

Synthesis, Crystal Structure Analyses, and Antibacterial Evaluation of the Cobalt(II) Complex with Sulfadiazine-Pyrazole Prodrug

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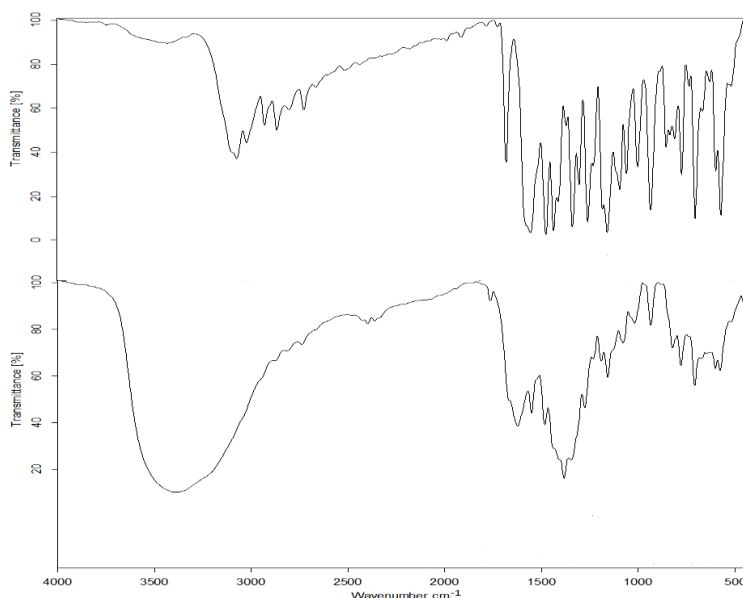


Figure S1. FTIR spectra of the ligand **L** (upper) and $[\text{Co}(\text{L})(\text{H}_2\text{O})_4](\text{NO}_3)_2$ complex (lower).

Method S1

The crystal of $[\text{Co}(\text{L})(\text{H}_2\text{O})_4](\text{NO}_3)_2$ was immersed in cryo-oil, mounted in a loop, and measured at a temperature of 170 K. The X-ray diffraction data were collected using a Bruker Kappa Apex II diffractometer with Mo K-radiation. The Denzo-Scalepack [35] software package was used for cell refinement and data reduction. A multi-scan absorption correction based on equivalent reflections (SADABS [36]) was applied to the intensities before the structure solution. The structure was solved using the intrinsic phasing method with SHELXT [37] software. Structural refinement was carried out using SHELXL [38] software with the SHELXLE [39] graphical user interface. All hydrogen atoms involved in hydrogen bonding were refined to avoid any geometric restrictions. Therefore, H_2O and NH hydrogen atoms were located from the difference Fourier map and refined isotropically. Other hydrogen atoms were positioned geometrically and constrained to ride on their parent atoms, with $\text{C-H} = 0.95\text{-}0.98 \text{ \AA}$ and $\text{Uiso} = 1.2\text{-}1.5 \text{ Ueq}$ (parent atom).

Method S2

1. Antibacterial assessment

The two chemical substances were evaluated for antibacterial efficacy against gram-positive bacteria; *Staphylococcus aureus* (ATCC 25923) and MRSA (1) clinical isolates as well against gram-negative bacteria including; *Klebsiella pneumonia* (ATCC 700603), *Escherichia coli* (ATCC 25922), *Proteus mirabilis* and *Acinetobacter baumannii* (8). Based on the CLSI reference [42], their minimum inhibitory concentrations (MIC) are determined.

2. Determination of minimum inhibitory concentration

Investigation of antibacterial activity of chemical compounds was performed by micro-broth dilution assay for determination of MIC. In summary, 100 μ L of Muller-Hinton broth (MHB) (Oxoid® Limited, Basingstoke, UK) were disseminated in 96 multi-well microtiter plates, followed by the addition of 100 μ L chemical compound into the first row of the microtiter plate. Then, from the first to the twelfth well, serial dilution was performed. Each well received 6 μ L of freshly prepared bacterial suspension (1.5×10^8 cfu/mL). For each bacterial strain, positive and negative controls were carried out. Plates were incubated for 18-24 hours at 37°C, with Amoxicillin 1000 μ g/mL serving as reference standard antibiotic. The MIC was estimated as the minimum concentration that demonstrated no detectable bacterial growth.

References

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