



Article Thermal Degradation of Vegetable Oils

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Abstract: Vegetable oils provide lipids and nutrition and provide foods with a desirable flavor, color, and crispy texture when used to prepare fried foods. However, the oil quality is degraded at elevated temperatures, and thus must be examined frequently because of the damage to human health. In this study, sunflower, soybean, olive, and canola oils were examined, and their properties were measured periodically at different elevated temperatures. The unsaturated triglyceride in oils reacted with the environmental oxygen or water vapor significantly changes in optical absorbance, viscosity, electrical impedance, and acid value. We used defect kinetics to analyze the evolution of these oil properties at elevated temperatures. The optical absorbance, viscosity, and electrical impedance follow the second-order, first-order, and zeroth-order kinetics, respectively. The rate constants of the above kinetics satisfy the Arrhenius equation. Olive oil has the lowest rate of color center and dynamic viscosity among the four oils, with the smallest pre-exponential factor and the largest activation energy, respectively. The rate constants of acid reaction also satisfy the Arrhenius equation. The activation energies of the polar compound and acid reaction are almost the same, respectively, implying that the rate constant is controlled by a pre-exponential factor if four oils are compared. Olive oil has the largest rate constant of acid reaction among the four oils, with the lowest pre-exponential factor.

Keywords: vegetable oil; optical absorbance; dynamic viscosity; electric impedance; acid value



Frying is a food process in which the foods are immersed in hot oils from 150 °C to 200 °C to achieve the desired flavor, color, and crispy structure. Thus, oils play a significant role in frying, contributing to the nutrition, taste, smell, and texture of foods.

A major component in vegetable oils is triglycerides, which consist of three fatty acid chains connected to a glycerol backbone through the carboxyl group [1]. All the double bonds are positioned in the cis configuration in fresh vegetable oils. The reaction of oils during frying in air and water moisture is complicated. Chemical reactions, such as hydrolysis, oxidation, and polymerization [2], produce harmful compositions such as aldehyde, ketone, trans-fatty acid, and polycyclic aromatic hydrocarbons due to triglyceride oxidation during frying. In addition to causing foul odor and opaqueness, those harmful compounds have been reported to be relevant to eicosanoid metabolism, elevated oxidation stress [3], lipid and glucose metabolism, liver malfunction in rats [4], and potential for cancers in humans [5,6]. In addition to the chef's experience in deciding whether the oils are discarded based on coloring, smoking, odd smells, or taste, we need instruments to determine the degradation of oils caused by heating.

Several instruments and methods are available to evaluate the quality of oil regarding its chemical compositions and physical properties. Fourier transform infrared spectroscopy (FTIR) with a database is a powerful instrument for identifying polar chemical compounds [7–9]. Chromatography with mass spectroscopy detects the quantity of total polar compounds of oils in vapor [10–12]. UV-vis spectrophotometry is another versatile instrument used to evaluate the compound change during frying via the color center [13–15].



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Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). As the thermal cycles of heating and cooling progress, the viscosity of oils increases due to the transformation of microstructure from unsaturated fat chains to long polymerization and blended chains. The viscosity meter effectively measures the change in the viscosity [16,17]. The titration determines the acid value of the fried oils, in which most governments enact standard values to discard deteriorated vegetable oils. In legislative regulation [13–19], the corrupted acid value ranges from 2 to 3 mg KOH/g, depending on the system of frying food and vegetable oil. However, the favored method reported in the literature [20–22] is an LCR meter with interdigital sensors of different geometries. The LCR measures electric impedance, capacitance, and dielectric constants, which are powerfully relevant to the degraded properties other than optical transmittance, viscosity, and acid value. A tested paper [19] and a hand-held device [23] are also commercially available to detect the acid value or total polar compounds in oils through color and digital monitoring, respectively.

Using commercially available electronic components, Wang et al. made a cost-effective capacitance sensor [24]. The voltages obtained from their column chromatography measurements had a linear relationship with the total polar compounds of the tested edible oils. Yang et al. [25] summarized a table comparing the advantages and disadvantages of various methods used to characterize oil quality. However, no literature explained why the changes in optical transmittance, viscosity, and electric impedance of oils during heating, which prompted us to study the thermal degradation of vegetable oils. Four types of oil, including sunflower, soybean, olive, and canola, were analyzed. Based on the viewpoint of materials science, we explained the evolution of optical transmittance, viscosity, and electric impedance of vegetable oils using defect mechanism theory. The activation energies of the oils were obtained, and their acid values were evaluated.

2. Experimental Section

Canola, sunflower, and olive oils were purchased from Taisun Enterprise Co. (Taipei City, Taiwan), and soybean oil from Taiwan Sugar Corp. (Hsinchu City, Taiwan). The oils were sealed and stored on a cool shelf to avoid exposure to the sun before the tests.

First, 125 mL of oil was poured into a 250 mL beaker in a silicon–oil bath to maintain a constant annealing temperature. The annealing temperatures were 140, 160, 180, and 200 °C, and the dwell time varied from 2 to 300 h, depending on the measurement method. The beaker was removed from the thermostatic bath and cooled to 25 °C for the test. When the test was not operated, the oil was stored in a refrigerator to slow the thermal degradation.

The UV-visible transmittance of oils was measured on an ultraviolet-visible spectrophotometer (U-3900, Hitachi High-Technologies Co., Tokyo, Japan). Two cuvettes were used to correct the baseline. The baseline correction of wavelengths was scanned from 190 nm to 1300 nm with a scan rate of 300 nm/min. After finishing baseline correction, the oil-filled cuvette and empty cuvette are executed for the transmittance measurement. The scanning wavelength ranges from 300 nm to 800 nm, and the scanning speed is 300 nm/min.

The dynamic viscosity of oils of 15 mL was determined at 25 °C on a 7990S Cannon-Ubbelohde glass viscometer (Ramin Corp., Magnolia, TX, USA). The reported viscosities are average values of five measurements. The measurement of the electrical impedance of heated oils was performed on an 8110G LCR meter (Good Well Electrics Industrial Co., New Taipei City, Taiwan) with a homemade interdigital sensor (see Reference [26] for the construction of the interdigital sensor).

A cosolvent of 25.0 mL ethanol (Taiwan Sugar Corp., Tainan City, Taiwan) and 25.0 mL diethyl ether (Echo Chemical Co., Miaoli, Taiwan) was prepared as a blank titration solvent to dissolve the oil. The oil of 10.0 g was mixed with 50.0 mL blank solvent. A few drops of phenolphthalein (Echo Chemical Co., Miaoli, Taiwan) were added to the mixed solution as an indicator. The transparent base titrant was 0.010 M KOH. The titration was ended after

the color of the solution changed to pink for more than 30 s. The acid value was calculated using Equation (1) below.

Acid value =
$$\frac{(\Omega - \Omega_0) \times C \times M}{W}$$
 (1)

where Ω is the volume of the consumed titrant KOH after the titration, Ω_0 is the volume of titrant KOH for a blank titration solvent, *C* is the concentration of the base titrant (0.010 mol/L), *M* is the molecular weight of titrant (56.1 g/mol), and W is the oil weight (10.0 g).

The chemical bonds of heated oils were analyzed on a Perkin Elmer Spectrum 100 FTIR (Perkin Elmer Co., Shelton, CT, USA) with an attenuated total reflection mode. The scanning wavenumber range was from 450 to 4000 cm⁻¹.

The experimental data of ultraviolet-visible spectroscopy, dynamic viscosity, electrical impedance, and acid values are analyzed using the least square method. The least square method with confidence interval R² was provided by the graphical software Origin (Electrical Art Inc., Redwood City, CA, USA).

3. Results and Discussion

3.1. Ultraviolet-Visible Spectroscopy

Figure 1a shows the transmittance spectra for soybean oil maintained at 180 °C at different times. The transmittance spectra exhibit S-like behavior with slow increase initially, rapid increase intermediately, and slow increase with increasing wavelength to reach a plateau. Increasing the heating time causes the right shift of the S-like spectrum. The transmittance spectra of the soybean oils heated at 140, 160, and 200 °C are presented in Figure S1a–c in Supplementary Information, respectively. According to Figures 1a and S1a–c, the time to reach the steady-state transmittance decreases with increasing the heating time, implying that the evolution of transmittance is associated with a thermally activated process. A similar trend for the transmittance spectra is observed for canola, sunflower, and olive oils, as shown in Figures S2a–d, S3a–d, and S4a–c in Supplementary Information, respectively. The small peak for the wavelength at 670 nm observed in the transmittance spectra of the olive oil, as shown in Figures 1b and S4a–c, is attributed to chlorophyll [14,27], whose peak increases with increasing time and temperature because chlorophyll is thermally unstable.



Figure 1. The variation of transmittance with the wavelength at 180 °C for different times: (**a**) soybean oil and (**b**) olive oil.

Define the cutoff wavelength as the maximum wavelength without transmittance. Figure 2 shows the temporal evolution of the cutoff wavelength for the sunflower oil at different temperatures, obtained directly from Figure 1a and Figure S3 in the Supplementary Material. The cutoff wavelength exhibits a redshift with increasing time and temperature. Similar behavior is observed for the cutoff wavelengths of the soybean, canola, and olive oil, as shown in Figure S5 in Supplementary Material, and are 320 nm to 440 nm. Canola



oil has the largest cutoff wavelength.

Figure 2. The relationship between the cutoff wavelength and times for the sunflower oil at different temperatures.

From the viewpoint of energy conservation, the summation of reflectance (I_R) , absorbance (A), and transmittance (I_T) is equal to unity. [28] Since the incident UV-visible light is normal on the specimen surface, the reflectance is negligible. Additionally, we are interested in the difference between absorbances at the initial state and the state after the heating for the dwell of t, which is equal to the difference between the corresponding two transmittances if other conditions are maintained at the same level. That is, the non-zero reflectance has a negligible effect on the absorbance difference. Figure 3 shows the variation in the absorbance difference $(A - A_0)$ at the wavelength of 445 nm with the heating time for the soybean oil heat-treated at 140, 1620,180, and 200 $^{\circ}$ C. Here, A₀ and A are the absorbances at the initial state and the heating time t. Note that the absorbance A is equal to $(1-I_T)$, which is different from that obtained using the Lambert–Beer law (i.e., $A = -Log I_T$). For each curve, the slope increases slowly initially, reaches constant, and then decreases to a plateau with the increasing heating time. The higher the temperature, the shorter the time to the plateau. Similar behavior is observed for other oils, as shown in Figures S6a-c in the Supplementary Information. At the same time, the absorbance increases with increasing temperature for all the oils.



Figure 3. The variation of absorbance $(A - A_0)$ of soybean oil at 445 nm with annealing time at different temperatures. The solid lines are obtained using Equation (4).

Assume that the evolution of absorbance is controlled by the generation of color centers, which second-order kinetics can describe. The differential equation for second-order kinetics is expressed as

$$\frac{dn}{dt} = \alpha_A (n_\infty - n)(1 - \beta(n_\infty - n)) \tag{2}$$

where *n* is the concentration of color centers, n_{∞} is the concentration at infinite time, α_A is the rate constant, β is the proportional constant related to absorbance, and t is the elapsed time. Note that the maximum absorbance is unity. The solution of Equation (2) with the initial condition, $n = n_0$ at $t = t_0$, is given as

$$\frac{n_{\infty} - n}{n_{\infty} - n_0} = \frac{e^{-\alpha_A t}}{1 + \beta (n_{\infty} - n_0)(e^{-\alpha_A t} - 1)}$$
(3)

Since $A = \beta n$ and the initial condition $A = A_0$ at t = 0, Equation (3) is rearranged as

$$\frac{A - A_0}{A_\infty - A_0} = 1 - \frac{e^{-\alpha_A t}}{1 + (A_\infty - A_0)(e^{-\alpha_A t} - 1)}$$
(4)

For the soybean oils, the solid lines in Figure 3 are obtained using Equation (4) with parameters listed in Table 1. Similarly, the parameters for the other oils used in Equation (4) fit the absorbance data in Figure S6a–c are shown in Tables S1–S3 in Supplementary Material, respectively. The rate constant of every vegetable oil increases with the annealing temperature.

Table 1. Parameters α_A , A_0 , and A_∞ used in Equation (4) to fit the absorbances of soybean oil at four annealing temperatures where R^2 is the confidence interval.

Temperature (°C)	$lpha_A$ (h $^{-1}$)	A ₀	A_∞	<i>R</i> ²
200	0.091	0.0362	0.9632	0.998
180	0.068	0.0852	0.9951	0.997
160	0.047	0.0867	0.9774	0.996
140	0.035	0.0321	0.9675	0.997

The logarithm plot of the rate constant with the reciprocal absolute temperature is shown in Figure 4. According to Figure 4, the rate constant satisfies the Arrhenius equation,

$$\alpha_A = \alpha_{A0} \exp\left(\frac{-Q_A}{RT}\right) \tag{5}$$

where α_{A0} is the pre-exponential factor, Q_A is the activation energy, R is the gas constant, and T is the Kelvin temperature. The activation energy and pre-exponential factor of absorbance for all oils are tabulated in Table 2. Canola oil has the largest pre-exponential factor and activation energy among the four types of oil. The descending activation energy of four types of vegetable oils is sunflower oil, canola oil, soybean oil, and olive oil, respectively. The higher the activation energy, the more resistance the chemical reaction induces to the color center and optical absorbance. However, the pre-exponential factor also plays a vital role in color center kinetics, which is a trade-off for the activation energy. Comparing Table 2 and Figure 5, canola oil has the lowest rate constant and largest pre-exponential factor and activation energy among the four oils, implying that the rate constant is dominated by activation energy of optical absorbance, suggesting that activation energy controls the rate constant.



Figure 4. The log (α_A) plots versus 1000/T for sunflower, canola, soybean, and olive oils.

Table 2. The activation energies and pre-exponential factors of optical absorbance for four types of oils.

Oil Type	$lpha_{A0}$ (h $^{-1}$)	Q_A (kJ/mol)	R^2
Soybean	58.34	25.44	0.995
Canola	69.18	26.17	0.994
Olive	14.58	20.99	0.998
Sunflower	31.98	23.40	0.997



Figure 5. The variations of dynamic viscosity with the annealing time for soybean oil at different temperatures.

3.2. Dynamic Viscosity

The variations of dynamic viscosity with the annealing time for soybean oils at different temperatures are exhibited in Figure 5. A similar trend is found for the other three types of vegetable oils, as shown in Figure S7a–c in the Supplementary Material. The dynamic viscosity increases exponentially with the annealing time at a given temperature, implying the defects hindering the oil flow. The higher the temperature, the shorter time needed for the oil to arrive at a given viscosity, e.g., 0.5 Pa·s.

The defect hindering the molecular motion is assumed to contribute to the dynamic viscosity. The defect concentration, n_v , follows the first-order generation kinetics as shown below

$$\frac{dn_v}{dt} = \alpha_v n_v \tag{6}$$

where α_v is the rate constant of dynamic viscosity and *t* is the annealing time. The solution of Equation (6) with the initial concentration, n_{v0} , at t = 0 is obtained as

$$n_v = n_{v0} \exp(\alpha_v t),\tag{7}$$

Further, we assumed that the total dynamic viscosity φ consists of a thermally independent component, φ_R , and the thermal component, $\varphi - \varphi_R$. The thermal component of dynamic viscosity is linearly proportional to the defect concentration as $\varphi - \varphi_R = \beta n_v$, where β is a constant. Using this relation, Equation (7) is rearranged as

$$\varphi = \varphi_R + (\varphi_0 - \varphi_R) e^{\alpha_v t} \tag{8}$$

The solid lines in Figure 5 for the soybean oil are obtained using Equation (8) with the parameters φ_0 , φ_R , and α_v listed in Table 3. The corresponding parameters used for the other three vegetable oils are in Tables S4–S6 in the Supplementary Material, respectively. According to Table 3 and Tables S4–S6 in the Supplementary Material, olive and canola oils are the first two largest φ_0 and φ_R among the four oils, likely attributed to chlorophyll and carotene [14,27].

Table 3. The parameters φ_0 , φ_R , and α_v yield the solid lines in Figure 6 for the soybean oil.

Annealing Time (°C)	$arphi_0$ (10 $^{-2}$ Pa \cdot s)	$arphi_R$ (10 $^{-2}$ Pa \cdot s)	$\varphi_{0-}\varphi_{R}(10^{-2} \text{ Pa}\cdot \text{s})$	α_v (10 ⁻² h ⁻¹)	R^2
200	5.743	5.015	0.728	4.434	0.993
180	5.208	4.455	0.753	3.476	0.992
160	5.377	4.582	0.795	2.625	0.991
140	5.648	5.011	0.637	1.699	0.998



Figure 6. The Arrhenius plots of dynamic viscosity for four vegetable oils.

Using Table 3 and Tables S4–S6 in Supplementary Material, the plots of Log α_v versus 1/T for the soybean, canola, olive, and sunflower oils are shown in Figure 6. The rate constants, α_v , are fitted by the following Arrhenius equation,

$$\alpha_v = \alpha_{v0} \exp\left(\frac{-Q_v}{RT}\right) \tag{9}$$

where α_{v0} and Q_v are the pre-exponential factor and activation energy of dynamic viscosity, respectively. *R* and *T* follow the conventional definition. The experimental data are in good agreement with the theoretical prediction. The corresponding activation energies and pre-exponential factors for the four types of vegetable oils are tabulated in Table 4. The oil with a higher pre-exponential factor possesses a greater amount of activation energy. According to Equation (9), the rate constant increases with the increasing pre-exponential factor and decreasing activation energy, implying that the pre-exponential factor is a trade-off for the

activation energy. Comparing Figure 6 and Table 4, the lower the rate constant, the greater the activation energy and pre-exponential factor. Olive oil has the lowest rate constant and largest activation energy and pre-exponential factor of dynamic viscosity, implying that the evolution of dynamic viscosity is controlled by the activation energy. Similarly, sunflower oil has the greatest rate constant and smallest pre-exponential factor and activation energy among the four oils, implying that the evolution of dynamic viscosity is controlled by thermally activated defect kinetics.

Oil Type	$lpha_{v0}$ (h $^{-1}$)	Q_v (kJ/mol)	<i>R</i> ²
Olive oil	88.7	29.90	0.996
Canola oil	43.8	27.14	0.997
Soybean oil	30.4	25.77	0.996
Sunflower oil	17.8	23.58	0.998

Table 4. The activation energies and pre-exponential factors of viscosity for four types of oils.

3.3. Electrical Impedance

The vegetable oils are dielectric materials, and their electrical impedances are measured by an LCR meter with an interdigital sensor of 100 μ m. The frequency range is selected from 1000 Hz to 3000 Hz. The relations between electrical impedance and frequency for sunflower oil under the applied voltage of 1 V at 200 °C are exhibited in Figure 7. The variations in impedance with frequency for the other three oils annealed at 200 °C and 160 °C for the different times are displayed in Figures S8 and S9 in the Supplementary Material. The impedance decreases with the increase in the annealing time and frequency for every oil. The impedance changing from *t* = 0 to 100 h at the given frequency is the largest for sunflower oil, followed by soybean oil, olive oil, and canola oil.



Figure 7. The impedance variations with frequency for sunflower oil annealed at 200 °C at different times.

From Figure 7 and Figure S8 in Supplementary Material, we plotted the impedance versus annealing time for the four oils measured at 1000 Hz and annealed at 200 °C in Figure 8, where I_0 is the initial impedance. Note that the initial impedances are listed in Table S7 in Supplementary Material, which was extrapolated from the impedances of the short times at 1000 Hz. The initial impedances are almost the same for all types of oil and the annealing temperatures. Figure S10a–c show the variations in impedance with annealing time for oils measured at 1000 Hz and annealed at (a) 180 °C, (b) 160 °C, and 140 °C, respectively. The impedance decreases linearly with the annealing time. Sunflower oil has the steepest slope, and canola oil has the smallest one.



Figure 8. The variations in impedance versus annealing time for the four types of oil annealed at 200 °C.

Assume that the positive charges (or negative charges) of electric dipole in the oil consist of two components: one arises from thermal annealing, P_A , and the other is generated from sources other than thermal annealing, P_0 . Then, the positive charges P are the sum of P_A and P_0 . Further, assume that P_A is linearly proportional to the concentration of defects carrying charges, i.e., $P_A = \beta n_E$, in which β and n_E are the proportional constant and defect concentration, respectively. The defect concentration contributing to the induced charges follows the zeroth-order kinetics, $dn_E/dt = \alpha_1$. The solution of this equation with the initial free concentration is $n_E = \alpha_1 t$. The positive charges can be written as $P = P_0 + \beta \alpha_1 t = P_0 + \beta_E t$ where β_E is the charge rate constant. The electrical impedance is expressed as $I = \sqrt{r^2 + 1/\omega^2 C^2}$, where r, ω , and C are resistance, angular frequency, and capacitance, respectively. Substituting $C = P/V = (P_0 + \beta_E t)/V$ into the above equation, we have $I = \sqrt{r^2 + V^2/\omega^2(P_0 + \beta_E t)^2}$. Note that V is the applied voltage. Using Taylor's series expansion to the first-order approximation, the impedance is as follows,

$$\frac{I}{I_0} = \frac{\sqrt{r^2 + \frac{V^2}{\omega^2 P_2^2}}}{\sqrt{r^2 + \frac{V^2}{\omega^2 P_0^2}}} \sim 1 - \frac{\beta_E t}{(1 + r^2 \omega^2 P_0^2 / V^2) P_0} = 1 - \alpha_E t$$
(10)

Since *r*, ω , *V*, and *P*₀ are temperature-independent parameters, α_E is a rate constant and satisfies the Arrhenius equation,

$$\alpha_E = \alpha_{E0} \exp(-Q_E / RT) \tag{11}$$

where Q_E and α_{E0} are the activation energy and pre-exponential factor of the impedance of the degraded oils, and R and T have their ordinary meanings.

The plots of the logarithm of the rate constant versus the reciprocal absolute temperature for different oils are shown in Figure 9. Using Equation (11) to fit the data in Figure 9, one obtains the pre-exponential factors and activation energies and lists them in Table 5. The activation energies of all oils are the same within the error bar, implying that the energy barriers for the polar compound reaction are identical for all oils. Comparing Figure 9 and Table 5, the rate constant of impedance increases with the increase in the pre-exponential factor. The oils with decreasing pre-exponential factors of the polar compound are sunflower, soybean, olive, and canola oils. Since the pre-exponential factor is time independent, sunflower oil always has the largest rate constant of impedance, and canola oil has the smallest.



Figure 9. The Arrhenius plots of the impedance for four types of vegetable oils.

Table 5. The pre-exponential factor and activation energy of electrical impedance for four types of oils.

Oil Type	α_{E0}	Q_E (kJ/mol)	R^2	
Soybean oil	1.27	28.44 (±0.53)	0.997	
Canola oil	0.90	$28.44 (\pm 0.49)$	0.998	
Olive oil	1.20	$28.66 (\pm 0.58)$	0.995	
Sunflower oil	1.32	28.32 (±0.45)	0.997	

3.4. Acid Value

The vegetable oils exposed to the harsh environment of high temperature for a long duration can generate free fatty acid and glycerin. The triglyceride in oils absorbs and reacts with the water vapor in the air, and its hydrolysis reaction is described as

$$C_3H_5(COOR)_3 + 3H_2O \rightarrow C_3H_5(OH)_3 + 3RCOOH$$

The higher the amount of free fatty acid in the oil, the worse the degraded oil, and the more consumable the titrant solution to neutralize the free acid. The variations in the acid value versus the time for the oils annealed at 200 °C are illustrated in Figure 10. The plots of acid value versus annealing time for all oils annealed at 180 °C, 160 °C, and 140 °C are shown in Figure S11a–c of the Supplementary Material, respectively. The acid value increases slowly in the beginning, then accelerates with the increase in the annealing time for all oils. Olive oil produces the easiest acid value, and soybean oil is the most difficult among the four types of oil.

According to the oil rancidification generation and legislation regulation [18,19], when the acid value of vegetable oil exceeds 2.0 mg KOH/g, the oil is discarded. Let t_A be the time when the oil reaches the acid value of 2.0 mg KOH/g. One obtains t_A from Figure 10 and Figure S11 in Supplementary Material and illustrates them in Figure 11. The rate constant of acid reaction equals $1/t_A$. Figure 11 shows the log $(1/t_A)$ plots versus (1/T)for all oils. It can be seen from Figure 11 that $1/t_A$ satisfies the Arrhenius equation, i.e., $\frac{1}{t_A} = \left(\frac{1}{t_{A0}}\right) \exp\left(-\frac{Q_{AC}}{RT}\right)$, where t_{A0} and Q_{AC} are the pre-exponential factor and activation energy for the acid reaction, and R and T are the gas constant and absolute temperature, respectively. Note that the definition of pre-exponential factor for the acid reaction is different from those for color center, dynamic viscosity, and polar compound. According to the above equation, the rate constant of an acid reaction is inversely proportional to the pre-exponential factor t_{A0} and activation energy. Table 6 lists the t_{A0} and activation energies of acid reaction for all oils obtained from Figure 11. The activation energies for olive, canola, soybean, and sunflower oil are almost equal within ± 1.0 kJ/mol. Comparing Table 6 and Figure 11, the pre-exponential factor independent of temperature controls the acid reaction. The oils with increasing pre-exponential factor t_{A0} of acid reaction are olive, canola, sunflower, and soybean. Olive oil has the largest rate constant because its t_{A0} is the smallest one.



Figure 10. The variations of the acid value with annealing time for the oils annealed at 200 °C.



Figure 11. The log $(1/t_A)$ variations with (1/T) for all oils.

Table 6. The activation energies and t_{A0} of the actual feaction for four ons.

Oil Type	<i>t</i> _{A0} (h)	Q _{AC} (kJ/mol)	<i>R</i> ²
Olive	0.0192	30.9	0.997
Canola	0.0239	30.4	0.996
Sunflower	0.0369	29.0	0.995
Soybean	0.0392	29.1	0.996

3.5. Fourier Transform Infrared Spectroscopy (FTIR)

The FTIR spectra of all oils without heat treatment are shown in Figure 12, with their chemical bonds listed in Table 7. Because triglyceride is a significant component in all vegetable oils, the spectral features of the FTIR spectroscopy look very similar, with slight minor differences in peak height and width to reflect the unique character of each oil type. The major peaks at wavenumbers 720, 1110, 1120, 1162, 1244, 1399, 1464, 1745, 2854, 2924, and 3008 cm⁻¹ coincided for all oils, implying that the oils exhibit the same molecular structure reported in references [9,28].



Figure 12. The FTIR spectra of all oils without heat treatment.

Table 7. The peak posit	ons with theiı	r chemical bonds
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Wavenumber (cm ⁻¹)	Description
3008 cm^{-1}	C-H stretching vibration of the <i>cis</i> -double bond (=CH)
2924 cm^{-1} , 2854 cm^{-1}	Symmetric and asymmetric stretching vibration of the aliphatic CH_2 group
$1745 {\rm cm}^{-1}$	Ester carbonyl functional group of the Triglycerides
1464 cm^{-1}	Bending vibrations of the CH ₂ and CH ₃ aliphatic groups
$1417 {\rm cm}^{-1}$	Rocking vibrations of CH bonds of <i>cis</i> -disubstituted olefins
1399 cm^{-1}	Bending in-ddplane vibrations of CH <i>cis</i> -olefinic groups
1323 cm^{-1}	The extra peak appears in mixed oil.
1244 cm^{-1} , 1162 cm $^{-1}$, 1100 cm $^{-1}$	Stretching vibration of the C-O ester Groups
1120 cm^{-1}	Aliphatic ethers
840.5 cm^{-1}	trisubstituted alkenes
$720 \mathrm{~cm}^{-1}$	Overlapping of the CH ₂ rocking vibration and the out-of-plane vibration of <i>cis</i> -disubstituted olefins

4. Conclusions

Edible oil quality is essential to human health. Soybean, canola, olive, and sunflower oils are studied. The UV-vis transmittance, chemical acid value (titration), dynamic viscosity, and electrical impedance measurements are used to evaluate the oil degradation at elevated temperatures. The cutoff wavelength is redshift, and its shifting rate increases with the increase in annealing time. The curves of the optical absorbance at a given wavelength versus annealing time grow slowly at short times, rapidly at intermediate times, and slowly again until they approach a plateau at long times. The dynamic viscosity of the vegetable oils increases exponentially with the annealing time at a given temperature. The electric impedance decreases with the frequency and annealing time. The acid value increases exponentially in the early stage and then keeps a roughly constant rate. We propose second-order, first-order, and zeroth-order kinetics to describe the evolutions of optical absorbance, dynamic viscosity, and electrical impedance. The rate constants satisfy the Arrhenius equation. Olive oil has the lowest rates of color centers and dynamic viscosity among the four oils because it has the smallest pre-exponential factor for the former and the largest activation energy for the latter, respectively. The reciprocal of time to reach acid value also satisfies the Arrhenius equation. The activation energies of the polar compound and acid reaction, respectively, are identical for all oils to reflect the same evolution path. Olive oil has the largest rate constant among the four oils with the smallest pre-exponential factor, t_{A0} .

Supplementary Materials: The following supporting information can be downloaded at: https: //www.mdpi.com/article/10.3390/foods12091839/s1, Figures S1–S4: UV-visible transmittance vs. wavelength for soybean, canola, sunflower, and olive oils, respectively. Figure S5: cutoff wavelengths, Figure S6: UV-visible absorbance, Figure S7: dynamic viscosity, Figures S8 and S9: electrical impedances vs. frequency at 200 and 160 °C, respectively. Figure S10: impedance vs. annealing time at frequency 1000 Hz. Figure S11: acid value vs. time. Tables S1–S3: optical absorbance parameters α , A_0 , and A_∞ for canola, olive, and sunflower, respectively, Tables S4–S6: viscosity parameters φ_0 , φ_R , and α_v for canola, olive, and sunflower oils, respectively, Tables S7: oil initial impedances at frequency 1000 Hz.

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References

- Dupain, X.; Costa, D.J.; Schaverien, C.J.; Makkee, M.; Moulijn, J.A. Cracking of a rapeseed vegetable oil under realistic FCC conditions. *Appl. Catal. B Environ.* 2007, 72, 44–61. [CrossRef]
- 2. Choe, E.; Min, D. Chemistry of deep-fat frying oils. J. Food Sci. 2007, 72, R77–R86. [CrossRef] [PubMed]
- Yen, P.-L.; Chen, B.-H.; Yang, F.L.; Lu, Y.-F. Effects of deep-frying oil on blood pressure and oxidative stress in spontaneously hypertensive and normotensive rats. *Nutrition* 2010, 26, 331–336. [CrossRef] [PubMed]
- 4. Li, X.; Yu, X.; Sun, D.; Li, J.; Wang, Y.; Cao, P.; Liu, Y. Effects of Polar Compounds Generated from the Deep-Frying Process of Palm Oil on Lipid Metabolism and Glucose Tolerance in Kunming Mice. *J. Agric. Food Chem.* **2016**, *65*, 208–215. [CrossRef] [PubMed]
- Cam, A.; Oyirifi, A.B.; Liu, Y.; Haschek, W.M.; Iwaniec, U.T.; Turner, R.T.; Engeseth, N.J.; Helferich, W.G. Thermally Abused Frying Oil Potentiates Metastasis to Lung in a Murine Model of Late-Stage Breast CancerThermally Abused Frying Oil Potentiates Me-tastasis to Lung. *Cancer Prevent. Res.* 2019, *12*, 201–210. [CrossRef]
- 6. Stott-Miller, M.; Neuhouser, M.L.; Stanford, J.L. Consumption of deep-fried foods and risk of prostate cancer. *Prostate* 2013, 73, 960–969. [CrossRef]
- 7. Moros, J.; Roth, M.; Garrigues, S.; de la Guardia, M. Preliminary studies about thermal degradation of edible oils through attenuated total reflectance mid-infrared spectrometry. *Food Chem.* **2008**, *114*, 1529–1536. [CrossRef]
- 8. Uncu, O.; Ozen, B.; Tokatli, F. Use of FTIR and UV–visible spectroscopy in determination of chemical characteristics of olive oils. *Talanta* **2019**, 201, 65–73. [CrossRef]
- De la Mata, P.; Dominguez-Vidal, A.; Bosque-Sendra, J.M.; Ruiz-Medina, A.; Cuadros-Rodríguez, L.; Ayora-Cañada, M.J. Olive oil assessment in edible oil blends by means of ATR-FTIR and chemometrics. *Food Control* 2012, 23, 449–455. [CrossRef]
- 10. Romano, R.; Giordano, A.; Le Grottaglie, L.; Manzo, N.; Paduano, A.; Sacchi, R.; Santini, A. Volatile compounds in intermittent frying by gas chromatography and nuclear magnetic resonance. *Eur. J. Lipid Sci. Technol.* **2013**, *115*, 764–773. [CrossRef]
- 11. Caldwell, J.D.; Cooke, B.S.; Greer, M.K. High Performance Liquid Chromatography–Size Exclusion Chromatography for Rapid Analysis of Total Polar Compounds in Used Frying Oils. *J. Am. Oil Chem. Soc.* **2011**, *88*, 1669–1674. [CrossRef]

- 12. Lazzerini, C.; Cifelli, M.; Domenici, V. Pigments in Extra-Virgin Olive Oil: Authenticity and Quality. In *Products from Olive Tree*; Books on Demand: McFarland, WI, USA, 2016. [CrossRef]
- 13. Zhang, W.; Li, N.; Feng, Y.; Su, S.; Li, T.; Liang, B. A unique quantitative method of acid value of edible oils and studying the impact of heating on edible oils by UV–Vis spectrometry. *Food Chem.* **2015**, *185*, 326–332. [CrossRef]
- Khosroshahi, M.E. Effect of Temperature on Optical Properties of Vegetable Oils Using UV-Vis and Laser Fluorescence Spectroscopy. Opt. Photon. J. 2018, 8, 247–263. [CrossRef]
- 15. Jolayemi, O.S.; Tokatli, F.; Ozen, B. UV–Vis spectroscopy for the estimation of variety and chemical parameters of olive oils. *J. Food Meas. Charact.* **2021**, *15*, 4138–4149. [CrossRef]
- Li, X.; Wu, X.; Liu, R.; Jin, Q.; Wang, X. Effect of frying conditions on fatty acid profile and total polar materials via viscosity. J. Food Eng. 2015, 166, 349–355. [CrossRef]
- 17. Lioumbas, J.S.; Ampatzidis, C.; Karapantsios, T.D. Effect of potato deep-fat frying conditions on temperature dependence of olive oil and palm oil viscosity. *J. Food Eng.* **2012**, *113*, 217–225. [CrossRef]
- 18. Firestone, D. Regulation of frying fat and oil. In Deep Frying; Elsevier: Amsterdam, The Netherlands, 2007; pp. 373–385.
- Advantec Company. Test Strips for Thermal Degradation of Frying Oils. Available online: https://www.advantec.co.jp/uploads/ en/service_support/download/catalog/107-E-01-20060_AV-CHECK_A4.pdf (accessed on 6 July 2022).
- Šegatin, N.; Pajk Žontar, T.; Poklar Ulrih, N. Dielectric properties and dipole moment of edible oils subjected to 'frying' thermal treatment. *Foods* 2020, 9, 900. [CrossRef]
- Fendri, A.; Kallel, A.Y.; Nouri, H.; Ghariani, H.; Kanoun, O. Measurement System for Lossy Capacitive Sensors: Application to Edible Oils Quality Assessment. Sensors 2019, 19, 4299. [CrossRef]
- Khaled, A.Y.; Aziz, S.A.; Rokhani, F.Z. Capacitive sensor probe to assess frying oil degradation. *Inf. Process. Agric.* 2015, 2, 142–148. [CrossRef]
- 23. ATAGO Company. Frying Oil Monitor DOM-24 | Products—Frying Oil Monitor | ATAGO Co., Ltd. Available online: https://www.atago.net/en/products-dom-top.php (accessed on 6 July 2022).
- 24. Wang, W.; Wang, Y.W.; Li, Z.X. Studies on Detection Device of Polar Components for Quality Evaluation of Frying Oil. *Adv. Mater. Res.* **2012**, 562–564, 1815–1818. [CrossRef]
- Yang, C.-C.; Li, Y.-T.; Chiang, D.; Chiu, P.-K.; Lin, Y.-C.; Hsiao, W.-T. Comparison of Sensing Methods for Characterization of Heated Oils Degradation. In Proceedings of the 2021 IEEE Sensors Applications Symposium (SAS), Sundsvall, Sweden, 23–25 August 2021.
- Chiang, D.; Ding, C.-F.; Li, Y.-T.; Lin, Y.-C.; Chiu, P.-K.; Tsai, H.-Y. (Eds.) Assessment of frying oil degradation using laser direct writing interdigital sensors. In Proceedings of the 2020 IEEE International Instrumentation and Measurement Technology Conference (I2MTC), Dubrovnik, Croatia, 25–28 May 2020.
- 27. Ricca, M.; Foderà, V.; Vetri, V.; Buscarino, G.; Montalbano, M.; Leone, M. Oxidation Processes in Sicilian Olive Oils Investigated by a Combination of Optical and EPR Spectroscopy. *J. Food Sci.* **2012**, *77*, C1084–C1089. [CrossRef] [PubMed]
- Vlachos, N.; Skopelitis, Y.; Psaroudaki, M.; Konstantinidou, V.; Chatzilazarou, A.; Tegou, E. Applications of Fourier transforminfrared spectroscopy to edible oils. *Anal. Chim. Acta* 2006, 573–574, 459–465. [CrossRef] [PubMed]

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