higher values of hematological and biochemical parameters with the 8 g/kg inclusion diet.

4.4. Immunostimulants

Algae feed supplements in aquatic animals play an important role in enhancing the immune system and infection resistance against viral pathogens. These supplements contain various bioactive compounds and nutrients that can bolster health and promote the disease resistance of aquatic animals [139–141]. Dietary incorporation of algae-derived astaxanthin at a concentration of 80 mg/kg improved hemolymph immunological indices, including total hemocyte counts, phagocytic activity of hemocytes, serum anti-superoxide radical activity, serum phenoloxidase activity, serum antibacterial activity, and serum bacteriolytic activity and disease resistance against WSSV, in Pacific white shrimp during a 4-week trial [142], showing the immunomodulatory and disease resistance-enhancing properties of astaxanthin derived from algae. Dietary incorporation of 5% S. platensis and Cladophora in a 60-day experiment significantly improved the immune system response in various fish species, including Nile tilapia, rainbow trout, channel catfish, and great sturgeon [143–146]. This improvement was evidenced by changes in parameters such as RBC and WBC counts, serum bacterial activity, phagocytosis, and lysozyme activity. In addition to immune system enhancement, the dietary supplementation also led to improved growth performance and infection resistance in these fish species, highlighting the potential of algae-based dietary supplements in enhancing the overall health and disease resistance of various fish species. In an 8-week feeding trial, dietary incorporation of *Laurencia caspica* extract at a concentration of 1.5% significantly improved the immune system of rainbow trout by improving immune gene expression, including IL-1 β , lysozyme II, Complement C3, and TNF- α [147]. Furthermore, the supplementation of *L. caspica* extracts improved resistance towards A. hydrophila infection. A recent study by Khanzadeh et al. [148] revealed significantly higher effects of L. caspica extract on immunoglobulin M and complement 3 as well as increased activity of alkaline phosphatase and alanine aminotransferase in Nile tilapia. Furthermore, extract administration considerably improved the survival of the fish challenged with Streptococcus agalactia. Dietary supplementation of sulphated polysaccharides derived from green algae, specifically Codium fragile, resulted in the upregulation of immune-related gene expression in olive flounder, increased expression of genes associated with the immune response, including TNF- α , IFN- γ , IL-1 β , IL-8, and enhanced lysozyme activity [149,150]. The dietary supplementation of green microalgae Chlorella vulgaris (10%) in a Nile tilapia diet revealed several beneficial effects, particularly when the fish was exposed to arsenic, as the supplementation resulted in enhanced serum biomarkers such as aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatase (ALP), creatinine, and urea [151]. Additionally, the immune gene expression was significantly upregulated, with TNF- α increasing 14-fold, TGF- β 1 increasing 13-fold, and IL-1 β increasing 7-fold, indicating the algae's ability to mitigate the negative impact of arsenic exposure in Nile tilapia [152]. Regarding the administration of S. platensis, a recent study [105] revealed that the supplementation in a Nile tilapia diet improved interleukin 10, lysozyme activity, complement 3, and IgM serum levels, as well as survival against A. hydrophila infection.

4.5. Rotifers and Algae

According to Gilbert [153], rotifers are subdivided into four categories as defined by types of food ingested: (1) nanoplankton, (2) microplankton, (3) 5–50 μ m algae, and (4) 5–

250 µm algae. Larval rearing of marine fish, for example, turbot (*Scophthalmus maximus*) and Atlantic halibut (*Hippoglossus hippoglossus*), can result in a bottleneck, and at this early life stage, it is important to transfer essential nutrients from algae to live food [154]. The reference to Yan et al. [155] likely provides valuable insights into the specific methodologies, experimental setups, and findings related to using rotifers for toxicity testing of harmful algae. Authors interested in this topic can benefit from consulting these papers to gain a deeper understanding of the experimental protocols, data interpretation, and potential applications of rotifer-based toxicity testing in the context of harmful algal blooms.

5. Conclusions

Microalgae have garnered attention for their potential benefits in aquaculture, contributing to various aspects of fish health and overall industry sustainability (as a growth enhancer, immunostimulant, antioxidant, and source of disease resistance). Incorporating microalgae into aquaculture practices aligns with the broader goals of sustainable and responsible aquaculture, addressing both economic and environmental considerations [155–159]. Microalgae in aquaculture practices offer a holistic approach to water quality management, nutrient cycling, and environmental sustainability. These organisms contribute to creating a more balanced and self-regulating aquatic ecosystem within aquaculture systems [160]. Consequently, microalgae play a pivotal role in supporting the early life stages of aquatic organisms in aquaculture. Their nutritional content, suitability for larviculture, and contribution to sustainable feed production make them a valuable component in the cultivation of various species, ultimately contributing to the overall success and sustainability of aquaculture operations [133,161–188] and contributing to the conservation of marine biodiversity. Microalgae also help alleviate the pressure on wild fisheries, support lower trophic levels, and reduce the ecological footprint of aquaculture operations, all of which are essential for maintaining the health and diversity of marine ecosystems. Another benefit of microalgae via biotechnology is decrease in the production cost by producing biomass using wastewater under the phytoremediation process. Microalgae's phytoremediation capabilities offer a cost-effective and sustainable solution for managing aquaculture wastewater. Microalgae biomass offers a wide range of positive impacts on aquaculture, spanning from the improvement of aquatic animal health to enhancing the quality of aqua feeds and wastewater treatment. It is important to note that specific regulations can vary between region and country. Aquaculture operators and scientists should be aware of and comply with local regulations to promote the responsible and environmentally friendly use of algae in aquaculture. The use of algae and algae-derived components in aquaculture is indeed a promising avenue for the future, contributing to more sustainable and environmentally responsible practices while simultaneously improving aquatic feed quality and animal health. This aligns with the growing emphasis on responsible and ethical food production, essential for meeting the increasing global demand for seafood.

Author Contributions: S.V.: conceptualization, writing—original draft, literature search, table preparation. E.R.: writing—review and editing, manuscript outline, supervision. H.G.: writing—review and editing. S.H.H.: writing—review and editing, validation. S.A.: writing—review and editing. C.-C.C.: writing—review and editing, validation, supervision, manuscript outline. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Data Availability Statement: Data are contained within the article.

Conflicts of Interest: The authors declare no conflicts of interest.

References

- 1. Turchini, G.M.; Trushenski, J.T.; Glencross, B.D. Thoughts for the future of aquaculture nutrition: Realigning perspectives to reflect contemporary issues related to judicious use of marine resources in aquafeeds. *N. Am. J. Aquacult.* **2019**, *81*, 13–39. [CrossRef]
- 2. FAO. The State of World Fisheries and Aquaculture 2016. In *Contributing to Food Security and Nutrition for All;* FAO: Rome, Italy, 2016; p. 200.
- Charoonnart, P.; Purton, S.; Saksmerprome, V. Applications of microalgal biotechnology for disease control in aquaculture. *Biology* 2018, 7, 24. [CrossRef]
- Markou, G.; Angelidaki, I.; Georgakakis, D. Microalgal carbohydrates: An overview of the factors influencing carbohydrates production, and of main bioconversion technologies for production of biofuels. *Appl. Microbiol. Biotechnol.* 2012, 96, 631–645. [CrossRef]
- 5. Shah, M.R.; Lutzu, G.A.; Alam, A.; Sarker, P.; Kabir Chowdhury, M.A.; Parsaeimehr, A.; Liang, Y.; Daroch, M. Microalgae in aquafeeds for a sustainable aquaculture industry. *J. Appl. Phycol.* **2018**, *30*, 197–213. [CrossRef]
- Spolaore, P.; Joannis-Cassan, C.; Duran, E.; Isambert, A. Commercial applications of microalgae. J. Biosci. Bioeng. 2006, 101, 87–96. [CrossRef] [PubMed]
- Sivaramakrishnan, T.; Swain, S.; Saravanan, K.; Sankar, K.; Roy, S.D.; Biswas, L. In vitro antioxidant and free radical scavenging activity and chemometric approach to reveal their variability in green macroalgae from south Andaman Coast of India. *Turk. J. Fish. Aquat. Sci.* 2017, 17, 639–648. [CrossRef]
- 8. Mukherjee, P.; Pal, R. Algae as antioxidants and effective fish feed. a review. Afr. J. Fish. Sci. 2021, 9, 001–003.
- 9. Reitan, K.I.; Rainuzzo, J.R.; Øie, G.; Olsen, Y. A review of the nutritional effects of algae in marine fish larvae. *Aquaculture* **1997**, 155, 207–221. [CrossRef]
- 10. Aranda-Burgos, J.A.; da Costa, F.; Nóvoa, S.; Ojea, J.; Martínez-Patiño, D. Effects of microalgal diet on growth, survival, biochemical and fatty acid composition of *Ruditapes decussatus* larvae. *Aquaculture* **2014**, 420–421, 38–48. [CrossRef]
- 11. Chen, F.; Leng, Y.; Lu, Q.; Zhou, W. The application of microalgae biomass and bio-products as aquafeed for aquaculture. *Algal Res.* **2021**, *60*, 102541. [CrossRef]
- 12. Van Hai, N. The use of medicinal plants as immunostimulants in aquaculture: A review. Aquaculture 2015, 446, 88–96. [CrossRef]
- 13. Yang, L.; Wang, R.; Lu, Q.; Liu, H. "*Algae aquaculture*" integrating algae-culture with aquaculture for sustainable development. *J. Clean. Prod.* 2020, 244, 118765. [CrossRef]
- 14. Apandi, N.M.; Mohamed, R.M.S.R.; Al-Gheethi, A.; Kassim, A.H.M. Microalgal biomass production through phycoremediation of fresh market wastewater and potential applications as aquaculture feeds. *Environ. Sci. Pollut. Res.* 2019, 26, 3226–3242. [CrossRef]
- 15. Ytrestøyl, T.; Struksnæs, G.; Rørvik, K.A.; Koppe, W.; Bjerkeng, B. Astaxanthin digestibility as affected by ration levels for Atlantic salmon, *Salmo salar. Aquaculture* **2006**, *261*, 215–224. [CrossRef]
- 16. Chen, F.; Xiao, Y.; Wu, X.; Zhong, Y.; Lu, Q.; Zhou, W. Replacement of feed by fresh microalgae as a novel technology to alleviate water deterioration in aquaculture. *RSC Adv.* **2020**, *10*, 20794–20800. [CrossRef]
- 17. Marudhupandi, T.; Inbakandan, D. Polysaccharides in aquatic disease management. Fish. Aquacult. J. 2015, 6, 3. [CrossRef]
- 18. Guerin, M.; Huntley, M.E.; Olaizola, M. *Haematococcus astaxanthin*: Applications for human health and nutrition. *TRENDS Biotechnol.* **2003**, *21*, 210–216. [CrossRef]
- 19. Rahman, M.M.; Khosravi, S.; Chang, K.H.; Lee, S.M. Effects of dietary inclusion of astaxanthin on growth, muscle pigmentation and antioxidant capacity of juvenile rainbow trout (*Oncorhynchus mykiss*). *Prev. Nutr. Food Sci.* **2016**, *21*, 281–288. [CrossRef]
- 20. Biswas, A.; Araki, H.; Sakata, T.; Nakamori, T.; Takii, K. Optimum fish meal replacement by soy protein concentrate from soymilk and phytase supplementation in diet of red sea bream, *Pagrus major. Aquaculture* **2019**, *506*, 51–59. [CrossRef]
- 21. Hamed, I.; Özogul, F.; Özogul, Y.; Regenstein, J.M. Marine bioactive compounds and their health benefits: A review. *Comp. Rev. Food Sci. Food Saf.* **2015**, *14*, 446–465. [CrossRef]
- Monteiro, M.; Santos, R.A.; Iglesias, P.; Couto, A.; Serra, C.R.; Gouvinhas, I.; Barros, A.; Oliva-Teles, A.; Enes, P.; Díaz-Rosales, P. Effect of extraction method and solvent system on the phenolic content and antioxidant activity of selected macro- and microalgae extracts. J. Appl. Phycol. 2020, 32, 349–362. [CrossRef]
- 23. Kolanjinathan, K.; Ganesh, P.; Govindarajan, M. Antibacterial activity of ethanol extracts of seaweeds against fish bacterial pathogens. *Eur. Rev. Med. Pharmacol. Sci.* 2009, 13, 173–177.
- 24. Natrah, F.M.I.; Harah, Z.M.; Sidik, B.J.; Izzatul, N.M.S.; Syahidah, A. Antibacterial activities of selected seaweed and seagrass from port Dickson coastal water against different aquaculture pathogens. *Sains Malays.* **2015**, *44*, 1269–1273. [CrossRef]
- Falaise, C.; François, C.; Travers, M.-A.; Morga, B.; Haure, J.; Tremblay, R.; Turcotte, F.; Pasetto, P.; Gastineau, R.; Hardivillier, Y.; et al. Antimicrobial compounds from eukaryotic microalgae against human pathogens and diseases in aquaculture. *Mar. Drugs* 2016, 14, 159. [CrossRef]
- 26. Benkendorff, K.; Davis, A.R.; Rogers, C.N.; Bremner, J.B. Free fatty acids and sterols in the benthic spawn of aquatic molluscs, and their associated antimicrobial properties. *J. Exp. Mar. Biol. Ecol.* **2005**, *316*, 29–44. [CrossRef]
- 27. Austin, B.; Austin, D.A.; Munn, C.B. *Bacterial Fish Pathogens: Disease of Farmed and Wild Fish*; Springer: Dordrecht, The Netherlands, 2007; Volume 26, p. 552.
- Desbois, A.P.; Mearns-Spragg, A.; Smith, V.J. A Fatty acid from the diatom *Phaeodactylum tricornutum* is antibacterial against diverse bacteria including multi-resistant *Staphylococcus aureus* (MRSA). *Mar. Biotechnol.* 2009, 11, 45–52. [CrossRef]

- 29. Kusumaningrum, H.P.; Zainuri, M. Detection of bacteria and fungi associated with *Penaeus monodon* post larvae mortality. *Proc. Environ. Sci.* 2015, 23, 329–337. [CrossRef]
- 30. Thitamadee, S.; Prachumwat, A.; Srisala, J.; Jaroenlak, P.; Salachan, P.V.; Sritunyalucksana, K.; Flegel, T.W.; Itsathitphaisarn, O. Review of current disease threats for cultivated penaeid shrimp in Asia. *Aquaculture* **2016**, 452, 69–87. [CrossRef]
- 31. Gao, F.; Li, C.; Yang, Z.-H.; Zeng, G.-M.; Feng, L.-J.; Liu, J.-Z.; Liu, M.; Cai, H.-W. Continuous microalgae cultivation in aquaculture wastewater by a membrane photobioreactor for biomass production and nutrients removal. *Ecol. Eng.* **2016**, *92*, 55–61. [CrossRef]
- 32. Ansari, F.A.; Singh, P.; Guldhe, A.; Bux, F. Microalgal cultivation using aquaculture wastewater: Integrated biomass generation and nutrient remediation. *Algal Res.* 2017, 21, 169–177. [CrossRef]
- 33. Han, P.; Lu, Q.; Fan, L.; Zhou, W. A review on the use of microalgae for sustainable aquaculture. *Appl. Sci.* 2019, *9*, 2377. [CrossRef]
- 34. Tibbetts, S.M.; Milley, J.E.; Lall, S.P. Chemical composition and nutritional properties of freshwater and marine microalgal biomass cultured in photobioreactors. *J. Appl. Phycol.* **2015**, *27*, 1109–1119. [CrossRef]
- Silva, B.F.; Wendt, E.V.; Castro, J.C.; de Oliveira, A.E.; Carrim, A.J.; Vieira, J.D.; Sassi, R.; da Costa Sassi, C.F.; da Silva, A.L.; de Oliveira Barboza, G.F.; et al. Analysis of some chemical elements in marine microalgae for biodiesel production and other uses. *Algal Res.* 2015, *9*, 312–321. [CrossRef]
- 36. Yarnold, J.; Karan, H.; Oey, M.; Hankamer, B. Microalgal aquafeeds as part of a circular bioeconomy. *Trends Plant Sci.* 2019, 24, 959–970. [CrossRef]
- Arun, J.; Raghu, R.; Hanif, S.S.M.; Thilak, P.G.; Sridhar, D.; Nirmala, N.; Pugazhendhi, A. A comparative review on photo and mixotrophic mode of algae cultivation: Thermochemical processing of biomass, necessity of bio-oil upgrading, challenges and future roadmaps. *Appl. Energy* 2022, 325, 119808. [CrossRef]
- Oladoja, N.A.; Adelagun, R.O.A.; Ahmad, A.L.; Ololade, I.A. Phosphorus recovery from aquaculture wastewater using thermally treated gastropod shell. *Process Saf. Environ. Prot.* 2015, 98, 296–308. [CrossRef]
- 39. Montero, M.; Lavrador, A.; Oliva-Teles, A.; Couto, A.; Carvalho, A.P.; Enes, P.; Diaz-Rosales, P. Macro- and microalgal extract as functional feed additives in diets for zebrafish juveniles. *Acuacult. Res.* **2021**, *52*, 6420–6433. [CrossRef]
- Kumar, B.R.; Mathimani, T.; Sudhakar, M.P.; Rajendran, K.; Nizami, A.S.; Brindhadevi, K.; Pugazhendhi, A. A state of the art review on the cultivation of algae for energy and other valuable products: Application, challenges, and opportunities. *Renew. Sustain. Energy Rev.* 2021, 138, 110649. [CrossRef]
- Emerenciano, M.G.C.; Martínez-Córdova, L.R.; Martínez-Porchas, M.; Miranda-Baeza, A. Biofloc technology (BFT): A tool for water quality management in aquaculture. *Water Qual.* 2017, 5, 92–109.
- Huang, L.; Li, M.; Ngo, H.H.; Guo, W.; Xu, W.; Du, B.; Wei, Q.; Wei, D. Spectroscopic characteristics of dissolved organic matter from aquaculture wastewater and its interaction mechanism to chlorinated phenol compound. *J. Mol. Liq.* 2018, 263, 422–427. [CrossRef]
- Huang, X.-F.; Ye, G.-Y.; Yi, N.-K.; Lu, L.-J.; Zhang, L.; Yang, L.-Y.; Xiao, L.; Liu, J. Effect of plant physiological characteristics on the removal of conventional and emerging pollutants from aquaculture wastewater by constructed wetlands. *Ecol. Eng.* 2019, 135, 45–53. [CrossRef]
- Tibbetts, S.M.; Yasumaru, F.; Lemos, D. In vitro prediction of the digestible protein content of marine microalgae (*Nannochloropsis granulata*) meals for Pacific white shrimp (*Litopenaeus vannamei*) and rainbow trout (*Oncorhynchus mykiss*). *Algal Res.* 2017, 21, 76–80. [CrossRef]
- 45. Silkina, A.; Kultschar, B.; Llewellyn, C.A. Far-red light acclimation for improved mass cultivation of cyanobacteria. *Metabolites* **2019**, *9*, 170. [CrossRef] [PubMed]
- 46. Guldhe, A.; Ansari, F.A.; Singh, P.; Bux, F. Heterotrophic cultivation of microalgae using aquaculture wastewater: A biorefinery concept for biomass production and nutrient remediation. *Ecol. Eng.* **2017**, *99*, 47–53. [CrossRef]
- 47. He, S.-B.; Xue, G.; Wang, B.-Z. Factors affecting simultaneous nitrification and de-nitrification (SND) and its kinetics model in membrane bioreactor. *J. Hazard. Mater.* **2009**, *168*, 704–710. [CrossRef]
- 48. Mohedano, R.A.; Costa, R.H.R.; Tavares, F.A.; Belli Filho, P. High nutrient removal rate from swine wastes and protein biomass production by full-scale duckweed ponds. *Bioresour. Technol.* **2012**, *112*, 98–104. [CrossRef]
- 49. Rawat, I.; Ranjith Kumar, R.; Mutanda, T.; Bux, F. Dual role of microalgae: Phycoremediation of domestic wastewater and biomass production for sustainable biofuels production. *Appl. Energy* **2011**, *88*, 3411–3424. [CrossRef]
- 50. Oswald, W.J.; Gotaas, H.B. Photosynthesis in sewage treatment. Trans. Am. Soc. Civil Eng. 1957, 122, 73–97. [CrossRef]
- Delanka-Pedige, H.M.K.; Munasinghe-Arachchige, S.P.; Cornelius, J.; Henkanatte-Gedera, S.M.; Tchinda, D.; Zhang, Y.; Nirmalakhandan, N. Pathogen reduction in an algal-based wastewater treatment system employing *Galdieria sulphuraria*. *Algal Res.* 2019, 39, 101423. [CrossRef]
- 52. Fatima, N.; Kumar, V.; Rawat, B.; Jaiswal, K. Enhancing algal biomass production and nutrients removal from municipal wastewater via a novel mini photocavity bioreactor. *Biointerface Res. Appl. Chem.* **2020**, *10*, 4714–4720.
- 53. Fatima, N.; Kumar, V.; Jaiswal, K.; Vlaskin, M.; Gururani, P.; Kumar, S. Toxicity of cadmium (Cd) on microalgal growth, (IC50 value) and its exertions in biofuel production. *Biointerface Res. Appl. Chem.* **2020**, *10*, 5828–5833.
- 54. Torres, M.A.; Barros, M.P.; Campos, S.C.G.; Pinto, E.; Rajamani, S.; Sayre, R.T.; Colepicolo, P. Biochemical biomarkers in algae and marine pollution: A review. *Ecotoxicol. Environ. Saf.* **2008**, *71*, 1–15. [CrossRef]

- 55. Wang, L.; Xiao, H.; He, N.; Sun, D.; Duan, S. Biosorption and biodegradation of the environmental hormone nonylphenol by four marine microalgae. *Sci. Rep.* **2019**, *9*, 5277. [CrossRef]
- 56. Sirakov, I.; Velichkova, K.; Stoyanova, S.; Staykov, Y. The importance of microalgae for aquaculture industry. *Review. Int. J. Fish. Aquat. Stud.* **2015**, *2*, 81–84.
- 57. Han, P.; Lu, Q.; Zhong, H.; Xie, J.; Leng, L.; Li, J.; Fan, L.; Li, J.; Chen, P.; Yan, Y.; et al. Recycling nutrients from soy sauce wastewater to culture value-added *Spirulina* maxima. *Algal Res.* **2021**, *53*, 102157. [CrossRef]
- 58. Velichkova, K.; Sirakov, I.; Stoyanova, S. Biomass production and wastewater treatment from aquaculture with *Chlorella vulgaris* under different carbon sources. *Sci. Bull. Ser. F Biotechnol.* **2014**, *18*, 83–88.
- Jais, N.M.; Apandi, W.M.; Asma, W.; Matias Peralta, H.M. Removal of nutrients and selected heavy metals in wet market wastewater by using microalgae *Scenedesmus* sp. *Appl. Mech. Mater.* 2015, 773, 1210–1214.
- 60. Lu, Q.; Han, P.; Chen, F.; Liu, T.; Li, J.; Leng, L.; Li, J.; Zhou, W. A novel approach of using zeolite for ammonium toxicity mitigation and value-added *Spirulina* cultivation in wastewater. *Bioresour. Technol.* **2019**, *280*, 127–135. [CrossRef] [PubMed]
- 61. Stevčić, Č.; Pulkkinen, K.; Pirhonen, J. Screening of microalgae and LED grow light spectra for effective removal of dissolved nutrients from cold-water recirculating aquaculture system (RAS) wastewater. *Algal Res.* **2019**, *44*, 101681. [CrossRef]
- 62. Tejido-Nuñez, Y.; Aymerich, E.; Sancho, L.; Refardt, D. Treatment of aquaculture effluent with *Chlorella vulgaris* and *Tetradesmus* obliquus: The effect of pretreatment on microalgae growth and nutrient removal efficiency. *Ecol. Eng.* **2019**, *136*, 1–9. [CrossRef]
- 63. Wuang, S.C.; Khin, M.C.; Chua, P.Q.D.; Luo, Y.D. Use of *Spirulina* biomass produced from treatment of aquaculture wastewater as agricultural fertilizers. *Algal Res.* 2016, 15, 59–64. [CrossRef]
- 64. Chen, H.; Zeng, F.; Li, S.; Liu, Y.; Gong, S.; Lv, X.; Liu, B. *Spirulina* active substance mediated gut microbes improve lipid metabolism in high-fat diet fed rats. *J. Funct. Foods* **2019**, *59*, 215–222. [CrossRef]
- 65. Stockenreiter, M.; Haupt, F.; Seppälä, J.; Tamminen, T.; Spilling, K. Nutrient uptake and lipid yield in diverse microalgal communities grown in wastewater. *Algal Res.* **2016**, *15*, 77–82. [CrossRef]
- Godwin, C.M.; Lashaway, A.R.; Hietala, D.C.; Savage, P.E.; Cardinale, B.J. Biodiversity improves the ecological design of sustainable biofuel systems. GCB Bioenergy 2018, 10, 752–765. [CrossRef]
- Tossavainen, M.; Katyal Chopra, N.; Kostia, S.; Valkonen, K.; Sharma, A.K.; Sharma, S.; Ojala, A.; Romantschuk, M. Conversion of biowaste leachate to valuable biomass and lipids in mixed cultures of *Euglena gracilis* and chlorophytes. *Algal Res.* 2018, 35, 76–84. [CrossRef]
- 68. Huo, S.; Liu, J.; Addy, M.; Chen, P.; Necas, D.; Cheng, P.; Li, K.; Chai, H.; Liu, Y.; Ruan, R. The influence of microalgae on vegetable production and nutrient removal in greenhouse hydroponics. *J. Clean. Prod.* **2020**, *243*, 118563. [CrossRef]
- Costache, T.A.; Acién Fernández, F.G.; Morales, M.M.; Fernández-Sevilla, J.M.; Stamatin, I.; Molina, E. Comprehensive model of microalgae photosynthesis rate as a function of culture conditions in photobioreactors. *Appl. Microbiol. Biotechnol.* 2013, 97, 7627–7637. [CrossRef] [PubMed]
- Kumar, V.; Jaiswal, K.K.; Verma, M.; Vlaskin, M.S.; Nanda, M.; Chauhan, P.K.; Singh, A.; Kim, H. Algae-based sustainable approach for simultaneous removal of micropollutants, and bacteria from urban wastewater and its real-time reuse for aquaculture. *Sci. Total Environ.* 2021, 774, 145556. [CrossRef]
- 71. Roy, S.S.; Pal, R. Microalgae in aquaculture: A review with special references to nutritional value and fish dietetics. *Proc. Zool. Soc.* **2015**, *68*, 1–8. [CrossRef]
- 72. Guerrero-Cabrera, L.; Rueda, J.A.; García-Lozano, H.; Navarro, A.K. Cultivation of *Monoraphidium* sp.; *Chlorella* sp. and *Scenedesmus* sp. algae in batch culture using Nile tilapia effluent. *Bioresour. Technol.* **2014**, *161*, 455–460. [CrossRef]
- Marinho-Soriano, E.; Nunes, S.O.; Carneiro, M.A.A.; Pereira, D.C. Nutrients' removal from aquaculture wastewater using the macroalgae *Gracilaria birdiae*. *Biomass Bioenergy* 2009, 33, 327–331. [CrossRef]
- 74. Das, J.; Ghosh, K. Nutrient profiling of five freshwater algae for their prospective use as fish feed ingredients. *Algae Res.* **2023**, *74*, 103173. [CrossRef]
- 75. Khatoon, N.; Sengupta, P.; Homechaudhuri, S.; Pal, R. Evaluation of algae based feed in Goldfish (*Carassius auratus*) nutrition. *Proc. Zool. Soc.* **2010**, *63*, 109–114. [CrossRef]
- Gunathilaka, B.E.; Veille, A.; Tharaka, K.; Shin, J.; Shin, J.; Jeong, J.-B.; Meallet, V.; Lee, K.-J. Evaluation of marine algal (*Ulva* spp./*Solieria* spp.) extracts combined with organic acids in diets for olive flounder (*Paralichthys olivaceus*). Aquacult. Res. 2021, 52, 3270–3279. [CrossRef]
- 77. Pezeshk, F.; Babaei, S.; Abedian Kenari, A.; Hedayati, M.; Naseri, M. The effect of supplementing diets with extracts derived from three different species of macroalgae on growth, thermal stress resistance, antioxidant enzyme activities and skin colour of electric yellow cichlid (*Labidochromis caeruleus*). *Aquacult. Nutr.* **2019**, *25*, 436–443. [CrossRef]
- 78. Duerr, E.O.; Molnar, A.; Sato, V. Cultured microalgae as aquacultured feeds. J. Mar. Biotechnol. 1998, 6, 65–70.
- 79. Tham, P.E.; Lim, H.R.; Khoo, K.S.; Chew, K.W.; Yap, Y.J.; Munawaroh, H.S.H.; Show, P.L. Insights of microalgae-based aquaculture feed: A review on circular bioeconomy and perspectives. *Algal Res.* **2023**, *28*, 103186. [CrossRef]
- 80. Norambuena, F.; Hermon, K.; Skrzypczyk, V.; Emery, J.A.; Sharon, Y.; Beard, A.; Turchini, G.M. Algae in fish feed: Performances and fatty acid metabolism in juvenile Atlantic salmon. *PLoS ONE* **2015**, *10*, e0124042. [CrossRef] [PubMed]
- Ahmad, M.T.; Shariff, M.; Md. Yusoff, F.; Goh, Y.M.; Banerjee, S. Applications of microalga *Chlorella vulgaris* in aquaculture. *Rev. Aquacult.* 2020, 12, 328–346. [CrossRef]
- 82. Raja, R.; Shanmugam, H.; Ganesan, V.; Carvalho, I. Biomass from microalgae: An overview. Oceanography 2014, 2, 1.

- Peixoto, M.J.; Salas-Leitón, E.; Pereira, L.F.; Queiroz, A.; Magalhães, F.; Pereira, R.; Abreu, H.; Reis, P.A.; Gonçalves, J.F.M.; de Ozório, R.O.A. Role of dietary seaweed supplementation on growth performance, digestive capacity and immune and stress responsiveness in European seabass (*Dicentrarchus labrax*). *Aquacult. Rep.* 2016, *3*, 189–197. [CrossRef]
- 84. Zhu, D.; Wen, X.; Xuan, X.; Li, S.; Li, Y. The green alga Ulva lactuca as a potential ingredient in diets for juvenile white spotted snapper *Lutjanus stellatus Akazaki*. J. Appl. Phycol. 2016, 28, 703–711. [CrossRef]
- Attalla, R.; Mikhail, S. Effect of replacement of fish meal protein with boiled full fat soybean seeds and dried algae on growth performance, nutrient utilization and some blood parameters of Nile tilapia (*Oreochromis niloticus*). *Egypt. J. Aquat. Biol. Fish.* 2008, 12, 41–61. [CrossRef]
- Passos, R.; Correia, A.P.; Pires, D.; Pires, P.; Ferreira, I.; Simões, M.; do Carmo, B.; Santos, P.; Pombo, A.; Afonso, C.; et al. Potential use of macroalgae *Gracilaria gracilis* in diets for European seabass (*Dicentrarchus labrax*): Health benefits from a sustainable source. *Fish Shellfish Immunol.* 2021, 119, 105–113. [CrossRef] [PubMed]
- 87. del Rocio Quezada-Rodriguez, P.; Fajer-Avilla, E.J. The dietary effect of ulvan from *Ulva clathrate* on hematological-immunological parameters and growth of tilapia (*Oreochromis niloticus*). *J. Appl. Phycol.* **2017**, *29*, 423–431. [CrossRef]
- 88. Neori, A. "Green water" microalgae: The leading sector in world aquaculture. J. Appl. Phycol. 2011, 23, 143–149. [CrossRef]
- Radhakrishnan, S.; Belal, I.E.; Seenivasan, C.; Muralisankar, T.; Bhavan, P.S. Impact of fishmeal replacement with *Arthrospira* platensis on growth performance, body composition and digestive enzyme activities of the freshwater prawn, *Macrobrachium* rosenbergii. Aquacult. Rep. 2016, 3, 35–44. [CrossRef]
- 90. Abdel-Tawwab, M.; Ahmad, M.H. Live *Spirulina (Arthrospira platensis)* as a growth and immunity promoter for Nile tilapia, *Oreochromis niloticus* (L.), challenged with pathogenic *Aeromonas hydrophila*. *Aquacult. Res.* **2009**, *40*, 1037–1046. [CrossRef]
- 91. Khani, M.; Soltani, M.; Mehrjan, M.S.; Foroudi, F.; Ghaeni, M. The effect of *Chlorella vulgaris* (*Chlorophyta, Volvocales*) microalga on some hematological and immune system parameters of Koi carp (*Cyprinus carpio*). *Iran. J. Ichthyol.* **2017**, *4*, 62–68.
- Radhakrishnan, S.; Saravana Bhavan, P.; Seenivasan, C.; Shanthi, R.; Muralisankar, T. Replacement of fishmeal with *Spirulina* platensis, *Chlorella vulgaris* and *Azolla pinnata* on non-enzymatic and enzymatic antioxidant activities of *Macrobrachium rosenbergii*. J. Basic Appl. Zool. 2014, 67, 25–33. [CrossRef]
- Supamattaya, K.; Kiriratnikom, S.; Boonyaratpalin, M.; Borowitzka, L. Effect of a *Dunaliella* extract on growth performance, health condition, immune response and disease resistance in black tiger shrimp (*Penaeus monodon*). *Aquaculture* 2005, 248, 207–216. [CrossRef]
- Wongprasert, K.; Rudtanatip, T.; Praiboon, J. Immunostimulatory activity of sulfated galactans isolated from the red seaweed Gracilaria fishery and development of resistance against white spot syndrome virus (WSSV) in shrimp. Fish Shellfish Immunol. 2014, 36, 52–60. [CrossRef] [PubMed]
- Younis, E.S.; Al-Quffail, A.S.; Al-Asgah, N.A.; Abdel-Warith, A.W.; Al-Hafedh, Y.S. Effect of dietary fish meal replacement by red algae, *Gracilaria arcuata*, on growth performance and body composition of Nile tilapia *Oreochromis niloticus*. *Saudi J. Biol. Sci.* 2018, 25, 198–203. [CrossRef] [PubMed]
- Mansour, E.; Mounes, H.; Ahmed, K. Effect of *Azolla pinnata* and *Nannochloropsis oculata* on growth performance and immunoresponse of Nile tilapia (*Oreochromis niloticus*) and its resistance to bacterial infection. *Egypt. J. Aquacult.* 2020, 10, 43–62.
- Shahabuddin, A.M.; Khan, M.N.D.; Mikami, K.; Araki, T.; Yoshimatsu, T. Dietary supplementation of red alga *Pyropia spheroplasts* on growth, feed utilization and body composition of sea cucumber, *Apostichopus japonicus* (Selenka). *Aquacult. Res.* 2017, 48, 5363–5372. [CrossRef]
- Sivagnanavelmurugan, M.; Marudhupandi, T.; Palavesam, A.; Immanuel, G. Antiviral effect of fucoidan extracted from the brown seaweed, *Sargassum wightii*, on shrimp *Penaeus monodon* postlarvae against *white spot syndrome virus*. J. World Aquacult. Soc. 2012, 43, 697–706. [CrossRef]
- 99. Zhang, Q.; Qiu, M.; Xu, W.; Gao, Z.; Shao, R.; Qi, Z. Effects of dietary administration of *Chlorella* on the immune status of gibel carp, *Carassius auratus gibelio. Ital. J. Anim. Sci.* 2014, 13, 3168. [CrossRef]
- Nandeesha, M.C.; Gangadhar, B.; Varghese, T.J.; Keshavanath, P. Effect of feeding *Spirulina platensis* on the growth, proximate composition and organoleptic quality of common carp, *Cyprinus carpio* L. *Aquacult. Res.* 1998, 29, 305–312. [CrossRef]
- 101. Knutsen, H.R.; Ottesen, O.H.; Palihawadana, A.M.; Sandaa, W.; Sørensen, M.; Hagen, Ø. Muscle growth and changes in chemical composition of spotted wolffish juveniles (*Anarhichas minor*) fed diets with and without microalgae (*Scenedesmus obliquus*). *Aquacult. Rep.* 2019, 13, 100175. [CrossRef]
- 102. Hart, B.; Schurr, R.; Narendranath, N.; Kuehnle, A.; Colombo, S.M. Digestibility of *Schizochytrium* sp. whole cell biomass by Atlantic salmon (*Salmo salar*). *Aquaculture* **2021**, *533*, 736156. [CrossRef]
- 103. Bélanger, A.; Sarker, P.K.; Bureau, D.P.; Chouinard, Y.; Vandenberg, G.W. Apparent digestibility of macronutrients and fatty acids from microalgae (*Schizochytrium* sp.) fed to rainbow trout (*Oncorhynchus mykiss*): A potential candidate for fish oil substitution. *Animals* 2021, 11, 456. [CrossRef] [PubMed]
- 104. Masha, B.; Moosavi, J.; Montajami, S. Assessment the effect of *Spirulina platensis* as supplemental feed on growth performance and survival rate in angel fish (*Pterophyllum scalare*). J. Fish. Int. **2013**, *8*, 74–77.
- 105. El-Araby, D.A.; Amer, S.A.; Attia, G.A.; Osman, A.; Fahmy, E.M.; Altohamy, D.E.; Tolba, S.A. Dietary *Spirulina platensis* phycocyanin improves growth, tissue histoarchitecture, and immune responses, with modulating immunoexpression of CD3 and CD20 in Nile tilapia, *Oreochromis niloticus*. *Aquaculture* **2022**, *546*, 737413. [CrossRef]

- 106. Lin, H.; Chen, X.; Yang, Y.; Wang, J.; Huang, X.; Huang, Z.; Qi, C. Effect of different levels of *Spirulina platensis* dietary supplementation on the growth, body color, digestion, and immunity of *Trachinotus ovatus*. *Isr. J. Aquac. Bamidgeh* **2016**, *68*, IJA_68.2016.1285.
- 107. Vizcaíno, A.J.; Saéz, M.I.; López, G.; Arizcun, M.; Abellán, E.; Martínez, T.F.; Alarcón, F.J. Tetraselmis suecia and Tisochrysis lutea meal as dietary ingredients for gilthead sea bream (Sparus aurata L.) fry. J. Appl. Phycol. 2016, 28, 2843–2855. [CrossRef]
- 108. Akbary, P.; Aminikhoei, Z. Effect of water-soluble polysaccharide extract from the green alga *Ulva rigida* on growth performance, antioxidant enzyme activity, and immune stimulation of grey mullet *Mugil cephalus*. J. Appl. Phycol. 2018, 30, 1345–1353. [CrossRef]
- 109. Yaakob, Z.; Ali, E.; Zainal, A.; Mohamad, M.; Takriff, M.S. An overview: Biomolecules from microalgae for animal feed and aquaculture. *J. Biol. Res.* 2014, 21, 6. [CrossRef] [PubMed]
- 110. Madeira, M.S.; Cardoso, C.; Lopes, P.A.; Coelho, D.; Afonso, C.; Bandarra, N.M.; Prates, J.A.M. Microalgae as feed ingredients for livestock production and meat quality: A review. *Livest. Sci.* 2017, 205, 111–121. [CrossRef]
- 111. Nonwachai, T.; Purivirojkul, W.; Limsuwan, C.; Chuchird, N.; Velasco, M.; Dhar, A.K. Growth, nonspecific immune characteristics, and survival upon challenge with *Vibrio harveyi* in Pacific white shrimp (*Litopenaeus vannamei*) raised on diets containing algal meal. *Fish Shellfish Immunol.* **2010**, *29*, 298–304. [CrossRef]
- Yao, X.; Lin, Y.; Shi, M.; Chen, L.; Qu, K.; Liu, Y.; Xie, S. Effect of *Schizochytrium limacinum* supplementation to a low fish-meal diet on growth performance, lipid metabolism, apoptosis, autophagy and intestinal histology of *Litopenaeus vannamei*. *Front. Mar. Sci.* 2022, *9*, 1090235. [CrossRef]
- Rajendran, P.; Subramani, P.A.; Michael, D. Polysaccharides from marine macroalga, *Padina gymnospora* improve the nonspecific and specific immune responses of *Cyprinus carpio* and protect it from different pathogens. *Fish Shellfish Immunol.* 2016, 58, 220–228. [CrossRef]
- 114. Goh, K.W.; Kari, Z.A.; Wee, W.; Zakaria, N.N.A.; Rahman, M.M.; Kabir, M.A.; Wei, L.S. Exploring the roles of phytobiotics in relieving the impacts of *Edwardsiella tarda* infection on fish: A mini-review. *Front. Vet. Sci.* **2023**, *10*, 1149514. [CrossRef] [PubMed]
- 115. Badwy, T.M.; Ibrahim, E.; Zeinhom, M. Partial replacement of fishmeal with dried microalga (*Chlorella* spp. and *Scenedesmus* spp.) in Nile tilapia (*Oreochromis niloticus*) diets. In Proceedings of the 8th International Symposium on Tilapia in Aquaculture, Cairo, Egypt, 12–14 October 2008; pp. 801–810.
- 116. Marudhupandi, T.; Kumar, T.T.A. Effect of fucoidan from *Turbinaria ornata* against marine ornamental fish pathogens. J. Coast. Life Med. 2013, 1, 282–286.
- 117. Ren, B.; Chen, C.; Li, C.; Fu, X.; You, L.; Liu, R.H. Optimization of microwave-assisted extraction of *Sargassum thunbergii* polysaccharides and its antioxidant and hypoglycemic activities. *Carbohydr. Polym.* **2017**, 173, 192–201. [CrossRef] [PubMed]
- 118. Rodrigues, D.; Sousa, S.; Silva, A.; Amorim, M.; Pereira, L.; Rocha-Santos, T.A.P.; Gomes, A.M.P.; Duarte, A.C.; Freitas, A.C. Impact of enzyme- and ultrasound-assisted extraction methods on biological properties of red, brown, and green seaweeds from the central west coast of Portugal. *J. Agricult. Food Chem.* 2015, 63, 3177–3188. [CrossRef] [PubMed]
- 119. Al-Asgah, N.A.; Younis, E.-S.M.; Abdel-Warith, A.-W.A.; Shamlol, F.S. Evaluation of red seaweed *Gracilaria arcuata* as dietary ingredient in African catfish, *Clarias gariepinus. Saudi J. Biol. Sci.* **2016**, *23*, 205–210. [CrossRef] [PubMed]
- 120. Arguelles, E. Proximate analysis, antibacterial activity, total phenolic content and antioxidant capacity of a green microalga *Scenedesmus quadricauda* (Turpin) Brébisson. *Asian J. Microbiol. Biotechnol. Environ. Sci.* **2018**, 20, 150–158.
- 121. Thanigaivel, S.; Chandrasekaran, N.; Mukherjee, A.; Thomas, J. Investigation of seaweed extracts as a source of treatment against bacterial fish pathogen. *Aquaculture* 2015, 448, 82–86. [CrossRef]
- 122. Pérez, M.J.; Falqué, E.; Domínguez, H. Antimicrobial action of compounds from marine seaweed. *Mar. Drugs* **2016**, *14*, 52. [CrossRef]
- 123. El-Habashi, N.; Fadl, S.E.; Farag, H.F.; Gad, D.M.; Elsadany, A.Y.; El Gohary, M.S. Effect of using *Spirulina* and *Chlorella* as feed additives for elevating immunity status of Nile tilapia experimentally infected with *Aeromonas hydrophila*. *Aquacult. Res.* 2019, 50, 2769–2781. [CrossRef]
- 124. Nakagawa, H.; Gómez-Díaz, G. Usefulness of *Spirulina* sp. meal as feed additive for giant freshwater prawn, *Macrobrachium rosenbergii*. *Aquacult. Sci.* **1995**, 43, 521–526.
- 125. Güroy, B.; Şahin, İ.; Mantoğlu, S.; Kayalı, S. *Spirulina* as a natural carotenoid source on growth, pigmentation and reproductive performance of yellow tail cichlid *Pseudotropheus acei*. *Aquacult. Int.* **2012**, *20*, 869–878. [CrossRef]
- 126. Maghawri, A.; Marzouk, S.S.; Ezz El-Din, H.M.; Nashaat, M. Effect of brown algae *Padina pavonica* as a dietary supplement on growth performance and health status of cultured *Oreochromis niloticus*. *Egypt. J. Aquat. Res.* **2023**, *49*, 379–385. [CrossRef]
- 127. Hanel, R.; Broekman, D.; De Graaf, S.; Schnack, D. Partial replacement of fishmeal by lyophilized powder of the microalgae *Spirulina platensis* in Pacific white shrimp diets. *Open Mar. Biol. J.* **2007**, *1*, 105. [CrossRef]
- 128. Hemaiswarya, S.; Raja, R.; Ravi Kumar, R.; Ganesan, V.; Anbazhagan, C. Microalgae: A sustainable feed source for aquaculture. *World J. Microbiol. Biotechnol.* 2011, 27, 1737–1746. [CrossRef]
- 129. Vizcaíno, A.J.; López, G.; Sáez, M.I.; Jiménez, J.A.; Barros, A.; Hidalgo, L.; Camacho-Rodríguez, J.; Martínez, T.F.; Cerón-García, M.C.; Alarcón, F.J. Effects of the microalga *Scenedesmus almeriensis* as fishmeal alternative in diets for gilthead sea bream, *Sparus aurata*, juveniles. *Aquaculture* 2014, 431, 34–43. [CrossRef]
- 130. Kandathil Radhakrishnan, D.; Velayudhannair, K.; Schmidt, B.V. Effects of bio-flocculated algae on the growth, digestive enzyme activity and microflora of freshwater fish *Catla catla* (Hamilton 1922). *Aquacult. Res.* **2020**, *51*, 4533–4540. [CrossRef]

- 131. Maisashvili, A.; Bryant, H.; Richardson, J.; Anderson, D.; Wickersham, T.; Drewery, M. The values of whole algae and lipid extracted algae meal for aquaculture. *Algal Res.* **2015**, *9*, 133–142. [CrossRef]
- Choi, Y.H.; Lee, B.-J.; Nam, T.J. Effect of dietary inclusion of *Pyropia yezoensis* extract on biochemical and immune responses of olive flounder (*Paralichthys olivaceus*). Aquaculture 2015, 435, 347–353. [CrossRef]
- 133. Sotoudeh, E.; Mardani, F. Antioxidant-related parameters, digestive enzyme activity and intestinal morphology in rainbow trout (*Oncorhynchus mykiss*) fry fed graded levels of red seaweed, *Gracilaria pygmaea*. Aquacult. Nutr. **2018**, 24, 777–785. [CrossRef]
- 134. Plaza, I.; García, J.L.; Galán, B.; De la Fuente, J.; Bermejo-Poza, R.; Villarroel, M. Effect of *Arthrospira* supplementation on *Oreochromis niloticus* gut microbiota and flesh quality. *Aquacult. Res.* **2019**, *50*, 1448–1458. [CrossRef]
- 135. Trevi, S.; Uren Webster, T.; Consuegra, S.; Garcia de Leaniz, C. Benefits of the microalgae *Spirulina* and *Schizochytrium* in fish nutrition: A meta-analysis. *Sci. Rep.* 2023, *13*, 2208. [CrossRef] [PubMed]
- Yeganeh, S.; Adel, M. Effects of dietary algae (*Sargassum ilicifolium*) as immunomodulator and growth promoter of juvenile great sturgeon (*Huso huso Linnaeus*, 1758). J. Appl. Phycol. 2019, 31, 2093–2102. [CrossRef]
- 137. Araújo, M.; Rema, P.; Sousa-Pinto, I.; Cunha, L.M.; Peixoto, M.J.; Pires, M.A.; Seixas, F.; Brotas, V.; Beltrán, C.; Valente, L.M.P. Dietary inclusion of IMTA-cultivated *Gracilaria vermiculophylla* in rainbow trout (*Oncorhynchus mykiss*) diets: Effects on growth, intestinal morphology, tissue pigmentation, and immunological response. J. Appl. Phycol. 2016, 28, 679–689. [CrossRef]
- Prabu, D.L.; Sahu, N.P.; Pal, A.K.; Dasgupta, S.; Narendra, A. Immunomodulation and interferon-gamma gene expression in sutchi catfish, *Pangasianodon hypophthalmus*: Effect of dietary fucoidan rich seaweed extract (FRSE) on pre and post-challenge period. *Aquacult. Res.* 2016, 47, 199–218. [CrossRef]
- 139. Abdelhamid, A.F.; Ayoub, H.F.; Abd El-Gawad, E.A.; Abdelghany, M.F.; Abdel-Tawwab, M. Potential effects of dietary seaweeds mixture on the growth performance, antioxidant status, immunity response, and resistance of striped catfish (*Pangasianodon hypophthalmus*) against *Aeromonas hydrophila* infection. *Fish Shellfish Immunol.* **2021**, *119*, 76–83. [CrossRef]
- Abdel-Latif, H.M.; Dawood, M.A.; Alagawany, M.; Faggio, C.; Nowosad, J.; Kucharczyk, D. Health benefits and potential applications of fucoidan (FCD) extracted from brown seaweeds in aquaculture: An updated review. *Fish Shellfish Immunol.* 2022, 122, 115–130. [CrossRef] [PubMed]
- 141. Trichet, V.V. Nutrition and immunity: An update. Aquacult. Res. 2010, 41, 356–372. [CrossRef]
- 142. Barman, D.; Nen, P.; Mandal, S.C.; Kumar, V. Immunostimulants for aquaculture health management. *J. Mar. Sci. Res. Dev.* 2013, 3, 134. [CrossRef]
- 143. Carton-Kawagoshi, R.J.; Caipang, C.M. Algal-derived products and their role in shrimp immunity. In *Biotechnological Advances in Shrimp Health Management in the Philippines*. 2015, pp. 73–88. Available online: https://www.semanticscholar.org/paper/Algal-derived-products-and-their-role-in-shrimp-Carton-Kawagoshi-Caipang/2531dde3db4fa5b6385eff75e198bb18e143edba (accessed on 25 January 2024).
- 144. Wang, H.; Dai, A.; Liu, F.; Guan, Y. Effects of dietary astaxanthin on the immune response, resistance to *white spot syndrome virus* and transcription of antioxidant enzyme genes in Pacific white shrimp *Litopenaeus vannamei*. *IFRO* **2015**, *14*, 699–718.
- 145. Promya, J.; Chanagun, C. The effects of *Spirulina platensis* and *Cladophora algae* on the growth performance, meat quality, and immunity-stimulating capacity of the African sharp tooth catfish (*Clarias gariepinus*). *Int. J. Agric. Biol.* **2011**, *13*, 77–82.
- 146. Ragap, H.M.; Khalil, R.H.; Mutawie, H.H. Immunostimulant effects of dietary *Spirulina platensis* on tilapia *Oreochromis niloticus*. J. *Appl. Pharm. Sci.* **2012**, *2*, 26–31.
- 147. Yeganeh, S.; Teimouri, M.; Amirkolaie, A.K. Dietary effects of *Spirulina platensis* on hematological and serum biochemical parameters of rainbow trout (*Oncorhynchus mykiss*). *Res. Vet. Sci.* **2015**, *101*, 84–88. [CrossRef]
- 148. Khanzadeh, M.; Beikzadeh, B.; Hoseinifar, S.H. The Effects of Laurencia caspica Algae Extract on Hemato-Immunological Parameters, Antioxidant Defense, and Resistance against Streptococcus agalactiae in Nile tilapia (Oreochromis niloticus). Aquac. Nutr. 2023, 4, 8882736. [CrossRef]
- Kiadaliri, M.; Firouzbakhsh, F.; Deldar, H. Effects of feeding with red algae (*Laurencia caspica*) hydroalcoholic extract on antioxidant defense, immune responses, and immune gene expression of kidney in rainbow trout (*Oncorhynchus mykiss*) infected with *Aeromonas hydrophila*. Aquaculture 2020, 526, 735361. [CrossRef]
- 150. Yang, Y.; Park, J.; You, S.G.; Hong, S. Immuno-stimulatory effects of sulfated polysaccharides isolated from *Codium fragile* in olive flounder, *Paralichthys olivaceus*. *Fish Shellfish Immunol.* **2019**, *87*, 609–614. [CrossRef] [PubMed]
- 151. Goh, K.W.; Abdul Kari, Z.; Wee, W.; Van Doan, H.; Reduan, M.F.H.; Kabir, M.A.; Seong Wei, L. The roles of polysaccharides in carp farming: A review. *Animals* 2023, *13*, 244. [CrossRef] [PubMed]
- Zahran, E.; Awadin, W.; Risha, E.; Khaled, A.A.; Wang, T. Dietary supplementation of *Chlorella vulgaris* ameliorates chronic sodium arsenite toxicity in Nile tilapia *Oreochromis niloticus* as revealed by histopathological, biochemical and immune gene expression analysis. *Fish. Sci.* 2019, *85*, 199–215. [CrossRef]
- 153. Gilbert, J.J. Food niches of planktonic rotifers: Diversification and implications. Limnol. Oceanogr. 2022, 67, 2218–2251. [CrossRef]
- 154. Ferreira, M.; Sousa, V.; Oliveira, B.; Canadas-Sousa, A.; Abreu, H.; Dias, J.; Valente, L.M. An in-depth characterisation of European seabass intestinal segments for assessing the impact of an algae-based functional diet on intestinal health. *Sci. Rep.* 2023, 13, 11686. [CrossRef] [PubMed]
- 155. Yan, T.; Wang, Y.; Wang, L.; Chen, Y.; Han, G.; Zhou, M. Application of rotifer *Brachionus plicatilis* in detecting the toxicity of harmful algae. *Chin. J. Oceanol. Limnol.* 2009, 27, 376–382. [CrossRef]

- 156. Wassef, E.A.; Saleh, N.E.; Abdel-Latif, H.M. Beneficial effects of some selected feed additives for European seabass (*Dicentrarchus labrax* L.): A review. *Int. Aquat. Res.* 2023, 15, 271–288.
- 157. Øie, G.; Makridis, P.; Reitan, K.I.; Olsen, Y. Protein and carbon utilization of rotifers (*Brachionus plicatilis*) in first feeding of turbot larvae (*Scophthalmus maximus*). Aquaculture **1997**, 153, 103–122. [CrossRef]
- 158. Li, X.D.; Wang, X.Y.; Xu, M.E.; Jiang, Y.; Yan, T.; Wang, X.C. Progress on the usage of the rotifer *Brachionus plicatilis* in marine ecotoxicology. *Aquat. Toxicol.* **2020**, 229, 105678. [CrossRef] [PubMed]
- Mendonça, A.J.C.D.; Rosas, V.T.; Monserrat, J.M.; Romano, L.A.; Tesser, M.B. The inclusion of algae *Gracilaria domingensis* in the diet of mullet juveniles (*Mugil liza*) improves the immune response. J. Appl. Aquacult. 2019, 31, 210–223. [CrossRef]
- Ma, K.; Chen, S.; Wu, Y.; Ma, Y.; Qiao, H.; Fan, J.; Wu, H. Dietary supplementation with microalgae enhances the zebrafish growth performance by modulating immune status and gut microbiota. *Appl. Microbiol. Biotechnol.* 2022, 106, 773–788. [CrossRef] [PubMed]
- 161. Bahi, A.; Ramos-Vega, A.; Angulo, C.; Monreal-Escalante, E.; Guardiola, F.A. Microalgae with immunomodulatory effects on fish. *Rev. Aquacult.* **2023**, *15*, 1522–1539. [CrossRef]
- Kashem, A.H.M.; Das, P.; AbdulQuadir, M.; Khan, S.; Thaher, M.I.; Alghasal, G.; Al-Jabri, H. Microalgal bioremediation of brackish aquaculture wastewater. *Sci. Total Environ.* 2023, 873, 162384. [CrossRef]
- 163. Yazıcı, M.; Naz, M.; Mazlum, Y. A Review of the utilization of laminarin, alginate, and fucoidan polysaccharides from macroalgae for promoting growth performance and health in aquatic organisms. In *Research & Reviews in Agriculture, Forestry and Aquaculture;* Gece Publishing: Ankara, Turkey, 2022; pp. 97–113.
- 164. Abdelrhman, A.M.; Ashour, M.; Al-Zahaby, M.A.; Sharawy, Z.Z.; Nazmi, H.; Zaki, M.A.; Goda, A.M. Effect of polysaccharides derived from brown macroalgae *Sargassum dentifolium* on growth performance, serum biochemical, digestive histology and enzyme activity of hybrid red tilapia. *Aquacult. Rep.* 2022, 25, 101212. [CrossRef]
- Abdulrahman, N.M. Evaluation of *Spirulina* spp. as food supplement and its effect on growth performance of common carp fingerlings. *Int. J. Fish. Aquat. Stud.* 2014, 2, 89–92.
- 166. Pradhan, B.; Bhuyan, P.P.; Ki, J.S. Immunomodulatory, antioxidant, anticancer, and pharmacokinetic activity of ulvan, a seaweedderived sulfated polysaccharide: An updated comprehensive review. *Mar. Drugs.* 2023, 21, 300. [CrossRef]
- 167. García-Márquez, J.; Domínguez-Maqueda, M.; Torres, M.; Cerezo, I.M.; Ramos, E.; Alarcón, F.J.; Balebona, M.C. Potential Effects of microalgae-supplemented diets on the growth, blood parameters, and the activity of the intestinal microbiota in *Sparus aurata* and *Mugil cephalus*. *Fishes* **2023**, *8*, 409. [CrossRef]
- 168. Arney, B.; Liu, W.; Forster, I.P.; McKinley, R.S.; Pearce, C.M. Feasibility of dietary substitution of live microalgae with spray-dried *Schizochytrium* sp. or *Spirulina* in the hatchery culture of juveniles of the Pacific geoduck clam (*Panopea generosa*). Aquaculture 2015, 444, 117–133. [CrossRef]
- 169. Bongiorno, T.; Foglio, L.; Proietti, L.; Vasconi, M.; Lopez, A.; Pizzera, A.; Parati, K. Microalgae from biorefinery as potential protein source for Siberian sturgeon (*A. baerii*) aquafeed. *Sustainability* **2020**, *12*, 8779. [CrossRef]
- Daneshvar, E.; Antikainen, L.; Koutra, E.; Kornaros, M.; Bhatnagar, A. Investigation on the feasibility of *Chlorella vulgaris* cultivation in a mixture of pulp and aquaculture effluents: Treatment of wastewater and lipid extraction. *Bioresour. Technol.* 2018, 255, 104–110. [CrossRef] [PubMed]
- 171. Dang, V.T.; Li, Y.; Speck, P.; Benkendorff, K. Effects of micro and macroalgal diet supplementations on growth and immunity of greenlip abalone, *Haliotis laevigata*. Aquaculture 2011, 320, 91–98. [CrossRef]
- 172. Duy, N.D.Q.; Francis, D.S.; Southgate, P.C. The nutritional value of live and concentrated micro-algae for early juveniles of sandfish, *Holothuria scabra*. Aquaculture 2017, 473, 97–104. [CrossRef]
- 173. Guo, Z.; Liu, Y.; Guo, H.; Yan, S.; Mu, J. Microalgae cultivation using an aquaculture wastewater as growth medium for biomass and biofuel production. *J. Environ. Sci.* 2013, 25, S85–S88. [CrossRef] [PubMed]
- 174. Hawrot-Paw, M.; Koniuszy, A.; Gałczyńska, M.; Zając, G.; Szyszlak-Bargłowicz, J. Production of microalgal biomass using aquaculture wastewater as growth medium. *Water* **2019**, *12*, 106. [CrossRef]
- 175. Hoshino, M.D.F.G.; Marinho, R.D.G.B.; Pereira, D.F.; Yoshioka, E.T.O.; Tavares-Dias, M.; Ozorio, R.O.D.A.; Faria, F.S.E.D.V.D. Hematological and biochemical responses of pirarucu (*Arapaima gigas, Arapaimidae*) fed with diets containing a glucomannan product derived from yeast and algae. *Acta Amaz.* 2017, 47, 87–94. [CrossRef]
- 176. Kiron, V.; Sørensen, M.; Huntley, M.; Vasanth, G.K.; Gong, Y.; Dahle, D.; Palihawadana, A.M. Defatted biomass of the microalga, *Desmodesmus* sp.; can replace fishmeal in the feeds for Atlantic salmon. *Front. Mar. Sci.* **2016**, *3*, 67. [CrossRef]
- 177. Kissinger, K.R.; García-Ortega, A.; Trushenski, J.T. Partial fish meal replacement by soy protein concentrate, squid and algal meals in low fish-oil diets containing *Schizochytrium limacinum* for longfin yellowtail *Seriola rivoliana*. *Aquaculture* 2016, 452, 37–44. [CrossRef]
- 178. Kousoulaki, K.; Mørkøre, T.; Nengas, I.; Berge, R.K.; Sweetman, J. Microalgae and organic minerals enhance lipid retention efficiency and fillet quality in Atlantic salmon (*Salmo salar* L.). *Aquaculture* **2016**, 451, 47–57. [CrossRef]
- 179. Maliwat, G.C.F.; Velasquez, S.F.; Buluran, S.M.D.; Tayamen, M.M.; Ragaza, J.A. Growth and immune response of pond-reared giant freshwater prawn *Macrobrachium rosenbergii* post larvae fed diets containing *Chlorella vulgaris*. *Aquacult. Fish.* **2021**, *6*, 465–470. [CrossRef]

- Nasir, N.M.; Bakar, N.S.A.; Lananan, F.; Hamid, S.H.A.; Lam, S.S.; Jusoh, A. Treatment of African catfish, *Clarias gariepinus* wastewater utilizing phytoremediation of microalgae, *Chlorella* sp. with *Aspergillus niger* bio-harvesting. *Bioresour. Technol.* 2015, 190, 492–498. [CrossRef]
- 181. Pascon, G.; Messina, M.; Petit, L.; Valente, L.M.P.; Oliveira, B.; Przybyla, C.; Tulli, F. Potential application and beneficial effects of a marine microalgal biomass produced in a high-rate algal pond (HRAP) in diets of European sea bass, *Dicentrarchus labrax*. *Environ. Sci. Pollut. Res.* **2021**, *28*, 62185–62199. [CrossRef]
- 182. Fonseca, F.; Fuentes, J.; Vizcaíno, A.J.; Alarcón, F.J.; Mancera, J.M.; Martínez-Rodríguez, G.; Martos-Sitcha, J.A. From invasion to fish fodder: Inclusion of the brown algae *Rugulopteryx okamurae* in aquafeeds for European sea bass *Dicentrarchus labrax* (L., 1758). *Aquaculture* 2023, 568, 739318. [CrossRef]
- 183. Patterson, D.; Gatlin, D.M., III. Evaluation of whole and lipid-extracted algae meals in the diets of juvenile red drum (*Sciaenops ocellatus*). *Aquaculture* **2013**, *416*, 92–98. [CrossRef]
- 184. Sarker, P.K.; Kapuscinski, A.R.; McKuin, B.; Fitzgerald, D.S.; Nash, H.M.; Greenwood, C. Microalgae-blend tilapia feed eliminates fishmeal and fish oil, improves growth, and is cost viable. *Sci. Rep.* **2020**, *10*, 19328. [CrossRef]
- 185. Siqwepu, O.; Richoux, N.B.; Vine, N.G. The effect of different dietary microalgae on the fatty acid profile, fecundity and population development of the calanoid copepod *Pseudodiaptomus hessei* (Copepoda: Calanoida). *Aquaculture* **2017**, *468*, 162–168. [CrossRef]
- 186. Southgate, P.C.; Braley, R.D.; Militz, T.A. Ingestion and digestion of micro-algae concentrates by veliger larvae of the giant clam, *Tridacna noae. Aquaculture* **2017**, 473, 443–448. [CrossRef]
- 187. Srimongkol, P.; Thongchul, N.; Phunpruch, S.; Karnchanatat, A. The ability of marine *Cyanobacterium Synechococcus* sp. VDW to remove ammonium from brackish aquaculture wastewater. *Agricult. Water Manag.* **2019**, 212, 155–161. [CrossRef]
- 188. Liu, T.S.; Wu, K.F.; Jiang, H.W.; Chen, K.W.; Nien, T.S.; Bryant, D.A.; Ho, M.Y. Identification of a far-red light-inducible promoter that exhibits light intensity dependency and reversibility in a *Cyanobacterium*. ACS Synth. Biol. 2023, 12, 1320–1330. [CrossRef] [PubMed]

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