



Article Enhanced Breaking of Lignin and Mesopore Formation in Zinc Chloride Assisted Hydrothermal Carbonization of Waste Biomasses

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Abstract: Hydrochars from hydrothermal carbonization of different biowaste materials (dried dandelion, sawdust, coconut shell powder) formed in the presence of aqueous salt solutions were compared to those obtained by the common method in pure water. Hydrochars with increased carbon contents, pore volume and surface areas were specifically obtained from coconut shell powder in the presence of zinc chloride. Compositional and structural changes within the hydrochar products caused by the process conditions and/or the additive were characterized by solid state ¹³C NMR spectroscopy, proving that cellulose and, in particular, lignin units in the biomass are more easily attacked in the presence of the salt. Under saline conditions, a distinct particle break-up led to the creation of mesoporosity, as observable from hysteresis loops in nitrogen adsorption isotherms, which were indicative of the presence of pores with diameters of about 3 to 10 nm. The obtained hydrochars were still rich in functional groups which, together with the mesoporosity, indicates the compounds have a high potential for pollutant removal. This was documented by adsorption capacities for the methylene blue and methyl orange dyes, which exceeded the values obtained for other hydrochar-based adsorbers. A subsequent physical activation of the mesoporous hydrochars in steam at different temperatures and times resulted in a further drastic increase in the surface areas, of up to about 750 m²/g; however, this increase is mainly due to micropore formation coupled with a loss of surface functionality. Consequently, the adsorption capacity for the quite large dyes does not provide any further benefit, but the uptake of smaller gas molecules is favored.

Keywords: hydrothermal carbonization; waste biomass; mesoporosity; salt additive; dye adsorption; steam activation

1. Introduction

Hydrothermal carbonization (HTC) provides a green and sustainable process, allowing the production of a variety of carbonaceous materials starting from cheap natural precursors, such as plant biomass, manure or digestates, under comparatively mild hydrothermal conditions [1]. In recent years, comparable processes such as the conversion of various biomasses by means of vapothermal carbonization (VTC), which uses a water–vapor phase instead of a liquid phase, or wet torrefaction, have also gained in importance. The latter process is particularly applicable because smaller amounts of highly contaminated process water are produced than in HTC [2–4]. Further processes to convert biomasses to biooils or high-energy density fuels are torrefaction and valorization [5,6].

The obtained hydrochars from HTC have received increasing interest in various relevant fields of the chemical industry, such as for catalysis, water purification, energy storage and CO_2 sequestration [7,8]. However, due to their low degree of porosity and small internal surface areas of less than $10 \text{ m}^2/\text{g}$ [9], the hydrochars mainly serve as precursors instead of competitive substitutes to the energy expensive activated carbons received by pyrolysis at high temperatures [10–13]. In principle, chars obtained from HTC exhibit great potential as substrates for the synthesis of highly porous carbon materials due to their preferable characteristics, such as their oxygenated functional groups which can catalyze



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Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). subsequent activation reactions [14]. The creation of carbons with high surface porosity by HTC was achieved from pure glucose in the presence of salt/water mixtures. The hydrophilic ions of the commonly available salts are supposed to alter the partial pressure of the water being in contact with the carbon phase [15]. With a eutectic mixture of lithium chloride and zinc chloride in the presence of water, a high surface area for carbons from glucose was obtained (673 m^2/g). However, due to their extreme structural complexity and diversity, natural raw lignocellulosic biomasses have not received much attention as substrates for HTC conversion to porous hydrochars. One of the available studies shows a slight increase in surface area to $69.45 \text{ m}^2/\text{g}$ if the hydrothermal carbonization of corncob was carried out under saline conditions [16]. In general, real biomass hydrochars have much smaller specific surface areas than chars obtained from pyrolysis [4,17,18]. A subsequent activation can act as a further carbonization step increasing the aromatic content, the porosity and the stability of the hydrochars [13]. Steam is used in physical activation tests [19-21]; for chemical activation KOH, NaOH or H₃PO₄ are used. For example, KOH acts as a catalyst and oxidizing agent, and the occurring reactions can be understood as in situ alkali-catalyzed gasification; specific surface areas up to $2.000 \text{ m}^2/\text{g}$ were obtained, which can mainly be attributed to the formation of small micropores with diameters less than 2 nm [22–24].

The main goal of our research, however, was to study the impact of the presence of ZnCl₂ salt on the formation of mesoporous hydrochars with pores having a size of about 3–10 nm resulting directly from the HTC of complex biomass substrates. For this, three easily available biowaste products were chosen, i.e., dried dandelion, sawdust and coconut shell powder. Elemental composition, morphology, functionality and surface areas of the obtained hydrochars were studied by scanning electron microscopy (SEM), infrared spectroscopy (FTIR), and nitrogen sorption, respectively. Dye adsorption studies on selected hydrochars were carried out to demonstrate the advantages of their porosity and surface functionality for a later application as an adsorbent in pollutant removal. It was shown before that hydrochars can have comparable adsorption capacities as oil-based activated carbons, indicating a potential application of low-cost chars obtained by HTC of waste biomasses. Alatalo et al. described the use of hydrochars from fructose conversion by HTC in the presence of a hypersaline salt mixture (LiCl/ZnCl₂) and found fast adsorption controlled by intraparticle diffusion [25]. By simple washing with ethanol, the dye could be desorbed, and the char could be recycled for further dye removal loops.

In a subsequent process we further treated the hydrochars by means of a physical activation with steam at different activation temperatures to elucidate the potential and drawbacks of this treatment.

2. Experimental

The HTC was performed at relatively mild conditions, i.e., 180–220 °C for reaction times of 4–20 h, either in water (further called "normal HTC") or in a salt–water mixture containing zinc chloride (further called "salt HTC"). After the HTC, the obtained hydrochars were collected by vacuum filtration, washed with water to remove the residual salt, dried at 105 °C overnight, ground and size-selected with a sieve having a mesh size of 0.63 mm.

2.1. Materials

Coconut shell powder (Cn) was kindly provided by Mahlwerk Neubauer-Friedrich Geffers GmbH as Coconut 0.2–0.4 and was used as received. Dried dandelion (Dd) was obtained from the online shop "kaninchenladen.de" and shredded in a Kenwood CH580 chopper. Sawdust (Sd) was picked up from the mechanical workshop section of the Carl von Ossietzky University Oldenburg, Germany; it represents a random mixture of wood being processed in the workshop at that time and was used as received. Lithium chloride (p.a.) was obtained from VWR and zinc chloride (\geq 98%) was acquired from Sigma Aldrich.

The dyes were received from AplliChem (methylene blue) and Alfa Aesar (methyl orange). All chemicals were used without further purification.

2.2. Hydrothermal Carbonization

The biomass substrates (9 g) were mixed either with 180 mL of pure water (Cn-HTC) or with 22.5 g freshly ground ZnCl₂ salt (Cn-HTC Zn) dissolved in the same volume of water. Each mixture was filled into a 200 mL PTFE inlet and placed into a stainless-steel autoclave, which was put in an oven at 180 °C for 12 h. For some HTC experiments the temperature and time were varied (200 or 220 °C and 4–20 h, respectively). After cooling to room temperature, the solid product was ground by hand in a porcelain mortar and stirred in 1 L of water overnight. Afterwards, the samples were collected by vacuum filtration, Soxhlet extracted with water and ethanol to remove any residual salts, and dried at 105 °C overnight. The obtained hydrochars were ground once more and passed through a sieve with a mesh size of 0.63 mm. Thus, all agglomerates were crushed to particles smaller than 0.63 mm.

2.3. Steam Activation

All the steam activated chars were previously processed by HTC. For the steam activation, 2–3 g of each hydrochar obtained by HTC was loaded into a ceramic crucible and placed into a glass inlet of a tube furnace and heated. Upon reaching the desired temperature, the steam originating from a simple heating of water in a connected 500 mL Erlenmeyer flask was allowed to flow through the glass inlet for 1 h. For the coconut substrate, two modifications of the activation were conducted: Activation at (a) 500 °C at different times of steam flow (t = 10, 30, 60 min, e.g., Cn-Act10min) or (b) for 60 min at different temperatures (T = 200, 350, 500, 650 °C, e.g., Cn-Act200C). After cooling, the thus-activated hydrochars were analyzed without further treatment except if otherwise noted under the specific characterization techniques.

2.4. Characterization Techniques

A EuroEA Elemental Analyser (HEKAtech GmbH) was used to determine the carbon, hydrogen and nitrogen contents of the hydrochars. Inorganic elemental concentrations were measured after incinerating 0.2 g of each sample in a muffle furnace at 550 °C for 7 h. Fourier transform infrared (FTIR) spectroscopy was conducted using a Bruker Tensor FTIR spectrometer in the range from 500 to 4000 cm⁻¹. Scanning electron microscopy (SEM) was used to determine the surface morphology of the samples with a Hitachi S3200N microscope being equipped with an INCA X-act silicon drift detector from Oxford Instruments for EDX measurements. Nitrogen sorption measurements were accomplished with N₂ at 77 K on a Micromeritics TRISTAR II or a Micromeritics ASAP 2020. Prior to the measurements, the samples were degassed under vacuum first at 80 °C for 1 h and subsequently at 150 °C overnight. The specific surface areas were calculated according to the Brunauer–Emmett–Teller (BET) formalism using adsorption data in the relative pressure range $p/p_0 < 0.3$ in further agreement with criteria proposed by Rouquerol et al. [26].

For the solid-state NMR measurements in this work, a Bruker Avance III HD spectrometer connected to a AscendTM 500 cryomagnet (11.75 T) was used for the characterization of different hydrochars. The corresponding Larmor frequencies were 500.45 and 125.84 MHz for ¹H and ¹³C, respectively. The rotating frequency used was 15 kHz; a standard 4.0 mm CP/MAS probe head (Bruker WVT-type) was employed. One-dimensional spectra were recorded with the *hahnecho* pulse sequence and with cross polarization technique (*CP* pulse sequence) to enhance ¹³C spectra. Therefore, the relaxation times of the protons were determined by classical *saturation recovery* experiments. Pulse lengths were 3.13 and 4.20 µs at power levels of 107.05 and 94.37 W for ¹H and ¹³C, respectively. Si(SiMe₃)₄ served as primary reference to determine the chemical shifts. The controlled temperature of the bearing gas (300.0 K) and a calibration curve of the reference material led to an estimated sample temperature of 338 ± 2 K. Up to 12,000 scans were accumulated using relaxation delays of 3 to 8 s, resulting in acquisition times between 2.5 h and 1 day for each spectrum. Peaks and peak areas were assigned by comparing constituents found in biomass building blocks (cellulose and lignin) with those found in the structurally complex biomass. In addition, they were compared to data from the literature [27–29].

2.5. Dye Adsorption Studies

The adsorption experiments with methylene blue (MB) and methyl orange (MO) were performed with selected hydrochars using a modified procedure based on a publication by Islam et al. [30]. For this, 40 mL of methylene blue or methyl orange solution (initial concentration of 100 mg/L) were added to a small amount (50 mg) of char in a 50 mL plastic tube. The mixture was shaken at 250 rpm for different time periods (up to 30 h) at room temperature with a Dual-Action Shaker KL 2 (Edmund Buehler GmbH). After the desired time, the residual concentration in the solution was analyzed using a Varian Cary 4000 spectrophotometer at 664 nm after filtering the adsorbent with syringe filters (0.2 μ m, Sarstedt GmbH) to make it carbon-free and diluting it with water by a factor of 20. The amount of dye removed from the solution is reported as *q_t* in mg/g; it was calculated according to Equation (1):

$$q_t = \frac{(C_0 - C_t)V}{m} \tag{1}$$

where C_0 and C_t are the initial dye concentration and the concentration after the desired time in mg/L, respectively, *V* is the volume of used dye solution (L), and *m* is the weight of the adsorbent (g). To reduce the error of the adsorption data, each sample was tested three times, and the average is reported. If the adsorption time was chosen long enough to reach the equilibrium state, C_t changes to C_e and the corresponding adsorption capacity is given as q_e .

3. Results and Discussion

3.1. Elemental Composition of the Hydrochars

Comparison of the carbon content as obtained by the elemental analysis of the raw materials and hydrochars (Table 1) shows that the HTC process leads to an increase in the carbon content by 4 to 11 wt.%, depending on the raw material used. From these, sawdust shows the lowest gain, indicating greater stability as the increase is normally due to cleavage and cracking of weak oxygen and hydrogen bonds in O and H-containing compounds which are preferentially lost due to their higher solubility [31]. If the salt mixture was present, the carbon content increased more, i.e., by 13 to 18 wt.% with respect to the original biomass, reaching 66 wt.% for Cn-HTC Zn.

By steam activation of the HTC hydrochars at 500 °C, the carbon content increased further up to 73 and 77 wt.% for chars from sawdust and coconut shells, respectively. In the case of the dried dandelion, the carbon content changed only slightly, i.e., by 0.1 to 2 wt.%. Coupled with a low yield, this indicates a high loss of carbon into the water or steam phase. Examples in Figure 1 for the hydrochars from coconut shells show the increase in the carbon content is, for all samples, visually supported by a change in color. As expected, the color of the hydrochars from normal HTC is dark brown, indicating that the typical peator lignite-like material is formed. The hydrochar obtained in the presence of the ZnCl₂, however, is much darker, nearly black, indicating the formation of a char which is mineral coal-like, with a much higher degree of polymerization and aromatic structures [15,31].

For coconut-based hydrochars, the variation in temperature has a more significant influence on the carbon content than the studied times, because only a small increase from 74–76 wt.% after 10 min, to around 77 wt.% after treatment for 60 min, takes place. With increasing temperature during steam activation, the carbon content gradually increases from 59 wt.% at 200 °C to 87 wt.% at 650 °C for HTC coal formed from coconut without salt. The contents of hydrogen and nitrogen were indirectly related to the process para-meters for all feedstocks, indicating mostly a loss in hydrogen from the dehydration reactions taking place in the hydrothermal carbonization.

Substrate	Sample	C [wt.%]	N [wt.%]	O [wt.%]	H [wt.%]	Ash [wt.%]
Dandelion	Dd (raw material)	37.5	2.16	44.3	5.5	10.76
	Dd-HTC	49.7	2.3	32.7	6.0	9.3
	Dd-HTC Zn	52.0	1.4	29.1	4.1	13.6
	Dd-Act500C	49.8	1.9	19.8	0.5	28.0
	Dd-Act500C Zn	53.6	1.6	17.4	1.5	25.9
Sawdust	Sd (raw material)	46.3	0.6	46.9	6.1	0.0
	Sd-HTC	50.0	0.4	43.8	5.9	0.0
	Sd-HTC Zn	60.3	0.4	35.6	3.7	0.0
	Sd-Act500C	66.1	1.8	30.4	0.9	0.9
	Sd-Act500C Zn	73.0	0.6	22.0	1.7	2.8
Coconut	Cn (raw material)	49.1	0.1	44.7	6.2	0.4
	Cn-HTC	59.5	0.1	35.0	5.4	0.1
	Cn-HTC Zn	66.2	0.1	29.6	4.0	0.1
	Cn-Act650C	86.9	0.2	11.8	1.2	0.00
	Cn-Act650C Zn	82.9	0.1	12.8	1.1	3.1
	Cn-Act500C	77.2	0.2	20.9	1.7	0.0
	Cn-Act500C Zn	77.0	0.1	18.8	2.0	2.2
	Cn-Act350C	69.2	0.3	27.4	3.1	0.0
	Cn-Act350C Zn	69.2	0.1	26.1	3.0	1.7
	Cn-Act200C	59.3	0.2	35.3	5.2	0.0
	Cn-Act200C Zn	65.1	0.8	31.1	3.2	0.5
	Cn-Act30min	76.8	0.2	21.4	1.6	0.0
	Cn-Act30min Zn	75.3	0.0	20.8	1.9	1.9
	Cn-Act10min	74.4	0.3	23.1	2.3	0.0
	Cn-Act10min Zn	75.9	0.1	19.8	2.2	2.1

Table 1. Elemental compositions of the raw materials, their hydrochars and activated carbons produced from dried dandelion (Dd), sawdust (Sd) and coconut shell powder (Cn). For all masses, errors up to 0.5 wt.% must be expected.



Figure 1. Digital photos of the original coconut shell powder (**a**) and hydrochars obtained by the normal (**b**) or the salt-HTC route (**c**).

The loss of hydrogen and oxygen in the different biomasses, i.e., coconut shell powder, dried dandelion leaves and sawdust, due to carbonization and steam activation is shown in a typical van Krevelen diagram in Figure 2. The van Krevelen diagram clearly shows that by HTC in the presence of salt, lower O/C and especially H/C, are obtained than by HTC with water. After steam activation, the obtained levels are closer to each other and the compositions are comparable to that of anthracite; that is, the black coal has the highest degree of condensation and lowest number of functional groups.



Figure 2. Van Krevelen diagram documenting the loss of oxygen and hydrogen from the biomasses (squares) during HTC with water (open circles) or HTC with salts (full circles) and subsequent steam activation (triangles). The representative areas for biomass, peat, lignite and black coal (anthracite) are collected from literature [32–34].

In the hydrochars received from dried dandelion leaves, the carbon content is limited to a level of around 50%; nevertheless, the H/C and O/C ratios are low, confirming that the ash content is high in hydrochars obtained from this substrate (up to 28 wt.%), due to the high amount already present in the raw material (10.6 wt.%). The ash contents of the sawdust and coconut samples are small, and the highest amounts (\leq 3 wt.%) of inorganic compounds remained after activation. Higher contents were observed if the salt mixture was used in the preceding HTC reactions, indicating that some remaining built-in ZnCl₂ was not washed out completely.

It is apparent from the yields that the sawdust showed the highest stability under HTC conditions, and also possessed the smallest amount of easily soluble compounds lost by leaching. Instead of complete destruction, the original carbon network seems to be preserved in wide parts and pores are created only in the regions with more soluble material. The stable network is very likely rich in lignin. Thus, the presence of a more stable morphology is a good prerequisite for the introduction of porosity into these structures. The hydrochars from dried dandelion could only be obtained with yields of 30 and 43%, underlining the low stability and high inorganic content of this substrate.

3.2. Surface Composition of the Compounds

FTIR spectra of the coconut shell powder, the related hydrochars from HTC with water or with salt, and the activated chars are shown in Figure 3. The samples from coconut shell powder were chosen as representative examples because all three tested biomasses and obtained hydrochars showed similar trends regarding their surface functionality. Signals of several functional groups in the coconut precursor, namely, hydroxyl groups (3500–3000 cm⁻¹), alkyl groups (3000–2800 cm⁻¹), ether groups (1300–1000 cm⁻¹), C-H groups in aldehydes (2900–2700 cm⁻¹), carbonyl groups in aldehydes and ketones (1850–1650 cm⁻¹) and carboxylic acid groups (1820–1750 cm⁻¹), in addition to aromatic C=C stretching (~1600 cm⁻¹) vibrations (for a more detailed description of IR bands see, e.g., [28,35]) become sharper after hydrothermal carbonization in pure water (Cn-HTC), indicating a relatively increasing amount of these groups in the samples. However, in the

presence of an aqueous salt mixture (Cn-HTC Zn), an opposite trend is observed with declined corresponding peaks, which are specifically seen by the flattening of the hydroxyl and ether bands at around 3300 and 1300 to 1000 cm^{-1} , respectively. This sample features only two prominent bands at 1700 and 1600 cm^{-1} , and a wider band at 1200 cm^{-1} becoming sharper in the course of the process, suggesting the presence of groups with mainly carbonyl and aromatic functionality. From Figure 2 it is also obvious that a thermal treatment with steam after HTC is accompanied with a significant loss of surface functionality, which is again stronger for the hydrochars obtained under saline conditions (Cn-Act500C Zn). This loss can be reduced by using lower activation temperatures; there is a linear correlation to the band flattening and finally complete disappearance with increasing temperatures. Many bands are still prominent after activation at 200 °C but, at 650 °C, with the exception of the double peak at around 2400 cm⁻¹ corresponding to gaseous CO₂, no peak remains. The time of steam activation has only a limited influence on the functional groups.



Figure 3. FTIR spectra of coconut shell powder and hydrochars obtained by HTC and subsequent steam activation.

3.3. Morphology of the Hydrochars

The SEM images (Figure 4a,b) reveal that the normal hydrochar morphology resembles the one seen for the substrate, i.e., heterogeneous structures with layers of rather smooth surfaces. The salt hydrochar, in contrast, shows a much rougher surface; the previously smooth surface is spotted with many holes in the still-maintained macroscopic structures of the biomass, indicating the creation of meso- and macroporosity in these materials (Figure 4c). This observation is confirmed by TEM images (Figure 4d,e), in which the hydrochar from normal HTC shows a vague structure with no identifiable primary building units. The hydrochars from salt HTC, however, are revealed to be built of small primary particles in the size range of 20–50 nm.

From our study it is obvious that all resulting hydrochars consist of very heterogeneous, irregular-shaped particles of different sizes together with larger, stacked structures. Smaller particles presumably originate from the breaking of the large frameworks, which represent the highest content in the initial coconut shell powder.

By HTC in aqueous salt mixture, the number of smaller fractions increases. Smaller particles are observable around and on top of macroscopic structures. This supports the idea that the present salt mixture either catalyzes dehydration reactions [36,37] or increases the solubility of parts of the biomass, i.e., the cellulose and hemicellulose [38]. This results in a partial breaking of the framework and the formation of smaller carbonized structures. However, it is also obvious from both images that the samples still possess large structures



resulting from components of the original biomass resistant to hydrothermal treatment such as cellulose and lignin [27].

Figure 4. Scanning electron micrographs (SEM, upper line) and transmission electron micrographs (TEM, lower line) of the original coconut shell powder (**a**) and hydrochars obtained by the normal (**b**,**d**) or the salt-HTC route (**c**,**e**).

After additional steam activation, the derived chars show a further increase in particle break-up. Moreover, with a progressing degree of carbonization, the roughness of the particle surfaces also increases. This shows that the steam breaks-up larger structures by oxidizing the different carbon compounds, while creating roughness and porosity on the surface and inside the particles.

Small traces of salt left after washing, as implied by the higher ash content in the salt-HTC samples (Table 1), may contribute with catalytic effects in the activation [16,39]. Energy-dispersive X-ray spectroscopy (EDX) indicates some remains of Cl^- and Zn^{2+} in traces of 0.32 and 0.34 wt.%, respectively.

Even after salt HTC and treatment with steam at high temperatures, parts of the natural macrostructure stay intact. Nevertheless, liquefaction and extraction of organic materials such as hemicellulose and non-structured amorphous cellulose leading to mass losses >50 wt.% [30] result in structural changes at the nanometer scale [13].

3.4. Mechanistics

The results indicate that, under the hydrothermal conditions of normal HTC (the autogenous pressure obtained is about 15 bar), the substrate is not significantly attacked at a temperature of 180 °C and stays mostly intact. In the salt HTC, the zinc chlorideenriched steam attacks the surface of the biomass significantly more strongly than the pure water steam, creating surface roughness and forming holes by degradation of the biomass compounds. However, it is worth mentioning that, because the main macrostructure stayed intact, parts of the framework seem to be resistant to the hydrothermal treatment at the chosen, rather soft HTC conditions, even in the presence of the salt. Biomass is mainly built by different ratios of the three components cellulose, hemicellulose and lignin, which all exhibit different stabilities upon hydrothermal carbonization. Lignin is usually claimed to be resistant to the HTC reactions up to 300 °C [40,41]. Thus, it is possible that the preserved macrostructures consist mostly of the lignin fraction of the biomass, and the holes and roughness were created by hydration, dissolution, decarboxylation and other reactions of the cellulose and/or hemicellulose, leading to a high extent of soluble compounds dissolved in the liquid HTC water phase. The remaining fragments, which are not soluble in water, may form some spherical-shaped carbon particles to minimize the contact area with their surroundings, which can settle on the char resulting from the skeleton frame of the original biomass, and thus create additional roughness [27].

More detailed information about the composition and structure changes from coconut biomass to hydrochars can be derived from ¹³C solid state CP/MAS NMR spectroscopy data. Figure 5 shows the spectra for different hydrochars obtained from coconut shell powder. As expected, signals of cellulose and lignin are found in the spectra of the hydrochars, as shown by use of the pure cellulose and pure lignin as reference materials (Figure 6). The corresponding signals are highlighted by colored boxes (orange: cellulose; grey: lignin) in Figure 5. Typical chemical shifts for cellulose are found between 55 and 105 ppm (Figure 5, orange box and Figure 6); the signals are mainly from aliphatic carbons linked to alcohol groups; the peak signature corresponds well to the literature data [42]. In contrast, for cellulose treated by normal HTC at 180 °C, almost no change in the ¹³C NMR spectrum is observable; nearly all original peaks of the cellulose are completely vanished after salt HTC, and the emerging new peaks indicate the formation of polymerization products (Figure 6 left). The NMR signature obtained for lignin shows two prominent peaks at 55 and 146 ppm, which can be attributed to lignin methoxy groups and aromatic carbons to which they are bound, respectively, and two broad peak regions between 0 and 90 ppm, and 100 and 140 ppm, typical for (Kraft) lignin (Figure 6 right). A complete assignment of these broad peaks is complicated; among others, methoxy groups, sp^2 hybridized and aromatic carbons contribute to them [43]. The NMR signature of these lignin related parts does not change significantly, even after normal HTC at 220 °C or salt HTC at 180 °C (Figure 5).

These NMR results confirm the discussion regarding the morphologies seen in the SEM and TEM images (Figure 4). In the hydrochars obtained from coconut, the cellulose related peaks obviously survive if normal HTC is undertaken at 180 °C, even if the reaction time is extended to 20 h (Figure 5a(I)). The relative increase in the signals related to lignin indicates that parts of the cellulose are disrupted and dissolved to the liquid phase, but a temperature of 180 °C is not sufficient to completely carbonize cellulose by HTC with water.

If the temperature is increased to 200 or 220 °C, the cellulose peaks disappear due to complete dissolution of the cellulose (Figure 5a(II)). The small cellulose signal left in the spectrum of the sample treated at 200 °C is completely gone at 220 °C and the lignin related peaks, which are small in the original shell powder, become more prominent. The broad signal from around 0 to 60 ppm results mostly from aliphatic carbons. The wide distribution indicates the presence of a variety of different CH_x sites. Weak peaks at 28 and 37 ppm, in addition to a shoulder at 13 ppm, can be attributed to quite rigid CH_2 , more mobile CH_2 and mobile CH_3 groups, respectively. A large number of signals now become visible in the aromatic (100–160 ppm) region, attributable to C=C-C (112 ppm), sp^2 hybridized carbons (126 ppm) and O-C=C (152 ppm), respectively, and even some carbonyl groups (174 and 208 ppm) are recognizable. These results hint that the structure of the hydrochar from salt HTC is mainly formed by carbon structures originating from the lignin parts of the biomass. The lignin parts form the macrostructure of the biomass as seen in the SEM and TEM images (Figure 4), whereas the holes and roughness are created by breaking of the cellulose particles. With increasing temperature, small changes in the lignin signals can also be seen. The two signals at around 55 and 147 ppm decrease in intensity. Moreover, the small signals at 114 and 129 ppm in the aromatic carbon region and signals



at 12 and 29 ppm in the aliphatic region become smaller. This shows that lignin is attacked if normal HTC is performed at temperatures of about 220 $^{\circ}$ C or higher.

Figure 5. ¹³C NMR spectra of hydrochars obtained from coconut shells by normal HTC (**a**) or salt HTC (**b**) obtained at different times (I) or temperatures (II).



Figure 6. ¹³C NMR spectra of cellulose (a) and lignin (b) obtained by normal HTC or salt HTC at 180 °C and 12 h reaction time.

By using the salt–water mixture, disruption of the cellulose is achieved already after 4 h at 180 °C (Figure 5b(I)). The ZnCl₂ enables reactions at 180 °C that are only possible at temperatures of at least 200 °C in the normal HTC reaction. Thus, the ¹³C-NMR signatures of Cn 200C 12 h and Cn 220C 12 h (Figure 5a(II)) are quite similar to those of Cn 180C 12 h

Zn (Figure 5b(II)). It seems that the zinc chloride acts as a kind of catalyst for hydrolysis, dehydration and decarboxylation [44]. For HTC experiments at 180 °C in the presence of strong mineral acids (i.e., H_2SO_4), a complete hydrolysis of the cellulose to glucose is reported [28]. However, an increased surface area in the hydrochars is not described in that study.

With longer reaction times in salt HTC at 180 °C, the lignin-related ¹³C-NMR signals change, and also indicate a starting conversion of the lignin (Figure 5b(I)). These changes are even more pronounced if the temperature is increased to 220 °C (Figure 5b(II)). Thus, the signal at around 128 ppm becomes more dominant. Because this signal represents sp² hybridized carbon in aromatic compounds, it indicates a high degree of polymerization and, thus, an advanced state of char formation.

This demonstrates that the lignin structures, which are always discussed as a problem compound in the biomass conversion due to their resistance in the fermentation-based biogas production, and in the normal HTC process and even in low-temperature pyrolysis [45], can in fact be easily destroyed by the addition of the zinc chloride to the HTC mixture. Therefore, the salt not only catalyzes the degradation of cellulose but also opens routes for reactions with the lignin compounds in the biomass. This additional attack of lignin is also the reason for the higher pore volumes and surface areas in all the salt-HTC treated hydrochars because the destruction of lignin polymer structures creates additional porosity.

3.5. Surface Area and Adsorption Studies

The surface areas as measured with N₂ for the different HTC-derived hydrochars, and the related steam-activated secondary products, are shown in Figure 7. The surface areas of hydrochars obtained by salt HTC are slightly increased compared to those after normal HTC, presumably due to the catalytic effects of the remaining ZnCl₂, which catalyzes stripping reactions of hydrogen and oxygen [16,27].



Figure 7. Specific surface areas (SSA) of hydrochars from different biomasses after normal HTC, salt HTC (both at 180 °C for 12 h) and subsequent steam activation for 1 h at 500 °C.

Additional steam activation increases the specific surface area of all hydrochars by a considerable factor. All activated samples showed an increased uptake of nitrogen in the pressure region $p/p_0 < 0.1$ with a type I(b) isotherm, indicating the high measured areas were due to the presence of micropores and possibly narrow mesopores (<~2.5 nm) [38]. The highest BET surface area, of well above 740 m²·g⁻¹, was obtained by activating the hydrochar from sawdust for 1 h at 500 °C with steam. During steam activation, a sufficiently high temperature ($T \ge 500$ °C) is required to increase the surface area; up to 350 °C, only SSAs of about 200 m²·g⁻¹ were observed. Interestingly, after steam activation, the chars

from dried dandelion and sawdust exhibited higher surface areas if no salt was previously

present in the HTC process, and was most distinct in the latter. The presence of more large structures in these samples presumably increases the chance to develop inner porosity. The high ash contents in the chars from dried dandelion, in general, lower the obtainable surface areas.

Figure 8 showcases typical nitrogen sorption isotherms for hydrochars received from HTC with water or the salt/water mixture along with those of their steam-activated followup products for the example of coconut shell biomass. The figure clearly shows the very flat isotherm leading to low SSA (<10 m²·g⁻¹) after normal HTC samples with water. Using the aqueous salt mixture in HTC, an increase in surface area by a factor of 16 is obtained. This increase could be derived from the breaking of large structures, as visible by SEM measurements. In the isotherms, a formation of some microporosity is indicated by the slightly sharper increase in the low relative pressure range < 0.04 p/p_0 (Figure 8 green). Most significant, however, is the formation of mesopores, ranging from 3 to about 10 nm in size, indicated by the small hysteresis loop in the isotherm. The type H3 hysteresis loop also points to the additional existence of macropores which are not completely filled with the N₂ condensate [46].



Figure 8. Nitrogen sorption isotherms of coconut shell-based hydrochars from normal HTC (Cn-HTC, red), salt HTC (Cn-HTC Zn, green) both at 180 °C for 12 h, and the follow-up products after steam activation at 500 °C for 1 h (Cn-Act500C, blue and Cn-Act500C Zn, orange).

Steam activating increases in the surface area to about 500 m²/g, mainly due to introduction of microporosity as evident from the typical type I shape of theN₂ isotherms in the low relative pressure range [36]. Micropore formation may occur by oxidation of surface carbon compounds caused by a rapid desorption of the formed CO₂. It is known that, for example, in some MOF materials, an increase in micropore and mesopore sizes is possible with compressed CO₂ [47]. The well-defined plateau visible for Cn-Act500 points to a microporous material with a relatively small external surface, whereas Cn-Act500 Zn shows a further increase at high relative pressure, also indicating the presence of an extended mesoporosity [37]. In the literature, a comparable but less pronounced gas uptake at high relative pressures is described as an indication of small interstitial voids between small particles [15].

3.6. Adsorption Studies with the Dyes Methylene Blue and Methyl Orange

Figure 9a shows the capacities q_e of the salt-HTC hydrochars (12 h, 180 °C) and their steam-activated (1 h, 500 °C) chars obtained from the three different substrates (coconut shell powder, dried dandelion leaves and sawdust) in the adsorption of the dye methylene blue. Surprisingly, the HTC hydrochars were able to remove more dye from the solution than their steam-activated follow-up products. The adsorption capacities for the salt-HTC hydrochars range between 60 and 70 mg·g⁻¹ but after steam activation the capacity drops to about 35 mg·g⁻¹ for the coconut shell powder-derived char and below 15 mg·g⁻¹ for the other two substrates.



Figure 9. (a) Adsorption capacity at equilibrium (q_e) of dyes onto different hydrochars (HTC at 180 °C for 12 h) and chars resulting from subsequent steam activation. (b) Hydrochars from coconut shell powder obtained under different treatments are compared.

It was supposed that the higher surface areas of the activated hydrochars (up to 740 m²/g from gas-phase N₂ adsorption) would offer more adsorption sites, leading to higher dye sorption. However, the formed micropores (Figure 8) are too small for dye uptake and, moreover, the observed loss of surface functionality (Figure 3) lowers the necessary interaction strength. Although having different surface areas (8.92 m²/g (Dd-HTC Zn) and 32.60 m²/g (Sd-HTC Zn)) their values for q_e MB were almost identical (~60 mg·g⁻¹). This supports the idea of higher importance of the surface roughness, functionality and available pore sizes of about 3 to 10 nm over the BET surface area, which is dominated by about 1 nm wide pores, for the dye adsorption.

This confirms literature data on the adsorption kinetics of methylene blue or malachite green onto tailored activated carbons from rice husk [48,49]. Although the full decolorization of a MB solution required more than 25 h when using microporous samples with surface areas of more than 2000 m²/g, less than 10 min was needed with carbon materials that exhibit lower BET surface areas (600 m²/g) but possess an increased amount of mesopores.

The dye adsorption on the carbons obtained from coconut shell powder (Figure 9b) shows the same trend; in line with the increase in the mesoporosity of the hydrochars obtained with the aqueous salt mixtures in the HTC reaction, the amount of dye removed from solution doubles compared to that with hydrochars from HTC with pure water. After the steam activation created micropores, the adsorption capacity decreased strongly for all samples, and mostly for the chars obtained without salt possessing or mesopores. An increase in the steam temperature during activation from 500 to 650 °C even increased the negative effect; the q_e of the salt-HTC hydrochars decreased further from about 35 mg·g⁻¹

to about 26 mg·g⁻¹, because a portion of the mesopores collapsed to micropores. However, even after activation, the dye adsorption is much higher on salt-HTC hydrochars than on those from the normal HTC process, due to the presence of mesopores which survive the steam activation (Figure 8).

The same trend holds if MO was used instead of MB, whereas the amount of dye removed from solution was always lower for MO than MB due to an inhibited surface coverage and pore accessibility for the larger MO, which has an average molecular size of 26.14 Å, compared to MB with 16.34 Å [50,51]. The necessity of mesopores is even more pronounced for the larger dye, as can be seen by the larger ratio in the q_e values of hydrochars from salt HTC and HTC (Figure 9b).

For both dyes, MB and MO, maximum adsorption was found for the salt HTCobtained hydrochar Cn-HTC Zn, of 70.13 and 42.15 mg·g⁻¹, respectively, demonstrating the high potential for pollutant removal of hydrochars obtained in the salt-assisted HTC process due to a good compromise between some mesoporosity (Figure 8) and some surviving functional groups at the inner surface (Figure 3).

If the temperature of the salt HTC is increased from 180 to 220 °C and the reaction time is prolonged from 12 to 20 h, for coconut shell-based hydrochars the BET surface areas increase from 88 m²/g to about 200 m²/g. This increase is mainly related to an increased mesopore volume. With such hydrochars an MB adsorption capacity q_e of 207 mg·g⁻¹ was obtained (Figure 10).



Figure 10. Pseudo-first-order kinetic plot resulting from the effect of contact time on the adsorption of MB onto a coconut shell based hydrochar from salt-HTC at 220 °C.

This adsorption capacity exceeds those reported by Saetea and Tippayawong, who determined for the removal of MB with steam-activated hydrochars from sewage sludge an adsorption capacity of around 160 mg·g⁻¹ [52]. In their study a much higher steam temperature of 900 °C was applied. Other reported adsorption capacities for MB with HTC salt-obtained hydrochars from pure fructose [25] and corncob [53] are with q_e values of 83 and 99, respectively, in the same range of our 180 °C salt-HTC treated chars. All three studies do not report on the adsorption of MO. For MO, Martin-Jimeno et al. reported a q_e of 29 for hydrochar from pure glucose, which was HTC treated in the presence of graphene oxide [54]. Although HTC was performed under different conditions with the more sophisticated and expensive additive graphene oxide, the q_e value is in the same range as for our 180 °C salt-HTC treatment (Figure 9b).

The adsorption of MB onto activated carbons is considered to take place via different mechanisms, such as electrostatic interactions (I), hydrogen bonding formation (II), electron

donor-acceptor (III) and π - π dispersion interaction (IV). In general, interaction IV might be the most important, because the basal plane of an activated carbon is constituted mainly of graphene layers with fewer functional groups present for the other mechanisms to take place [55].

However, an FTIR study after adsorption of the cationic dye methylene green showed that -OH and -C=O groups present on the hydrochar surface interact much more strongly than the aromatic C=C bonds [56]. By increasing the surface functionality via oxy-genation, the dye adsorption can be enhanced significantly. Moreover, metal cations, e.g., Cu^{2+} , are better adsorbed on hydrochars compared to pyrolytic chars obtained at high temperatures of 700 °C or above, even if the pore system of the hydrochars is less well developed [11]. The reason is again the presence of more oxygen-containing groups on the surface. These results are in good agreement with the fact that after the salt-HTC treatment, the highest MB adsorption is obtained, because in these samples oxygen-containing groups are still present on the inner surface of the hydrochars (Figure 3), which are lost after treatment with steam.

Regarding the adsorption kinetics of MB, it can be seen in Figure 10 that most of the dye was removed within the first minutes. After just 5 min, with $q_t = 47 \text{ mg} \cdot \text{g}^{-1}$, around one-quarter of the final equilibrium capacity ($q_e = 207 \text{ mg} \cdot \text{g}^{-1}$) was achieved. After 30 min, nearly 50% of the capacity was reached ($q_t = 94 \text{ mg} \cdot \text{g}^{-1}$) and after 8 h a value of 185 mg \cdot g⁻¹ was close to the final equilibrium capacity. When comparing pseudo-first-order, pseudo-second-order and Elovich kinetic models to fit the data, the pseudo-second-order model suits best, with a correlation coefficient $R^2 = 0.9979$. Therefore, the rate of adsorption probably depends less on the concentration of the adsorbate in the aqueous dye solution and more on the availability of the adsorption sites, as described by the pseudo-second-order model [57].

Additionally, the initial concentration can be an important driving force for the mass transfer from the solution to the solid. Therefore, the dependency of the initial dye concentration on the adsorption capacity was tested. The initial concentration c₀ ranged from 1 to 25 mg·L⁻¹ where, with increasing c_0 , the adsorption capacity q_e increased from 48 to 200–350 mg \cdot g⁻¹ (Figure 11a). Although there is a strong spread of data points due to the low weights and high dilutions of the different compounds, a quick saturation trend is visible. The obtained data points were fitted using three different isotherm models: Langmuir, Freundlich and Temkin. The obtained isotherms data are shown in Figure 11b–d. Table 2 summarizes the isotherm parameters determined from the slopes and intercepts of the linear fitting. The data was best fitted to the Langmuir isotherm model, with a correlation coefficient of 0.9682. The other two models showed a very poor representation of the data, with values for R^2 of around 0.5 (Freundlich: $R^2 = 0.5436$; Temkin $R^2 = 0.5031$). The good fit to the Langmuir isotherm indicates rather homogenous adsorption places on which the methylene blue will be adsorbed in a monolayer. It is highly probable that these adsorption places are the oxygen atoms of the functional groups still found in the non-activated samples by FTIR (Figure 3, sample Cn-HTC Zn) [57].

Model	Parameter	Value
Langmuir	$q_m [\mathrm{mg} \cdot \mathrm{g}^{-1}]$	277.01
	K_L [L·mg ⁻¹]	2.6159
	R^2	0.9682
Freundlich	$K_F [(mg \cdot g^{-1}) (L \cdot mg^{-1})^{1/n}]$	199.29
	1/n	0.1140
	R^2	0.5436
Temkin	$A_T \left[L \cdot g^{-1} \right]$	1455.59
	B	90.5810
	R^2	0.5031

Table 2. Parameters of the different isotherm models for the adsorption of MB onto a hydrochar.

The maximum monolayer adsorption capacity of the methylene blue (q_m) found from the Langmuir isotherm was determined to be about 277 mg·g⁻¹, significantly exceeding the above-mentioned values of different hydrochars [25,52,54].



Figure 11. Equilibrium study on the adsorption of MB onto a hydrochar (**a**) and the resulting Langmuir equilibrium isotherm (**b**), Freundlich isotherm (**c**), and Temkin isotherm (**d**).

4. Conclusions

It was shown that the approach of using aqueous salt mixtures as a reaction medium for the hydrothermal carbonization of complex biomass substrates results in altered particle size, surface morphology and functionality, in addition to an ultimately higher surface area of the obtained hydrochars when compared to those received from a normal HTC process with water only.

In addition to further improving the typical increase in carbon content in HTC, an enhancement of the otherwise low surface areas of the carbons (<10 m²/g) was observed for two of the three substrate tests (sawdust and coconut shell powder), of up to 88 m²/g at 180 °C carbonization temperature, and even 200 m²/g at 220 °C. These results were due to a breaking of larger char structures in the presence of the salts as a result of the degradation and dissolution of cellulose, hemicellulose and even parts of the lignin. This leads to a formation of mesopores and small amounts of micropores, as shown by SEM and N₂ sorption. Moreover, a shift of surface functionality to more carbonyl and aromatic functionality took place as seen by FTIR measurements. Only dried dandelion was identified as an inappropriate substrate for HTC processes due to its high ash content and low reaction yields.

We demonstrated that it is possible to alter the carbon content, surface morphology and surface area of hydrochars by use of zinc chloride in the HTC process as an easily removable and recyclable additive. It was proven that the salt could be re-used in at least three HTC runs without significant loss of surface area of the obtained hydrochars [58].

The ZnCl₂ helps to break cellulose and lignin structures in the biomass at significantly lower, and thus more energy-efficient, temperatures. This could be concluded from the analysis of the biomass substrate and the obtained hydrochars by solid state ¹³C NMR spectroscopy. In particular, the cracking of the usually HTC-resistant lignin structures indicates significant novel potential in the hydrothermal carbonization of lignocellulosic biomass materials.

Additional steam activation can significantly increase the surface areas of all hydrochars, by up to a value of about 740 m²·g⁻¹ for chars from sawdust. The duration of activation plays a minor role compared to the temperature and the presence of large biomass structures, which provide an extensive introduction to microporosity. The accompanied loss of surface functionality, however, is a drawback. Nonetheless, the formation of hydrochars under saline conditions is advantageous, as they show increased surface roughness and increased mesoporosity with preserved functionality, rendering them an interesting choice for pollutant removal, as shown by the results from the dye adsorption experiments. The dye adsorption takes place via external and intraparticle diffusion, and strongly depends on the presence of suitable oxygen-containing adsorption sites, as seen from the pseudo-second-order kinetics. This confirms the expectation that chemisorption processes are more important than pure physisorption.

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