

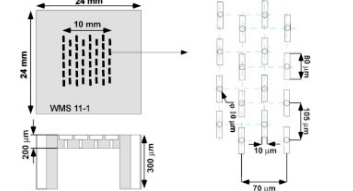
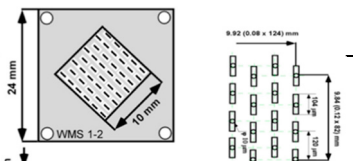
Table S1. Commercial microfluidic devices used to generate emulsion droplets.

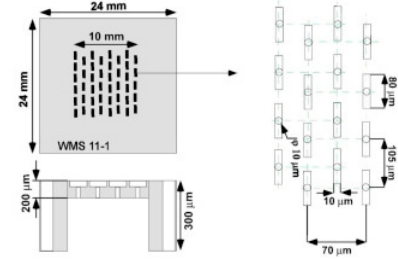
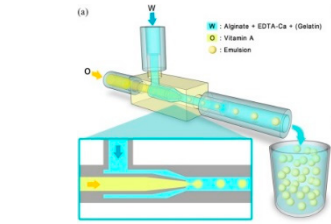
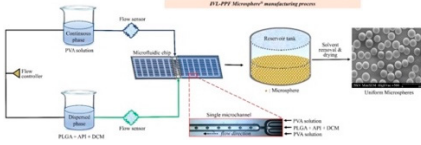
Company	Brand	Material	Characteristics	Applications	References
Dolomite	Telos ® 2 reagent Telos ® 1 reagent 3D flow focusing SC	Glass microfluidic device	-The device consists of 7 parallel junctions that can be combined with others 10. -Hydrophilic, hydrophobic, and fluorophilic coating options. -Available at 100 or 50 µm.	Designed for emulsion generation, foam generation, micro-particle synthesis and high throughput experimentation (for example, analysis of cells).	[1]
	Telos ® micromixer Chip		-The device consists of 7 independent micromixer channels on the chip (with 5 mixing stages each). -Hydrophilic and hydrophobic coating options. -Available at 30 or 50 µm.	Designed to create nano and micro particles and emulsions.	
	µEncapsulator 2 Reagent Droplet Chip		-Chip size 11.25mm x 15mm x 2mm. -Hydrophilic and fluorophilic coating surface options. -Junction of 100 µm scale.	Designed for rapid generation of double emulsion droplets (an aqueous core, surrounded by an oil shell)	
	µEncapsulator Sample Reservoir Chip		-Smooth channel with 100 µL storage volume. -Easy to clean and clear device. -Hydrophilic coating surface.	Designed for single emulsion droplet generation.	
	Large Droplet Junction Chip	Glass and quartz device	-Chip offers both a T- and X-junction. -Simple and easy to use. -High droplet production (12,000 per second). -Hydrophilic, hydrophilic and, fluorophilic coating options.	Designed for droplet emulsion generation.	
	Small Droplet Junction Chip		-Chip offers a flow focusing junction geometry with 14 x 17 µm cross-section at the junction.	Designed for generating small droplets in the size range of 5-30 µm,	

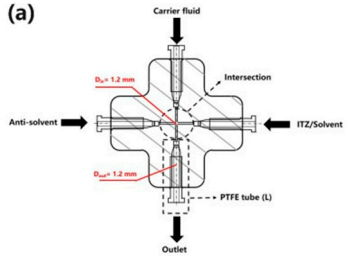
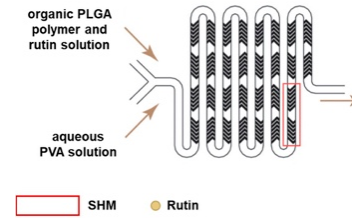
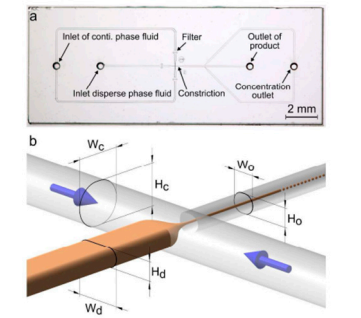
			-High droplet production (12,000 per second). -Hydrophilic, hydrophilic and, fluorophilic coating options.	improving control over the targeting and release of active compounds.	
	T-Junction Chip		-Extremely smooth channel surface. - Excellent chemical compatibility. -Hydrophilic, hydrophilic and, fluorophilic coating options.	Designed for a range of applications, including mixing fluids, microreactions and droplet formation.	
	6-Junction Droplet Chip	Glass device	-The device consists of 6 separate flow-focusing junctions. -Excellent chemical compatibility. -Hydrophilic, hydrophilic and, fluorophilic coating options.	Designed for the parallel generation of 20 µm - 60 µm oil-in-water droplets.	
Microfluidic ChipShop	Fluidic 162	Topas® COC (Cyclic olefin copolymer) or Polycarbonate PC	-One channel device. -Lid thickness 140 or 170 µm without surface treatment.	Designed for droplet generation on chip.	[2]
	Fluidic 537		-The flow-focusing device consists of 4 identical droplet generation units with a 38 µm nozzle size.		
	Fluidic 912		-The flow-focusing device consists of 8 identical droplet generation units with a channel dimension of 80 µm at the droplet formation region.		
	Fluidic 536		-The device consists of double-cross geometry with 37 µm nozzle size and 3 droplet generator units.	Designed for W/W/O double emulsion generation, allowing cell and particle encapsulation.	
	Fluidic 488		-The device consists of a combination of multiple T-junctions. -Storage module for capturing droplets. -Double cross nozzle type (74 µm).	Designed for generating single or multiple emulsions and storage.	

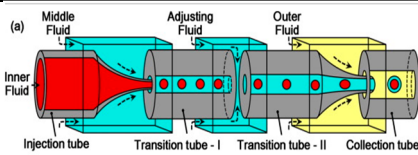
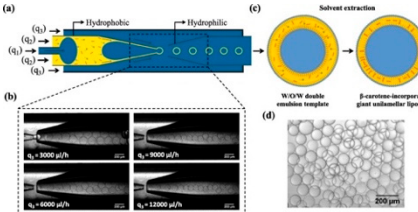
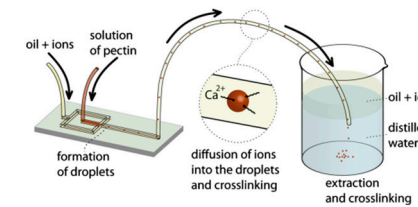
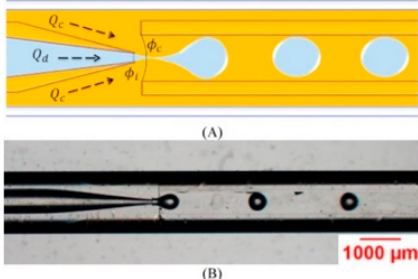
	Fluidic 285		-The multi-channel device presents various channels with different nozzle (50; 70; 80; 100 μm). -The main channel, as well as the entrance channel, vary in diameter, enabling a large set of experiments.	Designed for generating droplets with different volumes.	
	Fluidic 1196	Glass microfluidic device	-Several parallel microchannels in one unit. -Lid thickness of 250 μm -Channel dimensions 20 μm	Designed for droplet generation on chip.	

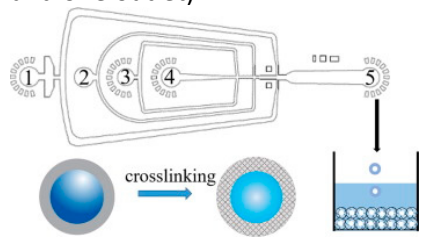
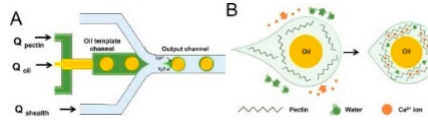
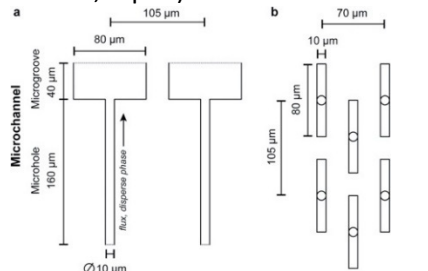
Table S2. Technological approaches and properties of the delivery systems based on food-grade emulsions assembled by microfluidic techniques.

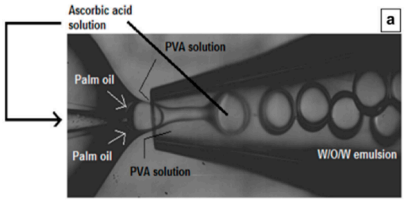
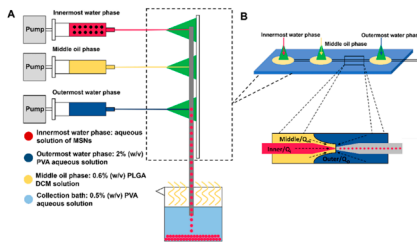
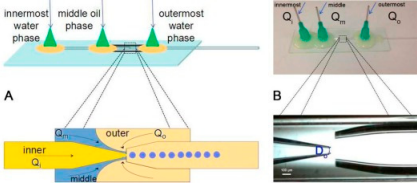
Emulsion type-Delivery system	Aqueous phase	Oil phase	Process conditions	Bioactive	Delivery system characterization	Microfluidic device type	Reference
O/W- Emulsion	Modified lecithin (ML), whey protein isolate (WPI), or Tween 20 (0.05-3 wt%)	Medium chain triacylglycerol oil	$Q_d = 0.1\text{-}10$ mL/h $Q_c = 100\text{-}500$ mL/h	Fucoxanthin extract (0-4 wt%)	Size= 30.3 ± 0.2 μm (ML), 32.3 ± 0.1 μm (WPI) and 29.0 ± 0.8 μm (Tween 20) DI= 0.5 EE= 100%(ML) 90.7% (WPI) 50.4% (Tween 20)	Silicon 24×24 mm ² microchannels array chip (Model: WMS 11-1; EP. Tech Co., Ltd., Hitachi, Japan) 	[3]
	Tween 20 (1% w/w), sodium salt of colic acid (Na-cholate), decaglycerol monolaurate, polyglyceryl-5-		$J_d = 10\text{-}300$ L/m.h	Quercetin (0.1-0.6 mg/mL)	Size= 28-29 μm DI < 0.21 EE= 80 % (4 °C) and 70 % (25 °C)	Silicon 24×24 mm ² microchannels array chip (Model WMS 1-2; EP. Tech Co., Ltd., Hitachi, Japan) 	[4]

	laurate (Sunsoft A-12E), or bovine serum albumin (BSA) (1% w/w)						
	Decaglycerol monolaurate (ML-750) or Tween 20 (1% w/w)						
	Sodium dodecyl sulfate (SDS), decaglycerol monolaurate (ML-750), decaglycerol monooleate (MO-7S), sodium casinate (Na-Cs), and modified lecithin (ML) (1% w/w)						
O/W-Microgel	Sodium alginate (1 wt%), gelatin (5 wt%), and EDTA-Ca (2 wt%)	Tert-butyl hydroquinone (TBHQ)	$Q_d = 1\text{--}14$ mL/h	β -sitosterol (0.5–4.0% w/w) and γ -oryzanol (0.5–4.0% w/w)	Size= 26–28 μm DI < 0.20 EE over 80 %	Silicon $24 \times 24 \text{ mm}^2$ microchannels array chip (Model: WMS 11–1; EP. Tech Co., Ltd., Hitachi, Japan)	[5]
			$Q_d = 0.25\text{--}5$ mL/h	AstaReal® (AR, astaxanthin purity 20%), Zanthin® (ZA, astaxanthin purity 10%), and Astaxanthin >97% (Sigma-Aldrich) (1–5% w/w)	Size= 35–37 μm DI < 0.25 EE over 98 %		[5]
	Poly (vinyl alcohol) (PVA) (0.25% w/v)	PLGA 7525A dissolved in dichloromethane (DCM)	-	Finasteride (28 mg, monthly doses)	Size around 30 μm DI= 23–28% EE= not specified CR= an initial burst, a moderate release, and then a plateau.	Photosensitive resin microfluidic device made using microscale 3D printing equipment	[6]
	Poly (vinyl alcohol) (PVA) (0.25% w/v)	PLGA 5050A (PURASORB® PDLG 5002A),	$P_d = 1,100$ mbar		Size= 40 μm (PLGA 7502A or 5002A) and		[7]
							[7]

		PLGA 7525A (PURASORB® PDLG 7502A), and PLA02A (PURASORB® PDL 02A) dissolved in dichloromethane (DCM)	$P_c = 2,200$ mbar		around $30\ \mu\text{m}$ (PLGA/PLA02A) DI= 0.28 and 0.16 EE > 96.5%		
O/W-Nanoparticle	D1: acetone or methanol D2: deionized water	3M fluorinated™ Fluid (FC-40)	$Q_c = 10$ to $250\ \mu\text{L}/\text{min}$. $Q_d =$ fixed at $50\ \mu\text{L}/\text{min}$.	Itraconazole (ITZ) nanoparticles (1 mg/mL)	Size= the volume was calculated and range from 1 to $6\ \mu\text{L}$. EE= encapsulation was confirmed by the increase of particle size.	316 stainless steel metal T-junction devices (a) 	[8]
	Poly (vinyl alcohol) (PVA; 1% w/v) solution	PLGA (13.4 mg/mL) in acetonitrile (ACN)	Total flow rate: 2-12 mL/min Flow rate ratio= 2:1 to 10:1, aqueous:oil phases	Rutin (10 mg/mL)	Size= $123.4\ \text{nm}$ DI= 0.16 ± 0.005 EE= $34 \pm 2\%$	NanoAssemblr® Benchtop Device 	[9]
	Water	Tween 20 dissolved in ethyl acetate	$P_c = 4.20$ bar $P_d = 3.81$ bar For the smallest particle size	Fenofibrate (0.5 %wt)	Size < $1\ \mu\text{m}$ DI= not specified	Glass microfluidic device 	[10]
O/W/O-	Middle phase: an aqueous solution	Outer phase: soybean oil,	$Q_i = 2.2\ \mu\text{L}/\text{h}$.	Thyme essential oil and lavender	Size= $182\text{--}342\ \mu\text{m}$ DI= 2.4-2.8%	Glass microfluidic device	[11]

Core-shell microcapsule	containing CaCl_2 ($100 \times 10^{-3} \text{ M}$), disodium-EDTA ($100 \times 10^{-3} \text{ M}$) and sodium alginate (2.0%; pH 7.0)	acetic acid (5% w/v) and PGPR (5.0% w/v) <i>Inner phase:</i> soybean oil, benzyl benzoate (1:1), and PGPR (2.0% w/v)	$Q_m = 5.0 \mu\text{L/h}$. $Q_o = 68.1 \mu\text{L/h}$	essential oil (33.33% v/v)	EE= not specified		
W/O/W- Giant liposome	<i>Outer phase:</i> poly (vinyl alcohol) (PVA; 10 % w/v) <i>Inner phase:</i> PVA (1% w/v) and dextran (9% w/v)	<i>Middle phase:</i> soybean lecithin (0.5% w/v) in the following organic solvent mixtures (1:1.8 v/v): chloroform/hexane; ethyl acetate/hexane or ethyl acetate/pentane.	$Q_i = 1000 \mu\text{L/h}$ $Q_m = 1000 \mu\text{L/h}$ $Q_o = 3,000\text{--}12,000 \mu\text{L/h}$	β -carotene (0.125% w/v)	Size= 100-180 μm DI= 3-6 % EE= not specified		[12]
W/O-Microgel	Pectin solution (0.5 and 1% w/w)	Acetic acid and CaCO_3 in rapeseed oil	$Q_d = 1 \text{ mL/h}$ $Q_c = 9 \text{ mL/h}$	Silver and gold nanoparticles (5.5 nm; 1.5 mg/mL)	The microgel was able to encapsulate nanoparticles		[13]
	Gellan gum (0.10–0.30% w/w) and <i>Jabuticaba</i> extract (diluted in 0.025 M potassium chloride pH 1 and 0.4 M sodium acetate pH 4.5)	PGPR (4% w/w) and calcium acetate (1% w/w) added to the soybean oil	$Q_d = 2\text{--}30 \text{ mL/h}$ $Q_c = 150\text{--}250 \text{ mL/h}$	<i>Jabuticaba</i> extract (10-30% v/v)	Size= 185-342 μm DI= 0.016 and 0.086		[14]

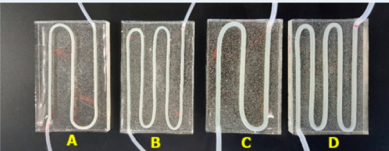
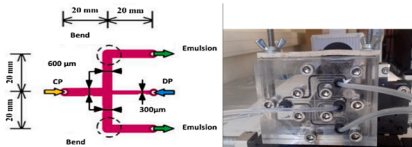
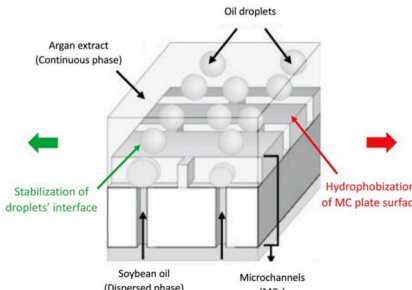
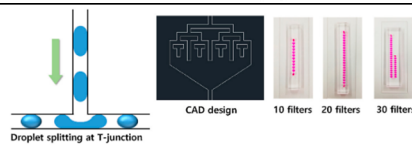
	<p>1: Dextran or protein aqueous solutions</p> <p>2: Alginate solution (2% wt%) containing CaCO_3 (200 mM). poly(ethyleneimine) (PEI; 0.3 wt%) or chitosan (1% v/v in acetic acid) coating</p>	<p>3: Mineral oil with Span 80 (3 wt%), used as oil phase A</p> <p>4: Mineral oil with Span 80 and acetic acid (0.2% v/v) used as oil phase B</p>	<p>$Q_1 = 15 \mu\text{L}/\text{min}$</p> <p>$Q_2 = 0.5 \mu\text{L}/\text{min}$</p> <p>$Q_3 = 10 \mu\text{L}/\text{min}$</p> <p>$Q_4 = 14 \mu\text{L}/\text{min}$</p>	Ovalbumin	<p>Size around $100 \mu\text{m}$</p> <p>DI= not specified</p> <p>EE= 88% (PEI coating) and 80% (chitosan coating)</p>	<p>Polydimethylsiloxane (PDMS) microfluidic device (four inlets and one outlet)</p> 	[15]
	<p>Alginate (2% w/v) and recombinant proteins in water</p> <p><i>Collection bath:</i> chitosan (0.5%) and CaCl_2 (0.1%)</p>	Span 80 in mineral oil	<p>$Q_d = 10 \mu\text{L}/\text{h}$</p> <p>$Q_c = 50 \mu\text{L}/\text{h}$</p>	eGFP+AvrA nanoparticles (2% w/v)	<p>Size= $339 \pm 52 \mu\text{m}$</p> <p>DI= 0.352 ± 0.153</p> <p>EE= not specified</p> <p>CR= 70 % of the encapsulated was released after 240 min</p>	<p>Polydimethylsiloxane (PDMS) microfluidic device</p>	[16]
	<p>Pectin (1 wt%)</p> <p>Sheath flow: CaCl_2 solution (1 wt%)</p>	<p>Mineral oil solution containing quercetin nanoparticles and retinyl palmitate, and Tween 80 (1 wt%)</p>	<p>$Q_d = 0.5 \mu\text{L}/\text{min}$</p> <p>$Q_c = 4.0 \mu\text{L}/\text{min}$</p> <p>$Q_{\text{sheath}} = 30 \mu\text{L}/\text{min}$</p>	<p>Quercetin nanoparticles (0.02 wt%) and retinyl palmitate (0.2 wt%)</p>	<p>Size around $67.3\text{--}93.1 \mu\text{m}$</p> <p>DI= not specified</p> <p>EE= not specified, encapsulation confirmed by fluorescence</p>	<p>Polydimethylsiloxane (PDMS) microfluidic device</p> 	[17]
W/O/W-Emulsion	<p><i>Inner phase:</i> phosphate buffer solution (100 mmol/L) containing betanin and D-glucose (1% w/w)</p> <p><i>Outer phase:</i> Tween 20 in water</p>	<p><i>Middle phase:</i> soybean oil and tetraglycerin monolaurate condensed ricinoleic acid ester (CR-310)</p>	<p>$J_d = 5\text{--}100 \text{ L}/\text{m.h}$</p>	E162, (mixture of beetroot extract and maltodextrin; 0.4% w/w betanin, TCI/ABCr), betanin, and spray dried beetroot juice) (0.1-1.0% w/w)	<p>Size < $50 \mu\text{m}$</p> <p>DI < 0.26 ± 0.01</p>	<p>Silicon $24 \times 24 \text{ mm}^2$ microchannels array chip (Model: WMS 11-1; EP. Tech Co., Ltd., Hitachi, Japan)</p> 	[18]

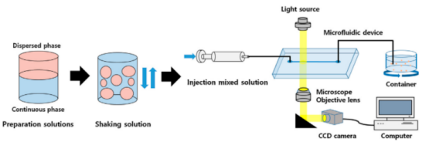
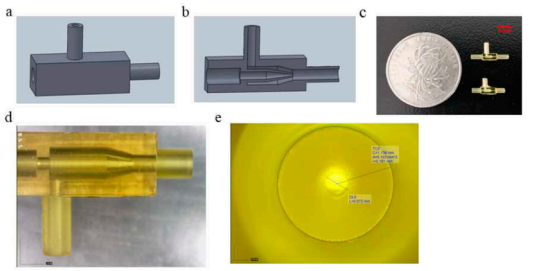
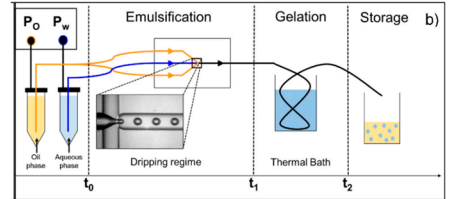
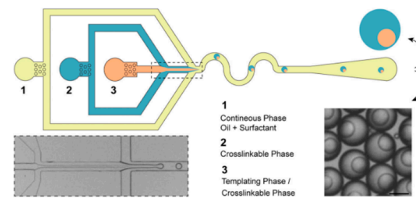
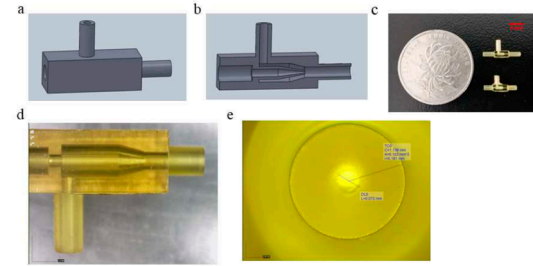
W/O/W- Solid lipid microparticle	<i>Outer phase:</i> water and polyvinyl alcohol (PVA; 10% (w/v) <i>Inner phase:</i> Ascorbic acid solution with or without CaCl ₂ , and chitosan.	<i>Middle phase:</i> palm fat oil	$Q_i = 3,000$ and $1,000 \mu\text{L/h}$. $Q_m = 2,500$ and $3,000 \mu\text{L/h}$. $Q_o = 14,000$ and $12,000 \mu\text{L/h}$	Ascorbic acid (3-20% w/w)	Size= 195- 342 μm DI= 73-95 % EE= 73-95 % CR decreased around 50% after 30 days	Glass microfluidic device 	[19]
W/O/W- Microcapsule	<i>Outer phase:</i> aqueous solution and PVA <i>Inner phase:</i> aqueous mesoporous silica nanoparticles (MSN) + Rhodamine B (RB) solution	<i>Middle phase:</i> PLGA oil	$Q_i = 1 \text{ mL/h}$. $Q_m = 2 \text{ mL/h}$. $Q_o = 4 \text{ mL/h}$	Mesoporous silica nanoparticles (MSN; 40% w/v) + Rhodamine B (RB; 10-500 mg/L)	Size= 119 nm (MSN) and 56 μm (PLGA-MSN) DI= 4.91% EE around 88% for PLGA-MSN and 95% for PLGA CR around 96% after 120 days for PLGA-MSN and 97% after 120 days for PLGA	Polymethyl methacrylate (PMMA) microfluidic device 	[20]
	<i>Outer phase:</i> Poly (vinyl alcohol) (PVA) aqueous solution (2 wt%) <i>Inner phase:</i> Poly (vinyl alcohol) (PVA) aqueous solution	<i>Middle phase:</i> PLGA (0.6 wt%) in dichloromethane)	$Q_i = 1000 \mu\text{L/min}$ $Q_m = 2000 \mu\text{L/min}$ $Q_o = 4000 \mu\text{L/h}$	2-[[[4-phenoxyphenyl]sulfonyl]methyl]-thiirane (SB-3CT; 0.5 mg/mL)	Size= 35-65 μm DI= 3% EE= 99%	Glass microfluidic device 	[21]
	<i>Middle phase:</i> poly (N-isopropyl acrylamide) (PNIPAM)	<i>Inner phase:</i> soybean oil solution added with camptothecin (0.4 mg/mL)	$Q_o = 1600-2400 \mu\text{L/min}$ $Q_m = 130-210 \mu\text{L/min}$ $Q_i = 20-100 \mu\text{L/min}$	Doxorubicin hydrochloride and camptothecin (2 mg/mL)	Size= 600-1000 μm DI= not specified EE= not specified CR= burst process	Glass microfluidic device	[22]

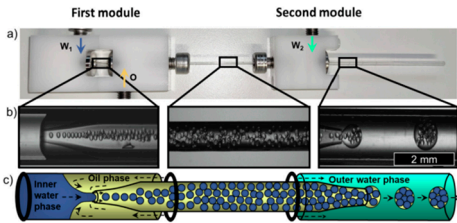
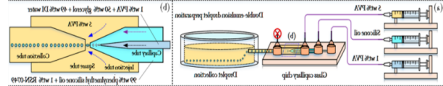
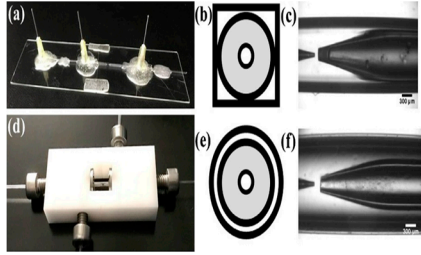
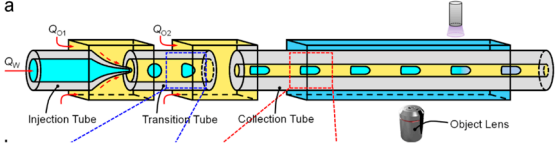
		Outer phase: soybean oil and PGPR					
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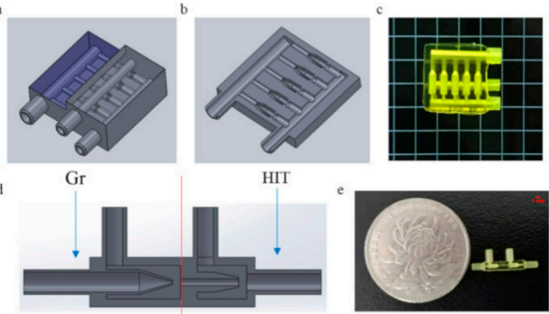
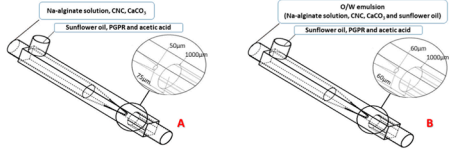
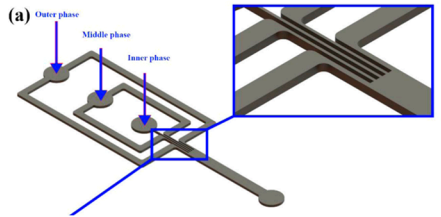
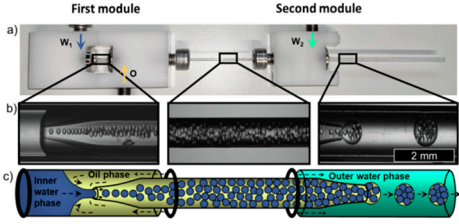
* Q_c : continuous phase flow rate; Q_d : disperse phase flow rate; Q_i : inner phase flow rate; Q_m : middle phase flow rate; Q_o : outer phase flow rate; J_d : disperse phase flux; P_d : disperse phase pressure; P_c : continuous phase pressure; Tween 20: polyoxyethylene (20) sorbitan monolaurate; EDTA-Ca: ethylenediaminetetraacetic acid calcium disodium salt hydrate; PGPR: polirricinoleato de poliglicerol; $CaCl_2$: calcium chloride; $CaCO_3$: calcium carbonate; PLGA: poly-lactic-co-glycolic acid; DI: dispersity index; EE: encapsulation efficiency; CR: compound release.

Table S3. Trends in droplet-based microfluidics approaches for fabricating delivery systems based on emulsions.

Emulsion type	Continuous phase	Disperse phase	Surfactant	Microfluidic device type	Process conditions	Microfluidic device design	Reference
O/W emulsion	Aqueous surfactant solution	Palm oil	Span 20	Combination of ultrasound and silicon/epoxy microchannel	Q_d and $Q_c = 4$ mL/min		[23]
	Xanthan gum or poloxamer in water	Sunflower oil	Tween 80 or Precirol™	Microsystem at High Throughput (MHT)	$Q_d = 20.8-95.2$ mL/min $Q_c = 500$ mL/min.		[24]
	Argan by-products extract	Soybean oil	Tween 80	Silicon	$Q_d = 2$ mL/h		[25]
	Mineral oil and glutaraldehyde (5 %wt)	Chitosan (2 %wt) in hydrochloric acid (1 %wt)	Span 80 (3 %wt)	PDMS microfluidic device	Q_d and $Q_c = 100$ to $500 \mu\text{L/h}$		[26]

							
	Silicon oil and surfactant	Deionized water	Span 80 (1% w/v)	Photosensitive resin Gr			[27]
	Water and PEG 2000	Sunflower oil	Tween 20	Glass microfluidic device	Q_d and $Q_c = 20$ to $280 \mu\text{L}/\text{min}$	-	[28]
W/O emulsion	Sunflower oil and surfactant	Whey protein isolated (WPI)	PGPR	Glass microfluidic device	Pressure ranged from 0-200 mbar for both phases		[29]
	Fluorocarbon oil (HFE 7500) and surfactant	PPGDA:Dextran (20% w/w) Dextran and PPGDA:PPGA (1:3 in 300 μL of ethanol)	Fluorosurfactant 008 (2% w/w)	PDMS microfluidic device	$Q_d = 60 \mu\text{L}/\text{h}$ for PPGDA and $30 \mu\text{L}/\text{h}$ for dextran		[30]
	Deionized water and surfactant	Mixture of mineral oil and heptane (50:50).	PVA (2% w/w).	Photosensitive resin Gr	Q_d and $Q_c = 100$ to $500 \mu\text{L}/\text{h}$		[27]

W/O/W emulsion	<i>Outer phase:</i> glycerol (5% w/v)	<i>Inner phase:</i> glycerol (50% w/v) in water <i>Middle phase:</i> middle chain triglyceride (MCT)	PGPR in MCT	Glass microfluidic device	$Q_i = 1-5 \text{ mL/h}$ $Q_m = 4-14 \text{ mL/h}$ $Q_o = 50-400 \text{ mL/h}$		[31]
	<i>Outer phase:</i> glycerol (25% w/w) in distilled water	<i>Inner phase:</i> Na-alginate (1.5 %wt) <i>Middle phase:</i> silicon oil	Span 80 (0.25% w/w) in middle phase. PVA (5% w/w) in outer phase	PDMS microfluidic device	$Q_i = 50 \mu\text{L/h}$. $Q_m = 500 \mu\text{L/h}$ $Q_o = 20000 - 100000 \mu\text{L/h}$		[32]
	<i>Outer phase:</i> aqueous surfactant solution	<i>Middle phase:</i> phenylmethyl silicone oil and silicone resin RSN-0749 <i>Inner phase:</i> glycerol, PVA, and water	PVA (5 wt%) in outer and inner phases.	Glass microfluidic device	$Q_i = 150 \text{ mL/h}$ $Q_m = 150-1750 \text{ mL/h}$ $Q_o = 1 \text{ mL/h}$		[33]
	<i>Outer phase:</i> glycerol (40 wt%) and surfactant	<i>Inner phase:</i> glycerol (5 %wt) and surfactant <i>Middle phase:</i> XIA-METER® RSN-0749 resin in Dow Corning® 200 fluid (2 wt%)	Hydrophilic: PVA Lipophilic: XIAMETER® RSN-074	Glass microfluidic device			[34]
O/W/O emulsion	<i>Outer phase 1 and 2:</i> oil solution containing PGPR (4% w/v)	<i>Inner phase:</i> poly(ethylene glycol) diacrylate (PEGDA; 50 %wt), 2-Hydroxy-methylpropio-phen one (HMPP; 5 %wt), and surfactant	Pluronic® F 127	Glass microfluidic device	$Q_i = 100-900 \mu\text{L/h}$ $Q_{o1} = 300 \text{ and } 200 \mu\text{L/h}$ $Q_{o2} = 500-800 \mu\text{L/h}$		[35]

	Outer phase: dimethicone	Inner phase: silicone oil. Middle phase: surfactant	PVA (2 % w/w) in the middle phase	Photosensitive resin Gr and HIT	$Q_i = Q_m = 2 \text{ mL/h}$ $Q_c = 5 \text{ mL/h}$		[27]
	Outer phase: sunflower oil and glacial acetic acid (0.1% w/w)	O/W emulsion prepared by ultrasound: Na- alginate (1% (w/w)) and CaCO_3 (0.1825 g/g of Na- alginate), cellulose nanocrystal and sunflower oil	PGPR (4 wt%)	Glass microfluidic device	$Q_c = 3000 - 15000 \mu\text{L/h}$. $Q_d = 600-3000 \mu\text{L/h}$		[36]
	Outer phase: silicone oil	Inner phase: silicone oil. Middle phase: glycerol (25% (w/w))	Span 80 (0.25% w/w) in the inner phase. PVA (10 % w/w) and in the middle phase. Span 80 (2.5% w/w) in the outer phase	PDMS microfluidic device	$Q_i = 50 \mu\text{L/h}$. $Q_m = 500 \mu\text{L/h}$ $Q_o = 20,000 - 100,000 \mu\text{L/h}$		[32]
	Outer phase: orange oil	Inner phase: middle chain triglyceride (MCT). Middle phase: PVA (1% w/v) in water	PVA in water and PGPR in MCT	Glass microfluidic device	$Q_o = 50-400 \text{ mL/h}$ $Q_m = 4-14 \text{ mL/h}$ $Q_i = 1-5 \text{ mL/h}$		[31]

* PVA: polyvinyl alcohol; PGPR: polyglycerol polyricinoleate; Span 20: sorbitan monooleate; PDMS: polydimethylsiloxane.

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