



Article

# Nutrient-Optimized Beef Enhances Blood Levels of Vitamin D and Selenium among Young Women

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Abstract: Bovine meat provides healthy nutrients but has also been negatively linked to greenhouse gases and non-communicable diseases. A double-blind intervention study was carried out to compare beef meat from bulls fed with feed supplemented with selenium, vitamin D, E, K (SeDEK-feed), and n-3, or REGULAR feed. Thirty-four young healthy women (19–29 years old) consumed 300 g of these beef types per day for 6 days in a cross-over design. Diet registrations, blood samples, anthropometric measurements, and clinical data were collected four times. Both beef diets were higher than their habitual diet in protein, fat, saturated fat, and several micronutrients; contained more vegetables and fewer carbohydrates and were followed by a higher feeling of satiety. The SeDEK beef had higher amounts of selenium, vitamin 25-hydroxyvitamin D3 (25(OH)D3), E, and K (MK4), and increased serum selenium and 25(OH)D3 from the participants' normal values if they were below 85  $\mu$ g/L of selenium and 30 nmol of 25(OH)D3/L, respectively. Our study showed that optimized beef increased serum selenium in young women having moderate selenium levels and improved blood 25(OH)D3 in a woman having low to normal 25(OH)D3. Meat should be optimized to increase specific consumer groups' needs for selenium and vitamin D.

Keywords: beef meat; human intervention; young women; vitamin D; selenium



Citation: Haug, A.; Vermeer, C.; Ruud, L.; Monfort-Pires, M.; Grabež, V.; Egelandsdal, B. Nutrient-Optimized Beef Enhances Blood Levels of Vitamin D and Selenium among Young Women. *Foods* **2022**, *11*, 631. https://doi.org/10.3390/foods11050631

Academic Editors: Carlotta Giromini and D. Ian Givens

Received: 12 January 2022 Accepted: 10 February 2022 Published: 22 February 2022

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# 1. Introduction

Bovine meat is a valuable food source, providing numerous nutrients and bioactive components that are vital for good health, such as proteins and peptides, B-vitamins, iron, and zinc [1]. Meat is palatable and gives a good satiety feeing and gastronomic pleasure. In the Scandinavian region, utilization of grassland in ruminant husbandry and the production of red meat occurs in areas unsuitable for the production of grain and vegetables. However, red meat has been given considerable negative attention due to concerns for greenhouse gasses and links between red meat and risk of non-communicable chronic diseases including cancer, cardiovascular diseases, diabetes, inflammation, and non-alcoholic fatty liver disease [2–7].

In the Western diet, low intakes of vegetables and fiber and a high intake of fat have been reported, while the contents of selenium, iodine, iron, vitamin D, K, and n-3 fatty acids are considered to be sub-optimal [8–10]. Adequate intake of long-chain n-3 fatty acids and a low ratio of n-6/n-3 fatty acids, for example, have documented positive effects on health parameters [9,11,12]. Micronutrients such as selenium and vitamin E are crucial for defense against oxidative damage and other important biological functions in the body [13,14].

Foods 2022, 11, 631 2 of 21

Vitamin K has an established cause-and-effect relationship with the maintenance of bone structure and blood coagulation [15,16].

In Europe, there is a gap between the current and recommended intake of vitamin D [17]. Insufficient vitamin D status is especially prevalent in children/adolescents in northern latitudes due to a lack of adequate sunlight in wintertime [18–20]. The vitamin D status in Norway has been evaluated [21]; the report claimed that a significant part of the population has suboptimal vitamin D levels (less than 50 nmol of 25-OH vitamin D/L serum). Vitamin D regulates many vital functions, including immune responses, inflammation, and proliferation associated with total mortality, fractures, falls, and CVD. For elderly persons, having a good vitamin D status is severely important [17,21–23].

Throughout the last decade, diets that avoid meat consumption have become more common among youth and young women [24–26], as well as vegans [27]. It has been pointed out that the omission of red meat may have negative effects on health, e.g., a lack of iron and vitamins [28,29]. Several of the nutrients in meat, such as selenium, iron, zinc, and some vitamins, have been shown to be more easily digested and absorbed compared to the same micronutrients from plants [30–32]. Selenium from meat has been shown to be retained more than twice as compared to plants' selenium (broccoli) [30]. Nevertheless, attempts to increase human serum concentration using selenium-fortified feed for production animals' meat have not been successful [33]. Iron as heme has approximately three times the bioactivity of inorganic iron [34]. The dominating form of vitamin D in beef meat is 25(OH)D3 [35], and this form has between 1.5 and 5 times higher bioactivity compared to vitamins D3 and D2 [36–38].

Supplementation of the bovine feed with extra nutrients can be an innovative strategy to increase the nutrient content in meat and thereby secure some nutrient and health claims related to compounds of importance to vulnerable groups [39]. Among the nutrients that have been discussed in relation to human health are vitamin D, E, K, *n*-3, and selenium. These five nutrients were deliberately attempted to be increased in the beef by providing a composite feed supplemented with *n*-3, selenium, vitamin D, E, and K to bulls [39], and this feed given to animals increased the concentration of these nutrients in the meat [39]. To our knowledge, there is limited research on how meat from animals that have been fed a diet supplemented with several different nutrients will affect human health markers. When producing bovine meat, all efforts should be made to obtain products with a nutrient composition that may benefit the consumers.

The aim of this double-blind cross-over (6-days  $\times$  2) intervention study in young female volunteers was as follows: (1) study the effects of ingesting 300 g of beef a day from bulls fed concentrate supplemented with extra vitamin D, E, K, n-3, and selenium (SeDEK-beef) compared to the regular composite feed (REGULAR); (2) compare the '300 g beef' diet with the habitual normal diet of the young healthy participants.

#### 2. Materials and Methods

#### 2.1. Subjects

A total of 34 free-living young healthy women, varying from 19 to 29 years, with normal weight (BMI 22.9  $\pm$  2.7; mean and standard deviation) participated. The trial was approved by the Norwegian Regional Committees for Medical and Health Research Ethics (REK 2016/620; ES498688; https://helseforskning.etikkom.no/ (accessed on 15 December 2021)) and registered in ISRCTN registry (ID ISRCTN25014465).

#### 2.2. Study Design

The study (32 days) design included a double-blind randomized cross-over intervention study (Figure 1) conducted at the Norwegian University of Life Sciences in Ås, Norway, at a latitude of  $60^{\circ}$  N during March when skin vitamin D synthesis is minimal.

Foods 2022, 11, 631 3 of 21

	Enrolment	Young female volunteers were assessed for eligibility (N = 40)
		Excluded for not meeting the inclusion criteria (N = 6) Randomization (N = 34)
Day		
1-3		Weighed dietary registration of food intake (N = 34) Habitual diet
4		Fasting blood and urine samples, BP and anthropometric measurements
5–10	1st intervention	Intake of 300 g SeDEK-beef per day (N = 17) Intake of 300 g REGULAR beef per day (N = 17)
8-10		Weighed dietary registration of food intake (N = 34)
11		Fasting blood and urine samples, BP and anthropometric measurements
11-24	Washout and crossover	Habitual diet
22-24		Weighed dietary registration of food intake (N = 34)
25		Fasting blood and urine samples, BP and anthropometric measurements
26-31	2nd intervention	Intake of 300 g REGULAR beef per day (N = 17) Intake of 300 g SeDEK-beef per day (N = 17)
29-31		Weighed dietary registration of food intake (N = 34)
32		Fasting blood and urine samples, BP and anthropometric measurements

**Figure 1.** Timeline for the experimental design of the cross-over double-blind study.

At baseline, energy content and nutrient composition of the normal, habitual diet were determined by weighted dietary registration (prospective registration) for three days using the "Kostholdsplanleggeren" [40].

The study contained two cross-over intervention periods each for 6 days (Figure 1) with an intake of 300 g raw minced beef meat from the forequarters, in combination with their chosen habitual food items, except fish. The raw minced beef meat corresponds to 240 g cooked beef, as about 20% weight is lost as meat juice during cooking.

The minced beef meat contained about 14% fat, contributing to about 42 g fat per day. The meat was from bulls fed regular control composite feed (REGULAR beef), and from bulls fed composite feed supplemented with vitamins D, E, K, *n*-3, and selenium (SeDEK-beef). Table 1 shows the values for myoglobin, fatty acids, selenium, vitamin D, E, and MK4 for both meats. SeDEK meat also had a lower *n*-6/*n*-3 ratio compared to the REGULAR beef (Table 1). SeDEK beef was a food item containing significant amounts of selenium, vitamin K (mainly as MK4) and vitamin D (mainly as 25(OH)D3) when classified according to the Council of the European Communities [41], as described earlier [39].

The participants received recipes on how to prepare healthy dishes with the meat, preferably adding vegetables. The participants were instructed to look for meat recipes at matprat.no (http://www.matprat.no/sok/#1/all/kj%C3%B8ttdeig/, accessed on 15 December 2021). They were free to use the beef in preparing hamburgers, tacos, or pizzas. The participants were asked to consume the meat in about three meals throughout the day.

The amount of beef given to participants was defined based on power calculation considering that it was a short intervention [42]. Because the amount of meat in the test diet was much higher than their habitual intake, the participants freely changed their diets to adapt to a new pattern with more meat, which included more regular meals and fewer snacks/sandwiches.

During the study, four weighed registrations of the complete food intake were performed, each of them for 3 days (Figure 1); before and during test periods. Then followed a two-week washout period. Samples of blood from the participants were collected in the morning (between 07:00 and 10:00) in fasting state, four times during the study, as well as four registrations of blood pressure, pulse, body weight, and height (only once) of the participants.

Foods 2022, 11, 631 4 of 21

**Table 1.** Measured nutrient contents in minced beef meat; control bulls (REGULAR) and bulls fed nutrient optimized composite feed (SeDEK) [39]. Reproduced with permission from Kaveri Thakuria, Meat Science, Elsevier, 2022.

I. 100 - 1	REGULAR	SeDEK	
In 100 g <sup>a</sup>	N = 6 Bulls	N = 6 Bulls	
Selenium, μg	10.0	12.6	
Myoglobin b, g	0.38	0.45	
Alpha-tocopherol, mg	0.16	0.65	
K <sub>1</sub> , μg	2.1	2.0	
MK4, μg	9.1	20.3	
Vit D3, μg	< 0.01	0.04	
25(OH)D3, μg	0.10	0.29	
Cholesterol, mg	73.0	57.5	
Fat, g	13.6	14.9	
C14:0, g	0.36	0.39	
C16:0, g	3.35	3.48	
C16:1 <i>n</i> -7, g	0.37	0.42	
C18:0, g	2.59	2.82	
C18:1 <i>n-</i> 9, g	5.01	5.56	
C18:1 trans, g	0.43	0.47	
C18:2 <i>n</i> -6, g	0.27	0.25	
C18:3 <i>n</i> -3, g	0.059	0.070	
C20:4 <i>n</i> -6, g	0.032	0.029	
C20:5 <i>n</i> -3, g	0.006	0.006	
C22:5 <i>n</i> -3, g	0.017	0.019	
n-6/n-3	3.6	2.9	

<sup>&</sup>lt;sup>a</sup> Analytical measurement errors are given in [35]. <sup>b</sup> hemin-based determination.

Forty young women were recruited to join the study but only 35 could start. The recruitment was conducted at the university by hand-outs and announcements via internet. One person was later removed due to extreme inflammatory markers (see method used below) that aligned with her survey comments on perceived sickness. The remaining 34 participants had an average age of 21.5 years. The 34 participants described themselves as healthy during the study, except a few who said they had a cold/flu for a few days. An a priori power calculation with a two-sided significance level of 5% concluded that 30 participants eating 300 g meat per day were required to observe significant differences in blood levels of selenium, vitamin K, and D. The recruitment was handled by personnel not directly involved in the study or in teaching of students. Exclusion criteria was use of medications, except birth control pills. The participants were asked not to take vitamin, mineral- or *n*-3 supplements for one month prior to the study and throughout the study period. The participants provided written informed consent before participating. They were informed that they were free to quit the study at any time.

Most participants joined studies in nutrition and food science and had earlier been trained in the method of three-day weighed dietary registration. The participants were informed by a physician if the results from the blood tests were out of the normal reference range.

#### 2.3. Anthropometric Measurements

Body weight was measured, without shoes and heavy clothing, to the nearest 0.1 kg using digital scales (Soehnle, Nassau, Germany). Height was measured to the nearest 0.1 cm using a portable stadiometer (Seca 217, Seca, Hamburg, Germany). Body mass index was calculated as weight (kg) divided by the square of height (m).

#### 2.4. Blood Analyses

Blood samples were obtained from the antecubital vein for whole blood, serum, and EDTA plasma. Medical personnel trained and authorized for blood sampling performed the

Foods 2022, 11, 631 5 of 21

sampling at a local medical center. The samples were marked by bar codes and processed within one hour of collection. Serum samples could clot for max 60 min, and plasma samples were put on ice. The tubes were centrifuged at  $1500 \times g$  for 15 min at 4 °C. Whole blood samples, plasma, and serum were stored at -20 °C, and the samples were analyzed at the end of the study, except hemoglobin and blood glucose, which were performed when the blood samples were taken. All blood samples were successfully collected except one blood sample taken from one participant that the phlebotomist failed to draw on one occasion.

# 2.4.1. Serum Lipids and Other Blood Markers

Total serum triacylglycerol, HDL-cholesterol, LDL-cholesterol, ALT, AST, insulin, C-peptide, and selenium were measured using accredited methods, at a commercial medical laboratory in Norway [43] who also provides the method codes and analytical coefficient of variation.

The inflammatory markers IL-6, IL-8, and IL-1 $\beta$  in EDTA plasma were analyzed using Quantikine ELISA kits Human IL-6, Human IL-8, and Human IL-1 $\beta$  (R&D Systems Europe, Bio-Techne, Abingdon OX14 3NB, UK).

Vitamins  $K_1$  (phylloquinone) and  $K_2$  (menaquinones, MK4, MK7, and MK8) in EDTA plasma were analyzed at VitaK laboratories (Maastricht, The Netherlands) by the following method: circulating levels of phylloquinone and menaquinones were measured with a standard HPLC technique using a C18 reversed-phase column and fluorometric detection after post-column electrochemical reduction [44]. Vitamin K1 (G L Synthesis, Worcester, MA, USA) was used as standard [44].

#### 2.4.2. Vitamin D3 and 25(OH)D3 Analyzed in Whole Blood

The analyses were performed at Fødevarestyrelsen, (8520 Lystrup, Denmark) using a 100  $\mu$ L whole blood where the protein was precipitated using 400  $\mu$ L 1% formic acid in cold acetonitrile. After mixing and centrifuging (12,000× g for 10 min), the supernatant was added to Oasis PRiME HLB 1cc (30 mg, manifold, Extraction Cartridges, Part no. 1860008055, Waters Corporation, Milford, MA 01757, USA) that was conditioned with 1 mL isopropanol/acetonitrile 60/40 plus 1 mL water. The vitamins were eluted with isopropanol/acetonitrile 2% NH3 80/20. Half a ml of the elute was evaporated under weak N2 flow at 25 °C for about 80 min, derivatized by 4-phenyl-1, 2, 4-triazoline-3.5-dione at room temperature for 1–2 h, dissolved in acetonitrile and separated by supercritical fluid chromatography (SFC-MS/MS) on ACQUITY UPC² Xevo TQ-S MS/MS from Waters Milford, MA 01757, USA. The contents of individual components were quantified by the following standards: Vitamin D3 (Cholecalciferol, 97%, ID DLM-8853-D) from Cambridge Isotope Laboratories, Inc. (Tewksbury, MA 01876, USA) and 25(OH)D3 (6,19,19-d3) (ID 705888-1MG) from Sigma-Aldrich A/S (2860 Søborg, Denmark).

#### 2.4.3. 25(OH)D3 in Whole Blood Compared to Serum, a Small Pilot Study

Due to lack of serum, vitamin D3 was analyzed in whole blood. A small pilot study was performed to estimate the conversion factor from blood concentrations to serum 25(OH)D3. Then, blood samples were drawn from five young women, and 25(OH)D3 was analyzed in whole blood and serum. This small pilot study showed a close association between the 25(OH)D3 in whole blood and serum (regression coefficient  $r=0.99,\,p<0.001$ ), and the concentration of 25(OH)D3 was lower in whole blood compared to serum. Data from the five women indicated that 25(OH)D3 in whole blood was 59.2% of the concentration in serum; the mean concentration of 25(OH)D3 in whole blood was 43.7 nmol/L and 73.8 nmol/L in serum. In the current study, the results for 25(OH)D3 were presented as 25(OH)D3 in serum (nmol/L serum), converted from blood values.

Foods 2022, 11, 631 6 of 21

#### 2.4.4. Determination of Blood Glucose and Hemoglobin

Determination of glucose and hemoglobin (Hb) was performed directly in blood droplets when the blood samples were taken. Blood glucose was determined by a blood glucose monitoring device, Accu-Chek, Roche, Germany. Hemoglobin was determined by HemoCue Hb 201<sup>+</sup>, (Brønshøj, Denmark) stationed at the medical center. If the results were outside the normal range, the participant was informed on location by the responsible clinical personnel or by the study responsible and advised to contact their personal physician.

# 2.5. Diet Registration

The nutrient intakes were calculated using data from the Norwegian Food Composition database (NFCD) [45] combined with an official diet tool (Kostholdsplanleggeren) [40]. The higher contents in the SeDEK beef of selenium, vitamin D3, and E were adjusted using own data. For the '300 g beef diet', the registration of vitamin D3 intake was calculated as the sum of vitamin D3 plus the biological amount of 25(OH)D3 in the beef; i.e., the concentration of analyzed 25(OH)D3 was multiplied by the factor 5. Information of 25(OH)D3 is not given in the NFCD, and no correction of biological activity was performed for other meat eaten. Information on vitamin K concentration in food is also not given in NFCD and estimates of vitamin K intake were performed by using the Danish Food Composition databases [38] and the corresponding US one [46].

# 2.6. Beef Production

Minced beef meat and fat tissue from quarter, left forepart, from one-year old Norwegian Red bulls reared at the Animal Production Experimental Farm (SHF) at NMBU were used in the study. From 6 months on, six of the bulls were given a regular composite feed, and the other six were given a composite feed supplemented with selenium and vitamin D, E, K, and *n*-3. The intake of feed concentrates, and roughages was registered individually for each animal. The bulls were slaughtered at a commercial slaughterhouse. Details of the study are published [39] and the complete diet is described regarding amounts and sources of ingredients except for the seeds that were from Vestfoldmøllene (Andebuveien 674, 3158 Andebu, Norway). The maximum content of total selenium authorized in feed was used for SeDEK [47].

Meat and fat tissue from each of the 12 animals were removed and minced to a homogenate containing about 14% fat as assessed by QVision (Tomra Systems ASA, Asker, Norway). The minced meat from all the six REGULAR and six SeDEK fed bulls, respectively, generated two batches. The minced meats were vacuum packaged, 300 g in each package, and marked with code 1 or 2 before frozen and stored in the dark at  $-20\,^{\circ}\mathrm{C}$  for 6 months. The chemical analyses of the meat were performed in the accredited laboratory of the Danish Veterinary and Food Administration (Fødevarestyrelsen, Århus, Denmark). Vitamin K of feed was determined at the National Institute of Nutrition and Seafood Research (NIFES), Bergen, Norway. Muscle myoglobin was assessed as hemin [39].

# 2.7. Nutrient Intake Calculations from Food Composition Databases and Our Own Analyses

The concentrations of selenium, alpha-tocopherol, vitamin  $K_1$ , MK4, vitamin D3, 25(OH)D3, fat, and fatty acids were determined in the forepart beef from REGULAR- and SeDEK-fed animals (Table 1).

The analyzed concentrations of selenium were numerically higher while vitamin D3 and E as alpha tocopherol equivalents ( $\alpha$ TE), n-3, and -6 fatty acids in the beef were numerically lower than in NFCD [45], respectively. 25(OH)D3 is not reported in NFCD.

#### 2.8. Estimation of Satiety in Habitual vs. Diets Holding 300 g Beef per Day

The participants were asked to describe their feeling of satiety (fullness) at four times during the study at days 4, 11, 25, and 32.

Foods 2022, 11, 631 7 of 21

#### 2.9. Statistical Analyses

Microsoft Office Excel was used for calculations of averages, standard deviations (SD), *t*-tests, and linear regressions. Minitab 18 (https://www.minitab.com/en-us/ (accessed on 15 December 2021) was used for calculations of one-way ANOVA (balanced or GLM) using Tukey's test as a post-hoc comparison test. Outlier detection was made with Grubbs' test (used on interleukin data).

Preprocessing of measured variables to fractional changes was also carried out as follows for measured variable i:  $(T_i - H_i)/H_i$  before statistical calculations. A test for values significantly different from zero change was performed using a one-sample Z-test based on the estimated standard deviation obtained for the population mean and standard deviation. A test for the validity of the cross-over design was used [48].

#### 3. Results

#### 3.1. Baseline Characteristics

The baseline anthropometric data and blood analyses of the 34 participants, standard deviations (SD), min and max values, and reference ranges are shown in Table 2. The baseline characteristics showed that the measured parameters, such as anthropometric registrations and the measured blood analyses at baseline, were within, or close to within, normal ranges (Table 2). One clear exception was their 25(OH)D3 levels.

One participant had hemoglobin (11.4 g/100 mL of blood) below the reference level (11.5 g/100 mL of blood). Four persons had serum C-peptide concentrations (205, 245, 245, and 258 pmol/L serum) below the lower reference level. In the present study, based on the average serum selenium concentrations, participants were close to being selenium deficient [8]. One participant had serum 25-OH vitamin D below 25 nmol/L serum, and 17 of the 34 participants were below 50 nmol/L serum, defined as vitamin D deficiency and the vitamin D lower limit for satisfactory vitamin D status in Norway, respectively [21]. Nine participants had phylloquinone below the reference level of 0.2  $\mu$ g/L plasma, one participant had MK7 higher than the reference level of 0.8  $\mu$ g/L, and two participants had MK8 higher than the reference level of 0.45  $\mu$ g/L plasma.

The baseline diet was as follows: Participant's habitual diet showed that the intake of products in natura or minimally processed (such as grains, fruits, and vegetables, but also animal products), corresponded to less than 40% of the daily energy intake. Moreover, the intake of fruits and vegetables corresponded to less than 30% of the energy intake from this food group (30% out of 40% of total energy intake), while rice, flour, and pasta represented more than 30%. The intake of non-processed meat was relatively low, representing less than 15% of the energy intake from minimally processed foods (data not shown). The baseline diet contained, on average, about 85 g per day of white-, red-, and processed meat, including liver-pate used as a bread spread, and only 13 g of beef meat. The diet contained in average 37 E% fat (from 27 to 46 E%), 13 E% saturated fat (from 9 to 18 E%), n-6/n-3 ratio was 5.5, 45 E% carbohydrates (from 29 to 62 E%), 18 E% protein (from 12 to 25 E%), 5 E% added sugar (from 1 to 16 E%), 27 g of fiber/day (from 10 to 58 g per day), and 6 g of salt/day (from 4 to 12 g per day).

Their baseline intakes of macro-nutrients were within Nordic recommendations [17] except for the following:

- Iodine; all the 34 persons had a lower intake than the recommended 150 μg per day, the average intake was 69 μg/day; only 46% of the recommended intake. The lower intake level is 70 μg per day [17], and 23 participants were below this level;
- Vitamin D; 29 of the 34. The lower intake level is 2.5 μg per day [17], and 9 of the participants were below this level;
- Iron; 28 of the 34 participants had a lower intake than the recommended 15 mg per day [17], and the average intake was 10.7 mg per day. The lower intake level was 9 mg per day, and 13 of the participants were below this level;

Foods 2022, 11, 631 8 of 21

• Selenium: 17 of the 34 participants had a lower intake than the recommended 50 μg per day, and the average intake was 52 μg per day. The lower intake level is 9 mg per day [17], and 13 of the participants were below this level.

**Table 2.** Baseline characteristics of 34 participants; anthropometric values and serum analyses, mean values at baseline, standard deviations (SD), min and max values and reference ranges.

N. O.	Baseline	Min	Max	Ref Values	
N = 34	$\overline{\text{Mean} \pm \text{SD}}$	IVIIII	Wiax		
Age, years	$21.4 \pm 2.0$	19	29		
Height, cm	$170 \pm 6$	157	180		
Body weight, kg	$66.1 \pm 8.5$	50	88		
BMI, kg/m <sup>2</sup>	$22.9 \pm 2.7$	18	29		
Systolic BP	$113 \pm 8$	94	132	90-130	
Diastolic BP	$70\pm7$	54	84	60-85	
Heart rate	$74\pm13$	48	102	60-100	
Blood glucose, mmol/L, blood	$5.0 \pm 0.5$	3.9	6.6	3.8-6.1	
Insulin, pmol/L serum	$72.6 \pm 31$	24	150	18-173	
C-peptide, pmol/L serum	$476\pm146$	205	850	270-1290	
Selenium μg/L serum	$84.9 \pm 15.1$	65.4	151.8	47-142	
AST, U/L serum	$18.3 \pm 4.5$	10	27	<35	
ALT, U/L serum	$14.7 \pm 5.7$	8	35	<45	
IL-6, ng/L plasma	$3.6 \pm 0.8$	2.2	6.8	<7 a	
IL-8, ng/L plasma	$21.5\pm1.2$	19.4	23.6	<16 <sup>b</sup>	
IL-1β, ng/L plasma	$12.3\pm2.6$	9.9	19.6	<12 <sup>c,d</sup>	
Hb, g/100 mL blood	$13.3 \pm 1.0$	11.4	15.6	11.5-16	
Triglycerides, mmol/L serum	$0.93 \pm 0.28$	0.5	1.6	<2.6	
LDL-cholesterol, mmol/L serum	$2.7\pm0.81$	0.9	4.8	1.2 - 4.3	
HDL-cholesterol, mmol/L serum	$1.6\pm0.41$	1.2	3.0	1.0-2.7	
Phylloquinone, μg/L plasma	$0.35 \pm 0.22$	nd	0.8	0.2-0.8 e,f,g,h	
MK4, μg/L plasma	n.d	n.d.	n.d.	<0.3 i	
MK7, μg/L plasma	$0.18 \pm 0.14$	nd	0.82	<0.8 j,k	
MK8, μg/L plasma	$0.13 \pm 0.18$	nd	0.48	<0.45 <sup>k</sup>	
25(OH)D3, nmol/L serum	$54.6^{1} \pm 19.6$	11.8	91.7	50-150	

 $<sup>^{</sup>a}$  [49];  $^{b}$  [50];  $^{cd}$  [51,52];  $^{e}$  [53];  $^{f}$  [54];  $^{g}$  [55];  $^{h}$  [56];  $^{i}$  [57];  $^{j}$  [58];  $^{k}$  [59];  $^{l}$  Only 6 samples out of 34 reached detection level for vitamin D3, and the vitamin D3 values are not presented.

No recommendations have been given for vitamin K in the Nordic Nutrition Recommendations [17]. EFSA has set the dietary reference value for vitamin K at 70  $\mu g/day$  for adults [60], but the U.S. Institute of Medicine has concluded that an adequate intake of vitamin K is 120 and 90  $\mu g/day$  for men and women, respectively [61].

# 3.2. Nutrient Intakes Were Calculated from Databases and Own Analyses

In Table 3, the nutrient intake was based on data reported in NFCD, while the calculated intakes of fat, *n*-3 and *n*-6, vitamin D, vitamin E, and selenium were corrected according to our own analyses of the SeDEK beef and REGULAR beef (Table 1). The vitamin D in Table 3 shows the sum of vitamin D as given in NFCD plus the amount of 25(OH)D3 as analyzed in the SeDEK beef and REGULAR beef, and then multiplied by 5, as suggested by [62], and as performed in Denmark [38]. Since NFCD does not contain 25(OH)D3, the intake of vitamin D shown for the Habitual diet in Table 3 may be underestimated.

Since vitamin K analyses are not given in NFCD, an estimation of the intake of vitamin K was made by us, based on the Danish and US food composition databases [38,46].

The intake of macro- and micronutrients at the baseline registration did not differ from the intake during the two-week washout period. Based on this, the average nutrient intake in these two registrations is presented in Table 3, left segment, named 'habitual diet'.

Foods 2022, 11, 631 9 of 21

**Table 3.** The calculated (databases and own values) nutrient intake of 34 participants, after the 3-day food registrations, mean values, and standard deviations (SD). The 'habitual diet' represents the mean nutrient intakes at baseline, and the washout period, the '300 g beef' represents the mean nutrient intakes when consuming SeDEK beef and REGULAR beef.

	Habitual Diet '300 g Beef'		42	SeDEK-Beef	REGULAR Beef	
N = 34	Mean ± SD	Mean $\pm$ SD	- <i>p</i> -	$\mathbf{Mean} \pm \mathbf{SD}$	$\mathbf{Mean} \pm \mathbf{SD}$	- <i>p</i>
Energy, kcal	$1848 \pm 336$	$1971 \pm 306$	0.117	$1894 \pm 313$	$2049 \pm 383$	0.071
Higher satiety, % of participants	3	77	< 0.001	-	-	-
Intake of beef, g	$12.7 \pm 2.3$	$312.2\pm8.4$	< 0.001	-	-	-
Intake of all meat, g	$84.8 \pm 17.8$	$391.0 \pm 28.8$	< 0.001	-	-	-
Intake of vegetables, g	$221\pm145$	$314 \pm 176$	0.020	-	-	-
Intake of cereals, g	$222 \pm 80$	$186 \pm 86$	0.080	-	-	-
Fat E%	$36.33 \pm 6.01$	$42.73 \pm 4.55$	< 0.001	$42.26 \pm 5.2$	$43.53 \pm 6.6$	0.385
Saturated fat E%	$12.53 \pm 3.10$	$17.60 \pm 2.69$	< 0.001	$17.65 \pm 3.3$	$17.56 \pm 3.7$	0.918
Trans unsaturated E%	$0.08 \pm 0.18$	$0.45 \pm 0.42$	< 0.001	$0.44 \pm 0.56$	$0.46 \pm 0.50$	0.909
Monounsaturated E%	$13.21 \pm 3.20$	$15.71 \pm 2.53$	< 0.001	$14.97 \pm 3.2$	$16.44 \pm 3.2$	0.062
Polyunsaturated fat E%	$6.40 \pm 1.89$	$4.56 \pm 1.53$	0.007	$4.24 \pm 1.5$	$4.88 \pm 1.7$	0.107
Carbohydrate (incl. fiber) E%	$45.68 \pm 7.11$	$35.54 \pm 5.13$	< 0.001	$36.15 \pm 5.8$	$34.94 \pm 6.7$	0.429
Added sugar E%	$5.65 \pm 4.98$	$3.97 \pm 3.34$	0.105	$4.03 \pm 4.4$	$3.88 \pm 3.7$	0.881
Protein E%	$18.12 \pm 3.67$	$21.37 \pm 2.30$	< 0.001	$21.53 \pm 3.0$	$21.21 \pm 2.3$	0.620
Fat, g	$73.7 \pm 21.1$	$91.5 \pm 15.8$	< 0.001	$90.0 \pm 15.9$	$95.0 \pm 21.1$	0.128
Saturated fat, g	$25.08 \pm 8.41$	$37.05 \pm 6.05$	< 0.001	$36.1 \pm 6.5$	$38.0 \pm 9.2$	0.333
Trans unsaturated fat, g	$0.51 \pm 0.33$	$1.07 \pm 0.25$	< 0.001	$1.01 \pm 0.3$	$1.14 \pm 0.4$	0.095
Monounsaturated fat, g	$26.85 \pm 8.73$	$34.62 \pm 6.27$	< 0.001	$31.91 \pm 6.0$	$35.92 \pm 9.2$	0.036
Polyunsaturated fat, g	$13.39 \pm 5.71$	$10.07 \pm 3.98$	0.007	$9.07 \pm 3.8$	$11.10 \pm 4.8$	0.055
n-3, g	$2.43 \pm 1.99$	$1.76 \pm 1.82$	0.151	$1.63 \pm 1.9$	$1.94 \pm 1.9$	0.599
n-6, g	$9.51 \pm 3.35$	$7.46 \pm 2.78$	0.008	$6.65 \pm 2.4$	$8.28 \pm 3.5$	0.032
n-6/n-3	$5.86 \pm 3.02$	$5.64 \pm 1.80$	0.722	$5.29 \pm 1.9$	$5.99 \pm 2.7$	0.220
Cholesterol, mg	$281 \pm 134$	$262 \pm 97$	0.514	$249 \pm 107$	$276\pm123$	0.341
Carbohydrate (incl. fiber) g	$219 \pm 47.8$	$185.3 \pm 46.9$	0.005	$178 \pm 51$	$193 \pm 60$	0.263
Starch, g	$108.5 \pm 29.7$	$95.3 \pm 33.4$	0.089	$91.6 \pm 35$	$99.0 \pm 45$	0.458
Mono- and disaccharides, g	$76.2 \pm 35.0$	$62.0 \pm 24.3$	0.057	$60.8 \pm 25$	$63.3 \pm 31$	0.714
Sugar, added, g	$26.6 \pm 25.1$	$18.1 \pm 15.3$	0.094	$16.63 \pm 16$	$19.49 \pm 17$	0.498
Fiber, g	$26.7 \pm 10.8$	$23.1 \pm 9.8$	0.149	$22.33 \pm 10$	$23.83 \pm 11$	0.560
Protein, g	$81.4 \pm 21.0$	$104.9 \pm 14.9$	< 0.001	$102 \pm 18$	$108 \pm 16$	0.214
Salt, g	$6.07 \pm 2.11$	$6.18 \pm 2.47$	0.843	$6.66 \pm 3.2$	$5.69 \pm 2.7$	0.175
Vitamin A, (RAE)	$1227 \pm 1327$	$634 \pm 349$	0.014	$593 \pm 436$	$675 \pm 491$	0.469
Retinol, µg	$648 \pm 1007$	$250 \pm 210$	0.028	$217 \pm 205$	$283 \pm 383$	0.379
Beta-carotene, µg	$4217 \pm 3562$	$3885 \pm 3589$	0.704	$3858 \pm 4350$	$3913 \pm 4252$	0.957
Vitamin D a, μg	$4.91 \pm 3.26$	$5.20 \pm 1.78$	0.650	$6.39 \pm 1.7$	$4.01 \pm 2.9$	< 0.001
Vitamin E, αTE	$13.16 \pm 4.75$	$11.49\pm3.70$	0.112	$11.38 \pm 3.3$	$11.61 \pm 5.0$	0.821
Vitamin K (total) b μg	$100.71 \pm 83.5$	$157.7 \pm 88.0$	0.008	$155.6 \pm 94$	$159.9 \pm 119$	0.869
MK4, μg	$10.57 \pm 7.05$	$54.15 \pm 5.63$	< 0.001	$69.29 \pm 2.8$	$39.00 \pm 10.2$	< 0.001
Thiamin, mg	$1.53 \pm 0.46$	$1.26 \pm 0.49$	0.022	$1.29 \pm 0.6$	$1.23 \pm 0.5$	0.648
Riboflavin, mg	$1.50 \pm 0.42$	$1.50 \pm 0.34$	0.923	$1.45 \pm 0.4$	$1.55 \pm 0.4$	0.288
Niacin, mg	$18.28 \pm 6.05$	$22.15 \pm 3.91$	0.003	$21.38 \pm 3.8$	$22.92 \pm 4.7$	0.141
Vitamin B6, mg	$1.80 \pm 0.57$	$1.72 \pm 0.49$	0.527	$1.64 \pm 0.5$	$1.79 \pm 0.6$	0.243
Folate, µg	$309 \pm 127$	$282 \pm 107$	0.336	$261 \pm 91$	$302 \pm 144$	0.173
Vitamin B12, μg	$5.16 \pm 2.22$	$7.10 \pm 1.31$	< 0.001	$6.96 \pm 1.5$	$7.24 \pm 1.7$	0.480
Vitamin C, mg	$104.4 \pm 67.1$	$119.5 \pm 70.8$	0.370	$120.5 \pm 95$	$118.6 \pm 72$	0.929
Calcium, mg	$835 \pm 328$	$737 \pm 254$	0.169	$726 \pm 357$	$748 \pm 327$	0.793
Iron, mg	$10.77 \pm 3.20$	$14.08 \pm 2.90$	< 0.001	$13.88 \pm 3.0$	$14.28 \pm 3.5$	0.616
Sodium, mg	$2405 \pm 871$	$2454\pm1016$	0.833	$2644 \pm 1297$	$2263 \pm 1074$	0.192
Potassium, mg	$3210 \pm 1071$	$3440 \pm 979$	0.358	$3382 \pm 913$	$3499 \pm 1242$	0.662
Magnesium, mg	$337 \pm 115$	$326 \pm 103$	0.693	$313 \pm 99$	$340 \pm 123$	0.316
Zinc, mg	$9.76 \pm 2.16$	$20.55 \pm 2.08$	< 0.001	$20.11 \pm 2.5$	$20.98 \pm 2.6$	0.162
Selenium, µg	$49.12 \pm 25.69$	$61.61 \pm 28.54$	0.023	$66.82 \pm 37.9$	$56.41 \pm 12.8$	0.134
Copper, mg	$1.31 \pm 0.53$	$1.30 \pm 0.42$	0.955	$1.24 \pm 0.4$	$1.36 \pm 0.5$	0.330
Phosphorous, mg	$1509 \pm 350$	$1573 \pm 307$	0.424	$1527 \pm 334$	$1620 \pm 389$	0.296
Iodine, μg	$76.38 \pm 34.81$	$55.72 \pm 27.45$	0.008	$51.33 \pm 29$	$60.10 \pm 37$	0.283

 $<sup>^{</sup>a}$  The vitamin D concentration in SeDEK and REGULAR beef is the sum of analyzed values for D3 + (25(OH)D3  $\times$  5). In the NFCD [45], only vitamin D is given.  $^{b}$  Vitamin K and MK4 are not given in the NFCD (Matvaretabellen.no) and the intake was estimated using values from 'the Danish Food Composition Database [38] and from the US Food Composition databases [46]. Total K is the sum of K1 + MK4.

# 3.3. Comparison of the '300 g Beef per Day' Diet to the Habitual Diet

# 3.3.1. Calculated Nutrient Intake of the Two Diets

The nutrient intake of the 300 g beef diet and the habitual diet is presented in Table 3. The beef diets were significantly different from the habitual diet (Table 3). The intake of beef and meat was significantly higher with the '300 g beef' diet; 312 g of beef and altogether 391 g of meat per day versus about 13 g of beef and altogether 85 g of meat per day when consuming the habitual diet. In addition, the intake of vegetables became significantly higher with the beef diets (p = 0.02), while the intake of grains was reduced

Foods 2022, 11, 631 10 of 21

and 66% of the participants reported that they were eating less bread when consuming the beef diet (p = 0.08). Higher satiety was reported when '300 g of beef' was consumed, 77% of the participants were significantly fuller (p < 0.001), compared to their habitual diet. However, no significant difference in energy intake between the diets was found (p = 0.12). The total content of saturated, monounsaturated, and trans fats, as well as protein intake, increased significantly (p < 0.001) during the beef intervention. The intake of polyunsaturated fat (p = 0.007), n-6 fatty acids (p = 0.008), and carbohydrates (p = 0.005) was significantly reduced when consuming the '300 g beef' diets compared to their habitual diet. The habitual intake of saturated fat was 12.5 E%, and this is higher than the recommended intake (<10 E%). With beef intervention, the average E% from fat and saturated fat were higher (42.7 E% and 17.6 E%) than recommended (40 E% and <10 E%), respectively [15]. The average intake of protein was higher (21.4 E%) than recommended with beef intervention (10–20 E%) [17]. Accordingly, the E% intake from carbohydrates (35.5 E%) was below the recommendation of 45–60 E% [15]. The intake of fiber was 26.7 g with the habitual diet and 23 g/day with the beef intervention; the latter below the recommended intake (≥25 g/day for women [17]).

The intakes of added sugar were 5.7~E% and 4~E% in habitual and beef intervention, respectively, which were within the recommendations (<10 E%) [15].

The calculated intake of micronutrients with the '300 g beef' diet compared to the habitual diet showed an increased intake of total vitamin K (p = 0.008), MK4 (p < 0.001), niacin (p = 0.003), vitamin B12 (p < 0.001), iron (p < 0.001), zinc (p < 0.001), and selenium (p = 0.023). The intake of vitamin A (p = 0.014), retinol (p = 0.028), thiamin (p = 0.022), and iodine (p = 0.008) were reduced during the '300 g beef' intervention compared to the habitual diet. The intake of vitamin D numerically increased (p = 0.65) with the beef diet compared to the habitual diet.

All of the participants had a habitual diet that did not meet one or more of the nutrients of the recommended intake [17] of iodine, vitamin D, K, iron, and selenium. Following the beef diets, a significant calculated increase in intake of iron, selenium, and vitamin K was shown (Table 3). The calculated intake of iodine worsened following the beef diet (Table 3); the average intake of iodine was  $56~\mu g$  during the beef intervention. Eating the beef diet had the following calculated consequences:

Iodine: The iodine intake decreased when consuming the beef diet. Beef meat is low in iodine ( $\sim$ 2 µg/100 g); i.e., gives minor contribution to the recommended intake of 150 µg/100 g.

Iron: 20 out of 34 participants were still below the recommended intake of 15 mg of iron per day, in contrast to 28 of the 34 participants at baseline.

Selenium: 5 of 34 participants had a lower calculated intake of selenium than the recommended 50 µg per day, in contrast to 17 of the 34 participants at baseline.

Vitamin K: Two participants were below the estimated intake of 70  $\mu$ g vitamin K per day during the beef intervention, in contrast to 13 participants at baseline/habitual diet.

3.3.2. Measured Changes in Physical and Blood Variables from Habitual Diets to a Diet including 300 g Beef per Day

Participant characteristics, anthropometric values, and serum analyses from the intervention periods are shown in Table 4, with the habitual diet shown in the left part of the table. The mean of two registrations with their SD following the intake of the beef and habitual diets are given with *p*-values for the comparison between the beef and habitual diets.

Body weight, BMI, blood pressure, and pulse were not affected by beef diets for 6 days, compared to the habitual status.

Blood glucose showed a tendency to increase following the intake of 300 g of beef per day (p = 0.078) compared to when consuming the habitual diet (Table 4, left part), but insulin showed no difference.

C-peptide and ALT were not affected by following beef diets compared to the habitual diet, while serum insulin and AST could not be properly interpreted since an effect of the

Foods 2022, 11, 631 11 of 21

presentation order of the two meats was found (Table 4). This could be interpreted as an additional factor that "accidently" co-varied with the test product, and the reason for this may, among other things, be due to intense exercise among some participants on a specific test diet [63].

Serum lipids (triglycerides, LDL cholesterol, and HDL cholesterol) were not significantly affected by the beef diet compared to the habitual diet (Table 4, left part).

The inflammatory variables in plasma were affected; IL-1 $\beta$ /IL-6 (the cardiac inflammation factor) was reduced, while IL-6 and IL-8 were increased when eating the beef diet compared to the habitual diet.

In addition, vitamin D, K, selenium, and hemoglobin were not significantly affected by daily intake of beef diets compared to the habitual diet (Table 4).

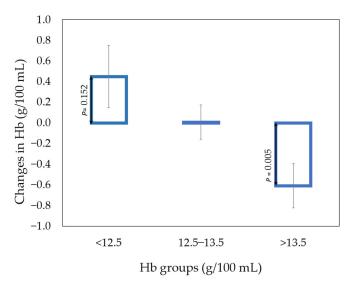
**Table 4.** Participant characteristics, anthropometric values, and serum analyses of 34 participants. Part I shows habitual and the '300 g beef' meat diets. Part II shows the 2 different meat diets in absolute values (upper part of cell) and as changes (lower part of cell). Part III tests for an effect of the 2 meat products' sequence.

	Part I				Part II			
	Habitual Diet	300 g Beef	3	SeDEK-Beef	REGULAR-Beef Mean $\pm$ SD	- <i>p</i> -	Cross Over	
Variable (N = 34)	Mean $\pm$ SD	Mean $\pm$ SD	— ра	Mean $\pm$ SD				
Body weight, kg	$66.35 \pm 8.58$	$66.64 \pm 8.64$	0.895	$66.45 \pm 8.71$	$66.47 \pm 8.83$	0.991	0.372	
body Weight, Rg	00.55 ± 0.56	00.04 ± 0.04	0.055	$0.001~^{\rm c}\pm0.011$	$0.002 \pm 0.010$	0.681	0.572	
BMI	$22.99 \pm 2.75$	$23.04 \pm 2.79$	0.944	$23.03 \pm 2.80$	$23.04 \pm 2.82$	0.993	0.839	
DIVII	22.77 ± 2.70	20.01 = 2.77	0.711	$0.001 \pm 0.010$	$0.002 \pm 0.010$	0.666	0.007	
Systolic BP	$113.3 \pm 7.16$	$115.8 \pm 7.17$	0.153	$116.4 \pm 9.02$	$115.4 \pm 7.32$	0.617	0.204	
,				$0.035 ** \pm 0.081$	$0.018 \pm 0.076$	0.372		
Diastolic PB	$70.4 \pm 5.46$	$71.3 \pm 5.88$	0.544	$71.3 \pm 7.49$ $0.011 \pm 0.120$	$71.4 \pm 5.95$ $0.025 * \pm 0.072$	0.957 0.543	0.175	
				$76.50 \pm 11.46$	$74.35 \pm 10.26$	0.543		
Pulse	$74.18 \pm 9.87$	$75.29 \pm 8.88$	0.625	$0.030 \pm 0.162$	$0.025 \pm 0.146$	0.419	0.378	
				$5.47 \pm 0.58$	$5.31 \pm 0.41$	0.334		
Blood glucose, mmol/L blood	$5.19 \pm 0.52$	$5.39 \pm 0.42$	0.078	$0.058 ** \pm 0.127$	$0.039 \pm 0.122$	0.538	0.125	
				$74.75 \pm 31.65$	$75.44 \pm 30.44$	0.928		
Insulin, pmol/L serum	$70.10 \pm 25.17$	$74.02 \pm 30.24$	0.564	$0.113 \pm 0.435$	$0.274 \pm 0.610$	0.221	0.038	
C (1) 1/I	100 1 444			$492 \pm 156$	$462 \pm 124$	0.386		
C-peptide, pmol/L serum	$459 \pm 111$	$471\pm138$	0.687	$0.078 \pm 0.290$	$0.085 \pm 0.318$	0.921	0.413	
ACT II /I	10.10   5.00	45.50   4.06		$17.62 \pm 5.10$	$18.03 \pm 5.79$	0.758		
AST, U/L serum	$19.13 \pm 5.23$	$17.59 \pm 4.86$	0.212	$0.047 \pm 0.212$	$0.052 \pm 0.207$	0.914	0.007	
ALT, U/L serum	15 41 1 6 64	1E EO   6 41	0.055	$15.53 \pm 6.45$	$15.91 \pm 8.16$	0.833	0.171	
ALI, U/L Serum	$15.41 \pm 6.64$	$15.50 \pm 6.41$	0.955	$0.023 \pm 0.191$	$0.040 \pm 0.215$	0.741	0.171	
AST/ALT	$1.31 \pm 0.26$	$1.22 \pm 0.26$	0.139	$1.22 \pm 0.34$	$1.21 \pm 0.25$	0.905	0.030	
A31/ALI	1.51 ± 0.20	1.22 ± 0.20	0.139	$-0.059 \pm 0.195$	$0.076 \pm 0.178$	0.702	0.030	
IL-1β, ng/L plasma	$11.3\pm2.7$	$10.5 \pm 3.0$	0.085	$10.7 \pm 3.6$	$10.3 \pm 2.2$	0.511	0.514	
12 1p/11g/ 2 p.moiim	11.5 ± 2.7	10.5 ± 5.0	0.003	$-0.065 \pm 0.245$	$0.089 *** \pm 0.141$	0.638	0.514	
IL-6, ng/L plasma	$3.6 \pm 0.8$	$4.1\pm1.3$	0.038	$4.2 \pm 1.4$	$4.0 \pm 1.2$	0.556	0.455	
, 8, 1	0.0 — 0.0		0.000	$0.119*\pm0.296$	$0.051 \pm 0.220$	0.294	0.100	
IL-8, ng/L plasma	$22.3 \pm 2.0$	$23.2 \pm 2.5$	0.037	$23.4 \pm 2.8$	$23.0 \pm 2.2$	0.519	0.252	
•				$0.049 *** \pm 0.074$	$0.027 * \pm 0.071$	0.221 0.500		
IL-1β/IL-6	$3.3 \pm 1.1$	$2.7 \pm 0.7$	0.001	$2.8 \pm 0.8 \\ -0.016 \pm 0.301$	$2.6 \pm 0.6 \\ 0.014 \pm 0.220$	0.500	0.379	
				$-0.016 \pm 0.301$ $13.13 \pm 1.14$	$13.32 \pm 0.90$	0.972		
Hb, g/100 mL blood	$13.17 \pm 0.90$	$13.23 \pm 0.92$	0.755	$-0.016 \pm 0.064$	$0.030 * \pm 0.078$	0.432	0.350	
				$0.89 \pm 0.26$	$0.90 \pm 0.34$	0.850		
Triacylglycerol, mmol/L serum	$0.89 \pm 0.27$	$0.89 \pm 0.29$	0.945	$0.033 \pm 0.262$	$0.025 \pm 0.220$	0.892	0.616	
				$2.72 \pm 0.85$	$2.81 \pm 0.85$	0.638		
LDL-cholesterol, mmol/L serum	$2.62 \pm 0.78$	$2.74 \pm 0.86$	0.556	$0.043 \pm 0.169$	$0.069 ** \pm 0.135$	0.492	0.021	
HDL-cholesterol, mmol/L serum	1 (2   0.27	1 (2   0 20	0.050	$1.65 \pm 0.34$	$1.67 \pm 0.41$	0.864	0.227	
ADL-cholesterol, mmol/ L serum	$1.62 \pm 0.37$	$1.63 \pm 0.38$	0.950	$0.015 \pm 0.098$	$0.043 ** \pm 0.072$	0.192	0.227	
Selenium, µg/L serum	$83.6 \pm 12.8$	$87.0 \pm 10.2$	0.093	$87.9 \pm 9.0$	$86.1 \pm 11.4$	0.472	0.410	
Scientini, µg/ L scium	05.0 ± 12.0	07.0 ± 10.2	0.053	$0.057 *** \pm 0.085$	$0.032*\pm0.079$	0.216	0.410	
Phylloquinone, µg/L plasma	$0.33 \pm 0.22$	$0.39 \pm 0.32$	0.410	$0.40 \pm 0.46$	$0.40 \pm 0.38$	0.989	0.302	
, Tamoric, pg, 2 pmonta	0.00 ± 0.22	0.00 ± 0.02	0.410	$0.38 \pm 1.15$	$0.36*\pm0.97$	0.938	0.002	
MK7, μg/L plasma	$0.19 \pm 0.12$	$9 \pm 0.12$ $0.17 \pm 0.11$ $0.436$	$0.436$ $0.16 \pm 0.11$	$0.18 \pm 0.17$	0.589	0.578		
	v — v <b>-</b>	**** — ****	0.200	$-0.125 \pm 0.402$	$-0.024 \pm 0.556$	0.456	3.370	
25(OH)D3, nmol/L serum	$52.0 \pm 18.1$	$49.2\pm16.8$	0.512	$50.4 \pm 17.2$	$48.4 \pm 16.6$	0.630	0.255	
. , ,	22.0 2 20.0 0.012		$-0.012 \pm 0.108$	$-0.064 ** \pm 0.131$	0.088 <sup>d</sup>			

<sup>&</sup>lt;sup>a</sup> The p represents t-tests; <sup>b</sup> The  $crossover\ p$  reveals probability of an effect of the presentation order [48]. <sup>c</sup> The fractional changes from habitual ( $H_i$ ) to each 300 g beef test diet intervention ( $T_i$ ) are calculated (( $T_i - H_i$ )/ $H_i$ ) and the significance (calculated by t-test) is indicated as: \* p < 0.05 \*\* p < 0.01 and \*\*\* p < 0.001); <sup>d</sup> This number was corrected (GLM analysis) for the effect of intervention week.

Foods 2022, 11, 631 12 of 21

Hb showed large variations (ranging from 11.4–15.5 g/100 g of blood at baseline), and the high blood Hb group revealed a significant decrease in Hb blood (Figure 2). The hemoglobin concentration for participants with <12.5 g Hb/100 mL blood (eight of 34 persons) numerically increased by consuming 300 g beef per day compared to eating the habitual diet (Figure 2). The average increase was 0.1 mg Hb/100 mL of blood per day.



**Figure 2.** The figure shows the changes in the blood Hb from habitual diet to the intake of 300 g of beef per day. The changes were allocated to three Hb groups Hb according to baseline values (<12.5 mg/100 mL, 12.5–13.5 mg/100 mL, and >13.5 mg/100 mL) where the number in the three groups were 8, 14, and 12, respectively. Mean values with standard errors are given and the p-values were obtained by one-sample Z-analysis indicating probability of obtained values being larger or lower than zero.

Regarding serum selenium, the average concentration at baseline (Table 2) was 84.9  $\mu$ g/L (1.07  $\mu$ mol/L), ranging from 65 to 152  $\mu$ g/L (0.8 to 1.9  $\mu$ mol/L), but only one person was below 70  $\mu$ g/L (0.85  $\mu$ mol/L) and only two persons were above 100  $\mu$ g/L (1.27  $\mu$ mol/L) of serum selenium at baseline.

# 3.4. Consuming SeDEK Beef Compared to REGULAR Beef, Calculated and Measured Data

Nutrient intake and study variables following the intake of 300 g of SeDEK beef compared to 300 g of REGULAR beef are shown in Tables 3 and 4, right parts of the tables, with mean, SD, and *p*-values. Tests based on fractional changes from baseline value, as well as the cross-over test are included in Table 4, right half of the table.

# 3.4.1. Calculated Nutrient Intake during the Intervention Consuming SeDEK Beef Compared to REGULAR Beef

Consumption of the SeDEK beef compared to REGULAR beef significantly reduced the intake of monounsaturated and n-6 fatty acids in grams (p = 0.036 and p = 0.032, respectively), although significant differences were not found when presented as E% (p = 0.062 and p = 0.173, respectively) (Table 3).

An increased intake of vitamin D and MK4 (p < 0.001) (Table 3), and selenium (p = 0.134) when consuming the SeDEK beef compared to REGULAR beef was found. As shown in Table 1, selenium concentration in the SeDEK and REGULAR beef were rather similar (38 µg and 30 µg/300 g beef, average selenium intake was 67 and 56 µg Se/day, respectively). The beef meat used in this intervention study was relatively rich in Se compared to most retail beef in Norway [35].

The daily intake of vitamin D was 60% higher when consuming SeDEK beef (6.4  $\mu$ g/day) compared with REGULAR beef (4.0  $\mu$ g/day), and it was a 75% higher intake of MK4 when consuming SeDEK beef (69  $\mu$ g/day) compared to REGULAR beef (39  $\mu$ g/day).

Foods 2022, 11, 631 13 of 21

3.4.2. Measured Study Variables following Consuming SeDEK Beef per Day Compared to REGULAR Beef

Body weight, BMI, Blood Pressure, and Pulse

Table 4 shows anthropometric registrations and study variables for participants consuming 300 g of SeDEK beef compared to 300 g of REGULAR beef per day. When tested for fractional changes from baseline values, significant increases in blood pressure were found (Table 4, right).

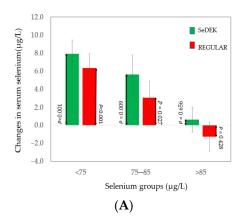
Serum lipids and Other Blood Markers

The concentration of blood lipids was not significantly different following the intake of SeDEK beef compared to REGULAR beef. When tested for fractional change, serum HDL cholesterol significantly increased for the REGULAR beef group (Table 4, right). Since serum LDL cholesterol was affected by the meat presentation order, the interpretation was less clear for LDL cholesterol.

A significant increase in blood glucose using SeDEK beef was observed, independent of the test order of the meats.

When the percentage of change from baseline was calculated (using a one-sample Z-test), the REGULAR beef intake increased hemoglobin concentration.

Serum selenium increased for both beef interventions when the percentage change from baseline was calculated, but the numerical value was highest when SeDEK beef was provided. Participants with serum selenium levels higher than  $\geq$ 1.15 µmol/L did not have an increase in serum selenium (Figure 3A. The uptake of selenium was larger the lower the initial blood selenium values were. In addition, serum selenium and calculated selenium intake correlated positively (r = 0.503, p < 0.01). Serum selenium correlated negatively to heart rate (r = -0.406, p < 0.01).



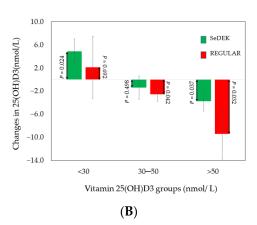


Figure 3. Mean values with standard errors are shown. The *p*-values were obtained by one-sample Z-test indicating probability of obtained values being larger or lower than zero. (**A**) Changes in the absolute serum values for three (<75  $\mu$ g/L, 75–85  $\mu$ g/L, and >85  $\mu$ g/L) different category groups of initial serum selenium. The average values in the three groups were 72 (N = 8), 80 (N = 13), and 98 (N = 13)  $\mu$ g/L, respectively. (**B**) Changes in the absolute values for three (<30 nmol/L, 30–50 nmol/L, and >50 nmol/L) different category groups of the initial 25(OH)D3 serum values. The average values in the three groups were 23 (N = 3), 40 (N = 15), and 73 (N = 16) nmol/L, respectively.

Due to the time of the year (March at  $60^{\circ}$  N, without sun exposure to increase serum vitamin D), vitamin 25(OH)D3 in the blood declined significantly (p = 0.012, GLM model) during the intervention period. The average concentration of 25(OH)D3 in the 34 participants on day 4, 11, 25, and 32 was the following: 55 nmol/L, 51 nmol/L, 49 nmol/L, and 48 nmol/L of serum, respectively. The reference range values for 25(OH)D3 in serum are 50–150 nmol/L [43].

Foods 2022, 11, 631 14 of 21

A difference in serum 25(OH)D3 between the two meats was found when the percent/fractional change from baseline was calculated, and the REGULAR beef intake would lead to a significant decline in 25(OH)D3 (Table 4, right and Figure 3B).

The absolute changes expected in 25(OH)D3 serum values depended on initial values (Figure 3B). For serum values above 50 nmol/L, the consumption of both types of meat induced a significant decline in 25(OH)D3. However, for serum values below 50 mmol/L, consumption of SeDEK beef showed no decline, while a significant decrease was detected for REGULAR beef when serum values were 30–50 nmol/L. The decrease of 25(OH)D3 may have occurred due to large variations within the group below 30 nmol/L.

There were no differences in total vitamin K, phylloquinone, MK7, and MK8 in plasma following intake of SeDEK beef compared to REGULAR beef. Plasma MK4, MK5, MK6, MK9, and MK10 were not detected in any of the plasma samples. The percent changes showed a 36–38 percent increase in plasma phylloquinone following both SeDEK and REGULAR meat intake, but the increase was only significant (p < 0.05) in the REGULAR beef group.

Vitamin E in beef meat, independent of feed fortification, seems too low to influence calculated intake (Table 3).

# • Inflammatory Markers

IL-6 and IL-8 showed small numerical increases with increased intake of meat. When fractional changes were used for analysis, these interleukins increased significantly for SeDEK-beef. Nevertheless, interleukin IL-1 $\beta$  only increased for REGULAR beef per day. We detected a trend (p < 0.1) for negative correlations between 25(OH)D3 and IL-6 and IL-8.

#### 4. Discussion

#### 4.1. The Nature of the Intervention (High Meat and Short Time)

The present cross-over intervention study reported improved vitamin D and selenium status of some groups of young women when consuming beef from bulls supplemented through feeding. High daily intake of beef (300 g of beef in raw weight per day) for 6 days resulted in increased vegetable consumption and did not adversely affect blood lipids, improved Hb levels for some groups, and reduced the cardiac inflammation ratio IL-1ß/IL-6, despite having increased the inflammatory markers IL-6 and IL-8. However, it is unknown whether other environmental factors, such as cold or flu during the winter season or low vitamin D status, could have affected the absolute values of the inflammatory markers. The weeks 3–11 in 2017 had an unusual high prevalence of the winter flu [64].

During the 6 days of 300 g of beef meat consumption, a drastic change in the diet occurred compared to the habitual diet. Apart from the high beef intake, they were free to choose their supplementary food intake and it is noteworthy that they significantly increased the intake of vegetables and reduced the intake of bread. When compared with their habitual diet, the intake of macronutrients during the beef interventions provided more fat and protein and less carbohydrate per day. By joining the study and eating 300 g of beef/day, the volunteers changed their diet towards some of the qualities of a Mediterranean diet, with a higher consumption of vegetables, less bread and refined grains, and moderate consumption of dairy products, which could have influenced some results, such as the lipid profile. The intake of micronutrients was also affected by eating high amounts of meat, more vegetables, and less bread, as shown by the increased intake of vitamin K, B12, niacin, iron, zinc, and selenium. This is in agreement with a study conducted in the US with data from NHANES in which the consumption of bread, as in sandwiches, was associated with fewer vegetable and fruit intakes [65].

Five nutrients (selenium, vitamin D, E, K, and *n*-3 fatty acids) were deliberately attempted to be increased to achieve healthier beef (SeDEK beef), compared to REGULAR beef, but only for selenium and vitamin D were significant blood changes observed. Supplemented feed gave minor differences in content of *n*-6 and *n*-3 fatty acids compared to REGULAR beef, due to bio-hydrogenation of PUFA in the rumen, but no significant effects on blood values were observed. Vitamin E was increased in the SeDEK beef, but the total

mean intake of vitamin E was many times higher than the contribution from the beef. The vitamin K3 that was added to the feed was bio-converted to MK4 in the bulls. An intake of 300 g of SeDEK beef could possibly have increased blood levels of MK4, but blood collected after 10 h fasting overnight showed no MK4 and no changes in other MK isomers.

# 4.1.1. Compliance

None of the 34 volunteers reported difficulties with compliance, although most of them were unfamiliar with such a high daily intake of red meat. The reported data showed high satiety, thus, no difficulties in consumption of the 300 g beef for 6 days  $\times$  2.

# 4.1.2. The Length of the Intervention Study

The 6-day intervention used in this study is shown to be long enough for determination of metabolic responses [42], but may be too short to register stable effects on some of the chosen study variables (such as vitamins, blood lipids, and even hemin/iron).

The present study (6 days) showed that a high intake of beef (three times higher than maximum recommendations) did not negatively affect the blood lipids and improved intakes of iron. On the other hand, beef intake slightly increased some inflammatory markers, and this should be further investigated

#### 4.2. Health Outcomes

# 4.2.1. Serum Lipids

LDL cholesterol is expected to increase with the extra intake of fat, especially saturated fat. However, consumption of 300 g of beef (with 40–45 g of fat, 20 g of saturated fat, and 12 g more saturated fat than the habitual diet) had no significant effect on the increase in the average concentration of serum lipids compared to the habitual diet. This could be due to a counterbalanced effect of increasing vegetable intake, although no increase in fiber intake was detected. Interestingly, only for the REGULAR beef group, the increase from baseline (%) following the diet showed a significant increase in LDL-cholesterol and HDL-cholesterol. However, the crossover p value for LDL cholesterol was 0.021, indicating an effect of the order of the diet. Thus, no strong conclusion can be drawn regarding LDL cholesterol. Although no adverse effects were detected in serum lipid after the beef diets, the short duration of the study (6 days  $\times$  2) and the alterations in several nutrients, such as higher intake of protein, selenium, vitamin B's, D, E, K, and n-3, and lower intake of carbohydrates altogether, may have had an effect on the results. Data from a recent meta-analysis indicates that an increased intake of some saturated fatty acids, such as lauric, myristic, or palmitic acid, can increase total, LDL, and HDL cholesterol levels and that the replacement of SFA for PUFA is able to reduce them [66].

# 4.2.2. Blood Glucose

The blood glucose concentration showed no significant changes from habitual to beef diets, although the intake of SeDEK beef led to a significant percentage (fractional) increase from the baseline. It is a topic of debate if a high protein intake increases blood glucose [67]. It might be suggested that the beef diets were more slowly degraded in the intestine compared to the carbohydrate-rich habitual diet, and a prolongation of the blood glucose levels might be an option.

#### 4.2.3. Micronutrients

# Blood Hb changes with increased meat intake

The increase in iron intake had no effect on average hemoglobin concentration following the intake of '300 g beef' compared to the habitual diet. Only participants having less than 12.5 g of hemoglobin/100 mL and consuming the '300 g beef' diet compared to the habitual diet tended to increase blood Hb after 6 days.

Heme-iron has approximately three times higher bioactivity than inorganic iron [34], and an improvement in heme-status could be expected following the beef diets, since the

Foods 2022, 11, 631 16 of 21

hemoglobin status of the volunteers was low-to-normal, and an improvement in hemestatus has been stated earlier [68]. This result may indicate individual differences between participants. Otherwise, it is possible that a 6-day intervention may not be long enough to obtain significant changes for the low blood Hb group [69]. It has also been shown that hemoglobin synthesis depends on selenium status [70].

# Iodine challenges with increased meat intake

The issue with the low iodine content in meat has recently been approached by feeding supplements of marine algae and seaweeds [71,72].

#### Selenium

The increase in serum selenium (%) from baseline levels was significantly higher after the intake of both beef diets, being numerically higher after SeDEK. The increase in serum selenium was high in subjects having serum selenium less than 85  $\mu$ g/L indicating that beef enriched with selenium (containing more than 10  $\mu$ g/100 g of beef) may increase serum values in those with lower serum selenium. This agrees with other studies [8].

The selenium concentrations in the two beef types, REGULAR and SeDEK beef (Table 1), were higher than anticipated (5  $\mu g$  Se/100 g according to the NFCD) [45]. The wheat used for both REGULAR and SeDEK-feed (imported) may have contained higher selenium content than the traditional local wheat. This information was not available during the feed processing, leading to a smaller difference between the REGULAR and SeDEK-feed than intended. A larger difference between the REGULAR and SeDEK feed might have led to larger differences in selenium, possibly giving larger differences in the present crossover intervention study, as has been shown in other selenium supplementation studies [73–75]. This shows the importance of feeding domestic animals with a feed with optimal nutritional quality regarding important nutrients such as selenium.

Selenium enriched meat could be a way of securing selenium status for groups having low selenium intake, i.e., individuals that do not consume fish. Ruminant feed enriched with selenium may provide meat products as a natural source of selenium kept at a healthy, beneficial, and harmless level.

#### Vitamin D

In general, beef diets contained 5–10 times more 25(OH)D3 than vitamin D3 (Table 1), and the SeDEK beef contained about three times more 25(OH)D3 than REGULAR beef. The importance of 25(OH)D3 in the diet has been assumed to be related to its higher bioactivity compared to vitamin D3.

This double-blind 6-day cross-over intervention study showed for the first time that 25(OH)D3 concentration in the blood improved in young women with a 25(OH)D3 concentration lower than 30 nmol/L when they consumed 300 g of SeDEK beef (containing 0.29  $\mu g$  of 25(OH)D3 g plus 0.04 ug D3/100 g). When consuming REGULAR beef (containing 0.10  $\mu g/100$  g), a significant decrease in blood 25(OH)D3 was found. This suggests that nutrient-optimized beef can enhance vitamin D status.

Intake of SeDEK beef did not improve blood 25(OH)D3 status if serum values were already >50 nmol/l at baseline. An obvious option is to aim towards the higher 25(OH)D3 values published [76]. Our target was set by the highest reported values in the locally produced meat [35,39]. Another possibility is that six days is too short a duration for observing larger differences, or that regulatory processes help to conserve the vitamin D level when the serum concentration is low.

#### • Vitamin K

Supplemented vitamin K3 added to the bull feed was bio-converted to MK4 in the bulls. Following the intake of 300 g of the SeDEK beef, compared to REGULAR beef, an increase in MK4 concentration in plasma was expected. However, MK4 was not detected at all in plasma in any of the participants. This may be due to its short plasma half-time [77]. Therefore, the effects of intake of MK4 on plasma could not be investigated in this study.

The phylloquinone (vitamin K1) concentration in SeDEK beef and REGULAR beef was nearly identical, and for the participants, the plasma levels of phylloquinone increased to about the same extent (38% and 36% increase) from baseline value following the intake of the two beef types, significantly only for the REGULAR group. The observed increase in plasma phylloquinone may be explained by the increase in consumption of vegetables when the participants were eating the beef diet, as plasma phylloquinone concentration has been shown to reflect the intake of vegetables [54].

With the '300 g beef' diet, total vitamin K intake significantly increased in 88% of participants and reached the recommended intake of 70 µg vitamin K/day.

# 4.2.4. Inflammation

The present study showed a percent increase from baseline in plasma cytokines with the intake of both beef diets. However, no difference between SeDEK beef and REGULAR beef was shown. The increase in IL-6 and IL-8 presents an important finding that indicates the potential impact of the high amount of beef on the inflammatory response. On the other hand, the reduced IL-1 $\beta$  /IL-6 ratio has been linked to reduced cardiac inflammation and to the expression of COX-2 [78].

#### 5. Conclusions

Selenium content above  $10~\mu g/100~g$  and 25(OH)D3 levels above  $0.29~\mu g/100~g$  in cattle meat will increase fasting serum values of young females with serum values below  $85~\mu g/L$  and 30~nmol/L, respectively. Nutrient-rich meat has market advantages for females that were deprived of these nutrients. The study points to the need for a higher awareness among meat producers regarding micronutrients. Even apparently small improvements in meat, as in SeDEK, may be of clinical significance for young women with a challenged nutrient status.

**Author Contributions:** Conceptualization: A.H. and B.E.; methodology, L.R.; validation: M.M.-P., A.H. and B.E.; formal analysis: L.R. and C.V.; investigation: V.G.; data curation: A.H., M.M.-P. and B.E.; writing—original draft preparation: A.H.; writing—review and editing: A.H., B.E., M.M.-P., C.V., V.G. and L.R.; visualization: A.H. and B.E., project administration: B.E.; funding acquisition: B.E. and A.H. All authors have read and agreed to the published version of the manuscript.

**Funding:** This study was funded by grants from the Norwegian Agriculture Agency Grant no 224794. Milena Monfort-Pires was funded by Grant no 281297 from the Norwegian Agriculture Agency.

**Institutional Review Board Statement:** The animal feeding study was conducted according to the guidelines/constraints previously described [39]. The feeding at The Department of Animal and Aquacultural Sciences and the commercial slaughtering was withing the general approval frame of the institutions. The human intervention was institutionally approved, formally by the Department Head when the external Ethical Committee (REK 2016/620; ES498688) had approved the intervention (9 January 2017).

**Informed Consent Statement:** Written informed consent has been obtained from the patient(s) to publish this paper.

**Data Availability Statement:** The data presented in this study made available https://archive.sigma2.no/pages/public/datasetDetail.jsf?id=10.11582/2022.00007, (accessed on 15 December 2021).

**Acknowledgments:** The authors are grateful to the technical staff at NMBU, especially Frank Sundby, Follo Bedriftshelsetjeneste (responsible medical doctor Hans Petter Storm Thorstensen, Ås, Norway), and Fürst Medisinske Senter (Oslo, Norway). Bolette Oxholm, Tor Lea, and Lars Fredrik Moen are thanked for analytical assistance, and Kristin Saarem for careful reading of the manuscript. Finally, Berit Johansen and Ingerid Arbo are thanked for suggestions regarding how to carry out this experiment.

Conflicts of Interest: The authors declare no conflict of interests.

#### References

1. Biesalski, H.K. Meat as a component of a healthy diet-are there any risks or benefits if meat is avoided in the diet? *Meat Sci.* **2005**, 70, 509–524. [CrossRef] [PubMed]

- 2. Bellavia, A.; Stilling, F.; Wolk, A. High red meat intake and all-cause cardiovascular and cancer mortality: Is the risk modified by fruit and vegetable intake? *Am. J. Clin. Nutr.* **2016**, *104*, 1137–1143. [CrossRef] [PubMed]
- 3. Wurtz, A.M.; Hansen, M.D.; Tjonneland, A.; Rimm, E.B.; Schmidt, E.B.; Overvad, K.; Jakobsen, M.U. Substitution of meat and fish with vegetables or potatoes and risk of myocardial infarction. *Br. J. Nutr.* **2016**, *116*, 1602–1610. [CrossRef] [PubMed]
- 4. InterAct, C. Adherence to predefined dietary patterns and incident type 2 diabetes in European populations: EPIC-InterAct Study. *Diabetologia* **2014**, *57*, 321–333.
- 5. Pan, A.; Sun, Q.; Bernstein, A.M.; Manson, J.E.; Willett, W.C.; Hu, F.B. Changes in red meat consumption and subsequent risk of type 2 diabetes mellitus: Three cohorts of US men and women. *JAMA Intern. Med.* **2013**, *173*, 1328–1335. [CrossRef]
- 6. Zelber-Sagi, S.; Ivancovsky-Wajcman, D.; Fliss Isakov, N.; Webb, M.; Orenstein, D.; Shibolet, O.; Kariv, R. High red and processed meat consumption is associated with non-alcoholic fatty liver disease and insulin resistance. *J. Hepatol.* **2018**, *68*, 1239–1246. [CrossRef]
- 7. Al-Shaar, L.; Satija, A.; Wang, D.D.; Rimm, E.B.; Smith-Warner, S.A.; Stampfer, M.J.; Hu, F.B.; Willett, W.C. Red meat intake and risk of coronary heart disease among US men: Prospective cohort study. *BMJ* **2020**, *371*, m4141. [CrossRef]
- 8. Rayman, M.P.; Winther, K.H.; Pastor-Barriuso, R.; Cold, F.; Thvilum, M.; Stranges, S.; Guallar, E.; Cold, S. Effect of long-term selenium supplementation on mortality: Results from a multiple-dose, randomised controlled trial. *Free Radic Biol. Med.* **2018**, 127, 46–54. [CrossRef]
- 9. Simopoulos, A.P. The importance of the ratio of omega-6/omega-3 essential fatty acids. *Biomed. Pharmacother.* **2002**, *56*, 365–379. [CrossRef]
- 10. Cashman, K.D.; Dowling, K.G.; Skrabakova, Z.; Gonzalez-Gross, M.; Valtuena, J.; De Henauw, S.; Moreno, L.; Damsgaard, C.T.; Michaelsen, K.F.; Mølgaard, C.; et al. Vitamin D deficiency in Europe: Pandemic? *Am. J. Clin. Nutr.* **2016**, *103*, 1033–1044. [CrossRef]
- 11. Haug, A.; Nyquist, N.F.; Mosti, T.J.; Andersen, M.; Hostmark, A.T. Increased EPA levels in serum phospholipids of humans after four weeks daily ingestion of one portion chicken fed linseed and rapeseed oil. *Lipids Health Dis.* **2012**, *11*, 104. [CrossRef]
- 12. Christophersen, O.A.; Haug, A. Animal products, diseases and drugs: A plea for better integration between agricultural sciences, human nutrition and human pharmacology. *Lipids Health Dis.* **2011**, *10*, 16. [CrossRef] [PubMed]
- 13. Comitato, R.; Ambra, R.; Virgili, F. Tocotrienols: A Family of Molecules with Specific Biological Activities. *Antioxidants* **2017**, *6*, 93. [CrossRef] [PubMed]
- 14. Hosnedlova, B.; Kepinska, M.; Skalickova, S.; Fernandez, C.; Ruttkay-Nedecky, B.; Malevu, T.D.; Sochor, J.; Baron, M.; Melcova, M.; Zidkova, J.; et al. A Summary of New Findings on the Biological Effects of Selenium in Selected Animal Species-A Critical Review. *Int. J. Mol. Sci.* 2017, *18*, 2209. [CrossRef] [PubMed]
- 15. EFSA. Scientific Opinion on the substantiation of health claims related to selenium and protection of DNA, proteins and lipids from oxidative damage (ID 277, 283, 286, 1289, 1290, 1291, 1293, 1751), function of the immune system (ID 278), thyroid function (ID 2). EFSA J. 2009, 7, 1220. [CrossRef]
- EFSA. Scientific Opinion on the substantiation of health claims related to vitamin K and maintenance of bone (ID 123, 127, 128, and 2879), blood coagulation (ID 124 and 126), and function of the heart and blood vessels (ID 124, 125 and 2880) pursuant to Article. EFSA J. 2009, 7, 2714.
- 17. Nordic Council of Ministers. *Nordic Nutrition Recommendations* 2012: *Integrating Nutrition and Physical Activity*, 5th ed.; Nordic Council of Ministers: Copenhagen, Denmark, 2014. [CrossRef]
- 18. Mortensen, C.; Damsgaard, C.T.; Hauger, H.; Ritz, C.; Lanham-New, S.A.; Smith, T.J.; Hennessy, A.; Dowling, K.; Cashman, K.D.; Kiely, M.; et al. Estimation of the dietary requirement for vitamin D in white children aged 4–8 y: A randomized, controlled, dose-response trial. *Am. J. Clin. Nutr.* **2016**, *104*, 1310–1317. [CrossRef]
- 19. Aspell, N.; Lawlor, B.; O'Sullivan, M. Is there a role for vitamin D in supporting cognitive function as we age? *Proc. Nutr. Soc.* **2018**, 77, 124–134. [CrossRef]
- Laird, E.; O'Halloran, A.M.; Carey, D.; Healy, M.; O'Connor, D.; Moore, P.; Shannon, T.; Molloy, A.M.; Kenny, R.A. The Prevalence of Vitamin D Deficiency and the Determinants of 25(OH)D Concentration in Older Irish Adults: Data From The Irish Longitudinal Study on Ageing (TILDA). J. Gerontol. A Biol. Sci. Med. Sci. 2018, 73, 519–525. [CrossRef]
- 21. Nasjonalt Råd for Ernæring. Vitamin D i Norge: Behov for Tiltak for å Sikre Vitamin D Status? Report Series 11/18. 2018, pp. 1–44. Available online: https://www.helsedirektoratet.no/rapporter/vitamin-d-i-norge-behov-for-tiltak-for-a-sikre-god-vitamin-d-status (accessed on 15 December 2021).
- 22. Berridge, M.J. Vitamin D deficiency accelerates ageing and age-related diseases: A novel hypothesis. *J. Physiol.* **2017**, *595*, 6825–6836. [CrossRef]
- 23. Wei, R.; Christakos, S. Mechanisms Underlying the Regulation of Innate and Adaptive Immunity by Vitamin, D. *Nutrients* **2015**, 7, 8251–8260. [CrossRef] [PubMed]
- 24. Fayet, F.; Flood, V.; Petocz, P.; Samman, S. Avoidance of meat and poultry decreases intakes of omega-3 fatty acids, vitamin B12, selenium and zinc in young women. *J. Hum. Nutr. Diet.* **2014**, 27 (Suppl. 2), 135–142. [CrossRef] [PubMed]

25. Rimstad, S.H. Interesse for et Sunnere Storfekjøtt Blant Forbrukere i Norge. Master's Thesis, Norwegian University of Life Sciences, Ås, Norway, 15 May 2016. Available online: https://brage.bibsys.no/xmlui/handle/11250/92927/browse?value=Rimstad%2C+Silje+Helene&type=author (accessed on 15 April 2021).

- 26. Segovia-Siapco, G.; Burkholder-Cooley, N.; Haddad Tabrizi, S.; Sabate, J. Beyond Meat: A Comparison of the Dietary Intakes of Vegetarian and Non-vegetarian Adolescents. *Front. Nutr.* **2019**, *6*, 86. [CrossRef] [PubMed]
- 27. Fallon, N.; Dillon, S.A. Low Intakes of Iodine and Selenium Amongst Vegan and Vegetarian Women Highlight a Potential Nutritional Vulnerability. *Front. Nutr.* **2020**, *7*, 72. [CrossRef]
- 28. Reddy, M.B.; Hurrell, R.F.; Cook, J.D. Meat consumption in a varied diet marginally influences nonheme iron absorption in normal individuals. *J. Nutr.* **2006**, *136*, 576–581. [CrossRef]
- 29. Samuelson, G.; Lonnerdal, B.; Kempe, B.; Elverby, J.E.; Bratteby, L.E. A follow-up study of serum ferritin and transferrin receptor concentrations in Swedish adolescents at age 17 age 15. *Acta Paediatr.* **2000**, *89*, 1162–1168. [CrossRef]
- 30. Finley, J.W.; Grusak, M.A.; Keck, A.-S.; Gregoire, B.R. Bioavailability of selenium from meat and broccoli as determined by retention and distribution of 75Se. *Biol. Trace Elem. Res.* **2004**, *99*, 191–209. [CrossRef]
- 31. Tetens, I.; Bendtsen, K.M.; Henriksen, M.; Ersboll, A.K.; Milman, N. The impact of a meat- versus a vegetable-based diet on iron status in women of childbearing age with small iron stores. *Eur. J. Nutr.* **2007**, *46*, 439–445. [CrossRef]
- 32. Krebs, N.F.; Westcott, J.E.; Culbertson, D.L.; Sian, L.; Miller, L.V.; Hambidge, K.M. Comparison of complementary feeding strategies to meet zinc requirements of older breastfed infants. *Am. J. Clin. Nutr.* **2012**, *96*, 30–35. [CrossRef]
- 33. Mrazova, J.; Gazarova, M.; Kopcekova, J.; Kolesarova, A.; Bucko, O.; Bobcek, B. The effect of consumption of pork enriched by organic selenium on selenium status and lipid profile in blood serum of consumers. *J. Environ. Sci. Health Part B* **2020**, *55*, 69–74. [CrossRef]
- 34. Bjorn-Rasmussen, E.; Hallberg, L.; Isaksson, B.; Arvidsson, B. Food iron absorption in man. Applications of the two-pool extrinsic tag method to measure heme and nonheme iron absorption from the whole diet. *J. Clin. Investg.* **1974**, *53*, 247–255. [CrossRef] [PubMed]
- Egelandsdal, B.; Oostindjer, M.; Hovland, E.M.; Okholm, B.; Saarem, K.; Bjerke, F.; Ruud, L.; Grabež, V.; Haug, A. Identifying labelling and marketing advantages of nutrients in minced beef meat: A case study. *Meat Sci.* 2020, 159, 107920. [CrossRef] [PubMed]
- 36. Cashman, K.D. The role of vitamers and dietary-based metabolites of vitamin D in prevention of vitamin D deficiency. *Food Nutr. Res.* **2012**, *56*, 5383. [CrossRef] [PubMed]
- 37. Jakobsen, J. Bioavailability and bioactivity of vitamin D3 active compounds–Which potency should be used for 25-hydroxyvitamin D3? *Int. Congr. Ser.* **2007**, 1297, 133–142. [CrossRef]
- 38. National Food Institute TUoD. FRIDA, DTU Foods Public Food Database; DTU Fødevareinstitutet: Kongens Lyngby, Denmark, 2019; Available online: https://frida.fooddata.dk/pdf/frida-dokumentation-da\_DK-august-2019.pdf (accessed on 15 April 2018).
- 39. Haug, A.; Vhile, S.; Berg, J.; Hove, K.; Egelandsdal, B. Feeding potentially health promoting nutrients to finishing bulls changes meat composition and allow for product health claims. *Meat Sci.* **2018**, 145, 461–468. [CrossRef]
- 40. Norwegian Directorate of Health; Norwegian Food Safety Authority. *A Diet Tool from the Norwegian Directorate of Health and the Norwegian Food Safety Authority*; Norwegian Directorate of Health and the Norwegian Food Safety Authority: Oslo, Norway, 2021. Available online: https://www.kostholdsplanleggeren.no/ (accessed on 1 September 2019).
- 41. Council of the European Communities. Council Directive 90/496/EEC of 24 September 1990 on Nutrition Labelling for Foodstuffs; EUR-Lex (European Union Law): Brussels, Belgium, 1990. Available online: https://eur-lex.europa.eu/legal-content/en/ALL/?uri=CELEX:31990L0496 (accessed on 2 February 2016).
- 42. Brattbakk, H.R.; Arbo, I.; Aagaard, S.; Lindseth, I.; de Soysa, A.K.; Langaas, M.; Kulseng, B.; Lindberg, F.; Johansen, B. Balanced caloric macronutrient composition downregulates immunological gene expression in human blood cells-adipose tissue diverges. *Omics A J. Integr. Biol.* **2013**, *17*, 41–52. [CrossRef]
- 43. Fürst Medisinsk Laboratorium. 2021. Available online: https://www.furst.no/analyse-og-klinikk/ (accessed on 1 November 2021).
- 44. Schurgers, L.J.; Vermeer, C. Determination of phylloquinone and menaquinones in food. Effect of food matrix on circulating vitamin K concentrations. *Haemostasis* **2000**, *30*, 298–307.
- 45. Authority NFS. The Norwegian Food Composition Database. 2021. Available online: https://matvaretabellen.no/?language=en (accessed on 3 January 2017).
- 46. Agricultural Research Service US Department of Agriculture. Food Data Central (2017). 2019. Available online: https://fdc.nal.usda.gov/ (accessed on 17 April 2018).
- 47. EFSA Panel on Additives and Products or Substances used in Animal Feed (FEEDAP); Bampidis, V.; Azimonti, G.; Bastos, M.D.L.; Christensen, H.; Dusemund, B.; Kouba, M.; Kos Durjava, M.; López-Alonso, M.; López Puente, S.; et al. Safety and efficacy of sodium selenate as feed additive for ruminants. *EFSA J.* **2019**, *17*, e05788.
- 48. Wellek, S.; Blettner, M. On the proper use of the crossover design in clinical trials: Part 18 of a series on evaluation of scientific publications. *Dtsch. Arztebl. Int.* **2012**, 109, 276–281.
- 49. Liu, Z.; Li, J.; Chen, D.; Gao, R.; Zeng, W.; Chen, S.; Huang, Y.; Huang, J.; Long, W.; Li, M.; et al. Dynamic Interleukin-6 Level Changes as a Prognostic Indicator in Patients With COVID-19. *Front. Pharmacol.* **2020**, *11*, 1093. [CrossRef]

Foods 2022, 11, 631 20 of 21

50. Biovendor Research and Diagnostics. Quantitative Determination Ofhuman Interleukin-8. 2016. Available online: https://www.biovendor.com (accessed on 26 August 2021).

- 51. Toh, J.; Gavrilova, T.; Rubinstein, A. Three Cases of Elevated IL-1beta in Common Variable Immunodeficiency (CVID) with Autoimmune Complications. *J. Allergy Clin. Immunol.* **2016**, *137*, AB19. [CrossRef]
- 52. Zou, Y.; Chen, X.; Liu, J.; Zhou, D.B.; Kuang, X.; Xiao, J.; Yu, Q.; Lu, X.; Li, W.; Xie, B.; et al. Serum IL-1beta and IL-17 levels in patients with COPD: Associations with clinical parameters. *Int. J. Chron. Obstruct. Pulmon. Dis.* 2017, 12, 1247–1254. [CrossRef] [PubMed]
- 53. Thane, C.W.; Bates, C.J.; Shearer, M.J.; Unadkat, N.; Harrington, D.J.; Paul, A.A.; Prentice, A.; Bolton-Smith, C. Plasma phylloquinone (vitamin K1) concentration and its relationship to intake in a national sample of British elderly people. *Br. J. Nutr.* **2002**, *87*, 615–622. [CrossRef]
- 54. Thane, C.W.; Wang, L.Y.; Coward, W.A. Plasma phylloquinone (vitamin K1) concentration and its relationship to intake in British adults aged 19–64 years. *Br. J. Nutr.* **2006**, *96*, 1116–1124. [CrossRef] [PubMed]
- 55. Schurgers, L.J.; Shearer, M.J.; Hamulyak, K.; Stocklin, E.; Vermeer, C. Effect of vitamin K intake on the stability of oral anticoagulant treatment: Dose-response relationships in healthy subjects. *Blood* **2004**, *104*, 2682–2689. [CrossRef]
- 56. Shearer, M.J.; Rahim, S.; Barkhan, P.; Stimmler, L. Plasma Vitamin-K1 in Mothers and Their Newborn Babies. *Lancet* **1982**, 2, 460–463. [CrossRef]
- 57. Thijssen, H.H.; Drittij, M.J.; Vermeer, C.; Schoffelen, E. Menaquinone-4 in breast milk is derived from dietary phylloquinone. *Br. J. Nutr.* **2002**, *87*, 219–226. [CrossRef]
- 58. Knapen, M.H.; Braam, L.A.; Teunissen, K.J.; Van't Hoofd, C.M.; Zwijsen, R.M.; van den Heuvel, E.G.H.M.; Vermeer, C. Steady-state vitamin K2 (menaquinone-7) plasma concentrations after intake of dairy products and soft gel capsules. *Eur. J. Clin. Nutr.* **2016**, 70, 831–836. [CrossRef]
- 59. Hodges, S.J.; Akesson, K.; Vergnaud, P.; Obrant, K.; Delmas, P.D. Circulating levels of vitamins K1 and K2 decreased in elderly women with hip fracture. *J. Bone. Miner. Res.* **1993**, *8*, 1241–1245. [CrossRef]
- 60. EFSA. Dietary Reference Values: EFSA Publishes Advice on Vitamin K. Available online: https://www.efsa.europa.eu/en/press/news/170522-12017 (accessed on 22 May 2017).
- 61. Institute of Medicine. Dietary Reference Intakes for Vitamin A, Vitamin K, Arsenic, Boron, Chromium, Copper, Iodine, Iron, Manganese, Molybdenum, Nickel, Silicon, Vanadium, and Zinc. In *Nutrients; SoURLo, Intakes; SolaUoDR, Intakes SCotSEoDR, Medicine*; Institute of Medicine (US) Panel on Micronutrients; National Academies Press: Washington, DC, USA, 2001.
- 62. Cashman, K.D.; Seamans, K.M.; Lucey, A.J.; Stocklin, E.; Weber, P.; Kiely, M.; Hill, T.R. Relative effectiveness of oral 25-hydroxyvitamin D3 and vitamin D3 in raising wintertime serum 25-hydroxyvitamin D in older adults. *Am. J. Clin. Nutr.* **2012**, *95*, 1350–1356. [CrossRef]
- 63. Pettersson, J.; Hindorf, U.; Persson, P.; Bengtsson, T.; Malmqvist, U.; Werkstrom, V.; Ekelund, M. Muscular exercise can cause highly pathological liver function tests in healthy men. *Br. J. Clin. Pharmacol.* **2008**, *65*, 253–259. [CrossRef]
- 64. Bragstad, K.; Paulsen, T.H.; Tønnessen, R.; Klüwer, B.; Rydland, K.M.; Aune, T.; Hungnes, O. *Influensasesongen i Norge* 2020-21; Norwegian Institute of Public Health: Oslo, Norway, 2021. Available online: https://www.fhi.no/globalassets/dokumenterfiler/influensa/influensaovervaking/influensasesongen-i-norge-2020-21 (accessed on 1 December 2021).
- 65. An, R.; Andrade, F.; Grigsby-Toussaint, D. Sandwich consumption in relation to daily dietary intake and diet quality among US adults, 2003–2012. *Public Health* **2016**, 140, 206–212. [CrossRef] [PubMed]
- 66. Mensink, R.P. Effects of Saturated Fatty Acids on Serum Lipids and Lipoproteins: A Systematic Review and Regression Analysis; World Health Organization: Geneva, Switzerland, 2016; Available online: https://apps.who.int/iris/handle/10665/246104 (accessed on 13 December 2021).
- 67. Ke, Q.; Chen, C.; He, F.; Ye, Y.; Bai, X.; Cai, L.; Xia, M. Association between dietary protein intake and type 2 diabetes varies by dietary pattern. *Diabetol. Metab. Syndr.* **2018**, *10*, 48. [CrossRef] [PubMed]
- 68. Gibson, S.; Ashwell, M. The association between red and processed meat consumption and iron intakes and status among British adults. *Public Health Nutr.* **2003**, *6*, 341–350. [CrossRef] [PubMed]
- 69. Alleyne, M.; Horne, M.K.; Miller, J.L. Individualized treatment for iron-deficiency anemia in adults. *Am. J. Med.* **2008**, *121*, 943–948. [CrossRef] [PubMed]
- 70. Houghton, L.A.; Parnell, W.R.; Thomson, C.D.; Green, T.J.; Gibson, R.S. Serum Zinc Is a Major Predictor of Anemia and Mediates the Effect of Selenium on Hemoglobin in School-Aged Children in a Nationally Representative Survey in New Zealand. *J. Nutr.* **2016**, 146, 1670–1676. [CrossRef]
- 71. Maehre, H.K.; Malde, M.K.; Eilertsen, K.E.; Elvevoll, E.O. Characterization of protein, lipid and mineral contents in common Norwegian seaweeds and evaluation of their potential as food and feed. *J. Sci. Food Agric.* **2014**, *94*, 3281–3290. [CrossRef]
- 72. Grabež, V.; Mydland, L.T.; Kidane, A.; Øverland, M.; Berg, P.; Fulladosa, E.; Øverland, M.; Mydland, M.T.; Brasas, E.C. *Inclusion of Intact Saccharina Latissima to Finishing Lamb Diet Improved Micronutrient Content in the Meat*; International Congress of Meat Science and Technology: Krakow, Poland, 2021; Available online: https://www.researchgate.net/publication/354544292\_SubID\_621528 08826\_Preferred\_session\_type\_VOS\_-virtual\_oral\_session\_only\_online\_presentation (accessed on 13 December 2021).
- 73. Rayman, M.P.; Blundell-Pound, G.; Pastor-Barriuso, R.; Guallar, E.; Steinbrenner, H.; Stranges, S. A randomized trial of selenium supplementation and risk of type-2 diabetes, as assessed by plasma adiponectin. *PLoS ONE* **2012**, 7, e45269. [CrossRef]

Foods 2022, 11, 631 21 of 21

74. Aro, A.; Alfthan, G.; Varo, P. Effects of supplementation of fertilizers on human selenium status in Finland. *Analyst* **1995**, *120*, 841–843. [CrossRef]

- 75. Hartikainen, H. Biogeochemistry of selenium and its impact on food chain quality and human health. *J. Trace Elem. Med. Biol.* **2005**, *18*, 309–318. [CrossRef]
- 76. Schmid, A.; Walther, B. Natural vitamin D content in animal products. Adv. Nutr. 2013, 4, 453–462. [CrossRef] [PubMed]
- 77. Schurgers, L.J.; Vermeer, C. Differential lipoprotein transport pathways of K-vitamins in healthy subjects. *Biochim. Biophys. Acta-Gen. Subj.* **2002**, 1570, 27–32. [CrossRef]
- 78. Cui, G.; Yuan, A.; Sun, Z.; Zheng, W.; Pang, Z. IL-1beta/IL-6 network in the tumor microenvironment of human colorectal cancer. *Pathol. Res. Pract.* **2018**, 214, 986–992. [CrossRef] [PubMed]