

Supplementary Tables.

Table S1. Strains used in this study.

Strain	Genotype	Source
<i>AkuB</i> ^{KU80}	<i>KU80Δ</i>	Da Silva Ferreira et al., 2006 [8]
Cyp51A G448S	<i>KU80Δ; cyp51A/G448S</i>	This study
Cyp51B G457S	<i>KU80Δ; cyp51B/G457S</i>	This study
Hmg1 F262del	<i>KU80Δ; hmg1/F262del</i>	This study
Cyp51B G457S	<i>KU80Δ; cyp51B/G457S;</i>	This study
/Hmg1 F262del	<i>hmg1/F262del</i>	
Cyp51B G457S/	<i>KU80Δ; cyp51B/G457S;</i>	This study
Hmg1 F262del/	<i>hmg1/F262del;</i>	
Cyp51A G448S	<i>cyp51A/G448S</i>	

Table S2. Gene amplification primers used in this study

Target gene	Primer name	Primer sequence (5' → 3')	Final construct
<i>cyp51A</i>	Cyp51A F	GGGCTGGAGATACTATGGCTTTCA	Mutated Cyp51A
	Cyp51A R	CAGGTTTTTCGCACGAGCTTC	G448S
<i>cyp51B</i>	Cyp51B F	ATGGGTCTCATCGCGTTTCATT	Mutated Cyp51B
	Cyp51B R	TCAGGCTTTGGTAGCGGACTC	G457S
<i>hmg1</i>	Hmg1 F	CAGCATCGAGTCGAGAGAATTT	Mutated Hmg1
	Hmg1 R	CTGCGTTACTCGGTCTTGGTAC	F262del
	Hmg1 OF	TATGCTGCCATATTTGCTGATG	Hmg1 F262del
	Hmg1 OR	ACGAGACAGTAGAGGTAGGCC	mutation area

Table S3. crRNAs used in this study

Target gene	5' crRNA	3' crRNA
<i>cyp51A</i>	GGTGCCGATGCTATGGCTTA	GAAGCCAAGCATCATCGGCT
<i>cyp51B</i>	ACATGGGTGCTTGTTGGAAT	AAAAGATCGGCCAAGCGGTT
<i>hmg1</i>	GCACCCTATACACACCATTG	GCATGGCGAAACATGAAGTA

Table S4. ARMP-PCR primers used in this study

Target gene	Primer name	Primer sequence (5' → 3')
<i>cyp51A</i>	Cyp51A OF	TGCTGAGACTGGCCTCACAGC
	Cyp51A WT	CACGTCAAGTCCCTATCTTCCGTGTG
	Cyp51A Mut	ATACAGCGGTGTCGGCCAGCCCT
	Cyp51A OR	GCCCTCGAGGGGCTGAATTAAGTAT
<i>cyp51B</i>	Cyp51B OF	CGTGTTCTCGGATCTGACTTG
	Cyp51B WT	CAATGCATCGGTGTCTACCTGATCC
	Cyp51B Mut	CCAATAGCCCGTACCTCCCGTGTA
	Cyp51B OR	TCAGGCTTTGGTAGCGGACTC

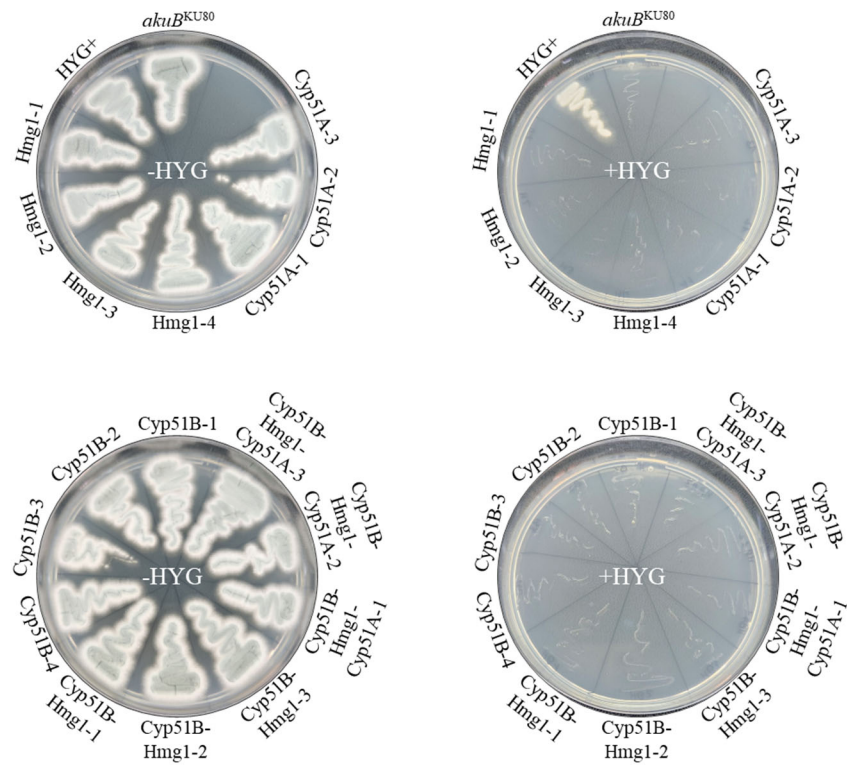


Figure S1. Analysis of mutants for loss of hygromycin resistance. Analysis of mutants Hmg1 F262del (Hmg1-1-4), Cyp51A G448S (Cyp51A1-3), Cyp51B G457S (Cyp51B1-4), Cyp51B G457S /Hmg1 F262del 1-3, and Cyp51B G457S/ Hmg1 F262del/ Cyp51A G448S 1-3 for loss of pTel-hyg^R and HYG resistance. Transformants were passaged twice on YAG absent hygromycin. Single isolates were then streaked on plates supplemented with (+HYG) or without (-HYG) hygromycin (350 µg/ml). Plates were photographed after 24 h incubation at 37C. Controls include parental strain *AkuB*^{KU80} and a positive control strain containing the hygromycin resistance cassette stably integrated into the genome (strain designated HYG+).

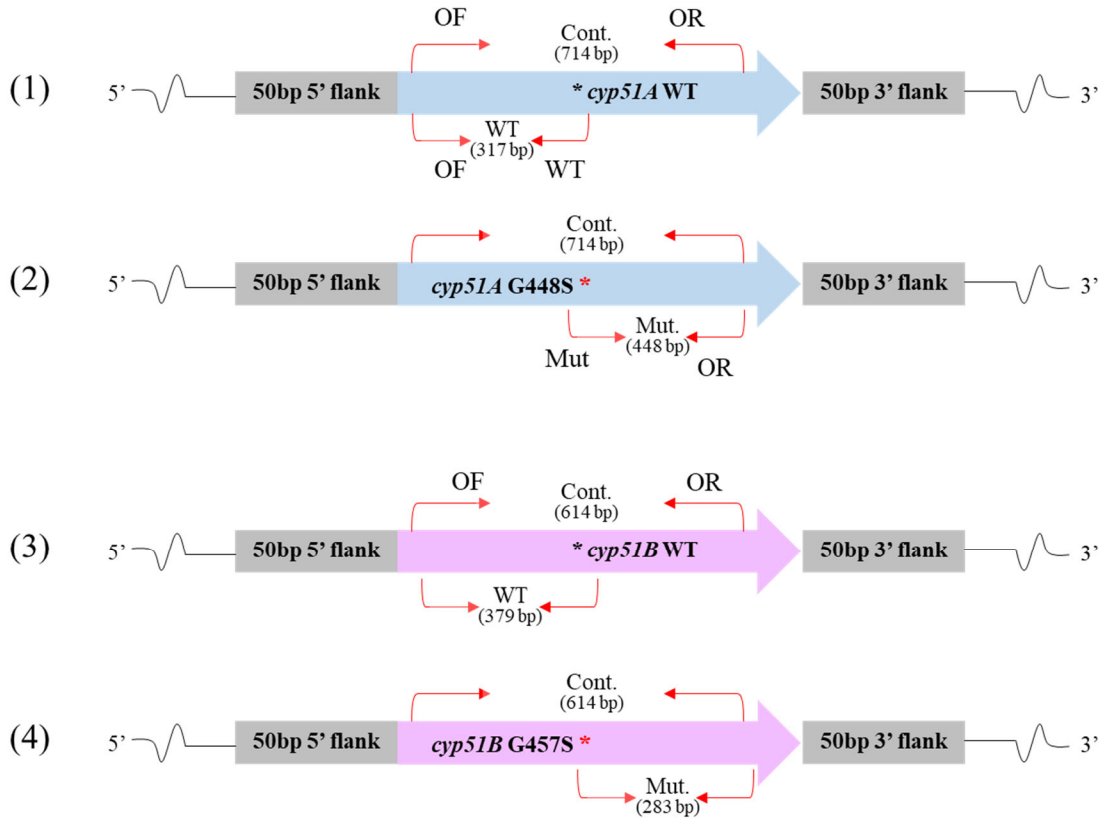


Figure S2. ARMS-PCR analysis of *cyp51A* and *cyp51B* point mutations. Cyp51A G448S detection uses three primer sets: Cyp51A OF and Cyp51A OR control primer set (S1.1), amplifying the 714 bp around the mutation (cont), Cyp51A OF and Cyp51A WT amplifying the 317 bp WT sequence if present (S1.1), and Cyp51A OR and Cyp51A Mut primer set, amplifying the 448 bp mutated (Mut.) sequence, if present (S1.2). Cyp51B G457S detection uses three primer sets: Cyp51B OF and Cyp51B OR control primer set (S1.3), amplifying the 614 bp around the mutation (cont), Cyp51B OF and Cyp51B WT amplifying the 379 bp WT sequence if present (S1.3), and Cyp51B OR and Cyp51B Mut primer set, amplifying the 283 bp mutated (Mut.) sequence, if present (S1.4).