



# Article Techno-Economic Analysis of the Production of Lactic Acid from Lignocellulosic Biomass

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Abstract: This study reports the results of an evaluation of the techno-economic feasibility of a biorefinery with an annual lactic acid production capacity of 100,000 metric tons using lignocellulosic biomass. Corn stover and miscanthus were considered as model feedstocks, and three different fermentation pathways involving bacteria, fungi, and yeast were compared with respect to their ability to convert biomass feedstocks to lactic acid. Equipment, raw materials, utilities and labor requirements, and lactic acid production costs were estimated. The minimum selling price (at a 10% internal rate of return) per metric ton of lactic acid produced from different feedstocks for lactic acid bacteria, fungi, and yeast-based pathways were in the range of USD 1243–1390, USD 1250–1392, and USD 993–1123, respectively, with lower costs for miscanthus. Lactic acid production using genetically engineered yeast strains can eliminate the need for the simultaneous neutralization and recovery of lactic acid, resulting in lower equipment, chemical, and utility requirements and lower lactic acid production costs. Lactic acid production costs were highly sensitive to the conversion rates of sugars into lactic acid, feedstock cost, production plant size, operation hours, and acid hydrolysis reactor costs. Improvements in process conditions and efficiencies and lower costs of equipment and consumables are necessary to utilize lignocellulosic biomass for lactic acid production at lower costs while remaining cost-competitive with respect to first-generation and petroleum-based feedstocks.

Keywords: bioproduct; biochemical; bioeconomy; lignocellulosic feedstock; process modeling

# 1. Introduction

Lactic acid (2-hydroxypropanoic acid; CH<sub>3</sub>–CH(OH)–COOH) is an organic acid found in many organisms and natural products [1]. Lactic acid is one of the most important acids in the industry due to its widespread application as a flavoring agent, bacterial inhibitor, and acidulant and its capacity to be converted to other useful products, such as esters, bio-solvents, and polymers [2,3]. It has been widely used in the food, pharmaceutical, cosmetic, and leather industries for several decades [4]. Lactic acid has also been identified as an important platform chemical that can be further converted into other important chemicals [5]. In recent years, the production of lactic-acid-derived biodegradable polylactic acid and the environmentally benign solvent ethyl lactate has given rise to numerous potential applications for lactic acid, constituting an industry with a projected growth of more than 400% by 2025 compared to 2016 [6–8]. The U.S. Department of Energy's National Renewable Energy Laboratory (NREL) and Pacific Northwest National Laboratory (PNNL) identified a few promising platform chemicals for biorefinery applications, one of which was lactic acid [9,10].

Lactic acid can be produced either through the chemical synthesis of petroleum derivatives or the biotechnological conversion of biobased feedstocks [11]. Lactic acid can be produced from petrochemical sources following the hydrolysis of lactonitrile with a strong acid. The lactic acid and byproduct mixture is then esterified using methanol to produce methyl lactate, which is hydrolyzed to obtain lactic acid [2,12,13]. However,



Citation: Manandhar, A.; Shah, A. Techno-Economic Analysis of the Production of Lactic Acid from Lignocellulosic Biomass. *Fermentation* 2023, 9, 641. https://doi.org/ 10.3390/fermentation9070641

Academic Editors: Adarsh Kumar, Bijoy Biswas and Jeff Czajka

Received: 5 June 2023 Revised: 1 July 2023 Accepted: 5 July 2023 Published: 8 July 2023



**Copyright:** © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). this petroleum-based route can only produce a racemic mixture of L- and D-lactic acid, constituting a major disadvantage of this approach. Optically pure L- or D-lactic acid can produce poly-lactic acid with higher crystallinity and a higher melting point than the racemic mixture [14,15]. Furthermore, the production of lactic acid using the petroleum-based pathway requires pure forms of lactonitrile and sulfuric acid, thus substantially increasing the production cost of the resulting lactic acid [6,16]. In addition, the limited reserves of fossil fuels that must be drawn upon to employ this pathway render it an unsustainable approach for lactic acid production.

In recent years, lactic acid production via the fermentation of renewable raw materials containing sugars has gained more attention from the research community and the lactic acid industry [11,17,18]. Compared to the petroleum route, the microbial lactic acid fermentation route utilizes renewable biomass, requires low production temperatures and lower amounts of energy, and produces single-isomer lactic acid at high purity [19]. The ability to produce pure L-lactic acid is important for the food and pharmaceutical industries because the human body can only incorporate L-lactic acid and D-lactic acid can be harmful to humans [12,17,20]. Presently, more than 90% of lactic acid produced globally is manufactured through the microbial fermentation pathway [13]. Biobased lactic acid is one of the few chemicals that holds great potential to be utilized for a variety of applications.

Lactic acid can be produced from different biomass sources, which include starch and sugar-based feedstocks such as corn grain, sugarcane, and lignocellulosic feedstocks including corn stover and miscanthus. Lactic acid production pathways differ for different biomass feedstocks. Lignocellulosic feedstock requires pretreatment and enzymatic hydrolysis to release the sugars prior to fermentation. In addition, different microorganisms, including lactic acid bacteria, fungi, or yeast strains, can be used for the fermentation of the sugars into lactic acid [5]. The lactic acid yield varies depending on the types of microorganisms and sugar used for fermentation [12]. Also, differences exist in the techniques used for the purification and recovery of lactic acid based on the microorganisms used for fermentation.

Many experimental studies [21–25] have shown that lactic acid can be produced from lignocellulosic feedstocks. However, there are only a few studies [26,27] on the techno-economic feasibility of this process and the potential of lignocellulosic feedstocks to be converted into lactic acid using different fermentation microorganisms. The corresponding lactic acid production costs and economic feasibility depend on the cost of the feedstocks, the pretreatment approach employed, the fermentation organisms used, and the feedstock-to-lactic-acid conversion yields. Prior research on the techno-economic feasibility of producing lactic acid from sugarcane bagasse indicated that cellulose-based processes lead to higher rates of lactic acid production and, consequently, lower costs compared to hemicellulose-based processes, while a yeast-based pathway in which the production of gypsum was avoided had the lowest cost [26]. Since lactic acid has enormous growth potential, exploring the techno-economic feasibility of different pathways could contribute to cost-effective lactic acid production from different biobased feedstocks. Incorporating multiple feedstock sources in a biorefinery would reduce the risks associated with the shortage of the supply of feedstocks for biorefineries in the future. Thus, the objective of this study was to evaluate the techno-economic feasibility of lactic acid production using a lignocellulosic feedstock. Corn stover and miscanthus were selected as the feedstocks for techno-economic modelling due to their potential in terms of being the main lignocellulosic feedstock for biobased industries in the U.S. in the near future [28]. This study considered three fermentation pathways using one of the three different lactic acid producing microorganisms: (1) bacteria, (2) fungi, or (3) yeast.

# 2. Materials and Methods

### 2.1. System Overview

The annual capacity of the existing petroleum- and starch-based lactic acid production facilities vary from a few thousand to about 140,000 t [29]. Larger lactic acid produc-

tion facilities with production capacities above 75,000 t have been established in recent years [30,31], and the lactic acid market is expected to grow substantially in the future [8]. Thus, a lignocellulosic-feedstock-based lactic acid biorefinery with an annual production capacity of 100,000 t, which is on the higher end of the capacity scale for existing petroleum-and starch-based biorefineries, was selected for this analysis.

The two feedstocks considered for this analysis were corn stover and miscanthus. These feedstocks were selected based on their availability and potential to be utilized as lignocellulosic feedstocks for lactic acid biorefinery in current and future scenarios according to the Billion Ton Report and other studies [21,22,24,25,28]. The cellulose, hemicellulose, and lignin compositions of corn stover and miscanthus considered for this analysis are provided in Table 1.

This study analyzed three pathways for fermentation using (1) lactic acid bacteria (LAB), (2) fungi, or (3) yeast. Traditionally, lactic acid has been produced from bacteria belonging to the genera Lactobacillus, Lactococcus, Streptococcus, Bacillus, and Enterococcus. The optimal production conditions for bacterial lactic acid fermentation include a pH in the range of 5 to 7 and a temperature in the 30–45 °C range [12,20]. The lactic acid yields and productivity of the LAB-based process are usually higher compared to those associated with fungal production (Table 1). However, this fermentation pathway (using bacteria for lactic acid production) has limitations. LAB require more nutrients, an organic source of nitrogen, and sterile conditions for operation [32]. Also, the lactic acid produced from the process needs to be neutralized in order to maintain the pH levels required for effective fermentation and recovery in the downstream processes [12,33,34]. Fungal species of the Rhizopus genus can also be used to convert the sugars in biomass to produce lactic acid. The use of fungi during fermentation offers several advantages compared to the use of LAB, such as their amylolytic characteristics enabling them to ferment both hexose and pentose sugars and low nutrient requirements resulting in minimal recovery and purification steps [35]. Studies have indicated that Rhizopus strains present better growth under nutrient-limited environments than LAB [36,37]. However, the corresponding lactic acid yield is usually lower, as other products, such as ethanol and fumaric acid, are also formed during fermentation [38]. In addition, the fermentation broth also needs to be aerated adequately to achieve higher lactic acid yields. The final fermentation pathway considered for this analysis consisted of the use of yeast for the fermentation of feedstock into lactic acid. Cargill Inc. has identified a yeast strain that can tolerate low pH during lactic acid fermentation [31]. This eliminates the need to neutralize the lactic acid as it is produced and discards the recovery step. Furthermore, the yeast strains selected using targeted engineering have higher lactic acid yields (Table 1) [31].

## 2.2. Discrete Production Processes, Sections, and Data Sources

The process model included all the unit operations required to convert the lignocellulosic feedstocks (corn stover and miscanthus) into lactic acid, which are grouped into multiple discrete sections: feedstock preparation, pretreatment, fermentation, product recovery, and stillage utilization (Figure 1). The different assumptions, distinct sections, and processes considered for this study are discussed in the following sections (Table 1).

**Table 1.** Technical parameters considered for lactic acid production and their ranges of values used for sensitivity analysis.

Parameters	Unit	Average Value	Pessimistic Value	Optimistic Value
Plant size	t/year	100,000	80,000	120,000
Annual operation hours	h	7920	7560	8280
Feedstock cost (corn stover) [39,40]	USD/t	119	150	87
Feedstock cost (miscanthus) [41,42]	USD/t	89	116	66
Feedstock moisture content	%	20	25	15
Cellulose content in corn stover [43,44]	%	38	35	40
Cellulose content in miscanthus [45–49]	%	46.7	35.1	52.2

#### Table 1. Cont.

Parameters	Unit	Average Value	Pessimistic Value	Optimistic Value
Hemi-cellulose content in corn stover [43,44]	%	30	17	35
Hemi-cellulose content in miscanthus [45–49]	%	29.3	19.2	34.0
Lignin content in corn stover [43,44]	%	18	7	21
Lignin content in miscanthus [45–49]	%	11.3	9.2	16.5
Pretreatment—Acid hydrolysis				
Cellulose to glucose conversion [50]	%	9.9		
Hemicellulose to xylose conversion [50]	%	90		
Lignin to soluble lignin [50]	%	5		
Sulfuric acid cost [51]	USD/t	70	94	57
Acid hydrolysis reactor cost [50]	USD	19,812,400	23,774,880	15,849,920
Pretreatment—Enzymatic hydrolysis				
Residence time for enzymatic hydrolysis [50]	h	84	100	60
Enzymatic hydrolysis temperature [50]	°C	45		
Total solids loading [50]	%	20		
Cellulase loading (mg enzyme protein/g cellulose) [50]	mg	20		
Cellulose to glucose conversion [50]	%	95		
Hemicellulose to xylose conversion [50]	%	60		
Enzyme cost [52]	USD/t	10.15	15	5
Ammonia cost [53]	USD/t	496	771	385
Enzymatic hydrolysis reactor cost [50]	USD	837,000	1,004,400	669,600
Fermentation		,	, ,	,
Fermentation duration [50]	h	48	65	30
Glucose to lactic acid conversion using LAB [54,55]	%	90	80	95
Xylose to lactic acid conversion using LAB [21,56]	%	70	60	85
Glucose to lactic acid conversion using fungi [38,57]	%	85	75	92
Xylose to lactic acid conversion using fungi [23,25]	%	80	70	88
Glucose to lactic acid conversion using yeast [31,58]	%	93	85	95
Xylose to lactic acid conversion using yeast [19]	%	60	46.4	69.6
Nutrient use [50]	%	0.35		
Nutrient cost [50]	USD/k	g 0.05	0.02	0.08
Diammonium phosphate fraction [50]	%	0.6		
Fermentation reactor cost [50]	USD	837,000	1,004,400	669,600
Product recovery				
Lime cost [59]	USD/t	110	150	90
Gypsum use cost *	USD/t	-50	-100	8
Distillation temperature (after esterification) [34]	°C	101		
Distillation temperature (after hydrolysis) [34]	°C	66		
Drying temperature [34]	°C	150		
Methanol cost [60]	USD/t	442	530.4	353.6
Stillage utilization				
Anaerobic digestion temperature [61]	°C	35		
Overall heat loss during steam generation [62]	%	5		
Multistage turbine efficiencies [62]				
Stage 1	%	67		
Stage 2	%	60		
Power generation efficiency [62]	%	90		

Note: \* Gypsum management approach. Gypsum can be either disposed of (shown as negative value) or utilized as a byproduct (shown as positive value) [63,64].

# 2.2.1. Feedstock Collection and Preparation

This study considered feedstocks with 20% average moisture content that are intended to be delivered to the production facility in bale format. The costs of the biorefinery gate-delivered feedstocks (Table 1) are estimated by considering the size of the lactic acid biorefinery established in the Midwestern U.S. and the costs of different production, harvest, and post-harvest operations, obtained from the literature, for delivering the feedstocks to a lactic acid biorefinery of the selected size. The potential yields of feedstocks are also considered to estimate the feedstock delivery costs. After the feedstocks are unloaded in the biorefinery, the corn stover and miscanthus feedstocks are stored in a biorefinery yard. The feedstocks are then ground to a particle size less than 6 mm using a hammer mill, which reduces the crystallinity of cellulose and improves thee feedstocks' digestibility during the hydrolysis process [50]. This study assumed that the cellulose, hemicellulose,



lignin, and ash fractions of the feedstock were within the ranges reported for corn stover and miscanthus (Table 1) [43–46,65].

Figure 1. Overview of steps in the conversion of lignocellulosic feedstock into lactic acid.

#### 2.2.2. Pretreatment

The feedstocks considered in this study are composed of complex carbohydrates that need to be broken down to their monomeric sugars before they can be fermented into lactic acid. Pretreatment breaks the lignin barriers and the polymeric bonds in the carbohydrates present in the feedstocks and renders the sugars readily available for fermentation. Among the different pretreatment methods available, the dilute acid pretreatment process is widely used in commercial production, mainly due to its lower cost and high sugar yields [66,67]; thus, it was considered for this study. Sulfuric acid concentrations of 0.5–2 wt.% were used for pretreatment of biomass based on different literature [68–71]. However, a sulfuric acid concentration of 1 wt.% and a solid loading ratio of 30 wt.% were considered for this study based on the recommendations from studies conducted by NREL [50,66]. The process conditions and conversion rates for cellulose and hemicellulose to glucose and xylose for acid hydrolysis were obtained from data on pretreatment and hydrolysis of biomass feedstocks from recent research [50,66,72] and are summarized in Table 1.

The pretreated slurry was then conditioned in a reactor for 30 min, where it was treated with ammonia to increase its pH to 5–6, which is a suitable range for enzymatic hydrolysis [50]. Despite its considerably higher cost, ammonia is preferred over lime for the conditioning of the hydrolysate slurry because it is more effective at conditioning due to its higher miscibility, its reduced propensity to degrade sugars (1–2% sugar loss) in the slurry compared to liming (up to 13% sugar loss), and the fact that is eliminates the separation step that would be required for removing gypsum produced while conditioning with lime [50].

The conditioned slurry includes complex carbohydrates, which are then hydrolyzed into soluble sugars using enzymes; subsequently, these sugars can be readily fermented by different microorganisms. Cellulases and hemicellulases are two general categories of enzymes used for enzymatic hydrolysis. A mixture of cellulases and hemicellulases are considered for effective hydrolysis leading to decreased hydrolysis times and process costs [73,74]. Sugar conversion rates during enzymatic hydrolysis (Table 1) were obtained from recent studies [50].

#### 2.2.3. Fermentation

The hydrolysate obtained after the hydrolysis of lignocellulosic biomass includes a mixture of cellulose and hemicellulose-derived sugars and traces of furfural, acetic acid, and 5-hydroxymethyl furfural, which are fermented in presence of either bacteria, fungi, or yeasts to produce lactic acid. As lactic acid is formed during fermentation, the pH of

the fermentation slurry progressively decreases, which can potentially affect the lactic acid yield depending on the type of microorganism used. Depending on the microorganism used and its tolerance to the low pH conditions present during fermentation, this study evaluated three pathways of fermentation using LAB, fungi, and yeast. The fermentation yields for different pathways are provided in Table 1.

## 2.2.4. Lactic Acid Separation and Recovery

Lactic acid is separated and purified before it can be used for other applications. Among the few routes suggested for lactic acid recovery and purification, the lactic acid separation process in which lime (calcium hydroxide) is used to neutralize the fermentation broth, followed by the recovery of lactic acid, is preferred for commercial scale production due to its low overall cost [34]; thus, it was considered for this study. Sulfuric acid is then added to the broth including calcium lactate to recover the lactic acid while simultaneously producing gypsum. However, lactic acid produced from genetically engineered yeast does not need to be neutralized and precipitated, as discussed above, and usually undergoes filtration. The recovered lactic acid also contains other impurities, such as other organic acids and residual sugars, and thus requires further purification [34,75]. Purification processes such as esterification, hydrolysis, and distillation have been selected for all three pathways [34,76]. Lactic acid is esterified using methanol to produce methyl lactate. The impurities in the methyl lactate broth are separated using distillation and used as stillage, whereas the distilled methyl lactate is hydrolyzed to produce lactic acid of higher purity.

## 2.2.5. Stillage Utilization

Commercial cellulosic lactic acid production produces large quantities of stillage, which includes wastewater, unutilized sugars, and lignin obtained during different conversion steps during lactic acid production. The quantities of wastewater, residual sugars, and lignin are based on the sugar conversion rates during pretreatment and fermentation and the requirements of different chemicals and their conversion rates during pretreatment, fermentation, and product recovery steps. This study considered stillage utilization via the anaerobic digestion of stillage followed by the combustion of biogas to produce steam. In this process, the stillage stream obtained from the purification and recovery unit is separated into solid and liquid fractions using a pressure filter [50]. The solid fraction along with lignin is sent to the boiler, and the liquid fraction is sent to an anaerobic digestion unit, where the sugars and wastewater are converted to biogas, which is sent to the boiler. The steam from the boiler is sent to a multistage turbine, which produces electricity and process steam. The electricity and steam produced are utilized in different processes.

# 2.3. Techno-Economic Modeling Overview

# 2.3.1. Process Modeling

The lactic acid production process for different pathways and feedstocks were modelled using SuperPro Designer v10. The assumptions made in this study were based on information from the literature. This study considered a lactic acid biorefinery operating 24 h/day and 330 days/year. The main input parameters for the techno-economic model included performance parameters (efficiencies and productivity of different processes and the equipment, consumables, and energy/fuel requirements for each process), temporal parameters (feedstock loading/unloading and preparation time; process residence times; and heating and cooling times), and quality parameters (conversion rates and yields for different conversion steps). Equipment types and size, labor and utilities requirements, and their unit costs were considered for each process in the model. The process model results included the details of materials requirements and flow through different processes and their utilities requirements.

### 2.3.2. Economic Analysis

The lactic acid production costs include total capital investments and annual operating costs. The total capital investment cost includes direct fixed cost (DFC), working capital, and start-up cost. The direct fixed cost includes direct costs incurred for total equipment purchases (PCs) and installation in a facility (Table 2). Equipment size and cost considered for this study were based on existing biochemical plants [50], which were adjusted for the analysis year 2022 to account for suitable equipment capacities and inflation (Supplementary Materials, Table S1). In addition, this study also estimated indirect costs such as construction and engineering costs, contractor's fees, and contingency. This study assumed a working capital for one month of operation. Working capital includes costs for raw materials, consumables, labor, and utilities and ensures that a biorefinery can continue its operations for a short-term. This study also considered start-up cost, which includes expenses incurred to establish and start a new biorefinery. Start-up cost is a one-time cost that covers registration and salaries/wages while developing the facility, and it is assumed to be 5% of direct fixed capital cost.

Table 2. Economic parameters for lactic acid production facility.

Time Parameters	Values	Capital Investment Parameters (Contd.)	Values
Analysis year <sup>a</sup>	2022	Buildings (% of PC) [62]	45
Year construction starts <sup>a</sup>	2022	Yard improvement (% of PC) [62]	15
Construction period (months)	18	Auxiliary facilities (% of PC) [62]	40
Start-up period (months)	12	Plants' indirect cost parameters	
Project lifetime (years)	30	Engineering [62]	20
Inflation rate (%) <sup>b</sup>	2.2	Construction [62]	20
Financing parameters		Contractor's fees [62]	5
Equity (%)	40	Contingencies [62]	10
Loan term (years)	12	Annual operating cost parameters	Values
Loan interest (%) [50]	8	Facility dependent costs	
Depreciation method [50]	Straight line	Equipment maintenance (% of PC) [62]	10
Depreciation period (years) [50]	15	Insurance (% of DFC) [62]	1
Income tax rate (%) [50]	40	Local taxes (% of DFC) [62]	2
Capital investment parameters		Overhead expense (% of DFC) [62]	5
Plants' direct fixed costs (DFC) parameters		Labor rate (USD/h) [62]	57
Process piping (% of total equipment purchase cost (PC)) [62]	35	Utilities costs	
Instrumentation (% of PC) [62]	40	Electricity (USD/kWh) [62]	0.07
Insulation (% of PC) [62]	5	Steam $(USD/t)$ [62]	12
Electrical (% of PC) [62]	10	Cooling water (USD/t) [62]	0.05

Note: <sup>a</sup> Modeling assumptions. <sup>b</sup> The inflation rate value is the average inflation rate in the U.S. from 2000 to 2021 [77].

The annual operating cost includes the costs associated with the facility, raw materials, consumables, utilities, labor, quality control, and waste management. The costs related to the facility include those related to the maintenance of the facility itself, equipment maintenance, and other costs such as insurance, taxes, and overhead expenses. The costs of feedstocks, consumables, and utilities were obtained from the literature and are summarized in Tables 1 and 2. The labor rate includes the basic rate, benefits, and administration. The economic lifetime of a lactic acid production facility is assumed to be 30 years for this analysis. An internal rate of return (IRR) of 10%, assumed to ensure some profitability of the production facility, and discounted cash flow analysis were adopted and employed, respectively, to estimate the minimum selling price of the lactic acid [50]. Considering the

same 10% IRR, financial analysis was conducted to estimate the net present value (NPV), return on investment (ROI), payback period, and gross margin.

#### 2.4. Sensitivity Analysis

The lactic acid production cost for the base case scenario was estimated using average values of different input parameters. Sensitivity analysis was performed to evaluate the effect of the most pessimistic and optimistic values of different input parameters on lactic acid production cost (Table 1). Sensitivity analysis was performed for a corn-stover-based lactic acid production facility, assuming similar trends for miscanthus based lactic acid production facility. This study considered a plant with an annual lactic acid production capacity of 100,000 t. However, variation in plant size could be possible depending on the production plant's location, availability of feedstock, and product demand. Thus, pessimistic and optimistic values for plant size were assumed to be -20% and +20% of the plant size for the base case, respectively. Variation in annual operation hours could be due to maintenance requirements, feedstock availability, and/or market demand. The delivery cost of feedstock can vary significantly depending on the availability of biomass, the location of the production facility, and the selected feedstock logistics systems. The moisture content of the delivered feedstock can vary from approximately 15 to 25%, depending on the time at which the biomass is harvested, the weather conditions during harvest, and storage conditions [78]. The cost of equipment and consumables such as enzymes, sulfuric acid, ammonia, nutrients, lime, and methanol can vary depending on their availability, market demand, and the location of the production facility. The gypsum produced in bacterial and fungal based pathways can be considered as a byproduct or waste, depending on the management practice, and can be sold [63] or disposed of. However, gypsum disposal adds to the lactic acid production costs [64].

#### 3. Results and Discussion

#### 3.1. Material Requirement

Depending on the material balances for the different conversion steps of lactic acid production (Figure 2), lactic acid production pathways using yeast for fermentation required higher quantities for both the corn stover and miscanthus feedstocks. Although the glucose-to-lactic-acid yield is high for the fermentation pathway using yeast, the xyloseto-lactic-acid yield is much lower compared to the fermentation pathways using bacteria and fungi. Thus, the overall feedstock requirements for lactic acid production using the yeast pathway were higher compared to the bacterial and fungal pathways. For all fermentation pathways, the feedstock requirements for miscanthus were lower compared to corn stover. The acid, ammonia, and enzyme requirements during feedstock pretreatment for all three fermentation pathways using both feedstocks were based on the quantity of the feedstocks and the sugars obtained from the feedstocks (Figure 2) and were similar to the requirements reported by previous studies based on corn stover feedstock [50,66,79]. The bacteria- and fungi-based fermentation pathways required similar quantities of lime (30% calcium hydroxide) and sulfuric acid to neutralize and recover the lactic acid and produced similar quantities of gypsum, as the lactic acid production was considered the same. As the neutralization of lactic acid was not necessary for the fermentation pathway using yeast tolerant to low pH conditions, lime and sulfuric acid were not required. Previous studies [26,34] also discussed and reported similar requirements for a neutralizing agent and acid for the two lactic acid fermentation pathways. For all pathways, the same quantity of methanol was used for lactic acid esterification. The water required for different conversion processes (Figure 2) varied and corresponded to the water required to maintain the solid loadings and concentrations for chemicals used for different processes [50]. Compared to corn stover, miscanthus had higher cellulose and hemicellulose content, resulting in higher sugar content. Thus, lower quantities of miscanthus feedstock than corn stover were required to produce the same quantity of lactic acid from a production facility. The miscanthus feedstock also produced less stillage than corn stover, as the miscanthus feedstock had

a higher fraction of fermentable sugars and less of the remaining fraction was transferred as stillage. This resulted in lower amounts of biogas, steam, and electricity production for lactic acid production.

### 3.2. Equipment, Utilities, and Labor Requirements

All three pathways required similar amounts and sizes of equipment for feedstock preparation, pretreatment (acid and enzymatic hydrolysis), and fermentation, as the feedstock and material flows were not significantly different. For the fungi-based pathway, an air compressor and a filtration unit were also required to provide aeration during fermentation. For the yeast-based pathway, lactic acid was not neutralized during fermentation; thus, equipment for the formation of gypsum from calcium lactate and the removal of gypsum from the mixture were not necessary. For the two feedstocks, there were differences in equipment sizes based on the material flow.



**Figure 2.** Material flow for lactic acid production using corn stover and miscanthus feedstocks for three pathways using bacteria, fungi, and yeast for fermentation of sugars into lactic acid. (Note: The material flows are presented in t/h; electricity production is presented in megawatts; the material flows for each component for different feedstocks and fermentation pathways are presented as follows: Line 1—corn stover–bacteria, corn stover–fungi, and corn stover–yeast; Line 2—miscanthus–bacteria, miscanthus–fungi, and miscanthus–yeast.)

The fungi-based pathway had the highest external electrical energy requirements due to the additional electrical power requirements of the air compressor and filtration unit required for fermentation (Table 3). The electricity requirements were higher for the fermentation and stillage processes used to operate large bioreactors, which required electricity to maintain temperature and mix the slurries. The yeast-based pathway was associated with the lowest amount of electricity consumed, as the electricity required for the reactors used for gypsum formation and the filtering unit for gypsum removal were not necessary. The steam and cooling water requirements were the lowest for the yeast-based pathway as a result of lower overall material flow during distillation. The recovery and purification units required the highest quantities of steam and cooling water for the distillation process conducted to recover the lactic acid. Lactic acid production using miscanthus had relatively lower utilities requirements compared to the corn-stover-based lactic acid production system. The lower feedstock requirements resulted in lower demand for water and other chemicals for different processes during lactic acid production, thereby reducing the demand for utilities.

The yeast-based pathway had the lowest labor requirements; this was mainly due to a reduction in the labor hours required to operate the reactors used for lactic acid neutralization and recovery (Table 4). The labor requirements for the fungi-based pathway were higher due to additional labor hours required to operate and oversee the air compressor and filtration units during the fermentation process. For all pathways, the labor requirements for fermentation and purification and recovery units were higher, as these processes required more equipment and frequent monitoring. The labor requirements for the production facility based on miscanthus were lower than those for corn stover due to a lower level of biomass handling.

Pathways	Baci	Bacteria Fungi		ngi	Yeast		
Feedstock	Corn Stover	Miscanthus	Corn Stover	Miscanthus	Corn Stover	Miscanthus	
Electricity (kWh/h)							
Feedstock preparation	514	454	504	450	535	466	
Pretreatment	1140	1013	1119	1005	1187	1041	
Fermentation	3900	3694	4780	4315	3881	3390	
Recovery and purification	527	484	520	479	304	279	
Stillage utilization	3082	2551	2903	2269	2866	2846	
Total amount of electricity used	9162	8196	9828	8519	8773	8023	
Total electricity produced	6360	4870	6040	4750	7200	5420	
Steam (t/h)							
Feedstock preparation	-	-	-	-	-	-	
Pretreatment	18.7	16.6	18.4	16.4	19.5	17.0	
Fermentation	-	-	-	-	-	-	
Recovery and	195.5	179.5	193.2	178.2	182.3	164.0	
Stillage utilization	1.3	1.1	1.2	0.9	1.0	1.3	
Total amount of steam used	215.5	197.1	212.8	195.4	202.8	182.2	
Total steam produced	33.7	26.3	32.1	25.7	37.9	29.0	
Cooling water (t/h)							
Feedstock preparation	-	-	-	-	-	-	
Pretreatment	448.9	398.9	440.9	395.7	467.4	409.9	
Fermentation	402.8	373.3	438.2	394.6	370.6	324.3	
Recovery and purification	3373.0	3280.5	3359.8	3272.0	2826.5	2717.6	
Stillage utilization	262.0	216.7	246.7	192.8	243.6	241.9	
Total amount of cooling water used	4486.6	4269.5	4485.5	4255.1	3908.0	3693.6	

Table 3. Utilities requirements for the selected lactic acid production pathways and feedstocks.

#### 3.3. *Capital Costs*

The capital investments for lactic acid production facilities with a production capacity of 100,000 t/year were in the range of USD 241–268 million, USD 244–266 million, and USD 236–257 million for the bacteria-, fungi-, and yeast-based pathways, respectively

(Figure 3). The pathway using yeast for fermentation had the lowest capital investment as lactic acid neutralization and recovery processes were not required in this pathway, thus reducing the costs associated with the purchase and installation of the equipment needed for these processes. The fungi-based pathway required higher capital investment since the fermentation process employing fungi required a continuous supply of air to be effective. This required additional air filters and compressor units, thus increasing the capital investment. The cost of different types of equipment and engineering operations were based on the shares of different components, as discussed in Table 2. For the facility using corn stover as a feedstock and an LAB-based fermentation pathway, the pretreatment stage contributed the most to the total equipment costs as it required acid-resistant reactors to hydrolyze the corn stover. The impact of the pretreatment process on the overall costs was also highlighted by a previous study [80]. The working capital for the yeast-based pathway was lower due to a reduction in the use of utilities, chemicals, and labor for the additional steps for lactic acid neutralization and recovery (Figure 3). For all three fermentation pathways, the miscanthus-based facility had slightly lower feedstock and other resources requirements; thus, it had lower equipment sizes and overall capital costs (USD 236–244 million) than the corn-stover-based facility (USD 257–268 million). The startup capital followed a trend similar to that of the direct fixed capital costs, as it was estimated as 5% of the direct fixed capital.

Table 4. Labor requirements (h/year) for different lactic acid production pathways and feedstocks.

Pathways	Pathways Bacteria			ngi	Yeast	
Feedstock	Corn Stover	Miscanthus	Corn Stover Miscanthus		Corn Stover	Miscanthus
Feedstock preparation	2489	2489	2489	2489	2489	2489
Pretreatment	22,383	22,383	22,383	22,383	22,383	22,383
Fermentation	23,166	23,422	24,763	24,595	23,488	23,278
Recovery and purification	26,219	26,219	26,219	26,219	22,629	22,629
Stillage utilization	7605	7605	7605	7605	7605	7605
Total labor	81,862	82,118	83,459	83,291	78,594	78,384



**Figure 3.** Capital investments for the facility producing 100,000 t/year of lactic acid from different fermentation pathways using corn stover and miscanthus feedstocks (**a**); share of different components of direct fixed capital for the facility using corn stover feedstock and the LAB-based pathway (**b**); contribution of different conversion steps to the facility costs (**c**).

## 3.4. Lactic Acid Production Costs

For a lactic acid production facility with an annual production capacity of 100,000 t/year, the per metric ton costs of lactic acid production were in the range of USD 1136–1281, USD 1137–1279, and USD 894–1033 for the bacteria-, fungi-, and yeastbased pathways, respectively (Figure 4). The production costs in this study were similar to the lactic acid production cost of USD 1.25/kg using ultra-filtered whey presented in a prior study [81]. The lactic acid production costs for the facility using miscanthus were lower than those for the facility using corn stover due to the higher lactic acid yields from miscanthus. Raw materials and facility-dependent costs were the major contributors to the total production costs of lactic acid. Raw materials accounted for 27–34% of the total lactic acid production costs for different production pathways using corn stover feedstock and 22–32% using miscanthus feedstock. The yeast-based pathway did not require the use of lime to neutralize the lactic acid in the fermentation broth nor sulfuric acid and water for the recovery of lactic acid from the neutralized mixture; thus, it had the lowest raw materials cost. The facility-dependent costs amounted to 37-44% and 37-47% of the total lactic acid production costs for different production pathways using corn stover and miscanthus feedstocks, respectively. The fungi-based pathway had higher facility-dependent costs due to the addition of air filter and compressor units required for fermentation. The yeast-based pathway did not require the use of reactors to produce and remove gypsum and thus had the lowest facility-dependent costs. Labor costs accounted for 4-5% of the lactic acid production cost and had a trend similar to that of the facility-dependent costs. Utilities contributed between 21 and 26% of the total lactic acid production costs using both feedstocks. The utilities costs were the lowest for the yeast-based pathway due to the elimination of the processes of producing and removing gypsum, which reduced the utilities requirements. The bacteria- and fungi-based pathways had higher waste treatment costs than the yeast-based pathway due to the additional gypsum removal process. Several studies have evaluated the technoeconomic feasibility of lactic acid production from different lignocellulosic feedstocks such as corn stover, sugarcane bagasse, leaves, and macroalgae [26,82,83]. Similar results were also reported in multiple studies evaluating lactic acid production from lignocellulosic feedstocks [26,83].



**Figure 4.** Lactic acid production costs for the facility producing 100,000 t/year of lactic acid from corn stover and miscanthus feedstocks and different pathways using bacteria, fungi, and yeast for fermentation.

The feedstock preparation steps contributed to 17–23% of the corn-stover- and 14–18% of the miscanthus-based lactic acid production costs. Purification and recovery had the highest shares of lactic acid production costs, amounting to 25–30% for the corn-stover-based and 27–32% for the miscanthus-based lactic acid production costs. Higher purification and recovery costs were due to higher costs for equipment and utilities required for distillation. Acid and enzymatic hydrolyses steps amounted to 21–28% of the total lactic acid production costs, which were mainly due to reactor, utilities, and chemical

costs. The fermentation step amounted to 10–20% of the corn-stover-based and 11–22% of the miscanthus-based lactic acid production costs. The costs of the fermentation reactor, utilities, and nutrient contributed to the fermentation costs. Similarly, stillage utilization amounted to 11–12% of the total costs for lactic acid production from corn stover and miscanthus.

## 3.5. Financial Analysis

The unit lactic acid production costs (USD/t) for the bacteria-, fungi-, and yeast-based pathways were between USD 1136–1281, USD 1137–1279, and USD 894–1033, respectively. Additional revenues from selling the electricity produced while producing a ton of lactic acid were between USD 54–70, USD 53–67, and USD 60–79 for the bacteria-, fungi-, and yeast-based pathways, respectively (Table 5). The yeast-based pathway had lower conversion rates for sugar fermentation; thus, the remaining organic matter was utilized in anaerobic digestion, which increased biogas, steam, and electricity production, resulting in higher revenues from selling electricity. Considering the production costs, revenues, and 10% IRR (after tax), the minimum selling prices per ton of lactic acid were USD 1243–1390 for the bacteria-based pathway, USD 1250–1392 for the fungi-based pathway, and USD 993–1123 for the yeast-based pathway, respectively. Based on the financial analysis of all three pathways (Table 5), the pathway for producing lactic acid using both corn stover and miscanthus feedstocks. The lactic acid produced using miscanthus had a lower minimum selling price than that when using corn stover as feedstock.

## 3.6. Sensitivity Analysis

The sensitivity analysis indicated that the conversion rates of glucose and xylose c to lactic acid, feedstock costs and moisture content, production plant capacities, annual operation hours, acid hydrolysis reactor costs, and enzyme costs were the most sensitive parameters affecting the lactic acid production costs for all three pathways (Figure 5). Lower conversion rates of sugar to lactic acid resulted in lower lactic acid production and thus increased the per ton lactic acid production costs, and vice versa. The lactic acid production cost was also highly sensitive to the feedstock costs. An increase in feedstock costs directly increased the costs of input materials and, ultimately, the unit cost of lactic acid production. Changes in the feedstock moisture content affected the wet feedstock quantity required to meet the lactic acid production demand. Using feedstock with lower moisture content reduced the quantity of wet feedstock, thereby reducing the size of equipment required in the production facility. Higher production plant size or biorefinery capacity increased the scale of operation and thus lowered the unit lactic acid production costs due to an economy of scale. An increase in annual operation hours subsequently increased the annual productivity of the biorefinery and thus increased the quantity of lactic acid produced with the same capital investment. Thus, increasing annual operation hours lowered the unit production cost of lactic acid. A higher cost of reactors increased the total costs of equipment and capital investment and, ultimately, the lactic acid production cost. In addition, the lactic acid production costs for the bacteria- and fungi-based pathways were highly sensitive to gypsum use and lime costs, as a substantial quantity of lime was required to maintain the pH conditions during fermentation. Similarly, a substantial quantity of gypsum was produced while recovering the lactic acid in the purification and recovery section. This resulted in sensitivity of lactic acid production costs based on the approach used to manage the gypsum, thus affecting the lactic acid production costs. The sensitivity for parameters is different for miscanthus and corn stover due to the differences in cellulose and xylose content.



**Figure 5.** Sensitivity analyses for lactic acid production via different fermentation pathways and feedstocks: (a) lactic acid bacteria–corn stover, (b) lactic acid bacteria–miscanthus, (c) fungi–corn stover, (d) fungi–miscanthus, (e) yeast–corn stover, and (f) yeast–miscanthus.

Pathways	Bacteria		Fı	ıngi	Yeast	
Feedstock	Corn Stover	Miscanthus	Corn Stover	Miscanthus	Corn Stover	Miscanthus
Unit production cost (USD/t)	1281	1136	1279	1137	1033	894
Revenues (USD/t)	69	53.7	66.0	52.4	78.4	60.0
Minimum selling price (USD/t)	1390	1243	1392	1250	1123	993
Gross margin (%)	12.2	12.4	12.3	12.4	14.0	15.1
Return on investment (%)	13.2	13.2	13.2	13.2	13.2	13.3
Payback period (years)	7.6	7.6	7.6	7.6	7.6	7.6
Net present value (million USD)	78.9	70.9	80.0	73.7	75.3	71.6

**Table 5.** Financial analysis of different pathways to produce lactic acid using corn stover and miscanthus feedstocks.

# 4. Conclusions

The cost of the lactic acid produced from the lignocellulosic feedstocks, miscanthus and corn stover, varied between USD 894 and USD 1281 per ton, depending on the three different fermentation pathways using LAB, fungi, and yeast. Lactic acid production using genetically engineered yeast strains can be fermented under lower pH conditions and thus does not require the simultaneous neutralization of lactic acid during fermentation and its recovery at later stages. This reduced the chemical, equipment, and utilities requirements for lactic acid production using yeast compared to that for LAB and fungi and, consequently, resulted in the lowest production costs. Miscanthus had higher sugar yields and lower feedstock production costs and thus lower lactic acid production costs. Raw materials and facilities contributed to more than 70% of the total cost of lactic acid production. For all three pathways, the sugar-into-lactic-acid conversion rates, feedstock costs, production plant capacity, annual operation hours, and acid hydrolysis reactor costs were the most sensitive to lactic acid production costs. The gypsum management approach also highly influenced the lactic acid production costs and the minimum selling price for the pathways using LAB and fungi. This study indicated the possibility of producing lactic acid using different lactic acid fermentation pathways and lignocellulosic feedstocks. Lactic acid production costs can be further reduced by improving process efficiencies, increasing sugar conversion rates, and lowering equipment and chemical costs. The identification of technically feasible and low-cost lactic acid production methods using biobased resources could contribute to a sustainable bioeconomy in the future.

**Supplementary Materials:** The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/fermentation9070641/s1, Table S1: Technical data related to different equipment used for lactic acid production [50,62,84,85].

**Author Contributions:** Conceptualization, A.M. and A.S.; data collection and analysis, A.M.; writing—original draft preparation, A.M.; review and editing, A.M. and A.S.; supervision, A.S.; funding acquisition, A.S. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by United States Department of Agriculture NIFA (Award no. 2019-67019-29310) and United States Department of Agriculture Hatch project (Project no. 1021119). The opinions expressed herein are those of the author(s) and do not necessarily reflect the views of the sponsoring agencies.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Supplementary data is provided with this submission.

**Conflicts of Interest:** The authors declare no conflict of interest.

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