



## **Early Post-Natal Immune Activation Leads to Object Memory Deficits in Female** *Tsc2*<sup>+/-</sup> **Mice: The Importance of Including Both Sexes in Neuroscience Research**

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Abstract: There is evidence that viral infections during pre-natal development constitute a risk factor for neuropsychiatric disorders and lead to learning and memory deficits. However, little is known about why viral infections during early post-natal development have a different impact on learning and memory depending on the sex of the subject. We previously showed that early post-natal immune activation induces hippocampal-dependent social memory deficits in a male, but not in a female, mouse model of tuberous sclerosis complex (TSC;  $Tsc2^{+/-}$  mice). Here, we explored the impact of a viral-like immune challenge in object memory. We demonstrate that early post-natal immune activation (during the first 2 weeks of life) leads to object memory deficits in female, but not male, mice that are heterozygous for a gene responsible for tuberous sclerosis complex ( $Tsc2^{+/-}$  mice), while no effect was observed in wild type (WT) mice. Moreover, we found that the same immune activation in Tsc2<sup>+/-</sup> adult mice was not able to cause object memory deficits in females, which suggests that the early post-natal development stage constitutes a critical window for the effects of immune challenge on adult memory. Also, our results suggest that mTOR plays a critical role in the observed deficit in object memory in female  $Tsc2^{+/-}$  mice. These results, together with previous results published by our laboratory, showing sex-specific memory deficits due to early post-natal immune activation, reinforce the necessity of using both males and females for research studies. This is especially true for studies related to immune activation, since the higher levels of estrogens in females are known to affect inflammation and to provide neuroprotection.

Keywords: Tsc2; female; immune activation; mTOR

### 1. Introduction

Evidence shows that viral infections during pre-natal development constitute a risk factor for neuropsychiatric disorders and lead to learning and memory deficits [1–15]. However, because most of those studies were carried out using almost exclusively male rodents, little is known about the impact of sex on the learning and memory effects of viral infections during early post-natal development [16–18].

Tuberous sclerosis complex (TSC) is an autosomal dominant disorder caused by mutations in either the *TSC2* gene located on chromosome 16p13.3 and encoding tuberin, or the *TSC1* gene located on chromosome 9q34 and encoding hamartin proteins [19–21]. TSC is the second most common neurocutaneous disorder (1 in 6000 people are affected worldwide), and is characterized by the growth of numerous benign tumors in many parts



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**Copyright:** © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). of the body, including the brain, skin, heart, lungs and kidneys [22]. Additionally, most TSC patients are affected by neuropsychiatric disorders [23].

Recent publications [24] from our laboratory showed that the challenge of an immune activation in the early post-natal period induces hippocampal-dependent social memory deficits in male, but not female,  $Tsc2^{+/-}$  mice due to the abnormal mammalian target of rapamycin (mTOR)-dependent interferon signaling and subsequent impairments in microglia function. Interestingly, those male  $Tsc2^{+/-}$  mice showed no deficits in object memory despite showing impairment in social memory. Here, we evaluate the effects of early post-natal immune activation on object memory of  $Tsc2^{+/-}$  mice, using the Novel Object Recognition (NOR) task. The NOR test is a cortical-dependent behavioral assay that relies on the animal's innate preference for novelty and is used to investigate non-spatial object memory. This task measures the animal's ability to distinguish a novel object from a familiar one [25–30].

### 2. Materials and Methods

## 2.1. Experimental Design and Subject Details

To generate the  $Tsc2^{+/-}$  mice used in this study to evaluate the impact of early postnatal immune activation in object memory of those mice, we crossed male  $Tsc2^{+/-}$  mice [31] with C57BL/6J wild type (WT) females (JAX, Cat.#: 000664).  $Tsc2^{+/-}$  male breeders were of a C57BL/6Ncrl genetic background (Charles River Laboratories, Cat.#: 027). The pregnant female mice were individually housed and not disturbed, except for a weekly cage change. The pregnancy was confirmed by checking for abdominal distension. Daily checks were conducted on the pregnant females to determine the exact day of parturition (designated as P0). To activate the immune system early post-natally, the pups were intraperitoneally injected with either 20 mg/kg Poly I:C or a vehicle on P3, P7 and P14. To assess the impact of immune activation in adult  $Tsc2^{+/-}$  mice, adult wild-type (WT) and mutant mice ( $Tsc2^{+/-}$ ) were injected with 20 mg/kg Poly I:C three times, followed by a similar schedule to that used during early post-natal development. Tail biopsies for genotyping were collected around P40.

## 2.2. Poly I:C Administration

The Poly I:C potassium salt (Sigma; Cat.#: P9582-50MG) was freshly dissolved in vehicle solution (0.9% sterile saline) before each use. It should be noted that Poly I:C is supplied at 10% of the total weight of the salt, and the dosage was based on the weight of Poly I:C itself.

- (a) Early post-natal administration: At the age of P3, P7 and P14, mutant *Tsc2*<sup>+/-</sup> mice and their WT littermates were injected intraperitoneally with 20 mg/kg Poly I:C. At the same time, mice for the control group were injected with the vehicle.
- (b) Administration in adults: Adult (4–6 months) *Tsc2<sup>+/-</sup>* mice and WT littermates were fed with PLX5622 or control chow for 21 days (see below). The mice were injected intraperitoneally with Poly I:C (20 mg/kg) 10 h, 4 days and 11 days after ending the treatment with PLX5622 or control chow. Thus, the schedule of Poly I:C injections overlapped with the period of microglial repopulation after PLX5622 treatment.

## 2.3. Novel Object Recognition (NOR) Test

Adults (4–6 months)  $Tsc2^{+/-}$  mice and WT littermates were tested using NOR.

The Novel Object Recognition (NOR) test was carried out as described previously [24,32]. This behavioral test involves several steps to assess object memory in mice. For the first step, the mice were handled for eight minutes daily for six consecutive days to acclimate them to the experimenter. Subsequently, over the next two days, they were habituated in an open field ( $41.5 \times 41.5 \times 40.5$  cm) for 12 min each day. During the training session, the mice were placed in the open field with two identical objects and were allowed to explore freely for seven minutes. After 24 h, the mice were tested for object recognition memory with both a previously presented object and a new object. To avoid any kind of bias during the test, the

new object's location was counterbalanced between trials. The mice were trained or tested only once per day, and the open field was cleaned after each session using 70% ethanol. The sessions were video recorded, and 1–2 blinded and experienced observers scored the object exploration time offline using stopwatches. The variation between observers was normally less than 2 s for the training and test sessions. Only when the mouse touched any of the objects with its nose, was exploration counted. All the stages of the experiments as well as the scoring were carried out blind to genotype and treatment condition. Small bottles and containers of different shapes were the objects used in this study.

### 2.4. PLX5622 Treatment

Plexxikon Inc. provided the PLX6622 and control rodent diet. Research Diets Inc. formulated that rodent diet in in AIN-76A standard chow. Over a period of 21 days, adult  $Tsc2^{+/-}$  mice, aged 4–6 months, were fed with PLX5622 (1200 mg/kg; Cat.#: D11100404i) or control chow (Cat.#: D10001i) [24,33], with each mouse receiving 5 g of chow per day.

### 2.5. Rapamycin Treatment

We freshly dissolved rapamycin (5 mg/kg; LC Laboratories, Cat.#: R-5000) in DMSO (Sigma-Aldrich, Cat.#: D5879-500ML) before use. Mice injected at an early post-natal age with Poly I:C were, as adults (4–6 months), treated with a daily intraperitoneal injection of rapamycin (5 mg/kg) or vehicle (DMSO) for 5 days prior to the Novel Object Recognition test. Mice were tested for object memory 18 h after the last injection of rapamycin or DMSO.

## 2.6. Immunohistochemistry

This methodology was carried out in accordance with a previously established protocol [34]. To begin, mice were transcardially perfused with a fixative containing 4% paraformaldehyde, and their brains were subsequently cryoprotected with 30% sucrose. Following this, sagittal free-floating brain sections, each 60  $\mu$ m in thickness, were incubated overnight at 4 °C with polyclonal rabbit anti-Iba1 (Wako Chemicals, Cat.#: 019-19741) at a 1:1000 dilution. Subsequently, the sections were incubated for 90 min with Alexa Fluor 488 goat anti-rabbit IgG (Invitrogen, Cat.#: A11011) at a 1:500 dilution. This was followed by a 15 min incubation in DAPI at a 1:1000 dilution, and a 14 min wash in PBS. The immunofluorescence labeling was then detected using a confocal microscope.

### 2.7. Statistical Analysis

The mouse behavioral data are presented as the mean  $\pm$  SEM and as individual data. Regarding the behavioral experiments, the statistics presented in the figures were initially based on Student's *t*-test. Additionally, the data were subjected to analysis using two-way ANOVA plus Holm–Sidak post hoc analyses, and the results obtained were found to be identical. The figure legends provide the *p*, *n* and *t* values, with a significance level of *p* < 0.05 considered significant. *p* values  $\geq$  0.05 were denoted as non-significant (n.s.), while different levels of significance were indicated as \*, \*\*, \*\*\* or \*\*\*\*, corresponding to *p* < 0.05, *p* < 0.01, *p* < 0.001 and *p* < 0.0001, respectively. The statistical analyses and the generation of graphical representations of the data in this manuscript were performed using GraphPad Prism 7 software.

## 3. Results

# 3.1. Early Post-Natal Immune Activation Induces Long-Lasting Object Memory Deficits in Adult Tsc2<sup>+/-</sup> Female Mice

Male and female WT and  $Tsc2^{+/-}$  mice were injected with either polyinosinic:polycytidylic acid (Poly I:C; 20 mg/kg) or saline (control) i.p. at post-natal day 3 (P3), P7 and P14 (Figure 1a). As adults (4–6 months old), those mice were tested using Novel Object Recognition (NOR) [25–30]. WT mice injected with Poly I:C showed normal object memory (Figure 1b). However, female, but not male,  $Tsc2^{+/-}$  mice injected with Poly I:C early post-natally ( $Tsc2^{+/-}$  Ep) showed object memory deficits (they showed no preference for

the novel object vs. the familiar object during the NOR test; Figure 1c). These results suggest that, in the first two weeks of life,  $Tsc2^{+/-}$  female (but not male) mice are especially sensitive to the impact of immune activation on object memory.



**Figure 1.** Early post-natal immune activation triggers object memory deficits in female  $Tsc2^{+/-}$  mice. (a) Timeline of injections of polyinosinic:polycytidylic acid (Poly I:C) or Saline and behavior approach. (b) Graph shows the percentage of time that mice spent actively exploring the novel and the familiar objects. All groups, male WT/Saline (n = 10; \*\*\*\* p < 0.0001, t = 9.64), female WT/Saline (n = 14; \*\*\*\* p < 0.0001, t = 10.28), male WT/Poly I:C (n = 14; \*\*\*\* p < 0.0001, t = 9.50) and female WT/Poly I:C (n = 15; \*\*\*\* p < 0.0001, t = 9.45) showed normal object memory (they explored significantly more the novel object than the familiar object). (c) Male  $Tsc2^{+/-}$ /Saline (n = 14; \*\*\*\* p < 0.0001, t = 14.29), female  $Tsc2^{+/-}$ /Saline (n = 8; \*\*\*\* p < 0.0001, t = 14.07) and male  $Tsc2^{+/-}$ /Poly I:C (n = 14; \*\*\*\* p < 0.0001, t = 14.29), female  $Tsc2^{+/-}$ /Saline (n = 8; \*\*\*\* p < 0.0001, t = 14.07) and male  $Tsc2^{+/-}$ /Poly I:C (n = 14; \*\*\*\* p < 0.0001, t = 13.88) showed normal object memory (they explored the novel object significantly more than the familiar object). Conversely, female  $Tsc2^{+/-}$ /Poly I:C (n = 14; n.s. p = 0.07, t = 1.88) showed deficits in object memory (showed no preference for the novel object). Data represent means  $\pm$  SEM as well as individual data.

## 3.2. The Administration of a mTOR Inhibitor (Rapamycin) in Adults Can Reverse the Object Memory Deficits of Female Tsc $2^{+/-}$ Ep Mice

The *Tsc2* gene plays a critical role in the regulation of mTOR signaling [35–38], leading to the upregulation of mTOR signaling in both rodents and humans in cases where *Tsc2* levels are reduced [31,39]. Previous findings have demonstrated that the administration of rapamycin, an mTOR inhibitor, has the ability to reverse multiple phenotypes observed in *Tsc2*<sup>+/-</sup> mice [40–42].

Rapamycin was initially developed as an immunosuppressant for organ transplant in humans. Its immunosuppressive effects has been studied in vivo and in vitro [43–45]. Rapamycin is also known for its effects as inhibitor of the mTOR signaling pathway [46,47], which integrates intracellular and extracellular signals to regulate the metabolism, growth, proliferation and survival of the cells [48]. Importantly, previous results in our laboratory [24] showed that the administration of rapamycin in male  $Tsc2^{+/-}$  Ep mice reversed the social memory deficit of these mice. To evaluate whether the inhibition of mTOR signaling in adult female  $Tsc2^{+/-}$  Ep mice was able to reverse the observed object memory deficits,  $Tsc2^{+/-}$  Ep mice were treated at an adult age with rapamycin (5 mg/kg) (Figure 2a). Female  $Tsc2^{+/-}$  Ep mice treated with rapamycin showed normal object memory (preference for the novel object vs. the familiar object; Figure 2b). These results suggest that mTOR signaling has a critical role in the object memory deficits of adult female  $Tsc2^{+/-}$  Ep mice.



**Figure 2.** The administration of a mTOR inhibitor (rapamycin) in adults can reverse the object memory deficits of female  $Tsc2^{+/-}$  Ep mice. (a) Timeline of injections of Poly I:C and Rapamycin (or DMSO in control mice) and behavior approach. (b) Graph shows the percentage of time that mice spent actively exploring the novel and familiar objects. Female  $Tsc2^{+/-}$  Ep/DMSO (n = 5; n.s. p = 0.21, t = 1.33) showed deficits in object memory (showed no preference for the novel object). However, female  $Tsc2^{+/-}$  Ep/Rapamycin (n = 5; \*\*\* p < 0.001, t = 5.19) showed normal object memory (they explored the novel object significantly more than the familiar object). Data represent means  $\pm$  SEM as well as individual data.

## 3.3. Adult Immune Activation Induces no Deficits in Object Memory in Tsc2<sup>+/-</sup> Mice

Next, we aimed to evaluate whether the administration of Poly I:C in  $Tsc2^{+/-}$  adult mice had similar outcomes regarding object memory as we observed in  $Tsc2^{+/-}$  Ep mice. Immune activation at adult age did not provoke object memory deficits, either in female (Figure 3c) or in male (Figure 3d)  $Tsc2^{+/-}$  mice.

PLX5622 is a Colony-Stimulating Factor 1 Receptor (CSF1R) inhibitor known for its effects on microglial depletion and its potential therapeutic benefits. Research has shown that the inhibition of the CSF1R results in the almost complete elimination of microglia brain-wide [24,49], followed by repopulation upon inhibitor withdrawal. Thus, such inhibitors represent a valuable tool for assessing the role of this glial cell population. Previous results in our laboratory [24] showed that the administration of Poly I:C in adults  $Tsc2^{+/-}$  during the repopulation of microglia lead to social memory deficits in both, male

and female  $Tsc2^{+/-}$  mice. Therefore, we used this depletion–repopulation strategy to determine whether the new/immature microglia may re-open a window of sensitivity to immune activation. For this purpose, adult male and female  $Tsc2^{+/-}$  mice were treated with PLX5622 (a CSF1R inhibitor) or control chow for 21 days. Then, all mice returned to normal chow, and were injected with Poly I:C (20 mg/kg) during the repopulation period (at 0, 4 and 11 days after PLX5622/control chow termination). All mice were tested for object memory at 6.5 weeks after the final Poly I:C injection (Figure 3a).





**Figure 3.** Adult immune activation induces no deficits in object memory in  $\text{Tsc2}^{+/-}$  mice. (a) Timeline for treatment with PLX5622 (PLX), for microglial depletion, or control chow, followed by injections of Poly I:C, and subsequent behavior approach. (b) IBA1 immunostaining of  $Tsc2^{+/-}$  Control and PLX5622-treated mice. Treatment with PLX5622 led to a massive reduction in the microglial population in the whole brain (hippocampus is shown here as example). Bar: 200 µm. (c) Female  $Tsc2^{+/-}$  /Poly I:C after PLX (n = 15; \*\*\*\* p < 0.0001, t = 6.66) and female  $Tsc2^{+/-}$  /Poly I:C after control chow (n = 12; \*\* p < 0.01, t = 3.52) showed normal object memory. (d) Male  $Tsc2^{+/-}$  /Poly I:C after PLX (n = 8; \*\*\*\* p < 0.0001, t = 5.73) and male  $Tsc2^{+/-}$  /Poly I:C after control chow (n = 6; \*\*\*\* p < 0.0001, t = 7.76) showed normal object memory. Data represent means  $\pm$  SEM as well as individual data.

By sacrificing sample mice after 21 days of PLX5622/control chow, we first confirmed that the administration of PLX5622 led to a considerable depletion of microglia (Figure 3b). Regarding object memory, neither female (Figure 3c) nor male Tsc2<sup>+/-</sup> mice (Figure 3d) showed any deficits. These results suggest that immune activation in adult Tsc2<sup>+/-</sup> mice does not induce deficits in object memory (as revealed mice treated with control chow). Moreover, the same immune challenge in  $Tsc2^{+/-}$  mice in the microglial repopulation/maturation stage (mice treated with PLX5622) yielded similar results in object recognition task, indicating that microglial immaturity does not seem to re-open a vulnerable window to immune activation for object memory. Taken together, these data indicate that during the first two weeks of life, the object memory of  $Tsc2^{+/-}$  female mice is especially sensitive to immune activation. Also, this sensitivity is sex-dependent, as male mice did not show object memory deficits.

### 4. Discussion

Sex-specific effects, even in animal studies, are often overlooked in neuroscience due to the increased workload necessary to balance male and female subjects. However, the observation of different results in males versus females can be extremely important because these findings help to generate hypotheses about the mechanisms of the observed biological phenomena. Previously [24], our laboratory showed that early post-natal immune activation induces social memory deficits in male, but not female,  $Tsc2^{+/-}$  mice due to an abnormal mammalian target of rapamycin (mTOR)-dependent interferon signaling and subsequent impairments in microglia function. Interestingly, male  $Tsc2^{+/-}$  mice showed no deficits in object memory. Here, we report that early post-natal immune activation triggers an object memory deficit in female, but not male,  $Tsc2^{+/-}$  mice (Table 1). Moreover, the same immune activation applied to adults did not provoke such memory deficits, highlighting the vulnerability of post-natal developmental stages to immune activation. We further showed that treatment with Rapamycin can reverse this object memory deficit, which suggests that mTOR plays an important role in the underlying mechanism. Together, these results strongly suggest that the early post-natal stages of development constitute a vulnerable window where immune activation can trigger learning and memory deficits in  $Tsc2^{+/-}$  mice. Moreover, it seems that mTOR plays a critical role in both phenotypes, as demonstrated by our finding that Rapamycin treatment can reverse both social [24] and object memory deficits. In fact, hamartin (TSC1) and tuberin (TSC2) protein complex negatively regulates mTOR signaling, so the dysregulation of such complex results in mTOR signaling overactivation [50]. Interestingly, clinical trials targeting mTOR inhibition have been carried out for certain symptoms (subependymal astrocytoma, angiomyolipoma) in tuberous sclerosis (TSC) patients [51-53] and is also considered an option for treating neurological symptoms [40,54,55]. In line with this idea, here we showed the reversal of object memory impairment by Rapamycin treatment, which further supports mTOR inhibition as a plausible strategy for treating neurological symptoms in TSC patients.

**Table 1.** Effects of early post-natal immune activation in  $Tsc2^{+/-}$  mice. Summary of our previous and current results. Early post-natal immune activation induces social memory deficits in male but not female  $Tsc2^{+/-}$  mice. Here we show that the same immune activation induces object memory deficits in female but not male  $Tsc2^{+/-}$  mice.  $\sqrt{:}$  Normal memory. X: Memory deficits.

Early Post-Natal Immune Activation		Social Memory (Hippocampal Dependent)	Object Memory (Cortical Dependent)
Male	WT Tsc2 <sup>+/-</sup>	√ X	
Female	WT Tsc2 <sup>+/-</sup>		√ X

Remarkably, the same immune challenge taking place at the same stage of development (i.e., post-natally) triggers different memory phenotypes in a sex-dependent manner in  $Tsc2^{+/-}$  mice. This may be due to the fact that those phenotypes can be associated with two different areas of the brain. The NOR test is a neurobehavioral assay known to be cortical-dependent [56,57], as shown by studies with cortical lesions [58–62], optogenetic stimulation [63] and neuronal activation [64]. On the other hand, social memory has been associated with other brain regions, including the hippocampus [65–67].

Our studies of  $Tsc2^{+/-}$  mice [24], including those presented here, show how early post-natal immune activation leads to social memory (hippocampal-dependent) deficits only in male mice, while the same immune challenge leads to object memory (cortical-dependent) deficits exclusively in female  $Tsc2^{+/-}$  mice. Thus, we speculate that early post-natal immune activation in  $Tsc2^{+/-}$  mice leads to different kinds and/or levels of response in the brain region in a sex-dependent manner. Future studies are required to evaluate this hypothesis and to determine the mechanisms underlying these memory deficits in  $Tsc2^{+/-}$  mice.

TSC is a genetic disorder with an incidence of 1 in 6000 people worldwide caused by mutations in the *TSC1* or *TSC2* genes [23]. Amongst a variety of symptoms, most patients (>90%) are affected by an ample spectrum of cognitive and/or behavioral problems, most of them related to neurodevelopment, including autism spectrum disorders (ASD) (25–50% of TSC cases; [23,68]). Our results support the already described the interactions of early immune activation (e.g., infections in early childhood) within a specific genetic background in neurodevelopment and underline the importance of preventing severe infections during this (early post-natal) developmental stage. Also, they provide further evidence of the sex-dependent differential susceptibility of different neurodevelopmental processes, which may underlie the sex-based bias in those cognitive and behavioral problems.

Historically, neuroscience research has preferentially used male instead of female mice for experiments, with the purpose of avoiding the putative and unknown impact of female cycling hormones on the phenomenon under study. To encourage scientists to use both male and female groups, in 2015 multiple institutions, including the U.S. National Institutes of Health (NIH), established policies to require applicants to report plans to balance male and female samples, cells and animals for their research [69]. Using female mice for research is especially important for neuroprotection studies because estrogen, which is primarily produced in the ovaries, plays an important neuroprotective and anti-inflammatory role [70] in neurological disorders and insults such as stroke, brain injury, Alzheimer's disease and Parkinson's disease [71].

This work emphasizes the necessity of considering both males and females for research studies. Clarifying the mechanisms underlying the different phenotypes observed between males and females, in response to the same challenge (such as immune activation), is critical for developing drugs and treatments for those phenotypes.

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**Data Availability Statement:** PLX5622 was obtained under a material transfer agreement with Plexxikon. All data needed to evaluate the conclusions in the paper are present in the paper.

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### References

- 1. Gogos, A.; Sbisa, A.; Witkamp, D.; van den Buuse, M. Sex differences in the effect of maternal immune activation on cognitive and psychosis-like behaviour in Long Evans rats. *Eur. J. Neurosci.* **2020**, *52*, 2614–2626. [CrossRef] [PubMed]
- Howland, J.G.; Cazakoff, B.N.; Zhang, Y. Altered object-in-place recognition memory, prepulse inhibition, and locomotor activity in the offspring of rats exposed to a viral mimetic during pregnancy. *Neuroscience* 2012, 201, 184–198. [CrossRef]
- Lante, F.; Meunier, J.; Guiramand, J.; Maurice, T.; Cavalier, M.; de Jesus Ferreira, M.C.; Aimar, R.; Cohen-Solal, C.; Vignes, M.; Barbanel, G. Neurodevelopmental damage after prenatal infection: Role of oxidative stress in the fetal brain. *Free Radic. Biol. Med.* 2007, 42, 1231–1245. [CrossRef] [PubMed]
- 4. Zhang, Y.; Cazakoff, B.N.; Thai, C.A.; Howland, J.G. Prenatal exposure to a viral mimetic alters behavioural flexibility in male, but not female, rats. *Neuropharmacology* **2012**, *62*, 1299–1307. [CrossRef] [PubMed]
- Smith, S.E.; Li, J.; Garbett, K.; Mirnics, K.; Patterson, P.H. Maternal immune activation alters fetal brain development through interleukin-6. J. Neurosci. 2007, 27, 10695–10702. [CrossRef] [PubMed]
- 6. Meyer, U.; Feldon, J. Neural basis of psychosis-related behaviour in the infection model of schizophrenia. *Behav. Brain Res.* 2009, 204, 322–334. [CrossRef]
- 7. Shi, L.; Fatemi, S.H.; Sidwell, R.W.; Patterson, P.H. Maternal influenza infection causes marked behavioral and pharmacological changes in the offspring. *J. Neurosci.* 2003, 23, 297–302. [CrossRef]
- 8. Malkova, N.V.; Yu, C.Z.; Hsiao, E.Y.; Moore, M.J.; Patterson, P.H. Maternal immune activation yields offspring displaying mouse versions of the three core symptoms of autism. *Brain Behav. Immun.* **2012**, *26*, 607–616. [CrossRef]
- 9. Choi, G.B.; Yim, Y.S.; Wong, H.; Kim, S.; Kim, H.; Kim, S.V.; Hoeffer, C.A.; Littman, D.R.; Huh, J.R. The maternal interleukin-17a pathway in mice promotes autism-like phenotypes in offspring. *Science* **2016**, *351*, 933–939. [CrossRef]
- 10. Atladottir, H.O.; Thorsen, P.; Ostergaard, L.; Schendel, D.E.; Lemcke, S.; Abdallah, M.; Parner, E.T. Maternal infection requiring hospitalization during pregnancy and autism spectrum disorders. *J. Autism Dev. Disord.* **2010**, *40*, 1423–1430. [CrossRef]
- Patterson, P.H. Immune involvement in schizophrenia and autism: Etiology, pathology and animal models. *Behav. Brain Res.* 2009, 204, 313–321. [CrossRef] [PubMed]
- 12. Brown, A.S.; Sourander, A.; Hinkka-Yli-Salomaki, S.; McKeague, I.W.; Sundvall, J.; Surcel, H.M. Elevated maternal C-reactive protein and autism in a national birth cohort. *Mol. Psychiatry* **2014**, *19*, 259–264. [CrossRef]
- 13. Atladottir, H.O.; Pedersen, M.G.; Thorsen, P.; Mortensen, P.B.; Deleuran, B.; Eaton, W.W.; Parner, E.T. Association of family history of autoimmune diseases and autism spectrum disorders. *Pediatrics* **2009**, *124*, 687–694. [CrossRef]
- 14. Ashwood, P.; Wills, S.; Van de Water, J. The immune response in autism: A new frontier for autism research. *J. Leukoc. Biol.* 2006, *80*, 1–15. [CrossRef] [PubMed]
- Lee, B.K.; Magnusson, C.; Gardner, R.M.; Blomstrom, A.; Newschaffer, C.J.; Burstyn, I.; Karlsson, H.; Dalman, C. Maternal hospitalization with infection during pregnancy and risk of autism spectrum disorders. *Brain Behav. Immun.* 2015, 44, 100–105. [CrossRef] [PubMed]
- Tchessalova, D.; Tronson, N.C. Memory deficits in males and females long after subchronic immune challenge. *Neurobiol. Learn. Mem.* 2019, 158, 60–72. [CrossRef] [PubMed]
- 17. Kohman, R.A.; Tarr, A.J.; Day, C.E.; McLinden, K.A.; Boehm, G.W. Influence of prenatal stress on behavioral, endocrine, and cytokine responses to adulthood bacterial endotoxin exposure. *Behav. Brain Res.* **2008**, *193*, 257–268. [CrossRef]
- Tronson, N.C.; Collette, K.M. (Putative) sex differences in neuroimmune modulation of memory. J. Neurosci. Res. 2017, 95, 472–486. [CrossRef]
- 19. European Chromosome 16 Tuberous Sclerosis Consortium. Identification and characterization of the tuberous sclerosis gene on chromosome 16. *Cell* **1993**, *75*, 1305–1315. [CrossRef]
- van Slegtenhorst, M.; de Hoogt, R.; Hermans, C.; Nellist, M.; Janssen, B.; Verhoef, S.; Lindhout, D.; van den Ouweland, A.; Halley, D.; Young, J.; et al. Identification of the tuberous sclerosis gene TSC1 on chromosome 9q34. *Science* 1997, 277, 805–808. [CrossRef]
- 21. Curatolo, P.; Bombardieri, R.; Jozwiak, S. Tuberous sclerosis. Lancet 2008, 372, 657–668. [CrossRef] [PubMed]
- Mak, B.C.; Yeung, R.S. The tuberous sclerosis complex genes in tumor development. *Cancer Investig.* 2004, 22, 588–603. [CrossRef] [PubMed]
- de Vries, P.J.; Whittemore, V.H.; Leclezio, L.; Byars, A.W.; Dunn, D.; Ess, K.C.; Hook, D.; King, B.H.; Sahin, M.; Jansen, A. Tuberous sclerosis associated neuropsychiatric disorders (TAND) and the TAND Checklist. *Pediatr. Neurol.* 2015, *52*, 25–35. [CrossRef] [PubMed]

- 24. Lopez-Aranda, M.F.; Chattopadhyay, I.; Boxx, G.M.; Fraley, E.R.; Silva, T.K.; Zhou, M.; Phan, M.; Herrera, I.; Taloma, S.; Mandanas, R.; et al. Postnatal immune activation causes social deficits in a mouse model of tuberous sclerosis: Role of microglia and clinical implications. *Sci. Adv.* **2021**, *7*, eabf2073. [CrossRef]
- Antunes, M.; Biala, G. The novel object recognition memory: Neurobiology, test procedure, and its modifications. *Cogn. Process.* 2012, 13, 93–110. [CrossRef]
- Leger, M.; Quiedeville, A.; Bouet, V.; Haelewyn, B.; Boulouard, M.; Schumann-Bard, P.; Freret, T. Object recognition test in mice. *Nat. Protoc.* 2013, *8*, 2531–2537. [CrossRef]
- Cohen, S.J.; Stackman, R.W., Jr. Assessing rodent hippocampal involvement in the novel object recognition task. A review. *Behav. Brain Res.* 2015, 285, 105–117. [CrossRef]
- Lueptow, L.M. Novel Object Recognition Test for the Investigation of Learning and Memory in Mice. J. Vis. Exp. 2017, 126, 55718.
  [CrossRef]
- Barbosa, F.F.; Silva, R.H. Chapter 18—Immediate-Early Gene Expression in Neural Circuits Related to Object Recognition Memory. In *Handbook of Behavioral Neuroscience*; Ennaceur, A., de Souza Silva, M.A., Eds.; Elsevier: Amsterdam, The Netherlands, 2018; Volume 27, pp. 261–271.
- 30. Denninger, J.K.; Smith, B.M.; Kirby, E.D. Novel Object Recognition and Object Location Behavioral Testing in Mice on a Budget. *J. Vis. Exp.* **2018**, *141*, 58593. [CrossRef]
- 31. Onda, H.; Lueck, A.; Marks, P.W.; Warren, H.B.; Kwiatkowski, D.J. Tsc2(+/-) mice develop tumors in multiple sites that express gelsolin and are influenced by genetic background. *J. Clin. Investig.* **1999**, *104*, 687–695. [CrossRef]
- Ennaceur, A.; Delacour, J. A new one-trial test for neurobiological studies of memory in rats. 1: Behavioral data. *Behav. Brain Res.* 1988, 31, 47–59. [CrossRef] [PubMed]
- Dagher, N.N.; Najafi, A.R.; Kayala, K.M.; Elmore, M.R.; White, T.E.; Medeiros, R.; West, B.L.; Green, K.N. Colony-stimulating factor 1 receptor inhibition prevents microglial plaque association and improves cognition in 3xTg-AD mice. *J. Neuroinflamm.* 2015, 12, 139. [CrossRef]
- Lopez-Aranda, M.F.; Acevedo, M.J.; Carballo, F.J.; Gutierrez, A.; Khan, Z.U. Localization of the GoLoco motif carrier regulator of G-protein signalling 12 and 14 proteins in monkey and rat brain. *Eur. J. Neurosci.* 2006, 23, 2971–2982. [CrossRef]
- Kwiatkowski, D.J.; Manning, B.D. Tuberous sclerosis: A GAP at the crossroads of multiple signaling pathways. *Hum. Mol. Genet.* 2005, 14 (Suppl. S2), R251–R258. [CrossRef] [PubMed]
- 36. Huang, J.; Manning, B.D. A complex interplay between Akt, TSC2 and the two mTOR complexes. *Biochem. Soc. Trans.* 2009, 37, 217–222. [CrossRef]
- Huang, J.; Dibble, C.C.; Matsuzaki, M.; Manning, B.D. The TSC1-TSC2 complex is required for proper activation of mTOR complex 2. *Mol. Cell. Biol.* 2008, 28, 4104–4115. [CrossRef] [PubMed]
- Li, Y.; Corradetti, M.N.; Inoki, K.; Guan, K.L. TSC2: Filling the GAP in the mTOR signaling pathway. *Trends Biochem. Sci.* 2004, 29, 32–38. [CrossRef]
- Prabowo, A.S.; Anink, J.J.; Lammens, M.; Nellist, M.; van den Ouweland, A.M.; Adle-Biassette, H.; Sarnat, H.B.; Flores-Sarnat, L.; Crino, P.B.; Aronica, E. Fetal brain lesions in tuberous sclerosis complex: TORC1 activation and inflammation. *Brain Pathol.* 2013, 23, 45–59. [CrossRef]
- Ehninger, D.; Han, S.; Shilyansky, C.; Zhou, Y.; Li, W.; Kwiatkowski, D.J.; Ramesh, V.; Silva, A.J. Reversal of learning deficits in a Tsc2+/- mouse model of tuberous sclerosis. *Nat. Med.* 2008, 18, 5.
- 41. Tsai, P.T.; Hull, C.; Chu, Y.; Greene-Colozzi, E.; Sadowski, A.R.; Leech, J.M.; Steinberg, J.; Crawley, J.N.; Regehr, W.G.; Sahin, M. Autistic-like behaviour and cerebellar dysfunction in Purkinje cell Tsc1 mutant mice. *Nature* **2012**, *488*, 647–651. [CrossRef]
- 42. Sato, A.; Kasai, S.; Kobayashi, T.; Takamatsu, Y.; Hino, O.; Ikeda, K.; Mizuguchi, M. Rapamycin reverses impaired social interaction in mouse models of tuberous sclerosis complex. *Nat. Commun.* **2012**, *3*, 1292. [CrossRef]
- 43. Martel, R.R.; Klicius, J.; Galet, S. Inhibition of the immune response by rapamycin, a new antifungal antibiotic. *Can. J. Physiol. Pharmacol.* **1977**, *55*, 48–51. [CrossRef] [PubMed]
- 44. Chang, J.Y.; Sehgal, S.N. Pharmacology of rapamycin: A new immunosuppressive agent. *Br. J. Rheumatol.* **1991**, 30 (Suppl. S2), 62–65.
- 45. Chen, Y.; Chen, H.; Rhoad, A.E.; Warner, L.; Caggiano, T.J.; Failli, A.; Zhang, H.; Hsiao, C.L.; Nakanishi, K.; Molnar-Kimber, K.L. A putative sirolimus (rapamycin) effector protein. *Biochem. Biophys. Res. Commun.* **1994**, 203, 1–7. [CrossRef] [PubMed]
- 46. Lamming, D.W. Inhibition of the Mechanistic Target of Rapamycin (mTOR)-Rapamycin and Beyond. *Cold Spring Harb. Perspect. Med.* **2016**, *6*, a025924. [CrossRef]
- 47. Ballou, L.M.; Lin, R.Z. Rapamycin and mTOR kinase inhibitors. J. Chem. Biol. 2008, 1, 27–36. [CrossRef] [PubMed]
- 48. Laplante, M.; Sabatini, D.M. mTOR signaling at a glance. J. Cell. Sci. 2009, 122, 3589–3594. [CrossRef]
- Elmore, M.R.; Najafi, A.R.; Koike, M.A.; Dagher, N.N.; Spangenberg, E.E.; Rice, R.A.; Kitazawa, M.; Matusow, B.; Nguyen, H.; West, B.L.; et al. Colony-stimulating factor 1 receptor signaling is necessary for microglia viability, unmasking a microglia progenitor cell in the adult brain. *Neuron* 2014, *82*, 380–397. [CrossRef]
- 50. Julich, K.; Sahin, M. Mechanism-based treatment in tuberous sclerosis complex. Pediatr. Neurol. 2014, 50, 290–296. [CrossRef]
- 51. Franz, D.N.; Belousova, E.; Sparagana, S.; Bebin, E.M.; Frost, M.; Kuperman, R.; Witt, O.; Kohrman, M.H.; Flamini, J.R.; Wu, J.Y.; et al. Efficacy and safety of everolimus for subependymal giant cell astrocytomas associated with tuberous sclerosis complex (EXIST-1): A multicentre, randomised, placebo-controlled phase 3 trial. *Lancet* 2013, *381*, 125–132. [CrossRef]

- 52. Bissler, J.J.; Kingswood, J.C.; Radzikowska, E.; Zonnenberg, B.A.; Frost, M.; Belousova, E.; Sauter, M.; Nonomura, N.; Brakemeier, S.; de Vries, P.J.; et al. Everolimus for angiomyolipoma associated with tuberous sclerosis complex or sporadic lymphangioleiomyomatosis (EXIST-2): A multicentre, randomised, double-blind, placebo-controlled trial. *Lancet* 2013, 381, 817–824. [CrossRef] [PubMed]
- 53. Davies, D.M.; de Vries, P.J.; Johnson, S.R.; McCartney, D.L.; Cox, J.A.; Serra, A.L.; Watson, P.C.; Howe, C.J.; Doyle, T.; Pointon, K.; et al. Sirolimus therapy for angiomyolipoma in tuberous sclerosis and sporadic lymphangioleiomyomatosis: A phase 2 trial. *Clin. Cancer Res.* **2011**, *17*, 4071–4081. [CrossRef] [PubMed]
- 54. Schneider, M.; de Vries, P.J.; Schonig, K.; Rossner, V.; Waltereit, R. mTOR inhibitor reverses autistic-like social deficit behaviours in adult rats with both Tsc2 haploinsufficiency and developmental status epilepticus. *Eur. Arch. Psychiatry Clin. Neurosci.* 2017, 267, 455–463. [CrossRef] [PubMed]
- 55. Ehninger, D.; de Vries, P.J.; Silva, A.J. From mTOR to cognition: Molecular and cellular mechanisms of cognitive impairments in tuberous sclerosis. *J. Intellect. Disabil. Res.* 2009, *53*, 838–851. [CrossRef] [PubMed]
- 56. Winters, B.D.; Saksida, L.M.; Bussey, T.J. Object recognition memory: Neurobiological mechanisms of encoding, consolidation and retrieval. *Neurosci. Biobehav. Rev.* 2008, 32, 1055–1070. [CrossRef] [PubMed]
- de Landeta, A.B.; Pereyra, M.; Medina, J.H.; Katche, C. Anterior retrosplenial cortex is required for long-term object recognition memory. *Sci. Rep.* 2020, *10*, 4002. [CrossRef] [PubMed]
- 58. Winters, B.D.; Forwood, S.E.; Cowell, R.A.; Saksida, L.M.; Bussey, T.J. Double dissociation between the effects of peri-postrhinal cortex and hippocampal lesions on tests of object recognition and spatial memory: Heterogeneity of function within the temporal lobe. *J. Neurosci.* 2004, 24, 5901–5908. [CrossRef]
- 59. Bussey, T.J.; Saksida, L.M.; Murray, E.A. Perirhinal cortex resolves feature ambiguity in complex visual discriminations. *Eur. J. Neurosci.* **2002**, *15*, 365–374. [CrossRef]
- 60. Barker, G.R.; Bird, F.; Alexander, V.; Warburton, E.C. Recognition memory for objects, place, and temporal order: A disconnection analysis of the role of the medial prefrontal cortex and perirhinal cortex. *J. Neurosci.* **2007**, *27*, 2948–2957. [CrossRef]
- 61. Aggleton, J.P.; Albasser, M.M.; Aggleton, D.J.; Poirier, G.L.; Pearce, J.M. Lesions of the rat perirhinal cortex spare the acquisition of a complex configural visual discrimination yet impair object recognition. *Behav. Neurosci.* **2010**, *124*, 55–68. [CrossRef]
- 62. Wilson, D.I.; Langston, R.F.; Schlesiger, M.I.; Wagner, M.; Watanabe, S.; Ainge, J.A. Lateral entorhinal cortex is critical for novel object-context recognition. *Hippocampus* **2013**, *23*, 352–366. [CrossRef] [PubMed]
- 63. Tamura, K.; Takeda, M.; Setsuie, R.; Tsubota, T.; Hirabayashi, T.; Miyamoto, K.; Miyashita, Y. Conversion of object identity to object-general semantic value in the primate temporal cortex. *Science* **2017**, *357*, *687–692*. [CrossRef]
- 64. Wan, H.; Aggleton, J.P.; Brown, M.W. Different contributions of the hippocampus and perirhinal cortex to recognition memory. *J. Neurosci.* **1999**, *19*, 1142–1148. [CrossRef] [PubMed]
- 65. Kogan, J.H.; Frankland, P.W.; Silva, A.J. Long-term memory underlying hippocampus-dependent social recognition in mice. *Hippocampus* **2000**, *10*, 47–56. [CrossRef]
- 66. Tzakis, N.; Holahan, M.R. Social Memory and the Role of the Hippocampal CA2 Region. *Front. Behav. Neurosci.* **2019**, *13*, 233. [CrossRef] [PubMed]
- 67. Lehr, A.B.; Kumar, A.; Tetzlaff, C.; Hafting, T.; Fyhn, M.; Stober, T.M. CA2 beyond social memory: Evidence for a fundamental role in hippocampal information processing. *Neurosci. Biobehav. Rev.* **2021**, *126*, 398–412. [CrossRef] [PubMed]
- 68. de Vries, P.J.; Belousova, E.; Benedik, M.P.; Carter, T.; Cottin, V.; Curatolo, P.; Dahlin, M.; D'Amato, L.; d'Augeres, G.B.; Ferreira, J.C.; et al. TSC-associated neuropsychiatric disorders (TAND): Findings from the TOSCA natural history study. *Orphanet J. Rare Dis.* **2018**, *13*, 157. [CrossRef]
- 69. Arnegard, M.E.; Whitten, L.A.; Hunter, C.; Clayton, J.A. Sex as a Biological Variable: A 5-Year Progress Report and Call to Action. J. Womens Health (Larchmt) 2020, 29, 858–864. [CrossRef]
- 70. Shvetcov, A.; Ruitenberg, M.J.; Delerue, F.; Gold, W.A.; Brown, D.A.; Finney, C.A. The neuroprotective effects of estrogen and estrogenic compounds in spinal cord injury. *Neurosci. Biobehav. Rev.* **2023**, *146*, 105074. [CrossRef]
- Brann, D.W.; Lu, Y.; Wang, J.; Sareddy, G.R.; Pratap, U.P.; Zhang, Q.; Tekmal, R.R.; Vadlamudi, R.K. Brain-Derived Estrogen and Neurological Disorders. *Biology* 2022, 11, 1698. [CrossRef]

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