



Article UFR2709, an Antagonist of Nicotinic Acetylcholine Receptors, Delays the Acquisition and Reduces Long-Term Ethanol Intake in Alcohol-Preferring UChB Bibulous Rats

Gabriel Gálvez ¹, Juan Pablo González-Gutiérrez ², Martín Hödar-Salazar ^{3,4}, Ramón Sotomayor-Zárate ⁵, María Elena Quintanilla ⁶, María Elena Quilaqueo ¹, Mario Rivera-Meza ^{1,*} and Patricio Iturriaga-Vásquez ^{4,*}

- ¹ Laboratory of Experimental Pharmacology, Faculty of Chemical Sciences and Pharmacy, University of Chile, Santiago 8380494, Chile; mj_galvez@hotmail.com (G.G.); maria.quilaqueo@ug.uchile.cl (M.E.Q.)
- ² Instituto de Ciencias Químicas Aplicadas, Facultad de Ingeniería, Universidad Autónoma de Chile, Talca 3467987, Chile; juan.gonzalez01@uautonoma.cl
- ³ Programa de Doctorado en Ciencias, Mención Biología Celular y Molecular Aplicada, Universidad de La Frontera, Temuco 4811230, Chile; martinhodar@gmail.com
- ⁴ Laboratorio de Farmacología Molecular y Química Medicinal, Facultad de Ingeniería y Ciencias, Universidad de La frontera, Temuco 4811230, Chile
- Centro de Neurobiología y Fisiopatología Integrativa (CENFI), Instituto de Fisiología, Facultad de Ciencias, Universidad de Valparaíso, Valparaíso 2360102, Chile; ramon.sotomayor@uv.cl
- Program of Molecular and Clinical Pharmacology, ICBM, Faculty of Medicine, University of Chile, Santiago 8380494, Chile; equintanilla@med.uchile.cl
- * Correspondence: mario.rivera@ciq.uchile.cl (M.R.-M.); patricio.iturriaga@ufrontera.cl (P.I.-V.)

Abstract: Alcoholism is a worldwide public health problem with high economic cost and which affects health and social behavior. It is estimated that alcoholism kills 3 million people globally, while in Chile it is responsible for around 9 thousand deaths per year. Nicotinic acetylcholine receptors (nAChRs) are ligand-gated ion channels expressed in the central nervous system, and they were suggested to modulate the ethanol mechanism involved in abuse and dependence. Previous work demonstrated a short-term treatment with UFR2709, a nAChRs antagonist, which reduced ethanol intake using a two-bottle free-choice paradigm in University of Chile bibulous (UChB) rats. Here, we present evidence of the UFR2709 efficacy in reducing the acquisition and long-term ethanol consumption. Our results show that UFR2709 (2.5 mg/kg i.p.) reduces the seek behavior and ethanol intake, even when the drug administration was stopped, and induced a reduction in the overall ethanol intake by around 55%. Using naïve UChB bibulous rats, we demonstrate that UFR2709 could delay and reduce the genetically adaptive impulse to seek and drink ethanol and prevent its excessive intake.

Keywords: UFR2709; voluntary ethanol intake; UChB rats; nicotinic antagonist

1. Introduction

Alcoholism is a worldwide public health problem with high economic cost and which affects health and social behavior. It is estimated that alcoholism kills 3 million people globally, while in Chile it is responsible for around 9 thousand deaths per year. Genetic and environmental conditions can lead to uncontrolled consumption of ethanol, one of the most used substances of abuse in the world [1]. Alcohol and nicotine are commonly co-abused [2,3] and many people who are considered heavy drinkers also smoke tobacco [4]. Specific common genes are suggested to regulate the susceptibility to both alcohol and nicotine dependence [2,5–9]. In this regard, there is evidence that shows a direct or indirect interaction between the nicotinic acetylcholine receptor (nAChR) and ethanol [10–12]. In addition, nAChRs were identified in the mesocorticolimbic dopamine system, and their role in alcoholism was proposed [2,13–15]. It was demonstrated that different nicotinic ligands



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Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). can reduce ethanol ingestion in animal models and humans. [16–21]. The reward system or mesocorticolimbic system projects dopamine neurons from the ventral tegmental area (VTA) to the nucleus accumbens (NAc) and prefrontal cortex (PFC) [22,23]. This system is regulated by several neurotransmitters such as serotonin, glutamate, and acetylcholine, among others, which stimulate the firing rate of VTA DA neurons [24,25]. The activation of the reward system increases the NAc DA release which is mediated by reinforcement produced by drugs of abuse, such as amphetamine, cocaine, nicotine, and ethanol [26]. Although many studies about the rewarding properties of ethanol have been carried out [26,27], the elucidation of a specific mechanism for ethanol action is not completely resolved. Moreover, ethanol has been demonstrated to interact with glycine and GABAA

receptors as a positive allosteric modulator and as a negative allosteric modulator of the N-methyl-D-aspartate (NMDA) receptor [28–30]. Additionally, it was shown that ethanol indirectly activates nAChR by increasing VTA acetylcholine levels [10–12,31], promoting the NAc DA release [32,33]. The activation of the reward system by nicotinic receptor agonism and its potentiation induced by ethanol [11,14,26,34] has opened a potential therapeutic target for treating nicotine addiction and alcoholism using nAChRs ligands.

nAChRs are ligand-gated ion channels that are present in the central nervous system (CNS), where the heteromeric $\alpha 4\beta 2$ and the homomeric $\alpha 7$ subtypes are the most abundant [35,36]. However, different heteromeric nAChRs subtypes are expressed in the brain, i.e., $\alpha 4\beta 2\alpha 5$, $\alpha 6\beta 2\beta 3$, $\alpha 4\beta 2\alpha 6$, $\alpha 4\beta 4$, $\alpha 3\beta 4$, and $\alpha 3\beta 2$, but they are less expressed or have a more specific distribution [35]. Several works have shown that ethanol effects involve different nAChRs subtype activation [37–39]. The α 7 [40] and the α 3 β 4 nAChRs [41,42] were implicated in ethanol intake, and the sedative effects of ethanol were associated with receptors containing the α 5 nAChR subunit [43]. However, due to the relevant role of the α 4 β 2 nAChR subtype in the brain reward system, it is suggested that this receptor subtype plays a pivotal function in ethanol intake. In this regard, it was established that a cholinergicdopaminergic reward axis exists [44], which is affected by ethanol [45]. Dopaminergic ligands (agonist, partial agonist, and antagonist) have shown the role of the dopaminergic system in ethanol intake. The dopamine D1 receptor partial agonist SFK 38393, and the D2 receptor agonist Bromocriptine reduce ethanol preference and intake [46,47], and the 7-OH-DPAT, a D2/D3 receptor agonist, increases ethanol intake at lower doses. However, the dopaminergic antagonists SCH 23990, raclopride, and risperidone do not affect ethanol intake and preference [46].

Pharmacological studies using different nAChRs ligands (partial agonist, noncompetitive and competitive antagonist) have established a key role of nAChR in the modulation of the rewarding effects of ethanol [48]. Cytisine and varenicline, two partial agonists of nAChRs [49–52], decrease ethanol intake in rodents after a single dose or short-term administration [53–55]. The inhibitory effects of varenicline and cytisine on alcohol intake in alcohol-preferring University of Chile Bibulous (UChB) rats were confirmed in our lab [17]. On the other hand, mecamylamine, a non-competitive antagonist of central and peripheral nAChR, decreases ethanol consumption in rats [12,56,57] and decreases ethanol-induced DA release in Nac [56–58]. Additionally, erysodine, a competitive antagonist of nAChR, was shown to noticeably decrease ethanol consumption in UChB rats [19]. Moreover, we recently demonstrated, using UChB rats, that UFR2709, a competitive nicotinic antagonist [59], reduces voluntary ethanol intake in a dose-dependent manner without affecting body weight or locomotor activity, and does not produce a DA release from the striatum under i.p. injection [20]. In this work, we studied the effect of UFR2709 administration in the acquisition and the maintenance of ethanol intake in UChB rats that were chronically exposed to ethanol consumption. We found that UFR2709 decreased alcohol consumption at both stages even after ceasing its administration to the animals.

2. Materials and Methods

2.1. Drugs and Drinking Solutions

UFR2709-HCL (M.W. 255.74 g/mol) was prepared in our lab as previously described [57]. The injection volume was calculated related to body weight to reach the applied dose of UFR2709-HCl. Absolute ethanol (Merck, Darmstadt, Germany) and tap water were used to prepare a 10% v/v of ethanol solution. This concentration was used based on previous studies using UChB rats [60,61].

2.2. Animals

The experiments were carried out in adult male Wistar-UChB rats (n = 14), a half (n = 7) for saline solution and the other half for UFR2709 administration. The UChB rat line has been bred for over 90 generations to ingest 10% v/v ethanol solutions in preference to water [60,61]. UChB rats are genetically adapted and are considered a high drinking ethanol model and are used to screen drugs to treat chronic ethanol intake [61]. UChB rats were individually housed with free access to food and water. Animals were maintained under temperature- and humidity-controlled conditions with a 12 h light-dark cycle (lights off at 19:00 h). The animal's weight was around 240 to 280 g. Ethanol intake experiments were performed under ARRIVE guidelines [62], at the faculty of Medicine, University of Chile, and were approved by the Animal Experimentation Ethics Committee of the University of Chile.

2.3. Effect of UFR2709 (2.5 mg/kg) on Voluntary Ethanol Intake by UChB Rats

Two-bottle free-choice experiments [17,60,61,63] were used to test the effects of a fixed dose of UFR2709 (2.5 mg/kg) on the voluntary ethanol intake of UChB rats. UChB rats were individually exposed to continuous access (24 h/day) of ethanol 10% v/v or distilled water in a two-bottle free-choice protocol. To prevent a potential position preference, bottles were daily alternated. All rats used were naïve but came from UChB bibulous mothers. The rats were then randomly divided into two groups (n = 7 per group), and a daily i.p. injection of UFR2709-HCl (2.5 mg/kg) or saline (1 mL/kg) was administered for 7 days at 15:00 h in a two-cycle paradigm, at days 1 to 7 and days 63 to 69 of the experiment (Figure 1). On days 1–2, animals were injected with UFR2709 or vehicle but were not given alcohol access, with the purpose to achieve pharmacologically effective drug concentration in the rats, before starting ethanol exposure. On day 3, ethanol access was started under 24 h continuous access two-bottle free-choice paradigm and maintained until day 100.



Figure 1. Arrows represent the days where UFR2709 and saline were administered, the blue square indicated the days where the drug was administered without access to ethanol, and the green timeline indicates the days where the ethanol intake protocol was conducted.

The second round of UFR2709 or saline administration was verified between days 63 to 69 (Figure 1). Animals were allowed *ad libitum* access to food and water. Ethanol and water intake were recorded at 14:00 h each day and expressed as g ethanol/kg/day

and g water/kg/day, respectively. The weight was recorded once a week until the end of the experiments.

2.4. Statistical Analysis

Differences in ethanol intake between UFR2709- and saline-treated animals were analyzed using two-way ANOVA with Bonferroni's multiple comparison test (Figures 2A, 3 and 4A,B). An unpaired *t*-test was used to analyze the effect of saline or UFR2709 administration on average ethanol intake (Figure 2B). Data are expressed as mean \pm SEM. Statistical analyses were performed using Graph Pad Prism 8.0 software (Graph Pad Software, San Diego, CA, USA), and the level of statistical significance was set at *p* < 0.05.



Figure 2. (**A**) A two-bottle free-choice experiment was used to test the effects of a fixed dose of UFR2709 (2.5 mg/kg) on the voluntary ethanol intake of UChB rats per 100 days. Each point represents the average ethanol intake for 7 animals per group during the experiment period. Rats (n = 7 per group) were administered a single i.p. injection of UFR2709 (2.5 mg/kg/day) or saline (1 mL/kg/day) at 15:00 h, and ethanol consumption was recorded at 14:00 h the next day. Drug administration was made in two stages, the first treatment was the days 1st to 7th and the second treatment was the days 63rd to 69th (labeled in gray in the graph). (**B**) Total average ethanol intake of the two stages of the experiments, days 3–62 in the first period and days 63–100 in the second one. An unpaired *t*-test was used to compare UFR2709 and saline group averages. Data are expressed as mean \pm SEM (g/kg/day), and the number inside the bar indicates the percentage of reduction in ethanol intake compared to the saline-treated group. **** p < 0.0001.



Figure 3. Preference for ethanol solution of the animals treated with UFR2709 or saline (gray bars). Ethanol preference data are expressed as mean \pm SEM (g/kg/day) and represent the percentage of total daily fluid intake that was ingested from the bottle containing the ethanol solution. Two-way ANOVA with Bonferroni's multiple comparison test was used to analyze the effect of UFR2709 treatment on ethanol preference. **** *p* < 0.0001.



Figure 4. (**A**) Total fluid intake of 2.5 mg/kg of UFR2709 compared to the saline group. A two-way ANOVA with Bonferroni's multiple comparison test between drug, saline, and days showed that UFR2709 produces significant differences in total fluid intake. (**B**) Data show the body weight of animal during the time of the experiment, all animal increases their body weight over the course of the experiment. Unpaired *t*-test was used to analyze the effect of UFR2709 treatment versus saline-treated group on the average ethanol consumption. *** *p* < 0.001.

3. Results

To determine the effect of a fixed dose of UFR2709 on the acquisition and long-term maintenance of voluntary ethanol intake, alcohol-preferring UChB rats were given a free choice between ethanol (10% v/v) and water. The dose of UFR2709 (2.5 mg/kg for 7 days) was selected from our previous work [20], where we demonstrated that this dose showed higher efficacy in reducing ethanol intake in this animal model of alcoholism. The first cycle of UFR 2709 administration (days 1-7) resulted in a marked delay in the acquisition of ethanol intake compared to the saline-treated animals (Figure 2A). Upon stopping UFR2709 administration, animals increase their ethanol intake steadily but did not reach the level of ethanol consumption shown by the saline-treated group. The second round of UFR2709 administration (days 63–69) resulted in a pronounced reduction of ethanol intake by the animals. Interestingly, after UFR2709 discontinuation, the rats showed a less pronounced increase in ethanol intake in comparison to the first cycle of UFR administration. A two-way ANOVA with Bonferroni's multiple comparison test between drug, saline and days showed that 2.5 mg/kg of UFR2709 significantly reduced ethanol intake (Figure 2A) compared with the saline group over all the time of the experiment (interaction $[F_{(99,1200)} = 1.992,$ p < 0.0001]; days [F (99,1200) = 5.171, p < 0.0001]; treatment [F (1,1200) = 2028, p < 0.0001]). For the first stage of the experiment (days 3 to 62) an unpaired t-test indicates a significant reduction of ethanol intake by UFR2709 (-52.77%) ([F (6.6) = 1.929, p < 0.0001]). Additionally, at the second stage of the experiment, UFR2709 administration induced a 57.22% of reduction in ethanol intake compared with the saline group ([F $_{(6,6)}$ = 1.596, p < 0.0001]) (Figure 2B).

Additionally, UFR2709 treatment significantly decreased the preference of the animals for the ethanol solution in the first stage of the experiment (days 3 to 62) compared to the saline-treated animals. In the second period of the experiment (days 63–100), the preference of the animals remained lower (approximately 60%) in comparison to the saline group, which showed ethanol preference close to 80% (Figure 3) (interaction [F (99,1200) = 1.442, p = 0.004]; days [F (99,1200) = 5.969, p < 0.0001]; treatment [F (1,1200) = 928.7, p < 0.0001]).

A two-way ANOVA with Bonferroni 's multiple comparison test between drug, saline and days showed that 2.5 mg/kg of UFR2709 produced significant differences on total fluid intake (Figure 4A) compared to the saline group (interaction [F $_{(99,1200)} = 1.748$, p < 0.0001]; days [F $_{(99,1200)} = 6.938$, p < 0.0001]; treatment [F $_{(1,1200)} = 350.7$, p < 0.0001]). In the first stage of the experiment, the UFR2709 treatment did affect the total fluid intake (days 3 to 62)

compared to the saline-treated animals. However, in the second period of the experiment (days 63–100), the total fluid intake of the animals treated with UFR2702 showed a lower total fluid intake in comparison to the saline group (Figure 4A). The administration of UFR2709 and the time course of the experiment did not affect the body weight compared to the saline group, and a normal increase in body weight was observed during the course of this study (interaction [F _(13,168) = 0.4239, *p* = 0.9596]; days [F _(13,168) = 21.12, *p* < 0.0001]; treatment [F _(13,168) = 32.55, *p* < 0.0001]) (Figure 4B).

4. Discussion

In this work, we studied the effects of a competitive nicotinic antagonist (UFR2709) on the acquisition and long-term ethanol consumption of alcohol-drinking UChB rats. This rat line has been bred for over 90 generations to ingest 10% v/v ethanol solution in preference to water [60,61] and could be considered a suitable model for screening drugs to treat chronic ethanol consumption [61]. We previously demonstrated that UFR2709 decreases ethanol intake at different doses in a short-term experiment (17 days), where we found that 2.5 mg/kg was the most effective dose of UFR2709 in reducing ethanol intake in UChB rats [20]. Therefore, we used this dose concentration to perform a long-term experiment.

One of the main differences between the protocol used in this experiment compared with our previous report is the fact that our rats, despite coming from bibulous mothers and fathers, were naïve to ethanol, and the acquisition of ethanol intake starts on day 3 after the drug administration. Previously, we used rats that were subjected to a homecage with free access to ethanol and after twenty days of ethanol intake, a stable plateau of ingestion was reached and the acquisition of ethanol intake behavior was established and this animal could be considered in a bibulous stage. Here, our protocol trends unveil that blocking the nicotinic systems associated with ethanol consumption before exposing rats to free access to ethanol, could decrease the instinct of the UChB rats to seek and ingest ethanol, considering that UChB rats have been genetically adapted for generations for the excessive consumption of ethanol. In this regard, rats were administered with the drugs two days before the start of ethanol access, only on the 3rd day were animals allowed ethanol consumption and water in the two-bottle free-choice paradigm, and the volumes of water and ethanol were recorded each day. Our results showed an interesting decrease in ethanol consumption mediated by UFR2709 administration compared to the saline-treated group. During the days of drug administration (days 1–7), animals reduce their impulsive behavior to ethanol intake significantly and when the drug administration was stopped, they progressively start to increase their ethanol ingestion but never reach the intake shown by the saline group. After the second period of UFR2709 administration, around the 60th day of the experiment, animals reduce their ethanol consumption to the levels observed after the first round of administration (~ 2.5 g ethanol/kg/day). When UFR2709 administration was stopped, animals slowly increase their ethanol intake but achieved a level of ethanol intake that was markedly lower than the level observed in vehicle-treated animals (~4.0 vs. 8.0 g ethanol/mg/kg), and the maximum level of ethanol consumption after the first round of UFR2709 (~ 6.0 g ethanol/kg/day) (Figure 2A). Our results indicate that UFR2709 significantly reduces the ingestion of ethanol even when the drug administration was stopped and produces a reduction of this behavior over all the time of the experiment. Additionally, we show that the body weight increases as rats become older, and no negative change was observed throughout the experiment at around 14 weeks (Figure 4B). On average, the total ethanol consumption of UChB rats treated with 2.5 mg/kg of UFR2709 was around 55% less compared with the saline group after two sessions of drug injection (Figure 2B) and a concomitant increase in water intake compared with the saline group. Our previous work shows that the effect of UFR2709 on ethanol intake does not result in inhibition of locomotor activity and no change in body weight was observed. Additionally, we demonstrated by microdialysis in the striatum that UFR2709 does not produce DA release, even more, it is possible to observe a small decrease in the DA levels [20]. Our study does not demonstrate the molecular mechanism involved

in the reduction of ethanol consumption mediated by UFR2709 but clearly shows that nicotinic antagonism delays the acquisition and reduces the ethanol intake in long-term experiments and could be used as therapeutic drugs to treat or prevent alcohol abuse.

5. Conclusions

Our findings show that UFR2709 decreases ethanol consumption in UChB bibulous rats, suggesting that blocking brain nicotinic receptors inhibits the behavior of seeking and ingesting ethanol. The UChB rat line is genetically conditioned to ingest ethanol in preference to water. UFR2709 administered before ethanol access shows that this drug could stop the impulsive/genetically adapted necessity of ethanol ingestion. These findings further support the view that nicotinic receptors are implicated in the regulation of alcohol intake and that blocking these receptors reduces the instinctive necessity of ethanol intake. Further studies are needed to understand the molecular and biochemical mechanism involved in alcoholism mediated by nicotinic receptor antagonists.

Author Contributions: M.R.-M. and P.I.-V. designed the experiments. G.G., M.H.-S., M.E.Q. (Maria Elena Quilaqueo), and J.P.G.-G. performed the experiments. R.S.-Z., M.R.-M. and P.I.-V. performed the statistical data analysis. M.R.-M., M.E.Q. (Maria Elena Quintanilla), R.S.-Z. and P.I.-V. made the figures. P.I.-V. wrote the manuscript. M.R.-M., R.S.-Z. and P.I.-V. edited the manuscript. All authors have read and agreed to the published version of the manuscript.

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References

- 1. WHO. Global Status Report on Alcohol and Health; World Health Organization: Geneva, Switzerland, 2018.
- Madden, P.A.F.; Heath, A.C. shared genetic vulnerability in alcohol and cigarette use and dependence. *Alcohol. Clin. Exp. Res.* 2002, 26, 1919–1921. [CrossRef] [PubMed]
- Talcott, G.W.; Poston, W.S.; Haddock, C.K. Co-occurrent use of cigarettes, alcohol, and caffeine in a retired military population. *Mil. Med.* 1998, 163, 133–138. [CrossRef] [PubMed]
- Moss, H.B.; Chen, C.M.; Yi, H.Y. Subtypes of alcohol dependence in a nationally representative sample. *Drug Alcohol Depend*. 2007, 91, 149–158. [CrossRef] [PubMed]
- Bergstrom, H.C.; Palmer, A.A.; Wood, R.D.; Burkhart-Kasch, S.; McKinnon, C.S.; Phillips, T.J. Reverse selection for differential response to the locomotor stimulant effects of ethanol provides evidence for pleiotropic genetic influence on locomotor response to other drugs of abuse. *Alcohol Clin. Exp. Res.* 2003, 27, 1535–1547. [CrossRef] [PubMed]
- 6. Collins, A.C.; Romm, E.; Selvaag, S.; Turner, S.; Marks, M.J. A comparison of the effects of chronic nicotine infusion on tolerance to nicotine and cross-tolerance to ethanol in long- and short-sleep mice. *J. Pharmacol. Exp. Ther.* **1993**, *266*, 1390–1397.
- de Fiebre, C.M.; Romm, E.; Collins, J.T.; Draski, L.J.; Deitrich, R.A.; Collins, A.C. Responses to cholinergic agonists of rats selectively bred for differential sensitivity to ethanol. *Alcohol Clin. Exp. Res.* 1991, 15, 270–276. [CrossRef]
- 8. de Fiebre, N.C.; Dawson, R., Jr.; de Fiebre, C.M. The selectively bred high alcohol sensitivity (HAS) and low alcohol sensitivity (LAS) rats differ in sensitivity to nicotine. *Alcohol Clin. Exp. Res.* **2002**, *26*, 765–772. [CrossRef]
- 9. Kendler, K.S.; Myers, J.; Prescott, C.A. Specificity of genetic and environmental risk factors for symptoms of cannabis, cocaine, alcohol, caffeine, and nicotine dependence. *Arch. Gen. Psychiatry* **2007**, *64*, 1313–1320. [CrossRef]

- 10. Blomqvist, O.; Soderpalm, B.; Engel, J.A. Ethanol-induced locomotor activity: Involvement of central nicotinic acetylcholine receptors? *Brain Res. Bull.* **1992**, *29*, 173–178. [CrossRef]
- 11. Davis, T.J.; de Fiebre, C.M. Alcohol's actions on neuronal nicotinic acetylcholine receptors. Alcohol Res. Health 2006, 29, 179–185.
- 12. Le, A.D.; Corrigall, W.A.; Harding, J.W.; Juzytsch, W.; Li, T.K. Involvement of nicotinic receptors in alcohol self-administration. *Alcohol Clin. Exp. Res.* **2000**, *24*, 155–163. [CrossRef] [PubMed]
- John, U.; Meyer, C.; Rumpf, H.J.; Schumann, A.; Thyrian, J.R.; Hapke, U. Strength of the relationship between tobacco smoking, nicotine dependence and the severity of alcohol dependence syndrome criteria in a population-based sample. *Alcohol Alcohol.* 2003, *38*, 606–612. [CrossRef]
- 14. Dani, J.A.; Harris, R.A. Nicotine addiction and comorbidity with alcohol abuse and mental illness. *Nat. Neurosci.* 2005, *8*, 1465–1470. [CrossRef] [PubMed]
- 15. Falk, D.E.; Yi, H.; Hiller-Sturmhöfel, S. An epidemiologic analysis of co-occurring alcohol and tobacco use and disorders. *Alcohol Res. Health* **2006**, *29*, 162–171. [PubMed]
- 16. Chatterjee, S.; Bartlett, S.E. Neuronal nicotinic acetylcholine receptors as pharmacotherapeutic targets for the treatment of alcohol use disorders. *CNS Neurol. Disord. Drug Targets* **2010**, *9*, 60–76. [CrossRef]
- Sotomayor-Zarate, R.; Gysling, K.; Busto, U.E.; Cassels, B.K.; Tampier, L.; Quintanilla, M.E. Varenicline and cytisine: Two nicotinic acetylcholine receptor ligands reduce ethanol intake in University of Chile bibulous rats. *Psychopharmacology* 2013, 227, 287–298. [CrossRef]
- Rahman, S.; Engleman, E.A.; Bell, R.L. Nicotinic receptor modulation to treat alcohol and drug dependence. *Front. Neurosci.* 2015, *8*, 426. [CrossRef]
- Quiroz, G.; Guerra-Díaz, N.; Iturriaga-Vásquez, P.; Rivera-Meza, M.; Quintanilla, M.E.; Sotomayor-Zárate, R. Erysodine, a competitive antagonist at neuronal nicotinic acetylcholine receptors, decreases ethanol consumption in alcohol-preferring UChB rats. *Behav. Brain Res.* 2018, 349, 169–176. [CrossRef]
- Quiroz, G.; Sotomayor-Zárate, R.; González-Gutierrez, J.P.; Vizcarra, F.; Moraga, F.; Bermudez, I.; Reyes-Parada, M.; Quintanilla, M.E.; Lagos, D.; Rivera-Meza, M.; et al. UFR2709, a Nicotinic Acetylcholine Receptor Antagonist, Decreases Ethanol Intake in Alcohol-Preferring Rats. *Front Pharmacol.* 2019, 10, 1429. [CrossRef]
- Mitchell, J.M.; Teague, C.H.; Kayser, A.S.; Bartlett, S.E.; Fileds, H.L. Varenicline decreases alcohol consumption in heavy-drinking smokers. *Psychopharmacology* 2012, 223, 299–306. [CrossRef]
- Kalivas, P.W.; Volkow, N.D. The neural basis of addiction: A pathology of motivation and choice. *Am. J. Psychiatry* 2005, 162, 1403–1413. [CrossRef] [PubMed]
- 23. Koob, G.F.; Volkow, N.D. Neurocircuitry of addiction. Neuropsychopharmacology 2010, 35, 217–238. [CrossRef] [PubMed]
- 24. Wise, R.A. Brain reward circuitry: Insights from unsensed incentives. Neuron 2002, 36, 229–240. [CrossRef]
- 25. Koob, G.F.; Arends, M.A.; Le Moal, M. Drugs, Addiction, and the Brain; Elsevier: Amsterdam, The Netherlands, 2014.
- 26. Di Chiara, G.; Imperato, A. Drugs abused by humans preferentially increase synaptic dopamine concentrations in the mesolimbic system of freely moving rats. *Proc. Natl. Acad. Sci. USA* **1988**, *85*, 5274–5278. [CrossRef] [PubMed]
- 27. Pierce, R.C.; Kumaresan, V. The mesolimbic dopamine system: The final common pathway for the reinforcing effect of drugs of abuse? *Neurosci. Biobehav. Rev.* 2006, *30*, 215–238. [CrossRef]
- Aguayo, L.G. Ethanol potentiates the GABAA-activated Cl⁻ current in mouse hippocampal and cortical neurons. *Eur. J. Pharmacol.* 1990, 187, 127–130. [CrossRef]
- 29. Lovinger, D.M.; White, G.; Weight, F.F. Ethanol inhibits NMDA-activated ion current in hippocampal neurons. *Science* **1989**, 243, 1721–1724. [CrossRef]
- 30. Weiner, J.L.; Gu, C.; Dunwiddie, T.V. Differential ethanol sensitivity of subpopulations of GABAA synapses onto rat hippocampal CA1 pyramidal neurons. *J. Neurophysiol.* **1997**, *77*, 1306–1312. [CrossRef]
- 31. Larsson, A.; Edstrom, L.; Svensson, L.; Soderpalm, B.; Engel, J.A. Voluntary ethanol intake increases extracellular acetylcholine levels in the ventral tegmental area in the rat. *Alcohol Alcohol.* **2005**, *40*, 349–358. [CrossRef]
- 32. Clarke, P.B.; Pert, A. Autoradiographic evidence for nicotine receptors on nigrostriatal and mesolimbic dopaminergic neurons. *Brain Res.* **1985**, *348*, 355–358. [CrossRef]
- Benwell, M.E.; Balfour, D.J. The effects of acute and repeated nicotine treatment on nucleus accumbens dopamine and locomotor activity. Br. J. Pharmacol. 1992, 105, 84956. [CrossRef] [PubMed]
- 34. Narahashi, T.; Aistrup, G.L.; Marszalec, W.; Nagata, K. Neuronal nicotinic acetylcholine receptors: A new target site of ethanol. *Neurochem. Int.* **1999**, *35*, 131–141. [CrossRef]
- Gotti, C.; Zoli, M.; Clementi, F. Brain nicotinic acetylcholine receptors: Native subtypes and their relevance. *Trends Pharmacol. Sci.* 2006, 27, 482–491. [CrossRef] [PubMed]
- Albuquerque, E.X.; Pereira, E.F.R.; Alkondon, M.; Rogers, S.W. Mammalian nicotinic acetylcholine receptors: From structure to function. *Physiol. Rev.* 2009, *89*, 73–120. [CrossRef] [PubMed]
- 37. Joslyn, G.; Brush, G.; Robertson, M.; Smith, T.L.; Kalmijn, J.; Schuckit, M.; White, R.L. Chromosome 15q25.1 genetic markers associated with level of response to alcohol in humans. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 20368–20373. [CrossRef]
- Saccone, N.L.; Wang, J.C.; Breslau, N.; Johnson, E.O.; Hatsukami, D.; Saccone, S.F.; Gruzca, R.A.; Sun, L.; Duan, W.; Budde, J.; et al. The CHRNA5-CHRNA3-CHRNB4 nicotinic receptor subunit gene cluster affects risk for nicotine dependence in african-americans and in european-americans. *Cancer Res.* 2009, 69, 6848–6856. [CrossRef] [PubMed]

- 39. Taslim, N.; Saeed Dar, M. The role of nicotinic acetylcholine receptor (nAChR) α7 subtype in the functional interaction between nicotine and ethanol in mouse cerebellum. *Alcohol. Clin. Exp. Res.* **2011**, *35*, 540–549. [CrossRef]
- Kamens, H.M.; Andersen, J.; Picciotto, M.R. Modulation of ethanol consumption by genetic and pharmacological manipulation of nicotinic acetylcholine receptors in mice. *Psychopharmacology* 2010, 208, 613–626. [CrossRef]
- Chatterjee, S.; Steensland, P.; Simms, J.A.; Holgate, J.; Coe, J.W.; Hurst, R.S.; Shaffer, C.L.; Lowe, J.; Rollema, H.; Bartlett, S.E. Partial agonists of the α3β4* neuronal nicotinic acetylcholine receptor reduce ethanol consumption and seeking in rats. *Neuropsychopharmacology* **2011**, *36*, 603–615. [CrossRef]
- 42. Miller, C.N.; Ruggery, C.; Kamens, H.M. The a3ß4 nicotinic acetylcholine receptor antagonist 18-Methoxycoronaridine decreases binge-like ethanol consumption in adult C57BL/6J mice. *Alcohol* **2019**, *79*, 1–6. [CrossRef]
- Santos, N.; Chatterjee, S.; Henry, A.; Holgate, J.; Bartlett, S.E. The α5 neuronal nicotinic acetylcholine receptor subunit plays an important role in the sedative effects of ethanol but does not modulate consumption in mice. *Alcohol. Clin. Exp. Res.* 2013, 37, 655–662. [CrossRef] [PubMed]
- 44. Xiao, C.; Cho, J.R.; Zhou, C.; Treweek, J.B.; Chan, K.; McKinney, S.L.; Yang, B.; Gradinaru, V. Cholinergic mesopontine signals govern locomotion and reward through dissociable midbrain pathways. *Neuron* **2016**, *90*, 333–347. [CrossRef] [PubMed]
- 45. Engel, J.A.; Jerlhag, E. Alcohol: Mechanisms along the mesolimbic dopamine system. *Prog. Brain Res.* 2014, 211, 201–233. [PubMed]
- 46. Silvestre, J.S.; O'Neill, M.F.; Fernandez, A.G.; Palacios, J.M. Effects of a range of dopamine receptor agonist and antagonist on ethanol intake in the rat. *Eur. J. Pharmacol.* **1996**, *318*, 257–265. [CrossRef]
- Mardones, J.; Quintanilla, M.E. Consumption of ethanol, water, and solid food by UChA and UChB rats. *Alcohol* 1996, 13, 355–357. [CrossRef]
- Hendrickson, L.M.; Guildford, M.J.; Tapper, A.R. Neuronal nicotinic acetylcholine receptors: Common molecular substrates of nicotine and alcohol dependence. *Front. Psychiatry* 2013, *4*, 29. [CrossRef]
- 49. Papke, R.L.; Heinemann, S.F. Partial agonist properties of cytisine on neuronal receptors containing the beta2 subunit. *Mol. Pharmacol.* **1993**, *45*, 142–149.
- 50. Rollema, H.; Shrikhande, A.; Ward, K.M.; Tingley, F.D.; Coe, J.W.; O'Neill, B.T.; Tseng, E.; Wang, E.Q.; Mather, R.J.; Hurst, R.S.; et al. Pre-clinical properties of the α4β2 nicotinic acetylcholine receptor partial agonists varenicline, cytisine and dianicline translate to clinical efficacy for nicotine dependence. *Br. J. Pharmacol.* 2010, *160*, 334–345. [CrossRef]
- 51. Rollema, H.; Chambers, L.K.; Coe, J.W.; Glowa, J.; Hurst, R.S.; Lebel, L.A.; Lu, Y.; Mansbach, R.J.; Mather, R.J.; Rovetti, C.C.; et al. Pharmacological profile of the α4β2 nicotinic acetylcholine receptor partial agonist varenicline, an effective smoking cessation aid. *Neuropharmacology* **2007**, *52*, 985–994. [CrossRef]
- 52. Coe, J.W.; Brooks, P.R.; Vetelino, M.G.; Wirtz, M.C.; Arnold, E.P.; Huang, J.; Sands, S.B.; Davis, T.I.; Lebel, L.A.; Fox, C.B.; et al. Varenicline: An alpha4beta2 nicotinic receptor partial agonist for smoking cessation. *J. Med. Chem.* **2005**, *48*, 3474–3477. [CrossRef]
- 53. Steensland, P.; Simms, J.A.; Holgate, J.; Richards, J.K.; Bartlett, S.E. Verenicline, an α3β4* nicotinic acetylcholine receptor partial agonist, selectively decreases ethanol consumption and seeking. *Proc. Natl. Acad. Sci. USA* 2007, 104, 12518–12523. [CrossRef] [PubMed]
- 54. Kamens, H.M.; Andersen, J.; Picciotto, M.R. The nicotinic acetylcholine receptor partial agonist varenicline increases the ataxic and sedative-hypnotic effects of acute ethanol administration in C57BL/6J mice. *Alcohol. Clin. Exp. Res.* 2010, *34*, 2053–2060. [CrossRef] [PubMed]
- 55. Sajja, R.K.; Rahman, S. Lobeline and cytisine reduce voluntary ethanol drinking behavior in male C57BL/6J mice. *Progr. Neuropsychopharmacol. Biol. Psycharity* 2011, 35, 257–264. [CrossRef] [PubMed]
- 56. Blomqvist, O.; Ericson, M.; Engel, J.A.; Soderpalm, B. Accumbal dopamine overflow after ethanol: Localization of the antagonizing effect of mecamylamine. *Eur. J. Pharmacol.* **1997**, *334*, 149–156. [CrossRef]
- 57. Ericson, M.; Blomqvist, O.; Engel, J.A.; Soderpalm, B. Voluntary ethanol intake in the rat and the associated accumbal dopamine overflow are blocked by ventral tegmental mecamylamine. *Eur. J. Pharmacol.* **1998**, 358, 189–196. [CrossRef]
- 58. Tizabi, Y.; Copeland, R.L., Jr.; Louis, V.A.; Taylor, R.E. Effects of combined systemic alcohol and central nicotine administration into ventral tegmental area on dopamine release in the nucleus accumbens. *Alcohol Clin. Exp. Res.* 2002, *26*, 394–399. [CrossRef]
- Faundez-Parraguez, M.; Farias-Rabelo, N.; Gonzalez-Gutierrez, J.P.; Etcheverry-Berrios, A.; Alzate-Morales, J.; Adasme-Carreño, F.; Varas, R.; Bermudez, I.; Iturriaga-Vasquez, P. Neonicotinic analogues: Selective antagonists for α4β2 nicotinic acetylcholine receptors. *Bioorg. Med. Chem.* 2013, 21, 2687–2694. [CrossRef]
- 60. Mardones, J.; Segovia-Riquelme, N. Thirty-two years of selection of rats by ethanol preference: UCHA and UCHB strains. *Neurobehav. Toxicol. Teratol.* **1983**, *5*, 171–178.
- 61. Quintanilla, M.E.; Israel, Y.; Sapag, A.; Tampier, L. The UChA and UChB rat lines: Metabolic and genetic differences influencing ethanol intake. *Addict. Biol.* 2006, *11*, 310–323. [CrossRef]
- 62. Kilkenny, C.; Browne, W.; Cuthill, I.C.; Emerson, M.; Altman, D.G. Animal research: Reporting in vivo experiments: The ARRIVE guidelines. *Br. J. Pharmacol.* 2010, 160, 1577–1579. [CrossRef]
- 63. Tampier, L.; Quintanilla, M.E. UChA and UChB rats: An animal model for the study of alcoholism. *Rev. Soc. Farmacol. Chile* **2010**, *3*, 5–11.