

Editorial

Host–Pathogen Interactions: Insects vs. Fungi

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Although many insects successfully live in dangerous environments exposed to diverse communities of microbes, they are often exploited and killed by specialist pathogens. In the process of co-evolution of insects and entomopathogenic microorganisms, they develop various adaptive systems that determine the sustainable existence of dynamic host–parasite interactions at both the organismic and population levels. Many different species of fungi are associated with insects. It should be noted that the diversity of fungi largely depends on the specific insect–fungus system. Thus, in the population of *Chilo suppressalis*, a serious pest of rice, in northern Iran *Beauveria bassiana*, *Akanthomyces lecanii*, *Akanthomyces muscarius*, *Metarhizium anisopliae*, *Hirsutella subulata*, and *Trichoderma* sp. persisted [1]. Lepidopteran forest-pest species *Ematurga atomaria*, *Cabera pusaria*, *Hypomecis punctinalis*, and *Orthosia gothica* were associated with members of Cordycipitaceae (*Akanthomyces muscarius* and *Cordyceps farinosa*) and fungi from families *Aspergillaceae*, *Nectriaceae*, *Mortierellaceae*, *Hypocreaceae*, etc. [2]. The host defences are designed to exclude the pathogen or mitigate the damage inflicted, while the pathogen counters with immune evasion and utilization of host resources. Transcriptome (RNAseq) analysis of immune response uncovers new abilities to study host–parasite systems. Study of cricket *Gryllus bimaculatus* transcriptome demonstrated high tissue-specific variety in inducing antifungal immune factors [3]. Entomopathogenic fungi (EPF) neutralize their immediate surroundings on the insect integument and benefit from the physicochemical properties of the cuticle and its compounds that exclude competing microbes. Interestingly, in some cases EPF have low virulence because plant phytochemicals can demonstrate antimicrobial activity on insects cuticle [4]. EPF interplay host defence with factors which regulate adhesion to the cuticle, cuticle degradation, stress management and toxins [5]. Thus *B. bassiana* express bassianolide and beauvericin toxins during infection of the bug *Triatoma infestans* [6] and proteases, chitinases and lipase in the presence of *C. suppressalis* cuticle probably to pass the insects defence faster [1]. It was found that EPF peroxisome-type and hexagonal crystal-like organelles (Woronin bodies) are required for appressorium differentiation and the topical infection of insect hosts [7]. Insects' immune, detoxification, and antioxidant systems work synergistically to combat infections and mitigate stress. Some proteins demonstrate multifunctional properties, participating in metabolism, homeostasis, and pathogen recognition [8]. Besides, insect hormones such as juvenile hormone [9] and dopamine [10] have been suggested to be a potential mediator in the insects' immunity against fungi.

The application of EPF in the field needs high-quality scientific support to establish the mechanisms of action and ways to improve fungal biological preparations [11,12]. There are some cases in which an insect's microbiota [13] and nematodes [14] may influence the development of fungal infections. These facts could open new abilities for the development of a complex approach to plant biological protection.

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References

1. Shahriari, M.; Zibae, A.; Khodaparast, S.A.; Fazeli-Dinan, M. Screening and virulence of the entomopathogenic fungi associated with chilo suppressalis walker. *J. Fungi* **2021**, *7*, 34. [[CrossRef](#)] [[PubMed](#)]
2. Gielen, R.; Meister, H.; Tammaru, T.; Põldmaa, K. Fungi recorded on folivorous lepidoptera: High diversity despite moderate prevalence. *J. Fungi* **2021**, *7*, 25. [[CrossRef](#)] [[PubMed](#)]
3. Hussain, A.; Ali, M.W.; Aljabr, A.M.; Al-Kahtani, S.N. Insights into the gryllus bimaculatus immunerelated transcriptomic profiling to combat naturally invading pathogens. *J. Fungi* **2020**, *6*, 232. [[CrossRef](#)] [[PubMed](#)]
4. Zemek, R.; Konopická, J.; Abdin, Z.U. Low efficacy of isaria fumosorosea against box tree moth cydalima perspectalis: Are host plant phytochemicals involved in herbivore defence against fungal pathogens? *J. Fungi* **2020**, *6*, 342. [[CrossRef](#)] [[PubMed](#)]
5. Grizanov, E.V.; Coates, C.J.; Dubovskiy, I.M.; Butt, T.M. Metarhizium brunneum infection dynamics differ at the cuticle interface of susceptible and tolerant morphs of Galleria mellonella. *Virulence* **2019**, *10*, 999–1012. [[CrossRef](#)] [[PubMed](#)]
6. Baldiviezo, L.V.; Pedrini, N.; Santana, M.; Mannino, M.C.; Nieva, L.B.; Gentile, A.; Cardozo, R.M. Isolation of Beauveria Bassiana from the Chagas disease vector Triatoma Infestans in the gran Chaco region of Argentina: Assessment of gene expression during host-pathogen interaction. *J. Fungi* **2020**, *6*, 219. [[CrossRef](#)] [[PubMed](#)]
7. Tang, G.; Shang, Y.; Li, S.; Wang, C. Mrhex1 is required for woronin body formation, fungal development and virulence in metarhizium robertsii. *J. Fungi* **2020**, *6*, 172. [[CrossRef](#)] [[PubMed](#)]
8. Butt, T.M.; Coates, C.J.; Dubovskiy, I.M.; Ratcliffe, N.A. Entomopathogenic Fungi: New Insights into Host-Pathogen Interactions. *Adv. Genet.* **2016**, *94*, 307–364. [[CrossRef](#)] [[PubMed](#)]
9. Rantala, M.J.; Dubovskiy, I.M.; Pölkki, M.; Krama, T.; Contreras-Garduño, J.; Krams, I.A. Effect of juvenile hormone on resistance against entomopathogenic fungus metharizium robertsii differs between sexes. *J. Fungi* **2020**, *6*, 298. [[CrossRef](#)] [[PubMed](#)]
10. Chertkova, E.A.; Grizanov, E.V.; Dubovskiy, I.M. Bacterial and fungal infections induce bursts of dopamine in the haemolymph of the Colorado potato beetle Leptinotarsa decemlineata and greater wax moth Galleria mellonella. *J. Invertebr. Pathol.* **2018**, *153*, 203–206. [[CrossRef](#)] [[PubMed](#)]
11. Putnok-Csicsó, B.; Tonk, S.; Szabó, A.; Márton, Z.; Bogdányi, F.T.; Tóth, F.; Abod, É.; Bálint, J.; Balog, A. Effectiveness of the entomopathogenic fungal species metarhizium anisopliae strain ncaim 362 treatments against soil inhabiting melolontha melolontha larvae in sweet potato (Ipomoea batatas L.). *J. Fungi* **2020**, *6*, 116. [[CrossRef](#)] [[PubMed](#)]
12. Karthi, S.; Vasanth-Srinivasan, P.; Ganesan, R.; Ramasamy, V.; Senthil-Nathan, S.; Khater, H.F.; Radhakrishnan, N.; Amala, K.; Kim, T.J.; El-Sheikh, M.A.; et al. Target activity of isaria tenuipes (Hypocreales: Clavicipitaceae) fungal strains against dengue vector aedes aegypti (lin.) and its non-target activity against aquatic predators. *J. Fungi* **2020**, *6*, 196. [[CrossRef](#)]
13. Kryukov, V.Y.; Kosman, E.; Tomilova, O.; Polenogova, O.; Rotskaya, U.; Tyurin, M.; Alikina, T.; Yaroslavtseva, O.; Kabilov, M.; Glupov, V. Interplay between fungal infection and bacterial associates in the wax moth galleria mellonella under different temperature conditions. *J. Fungi* **2020**, *6*, 170. [[CrossRef](#)] [[PubMed](#)]
14. Zhang, Y.; Li, S.; Li, H.; Wang, R.; Zhang, K.Q.; Xu, J. Fungi–nematode interactions: Diversity, ecology, and biocontrol prospects in agriculture. *J. Fungi* **2020**, *6*, 206. [[CrossRef](#)] [[PubMed](#)]