

Supporting Information:

Contributions of a histone deacetylase (*SirT2/Hst2*) to *Beauveria bassiana* growth, development, and virulence

Qing Cai^{1,2*}, Li Tian³, Jia-Tao Xie¹, Dao-Hong Jiang¹ and Nemat O. Keyhani^{2*}

¹ State Key Laboratory of Agricultural Microbiology, College of Plant Science and Technology, Huazhong Agricultural University, Wuhan, Hubei, 430070, China.

² Department of Microbiology and Cell Science, University of Florida, Bldg. 981, Museum Rd., Gainesville, FL, 32611, USA.

³ Shandong Provincial Key Laboratory of Microbial Engineering, Department of Bioengineering, Qilu University of Technology, Jinan, Shandong, 250353, China

* Correspondence: **author:** Qing Cai, E-mail: caiqing@mail.hzau.edu.cn; Nemat, O. Keyhani, E-mail: keyhani@ufl.edu.

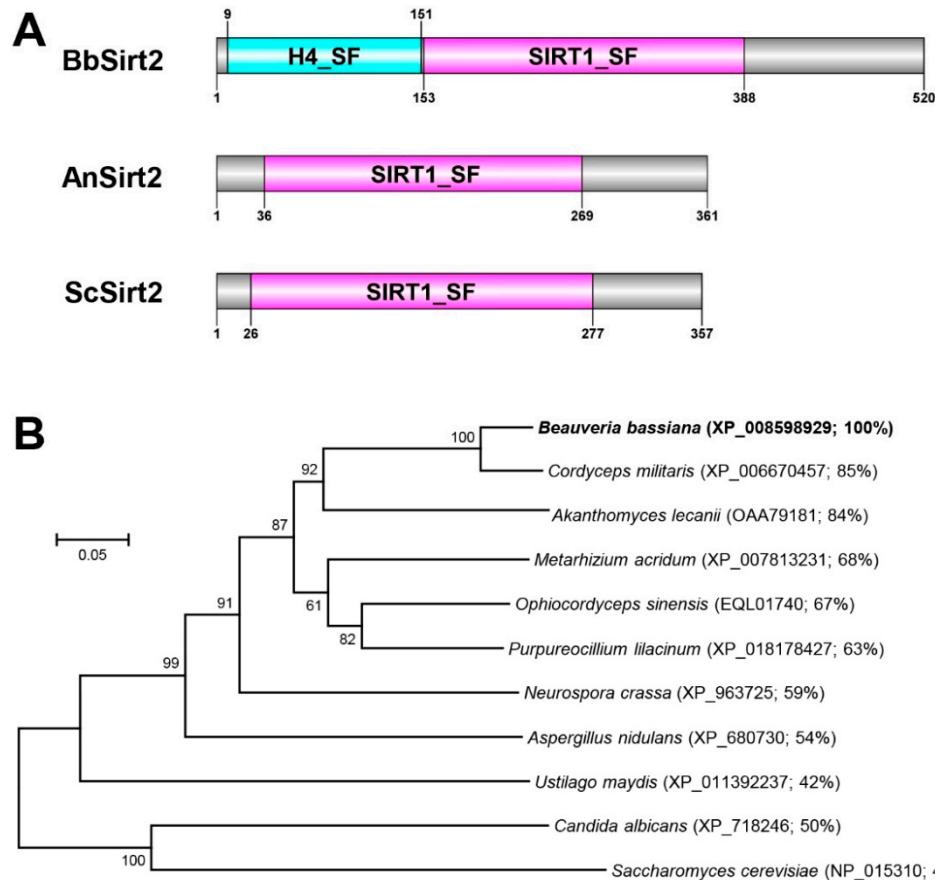


Figure S1. *B. bassiana* SirT2 phylogenetic analysis with homologs identified in other representative organisms. (A) Conserved domain of Sirt2 homologs found in *Beauveria bassiana* (*Bb*), *Aspergillus nidulans* (*An*), and *Saccharomyces cerevisiae* (*Sc*) predicted at <https://blast.ncbi.nlm.nih.gov/Blast.cgi>. (B) Phylogenetic analyses of *B. bassiana* SirT2 with the homologs found in other representative fungi. A neighbor-joining method in MEGA7 software at <http://www.megasoftware.net> was used in the phylogenetic analysis. Poisson model was performed with 1000 bootstrap replications in uniform rates. Each fungal name is followed by the NCBI accession code of each protein and its sequence identity (%) to *B. bassiana* SirT2 in parentheses. Scale bar: branch length proportional to genetic distance assessed with the neighbor-joining method.

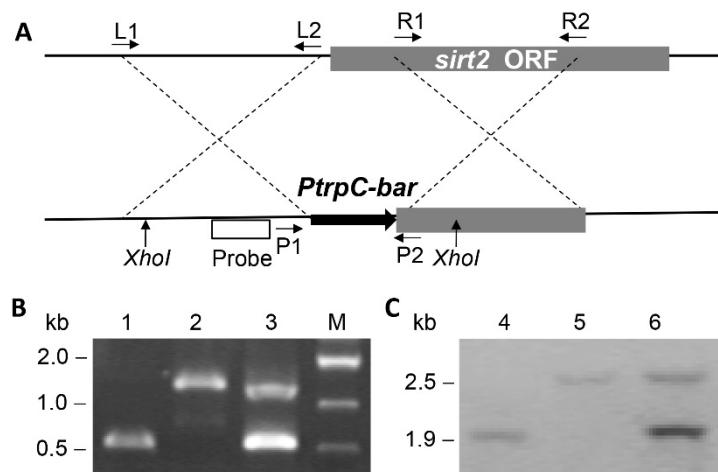


Figure S2. Construction and verification of *B. bassiana* SirT2 mutants. (A) Schematic diagram for the strategy of *Sirt2* deletion. (B, C) The *Sirt2* mutants identified via PCR (lanes 1–3) and Southern blotting (lanes 4–6) analyses with paired primers and amplified probe (Table S1). Lanes 1 and 4: wild-type. Lanes 2 and 5: Δ *Sirt2* mutant. Lanes 3 and 6: Δ *Sirt2*:: Δ *Sirt2*:: Δ *Sirt2*.

Sirt2 mutant. Genomic DNAs were digested with *Xba*I/*Xba*I at the marked sites for the Southern blotting of *Sirt2*.

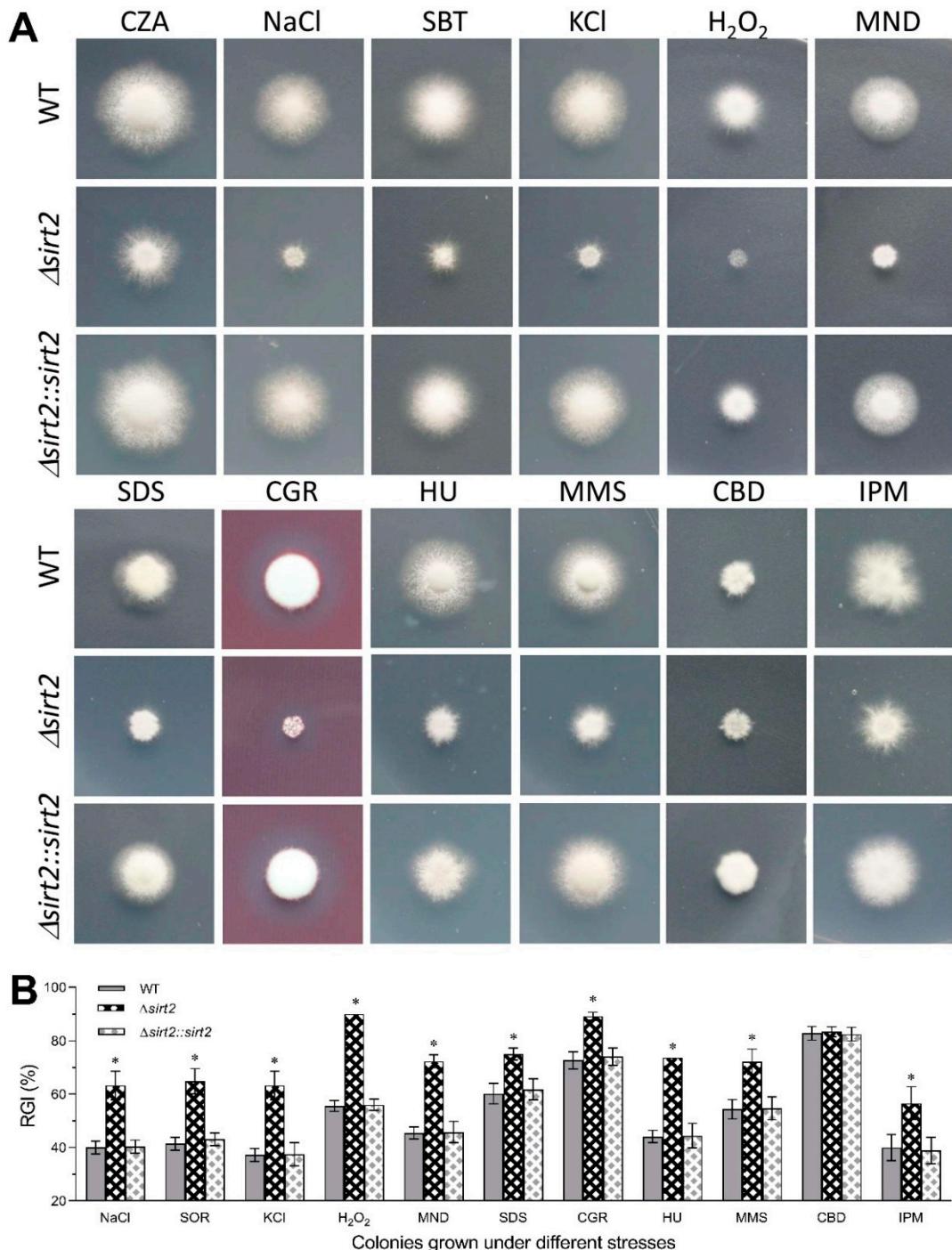


Figure S3. Stress response phenotype of *ΔBbSirT2* mutant and control strains. (A, B) Images and RGI (relative growth inhibition) of fungal colonies grown at 25°C for 7 d on CZA supplemented with either, (i) CZA unamended, control, (ii) H₂O₂ (2 mM) or menadione (0.02 mM), oxidative stress (ii) 0.4 M NaCl, 0.4 M KCl or 0.8 M sorbitol, osmotic stress; (iii) H₂O₂ (2 mM) or menadione (0.02 mM), oxidative stress; (iv) carbendazim (CBD) (10 µg ml⁻¹) or iprodione metabolite (IPM, 10 µg ml⁻¹), drug resistance analysis; (v) hydroxyurea (HU) (10 mM) or methyl methanesulfonate (MMS) (0.05%), DNA damage stress, (vi) SDS (100 µg ml⁻¹) or Congo red (10 µg ml⁻¹), cell wall perturbing stress. The asterisked bar in each three-bar group differ significantly from those unmarked (Tukey's HSD, *p* < 0.05). Error bars: SD.

Table S1. Lists of primers used for the *BbSirT2* mutant strains construction.

| Primers | Paired sequences (5'-3')* | Purpose |
|-------------|--|--|
| Sirt2up-F/R | AAAAACCCGGGCACTGTCTTGCTTCTCCGTC/ AAAAA <u>GGA</u> TCCGACAAAGTAAGCGTGCCTGG | Cloning <i>Sirt2</i> 5'-end (1579 bp) for <i>Sirt2</i> deletion |
| Sirt2dn-F/R | AAAAA <u>TCT</u> ACTGACCTACAAACAGCTACCGT/ AAAAAA <u>ACTAG</u> TCGTCAAGGAACCTTTTG <u>gggg</u> ACCACTTGATACAAGAAAGCTGGGTNCATCAC- | Cloning <i>Sirt2</i> 3'-end (1576 bp) for <i>Sirt2</i> deletion |
| Sirt2fl-F/R | GGTCCTCATT- GTC/ <u>gggg</u> ACAAGTTGTACAAAAAAGCAGGCTNGTCCAT CGGTAGCGTTATT CCCGGGACTAGT <u>GATAT</u> CATGTCCGACCAC- | Cloning full-length <i>Sirt2</i> (5102 bp) for <i>Sirt2</i> complementation |
| cSirt2-F/R | GAATTGG/CTTGCTCACCAT <u>GAATT</u> CAAGGGCCGACTT GGGAAAAC | Cloning <i>Sirt2</i> cDNA (1560 bp) |
| pSirt2-F/R | ACCTATTACTCGCCGTCGCTC / GCAGCATTCTATGAG-CAGGTC | PCR detecting <i>Sirt2</i> |
| sbSirt2-F/R | AGCTCGACTCGGTCAATT / TGCTTGTGCCTCTTATTTC | Southern probe of <i>Sirt2</i> (418 bp) |
| qSirt2-F/R | AAGACTGGGCTGTATAAC / GTAGAATGGCTCAGGATT | qPCR detecting <i>Sirt2</i> |
| q18S-F/R | TGGTTCTAGGACC CGCCGTAA / CCTTGG-CAAATGCTTCGC | qPCR detecting 18S RNA |

* Underlined regions denote the restriction enzyme sites for the deletion of *Sirt2* (*Xba*I/*Bam*HI and *Xba*I/*Spe*I) and the cloning of *Sirt2* cDNA (*Eco*RV/*Eco*RI) or the fragments of gateway exchange for *Sirt2* complementation.

Table S2. Primer list used for qPCR transcriptional profiling.

| Tag loci* | Gene | Annotation | Paired primers used in qRT-PCR |
|-----------|-------------|--|---|
| BBA_04942 | <i>FluG</i> | developmental protein | CCTCCCTAGTTGGTCGCTTCTC / CGCTGTCGGAATCTGCTCCTC |
| BBA_02968 | <i>FlbA</i> | developmental regulator | CCAATCCACTCGCCGCTCTC / CGGAGGAAA-GAGAATCGGTAGAGG |
| BBA_06988 | <i>FlbB</i> | bZIP transcription factor | GCACTGACACGCCGACAAGAGC / CCGCCGCCGAAGCCTGTTG |
| BBA_03181 | <i>FlbC</i> | C ₂ H ₂ conidiation transcription factor | TCCATCTCCAACTTGCTGGGTCTC / GGCGGCG-TAGGCGGAAGG |
| BBA_07259 | <i>FlbD</i> | MYB conidiophore development protein | CGGCAAGCGATGGGCAGAGATTG / ACGAG-CAAGGTGACGGTAGAGGTG |
| BBA_01716 | <i>FlbE</i> | conidiophore development protein | CAGACGATGAGACAGAGA / GGGCTTATATGCGAGTAG |
| BBA_07544 | <i>BrlA</i> | C ₂ H ₂ conidiation transcription factor | GACCAGTTAACAGACAAG / CAG-TAATCTCGTGCTTCTC |
| BBA_00300 | <i>AbaA</i> | Conidiation factor | GCAAGTCTCCAGCCATAT / CTCCTCTCGTCATACTAGTC |
| BBA_06126 | <i>WetA</i> | Conidial maturation factor | CGCAGACGAATTGACTT / GCTGGTGGTT-GAATACAT |
| BBA_01023 | <i>VosA</i> | Velvet protein | GGACAGACGAGTGATTGA / GGCATATACGAC-GCATCT |

* Gene accession codes in the genome database of *B. bassiana* under the NCBI accession NZ_ADAH00000000.