

SUPPLEMENTARY Tables

Supplementary Table S1. List of the genes targeted in the transcriptomic study.

Function	Gene	Primer Sequences	GenBank or TC TIGR* Accession Number
Reference genes	<i>EF1</i> (EF1- α elongation factor)	5'-GAACTGGGTGCTTAGGC-3' 5'-AACCAAAATATCCGGAGTAAAAGA-3'	GU585871
	<i>60SRP</i> (60S ribosomal protein L18)	5'-ACTACCTCAAGCTCTAGTC-3' 5'-CAATCTTGTCTCTCTTCTC-3'	XM_002270599
	<i>39SRP</i> (39S ribosomal protein L41)	5'-AACCAAAATATCCGGAGTAAAAGA-3' 5'-GACTGACTTCAAGCTTAAACC-3'	XM_002285709
Phenylpropanoid metabolism	<i>PAL</i> (Phenylalanine ammonia lyase)	5'-TCCTCCCGGAAACAGCTG-3' 5'-TCCTCCAATGCTCAAATCA-3'	X75967
	<i>STS</i> (Stilbene synthase)	5'-AGGAAGCAGCATTTGAAGGCT-3' 5'-TGACCCAGGCATTCTACACC-3'	FJ851185
Defense protein	<i>CHIT4C</i> (Chitinase class V)	5'-TCGAATGGGATGGTGGAAA -3' 5'-TCCCCTGTCGAAACACCAAG -3'	NM_001281244
	<i>GLUC</i> (β -1,3 glucanase)	5'-TCAATGGCTGCAATGGTC-3' 5'-CGGTCGATGTTGGAGATTAA-3'	DQ267748
	<i>PR1</i> (pathogenesis-related protein 1)	5'- GGAGTCCATTAGCACTCCTTG -3' 5'- CATAATTCTGGCGTAGGCAG -3'	XM_002273752
	<i>PR10</i> (pathogenesis-related protein 10)	5'- CGTTAAGGGCGCAAAGAG -3' 5'- GCATCAGGGTGTGCCAAGA -3'	DQ396809
	<i>GST1</i> (Glutathione s-transferase 1)	5'- TGCATGGAGGAGGAGTTCTG -3' 5'- CAAGGCTATATCCCCATTTCTC -3'	NM_001281248
Photosynthesis	<i>PsbP1</i> (oxygen-evolving enhancer <i>PsbP</i> subunit of photosystem I)	5'-TGTCCCCAGCTGTACCTTG-3' 5'-GCTGACGGAGATGAAGGTGG-3'	XM_002283012.4
	<i>RbcL</i> (<i>RuBisCo</i> large sub-unit)	5'-AATTTCTCCAGGCATA -3' 5'-ATCTGCGCCCGCTTATA -3'	TC57584
Arsenite recovery genes	<i>MSR</i> (Peptide methionine sulfoxide reductase))	5'-GCATTGGCGTGTAGATGG-3' 5'-GCTTCAGTGTGGCTGTTCT -3'	XM_010662104.2
	<i>WRKY</i> (transcription factors)	5'-GGGCAGAAGGACATTCTTGA -3' 5'-AGGGATCTTCATCCGAACGC -3'	XM_002272684.3
	<i>HYD2</i> (ABA 8' hydroxylase 2)	5'-GAAGCTTGTCTCCGAGCCT -3' 5'-TATTGATCGACCGGCCCTCC -3'	NM_001281052.1

* see <http://www.jcvi.org/cms/research/projects/tdb/overview/>

Supplementary Table S2: Identified asymptomatic Leaves metabolites from treated and non-treated vines (DAT-la) versus (DS-la). From 16 significant *m/z* (*P*: ANOVA p value) discriminating the two conditions DAT and DS asymptomatic leaves (see Volcano in Figure 13A) the 5 annotated metabolites are ranked according to the normalized amount difference between DAT-DS (Diff). Negative or positive differences means that the metabolites are either more accumulated in DS or in DAT, respectively. Annotations names were obtained from MassMatrix queries on *Vitis vinifera* database (with 1 ppm error), which assigned theoretical masses (ion and neutral) associated to the corresponding raw formula and predicted structure (MSCC).

ID	<i>P</i>	Diff	Th Mass		Error (ppm)	Names		
			(Ion)	Structure				
1581	0.000272	-4.60	338.088130	Ph	C ₁₅ H ₁₇ NO ₈	339.095420	0.441	6-Hydroxy-5-methoxyindole glucuronide (see KEGG C03033); (2S,3S,4S,5R,6S)-3,4,5-trihydroxy-6-[(5-methoxy-1H-indol-6-yl)oxy]oxane-2-carboxylic acid [acetal]
3316	0.004486	-4.16	413.218150	Lp	C ₂₁ H ₃₄ O ₈	414.225589	0,535	methyl 5-hydroperoxy-6,8,9,11-bisepidioxy-12,14-eicosadienoate [Hydroperoxy fatty acids [FA0104]]
3963	0.002901	-3.60	437.436393	Lp	C ₂₉ H ₅₈ O ₂	438.443680	-0,001	Nonacosanoic acid; nonacosanoic acid [carboxylic acid]
189	0.000046	-1.18	219.051039	NM	C ₈ H ₁₂ O ₇	220.058305	0,012	(R)-(Homo) 2-citrate; 1-Hydroxypentane-1,2,5-tricarboxylate; (-) threo-iso(homo) 2 citrate; 3-Hydroxy-3-carboxymethyl-adipic acid
2193	0.000682	+0.95	365.064435	NM	C ₁₃ H ₁₉ O ₁₀ P	366.071588	0,013	Salicin 6-phosphate; Salicin-6P

Supplementary Table S3 : Identified Canes metabolites from treated and non-treated vines (DAT-c) versus (DS-c). From 113 significant *m/z* (*P*: ANOVA p value) discriminating the two conditions DAT and DS canes (see Volcano in Figure 13B) the 32 annotated metabolites are ranked according to the normalized amount difference between DAT-DS (Diff). Negative or positive differences means that the metabolites are either more accumulated in DS or in DAT, respectively. Annotations names were obtained from Masstrix queries on *Vitis vinifera* database (with 1 ppm error), which assigned theoretical masses (ion and neutral) associated to the corresponding raw formula and predicted structure (MSCC).

ID	P	Diff	Th Mass (Ion)	Structure	Formula	Th Mass (Neutral)	Error (ppm)	Names
1913	0.010626	-4.21	353.103146	Ph	C ₂₀ H ₁₈ O ₆	354.110340	0.006	Cyclokievitone
212	0.009158	-3.62	223.13398	Lp	C ₁₃ H ₂₀ O ₃	224.141245	0.004	Vomifoliol; (6S,9R)-6-hydroxy-3-oxo- α -ionol
9268	0.007012	-3.55	629.172184	Ph	C ₂₇ H ₃₄ O ₁₇	630.179591	-0.012	Leucodelphinidin 3-O-(β -D-glucopyranosyl-(1->4)- β -L-rhamnopyranoside) [Flavans, Flavanols and Leucoanthocyanidins [PK1202]]
4910	1.18E-07	-3.40	471.384416	Lp	C ₃₁ H ₅₂ O ₃	472.391645	-0.000	Soyasapogenol D
9149	0.027266	-3.39	623.161585	Ph	C ₂₈ H ₃₂ O ₁₆	624.168680	-0.568	Tectorigenin 7-O-gentioside [Isoflavonoids [PK1205]]
8162	0.041731	-2.93	575.452665	Lp	C ₃₂ H ₆₄ O ₈	576.460120	0.002	1-(O- β -D-glucopyranosyl)-(1,3R,25R)-hexacosanetriol [Fatty acyl glycosides of mono- and disaccharides [FA1301]]
8734	0.015264	-2.76	603.484088	Lp	C ₃₄ H ₆₈ O ₈	604.491420	0.002	1-(O- β -D-glucopyranosyl)-(1,3R,27R)-octaconsanetriol [Fatty acyl glycosides of mono- and disaccharides [FA1301]]
2470	0.000113	-2.75	377.160518	Ph	C ₂₀ H ₂₆ O ₇	378.167631	-0.587	Chaparrinone
6583	0.022281	-2.72	517.389808	Lp	C ₃₂ H ₅₄ O ₅	518.397125	0.001	7 β -acetoxy-gorgostan-3 β ,5 β ,6 β -triol [Gorgosterols and derivatives [ST0106]]
3850	0.034607	-2.45	433.114036	Ph	C ₂₁ H ₂₂ O ₁₀	434.121138	-0.365	Naringenin 7-O- β -D-glucoside; Prunin
168	0.003001	-2.39	215.032805	Cb	C ₅ H ₁₃ O ₇ P	216.040068	0.832	2-C-Methyl-D-erythritol 4-phosphate
398	0.017330	-2.29	252.051365	Ph	C ₁₁ H ₁₁ NO ₆	253.058639	0.009	N-Pyruvoyl-5-methoxy-3-hydroxyanthranilate; N-Pyruvoyl-5-methoxy-3-hydroxyanthranilic acid
4275	0.010541	-2.17	449.108874	Ph	C ₂₁ H ₂₂ O ₁₁	450.116228	0.037	Neoastilbin; (2S,3S)-Taxifolin 3-rhamnoside
3963	0.005772	-2.03	437.436392	Lp	C ₂₉ H ₅₈ O ₂	438.4443680	-0.001	Nonacosanoic acid; nonacosanoic acid [carboxylicacid]
9827	0.043267	-2.00	659.416826	Lp	C ₃₈ H ₆₀ O ₉	660.424359	0.949	12-O-Palmitoyl-16-hydroxyporphol 13-acetate; Crotonfactor F1
3595	0.019732	-1.86	423.420748	Lp	C ₂₈ H ₅₆ O ₂	424.428030	-0.001	Octacosanoic acid; octacosanoic acid [carboxylicacid]
2193	0.019086	-1.70	365.064434	Cb	C ₁₃ H ₁₉ O ₁₀ P	366.071588	0.013	Salicin 6-phosphate; Salicin-6P
4370	0.021845	-1.65	451.452007	Lp	C ₃₀ H ₆₀ O ₂	452.459330	-0.001	Melissic acid; triacontanoic acid [Straight chain fatty acids [FA0101]]
3437	0.041210	-1.65	419.098345	Ph	C ₂₀ H ₂₀ O ₁₀	420.105480	-0.396	5,3',5'-Trihydroxy-3,6,7,8,4'-pentamethoxyflavone [Flavones and Flavonols [PK1211]]
2711	0.035887	-1.38	387.129652	Lp	C ₁₇ H ₂₄ O ₁₀	388.136901	-0.117	Secologanin; (-)-Secologanin
1466	0.026006	-1.25	333.061614	Ph	C ₁₆ H ₁₄ O ₈	334.068821	-0.137	6-Methoxytaxifolin
8192	0.041414	-1.24	577.135010	Ph	C ₃₀ H ₂₆ O ₁₂	578.142443	0.029	Epicatechin-(4 β ->8)-ent-epicatechin
1341	0.020785	-1.21	327.10854	Ph	C ₁₅ H ₂₀ O ₈	328.115820	0.008	Anisatin
8092	0.034853	-1.01	571.327453	Lp	C ₃₃ H ₄₈ O ₈	572.334750	-0.294	6,8a-Seco-6,8a-deoxy-5-oxoavermectin"2b"glycone
6493	0.035506	-0.93	515.192200	Pp	C ₁₄ H ₃₃ N ₁₀ O ₇ PS	516.199536	0.651	Phaseolotoxin
1699	0.009584	-0.73	343.124559	Cb	C ₁₂ H ₂₄ O ₁₁	344.131972	0.323	Melibitol; 6-O- β -D-Galactosyl-D-glucitol
476	0.041925	-0.67	263.128880	Ph	C ₁₅ H ₂₀ O ₄	264.136160	0.005	Abscisate; Abscisic acid; (+)-Abscisic acid
1291	0.025648	-0.45	325.114000	Cb	C ₁₂ H ₂₁ O ₁₀ R	326.121300	0.010	R replaced by H in β -L-Fucosyl-1,2- β -D-galactoside; Glycoprotein β -L-fucosyl-(1,2)-D-galactose; Glycolipid β -L-fucosyl-1,2- β -D-galactose; β -L-Fucosyl-(1->2)-D-galactosyl-R
2044	0.034599	-0.45	359.098344	NM	C ₁₅ H ₂₀ O ₁₀	360.105488	-0.441	3-Methoxy-4-hydroxyphenylglycol glucuronide (see KEGG C03033); (2S,3S,4S,5R,6R)-3,4,5-trihydroxy-6-[2-hydroxy-2-(4-hydroxy-3-methoxy-phenyl) ethoxy]oxane-2-carboxylic acid [acetal]

4909	1.26E-05	+4.75	471.38413	Lp	C ₃₁ H ₅₂ O ₃	472.391300	-0.732	Soyasapogenol D
5268	0.034407	+0.89	481.135032	Ph	C ₂₂ H ₂₆ O ₁₂	482.142070	-0.739	Catalposide
5975	0.012284	+0.26	501.145887	Cb	C ₁₈ H ₃₀ O ₁₆	502.153390	0.011	β-L-Rhamnopyranosyl-(1->2)-β-D-galactopyranosyl-(1->2)-β-D-glucuronopyranoside; β-Fabatriose

Supplementary Table S4. List of fungal species detected in the woody subsamples of vines belonging to the three different conditions analyzed in 2019. *In bold the genera/species associated to GTDs.*

<i>Curvularia gladioli</i>			
<i>Cylindrotrichum gorii</i>		Detected	
<i>Cystobasidium slooffiae</i>	Detected		
<i>Cystodendron sp.</i>			Detected
<i>Cystofilobasidium capitatum</i>			
<i>Cystofilobasidium macerans</i>			
<i>Dactylolectria alcacerensis</i>			
<i>Dactylolectria pauciseptata</i>			
<i>Dendryphion europaeum</i>			
<i>Devriesia sp.</i>		Detected	
<i>Didymella sp.</i>			
<i>Didymosphaeria futilis</i>			
<i>Didymosphaeria sp.</i>			
<i>Diplodia seriata</i>			
<i>Entoleuca sp.</i>			
<i>Entoloma platyphyloides</i>			
<i>Epicoccum nigrum</i>			
<i>Erysiphe arcuata</i>			
<i>Erysiphe heraclei</i>			
<i>Erythrobasidium hasegavianum</i>			
<i>Eutypa lata</i>			
<i>Eutypa leptoplaca</i>			
<i>Exophiala brunnea</i>			
<i>Exophiala equina</i>			
<i>Exophiala moriliae</i>			
<i>Exophiala opportunistica</i>			
<i>Exophiala sp.</i>			
<i>Exophiala xenobiotica</i>			
<i>Fellomyces horovitziae</i>			
<i>Fellomyces mexicanus</i>			
<i>Fellomyces sp.</i>			
<i>Fellomyces thailandicus</i>			
<i>Fibulobasidium inconspicuum</i>			
<i>Filibasidium floriforme</i>			
<i>Flagelloscypha minutissima</i>			
<i>Flagelloscypha sp.</i>			
<i>Flavoplaca sp.</i>			
<i>Fomitiporia aethiopica</i>			
<i>Fomitiporia mediterranea</i>			
<i>Fomitiporia polymorpha</i>			
<i>Fusarium acuminatum</i>			
<i>Fusarium beomiforme</i>			
<i>Fusarium boothii</i>			
<i>Fusarium pseudensiforme</i>			
<i>Fusarium sp.</i>			
<i>Fusidium griseum</i>			
<i>Geopora cervina</i>			
<i>Geopyxis sp.</i>			
<i>Geotrichum klebahnii</i>			
<i>Guehomycetes pullulans</i>			
<i>Gymnascella aurantiaca</i>			
<i>Gymnopus dryophilus</i>			
<i>Gymnstellatospora frigida</i>			
<i>Hannaella oryzae</i>			
<i>Hannaella surugaensis</i>			
<i>Hannaella zaea</i>			
<i>Hanseniaspora thailandica</i>			
<i>Hanseniaspora uvarum</i>			
<i>Helgardia anguiooides</i>			
<i>Helicoma sp.</i>			
<i>Hemimycena cucullata</i>			
<i>Herpotrichia parasitica</i>			
<i>Heteroacanthella acanthophysa</i>			
<i>Holtermannia corniformis</i>			
<i>Humicola grisea</i>			
<i>Hydropisphaera sp.</i>			
<i>Hyphodermella corrugata</i>			
<i>Hyphodontia radula</i>			

<i>Hypholoma fasciculare</i>			Detected					
<i>Ilyonectria lirioidendri</i>		Detected	Detected					
<i>Kavinia alboviridis</i>								
<i>Lachancea thermotolerans</i>				Detected				
<i>Lachnum cerinum</i>					Detected			
<i>Lachnum sp.</i>								
<i>Lalaria carpini</i>				Detecte				
<i>Lasiobelonium nidulum</i>								
<i>Lecania cyrtella</i>								
<i>Lecania cyrtellina</i>				Detecte				
<i>Lecania proteiformis</i>								
<i>Lecanicillium muscarium</i>				Detecte				
<i>Lecanicillium saksenae</i>								
<i>Lecanicillium sp.</i>				Detecte				
<i>Lecanora albescens</i>								
<i>Lecanora dispersa</i>				Detecte				
<i>Lecanora hagenii</i>								
<i>Lecythophora sp.</i>				Detecte				
<i>Leohumicola minima</i>								
<i>Lepiota brunneoincarnata</i>				Detecte				
<i>Lepiota echinella</i>								
<i>Lepista inversa</i>				Detecte				
<i>Leptosphaeria sp.</i>				Detecte				
<i>Leptosphaerulina australis</i>								
<i>Lewia infectoria</i>				Detecte				
<i>Limonomycetes roseipellis</i>								
<i>Lipomyces chichibuensis</i>								
<i>Lophiostoma sp.</i>				Detecte				
<i>Malassezia globosa</i>								
<i>Malassezia restricta</i>				Detecte				
<i>Malassezia sp.</i>								
<i>Massarina sp.</i>								
<i>Metarhizium anisopliae</i>								
<i>Metschnikowia chrysoperiae</i>				Detecte				
<i>Metschnikowia pulcherrima</i>								
<i>Metschnikowia sp.</i>				Detecte				
<i>Meyeromyza guilliermondii</i>								
<i>Microascus sp.</i>				Detecte				
<i>Microdiplosid sp.</i>								
<i>Monographella cucumerina</i>								
<i>Monographella nivalis</i>								
<i>Morchella semilibera</i>								
<i>Mortierella camargensis</i>								
<i>Mortierella clonocystis</i>								
<i>Mortierella elongata</i>				Detecte				
<i>Mortierella rishikeshia</i>								
<i>Mortierella sp.</i>								
<i>Mortierella stylospora</i>								
<i>Mucor brunneogriseus</i>								
<i>Mycena capillaris</i>								
<i>Mycena cicognanii</i>				Detecte				
<i>Mycena sp.</i>								
<i>Mycena terena</i>								
<i>Mycenella bryophiloides</i>								
<i>Mycenella trachyspora</i>								
<i>Mycoleptodiscus sp.</i>				Detecte				
<i>Nectria cinnabarinaria</i>								
<i>Nectria ramulariae</i>								
<i>Nematoctonus leiosporus</i>								
<i>Neoascochyta exitialis</i>								
<i>Neopeckia coulteri</i>				Detecte				
<i>Neophaeoasphaeria sp.</i>								
<i>Neosetophoma sp.</i>								
<i>Neurospora terricola</i>								
<i>Nidulariopsis iowensis</i>				Detecte				
<i>Nigroporus vinosus</i>								
<i>Ochroconis sp.</i>								
<i>Oidiodendron griseum</i>				Detecte				

Supplementary Table S5: Grape juices enological parameters obtained from the analysis of two distinct grape juices from five distinct vine trunks, for each of the three vine conditions (only 4 vine trunks for the H condition).

	pH ^a		α -amino Nitrogen ^b (mg/L)		Ammoniacal Nitrogen ^a (mg/L)		TA ^a (g/L Sulfuric acid)		Sugars ^a (g/L)	
Sample	Average	STD	Average	STD	Average	STD	Average	STD	Average	STD
DS1	2.74	0.06	90	15	99.55	0.05	8.7	0.7	100	12
DS2	2.91	0.01	84	5	85	3	7.9	0.1	169	4
DS3	2.83	0.03	93	7	86	6	9.1	0.1	133	2
DS4	2.94	0.00	241	17	361	11	11.7	0.4	56	3
DS5	2.81	0.10	55	1	82	3	7.4	0.5	155	14
DAT1	2.85	0.03	61	2	74	2	6.07	0.18	167	6
DAT2	2.83	0.01	79	5	123	7	6.69	0.12	156	3
DAT3	2.87	0.01	78.2	0	93	2	6.6	0.1	171	3
DAT4	2.95	0.01	46	9	45	2	6.96	0.09	145.1	3.0
DAT5	2.82	0.04	81	7	82	6	6.9	0.3	164	9
H1	2.98	0.01	68	4	66.9	0.9	7.6	0.1	176	3
H2	3.06	0.03	210	29	237.1	0.5	6.5	0.0	178	2
H3	2.93	0.01	95	9	88	5	7.54	0.05	175.1	1.8
H5	2.99	0.02	19	3	45	0.4	7.0	0.1	154	5

a. FTIR : Fourier Transformed Infra Red; b. enzymatic kit ; TA :total Acidity ; STD : Standard Deviation