



Supplementary Information for: Vibrational Spectroscopy Coupled to a Multivariate Analysis Tiered Approach for Argentinean Honey Provenance Confirmation



Figure S1. Geographical map of the three Argentinian provinces included in the study.

HN2014	Prediction	
	J>	HN2015
HN2014; HN2015	Prediction	HN2016
HN2014; HN2015; HN2016	Prediction	HN2017
	HN2014; HN2015 HN2014; HN2015; HN2016	HN2014; HN2015 HN2014; HN2015; HN2016 Prediction

**Figure S2.** Scheme of the adopted validation strategy.



**Figure S3.** Recorded raw spectra of FT-MIR (**a**), FT-Raman (**b**) and NIR (**c**). Grey bands show spectral regions excluded from the data analysis.



**Figure S4.** Line plots of PC1 and PC2 loadings obtained from FT-MIR (a), FT-Raman (b) and NIR (c) data.

## **Band** assignment

Investigation of the main peaks/bands relevant for the samples segregation was carried out with the aid of Variable Importance in Projection (VIP) extracted from a PLS model. VIP scores summarize the influence that the original descriptors have had on the PLS model generation. In addition, these scores are scaled in a way that variables exhibiting VIP values greater than 1 can be considered statistically-relevant in a given model. In order to obtain such importance estimates, PLS2-DA models were generated considering all the geographical regions at once (i.e. BA vs Cat vs Mis) on the data coming from the different instrumental techniques. A 5-fold cross validation (venetian blinds) was adopted. Illustration of the VIP scores for the three classification models are reported in **S. Fig 5**.



**Figure S5.** Line plots of the VIP scores extracted from the PLS2-DA (i.e. BA vs Cat vs Mis) model generated on FT-MIR (a), FT-Raman (b) and NIR (c) data. The red dashed line represents the threshold value for significant contribution (VIP=1). Wavenumber/wavelength corresponding to the maximum band absorbance are highlighted.

Concerning the FT-MIR data, as can be seen, Region 1 contains the highest VIP scores, with peaks at 956, 983 and 1006, 1024, 1040, 1061, 1086 and 1104 cm<sup>-1</sup>. According to the literature, Region 1 (1120–940 cm<sup>-1</sup>) corresponds to the C–O and C–C stretching of the three major carbohydrates of honey: fructose, glucose and sucrose <sup>1</sup>. Indeed, the peak at 1040 cm<sup>-1</sup> has been assigned to the C–O stretch in the C–OH group in the carbohydrate structure <sup>2,3</sup>. Furthermore, the small peak at 1104 cm<sup>-1</sup> corresponds to the C–O–C linkage (the C–O–C is present in sucrose as a glycoside bond) <sup>2</sup>. High VIP values can be also observed in Region 2 (920–750 cm<sup>-1</sup>) with peaks at 767, 784, 806, 824, 854 872, 889 and 912 cm<sup>-1</sup>. This is the anomeric region, characteristic of the saccharide configuration <sup>1,2</sup>. In particular, the peak at 912 cm<sup>-1</sup> is due to the C–H bending of the carbohydrate. No statistically relevant bands were highlighted in the 1540–1175 cm<sup>-1</sup> range, which has been assigned by Gok and co-workers to the O–H stretching/bending, C=O stretching of ketones and the C–O & C–H stretching of carbohydrates <sup>4</sup>.

With regard to FT-Raman data, the two peaks at 2936 and 2902 cm<sup>-1</sup> are related to the asymmetric stretching of CH<sub>2</sub> and C-H stretching vibrations, respectively. In addition, several peaks characteristic

of different chemical groups can be observed in the fingerprint region (200-1500 cm<sup>-1</sup>). Most of these signals are consistent with the literature and thorough band assignment can be found in our previous work <sup>5</sup>.

Finally, VIP extracted from NIR data highlighted two statistically-relevant bands, 1400-1490 nm and 1870-1930 nm, both corresponding to O–H, C–H, and C–H<sup>2</sup> deformations <sup>6</sup>. Besides, the spectral range between 2000 and 2100 nm corresponds to the C–H combinations <sup>6,7</sup>, the minor peak at 2332 nm has been related to the C–H bonds <sup>8</sup> and the 2430-2490 nm band is characteristic of the O–H, N–H, and C–H deformation <sup>6</sup>. Furthermore, signals at 1470 and 1960 nm have been associated with the O-H stretch (first and second overtone, respectively) <sup>9</sup>.

Year	Buenos Aires	Catamarca	Misiones
2014	78	25	31
2015	93	11	28
2016	107	15	25
2017	45	17	28

Table	S1.	Samp	le set	overview.
-------	-----	------	--------	-----------

Table S2. PLS-DA prediction results expressed as correct classification rates (FT-Raman data).

Predicted harvest	Correct classification rate (%)		
	BA vs Cat	BA. vs Mis	Cat vs Mis
2015	80.7	95.8	79.4
2016	81.9	96.2	80.0
2017	82.3	97.2	84.4

Table 3. PLS-DA prediction results expressed as correct classification rates (NIR data).

Predicted harvest	Correct classification rate (%)		
	BA vs Cat	BA. vs Mis	Cat vs Mis
2015	70.1	96.6	76.5
2016	80.3	87.1	65.0
2017	88.7	94.5	82.2

## References

- 1. Subari, N.; Mohamad Saleh, J.; Md Shakaff, A.; Zakaria, A. A Hybrid Sensing Approach for Pure and Adulterated Honey Classification. *Sensors* **2012**, *12* (10), 14022–14040. https://doi.org/10.3390/s121014022.
- Gallardo-Velázquez, T.; Osorio-Revilla, G.; Loa, M. Z. de; Rivera-Espinoza, Y. Application of FTIR-HATR Spectroscopy and Multivariate Analysis to the Quantification of Adulterants in Mexican Honeys. *Food Res. Int.* 2009, 42 (3), 313–318. https://doi.org/10.1016/j.foodres.2008.11.010.
- 3. Tewari, J. C.; And, †; Irudayaraj, J. M. K. Floral Classification of Honey Using Mid-Infrared Spectroscopy and Surface Acoustic Wave Based z-Nose Sensor. **2005**. https://doi.org/10.1021/jf050139z.
- Gok, S.; Severcan, M.; Goormaghtigh, E.; Kandemir, I.; Severcan, F. Differentiation of Anatolian Honey Samples from Different Botanical Origins by ATR-FTIR Spectroscopy Using Multivariate Analysis. *Food Chem.* 2015, 170, 234–240. https://doi.org/10.1016/j.foodchem.2014.08.040.
- 5. Fernándz Pierna, J. A.; Abbas, O.; Dardenne, P.; Baeten, V. Discrimination of Corsican Honey by FT-Raman Spectroscopy and Chemometrics. *Biotechnol. Agron. Soc. Environ.* **2011**, *15* (1), 75–84.
- Bisutti, V.; Merlanti, R.; Serva, L.; Lucatello, L.; Mirisola, M.; Balzan, S.; Tenti, S.; Fontana, F.; Trevisan, G.; Montanucci, L.; Contiero, B.; Segato, S.; Capolongo, F. Multivariate and Machine Learning Approaches for Honey Botanical Origin Authentication Using near Infrared Spectroscopy. J. Near Infrared Spectrosc. 2019, 27 (1), 65–74. https://doi.org/10.1177/0967033518824765.
- Woodcock, T.; Downey, G.; Kelly, J. D.; O'Donnell, C. Geographical Classification of Honey Samples by Near-Infrared Spectroscopy: A Feasibility Study. J. Agric. Food Chem. 2007, 55 (22), 9128–9134. https://doi.org/10.1021/jf072010q.
- Latorre, C. H.; Crecente, R. M. P.; Martín, S. G.; García, J. B. A Fast Chemometric Procedure Based on NIR Data for Authentication of Honey with Protected Geographical Indication. *Food Chem.* 2013, 141 (4), 3559– 3565. https://doi.org/10.1016/j.foodchem.2013.06.022.
- Corbella, E.; Cozzolino, D. The Use of Visible and near Infrared Spectroscopy to Classify the Floral Origin of Honey Samples Produced in Uruguay. *J. Near Infrared Spectrosc.* 2005, 13 (2), 63–68. https://doi.org/10.1255/jnirs.458.