

Article

Rust Fungi on Medicinal Plants in Guizhou Province with Descriptions of Three New Species

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Abstract: During the research on rust fungi in medicinal plants of Guizhou Province, China, a total of 9 rust fungal species were introduced, including 3 new species (*Hamasporea rubi-alceifolii*, *Nyssopsora altissima*, and *Phragmidium cymosum*), as well as 6 known species (*Melampsora laricis-populina*, *Melampsoridium carpini*, *Neophysopella ampelopsidis*, *Nyssopsora koelrezidis*, *P. rosae-roxburghii*, *P. tormentillae*). Notably, *N. ampelopsidis* and *P. tormentillae* were discovered for the first time in China, while *M. laricis-populina*, *Me. carpini*, and *Ny. koelreuteriae* were first documented in Guizhou Province. Morphological observation and molecular phylogenetic analyses of these species with similar taxa were compared to confirm their taxonomic identities, and taxonomic descriptions, illustrations and host species of those rust fungi on medicinal plant are provided.

Keywords: medicinal plant; molecular phylogeny; new taxa; phytopathogen; *Pucciniales*



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1. Introduction

Rust fungi comprise the largest and most ubiquitous group of obligately biotrophic fungi on vascular plants [1]. The economic impact of rust fungi cannot be ignored. Because of rust fungi, many economic plants suffer diseases and reduce yields [2]. For example, when wheat is harmed by *Puccinia striiformis* Westend., its yield can be reduced up to 50% [3]. The impact of rust fungi on tropical crops is also immeasurable [4]. The coffee rust fungus has a global distribution and is often found in the coffee growing areas of China, with a greater impact on the main coffee producing countries [2,5,6].

More than 8000 species of rust fungi have been identified, mainly on the basis of their morphological characteristics of teliospores and spermonogia, however, there are still a larger number of genera *incertae sedis* [7–11]. Rust occurs on ferns to advanced monocots and dicots, and they are obligate biotrophic phytopathogens that produce not only basidiospore but also four other different types of spores: aeciospore, urediniospore, teliospore, and spermatia [8,12,13]. Many species in the order *Pucciniales* were not described with all types of spores, and have various lifestyles (micro-, hemi-, demi-, or macrocyclic), with alternation on single (autoecious) or two unrelated host plants (heteroecious) [7,14]. To date, approximately 8400 rust species are currently recognized worldwide, and 71 genera and 1175 species have been discovered in China so far [11]. However, species diversity, host alternation and geographic distribution of rust fungi in China remain poorly understood.

Medicinal plants are also infected by rust fungi on a large scale [15]. By the end of 2020, a total of 79 rust species have been reported on 76 medicinal plant species from 33 families, and these rust fungi restricts the development and utilization of medicinal plants and affects the quality of botanicals [16,17]. In recent years, as many as 3924 species of medicinal plants are cultivated in Guizhou province, and the number of medicinal plants is increasing number year by year [18]. As important pathogenic fungi, rust can infect the leaf and stem of a variety of medicinal plants and affect their quality and yield which eventually hampered the development and utilization of medicinal plant resources [19]. For example, rust diseases are frequently found on pepper leaves in Guizhou province, and those diseases seriously affect the normal development of pepper, with their incidence reaching 90% in serious infections [20]. However, there are few researches on rust species infecting medicinal plant diseases in Guizhou province [21]. Therefore, it is of great significance to investigate the species diversity of rust fungi on important medicinal plants for local medicinal production.

In 2021, an investigation of rust fungi on the medicinal plants was carried out in Guizhou province, China. Nine species including three new species were found on medicinal plants. Detailed descriptions and illustrations of all those novel species and other species on the medicinal plants are provided.

2. Materials and Methods

2.1. Sample Collections

Rust infected specimens were collected from Guiyang, Qingzhen and Anshun cities in Guizhou province, China. All hosts and habitats information of specimens was recorded. For each specimen, part of specimens was kept in a refrigerator at 4 °C, and the other part was made as a dry specimen [22]. Specimens were deposited in both Mycological Herbarium of the Chifeng University, Inner Mongolia, China (CFSZ) and Herbarium of Guizhou Medical University (GMB).

2.2. Morphology

The specimens were observed under a stereomicroscope (Nikon SMZ745T, Nikon Corporation, Tokyo, Japan) and shot with a Canon digital camera (Canon EOS 1500D, Canon Inc., Tokyo, Japan) fitted on. Microscope images of the samples were taken by a Canon EOS 700D digital camera fitted on the Nikon ECLIPSE Ni compound microscope (Nikon, Japan). Measurements were taken with the Tarosoft (R) Image Frame Work (v.0.9.7). More than 30 morphological characteristics such as teliospores, urediniospores, and paraphyses were measured for each specimen. Photo plates were arranged by using Adobe Photoshop CS6 v. 13 (Adobe Systems Software Ireland Ltd, San Jose, USA). The different spore stages of rust fungi are designated by the following Roman numerals: spermogonia/spermatia (0), aecia/aeciospores (I), uredinia/urediniospores (II), telia/teliospores (III), and basidia/basidiospore (IV). We applied the definitions of spore stage based on Cummins and Hiratsuka [7], and followed morphological types of spermogonia designated by Hiratsuka and Hiratsuka [23].

2.3. DNA Extraction, Polymerase Chain Reaction (PCR), and Sequencing

The rust sori were picked out into a sterilized centrifuge tube with a sterilized fine needle for DNA extraction. Genomic DNA was extracted following the manufacturer's protocol of the OMEGA E.Z.N.A.[®] Fungal Genomic DNA Extraction Kit (D3390, Guangzhou Feiyang Bioengineering Co., Ltd., Guangzhou, China). DNA extracts were stored at −20 °C. PCR was carried out in a volume of 25 µL containing 9.5 µL of ddH₂O, 12.5 µL of 2 × Taq PCR Master Mix (2 × Taq Master Mix with dye, TIANGEN, China), 1 µL of DNA extraction and 1 µL of forward and reverse primers (10 µm each) in each reaction. Primers pairs, ITS4/ITS5 and LR0R/LR5 (Sangon Biotech, Shanghai, China) were used to amplify the regions of internal transcribed spacer (ITS) and large subunit ribosomal (LSU), respectively [8,24–26]. PCR profiles for the ITS and LSU were: initially at 95 °C for

5 min, followed by 35 cycles of denaturation at 94 °C for 1 min, annealing at 52 °C for 1 min, polymerization at 72 °C for 1.5 min and a final extension at 72 °C for 10 min. PCR products were sequenced by Sangon Biotech (Shanghai) Co., Ltd., China.

2.4. Phylogenetic Studies

All sequences used for phylogenetic tree construction were listed in Table 1. Sequences were aligned by MAFFT v. 7.394 (<https://www.ebi.ac.uk/Tools/msa/mafft/>) (accessed on 12 September 2023) [27] and adjusted to ensure maximum similarity using TrimAl v1.4.1 [28]. Alignments were converted from FASTA to PHYLIP format by using Alignment Transformation Environment online (<https://sing.ei.uvigo.es/ALTER/>) (accessed on 12 September 2023) [29]. Maximum Likelihood (ML) analyses and Bayesian posterior probabilities (BYPP) based on a combination of ITS and LSU sequence data were performed using RAxML-HPG 7.4.2 BlackBox [30] and MrBayes v. 3.2.7 tools in the CIPRES Science Gateway platform [31,32]. GTR+I+G was estimated as the best-fit substitution model by jModelTest2 on XSEDE v.2.1.6 [33,34]. The Bootstrap values of ML analyses were obtained by running 1000 replicates by using a Markov chain Monte Carlo (MCMC) method to approximate the posterior probabilities of trees. Six simultaneous Markov Chains were run for 3,000,000 generations and trees were sampled every 1000th generation. Finally, the trees were visualized in FigTree v.1.4.4 [35] and edited by using Adobe Photoshop CS6 v. 13 software. The final alignment and phylogenetic trees were deposited in TreeBASE v. 2 under the submission ID30041 (<http://www.treebase.org/>) (accessed on 12 September 2023).

Table 1. GenBank accession number and information of taxa used for phylogenetic analyses.

Family Name	Species	Specimen No.	Host	Province, Country	GenBank Accession No. ITS	GenBank Accession No. LSU	Reference
<i>Chrysomyxaceae</i>	<i>Chrysomyxa empetri</i>	QFB 25015	<i>Empetrum nigrum</i>	Ste-Anne-des-Monts, Quebec, Canada	GU049434	GU049526	[36]
	<i>Rossmatomyces monesis</i>	DAOM 221982	<i>Moneses</i> (=Pyrola) <i>uniflora</i>	Graham Island, British Columbia, Canada	GU049476	GU049547	[36]
<i>Gymnosporangiaceae</i>	<i>Gymnosporangium asiaticum</i>	CUP-0016	<i>Juniperus chinensis</i>	Gifu prefecture, Japan	MN642593	MN642617	[37]
	<i>Gymnosporangium clavariiforme</i>	BRIP 59471	<i>Crataegus</i> sp.	—	—	MW049261	[10]
	<i>Gymnosporangium confusum</i>	LD 1021	<i>Crataegus monogyna</i>	Turkey	HM114219	HM114219	—
	<i>Gymnosporangium cupressi</i>	RSP9998	—	—	KJ720169	KJ720169	—
	<i>Gymnosporangium juniperi-virginianae</i>	MCA3585	<i>Cupressaceae juniperus</i>	Virginia, America	—	MG907217	[10]
	<i>Gymnosporangium sabinae</i>	TNM F0030477	<i>Pyrus communis</i>	Bulgaria: Sofia	KY964764	KY964764	[38]
	<i>Gymnosporangium trachysorum</i>	RN91	—	—	KJ720184	KJ720184	—
<i>Hyalopsoraceae</i>	<i>Coleopuccinia sinensis</i>	BJFC:R02506	<i>Cotoneaster microphyllus</i>	China	—	MF802285	[39]
	<i>Hyalopora aspidiotus</i>	PUR N4641	<i>Gymnocarpium dryopteris</i>	China	—	MW049264	[40]
	<i>Melampsoridium alni</i>	H 7019539	<i>Alnus mandshurica</i>	Finland	KF031557	KF031534	[41]
	<i>Melampsoridium betulinum</i>	BPI 871107 (MCA2884)	<i>Alnus</i> sp.	Costa Rica	—	DQ354561	[8]
	<i>Melampsoridium hiratsukanum</i>	PDR #1480181	<i>Alnus rhombifolia</i>	USA: Santa Cruz County, California	KC313888	KC313888	[42]
	<i>Melampsoridium carpini</i>	KR-M-0048587	<i>Carpinus betulus</i>	—	MH908486	MH908486	[43]
	<i>Melampsoridium carpini</i>	GMB0112	<i>Carpinus turczaninowii</i>	Guizhou, China	OQ067091	—	This study
<i>Melampsoraceae</i>	<i>Melampsora abietis-canadensis</i>	1399MEA-POG-USA	<i>Populus grandidentata</i>	Wisconsin, America	JN881733	JN934918	[44]
	<i>Melampsora abietis-populi</i>	TSH-R20042 *	<i>Populus yunnanensis</i>	Yunnan, China	JN881739	JN934933	[44]
	<i>Melampsora apocyni</i>	LYR3	<i>Apocynum venetum</i>	Xinjiang, China	KR296802	KR296803	—
	<i>Melampsora caprearum</i>	NYS-F-003819	<i>Salix caprea</i>	Germany	KU550034	KU550033	[45]
	<i>Melampsora coleosporioides</i>	HNMAP3114	<i>Salix</i> sp.	Japan	KF780755	KF780638	[46]
	<i>Melampsora epiphylla</i>	TSH-R3884	<i>Salix bakko</i>	Japan	KF780787	KF780670	[45]
	<i>Melampsora epitea</i>	TNS-F-121034	<i>Salix viminalis</i>	Germany	KX386070	KX386097	[35]
	<i>Melampsora euphorbiae</i>	HMAS350000	<i>Euphorbia kansui</i>	Guangxi, China	MK518875	MK518545	[11]
	<i>Melampsora euphorbiae</i>	BRIP 39560	<i>Euphorbia peplus</i>	Queensland, Australia	—	EF192199	[40]
	<i>Melampsora euphorbiae</i>	BPI 871135 (U681)	<i>Euphorbia heterophylla</i>	Oman	—	DQ351722	[47]
	<i>Melampsora humilis</i>	TSH-R7550	<i>Salix koriyanagi</i>	Miyagi, Japan	KF780812	—	[46]
	<i>Melampsora hyperici-sampsonii</i>	HMAS350001 *	<i>Hypericum sampsonii</i>	Guangxi, China	MK518877	MK518547	[11]

Table 1. Cont.

Family Name	Species	Specimen No.	Host	Province, Country	GenBank Accession No. ITS	LSU	Reference
	<i>Melampsora iranica</i>	HMAAC4055	<i>Salix</i> sp.	China	MK372158	MK372191	[48]
	<i>Melampsora kamikotica</i>	HNMAP3186	<i>Chosenia arbutifolia</i>	China	KF780760	KF780643	[46]
	<i>Melampsora laricis-pentandrae</i>	HNMAP3201	<i>Salix pentandra</i>	Inner Mongolia, China	KF780801	KF780684	[46]
	<i>Melampsora laricis-populina</i>	HMAS247977	<i>Populus simonii</i>	Haixi Qinghai, China	MK028583	MK064524	[49]
	<i>Melampsora laricis-populina</i>	GMB0097	<i>Populus lasiocarpa</i>	Guizhou, China	OQ067085	—	This study
	<i>Melampsora larici-tremulae</i>	PFH04-5	<i>Populus tremula</i>	France	JN881744	JN934956	[44]
	<i>Melampsora medusae</i>	BPI 0021209	—	America	JX416848	JX416843	[44]
	<i>Melampsora medusae</i> f.sp. <i>deltoidis</i>	98D10	<i>Populus × euramericana</i>	South Africa	GQ479307	JN934962	[44]
	<i>Melampsora medusae</i> f.sp. <i>tremuloidis</i>	1028ME-LAL-LJ	<i>Larix laricina</i>	Quebec, Lac Saint Jean, Peribonka, Canada	GQ479883	JN934965	[44]
	<i>Melampsora microsora</i>	TSH-R7335	<i>Salix subfragilis</i>	Hiroshima, Japan	KF780833	KF780730	[46]
	<i>Melampsora microspora</i>	PUR:F17540	<i>populus nigra</i>	Iraq	JN881737	JN934931	[44]
	<i>Melampsora occidentalis</i>	MO96H	<i>Populus trichocarpa</i>	Idaho, America	JN881740	JN934934	[44]
	<i>Melampsora pakistanica</i>	BA13c	<i>Euphorbia helioscopia</i>	Pakistan	KX237555	KX237556	[50]
	<i>Melampsora populnea</i>	664ME-POA-BC45.1	<i>Populus alba</i>	British Columbia, Canada	EU808021	FJ666510	[51]
	<i>Melampsora populnea</i>	97MP10	—	France	EU808035	FJ666523	[51]
	<i>Melampsora populnea</i>	AAH00-1	<i>Populus alba</i>	England	AY444772	AY444786	[52]
	<i>Melampsora pruinosa</i>	BPI 1109446	<i>Populus diversifolia</i>	Xinjiang Uygur Zizhiqu, Shule, China	GQ479899	JN934938	[44]
	<i>Melampsora pulcherrima</i>	O8ZK4	<i>Mercurialis annua</i>	Italy	GQ479320	JN934941	[44]
	<i>Melampsora ribesii-purpureae</i>	PURP897-1	<i>Salix purpurea</i>	—	AY444770	AY444791	[52]
	<i>Melampsora euphorbiae</i>	PDD 98363	<i>Ricinus communis</i>	—	—	KJ716352	[36]
	<i>Melampsora rostrupii</i>	01G1.1	<i>Mercurialis perennis</i>	France	EU808038	JN934942	[44]
	<i>Melampsora salicis-albae</i>	NWC-06210	<i>Salix alba</i>	Rothamsted, England	KF780757	KF780640	[46]
	<i>Melampsora salicis-argyreae</i>	HMAS52894 *	<i>Salix argyreae</i>	Xinjiang, China	KF780733	KF780616	[46]
	<i>Melampsora salicis-bakko</i>	TSH-R3879 *	<i>Salix bakko</i>	—	KC631854	KC685611	[45]
	<i>Melampsora salicis-cavaleriei</i>	HMAAC4043	<i>Salix serrulatifolia</i>	China	MK277296	MK277301	[48]
	<i>Melampsora salicis-delavayanae</i>	HMAS248120 *	<i>Salix delavayana</i>	Yunnan, China	MK518954	MK518651	[11]
	<i>Melampsora salicis-purpureae</i>	HMAS62584 *	<i>Salix purpurea</i>	Shandong, China	KF780766	KF780649	[46]
	<i>Melampsora salicis-viminalis</i>	HMAS38658 *	<i>Salix viminalis</i>	Tibet, China	KF780732	KF780615	[46]
	<i>Melampsora</i> sp.	HMAS350003		Xinjiang, China	MK518844	MK518499	[11]
<i>Neophysopellaceae</i>	<i>Neophysopella ampelopsidis</i>	IBA-8618	<i>Ampelopsis brevipedunculata</i>	Kagoshima, Japan	AB354774	AB354741	[11]

Table 1. Cont.

Family Name	Species	Specimen No.	Host	Province, Country	GenBank Accession No. ITS	LSU	Reference
Phakopsoraceae	<i>Neophysopella ampelopsidis</i>	GMB0110	<i>Ampelopsis sinica</i>	Guizhou, China	OQ067090	—	This study
	<i>Neophysopella meliosmae</i>	IBAR-10037	<i>Meliosma myriantha</i>	Ibaraki, Japan	KC815591	KC815650	[53]
	<i>Neophysopella meliosmae-myrianthae</i>	AGLR56	—	—	MK290819	MK290819	—
	<i>Neophysopella meliosmae-myrianthae</i>	IBA-8584	<i>Vitis coignetiae</i>	Tochigi, Japan	AB354785	AB354748	[53]
	<i>Neophysopella orientalis</i>	IBAR-9942	<i>Meliosma tenuis</i>	Tochigi, Japan	KC815597	KC815656	[54]
	<i>Neophysopella vitis-davidii</i>	HMAS350005	<i>Viola fauriana</i>	Chongqing, China	MK518870	MK518536	[11]
	<i>Neophysopella vitis-davidii</i>	HMAS248115 *	<i>Vitis davidii</i>	Yunnan, China	—	MK518593	[11]
	<i>Masseëlla capparisi</i>	RIP 56844	<i>Flueggea virosa</i>	Northern Territory, Australia	JX136798	JX136798	[13]
	<i>Nothoravenelia japonica</i>	HMJAU8598	—	China	—	MK296509	—
	<i>Phakopsora argentinensis</i>	ZT:RB 8248	<i>Croton cf. anisodontus</i>	—	KF528009	KF528009	[55]
	<i>Phakopsora crcis-filii</i>	ZT Myc 48990	<i>Annona paludosa</i>	Sinamary, French Guiana	KF528016	KF528016	[55]
	<i>Phakopsora myrtacearum</i>	PREM 61155 *	<i>Eucalyptus grandis</i>	Maragua, Kenya	NR_132913	KP729473	[56]
	<i>Phakopsora neocheimoliae</i>	RB 3096 in ZT	<i>Annona cherimola</i>	San José Costa Rica	KF528011	KF528011	[55]
	<i>Phakopsora pachyrhizi</i>	BRIP 56941	<i>Neonotonia wightii</i>	Warrumbungle, New South Wales, Australia	—	KP729475	[56]
	<i>Phakopsora phyllanthi</i>	RB 8581	<i>Phyllanthus acidus</i>	Ceará, Brazil	KF528025	KF528025	[55]
	<i>Phakopsora pistila</i>	ZT Myc 48992	<i>Annona sericea</i>	French GuianaIracubo, French Guiana	KF528026	KF528026	[55]
	<i>Phakopsora sophorae</i>	HMAS248098 *	<i>Leptopus chinensis</i>	Beijing, China	—	MK518628	[11]
	<i>Arthuriomyces peckianus</i>	BPI 879271	<i>Rubus occidentalis</i>	New York, America	GU058010	GU058010	[57]
Phragmidiaceae	<i>Gerwasia pittieriana</i>	BPI 843556	<i>Rubus</i> sp.	—	KY764065	KY764065	—
	<i>Gerwasia rubi</i>	HMAS249978	<i>Rubus parkeri</i>	Yunnan, China	MK519039	MK518737	[11]
	<i>Gerwasia rubi</i>	HMAS249983	<i>Rubus setchuenensis</i>	Fujian, China	—	MK518734	[11]
	<i>Gerwasia rubi</i>	BRIP:58369	<i>Rubus</i> sp.	—	—	KT199397	[13]
	<i>Gerwasia rubi-playfairianus</i>	HMAS249840 *	<i>Rubus playfairianus</i>	Guangxi, China	MK518976	—	[11]
	<i>Hamaspora acutissima</i>	BRIP:55606	<i>Rubus rolfei</i>	Philippines	—	KT199398	[13]
	<i>Hamaspora rubi-alceifolii</i>	GMB0116	<i>Rubus alceaefolius</i>	Guizhou, China	—	OQ067533	This study
	<i>Hamaspora rubi-alceifolii</i>	GMB0109 *	<i>Rubus alceaefolius</i>	Guizhou, China	—	OQ067532	This study
	<i>Hamaspora longissima</i>	BPI 871506	<i>Rubus rigidus</i>	Eastern Cape, South Africa	—	MW049262	[40]

Table 1. Cont.

Family Name	Species	Specimen No.	Host	Province, Country	GenBank Accession No. ITS	LSU	Reference
	<i>Hamaspora sinica</i>	HMAS249989	<i>Rubus setchuenensis</i>	Guangdong, China	MK519049	MK518636	[11]
	<i>Kuehneola japonica</i>	BPI 910185	<i>Rosa</i> sp.	—	KY764067	—	—
	<i>Kuehneola uredinis</i>	LD1029	<i>Rubus</i> sp.	New York, America	GU058013	GU058013	[57]
	<i>Kuehneola uredinis</i>	R216	<i>Rubus fruticosus</i>	Belgium	EU014068	EU014068	[58]
	<i>Phragmidium andersonii</i>	HMAS53231	<i>Potentilla fruticosa</i>	China	—	MG669120	[59]
	<i>Phragmidium barclayi</i>	HMAS67281	<i>Rubus austrotibetanus</i>	China	—	MG669117	[59]
	<i>Phragmidium barnardii</i>	BRIP:56945	<i>Rubus multibracteatus</i>	Queensland, Australia:	—	KT199402	[13]
	<i>Phragmidium biloculare</i>	U564	<i>Potentilla flabellifolia</i>	Washington, America	—	JF907670	[60]
	<i>Phragmidium brevipedicellatum</i>	HMUT100463	<i>Potentilla multifida</i>	Xinjiang, China	—	KU059170	—
	<i>Phragmidium butleri</i>	HMAS67841	<i>Rosa macrophylla</i>	China	—	MG669118	[59]
	<i>Phragmidium chayuiensis</i>	BJFC:R02532 *	<i>Rosa duplicata</i>	China	MH128374	NG_064492	[59]
	<i>Phragmidium cibantum</i>	BJFC:R02528 *	<i>Rubus niveus</i>	China	MH128370	NG_064491	[59]
	<i>Phragmidium cymosum</i>	GMB0115	<i>Rosa cymosa</i>	Guizhou, China	OQ067097	OQ067531	This study
	<i>Phragmidium cymosum</i>	GMB0108 *	<i>Rosa cymosa</i>	Guizhou, China	OQ067096	OQ067530	This study
	<i>Phragmidium duchesnea-indica</i>	HMAS249846 *	<i>Duchesnea indica</i>	Yunnan, China	—	MK518681	[11]
	<i>Phragmidium fragariae</i>	WM 1317	<i>Potentilla sterilis</i>	—	—	AF426217	[61]
	<i>Phragmidium fructigenum</i>	HMUT100472	<i>Rosa glomerata</i>	Chongqing, China	—	KU059168	—
	<i>Phragmidium fusiforme</i>	BJFC:R00942	<i>Rosa hugonis</i>	China	—	KP407632	[62]
	<i>Phragmidium griseum</i>	HMAS56906	<i>Rubus crataegifolius</i>	Beijing, China	MH128377	MG669115	[59]
	<i>Phragmidium griseum</i>	HMAS350016	<i>Rosa</i> sp.	Beijing, China	—	MK518530	[11]
	<i>Phragmidium handelii</i>	BJFC:R01030	<i>Rosa webbiana</i>	China	—	KP407631	[62]
	<i>Phragmidium ivesiae</i>	U35	<i>Potentilla gracilis</i>	Utah, America	—	JF907672	[60]
	<i>Phragmidium japonicum</i>	HMAS41585	<i>Rosa laevigata</i>	Fujian, China	MN264716	MN264734	[63]
	<i>Phragmidium jiangxiense</i>	BJFCR 03453 *	<i>Rosa laevigata</i>	Jiangxi, China	MN264715	MN264733	[63]
	<i>Phragmidium kanasense</i>	HMAS248114 *	<i>Rosa fedtschenkoana</i>	Xinjiang, China	—	MK518464	[11]
	<i>Phragmidium leucoaetium</i>	BJFCR 02118 *	<i>Rosa</i> sp.	Yunnan, China	MN264719	MN264737	[63]
	<i>Phragmidium mexicanum</i>	HMAS350019	<i>Phoenix acaulis</i>	Yunnan, China	MK518980	MK518678	[11]
	<i>Phragmidium mexicanum</i>	HMAS350020	<i>Duchesnea indica</i>	Yunnan, China	MK518982	MK518680	[11]
	<i>Phragmidium mexicanum</i>	MCA2496	<i>Potentilla indica</i>	Maryland, America	—	JF907660	[60]
	<i>Phragmidium montivagum</i>	FO 47828	<i>Rosa</i> cf. <i>woodsii</i>	—	—	AF426213	[61]
	<i>Phragmidium mucronatum</i>	U-671	<i>Rosa</i> sp.	Oman	—	HQ412646	[64]
	<i>Phragmidium mucronatum</i>	BRIP:60097	<i>Rosa rubiginosa</i>	—	—	MW049275	[40]
	<i>Phragmidium nambuanum</i>	HMUT100465	<i>Rubus innominatus</i>	Chongqing, China	—	KU059165	—
	<i>Phragmidium octoloculare</i>	HMAS140416	<i>Rubus biflorus</i>	China	MH128376	MG669119	[59]

Table 1. Cont.

Family Name	Species	Specimen No.	Host	Province, Country	GenBank Accession No. ITS	LSU	Reference
Pucciniastraceae	<i>Phragmidium pauciloculare</i>	HMUT100466	<i>Rubus innominatus</i>	Chongqing, China	—	KU059162	—
	<i>Phragmidium pauciloculare</i>	HMAS350023	<i>Rubus corchorifolius</i>	Guangxi, China	MK518874	MK518542	[11]
	<i>Phragmidium potentillae</i>	BRIP:60089	<i>Acaena novae-zelandiae</i>	Tasmania, Australia	—	KT199403	[13]
	<i>Phragmidium potentillae-canadensis</i>	MCA2858	<i>Potentilla</i> sp.	New York, America	—	JF907666	[60]
	<i>Phragmidium punjabense</i>	BA65A *	<i>Rosa</i> sp.	Murree, Ghora Gali, Pakistan	KX358856	KX358854	[50]
	<i>Phragmidium rosae-californicae</i>	MVAP0000154	<i>Rosa californica</i>	Putah Creek Reserve, Davis, California, America	MK045315	MK045315	—
	<i>Phragmidium rosae-moschatae</i>	BPI 893257	<i>Rosa macrophylla</i>	—	—	KY798368	—
	<i>Phragmidium rosae-multiflorae</i>	HMAS71053	<i>Rosa multiflora</i>	Shanxi, China	—	KU059174	[65]
	<i>Phragmidium rosae-multiflorae</i>	HMAS94924	<i>Rosa multiflora</i>	Zhejiang, China	—	KU059175	[65]
	<i>Phragmidium rosae-multiflorae</i>	BJFCR 03454 *	<i>Rosa multiflora</i>	Jiangxi, China	MN264721	MN264739	[63]
	<i>Phragmidium rosae-roxburghii</i>	HGUP21025 *	<i>Rosa roxburghii</i>	Guizhou, China	OL684818	OL684831	[65]
	<i>Phragmidium rosae-roxburghii</i>	GMB0104	<i>Rosa xanthina</i>	Guizhou, China	OQ067092	—	This study
	<i>Phragmidium rubi-idaei</i>	HMUT100470	<i>Rubus saxatilis</i>	Chongqing, China	—	KU059163	—
	<i>Phragmidium rubi-oldhami</i>	HMAS64306	—	China	—	MG669116	[59]
	<i>Phragmidium tormentillae</i>	GMB00114	<i>Potentilla simulatrix</i>	Guizhou, China	OQ067093	—	This study
	<i>Phragmidium tormentillae</i>	BPI 843392	<i>Duchesnea</i> sp.	Maryland, America	—	DQ354553	[8]
	<i>Phragmidium tuberculatum</i>	BPI 877977	<i>Rosa floribunda</i>	Massachusetts, America	—	KJ841923	[66]
	<i>Phragmidium tuberculatum</i>	BPI 877980	<i>Rosa floribunda</i>	Oregon, America	—	KJ841922	[66]
	<i>Phragmidium violaceum</i>	PDD 99246	<i>Rubus</i> sp.	New Zealand	—	KJ716351	[36]
	<i>Phragmidium yangii</i>	BJFC:R00338 *	<i>Rosa</i> sp.	China	—	NG_060138	[62]
	<i>Phragmidium zangdongii</i>	BJFC:R02447 *	<i>Rosa tibetica</i>	Tibet, China	MH128372	NG_064490	[59]
	<i>Phragmidium zhouquense</i>	BJFC:R01516 *	<i>Rosa omeiensis</i>	China	—	NG_060139	[62]
	<i>Trachyspora alchemillae</i>	WM 1019	<i>Alchemilla vulgaris</i>	—	—	AF426220	[61]
	<i>Trachyspora alchemillae</i>	BPI 843828	<i>Alchemilla vulgaris</i>	Switzerland	—	DQ354550	[8]
	<i>Xenodochus carbonarius</i>	PUR N15566	<i>Sanguisorba officinalis</i>	—	—	MW049289	[40]
	<i>Melampsorella elatina</i>	WM 1092	<i>Abies alba</i>	—	—	AF426232	[61]
	<i>Melampsorella elatina</i>	PUR 82	<i>Cerastium fontanum</i>	Minnesota, America	—	MG907233	[10]
	<i>Pucciniastrum circaeae</i>	RB 2098	<i>Circaea lutetiana</i>	—	—	AF426227	[61]
	<i>Pucciniastrum corni</i>	TSH-R13510	<i>Cornus kuosa</i>	Tottori, Japan	AB221437	AB221409	[67]
	<i>Pucciniastrum epilobii</i>	PUR N11088(MCA5308)	<i>Epilobium angustifolium</i>	—	—	MW147083	[40]

Table 1. Cont.

Family Name	Species	Specimen No.	Host	Province, Country	GenBank Accession No. ITS	LSU	Reference
Sphaerophragmiaceae	<i>Pucciniastrum fagi</i>	TSH-R10724	<i>Fagus crenata</i>	Tochigi, Japan	AB221425	AB221378	[67]
	<i>Pucciniastrum hikosanense</i>	TSH-R4287 (IBA2565)	<i>Acer rufrinerve</i>	Yamanashi Japan	AB221441	AB221388	[67]
	<i>Pucciniastrum hydrangeae</i>	MCA2837	<i>Hydrangea arborescens</i>	North Carolina, America	—	MG907240	[10]
	<i>Pucciniastrum kusanoi</i>	HiroU21509	<i>Clethra barbinervis</i>	Miyazaki, Japan	AB221426	AB221402	[67]
	<i>Pucciniastrum yoshinagai</i>	TSH-R4272 IBA8430	<i>Stewartia monadelphae</i>	Nara, Yoshino-gun, Japan	AB221434	AB221411	[67]
	<i>Thekopsora minima</i>	MCA2988	<i>Vaccinium angustifolium</i>	Maryland, America	—	MG907243	[10]
	<i>Austropuccinia psidii</i>	BRIP:58164	<i>Rhodamnia angustifolia</i>	Brisbane, Queensland. Australia	—	KF318449	[67]
	<i>Dasyscypha amazonica</i>	BPI US0116382 *	<i>Xylopia amazonica</i>	Brazil	JF263460	JF263460	[68]
	<i>Dasyscypha gregaria</i>	ZT Myc 3397	<i>Xylopia cayennensis</i>	French Guiana	JF263477	JF263477	[68]
	<i>Dasyscypha guianensis</i>	ZT Myc 3413 *	<i>Xylopia benthamii</i>	French Guiana	JF263479	JF263479	[68]
	<i>Dasyscypha mesoamericana</i>	PUR 42390 *	<i>Xylopia frutescens</i>	Panama	JF263480	JF263480	[68]
	<i>Dasyscypha segregaria</i>	PMA MP4941	<i>Xylopia aromatica</i>	Panama	JF263488	JF263488	[68]
	<i>Dasyscypha winteri</i>	S F30078	<i>Xylopia sericea</i>	Brazil	JF263492	JF263492	[68]
	<i>Puccorchidium polyalthiae</i>	ZT HerB 251	<i>Polyalthia longifolia</i>	India	JF263493	JF263493	[68]
Uredinineae incertae sedis	<i>Sphaerophragmium acaciae</i>	BRIP:56910	<i>Albizia</i> sp.	Kununurra, Western Australia, Australia	—	KJ862350	[69]
	<i>Sphaerophragmium longicorne</i>	PUR N16513	<i>Dalbergia hostilis</i>	—	—	MW147053	[40]
	<i>Sphenorchidium xylopieae</i>	PC 0096711	<i>Xylopia aethiopica</i>	western Central Africa.	KM217355	KM217355	[70]
	<i>Allodus podophylli</i>	BPI 842277	<i>Podophyllum peltatum</i>	Maryland, America	DQ354543	DQ354543	[67]
	<i>Neopuccinia bursa</i>	RB 75707	<i>Protium heptaphyllum</i>	Brazil	—	MH047186	—
	<i>Nyssopsora altissima</i>	GMB0103 *	<i>Ailanthus altissima</i>	Guizhou, China	OQ067089	OQ067529	This study
	<i>Nyssopsora echinata</i>	KR0012164	<i>Meum athamanticum</i>	—	—	MW049272	[40]
	<i>Nyssopsora koelreuteriae</i>	BBSW-1	<i>Koelreuteria bipinnata</i>	—	KT750965	KT750965	[71]
	<i>Nyssopsora koelreuteriae</i>	GMB0105	<i>Koelreuteria bipinnata</i>	Guizhou, China	OQ067088	—	This study
	<i>Nyssopsora thwaitesii</i>	AMH:9528	<i>Schefflera wallichiana</i>	India	KF550283	—	[72]

Notes: —: no data available; *: type specimens.

3. Results

In this study, 11 samples were collected from 9 species of medicinal plants in nine genera in seven families in Guizhou Province. Ten species including three new species were identified based on morphological and molecular phylogenetic studies. Morphological and phylogenetically allied taxa were selected for the final phylogenetic analyses, mainly following Zhao et al. [11]. The alignment for *Melampsoraceae* includes 1200 character (ITS: 532 bp, LSU: 1200 bp) (Figure 1). The dataset of phylogenetic tree from *Neophysopellaceae* and *Phakopsoraceae* has 1162 characters (ITS: 474 bp, LSU: 1162 bp) including gaps (Figure 2). The alignment for *Neophysopellaceae* and *Phakopsoraceae* phylogenetic tree contained 20 taxa. The taxa from genera *Gerwasia*, *Hamasporea*, *Kuehneola*, *Phragmidium*, *Trachyspora*, and *Xenodochus* were included in the phylogenetic tree of the family *Phragmidiaceae* (Figure 3). The alignment is made up of 71 species and has 1244 characters including gaps (ITS: 392 bp, LSU: 1244 bp). The phylogenetic tree of the families *Pucciniastraceae* and *Hyalopsoraceae* includes the taxa from genera *Coleopuccinia*, *Hyalopora*, *Melampsorella*, *Melampsoridium*, and *Pucciniastrum* (Figure 4). In the phylogenetic tree of *Pucciniastraceae*, there are 20 taxa with 1244 characters, including gaps (ITS: 351 bp, LSU: 1244 bp). Twenty-seven representative species from the *Gymnosporangiaceae*, *Sphaerophragmiaceae*, and *Uredinineae incertae sedis* were chosen for Figure 5, which included 1272 characters including gaps (ITS: 398, LSU: 1272). Based on morphological and multi-locus phylogenetic characterisation, 13 specimens were identified to 9 species in 6 genera 5 families (*Hamasporea rubi-alceifolii* sp. nov., *Nyssopsora altissima* sp. nov., *Phragmidium cymosum* sp. nov., *Melampsora laricis-populina*, *Melampsoridium carpini*, *Neophysopella ampelopsidis*, *Ny. koelreuteriae*, *P. rosae-roxburghii* and *P. tormentillae*). Among them, three novel species were recognized, moreover, 2 species were reported first in China, and 4 species were first reported on medicinal plant.

Taxonomy

Based on morphology and molecular phylogeny, all collected rust specimens on medicinal plants were identified as three new species (*Hamasporea rubi-alceifolii* sp. nov., *Nyssopsora altissima* sp. nov., *Phragmidium cymosum* sp. nov.) and six known species (*Melampsora laricis-populina*, *Melampsoridium carpini*, *Neophysopella ampelopsidis*, *Ny. koelreuteriae*, *P. rosae-roxburghii* and *P. tormentillae*). All species parasitic on medicinal plants are described and illustrated below.

Hamasporea rubi-alceifolii Q. Z. Wu, T. Z. Liu, P. Zhao & Q. R. Li, sp. nov. Figure 6
Mycobank number: MB847104

Etymology: Epithet follows the epithet of host species, *Rubus alceifolius* Poir.

Holotype: GMB0109

Parasitic on the leaf and stem of *R. alceifolius*. *Telia* up to 5 mm long, mostly hypophyllous, occasionally amphigenous and stem, caespitose, filiform, fluffy, golden yellow when fresh, white when dry; *Teliospores* 116–230 × 20–10 µm (av. = 184 × 17 µm, n = 30), fusiform, hyaline 3–6 septate, mostly 5–6 septate, smooth, the contents yellow when fresh, solid apex 9–30 µm (av. = 23 µm, n = 30). Spermogonia, aecia and uredinia were not observed.

Materials examined: CHINA, Guizhou Province, Guiyang City, Campus of Guizhou Medical University (26°22′48.37″ N, 106°37′30.33″ E), III on leaf of *R. alceifolius* Poir., 6 October 2021, Q. Z. Wu and L. L. Liu, GMB0109, holotype, CFSZ 50531, isotype; CHINA, Guizhou Province, Guiyang City, Huaxi District, III on leaf of *R. alceifolius* Poir., 20 October 2021, Q. Z. Wu and L. L. Liu, GMB0116.

Notes: *Hamasporea rubi-alceifolii* is characterized by 5–6 septate teliospore with long solid apex up to 30 µm, and hypophyllous telia (Figure 6). Phylogenetically, it formed a distinct clade sister to *H. acutissima* with high support values (100% ML, 1 BYPP; Figure 3). Morphologically, the differences between *H. rubi-alceifolii* and *H. acutissima* are in the number of septa in the teliospores (5–6 vs. 2–3), smaller teliospores (116–230 × 10–20 µm vs. 158–205 × 18–25 µm), and smaller solid apex (9–30 µm vs. 20–40 µm). Both *H. rubi-sieboldii* and *H. rubi-alceifolii* exhibit similar teliospore morphology with consistent solid apex size. However, the difference between *H. rubi-alceifolii*

and *H. rubi-sieboldii* is that the former has more cells (5–6 vs. 4) and smaller solid apex ($116\text{--}230 \times 10\text{--}20 \mu\text{m}$ vs. $118\text{--}240 \times 15\text{--}23 \mu\text{m}$) [73–76].

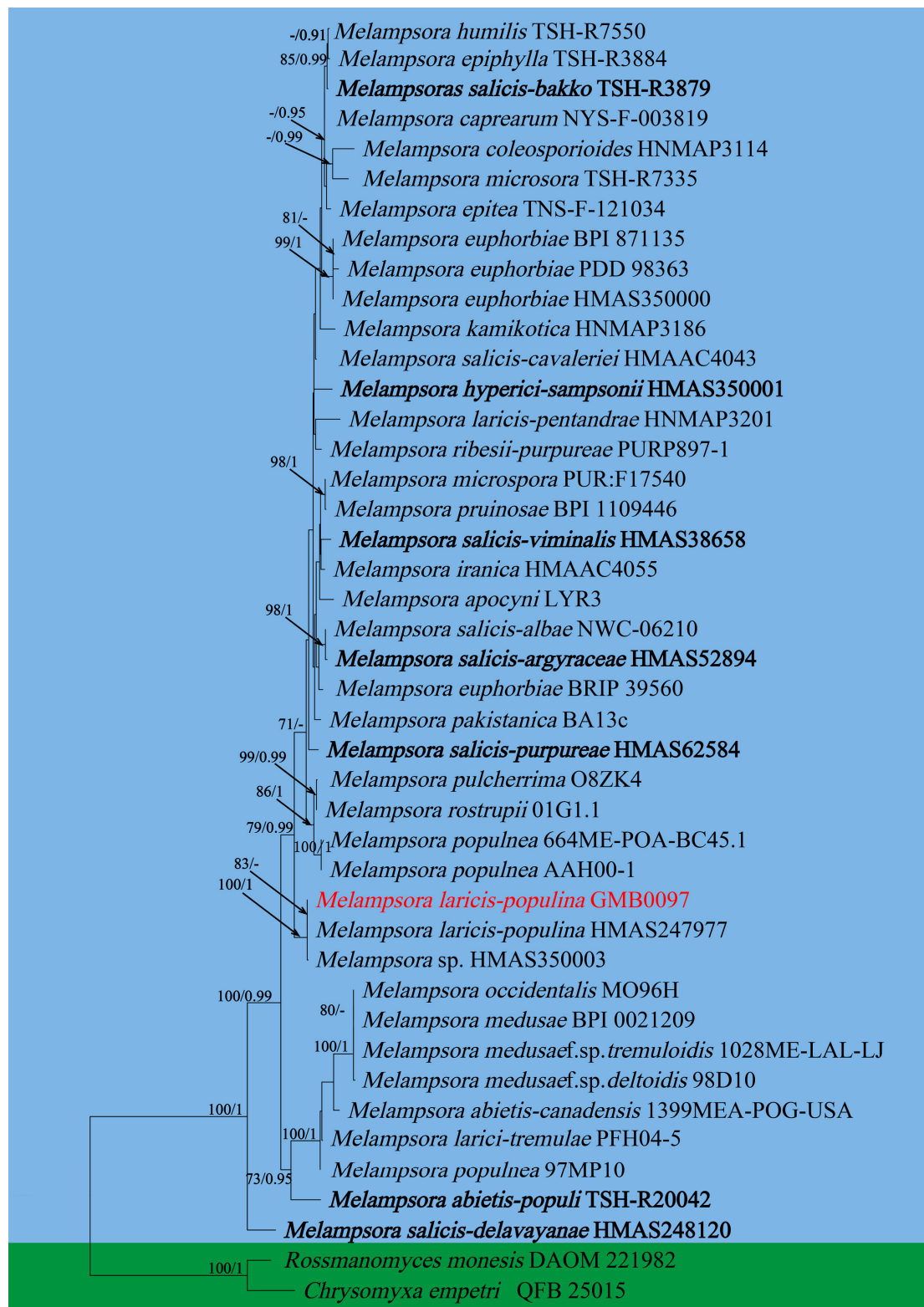


Figure 1. RAxML tree of the family Melampsoraceae based on rDNA ITS and LSU sequence. ML bootstrap supports ($\geq 70\%$) and Bayesian posterior probability (≥ 0.90) are indicated as ML/BYPP. The tree is rooted to *Rossmatomyces monesis* and *Chrysomyxa empetri* [11]. The type specimens are shown as boldface. New sequences are in red.

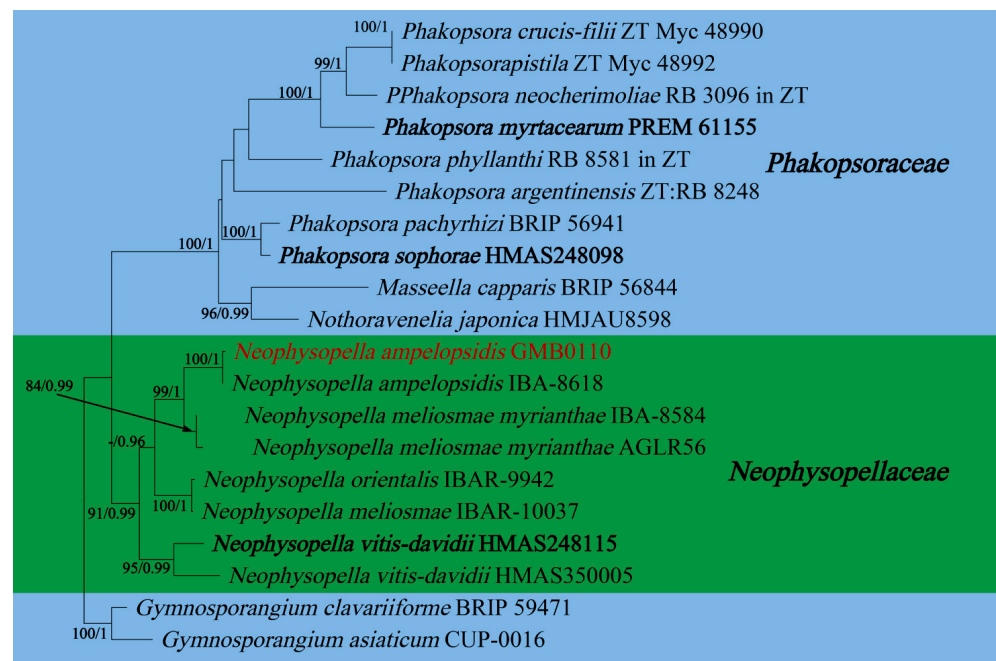
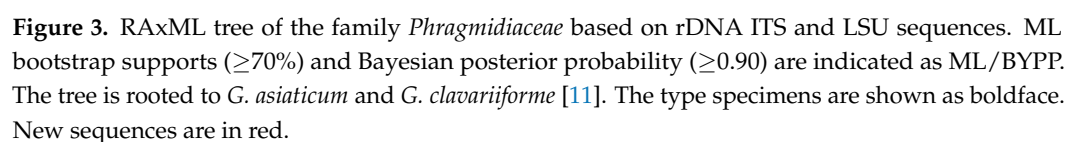


Figure 2. The RAxML tree of the family *Neophysopellaceae* and *Phakopsoraceae* based on rDNA ITS and LSU sequences. ML bootstrap supports ($\geq 70\%$) and Bayesian posterior probability (≥ 0.90) are indicated as ML/BYPP. The tree is rooted to *G. asiaticum* and *G. clavariiforme* [11]. The type specimens are shown as boldface. New sequences are in red.



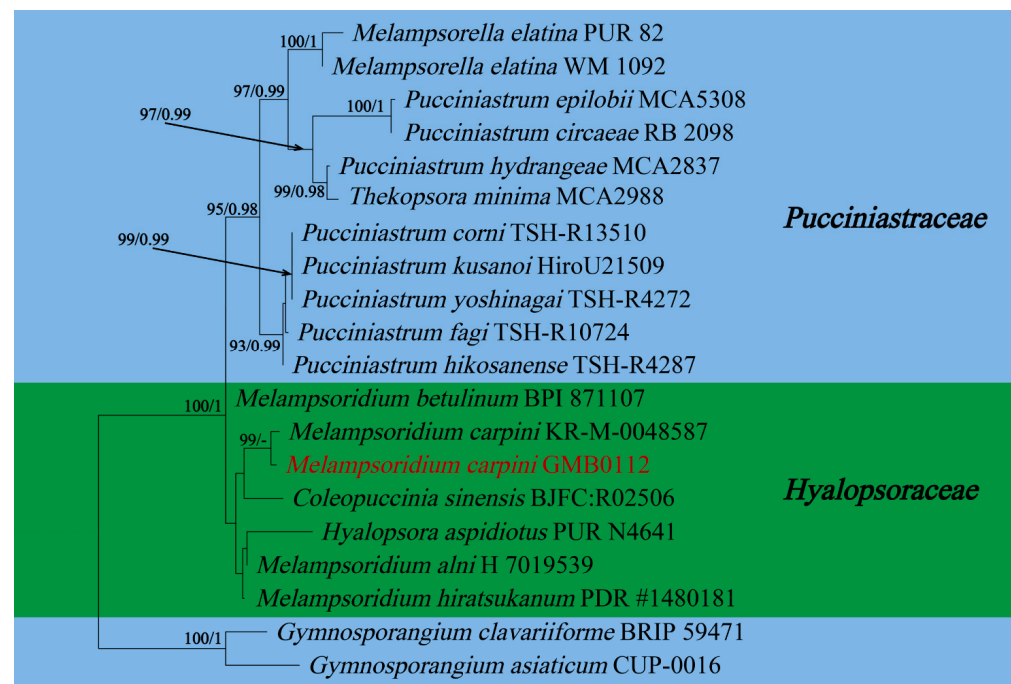


Figure 4. RAxML tree of the family Pucciniastraceae and Hyalopsoraceae based on rDNA ITS and LSU sequence. ML bootstrap supports ($\geq 70\%$) and Bayesian posterior probability (≥ 0.90) are indicated as ML/BYPP. The tree is rooted to *G. asiaticum* and *G. clavariiforme* [11]. New sequences are in red.

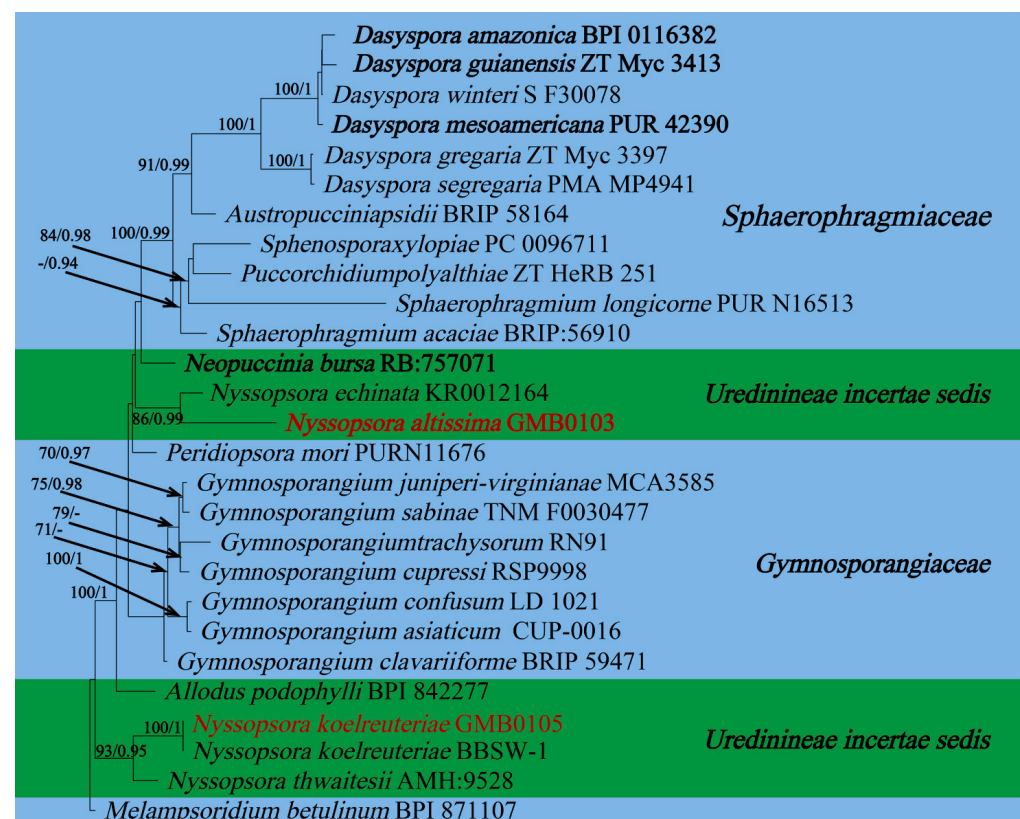


Figure 5. The RAxML tree of the families Gymnosporangiaceae, Sphaerophragmiaceae and Uredinineae incertae sedis based on rDNA ITS and LSU sequences. ML bootstrap supports ($\geq 70\%$) and Bayesian posterior probability (≥ 0.90) are indicated as ML/BYPP. The tree is rooted to *Melampsoridium betulinum* [40]. The type specimens are shown as boldface. New sequences are in red.

Roots and leaves of *Rubus alceifolius* is a traditional Chinese medicine, which can be used for treatment of acute and chronic hepatitis, hepatosplenomegaly and other liver damage diseases [77–81]. Previously, thirteen *Hamasporea* species have already been reported on *Rubus* species [76]. For the convenience of recognition, a worldwide identification key for the *Hamasporea* has been provided.

Key to the Species of *Hamasporea*

1. Uredia present 2
1. Uredia not present 9
2. Paraphyses brown, short *H. rubi-sieboldii*
2. Paraphyses hyaline or absent 3
3. Apex of thick-walled teliospore, not over 5 µm in length;
urediospores 32–56 × 12–42 µm *H. hashiokai*
3. Apex of thick-walled teliospore, usually over 5 µm in length 4
4. Teliospores mostly 2–3 septate 5
4. Teliospores mostly either 3–4 or more septate 7
5. Teliospores 93–138 µm long, solid apex of 8 µm; urediospores 23–30 × 18–28 µm *H. longissima*
5. Teliospores longer than 138 µm 6
6. Teliospores mostly 105–193 µm long, solid apex of 18 µm;
urediospores 28–35 × 25–30 µm with a 3–5 µm thick wall *H. engleriana*
6. Teliospores mostly 158–205 µm long with solid apex of 28–40 µm;
urediospores 18–28 × 13–28 µm with a thin wall *H. acutissima*
7. Teliospores mostly 3–4 septate, 77–199 × 17–24 µm long, solid apex 5–17 µm;
urediospores 19–29 × 14–22 µm *H. australis*
7. Teliospores mostly either 4–5 more septate 8
8. Teliospores mostly 4–5 septate, 148–225 µm long, solid apex 15–30 µm;
urediospores 20–30 × 15–28 µm *H. sinica* var. *sinica*
8. Teliospores mostly 5–6 septate, 117–185 × 12–15 µm long, solid apex 6–12 µm;
urediospores 19–25 × 14–17 µm *H. rubus-pirifolius*
9. Teliospores 3 septate, solid apex 5 – 25 µm *H. sinica* var. *trisepta*
9. Teliospores 2–3 septate or more than 4 septate 10
10. Teliospores mostly 2–3 septate, 79–151 µm long, solid apex 7–24 µm *H. okinawensis*
10. Teliospores 4 or more septate 11
11. Teliospores with solid apex longer than 10 µm *H. gedeana*
11. Teliospores with solid apex not exceeding 10 µm 12
12. Teliospores mostly 90–123 µm long *H. ozeensis*
12. Teliospores longer than 123 µm 13
13. Teliospores mostly 4 septate, 135–178 × 13–18 µm long, solid apex 5–10 µm *H. taiwaniana*
13. Teliospores mostly 5–6 septate, 116–230 × 20–10 µm long, solid apex 9–30 µm *H. rubi-alceifolii*

Melampsora laricis-populina Kleb., Z. PflKrankh. 12: 43 (1902) Figure 7 Mycobank number: MB18930

Uredinia, mostly hypophyllous, seldom epiphyllous, in little groups, 0.25 mm, Bright yellow. Urediniospores oblong to broadly ellipsoid or obovoid, hyaline to light yellow with yellowish granules, 33–45 × 21–26 µm (av. = 37 × 24 µm, n = 30), wall 1.8–3 µm thick, echinulate except at the smooth apex; Paraphyses clavate to capitate, hyaline to pale yellow, 50–71 × 12–20 µm (av. = 62 × 17 µm, n = 30), wall thick, up to 9 µm at the apex.

Materials examined: CHINA, Guizhou Province, Kaili City, Xiasi Town (26°26′29.48″ N, 107°47′30.33″ E) II on *Populus lasiocarpa* Oliv., 1 October 2021, Q. Z. Wu, GMB0097.

Notes: *Melampsora laricis-populina* is characterized by the echinulate urediniospores that are obovate or oval, with golden yellow cytoplasm; roughly spherical paraphyses with swollen tips. The uredinial morphologies of our specimen (GMB0097) are consistent with those of *M. laricis-populina* [82,83]. Phylogenetic tree (Figure 1) showed that our specimen (GMB0097)

was clustered with *M. laricis-populina* with the high bootstrap supports (83/-). Thus, here we confirmed the rust fungus on *Populus lasiocarpa* as *M. laricis-populina*. *Populus lasiocarpa* has hemostatic function and mainly used for treatment for bleeding from trauma [84,85]. Previously, *M. laricis-populina* has already been reported on *Populus lasiocarpa* in Japan [86] and Norway [87], and it has been reported in northwest region of China [88].

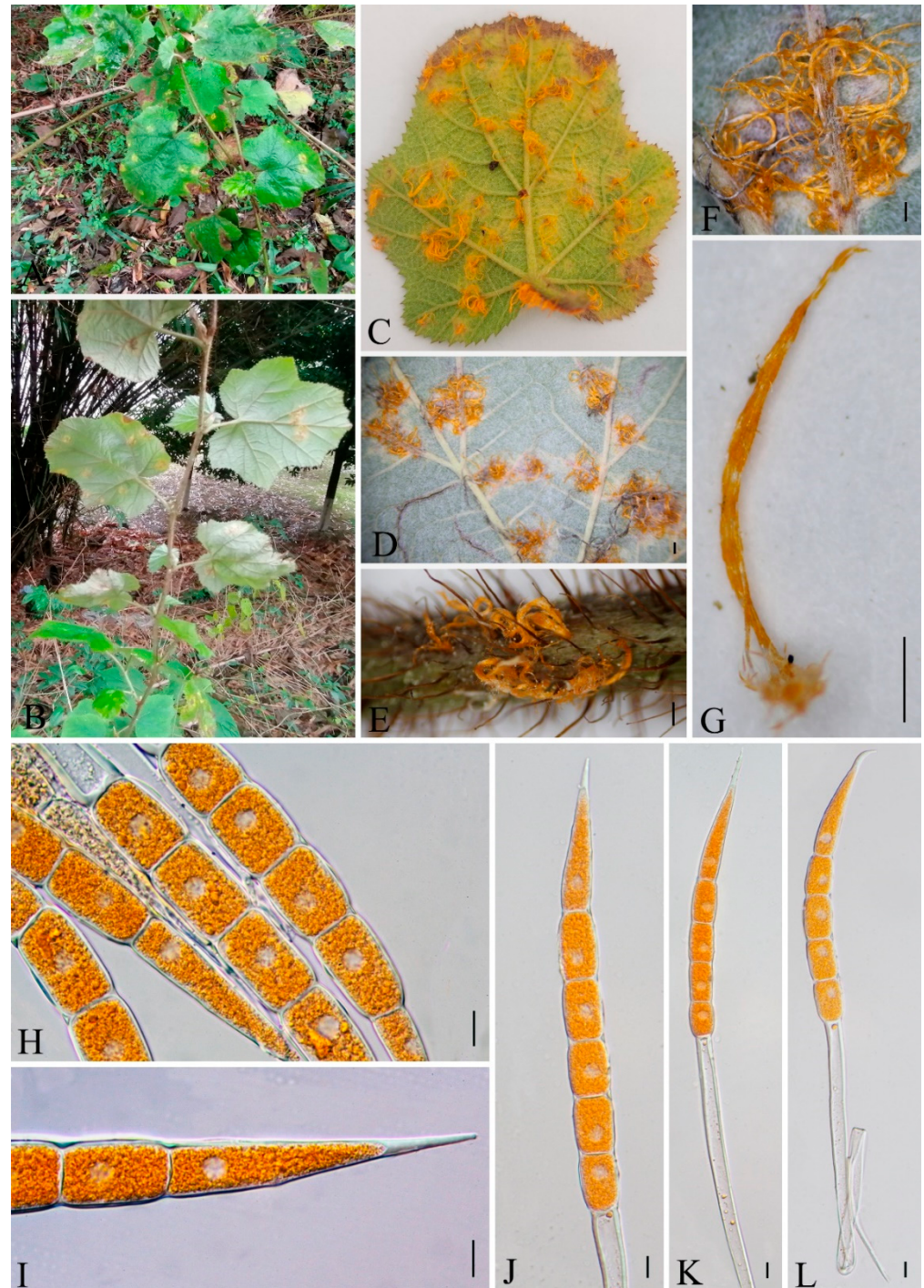


Figure 6. *Hamaspora rubi-alceifolii* (GMB0109). (A,B) host and its habitat; (C–F) telia on the hypophyllous leaf and stem; (G) teliospore cluster; (H–L) fusiform teliospores with 5–6 septa. Scale bars: (D) = 1 mm; (E–G) = 0.5 mm; (H–L) = 10 μ m.

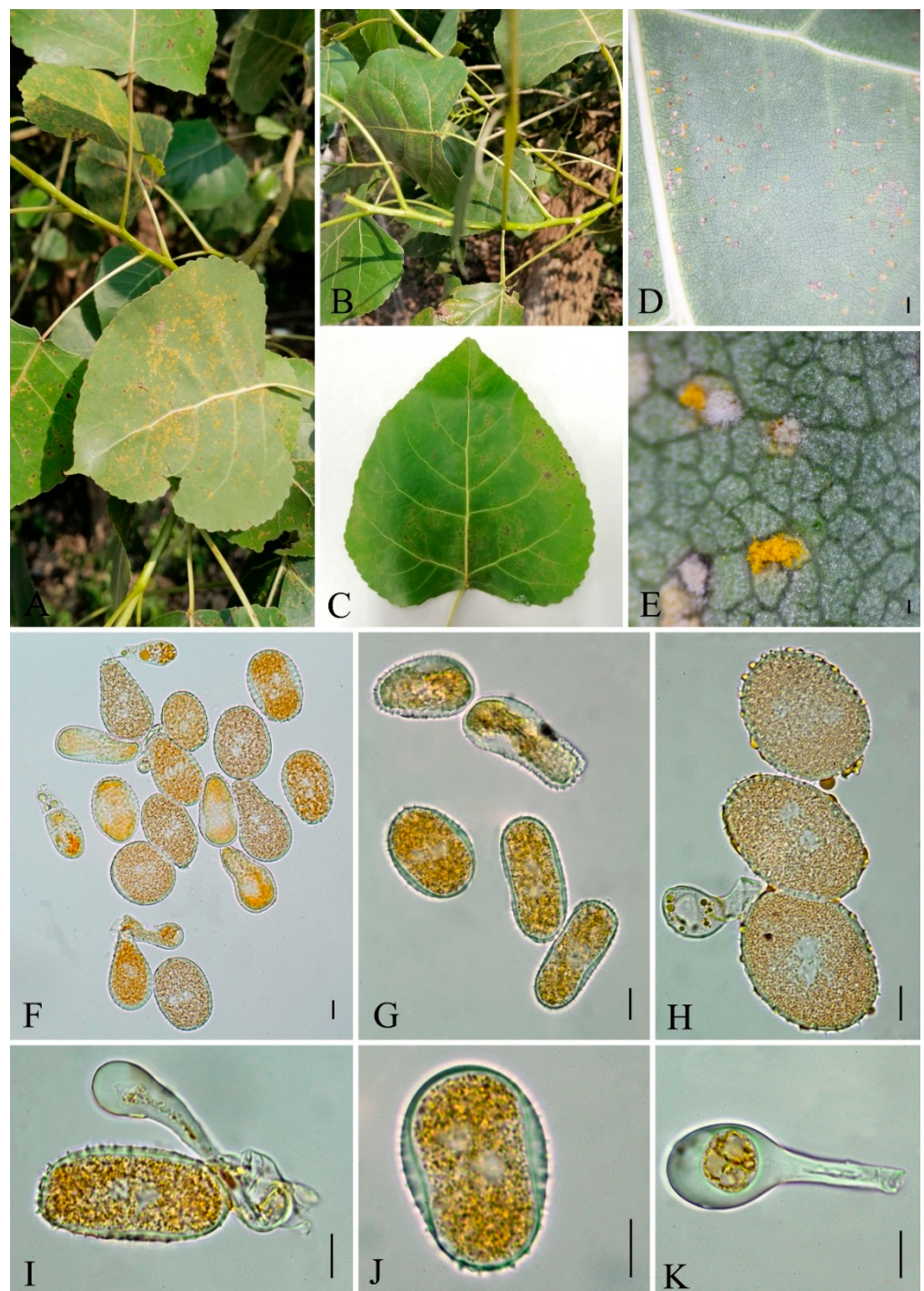


Figure 7. *Melampsora laricis-populina* (GMB0097) (A,B) host and its habitat; (C–E) uredinia; (F–K) urediniospores and paraphyses. Scale bars: (D) = 1 mm; (E) = 0.1 mm; (F–K) = 10 μ m.

Melampsoridium carpini (Nees) Dietel, in Engler & Prantl, Nat. Pflanzenfam., Teil. I (Leipzig) 1(1**): 551 (1900) Figure 8

Basionym: *Caeoma carpini* Nees, Syst. Pilze (Würzburg): 16 (1816) [1816–17]

Mycobank number: 205589

Uredinia, hypophyllous, scattered or grouped on yellow spots, 0.2 mm diam., yellow. Urediniospores long obovoid, clavate or pear shaped, $19\text{--}31 \times 13\text{--}18 \mu\text{m}$ (av. = $26 \times 15 \mu\text{m}$, $n = 30$), yellow; spore wall hyaline, echinulate, on apex smooth, 0.8–1.5 μm thick.

Materials examined: CHINA, Guizhou Province, Qingzhen City, Xiasi Town (26°27'18.48" N, 107°20'7.33" E) II on *Carpinus turczaninowii* Hance, 8 October 2021, Q. Z. Wu and L. L. Liu, GMB0112, CFSZ 50543.

Notes: *Carpinus turczaninowii* is commonly used to treat bruises, canker sores and swellings, as recorded in the Pharmacopoeia of the People's Republic of China. *Carpinus turczaninowii* can alleviate arterial damage and inflammation caused by hyperglycemia [89]. It contains *Pheophorbide A*, which has anti-cancer and anti-inflammatory activity [90,91]. We collected rust infected *Carpinus turczaninowii* in Guizhou province in China, and the urediospores of our specimen (GMB0112) are consistent with those of *Me. carpini*, which is characterized by sparse thorns on the surface and a smooth top of urediospores [92–94]. According to the phylogenetic tree (Figure 4.), the new collection (GMB0112) was clustered with *Me. carpini* with the high bootstrap values (99/-). This species has been found in Anhui, Chongqing, Sichuan, Taiwan and other provinces in China [94], which is the first record of *Me. carpini* from Guizhou province, China.

Neophysopella ampelopsidis (Dietel & P. Syd.) Jing X. Ji & Kakish., in Ji, Li, Li & Kakishima, Mycol. Progr. 18(6): 863 (2019) Figure 9

Basionym: *Phakopsora ampelopsidis* Dietel & P. Syd. [as 'ampelosidis'], in Dietel, Hedwigia 37: 217 (1898)

Mycobank number: MB830298

Uredinia hypophyllous, grouped on brown or yellow spots. the paraphyses, incurved, 30–43 × 6–14 µm (av. = 37 × 11 µm, n = 30), wall 2–3 µm thick (av. = 2.5 µm, n = 30). *Urediniospores*, obovoid or obovoid-ellipsoid, dark yellow or brown, 20–32 × 15–21 µm (av. = 25 × 18 µm, n = 30), walls hyaline, echinulate. The wall was colorless or pale yellowish, equally ca 0.8–1.5 µm thick, and evenly echinulate.

Materials examined: CHINA, Guizhou, Qingzhen City, Xiasi Town (26°27'18.18" N, 107°20'7.13" E) II on *Ampelopsis sinica* (Mig.) W.T. Wang., 22 July 2021. Q. Z. Wu and L. L. Liu, GMB0110, CFSZ 50532.

Notes: *Ampelopsis sinica* root (ASR) is a traditional Chinese medicine known to have a hepatoprotective function. Moreover, it has been proven having anti-hepatocellular carcinogenic activity and to inhibit Hepatitis B virus activity [95–97]. Our rust collection (GMB00110) on *Ampelopsis sinica* is compatible with *Neophysopella ampelopsidis* [98]. Phylogenetically, our collection clustered with *N. ampelopsidis* (IBA-8618) with the high bootstrap supports (100/1, in Figure 2). *N. ampelopsidis* has been previously introduced from Japan, Philippines and Taiwan provinces of China [98]. This is the first record of *N. ampelopsidis* from the Chinese mainland.

Nyssopsora altissima Q. Z. Wu, T. Z. Liu, P. Zhao & Q. R. Li, **sp. nov.** Figure 10

Mycobank number: MB847103

Etymology: Epithet follows the epithet of host species, *Ailanthus altissima* (Mill.) Swingle.

Holotype: GMB0103

Parasitic on the leave of *A. altissima*. *Uredinia*, usually amphigenous, sparse or aggregated, pulverulent, golden; *Urediniospores* subglobose or ellipsoid, 18–23 × 16–24 µm (av. = 21 × 20 µm, n = 30), cell wall 1.2–2.4 µm (av. = 1.5 µm, n = 30) thick, echinulate. *Telia* soft hypophyllous, aggregated, rounded, pulverulent, dark; *Teliospores* subglobose or globose-trigonal, septa constricted slightly, 2–3 cells are mostly 3, reddish-brown to opaque, 28–38 × 20–39.5 µm (av. = 33.5 × 33 µm, n = 30), projections up to 14 (av. = 12, n = 30), 1–6 (av. = 2, n = 30) apical furcations, 2.8–7.8 µm (av. = 4.5 µm n = 30) long and 1.3–3.3 µm (av. = 2 µm, n = 30) thick, each cell has 1–2 germination pores; pedicel hyaline, persistent, about 33–57 µm (av. = 44 µm, n = 30) long, 5.5–9 µm (av. = 7.8 µm, n = 30) thick.

Materials examined: CHINA, Guizhou Province, Guiyang City, Campus of Guizhou Medical University (26°22'48.37" N, 106°37'30.33" E), II, III on leave of *A. altissima*, 8 July 2021, Q.Z. Wu and L.L. Liu, GMB0103, holotype, CFSZ 50535, isotype.

Notes: Phylogenetically, *Nyssopsora altissima* was phylogenetically allied to *Ny. echinata* with a high bootstrap support (86/0.99, Figure 5). Morphologically, *Ny. altissima* differs from *Ny. echinata* by the number of germination pores in teliospores (3 vs. 2), the bigger teliospores

($28\text{--}38 \times 20\text{--}39.5 \mu\text{m}$ vs. $25\text{--}30 \times 23\text{--}27 \mu\text{m}$), and the shorter processes ($2.8\text{--}7.8 \mu\text{m}$ vs. $6.6\text{--}12 \mu\text{m}$). Morphologically, *Ny. altissima* can be distinguished from *Ny. cedrelae* by several morphological differences that bigger urediniospores ($18\text{--}23 \times 16\text{--}24 \mu\text{m}$ compared to $15\text{--}19 \times 13\text{--}17 \mu\text{m}$), smaller teliospores ($28\text{--}38 \times 20\text{--}39.5 \mu\text{m}$ vs. $28\text{--}45 \times 18\text{--}43 \mu\text{m}$), and fewer projections ($1\text{--}14$ vs. $15\text{--}22$). Additionally, *Ny. altissima* has shorter hyaline ($33\text{--}57 \mu\text{m}$ vs. $40\text{--}65 \mu\text{m}$) and finer hyaline ($5.5\text{--}9 \mu\text{m}$ vs. $10\text{--}12 \mu\text{m}$) than those of *Ny. cedrelae* [76,99].

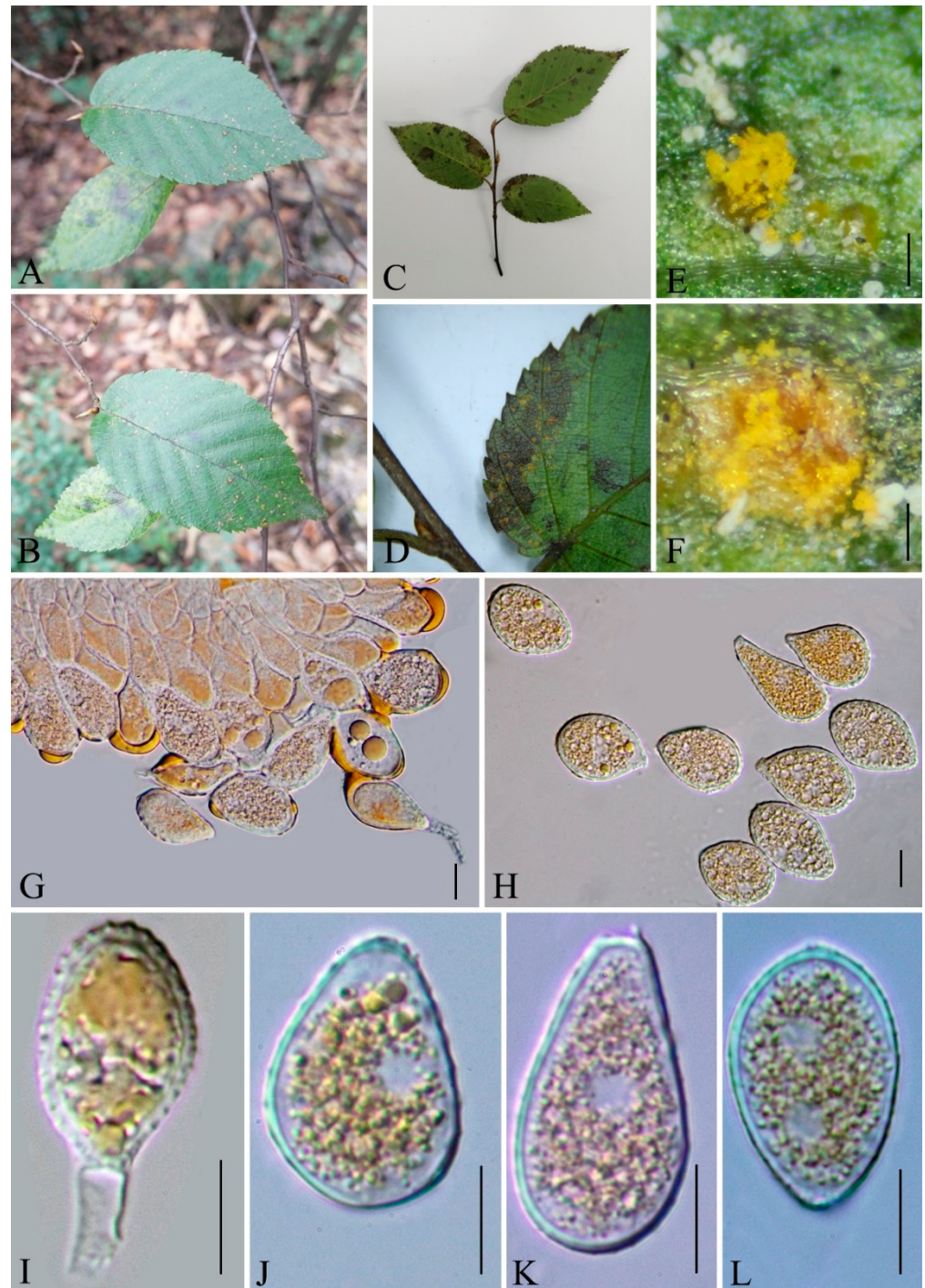


Figure 8. *Melampsoridium carpini* (GMB0112) (A,B) host; (C–F) uredinia; (G–L) urediniospores. Scale bars: (E,F) = 0.1 mm; (G–L) = 10 μm .

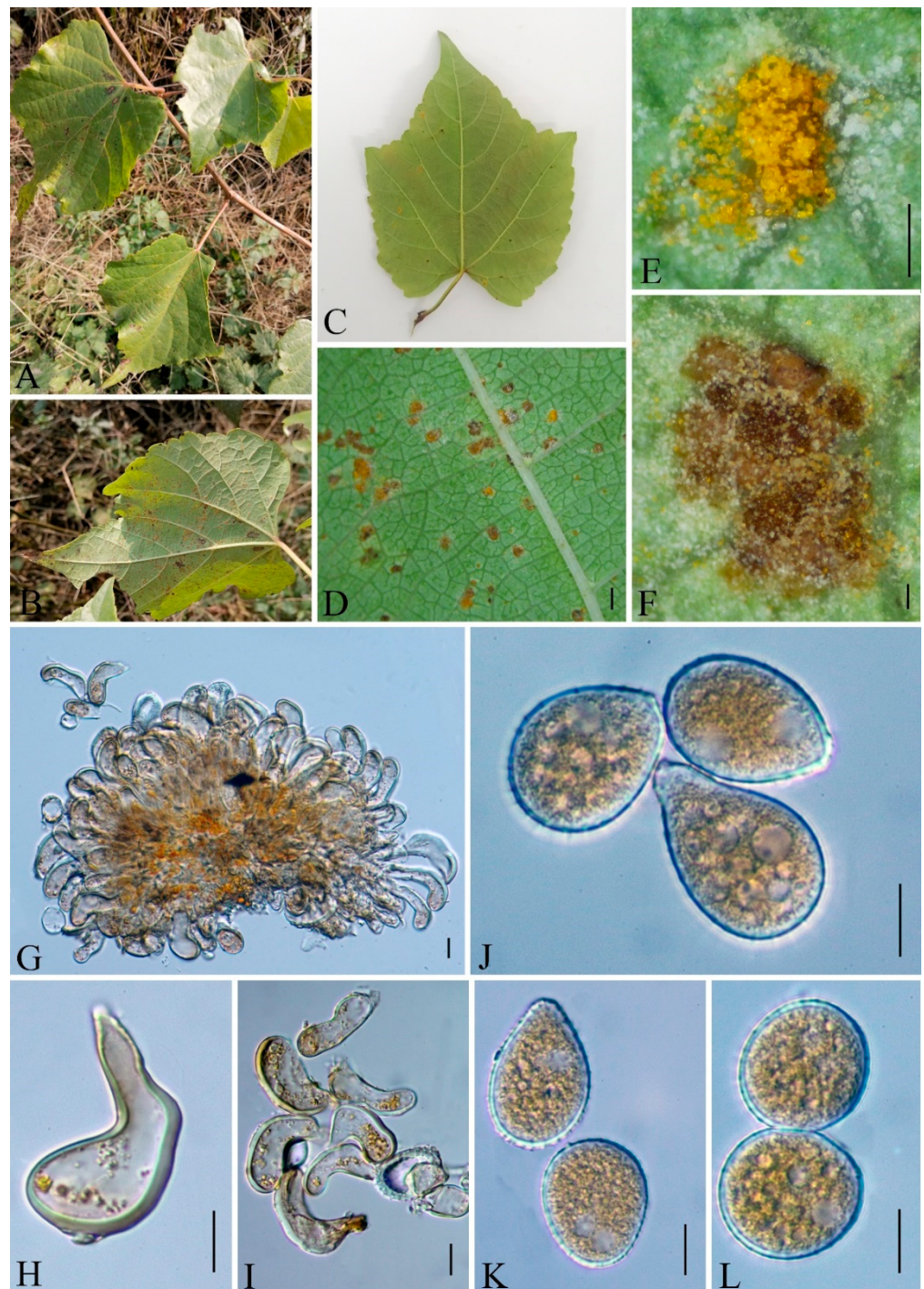


Figure 9. *Neophysopella ampelopsidis* (GMB0110). (A,B) host and its habitat; (C–F) uredinia.; (G–L) urediniospores and paraphyses. Scale bars: (D–F) = 0.1 mm, (G–L) = 10 µm.

Ailanthus altissima has been used as a medicinal herb to hemostasis and Anti-diarrhea documented in Chinese pharmacopoeia [100]. *Ailanthus altissima* has the anti-malarial, anti-viral and anti-tumor active ingredient and shows potential as a novel drug for the treatment of prostate cancer [101–103]. To date, another rust species, *Ny. cedrelae*, has already been reported on *Ailanthus altissima* [76], here we reported another *Nyssopsora* species on *Ailanthus altissima*. Compared to *Ny. cedrelae*, *Ny. altissima* exhibits larger-sized urediniospores ($18\text{--}23 \times 16\text{--}24 \mu\text{m}$ vs. $15\text{--}19 \times 13\text{--}17 \mu\text{m}$), shorter apical furcations ($2.8\text{--}7.8 \mu\text{m}$ vs. $2\text{--}10 \mu\text{m}$), and a shorter pedicel ($33\text{--}57 \mu\text{m}$ vs. $120 \mu\text{m}$).

Nyssopsora koelreuteriae (Syd. & P. Syd.) Tranzschel, J. Soc. bot. Russie 8: 132 (1925) [1923] Figure 11

Basionym: *Triphragmium koelreuteriae* Syd. & P. Syd. 1913

Mycobank number: MB335240

Parasitic on leaves of *Koelreuteria bipinnata*, surrounded by yellowish margins spermatogonia, aecia and uredinia unknown. *Telia* amphigenous, scattered or slightly clustered, densely aggregated in groups, confluent, naked, erumpent, blackish, chestnut-brown. *Teliospores* 27–31 × 24–32 µm (av. = 28 × 27 µm, n = 30), 3-celled with a single proximal cell and two collateral distal cells, triquetrous pyriform, strongly constricted at septa, blackish-brown; walls uniformly 1.5–3 µm thick, pale yellow when young and becoming blackish-brown when older. Projections up to 18 and branched at the tips, 2–3 branched at apex. *Pedicle* 20–26 × 4.5–6.5 µm (av. = 23 × 6 µm, n = 30), persistent, hyaline.

Materials examined: CHINA, Guizhou Province, Anshun City, Longdong Scenic Area (26°6′32.28″ N, 105°52′30.23″ E) III on *Koelreuteria bipinnata* Franch., 10 July 2021 Q.Z. Wu, GMB0105.

Notes: The leaves of *Koelreuteria bipinnata* have strong antimicrobial activity, and their extracts contain a variety of components that can inhibit bacteria and fungi [104,105]. We collected rust samples on the leaves of *Koelreuteria bipinnata*, and it can be identified as *Ny. koelreuteriae* based on both morphological and molecular evidences (Figures 5 and 11). This rust fungus has already been reported in Zhejiang province, China in 1928 [106], and here we reported it on the same host species in Guizhou province.

Phragmidium cymosum Q. Z. Wu, T. Z. Liu, P. Zhao & Q. R. Li, **sp. nov.** Figure 12

Mycobank number: MB847102

Etymology: Epithet follows to the epithet of host species, *Rosa cymosa* Tratt.

Holotype: GMB0108

Parasitic on the leaves of *Rosa cymosa*. *Uredinia* *Uredo*-type (*Uraecium* type II), hypophyllous, yellowish, aggregated, 0.1–0.4 mm (av. = 0.2 mm, n = 30), with thick-walled, gradual thickening by the roots of the paraphyses, incurved, intermixed paraphyses, 25–70 × 9–15 µm (av. = 44 × 11 µm, n = 30), wall 3–11 µm thick (av. = 7 µm, n = 30), top width 5–13 µm thick (av. = 8.5 µm, n = 30), *Urediniospores* borne singly, mostly echinulate, globose, broadly or obovoid, echinulate, 19–27 × 21–29 µm (av. = 24 × 26 µm, n = 30), cytoplasm orange, wall 1–2.5 µm thick (av. = 1.5 µm, n = 30).

Materials examined: CHINA, Guizhou Province, Guiyang City, Campus of Guizhou Medical University (26°22′46.37″ N, 106°37′29.33″ E), II on *R. cymosa*, 6 October 2021, Q. Z. Wu, GY-XGQW GMB0108, holotype, CFSZ 50542, isotype; CHINA, Guizhou Province, Guiyang City, Campus of Guizhou Medical University (26°22′48.22″ N, 106°37′30.12″ E), II on *R. cymosa*, 6 October 2021, Q. Z. Wu, GMB0115.

Notes: *Phragmidium cymosum* formed a distinct lineage in the phylogenetic tree (Figure 3) with a high support rate (98/1). Morphologically, it differs from those species by its paraphyses with relatively thick wall, by the width of the urediniospores (21–29 µm in *P. cymosum*; 18–21 µm in *P. japonicum*; 11–18 µm in *P. jiangxiense*; 15–20 in *P. rosae-multiflorae*) [63,76]. In addition, the paraphyses of *P. cymosum* (25–70 × 9–15 µm) are thicker compared to those of *P. jiangxiense* (22–41 × 6–10 µm), and the cell walls of the paraphyses in *P. cymosum* (3–11 µm) are thicker than those of *P. rosae-multiflorae* (1 µm). *Phragmidium rosae-multiflorae* was once reported on the same host species in China [107].

The leaves, flowers and roots of *Rosa cymosa* can be used as the Chinese herbal medicine. Moreover, it has anti-inflammatory components, can be used to treat burns, analgesic [108–110].

Phragmidium rosae-roxburghii J.E. Sun & Yong Wang **sp. nov.** Figure 13

Mycobank number: MB845041

Parasitic on the leaf of *Rosa roxburghii* Tratt. *Uredinia* *Uredo*-type (*Uraecium* type II), hypophyllous, yellowish, aggregated, with thick-walled, gradual thickening by the roots of the paraphyses, incurved, intermixed paraphyses, 23–50 × 8–13.5 µm (av. = 36.5 × 10 µm,

n = 30), wall 2–8.5 μm thick (av. = 5.5 μm , n = 30), *Urediniospores* borne singly, mostly echinulate, globose, broadly or obovoid, echinulate, 18–29 $\times$ 17–24 μm (av. = 24 $\times$ 20 μm , n = 30), cytoplasm orange, wall 0.5–1.5 μm thick (av. = 1 μm , n = 30).

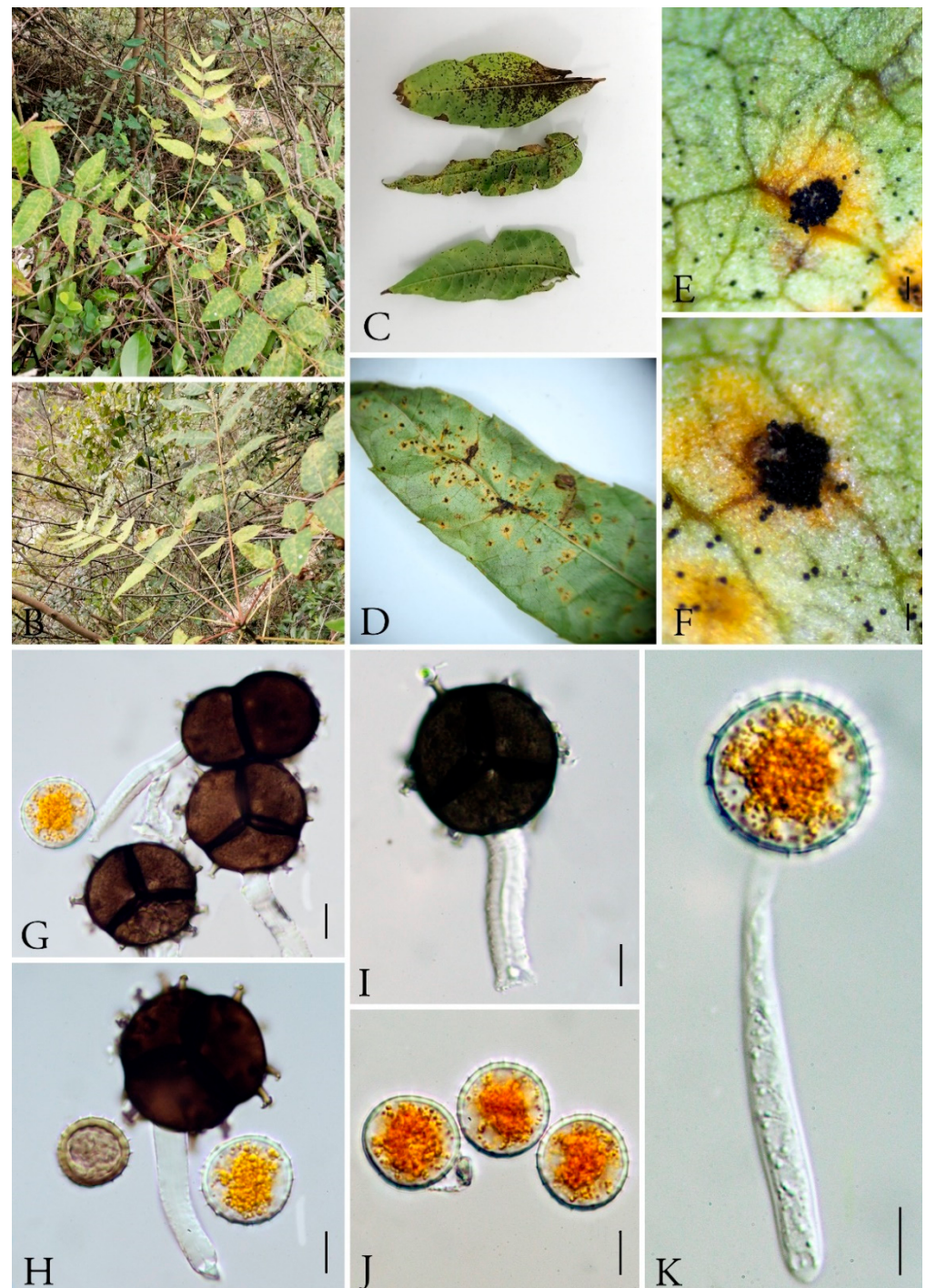


Figure 10. *Nyssopsora altissima* (GMB0103). (A,B) host and its habitat; (C,D) telia on the amphigenous leaf; (E,F) telia; (G–I) teliospores; (J,K) urediniospores with handle. Scale bars: (E,F) = 0.1 mm; (G–K) = 10 μm .

Materials examined: CHINA, Guizhou province, Anshun City, Longdong Scenic Area (26°6′46.32″ N, 105°52′32.23″ E) II on *Rosa roxburghii* Tratt, 10 July 2021 Q.Z. Wu, GMB0104, CFSZ 50540.

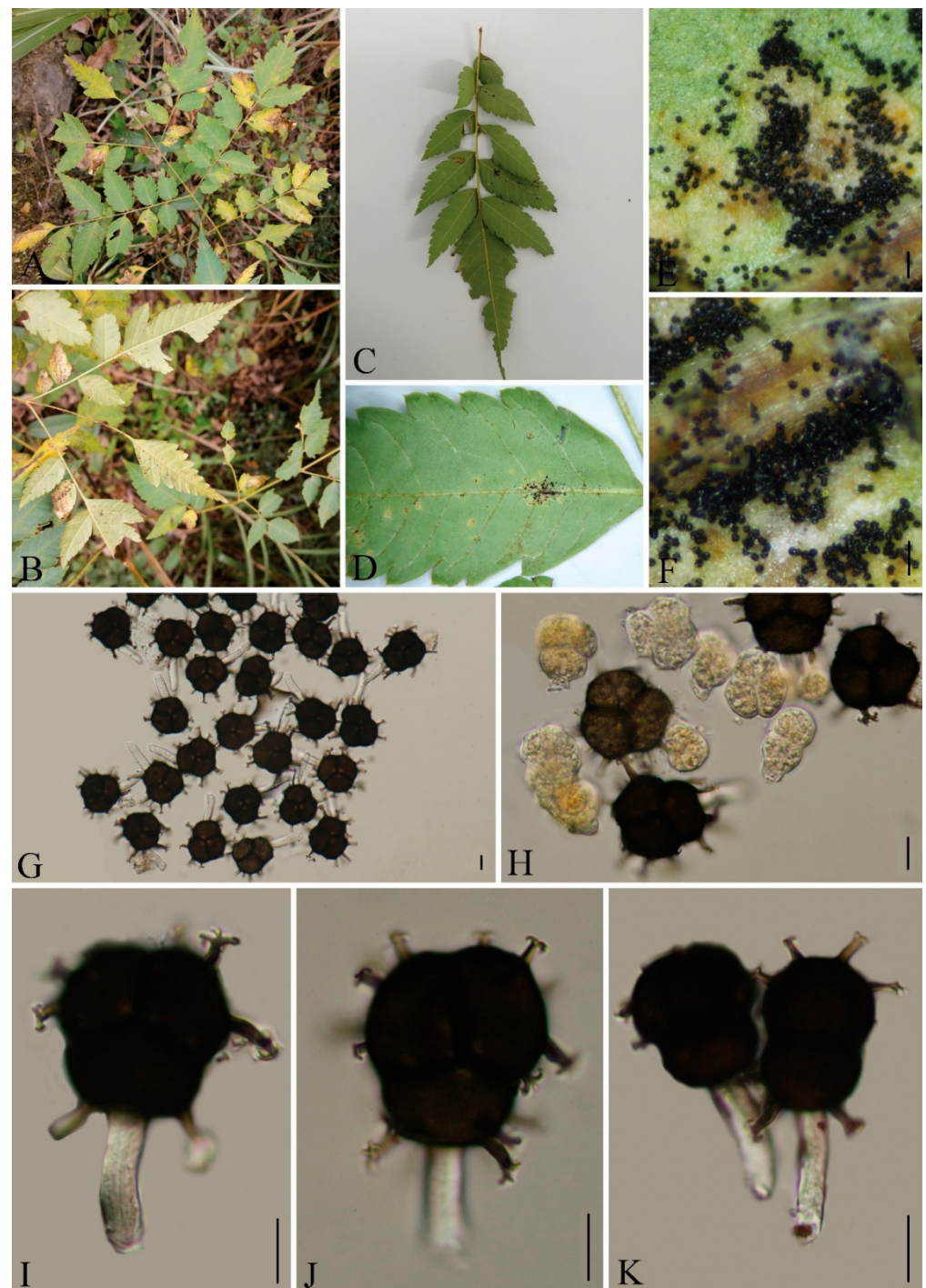


Figure 11. *Nyssopsora koelreuteriae* (GMB0105). (A,B) Host and its habitat. (C–F) Telia (G–K) Teliospores on the stem. Scale bars: (E,F) = 0.1 mm; (G–K) = 10 μ m.

Notes: *Rosa roxburghii* as a Chinese herbal medicine is used as a remedy for respiratory diseases. It has been recently reported to be an antioxidant and anticoagulant, and also be used to treat dyspepsia, dysentery, hypo immunity, and neurasthenia [65,111]. Our collection (GMB0104) and *P. rosae-roxburghii* form a clade in phylogenetic tree with a high support value (100/1). The morphological characteristics of GMB0104 are consistent with those of *P. rosae-roxburghii*. *Phragmidium rosae-roxburghii* was introduced based on the specimen collected from Guizhou province [65]. Here we reported it on medicinal plant in the same province.

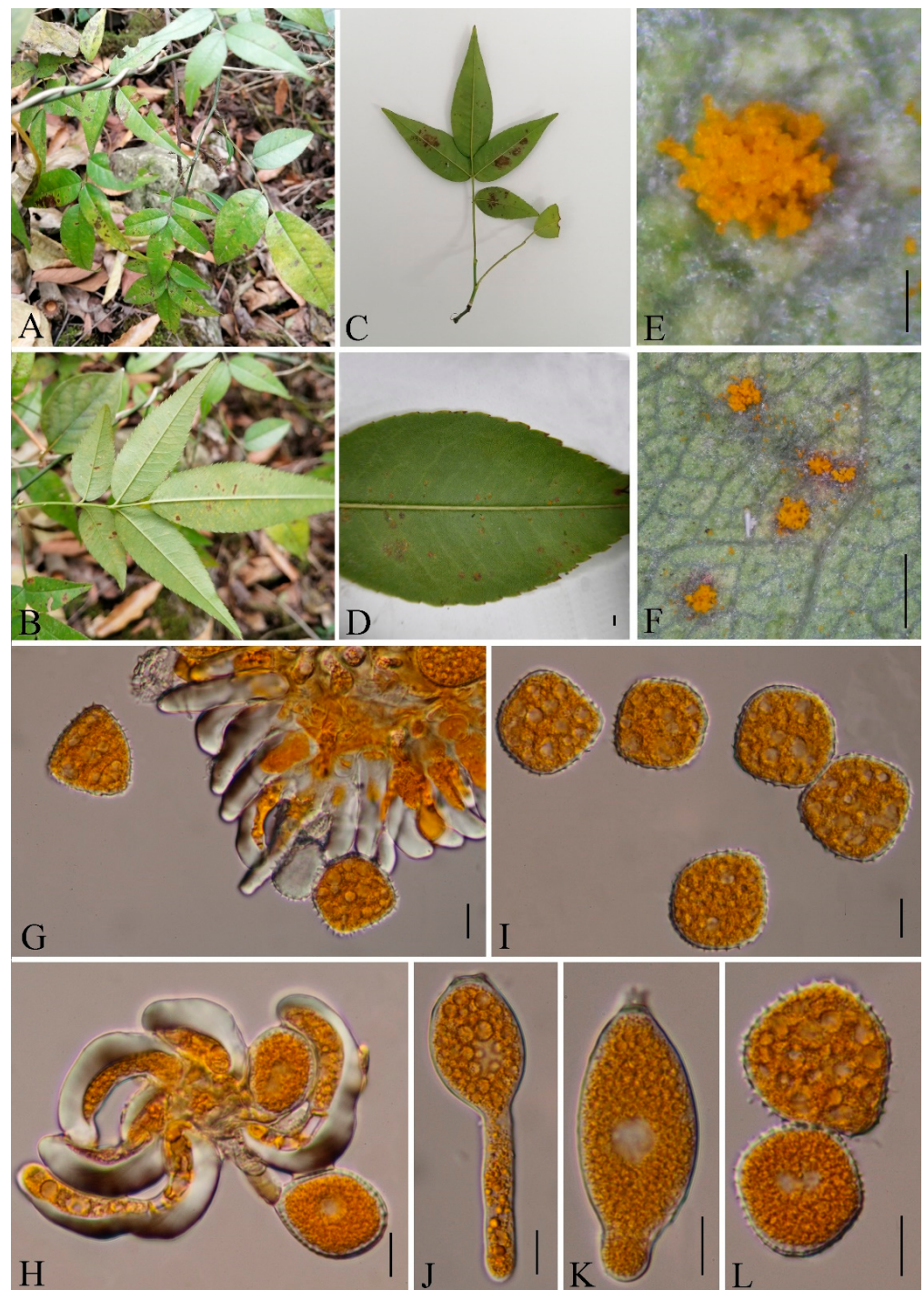


Figure 12. *Phragmidium cymosum* (GMB0108). (A,B) host and its habitat; (C,D) uredinia on the hypophyllous leaf surfaces; (E,F) uredinia; (G–L) globose or obovoid urediniospores with echinulate spines or with abnormal protrusions and paraphyses. Scale bars: (D) = 1 mm; (E) = 0.1 mm; (F) = 0.5 mm; (G–L) = 10 µm.

Phragmidium tormentillae Fuckel, Jb. nassau. Ver. Naturk. 23–24: 46 (1870) [1869–70] Figure 14

Mycobank number: MB177066

Spermogonia were not observed. *Uredinia* hypophyllous, scattered, subepidermal and erumpent becoming pulverulent, 0.5 mm diam. *Urediniospores* circular or obovoid,

pale yellow, $18\text{--}24 \times 14\text{--}20 \mu\text{m}$ (av. = $211 \times 17 \mu\text{m}$, $n = 30$), walls hyaline, echinulate, 1–1.5 mm thick.

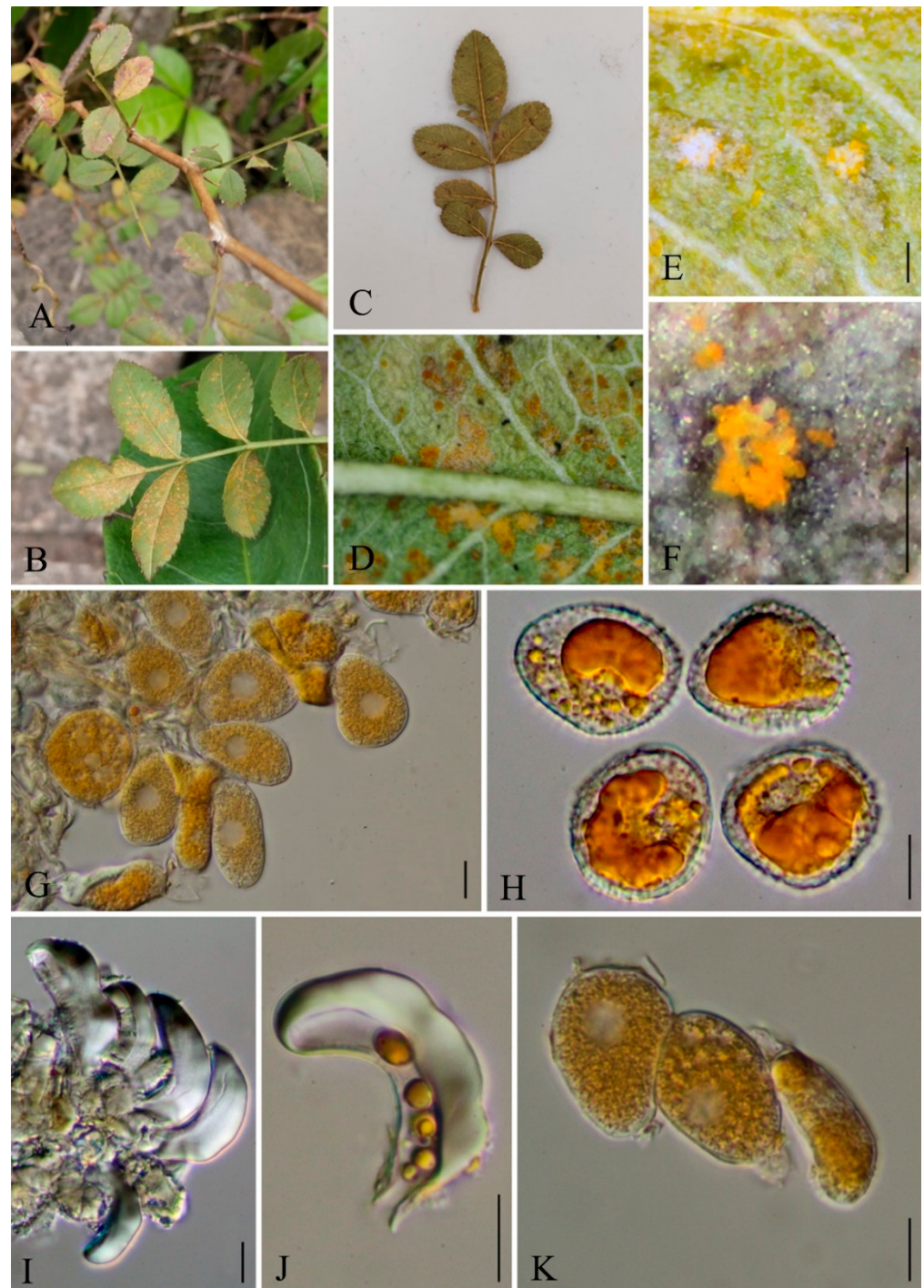


Figure 13. *Phragmidium rosae-roxburghii* (GMB0104). (A,B) host and its habitat; (C,D) uredinia on the hypophyllous leaf surfaces; (E,F) uredinia; (G–K) globose or oval urediniospores with echinulate spines and paraphyses. Scale bars: (E,F) = 0.5 mm, (G–K) = 10 μm .

Materials examined: CHINA, Guizhou Province, Qingzhen City, Xiasi Town ($26^{\circ}27'16.48''$ N, $107^{\circ}20'8.23''$ E) II on *Potentilla simulatrix* Th. Wolf, 22 July 2021 Q.Z. Wu, GMB0114.

Notes: *Potentilla* is documented in most areas of China, which was used as a traditional Chinese medicine for hemostasis and treatment of malaria [112]. The pharmacological

activities of *Potentilla* are mainly related to antioxidant, hypoglycemic, anti-inflammatory, antibacterial, antitumor, and cardiovascular system protective effects [113]. It also has positive effects on hemorrhagic cystitis [114]. Our collection on *Potentilla simulatrix* (GMB0110) is located in the same clade with *P. tormentillae* (Figure 3). Based on the size, shape and wall thickness of the urediniospores, as well as the presence of sparse spines on the surface of the urediniospores, new collection is identified as *P. tormentillae* [60,115,116]. However, no spermatozoa and paraphyses were observed in new collections. *Phragmidium tormentillae* was firstly collected in Norway in 1895 [116]. This is the first record for Chinese mainland.

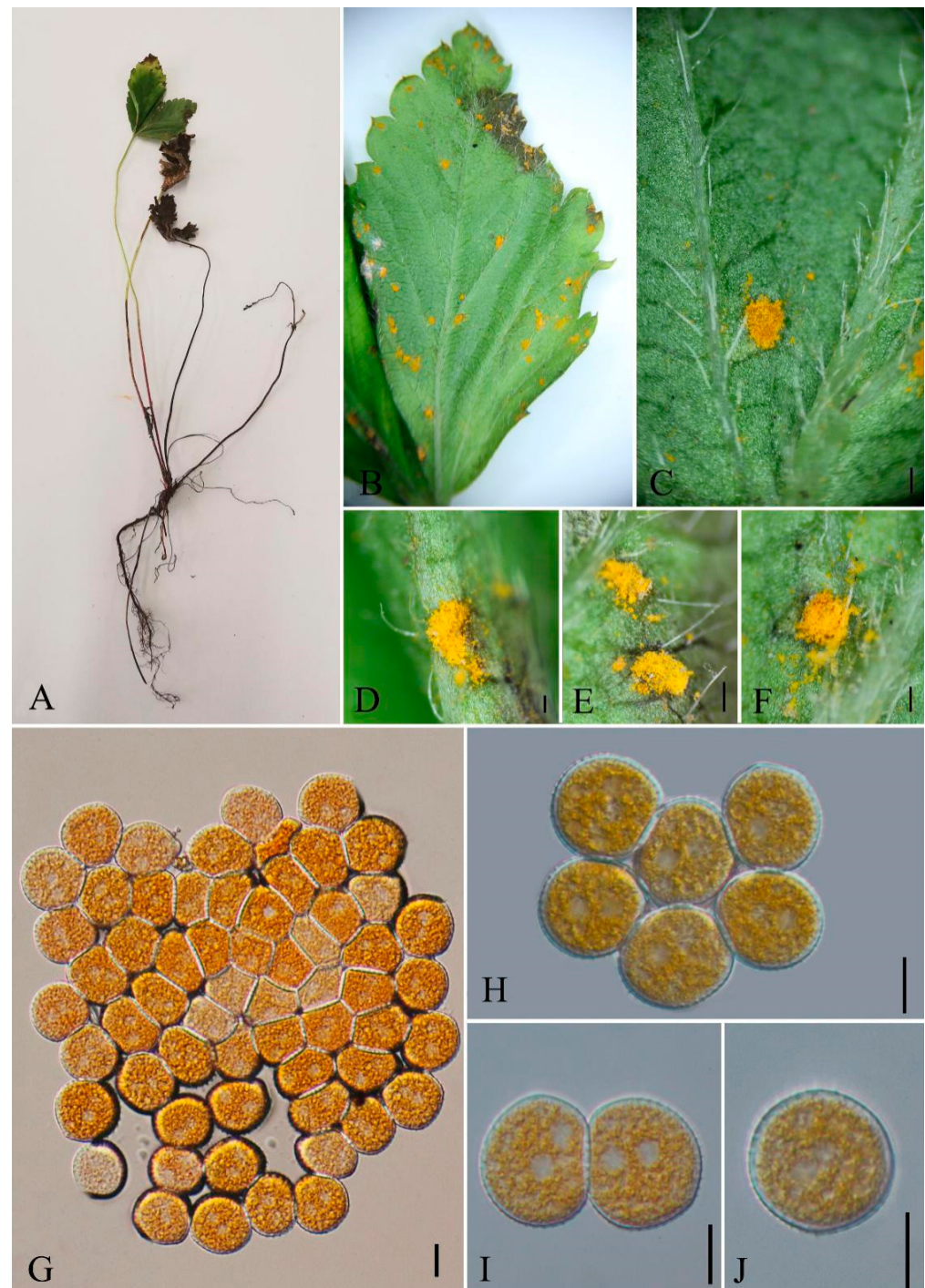


Figure 14. *Phragmidium tormentillae* (GMB0114). (A) host; (B–F) uredinia; (G–J) urediniospores. Scale bars: (C–F) = 0.1 mm; (G–J) = 10 μ m.

4. Discussion

The research on rust diseases in medicinal plants has been relatively less compared to those in economic crops, mainly because in the past, medicinal plants were mostly sourced from the wild, with fewer incidents of rust diseases and thus had not received much attention. However, as the cultivation area of medicinal plants continues to increase, rust diseases have gradually become one of the important diseases affecting the quality and yield of medicinal materials. Attention on rust diseases in medicinal plants have been steadily increasing over time [17].

A total of 79 rust species were found to cause diseases on 76 species of medicinal plants from 33 families in China [16,17]. Rust diseases have become the primary diseases on some important medicinal plants in their primary growth regions, with strong prevalence and large damage areas, such as safflower rust disease (*Puccinia carthami*), Japanese yam rust (*Puccinia dioscoreae*), Radix glehniae rust (*Puccinia phellopteri*), and Rust on bulbous fritillariae ussuriensis (*Uromyces aecidiiformis*) [117–120]. Most researches on rusts on medicinal plants have been focused on the descriptions of symptoms, the incidence scopes and geographic distribution and the rough morphological descriptions of some spores [17]. However, high morphological variations, wide host range and complicated life cycles, identification of rust fungi is very difficult solely based on morphologies or host specificity. Herein, with the aid of morphological and molecular data, ten rust species have been found on medicinal plants collected from Guizhou province, including three new species and six known species. Among them, *Hamasporea rubi-alceifolii*, *Nyssopsora altissima* and *Phragmidium cymosum* were introduced as new to science. *Neophysopella ampelopsidis*, *Phragmidium tormentillae* was firstly introduced in Chinese mainland. *Melampsora laricis-populina*, *Melampsoridium carpini*, and *Nyssopsora koelreuteriae* were documented for the first time in Guizhou province. The accurate identification of rust fungi on medicinal plants will lay the foundation for disease control of medicinal plants.

The use of DNA sequences is becoming more and more important in the identification of rust fungi. Despite early research on rust fungi, the taxonomic system remains perplexing [121]. Distinguishing between individual rust fungi based solely on morphology is challenging [13,122,123]. Because the vast majority of rust fungi cannot be cultured on the artificial medium, pure culture strain cannot be obtained. Therefore, there is not enough DNA sequence available for a large number of rust species for a long time. As DNA extraction techniques continue to improve, valid DNA sequences will become increasingly available [61], and phylogenetic and morphological-based approaches will resolve the taxonomic confusion in rust fungi [11,124]. There were 337 species of rust fungi in 76 genera of 14 families using both morphological and molecular data from 86 natural reserves and national parks in the past five years [11]. Because molecular phylogenetic approaches can be used to connect the telial and aecial stages of rust fungi, they used more additional characters for species recognition [125]. Thus, their studies using DNA-based phylogenetic approach have facilitated precise identification of rust fungi at familial, generic, and species level. These studies can present a significant contribution to the knowledge of rust flora in China, especially those on medicinal plants.

Author Contributions: Conceptualization, Q.L. and L.L.; investigation, Q.W., L.L. and H.H.; morphological examinations, molecular sequencing, and phylogenetic analyses, Q.W. and M.H.; specimen identification, T.L., P.Z. and Q.W.; writing—original draft preparation, Q.W.; writing—review and editing, P.Z. and T.L.; supervision, Q.L. All authors have read and agreed to the published version of the manuscript.

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Informed Consent Statement: Not applicable.

Data Availability Statement: All newly generated sequences were deposited in GenBank (<https://www.ncbi.nlm.nih.gov/genbank/>, accessed on 16 December 2022; Table 1). All new taxa were deposited in MycoBank (<https://www.mycobank.org/>, accessed on 3 January 2023; MycoBank identifiers follow new taxa).

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Conflicts of Interest: The authors declare no conflict of interest.

References

- Kolmer, J.A.; Ordonez, M.E.; Groth, J.V. The rust fungi. In *Encyclopedia of Life Sciences (ELS)*; John Wiley & Sons, Ltd.: Chichester, UK, 2009.
- Rhiney, K.; Guido, Z.; Knudson, C.; Avelino, J.; Bacon, C.M.; Leclerc, G.; Aime, M.C.; Bebb, D.P. Epidemics and the future of coffee production. *Proc. Natl. Acad. Sci. USA* **2021**, *118*, e2023212118. [PubMed]
- Zatybekov, A.; Genievskaya, Y.; Rsaliyev, A.; Maulenbay, A.; Yskakova, G.; Savin, T.; Turuspekoy, Y.; Abugaliev, S. Identification of quantitative trait loci for leaf rust and stem rust seedling resistance in bread wheat using a genome-wide association Study. *Plants* **2021**, *11*, 74. [CrossRef] [PubMed]
- Fernandez, D.; Tisserant, E.; Talhinas, P.; Azinheira, H.; Vieira, A.; Petitot, A.S.; Loureiro, A.; Poulain, J.; Da Silva, C.; Silva Mdo, C.; et al. 454-pyrosequencing of *Coffea arabica* leaves infected by the rust fungus *Hemileia vastatrix* reveals in planta-expressed pathogen-secreted proteins and plant functions in a late compatible plant-rust interaction. *Mol. Plant Pathol.* **2012**, *13*, 17–37.
- Chen, Z. Investigation of coffee leaf rust (*Hemileia vastatrix*) physiological races in China. *Mycosystema* **1998**, *17*, 21–28.
- Li, L.; Várzea, V.M.P.; Xia, Q.; Xiang, W.; Tang, T.; Zhu, M.; He, C.; Pereira, A.P.; Silva, M.C.; Wu, W.; et al. First report of *Hemileia vastatrix* (Coffee leaf rust) physiological races emergent in coffee germplasm collections in the coffee-cropping regions of China. *Plant Dis.* **2021**, *105*, 4162. [CrossRef] [PubMed]
- Cummins, G.B.; Hiratsuka, Y. *Illustrated Genera of Rust Fungi*, 2nd ed.; American Phytopathological Society (APS Press): St Paul, MN, USA, 1983.
- Aime, M.C. Toward resolving family-level relationships in rust fungi (*Uredinales*). *Mycoscience* **2006**, *47*, 112122.
- Spatafora, J.W.; Aime, M.C.; Grigoriev, I.V.; Martin, F.; Stajich, J.E.; Blackwell, M. The fungal tree of life: From molecular systematics to genome-scale phylogenies. In *The Fungal Kingdom*; Wiley: Hoboken, NJ, USA, 2017; pp. 1–34.
- Aime, M.C.; Bell, C.D.; Wilson, A.W. Deconstructing the evolutionary complexity between rust fungi (*Pucciniales*) and their plant hosts. *Stud. Mycol.* **2018**, *89*, 143–152. [CrossRef]
- Zhao, P.; Zhang, Z.F.; Hu, D.M.; Tsui, K.M.; Qi, X.H.; Phurba, D.; Gafforov, Y.; Cai, L. Contribution to rust flora in China I, tremendous diversity from natural reserves and parks. *Fungal Divers.* **2021**, *110*, 1–58.
- Zhuang, J.Y.; Wei, S.X.; Wang, Y.C. *Flora Fungorum Sinicorum*; Science Press: Beijing, China, 1998; Volume 10.
- McTaggart, A.R.; Shivas, R.G.; van der Nest, M.A.; Roux, J.; Wingfield, B.D.; Wingfield, M.J. Host jumps shaped the diversity of extant rust fungi (*Pucciniales*). *New Phytol.* **2016**, *209*, 1149–1158. [CrossRef]
- Laundon, G.F.A. *Coleosporium* rust new to New Zealand. *Plant Dis. Rep.* **1978**, *62*, 796–797.
- Demers, J.E.; Liu, M.; Hambleton, S.; Castlebury, L.A. Rust fungi on *Panicum*. *Mycologia* **2017**, *109*, 1–17. [CrossRef] [PubMed]
- Delate, K.; McKern, A.; Burcham, R. Evaluation of organic soybean rust treatments for organic production neely-kinyon trial, 2005. *Iowa State Univ. Res. Demonstr. Farms Prog. Rep.* **2006**, *2005*, 1.
- Yu, Z.L.; Yang, J.; Lei, M.Y.; Quan, J.; Yang, T.J.; Yang, C.Q. Status quo of and challenges for research on rust disease in medicinal plants. *Zhongguo Zhong Yao Za Zhi-China J. Chin. Mater. Medica* **2021**, *46*, 3566–3576. (In Chinese)
- Zou, T.C. Research on sustainable utilisation of medicinal plant germplasm resources in Guizhou. *J. Pharm. Pract.* **2000**, *5*, 316–323. (In Chinese)
- Xia, Z.M.; Zhang, Z.M. Preliminary report on the survey of pest and weed species and damage of medicinal plants in Guizhou Province. *Plant Dr.* **2003**, *16*, 26–28. (In Chinese)
- Xia, Z.P.; Hu, X.P.; Wei, C.S. The occurrence and control of pepper rust in the top altar of Guizhou Province. *Mod. Agric. Technol.* **2018**, *16*, 117. (In Chinese)
- Huang, J.; Zang, Z.G.; Huang, R.M.; Song, B.A. Survey and research on diseases of key medicinal plants in Guizhou Province. *Agrochemicals* **2002**, *10*, 6–9.
- Wei, G.R.; Wang, X.J. Purification and storage of *Puccinia striiformis* f. sp. *Tritici*. *J. Henan Agric. Sci.* **2011**, *40*, 90–92.
- Hiratsuka, Y.; Hiratsuka, N. Morphology of spermogonia and taxonomy of rust fungi. *Rep. Tottori Mycol. Inst.* **1980**, *18*, 257–268.

24. Vilgalys, R.; Hester, M. Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *Cryptococcus* species. *J. Bacteriol.* **1990**, *172*, 4238–4246. [\[CrossRef\]](#)
25. Beenken, L.; Lutz, M.; Scholler, M. DNA barcoding and phylogenetic analyses of the genus *Coleosporium* (*Pucciniales*) reveal that the North American goldenrod rust *C. solidaginis* is a neomycete on introduced and native *Solidago* species in Europe. *Mycol. Prog.* **2017**, *16*, 1073–1085. [\[CrossRef\]](#)
26. Colmán, A.; Evans, H.; Salcedo, S.; Braun, U.; Belachew–Bekele, K.; Barreto, R. A fungus–eat–fungus world: Digitopodium, with particular reference to mycoparasites of the coffee leaf rust, *Hemileia vastatrix*. *IMA Fungus* **2021**, *12*, 1. [\[CrossRef\]](#) [\[PubMed\]](#)
27. Katoh, K.; Rozewicki, J.; Yamada, K. MAFFT online service: Multiple sequence alignment, interactive sequence choice and visualization. *Brief. Bioinform.* **2019**, *20*, 1160–1166. [\[CrossRef\]](#)
28. Capella–Gutiérrez, S.; Silla–Martínez, J.M.; Gabaldón, T. TrimAl: A tool for automated alignment trimming in large–scale phylogenetic analyses. *Bioinformatics* **2009**, *25*, 1972–1973. [\[CrossRef\]](#) [\[PubMed\]](#)
29. Glez–Peña, D.; Gómez–Blanco, D.; Reboiro–Jato, M.; Fdez–Riverola, F.; Posada, D. ALTER: Program–oriented conversion of DNA and protein alignments. *Nucleic Acids Res.* **2010**, *38*, W14–W18. [\[CrossRef\]](#) [\[PubMed\]](#)
30. Stamatakis, A. RAxML–VI–HPC: Maximum likelihood–based phylogenetic analyses with thousands of taxa and mixed models. *Bioinformatics* **2006**, *22*, 2688–2690. [\[CrossRef\]](#)
31. Miller, M.A.; Pfeiffer, W.; Schwartz, T. Creating the CIPRES Science Gateway for inference of large phylogenetic trees. In Proceedings of the 2010 Gateway Computing Environments Workshop (GCE), New Orleans, LA, USA, 14 November 2010; Volume 2010, pp. 1–8.
32. Huelsenbeck, J.P.; Ronquist, F. MRBAYES: Bayesian inference of phylogenetic trees. *Bioinformatics* **2001**, *17*, 754–755. [\[CrossRef\]](#)
33. Posada, D.; Crandall, K.A. MODELTEST: Testing the model of DNA substitution. *Bioinformatics* **1998**, *14*, 817–818. [\[CrossRef\]](#)
34. Nylander, J.J.E.B.C. *MrModeltest v2. Program Distributed by the Author, Version 2*; Uppsala University: Uppsala, Sweden, 2004.
35. Rambaut, A.; Drummond, A. *FigTree: Tree Figure Drawing Tool, v1. 4.2*; University of Edinburgh: Edinburgh, UK, 2012.
36. Feau, N.; Vialle, A.; Allaire, M.; Maier, W.; Hamelin, R.C. DNA barcoding in the rust genus *Chrysomyxa* and its implications for the phylogeny of the genus. *Mycologia* **2011**, *103*, 1250–1266. [\[CrossRef\]](#)
37. Zhao, P.; Qi, X.H.; Crous, P.W.; Duan, W.J.; Cai, L. *Gymnosporangium* species on *Malus*: Species delineation, diversity and host alternation. *Persoonia* **2020**, *45*, 68–100. [\[CrossRef\]](#)
38. Shen, Y.M.; Chung, W.H.; Huang, T.C.; Rodeva, R.; Hung, T.H. Unveiling *Gymnosporangium corniforme*, *G. unicornae*, and *G. niitakayamense* sp. nov. in Taiwan. *Mycoscience* **2017**, *59*, 218–228. [\[CrossRef\]](#)
39. Cao, B.; Tao, S.; Tian, C.; Liang, Y. *Coleopuccinia* in China and its relationship to *Gymnosporangium*. *Phytotaxa* **2018**, *347*, 235–242. [\[CrossRef\]](#)
40. Aime, M.C.; McTaggart, A.R. A higher–rank classification for rust fungi, with notes on genera. *Fungal Syst. Evol.* **2021**, *7*, 21–47. [\[CrossRef\]](#)
41. McKenzie, E.H.C.; Padamsee, M.; Dick, M. First report of rust on *Alnus* in New Zealand is *Melampsoridium betulinum*, not *M. hiratsukanum*. *Plant Pathol. Quar.* **2013**, *3*, 59–65. [\[CrossRef\]](#)
42. Blomquist, C.L.; Scheck, H.J.; Haynes, J.; Woods, P.W.; Bischoff, J. First published report of rust on white alder caused by *Melampsoridium hiratsukanum* in the United States. *Plant Dis.* **2014**, *98*, 155. [\[CrossRef\]](#)
43. Bubner, B.; Buchheit, R.; Friedrich, F.; Kummer, V.; Scholler, M. Species identification of European forest pathogens of the genus *Milesina* (*Pucciniales*) using urediniospore morphology and molecular barcoding including *M. woodwardiana* sp. nov. *MycKeys* **2019**, *48*, 1–40. [\[CrossRef\]](#)
44. Vialle, A.; Feau, N.; Frey, P.; Bernier, L.; Hamelin, R.C. Phylogenetic species recognition reveals host–specific lineages among poplar rust fungi. *Mol. Phylogenetics Evol.* **2013**, *66*, 628–644. [\[CrossRef\]](#)
45. Zhao, P.; Wang, Q.H.; Tian, C.M.; Wang, Q.; Kakishima, M. *Melampsora salicis–bakko*, a new species on willows in Japan evidenced by morphological and molecular phylogenetic analyses. *Mycol. Prog.* **2016**, *15*, 13. [\[CrossRef\]](#)
46. Zhao, P.; Wang, Q.H.; Tian, C.M.; Kakishima, M. Integrating a numerical taxonomic method and molecular phylogeny for species delimitation of *Melampsora* species (*Melampsoraceae*, *Pucciniales*) on willows in China. *PLoS ONE* **2015**, *10*, e0144883. [\[CrossRef\]](#)
47. Deadman, M.L.; Al–Sa’di, A.M.; Al–Maqbali, Y.M.; Al–Jahdhami, S.; Patzelt, A.; Aime, M.C. First report of the rust *Melampsora euphorbiae* on *Euphorbia heterophylla* in Oman. *J. Plant Pathol.* **2006**, *88*, 229.
48. Wang, L.L.; Li, K.M.; Liu, Y.; Tian, C.M. *Melampsora salicis–michelsonii* sp. nov. on *Salix michelsonii* and *Melampsora salicis–cavaleriei* on *Salix serrulatifolia* from China. *Phytotaxa* **2020**, *435*, 280–292. [\[CrossRef\]](#)
49. Zheng, W.; Newcombe, G.; Hu, D.; Cao, Z.; Yu, Z.; Peng, Z. The first record of a north American poplar leaf rust fungus, *Melampsora medusae*, in China. *Forests* **2019**, *10*, 182. [\[CrossRef\]](#)
50. Ali, B.; Sohail, Y.; Toome–Heller, M.; Mumtaz, A.S. *Melampsora pakistanica* sp. nov.; a new rust fungus on *Euphorbia helioscopia* (Sun spurge) from Pakistan. *Mycol. Prog.* **2016**, *15*, 1285–1292. [\[CrossRef\]](#)
51. Vialle, A.; Feau, N.; Allaire, M.; Didukh, M.; Martin, F.; Moncalvo, J.M.; Hamelin, R.C. Evaluation of mitochondrial genes as DNA barcode for *Basidiomycota*. *Mol. Ecol. Resour.* **2009**, *9*, 99–113. [\[CrossRef\]](#) [\[PubMed\]](#)
52. Pei, M.H.; Bayon, C.; Ruiz, C. Phylogenetic relationships in some *Melampsora* rusts on *Salicaceae* assessed using rDNA sequence information. *Mycol. Res.* **2005**, *109*, 401–409. [\[CrossRef\]](#)
53. Chatasiri, S.; Ono, Y. Phylogeny and taxonomy of the Asian grapevine leaf rust fungus, *Phakopsora euvitis*, and its allies (*Uredinales*). *Mycoscience* **2008**, *49*, 66–74. [\[CrossRef\]](#)

54. Pota, S.; Chatasiri, S.; Unartngam, J.; Yamaoka, Y.; Hosaka, K.; Ono, Y. Taxonomic identity of a *Phakopsora* fungus causing the grapevine leaf rust disease in Southeast Asia and Australasia. *Mycoscience* **2014**, *56*, 198–204. [\[CrossRef\]](#)
55. Beenken, L. *Pucciniales* on *Annona* (Annonaceae) with special focus on the genus *Phakopsora*. *Mycol. Prog.* **2014**, *13*, 791–809. [\[CrossRef\]](#)
56. Maier, W.; McTaggart, A.R.; Roux, J.; Wingfield, M.J. *Phakopsora myrtacearum* sp. nov.; a newly described rust (*Pucciniales*) on eucalypts in eastern and southern Africa. *Plant Pathol.* **2016**, *65*, 189–195. [\[CrossRef\]](#)
57. Dixon, L.J.; Castlebury, L.A.; Aime, M.C.; Glynn, N.C.; Comstock, J.C. Phylogenetic relationships of sugarcane rust fungi. *Mycol. Prog.* **2010**, *9*, 459–468. [\[CrossRef\]](#)
58. Alaei, H.; De Backer, M.; Nuytinck, J.; Maes, M.; Höfte, M.; Heungens, K. Phylogenetic relationships of *Puccinia horiana* and other rust pathogens of *Chrysanthemum × morifolium* based on rDNA ITS sequence analysis. *Mycol. Res.* **2009**, *113*, 668–683. [\[CrossRef\]](#)
59. Liu, Y.; Cao, B.; Tao, S.; Tian, C.; Liang, Y. *Phragmidium* species parasitizing species of *Rosaceae* in Tibet, China, with descriptions of three new species. *Mycol. Prog.* **2018**, *17*, 967–988. [\[CrossRef\]](#)
60. Yun, H.Y.; Minnis, A.M.; Kim, Y.H.; Castlebury, L.A.; Aime, M.C. The rust genus *Frommeëlla* revisited: A later synonym of *Phragmidium* after all. *Mycologia* **2011**, *103*, 1451–1463. [\[CrossRef\]](#) [\[PubMed\]](#)
61. Maier, W.; Begerow, D.; Weiß, M.; Oberwinkler, F. Phylogeny of the rust fungi: An approach using nuclear large subunit ribosomal DNA sequences. *Can. J. Bot.* **2003**, *81*, 12–23. [\[CrossRef\]](#)
62. Yang, T.; Chang, W.; Cao, B.; Tian, C.; Zhao, L.; Liang, Y. Two new *Phragmidium* species identified on *Rosa* plants native to China. *Phytotaxa* **2015**, *217*, 182–190. [\[CrossRef\]](#)
63. Liu, Y.; Liang, Y.M.; Ono, Y. Taxonomic revision of species of *Kuehneola* and *Phragmidium* on *Rosa*, including two new species from China. *Mycologia* **2020**, *112*, 742–752. [\[CrossRef\]](#) [\[PubMed\]](#)
64. Deadman, M.L.; Al Sadi, A.M.; Al Maqbali, Y.M.; Farr, D.F.; Aime, M.C. Additions to the rust fungi (*Pucciniales*) from northern Oman. *Sydowia* **2011**, *63*, 155–168.
65. Sun, J.E.; Zhang, Q.; Luo, W.M.; Yang, Y.Q.; An, H.M.; Wang, Y. Four new *Phragmidium* (*Phragmidiaceae*, *Pucciniomycetes*) species from *Rosaceae* plants in Guizhou Province of China. *Mycologia* **2022**, *93*, 193–213. [\[CrossRef\]](#)
66. Wilson, A.W.; Aime, M.C. The rose rust fungus, *Phragmidium tuberculatum*, is widespread in the Americas: First reports from California, Oregon, Massachusetts, and Honduras. *Plant Dis.* **2014**, *98*, 1581. [\[CrossRef\]](#)
67. Liang, Y.M.; Tian, C.M.; Kakishima, M. Phylogenetic relationships on 14 morphologically similar species of *Pucciniastrum* in Japan based on rDNA sequence data. *Mycoscience* **2006**, *47*, 137–144. [\[CrossRef\]](#)
68. Beenken, L.; Zoller, S.; Berndt, R. Rust fungi on *Annonaceae* II: The genus *Dasyscypha* Berk. MA Curtis. *Mycologia* **2012**, *104*, 659–681. [\[CrossRef\]](#) [\[PubMed\]](#)
69. McTaggart, A.R.; Doungsa-Ard, C.; Geering, A.D.W.; Aime, M.C.; Shivas, R.G. A co-evolutionary relationship exists between *Endoraecium* (*Pucciniales*) and its *Acacia* hosts in Australia. *Persoonia* **2015**, *35*, 50–62. [\[CrossRef\]](#) [\[PubMed\]](#)
70. Beenken, L.; Wood, A.R. *Puccorchidium* and *Sphenorchidium*, two new genera of *Pucciniales* on *Annonaceae* related to *Puccinia psidii* and the genus *Dasyscypha*. *Mycol. Prog.* **2015**, *14*, 49. [\[CrossRef\]](#)
71. Yang, Y.; Yang, Y.; Yu, Y.; Bi, C. First report of rust disease on *Koeleruteria bipinnata* caused by *Nyssopsora koeleruteriae* in China. *Plant Dis.* **2016**, *100*, 1014. [\[CrossRef\]](#)
72. Baiswar, P.; Ngachan, S.V.; Chandra, S. Identification of *Nyssopsora thwaitesii* on *Schefflera* in northeast India. *Australas. Plant Dis. Notes* **2014**, *9*, 124. [\[CrossRef\]](#)
73. Cummins, G.B. *Uredinales* of continental China collected by SY Cheo. I. *Mycologia* **1950**, *42*, 779–797. [\[CrossRef\]](#)
74. Monoson, H.L. The species of *Hamaspora*. *Mycopathol. Mycol. Appl.* **1969**, *37*, 263–272. [\[CrossRef\]](#)
75. Chen, Z.C. Additional materials for a rust-flora of Taiwan. *Taiwania* **1980**, *25*, 185–186.
76. Zhuang, J.Y.; Wei, S.X.; Wang, Y.C. *Flora Fungorum Sinicorum*; Science Press: Beijing, China, 2012; Volume 41.
77. Yao, Z.S.; Yang, W.L. The medicinal plants of the genus *Rubus* in Jiangxi Province and suggestions for their utilization. *J. Chin. Med. Mater.* **1995**, *11*, 551–554. (In Chinese)
78. Ye, H.Z.; Hong, Z.F.; Wang, Y.H.; Chen, Y.H. Study of the therapeutic effect of *Rubus alceaefolius* on experimental liver injury. *Chin. Arch. Tradit. Chin. Med.* **2005**, *23*, 829–831. (In Chinese)
79. Niero, R.; Filho, V.C. Therapeutic potential and chemical composition of plants from the genus *Rubus*: A mini review of the last 10 years. *Nat. Prod. Comm.* **2008**, *3*, 437–444. [\[CrossRef\]](#)
80. Zhao, H.Y.; Zhao, J.Y.; Liu, Y.; Zheng, Y.Q.; Wu, J.; Hong, Z.F. Effect of total alkaloids of *Rubus alceaefolius* on oxidative stress in rats with non-alcoholic fatty liver disease. *China J. Chin. Med. Mater. Medica* **2011**, *36*, 2383–2387. (In Chinese)
81. Li, J.; Du, L.F.; He, Y.; Yang, L.; Li, Y.Y.; Wang, Y.F.; Gao, X.M. Chemical constituents and biological activities of plants from the genus *Rubus*. *Chem. Biodivers.* **2015**, *12*, 1809–1847. [\[CrossRef\]](#)
82. Kuprevich, V.J.; Tranzhel, V.G. *Cryptogamic Plants of the USSR, IV, Fungi (1), Rust Fungi No. 1. Family Melampsolaceae*; Translated from Russian (1970); Israel Program for Scientific Translations: Jerusalem, Israel, 1957; pp. 362–379.
83. Spiers, A.G.; Hopcroft, D.H. Comparative studies of the poplar rusts *Melampsora medusae*, *M. larici-populina* and their interspecific hybrid *M. medusae-populina*. *Mycol. Res.* **1994**, *98*, 889–903. [\[CrossRef\]](#)
84. Yue, Y.J. *Populus lasiocarpa* leaves for diabetic foot ulcers in 5 cases of observation and care. *Diabetes New World* **2015**, *35*, 142. (In Chinese)

85. Yue, Y.J. Care of one case of severe pressure ulcer treated with leaf dressing of *Populus lasiocarpa*. *Chin. J. Misdiagn.* **2008**, *8*, 8003. (In Chinese)
86. Hiratsuka, N.; Sato, S.; Katsuya, K.; Zhao, M.; Hiratsuka, Y.; Kaneko, S.; Ono, Y.; Sato, T.; Harada, Y.; Hiratsuka, T. *The Rust Flora of Japan*; Tsukuba Shuppankai: Tsukuba-shi, Japan, 1992.
87. Spaulding, P. *Foreign Diseases of Forest Trees of the World*; US Department of Agriculture: Washington, DC, USA, 1961; Volume 197.
88. Zhuang, J.Y.; Wei, S.X.; Wang, Y.C. *Flora Fungorum Sinicorum*; Science Press: Beijing, China, 2005; Volume 25.
89. Son, Y.K.; Yoon, S.R.; Bang, W.Y.; Bae, C.H.; Yeo, J.H.; Yeo, R.; An, J.; Song, J.; Kim, O.Y. *Carpinus turczaninowii* extract modulates arterial inflammatory response: A potential therapeutic use for atherosclerosis. *Nutr. Res. Pract.* **2019**, *13*, 302. [[CrossRef](#)] [[PubMed](#)]
90. Cieckiewicz, E.; Angenot, L.; Gras, T.; Kiss, R.; Frederich, M. Potential anticancer activity of young *Carpinus betulus* leaves. *Phytomedicine* **2012**, *19*, 278–283. [[CrossRef](#)] [[PubMed](#)]
91. Qianqian, S.; Fang, X.; Zhu, Z.; Xiao, W.; Wang, Z.; Ding, G.; Zhao, L.; Li, Y.; Yu, P.; Ding, Z.; et al. Seasonal variation of pheophorbide a and flavonoid in different organs of two *Carpinus* species and its correlation with immunosuppressive activity. *Vitr. Cell. Dev. Biol.-Anim.* **2016**, *52*, 654–661.
92. Majewski, T. *Flora polska*. In *Grzyby (Mycota) 9: Basidiomycetes, Uredinales I*; PWN: Warszawa, Poland, 1977.
93. Zhuang, W.Y.; Guo, Y.L.; Wen, H.A.; Zhang, X.Q.; Lin, G.; Ning, Z.; Zhuang, J.Y.; Wei, S.X. Fungal flora of the Daba Mountains: *Uredinales*. *Mycotaxon* **1997**, *61*, 49–79.
94. Zhuang, J.Y.; Wei, S.X.; Wang, Y.C. *Flora Fungorum Sinicorum*; Science Press: Beijing, China, 2021; Volume 62.
95. Xi, Z.H.; Liu, X.J.; Mu, M.J.; Liang, Q.Y. Studies on the chemical composition of *Ampelopsis sinica* root. *Chin. Tradit. Herb. Drugs* **1992**, *23*, 611. (In Chinese)
96. Pang, R.; Tao, J.; Zhang, S.-L.; Chen, K.-L.; Zhao, L.; Yue, X.; Wang, Y.-F.; Ye, P.; Hua, D.; Zhu, Y.; et al. Corrigendum to “Ethanol Extract from *Ampelopsis sinica* Root Exerts Anti-Hepatitis B Virus Activity via Inhibition of p53 Pathway In Vitro”. *Evid.-Based Complement. Altern. Med.* **2015**, *2015*, 603232-1. [[CrossRef](#)] [[PubMed](#)]
97. Wang, J.Z.; Huang, B.S.; Cao, Y.; Chen, K.L.; Li, J. Anti-hepatoma activities of ethyl acetate extract from *Ampelopsis sinica* root. *Oncol. Rep.* **2017**, *37*, 2227–2236. [[CrossRef](#)] [[PubMed](#)]
98. Ono, Y. Taxonomy of the *Phakopsora ampelopsidis* species complex on vitaceous hosts in Asia including a new species, *P. euvitis*. *Mycologia* **2000**, *92*, 154–173. [[CrossRef](#)]
99. Monoson, H. The species of *Triphragmium*, *Nyssopsora* and *Triphragmiopsis*. *Mycopathologia* **1974**, *52*, 115–131. [[CrossRef](#)]
100. Ye, S.Y.; Lu, J.; Luo, S.M. Biological characteristics of *Ailanthus altissima* and its exploitation. *Beijing Agric.* **2013**, *33*, 79–80. (In Chinese)
101. Tusculum, Li, T.; Wang, Y.L.; Zhao, M. Study on the chemical composition of the roots of *Ailanthus altissima*. *Chin. Tradit. Herb. Drugs* **2015**, *46*, 1426–1430. (In Chinese)
102. Peng, S.; Yi, Z.; Liu, M. Ailanthone: A new potential drug for castration-resistant prostate cancer. *Chin. J. Cancer* **2017**, *36*, 25. [[CrossRef](#)]
103. Bao, X.Z.; Yv, X.C.; Niu, Z.S.; Fu, L.; Li, Q. Effect of Ailanthinone on endogenous AR of prostate cancer 22RV1 cells. *Northwest Pharm. J.* **2022**, *37*, 62–66. (In Chinese)
104. Li, X.; Jiang, W.B.; Weng, M.L. Landscape characteristics of *Koeleruteria bipinnata* and its development and utilization. *Chin. Agric. Sci. Bull.* **2009**, *25*, 141–146. (In Chinese)
105. Cao, L.M.; Tuo, J.Y.; He, J.; Ten, T.; Cao, M. nalysis of fatty acid and amino acid composition of seed kernels of *Koeleruteria bipinnata*. *China Oils Fats* **2020**, *45*, 134–137. (In Chinese)
106. Tranzschel, W. Contribution à l'étude du genre *Triphragmium* auct. (*Triphragmium* Link, *Triphragmiopsis* Naumov, *Nyssopsora* Arthur). *J. Soc. Bot. Russ.* **1925**, *8*, 132.
107. Zhuang, J.Y. A general survey of rust flora in the Mt. Namjagbarwa region. *Mauntain Res.* **1984**, *2*, 198–203.
108. Zhang, L.W.; ZHhao, J.N.; Xu, J.W. Study on the anti-inflammatory effect of aqueous extracts of *Rosa cymosa* roots. *J. Jilin Agric. Univ.* **2010**, *32*, 37–39. (In Chinese)
109. Ouyang, L.M.; Huang, S.C.; Huang, X.L.; Huang, Y. Pharmacodynamic study on the anti-inflammatory and analgesic, hemostatic and intestinal motility inhibiting effects of aqueous extracts of roots and stems of *Rosa minor*. *Mod. Chin. Med.* **2012**, *14*, 4–8. (In Chinese)
110. Fan, X.; Bai, L.; Lin, L.; Liao, D.; Gong, Y.; Liu, X.; Wang, Z.; Li, C. Studies on the chemical constituents and quality evaluation of *Rosa cymosa* Tratt. root. *J. Sep. Sci.* **2020**, *43*, 4379–4389. [[CrossRef](#)] [[PubMed](#)]
111. Chen, F.; Wang, L.; Jin, F.; Li, L.; Wang, T.; Gao, M.; Yang, X. Neuroprotection of Kaji-Ichigoside F1 via the BDNF/Akt/mTOR Signaling Pathways against NMDA-Induced Neurotoxicity. *Int. J. Mol. Sci.* **2022**, *23*, 16150. [[CrossRef](#)] [[PubMed](#)]
112. Yu, D.X. *Flora of China*; Science Press: Beijing, China, 1990; Volume 37.
113. Wu, J.; Zhang, Z.Q.; Yu, H.H.; Huang, F.B.; Chen, Z.L.; Chu, L.L.; Li, B.; Wang, W. Research progress on chemical constituents and pharmacological activities of *Potentilla*. *Zhongguo Zhongyao Zazhi-China J. Chin. Mater. Med.* **2022**, *47*, 1509–1538. (In Chinese)
114. Juszczak, K.; Adamowicz, J.; Zapała, Ł.; Kluz, T.; Adamczyk, P.; Wdowiak, A.; Bojar, I.; Misiek, M.; Grzybowska, M.E.; Stangel-Wójcikiewicz, K.; et al. *Potentilla chinensis* aqueous extract attenuates cyclophosphamide-induced hemorrhagic cystitis in rat model. *Sci. Rep.* **2022**, *12*, 13076. [[CrossRef](#)]

115. McKenzie, E.H.C.; Dingley, J.M. New plant disease records in New Zealand: Miscellaneous fungal pathogens III. *N. Z. J. Bot.* **1996**, *34*, 263–272. [[CrossRef](#)]
116. Lye, K.A. Norwegian rust fungi-The genus *Frommeëlla*. *Agarica* **2012**, *35*, 59–66.
117. Zhao, L.S.; Tian, X.J.; Zang, S.X.; Li, Y.Q.; Zhang, Z.M. Discovery of pycnospore stage of *Puccinia phellopteri* and study on rust of *glehnia littoralis*. *Acta Phytopathol. Sin.* **1996**, *26*, 49.
118. Fu, J.F.; Zhou, R.J.; Yan, X.R.J. Occurrence of the rust disease on *Dioscorea nipponica* Makino in Liaoning Province. *Plant Prot.* **2004**, *30*, 90–92.
119. Zhu, R.J.; Zheng, C.J. Characteristics and control measures of rust disease of *Fritillaria ussuriensis*. *China Plant Prot.* **2008**, *28*, 32–33. (In Chinese)
120. Liao, A.P. Study on safflower rust caused by *Puccinia carthami* (Hutz) Corda. Master's Thesis, Sichuan Agricultural University, Ya'an, China, 2008.
121. Schultze-Kraft, R.; Cook, B.; Ciprián, A. Clearing confusion in *Stylosanthes* taxonomy. 2. *S. macrocephala* M.B. Ferreira & Sousa Costa vs. *S. capitata* Vogel and *S. bracteata* Vogel. *Trop. Grassl. –Forrajes Trop.* **2020**, *8*, 250–262.
122. Uchida, J.; Zhong, S.; Killgore, E. First report of a rust disease on ohia caused by *Puccinia psidii* in Hawaii. *Plant Dis.* **2006**, *90*, 524. [[CrossRef](#)]
123. Ono, Y. Life cycle and nuclear behavior in three rust fungi (*Uredinales*). *Mycoscience* **2002**, *43*, 0037–0045. [[CrossRef](#)]
124. Piškur, B.; Jurc, D. New findings of rusts (*Pucciniales*) on trees and other plants in Slovenia. In Proceedings of the Zbornik Predavanj in Referatov, 13. Slovenskega Posvetovanja o Varstvu Rastlin z Mednarodno Udeležbo, Rimske Toplice, Slovenia, 7–8 March 2017; Volume 2017, pp. 222–229.
125. Zhao, P.; Li, Y.; Li, Y.; Liu, F.; Liang, J.; Zhou, X.; Cai, L. Applying early divergent characters in higher rank taxonomy of *Melampsorineae* (*Basidiomycota*, *Pucciniales*). *Mycology* **2022**, *14*, 11–36. [[CrossRef](#)]

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