

Supplementary Material



Figure S1. Experimental installation used for microalgal growth.

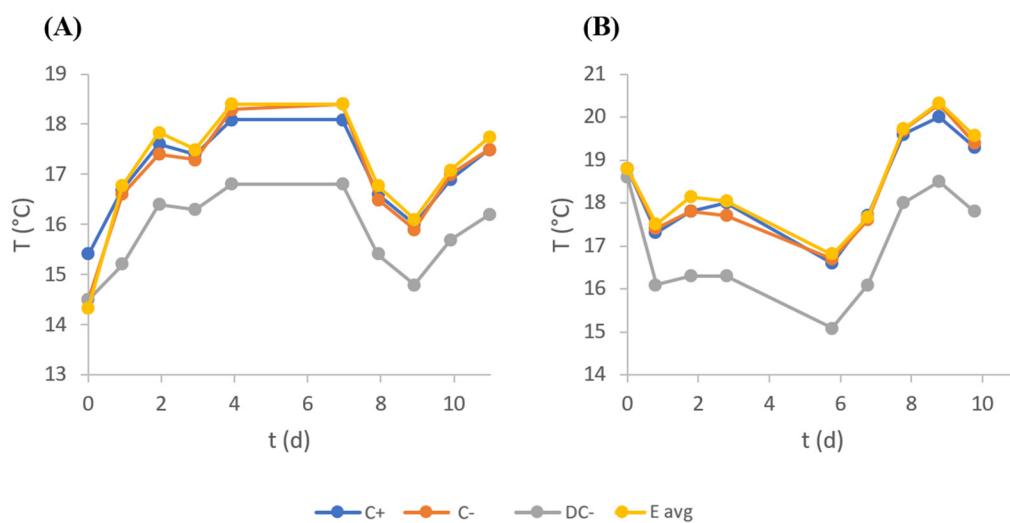


Figure S2. Time-course evolution of temperature inside each flask (A and B represent effluent 1 and 2, respectively).

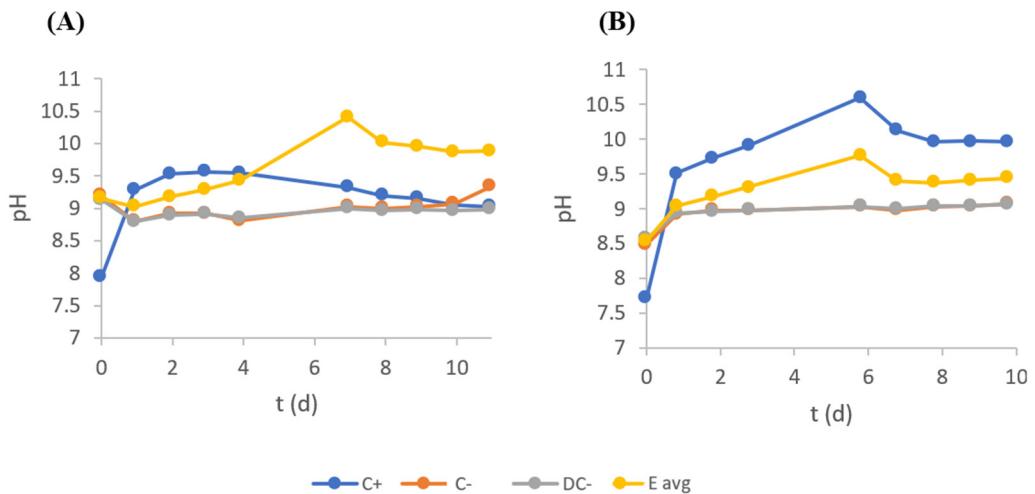


Figure S3. Time-course evolution of pH inside each flask (A and B represent effluent 1 and 2, respectively).

Table S1. All the calibration curves used throughout this study.

Calibration Curve	y	a ± sa	b ± sb	R ²	LOD	LOQ
Biomass concentration (mg _{dw} L ⁻¹)	OD _{680nm}	2.67×10 ⁻³ ± 4.4×10 ⁻⁵	-0.13064 ± 8.83×10 ⁻³	0.999	9.917	33.058
NO ₃ ⁻ (mg L ⁻¹)	Abs _{410nm}	0.0510 ± 0.0008	0.0176 ± 0.0044	0.998	0.26	0.87
PO ₄ ³⁻ (mg L ⁻¹)	Abs _{820nm}	0.184 ± 0.002	0.004 ± 0.003	1.000	0.03	0.19
Colour	Abs _{400nm}	9.28×10 ⁻⁴ ± 7.66×10 ⁻⁶	-0.00223 ± 0.001306	0.999	4.222	14.075
COD – Low Range (mgO ₂ L ⁻¹)	Abs _{420nm}	0.00273 ± 0.000113	-0.0160 ± 0.00587	0.993	6.441	21.470
COD – High Range (mgO ₂ L ⁻¹)	Abs _{600nm}	0.000412 ± 2.75×10 ⁻⁶	-0.00187 ± 0.00148	0.998	10.803	36.011

a: slope of the calibration curve; Abs: Absorbance; b: Intercept; COD: Chemical oxygen demand; LOD: Limit of detection; LOQ: Limit of quantification; OD: Optical density; R²: Coefficient of determination; sa: Standard deviation of the slope; sb: Standard deviation of the intercept.

Table S2. Chemical characterization of each effluent. Values are presented as mean \pm standard deviation.

	Effluent 1	Effluent 2
[NO ₃ -N] (mg L ⁻¹)	0.79 \pm 0.03	1.34 \pm 0.07
[NH ₄ -N] (mg L ⁻¹)	1.30 \pm 0.04	6.8 \pm 0.2
[PO ₄ -P] (mg L ⁻¹)	7.2 \pm 0.2	3.25 \pm 0.06
COD (mg L ⁻¹)	60 \pm 1	583 \pm 14
Turbidity (NTU)	135 \pm 7	100 \pm 0
pH	9.13	8.57
TS (mg L ⁻¹)	4.19 \pm 0.04	5.5 \pm 0.2
TSS (mg L ⁻¹)	0.18 \pm 0.02	0.12 \pm 0.02
Colour (uH)	675 \pm 12	956 \pm 18
DOC (mg L ⁻¹)	135 \pm 7	126 \pm 8
TN (mg L ⁻¹)	11.4 \pm 0.2	14.6 \pm 0.7

COD: Chemical oxygen demand; DOC: Dissolved organic carbon; NH₄-N: Ammonium-nitrogen; NO₃-N: nitrate-nitrogen; PO₄-P: Phosphate-phosphorus; TN: Total nitrogen; TS: Total solids; TSS: Total suspended solids.

Table S3. Kinetic parameters correspondent to the obtained modified Gompertz models.

	Parameter	Effluent 1	Effluent 2
NO ₃ -N	S ₀ (mg L ⁻¹)	24.05	11.10
	k (d ⁻¹)	1.15	0.78
	λ (d)	0.68	2.30
	R ²	0.997	0.998
NH ₄ -N	RMSE (mg L ⁻¹)	0.631	0.292
	S ₀ (mg L ⁻¹)	1.12	6.99
	k (d ⁻¹)	8.35	2.83
	λ (d)	0.00005	0.00005
PO ₄ -P	R ²	1.000	1.000
	RMSE (mg L ⁻¹)	0.001	0.030
	S ₀ (mg L ⁻¹)	7.61	3.02
	k (d ⁻¹)	0.43	2.12
	λ (d)	0.82	0.27
	R ²	0.996	1.000
	RMSE (mg L ⁻¹)	0.338	0.008

k: Uptake rate; RE: Removal efficiency; RMSE: Root mean squared error; R²: Coefficient of determination; S₀: Initial nutrient concentration; λ : Lag time.

Table S4. Biomass specific yields obtained for C+ and E assays. Values are presented as mean \pm standard deviation. Within the same column and Y_{X/S}, average values sharing the same letter (a, b) are statistically different (p < 0.05).

		Effluent 1	Effluent 2
Y _{X/S} (N, g _{biomass} /g _{substrate})	C+	31 \pm 2 ^a	47 \pm 2 ^a
	E	49 \pm 11 ^b	45 \pm 4 ^a
Y _{X/S} (P, g _{biomass} /g _{substrate})	C+	81 \pm 2 ^a	140 \pm 4 ^a
	E	159 \pm 3 ^a	257 \pm 10 ^a

Y_{X/S}: Biomass specific yield.

Table S5. Initial and final COD values obtained for C+, E, C- and DC- assays. Values are presented as mean \pm standard deviation.

	Effluent 1		Effluent 2	
	t_0 (mg L ⁻¹)	t_f (mg L ⁻¹)	t_0 (mg L ⁻¹)	t_f (mg L ⁻¹)
C+	22 \pm 2	86.7 \pm 0.9	42 \pm 13	224 \pm 15
E	75 \pm 2	75 \pm 1	458 \pm 30	556 \pm 34
C-	71 \pm 3	79 \pm 3	456 \pm 18	220 \pm 17
DC-	72 \pm 2	78 \pm 2	497 \pm 6	218 \pm 17

t_x : value obtained on the day x.

Table S6. Initial and final turbidity values obtained for C+, E, C- and DC- assays. Values are presented as mean \pm standard deviation.

	Effluent 1		Effluent 2	
	t_0 (NTU)	t_f (NTU)	t_0 (NTU)	t_f (NTU)
C+	0.90 \pm 0.07	13 \pm 0	2.0 \pm 0.4	26 \pm 3
E	73 \pm 7	25 \pm 2	47 \pm 4	11 \pm 1
C-	60 \pm 0	19.5 \pm 0.7	50 \pm 0	11.5 \pm 0.7
DC-	65 \pm 0	19.5 \pm 0.7	45 \pm 0	13.5 \pm 0.7

t_x : value obtained on the day x.

Table S7. Initial and final colour values obtained for C+, E, C- and DC- assays.

	Effluent 1		Effluent 2	
	t_0 (uH)	t_{11} (uH)	t_0 (uH)	t_{10} (uH)
C+	19	115	23	160
E	480	280	600	440
C-	400	230	680	460
DC-	420	240	520	440

t_x : value obtained on the day x.

Table S8. Initial and final pigment content (in mass percentage) in the microalgal biomass for C+ and E assays. Values are presented as mean \pm standard deviation. Within the same column and for each pigment, average values sharing the same letter (a, b) are statistically different ($p < 0.05$).

		S_0 (% m m ⁻¹)	S_f (% m m ⁻¹)
Chl-a	Effluent 1	C+ 1.49 ± 0.02^a	0.346 ± 0.001^a
		E 0.80 ± 0.02^a	0.62 ± 0.02^a
	Effluent 2	C+ 2.29 ± 0.03^a	0.87 ± 0.03^a
		E 2.4 ± 0.2^b	1.15 ± 0.06^a
Chl-b	Effluent 1	C+ 0.67 ± 0.01^a	0.19 ± 0.01^a
		E 0.41 ± 0.02^a	0.34 ± 0.02^a
	Effluent 2	C+ 0.97 ± 0.01^a	0.41 ± 0.04^a
		E 1.04 ± 0.06^b	0.50 ± 0.01^a
Chl-a + Chl-b	Effluent 1	C+ 2.17 ± 0.03^a	0.53 ± 0.01^a
		E 1.211 ± 0.001^a	0.96 ± 0.04^a
	Effluent 2	C+ 3.25 ± 0.03^a	1.28 ± 0.07^a
		E 3.4 ± 0.2^b	1.66 ± 0.07^a
Carotenoids	Effluent 1	C+ 0.41 ± 0.01^a	0.18 ± 0.01^a
		E 0.22 ± 0.01^a	0.17 ± 0.01^b
	Effluent 2	C+ 0.48 ± 0.01^a	0.25 ± 0.01^a
		E 0.53 ± 0.04^a	0.31 ± 0.02^a

S_0 : Concentration obtained at day 0; S_f : Concentration obtained at the last day.

Table S9. Experimental results obtained in previous studies for textile wastewater treatment using microalgae.

Microalgae Species	Operating Conditions	Wastewater Characteristics	Results Obtained	Ref.
<i>C. vulgaris</i>	Location: Portugal; No dilution (100 %ww); LI = 214 ± 5 μmol m⁻² s⁻¹ ; LDR: 24:0 ; V: 1 L ; Time: 11 d	$\text{NO}_3\text{-N}^*$: $0.79 \pm 0.03 \text{ mg L}^{-1}$; $\text{NH}_4\text{-N}$: $0.290 \pm 0.003 \text{ d}^{-1}$; $P_{\max} = 176 \pm 3.25 \pm 0.06 \text{ mg L}^{-1}$; COD: $60 \pm 1 \text{ mg NO}_3\text{-N RE: } 100 \pm 0 \%$; $\text{NH}_4\text{-N RE: } 1.30 \pm 0.04 \text{ mg L}^{-1}$; Color: $956 \pm 18 \text{ uH}$; TS: $4.19 \pm 0.4 \%$; PO₄-P RE: $99.062 \pm 0.04 \text{ mg L}^{-1}$; Turbidity: $135 \pm 7 \text{ NTU}$	$\mu_{\max}: 0.290 \pm 0.003 \text{ d}^{-1}$; $P_{\max} = 176 \pm 0.005 \%$; Color RE: 42% ; Valorization product: pigments	This study
<i>C. vulgaris</i>	Location: Egypt; Dilutions (5, 8.5, 17.5, 26.5, 30 %ww); LI = 100 μmol m⁻² s⁻¹ ; T = 25 ± 1 °C ; LDR: 12:12 ; pH: 8.05 ; ranging from 6.33 to 380.4 mg L⁻¹; Time: 10 d	COD: $51.2 \text{ mg O}_2 \text{ L}^{-1}$; TS: 735 mg L^{-1} ; TP: 1.51 mg L^{-1} ; Heavy Metals: (Cu, Zn, Cr, Mn, Fe)	$\mu_{\max}: 0.89 \text{ d}^{-1}$ (8.5 %ww); COD RE_{max}: 69.90% (17.5 %ww); Color RE_{max}: 76.32% (17.5 %ww); Valorization product: not quantified	[28]
<i>C. vulgaris</i>	Reactor: Bubble Column; Volume: 1 L; T = 25 ± 2 °C ; Dilutions (0, 25, 50, 75 %ww); Time: 12 d ; pH: 6.5	COD: $755 \pm 20 \text{ mg O}_2 \text{ L}^{-1}$; TS: $6267 \pm 84 \text{ mg L}^{-1}$	$\mu_{\max}: 0.28 \pm 0.07 \text{ d}^{-1}$ (25 %ww); $P_{\max} = 2.91 \pm 0.01 \text{ g L}^{-1} \text{ d}^{-1}$ (25 %ww); COD RE_{max}: 82% (25 %ww); Color RE_{max}: 99 ± 0.13 % (25 %ww); Valorization product: biodiesel	[27]
<i>Chlorella sp. KU211b</i>	Dilutions: (0.5, 1, 2 %ww); LI = 60 μmol m⁻² s⁻¹ ; Agitation: 110 rpm; V: 250 ml ; Time: 2 weeks ; pH: 13	COD: $42442 \pm 453 \text{ mg O}_2 \text{ L}^{-1}$; TN: $374 \pm 12 \text{ mg L}^{-1}$; TP: $79 \pm 4 \text{ mg L}^{-1}$; Turbidity: $2675 \pm 89 \text{ NTU}$; Heavy metals: (Al, Cu, Pb, Se, among others)	$X_{\max}: 0.90 \text{ g L}^{-1}$ (0.5 %ww); Heavy metal RE: 100% (Pb, Se), 45% (Al), 50% (Cu); Color RE_{max}: 71.16% (2 %ww); Valorization product: not quantified	[17]
<i>Chlorella pyrenoidosa</i>	Location: India; Dilution: 75 %ww; V: 500 mL ; T: 28 °C ; LDR: 24:0 ; Time: 15 d	pH: 6.8 ; TS: 5400 ; BOD: $710 \text{ mg O}_2 \text{ L}^{-1}$; TP: 4.7 ; TN: 360	TN RE: $81 \pm 1 \%$; TP RE: $36 \pm 2 \%$; BOD RE: $73 \pm 1.6 \%$; Valorization product: not quantified	[29]

LDR: Light:dark ratio; LI: Light intensity; COD: Chemical oxygen demand; P: Biomass productivity; RE: Removal efficiency; T: Temperature; TS: Total solids; TP: Total phosphorus; TN: Total nitrogen; V: Volume; X: Biomass concentration; WW: Wastewater; μ : specific growth rate; * - nutrient was added before microalgal treatment.