

Supplementary Information for

Article

Synthesis of Peptide-Immobilized Magnetic Beads, and Peptide Reactivity Assay for Assessing Skin Sensitization Utilizing Chromophore

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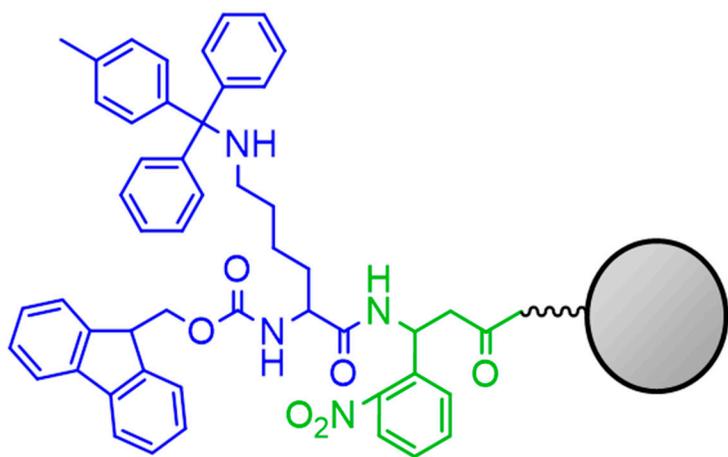
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Fmoc-Lys(Mtt)-npp-beads



Ac-Lys(Flu)-npp-beads

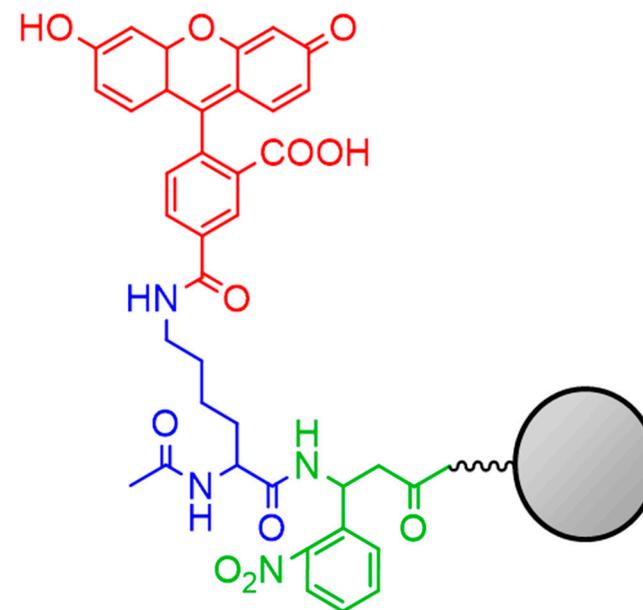


Figure S1. Structures of Fmoc-Lys(Mtt)-npp-beads and fluorescein tagged Ac-Lys(Flu)-npp-beads

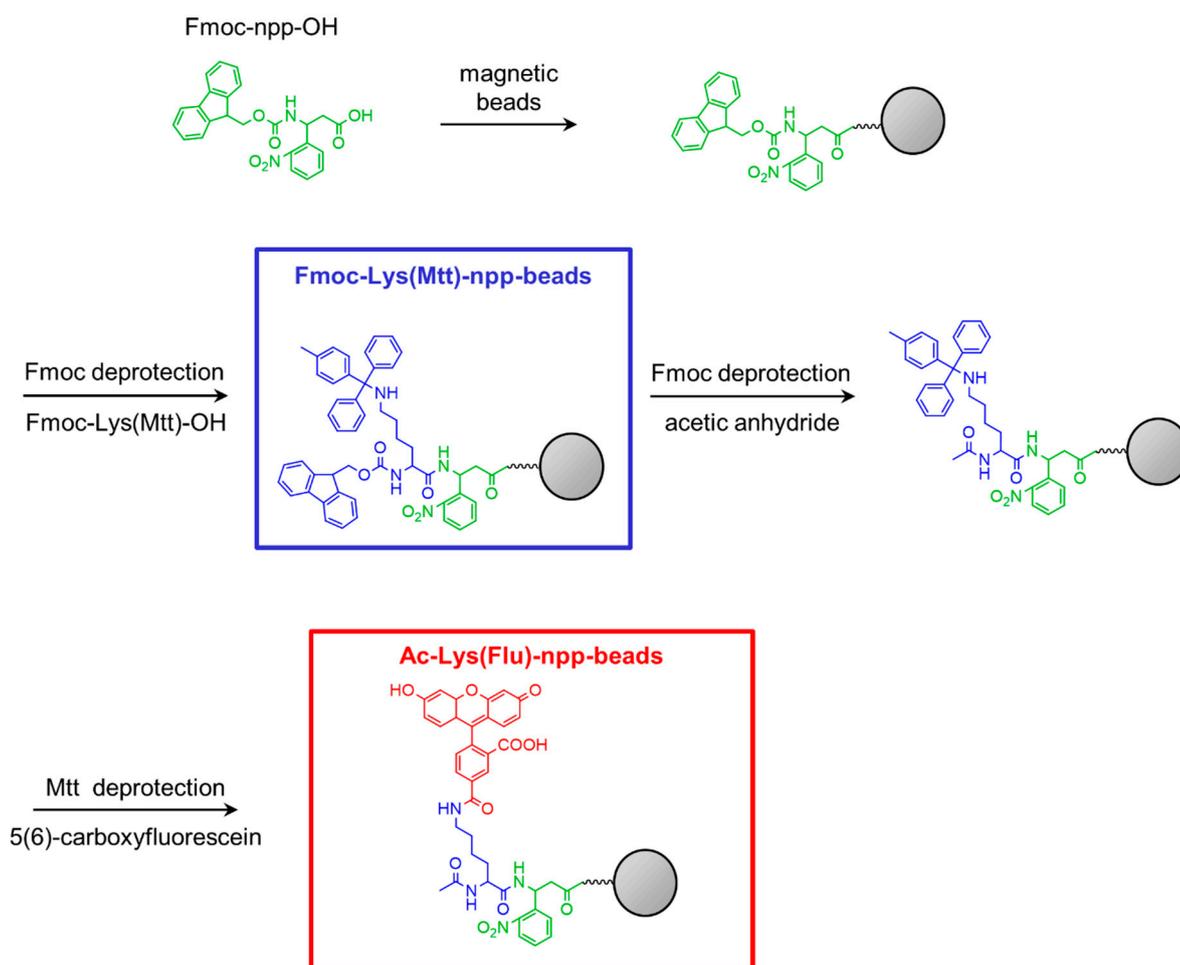


Figure S2. Synthesis of Fmoc-Lys(Mtt)-npp-beads and Ac-Lys(Flu)-npp-beads.

Table S1.Reactivity of test chemicals to Ac-Lys-beads determined by percent depletion

Test Chemicals	log K _o w	LLNA Potency category	Mechanism	DPRA ^{a,b}		ADRA ^d		C-SPRA-MB	
				Mean depletio n ratio (%)	Results ^c	Depletio n ratio (%)	Results ^e	Depletio n ratio (%)	Results ^f
<i>p</i> -Benzoquinone (BQ)	0.25	Extreme	Michael acceptor	95.0 ^a	P	98.2 ^d	P	92.5	P
Fluorescein-5-isothiocyanate (FITC)	4.69	Strong	Acyl-transfer	80.6 ^a	P	100.0 ^d	P	84.6	P
Benzylidene acetone (BA)	2.04	Moderat e	Michael acceptor	48.1 ^a	P	55.1 ^d	P	75.0	P
5-Methyl-2-phenyl-2-hexenal (MPH)	3.77	Moderat e	Michael acceptor /Schiff base	-	-	-	-	46.2	P
Undec-10-enal (UE)	4.12	Moderat e	Schiff base	0.00 ^b	N ^g	-	N ^g	67.5	P
-Amyl cinnamic aldehyde (ACA)	4.33	Weak	Michael acceptor /Schiff base	2.25 ^a	N ^g	4.1 ^d	N ^g	40.0	P
Dibutyl phthalate (DP)	4.61	Non- sensitizer	Non-binding	0.00 ^b	N	-	N	9.7	N

^aData from Ref. 1. ^bData from Ref. 2. ^cThreshold of 6.38% average peptide depletion was used to discriminate between ‘P’ (positive) and ‘N’ (negative). ^dData from Ref. 3. ^eThreshold of 4.9 % mean peptide depletion was used to discriminate between ‘P’ (positive) and ‘N’ (negative). ^f Threshold of 20 % mean peptide depletion was used to discriminate between ‘P’ (positive) and ‘N’ (negative).

^gAlthough they are sensitizers, they showed “false negatives” in DPRA and ADRA.

References

1. Natsch, A., Ryan, C. A., Foertsch, L., Emter, R., Jaworska, J., Gerberick, F. and Kern, P.: A dataset on 145 chemicals tested in alternative assays for skin sensitization undergoing prevalidation, *J. Appl. Toxicol.*, **2013**, 33, 1337–1352.
2. Otsubo, Y., Nishijo, T., Miyazawa, M., Saito, K., Mizumachi, H. and Sakaguchi, H.: Binary test battery with KeratinoSens™ and h-CLAT as part of a bottom-up approach for skin sensitization hazard prediction, *Regul. Toxicol. Pharmacol.*, **2017**, 88, 118–124.
3. Fujita, M., Yamamoto, Y., Tahara, H., Kasahara, T., Jimbo, Y. and Hioki, T.: Development of a prediction method for skin sensitisation using novel cysteine and lysinederivatives, *J. Pharmacol. Toxicol. Methods*, **2014**, 70, 94–105.