

Supplementary material

Supplementary material: Molecular diagnosis of paracoccidioidomycosis: current status and future perspectives

Breno Gonçalves Pinheiro¹, Rosane Christine Hahn², Zoilo Pires de Camargo^{1,3}, Anderson Messias Rodrigues^{1,*}

¹ Laboratory of Emerging Fungal Pathogens, Department of Microbiology, Immunology, and Parasitology, Discipline of Cellular Biology, Federal University of São Paulo (UNIFESP), São Paulo, Brazil.

² Federal University of Mato Grosso, Faculty of Medicine, Laboratory of Mycology/Research, Cuiabá, Mato Grosso, Brazil, Federal University of Mato Grosso, Júlio Muller University Hospital, Mato Grosso, Brazil.

³ Department of Medicine, Discipline of infectious Diseases, Federal University of São Paulo (UNIFESP), São Paulo, Brazil.

* Correspondence: amrodrigues.amr@gmail.com; Tel.: +55-1155764551 Ext. 1540.

1. Search strategy

A search of PubMed was undertaken in September 2020. The search terms used were: (1) *Paracoccidioides* AND molecular diagnosis; (2) Paracoccidioidomycosis AND molecular diagnosis; (3) *Paracoccidioides* AND PCR; (4) *Paracoccidioides* AND MLSA; (5) *Paracoccidioides* AND barcoding; (6) *Paracoccidioides* AND internal transcribed spacer; (7) *Paracoccidioides* AND restriction fragment length polymorphism; (8) *Paracoccidioides* AND restriction amplified polymorphic DNA; (9) *Paracoccidioides* AND loop-mediated isothermal amplification; (10) *Paracoccidioides* AND qPCR; (11) *Paracoccidioides* AND *in situ* hybridization; (12) *Paracoccidioides* AND MALDI-ToF; (13) *Paracoccidioides* AND FTIR; (14) *Paracoccidioides* AND transposable elements; (15) *Paracoccidioides* AND microsatellites, and 73-72-116-0-1-16-6-2-2-29-3-3-3-3-6 results were returned, respectively. These results were manually filtered to recover articles that describe new methods aiming to be used in the molecular diagnosis of paracoccidioidomycosis.

Search #	Terms	Period covered	N. of articles
1	<i>Paracoccidioides</i> AND molecular diagnosis	1986-2020	73
2	Paracoccidioidomycosis AND molecular diagnosis	1986-2020	72
3	<i>Paracoccidioides</i> AND PCR	1995-2020	116
4	<i>Paracoccidioides</i> AND MLSA	-	0
5	<i>Paracoccidioides</i> AND barcoding	2019-2020	1
6	<i>Paracoccidioides</i> AND internal transcribed spacer	1998-2020	16
7	<i>Paracoccidioides</i> AND restriction fragment length polymorphism	2000-2020	6
8	<i>Paracoccidioides</i> AND restriction amplified polymorphic DNA	1999-2020	2
9	<i>Paracoccidioides</i> AND loop-mediated isothermal amplification	2004-2020	2
10	<i>Paracoccidioides</i> AND qPCR	2004-2020	29
11	<i>Paracoccidioides</i> AND <i>in situ</i> hybridization	1999-2020	3
12	<i>Paracoccidioides</i> AND MALDI-ToF	2010-2020	3
13	<i>Paracoccidioides</i> AND FTIR	2013-2020	3
14	<i>Paracoccidioides</i> AND transposable elements	2009-2020	3
15	<i>Paracoccidioides</i> AND microsatellites	2000-2020	6

The articles described in the Supplementary Table S1 were included in this review.

Table S1: Summary of molecular diagnosis methods in paracoccidioidomycosis.

Technique	Identification ¹	Species ²	Typing ³	Sample	Sensitivity	Specificity	Target	Ref. ⁴
PCR	Yes	Pcx	No	Culture	N/A	N/A	β-actin	[1]
PCR	Yes	Pcx	No	Culture	N/A	100%	ITS 28S	[2]
PCR	Yes	Pcx	No	Culture, soil armadillo tissue	3 pg	N/A	P27	[3]
PCR	Yes	Pcx	No	Culture	N/A	100%	ITS 5.8S	[4]
PCR	Yes	Pcx	No	Culture	N/A	N/A	ITS 5.8S 28S	[5]
PCR	Yes	Pcx	No	Sputum	10 cells/ml	N/A	GP43	[6]
PCR	Yes	Pcx	No	Sputum, CSF	10 pg	N/A	0.72 kb fragment	[7]
PCR	Yes	PI	No	Culture	N/A	N/A	HSP70	[8]
PCR	Yes	PI	No	Serum	1.1 pg/μl	N/A	ITS	[9]
Nested-PCR	Yes	Pcx	No	Tissue	N/A	N/A	GP43	[10]
Nested-PCR	Yes	Pcx	Yes	FFPE	N/A	N/A	GP43	[11]
Nested-PCR	Yes	Pcx	No	Culture, lymph nodes	N/A	N/A	Ceja-1	[12]
Semi-nested-PCR	Yes	Pcx	No	Biopsy	0.25 pg	100%	ITS	[13]
Semi-nested-PCR	Yes	Pcx	No	Sputum	2.5 pg	100%	ITS	[14]
Nested-PCR	Yes	Pcx	No	Culture, sputum, CSF, body fluids	1 fg	100%	GP43	[15]
Nested-PCR	Yes	<i>Paracoccidioides</i>	No	Soil	N/A	N/A	ITS	[16]
RFLP	No	<i>Paracoccidioides</i> (5 groups)	Yes	Culture	N/A	N/A	Whole-genome	[17]
PCR-RFLP	Yes	S1, PS2, PS3, PI	Yes	Culture	N/A	N/A	TUB-1	[18]
RAPD	No	<i>Paracoccidioides</i> (2 groups)	Yes	Culture	N/A	N/A	Whole-genome	[19]
RAPD	No	<i>Paracoccidioides</i> (5 groups)	Yes	Culture	N/A	N/A	Whole-genome	[20]
RAPD	No	<i>Paracoccidioides</i> (2 groups)	Yes	Culture	N/A	N/A	Whole-genome	[21]
RAPD	No	<i>Paracoccidioides</i> (3 groups)	Yes	Culture	N/A	N/A	Whole-genome	[22]
RAPD	No	<i>Paracoccidioides</i>	Yes	Culture	N/A	N/A	Whole-genome	[23]

Technique	Identification ¹	Species ²	Typing ³	Sample	Sensitivity	Specificity	Target	Ref. ⁴
RAPD	No	(2 groups) <i>Paracoccidioides</i>	Yes	Culture	N/A	N/A	Whole-genome	[24]
RAPD	No	(2 groups) <i>Paracoccidioides</i>	Yes	Culture	N/A	N/A	Whole genome	[25,26]
RAPD	No	(2 groups) <i>Paracoccidioides</i>	Yes	Culture	N/A	N/A	Whole-genome	[27]
LAMP	Yes	Pcx	No	FFPE	100 fg	N/A	GP43	[28]
LAMP	Yes	Pcx	No	Sputum	N/A	N/A	GP43	[29]
LAMP	Yes	PI	No	Culture	N/A	100%	GP43	[30]
qPCR	Yes	Pcx	No	Culture	N/A	100%	GP43	[31]
qPCR	Yes	Pcx	No	Culture, Sputum, tissue, serum	N/A	100% culture, 50% serum	ITS	[32]
qPCR	Yes	S1, PS2, PS3, PI	No	Culture	N/A	N/A	GP43, ARF, PRP8	[33]
qPCR	Yes	<i>Paracoccidioides</i>	No	Culture, tissue, serum	10 fg	100%	PB27	[34]
qPCR	Yes	Pcx	No	Mice	2 fg	N/A	GP43	[35]
ISH	Yes	Pcx	No	Oral biopsies	N/A	N/A	ITS	[36]
ISH	Yes	Pcx, PI	No	Aerosol	N/A	N/A	ITS	[16]
FISH	Yes	Pcx, PI	No	Slides	N/A	N/A	ITS	[37]
MALDI-ToF	Yes	Pcx, PI	Yes	Culture	N/A	N/A	-	[38]
FT-IR	Yes	Pcx, PI	Yes	Culture	N/A	N/A	-	[39]
Transposable Elements	Yes	Pcx, PI	Yes	Culture	N/A	N/A	Mariner Transposon	[40]
Microsatellites	No	<i>Paracoccidioides</i>	Yes	Culture	N/A	N/A	Whole-genome	[41]
Microsatellites (SSR)	No	<i>Paracoccidioides</i> (3 groups)	Yes	Culture	N/A	N/A	Whole-genome	[42]
Microsatellites (SSR)	No	<i>Paracoccidioides</i> (4 groups)	Yes	Culture	N/A	N/A	Whole-genome	[43]
MLSA	Yes	S1, PS2, PS3	Yes	Culture	N/A	N/A	CHS2, FKS, TUB, ARF, PBGP43	[44]
MLSA	Yes	S1, PS2, PS3, PI	Yes	Culture	N/A	N/A	HYD1, HSP, KEX, ITS	[45]

Technique	Identification ¹	Species ²	Typing ³	Sample	Sensitivity	Specificity	Target	Ref. ⁴
MLSA	Yes	S1, PS2, PS3, Pl	Yes	Culture	N/A	N/A	GP43, ARF, β -TUB, HSP70	[8]
ITS Barcoding	Yes	Pcx	Yes	Culture	N/A	N/A	ITS	[46,47]
ITS Barcoding	Yes	Pcx, Pl	Yes	Culture	N/A	N/A	ITS, GP43	[48]

¹Does the method allow identification? ²Speciation level allowed; ³Does the method allows for molecular typing? ⁴Reference. PCM: paracoccidioidomycosis; Pcx: *P. brasiliensis* complex = *P. brasiliensis sensu lato*; Pl: *P. lutzii*; CSF: cerebrospinal fluid; PCR: polymerase chain reaction; qPCR: quantitative real-time polymerase chain reaction; LAMP: loop-mediated isothermal amplification; FFPE: formalin-fixed paraffin-embedding; MLSA: multilocus sequence analysis; SSR: single sequence repeats; Pb: *Paracoccidioides brasiliensis*; SnaPshot: single-nucleotide polymorphism (SNP) genotyping; MALDI-ToF: matrix-assisted laser desorption/ionization time-of-flight mass spectrometry; PCR-RFLP: polymerase chain reaction-restriction fragment length polymorphism; TUB1: tubulin alpha-1 chain; FISH: fluorescence *in situ* hybridization; FT-IR: Fourier-transform infrared spectroscopy; ITS: Internal transcribed spacer; GP43: 43-kDa glycoprotein; ARF: ADP-ribosylation factor ; PRP8: Pre-mRNA processing 8; P27: 27-kDa protein; HSP70: 70kDa-heat-shock protein; β -actin: Beta-actin; CHS2: Chitin synthase; FKS: β -glucan synthase; TUB: tubulin; HYD1: Hydrophobin 1; KEX: Kex protein gene; N/A: not available.

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