



Review

Non-Invasive Pulsatile Shear Stress Modifies Endothelial Activation; A Narrative Review

Jose A. Adams ^{1,*} , Arkady Uryash ¹ and Jose R. Lopez ²¹ Division of Neonatology, Mount Sinai Medical Center, Miami Beach, FL 33140, USA² Department of Research, Mount Sinai Medical Center, Miami Beach, FL 33140, USA

* Correspondence: tony@msmc.com

Abstract: The monolayer of cells that line both the heart and the entire vasculature is the endothelial cell (EC). These cells respond to external and internal signals, producing a wide array of primary or secondary messengers involved in coagulation, vascular tone, inflammation, and cell-to-cell signaling. Endothelial cell activation is the process by which EC changes from a quiescent cell phenotype, which maintains cellular integrity, antithrombotic, and anti-inflammatory properties, to a phenotype that is prothrombotic, pro-inflammatory, and permeable, in addition to repair and leukocyte trafficking at the site of injury or infection. Pathological activation of EC leads to increased vascular permeability, thrombosis, and an uncontrolled inflammatory response that leads to endothelial dysfunction. This pathological activation can be observed during ischemia reperfusion injury (IRI) and sepsis. Shear stress (SS) and pulsatile shear stress (PSS) are produced by mechanical frictional forces of blood flow and contraction of the heart, respectively, and are well-known mechanical signals that affect EC function, morphology, and gene expression. PSS promotes EC homeostasis and cardiovascular health. The archetype of inducing PSS is exercise (i.e., jogging, which introduces pulsations to the body as a function of the foot striking the pavement), or mechanical devices which induce external pulsations to the body (Enhanced External Pulsation (EEXP), Whole-body vibration (WBV), and Whole-body periodic acceleration (WBPA aka pGz)). The purpose of this narrative review is to focus on the aforementioned noninvasive methods to increase PSS, review how each of these modify specific diseases that have been shown to induce endothelial activation and microcirculatory dysfunction (Ischemia reperfusion injury-myocardial infarction and cardiac arrest and resuscitation), sepsis, and lipopolysaccharide-induced sepsis syndrome (LPS)), and review current evidence and insight into how each may modify endothelial activation and how these may be beneficial in the acute and chronic setting of endothelial activation and microvascular dysfunction.



Citation: Adams, J.A.; Uryash, A.; Lopez, J.R. Non-Invasive Pulsatile Shear Stress Modifies Endothelial Activation; A Narrative Review. *Biomedicines* **2022**, *10*, 3050. <https://doi.org/10.3390/biomedicines10123050>

Academic Editor: Virginia Tancredi

Received: 17 October 2022

Accepted: 21 November 2022

Published: 28 November 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Keywords: pulsatile shear stress; whole body periodic acceleration; exercise; enhanced external counterpulsation; whole body vibration; endothelial activation; nitric oxide; cytokines

1. Introduction

The luminal surface of all the vasculature and the heart is lined by endothelial cells (EC), encompassing more than 5000 m². Furthermore, the response of EC to external signals and the synthesis and production of various mediators is heterogeneous and adaptive based on location and signals [1–11]. EC membranes are the sensing mechanism, responsive to mechanical (shear stress) and biochemical signaling (chemosensor) [6,12]. EC output is important for blood fluidity, coagulation, vasoreactivity, vasculogenesis, barrier function, and inflammation [13]. Endothelial cell activation is the process by which EC changes from a quiescent cell phenotype, which maintains cellular integrity, antithrombotic, and anti-inflammatory properties, to a prothrombotic, pro-inflammatory, and permeable phenotype, also at the site of injury or infection, involved in repair and leukocyte trafficking. Endothelial activation is triggered by a multitude of stimuli that include inflammatory cytokines (interleukins, tumor necrosis factor, and interferon- γ), bacterial endotoxins, and

pattern recognition receptor activation (PRR) after recognition of pathogen-associated molecular patterns (PAMP) or damage-associated molecular patterns (DAMP) [14–16]. Pathological activation of EC leads to increased vascular permeability, thrombosis, and an uncontrolled inflammatory response leading to endothelial dysfunction; the latter can be contained at the local level or participate in a more profound systemic response leading to multiorgan dysfunction and death [14,17–22].

The sensing capabilities of EC include tangential, radial, axial, pulsatile, and oscillatory shear flow patterns [23,24]. Pulsatile shear stress (PSS) is a periodical laminar flow that occurs predominantly in the straight part of the blood vessel; it has a positive mean flow rate and fluid velocity that oscillates at the frequency of the heart rate [25–28]. PSS promotes a cytoprotective, non-inflammatory EC phenotype and vascular health [20,24], in contrast to oscillatory shear (mainly seen in arterial bifurcations and curvatures and where blood flow is uneven), which promotes an inflammatory and atherogenic phenotype [28–35].

The balance between vasoconstriction/relaxation is also governed by EC through vasoconstriction mediators (endothelin-1 and thromboxane-A₂) and vasodilators (nitric oxide (NO), prostacyclin, and endothelium-derived hyperpolarizing factor (EDHF)). Nitric oxide (NO) is a gas that is important not only for vasodilation, but also critical in signal transduction [36–39]. NO production occurs through the oxidation of L-arginine to L-citrulline using the enzyme nitric oxide synthase (NOS) and tetrahydrobiopterin (BH₄). There are three nitric oxide synthases: (a) endothelial nitric oxide synthase derived from the endothelium (eNOS also called NOS3) is constitutively expressed, calcium-dependent, and produces nanomolar amounts of NO and produced in response to PSS; (b) inducible nitric oxide synthase (iNOS also called NOS2) is not constitutively expressed, produces large quantities of NO, and is usually produced by macrophages and inflammatory cells; (c) neuronal nitric oxide synthase (nNOS also called NOS1) is found mostly in both neuronal and cardiovascular tissue and has a role in neuronal signal transduction and chronotropicity of the heart [37,40].

Experiments performed in the early 1990s by Hutchenson et al. found that NO is produced by EC as a function of pulsatility with an optimal frequency of pulsation of 2–8 Hz (120–480 cpm) [41]. PSS activates eNOS through phosphorylation, thus producing eNO, which is important for increasing blood flow and an important signaling molecule that down-regulates the inflammatory cascade and improves microvascular dysfunction [37,42–49]. eNO also serves to counterbalance signals that mediate endothelial activation [50].

The baseline pulsations in the human circulation are in the frequency range of 1 to 2 Hz and additional pulsations beyond these increase NO bioavailability via eNOS [41,51]. The concept of modulating the endothelium using noninvasive methods that induce PSS is a novel paradigm shift in thinking. In this narrative, we will focus on the modification of endothelial activation using noninvasive PSS, of which the archetype is exercise. Therefore, we review simple methods for inducing PSS under conditions known to produce endothelial cell activation.

This review will focus on four noninvasive and nonpharmacologic methods to increase PSS that have been shown to confer a protective role in ischemia reperfusion injury (IRI), sepsis or *Escherichia Coli* endotoxemia, lipopolysaccharide-induced sepsis-like syndrome (LPS), exercise (EXER), enhanced external counterpulsation (EECP), whole-body vibration (WBV), and whole-body periodic acceleration (WBPA, also known as, pGz) (Figure 1).

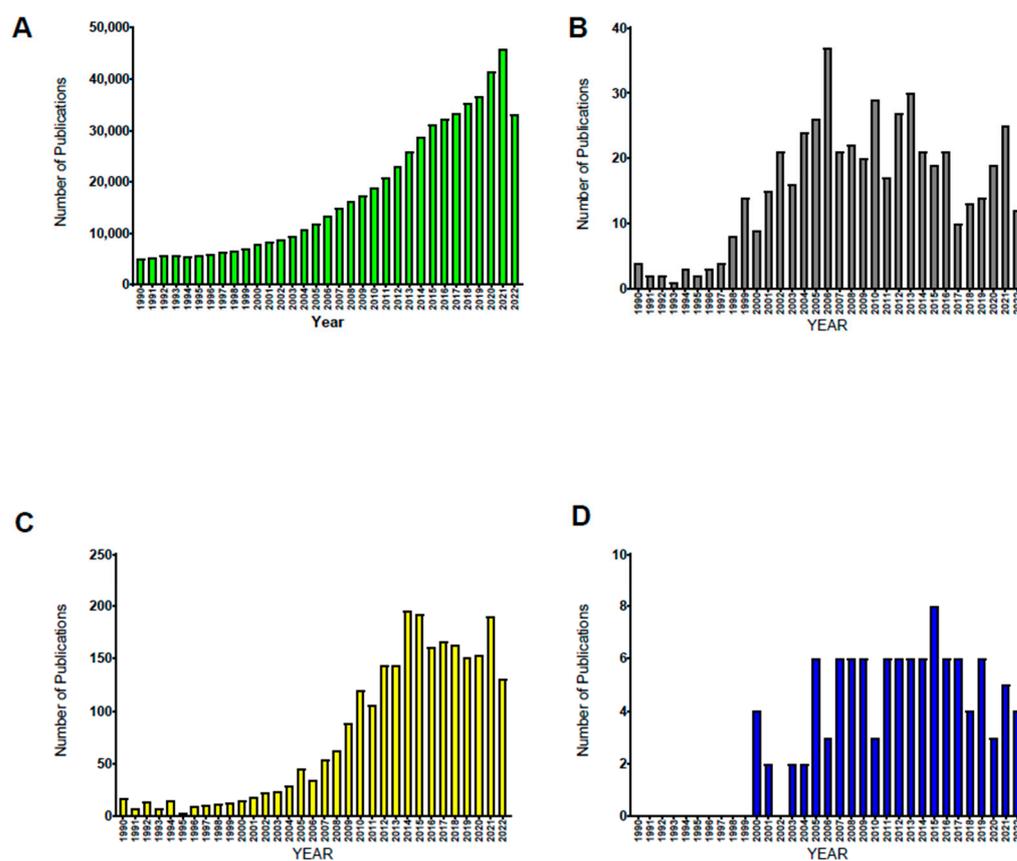


Figure 1. Publications in PUBMED from 1990 to 2022 for Four Methods of Noninvasive Pulsatile Shear Stress. Number of publications in PUBMED by year for the four methods described in this manuscript of noninvasive pulsatile shear stress: (A) Exercise, (B) Enhanced External Counterpulsation, (C) Whole-Body Vibration, (D) Whole-Body Periodic Acceleration (also known as pGz).

2. Methods

A review of the literature on endothelial activation was carried out using the following databases: EMBASE, PUBMED, SCOPUS. Each database was searched for the following key words [Endothelial activation] or [endothelium] and one of the three modalities of inducing noninvasive PSS: Exercise, WBV, WBPA, or pGz. The latter combination was combined with either; (a) sepsis, infection or inflammation, or lipopolysaccharide (LPS), or (b) ischemia reperfusion or myocardial infarction or cardiac arrest or resuscitation or post-resuscitation injury (Supplementary Material File contains search strategy and the number of articles). The studies were limited to those in English. The titles and abstracts were reviewed for relevant information on the various interventions of pulsatile shear stress in relation to disease models. Search dates: January 1990 to September 2022.

3. Models of Endothelial Activation

3.1. Ischemia Reperfusion Injury (IRI)-Cardiac Arrest (CA) and Myocardial Infarction (MI)

Estimates indicate that in the United States there are more than 500,000 cases annually of outpatient and in-hospital cardiac arrest, with a return to spontaneous circulation (ROSC) of 40–50% and a survival rate to hospital discharge of 10.5% and 26.7% for outpatient and in-hospital cardiac arrest, respectively [52]. Postcardiac arrest syndrome (PCAS) is characterized by reperfusion injury from systemic ischemia, myocardial dysfunction, brain, and other vital organ injury, superimposed on underlying diseases, all of which explain the low survival rate of hospital discharge. After CA, a systemic inflammatory response occurs that has been shown to occur in the reperfusion stage, with the release of pro-inflammatory cytokines by leukocytes and endothelial cells through the activation of leukocytes and endothelial cells and the release of secondary cytokines [53–60]. EC

expresses a wide spectrum of cytokines and chemokines, including pro-inflammatory interleukins; IL-1 β , IL-3, IL-5, IL-6, IL-8, IL-11, IL-15, and tumor necrosis factor (TNF- α), as well as anti-inflammatory cytokines such as IL-1 receptor antagonist (IL-1ra), IL-10, IL-13, and transforming growth factor beta (TGF- β) [61–63].

Recent data in mice show that myocardial infarction (MI) produces remote global endothelium activation, with up-regulation of the vascular cell adhesion molecule (VCAM-1), the cell adhesion molecule (P-Selectin) and platelet adhesion in remote arterial and microvascular beds, which persists for longer periods in animals with preexisting atherosclerosis [64,65]. Several reviews have outlined the role of NO in the cardiovascular system [47,48]. CA and resuscitation are models of total-body IRI, while MI with and without reperfusion are models of focal injury. These two models offer the opportunity to study what has been termed “sterile” endothelial activation [66].

3.2. Sepsis and Lipopolysaccharide-Induced Sepsis Syndrome (LPS)

In 2017, the estimated global burden of sepsis was 48.9 million people worldwide, with a mortality of 11 million. In the United States, for example, sepsis is the most common cause of in-hospital deaths and costs more than USD 2 billion annually [67]. Sepsis is a dysregulated response to infection that triggers a complex set of pathways and a cellular response that includes endothelial activation, macro, and micro circulatory failure, ultimately leading to organ failure and death. The systemic response may be triggered by the recognition of a pathogen (Pathogen Associated molecular pattern, PAMP) and or cellular injury proteins (damage associated molecular pattern, DAMP) recognition. The initial response predominantly by immune cells is the release of cytokines, interleukins, chemokines, interferons, tumor necrosis factor (TNF- α), and growth factors. The initiation of the inflammatory cascade is orchestrated by nuclear factor kappa beta (NF κ - β). The target of cytokines is the EC; however, these are also capable of secreting cytokines. During sepsis, the role of EC is to amplify the immune response and activate the coagulation system, with endothelial activation ultimately contributing to end organ damage and micro-circulatory failure [66,68,69]. In endotoxin (*Escherichia Coli*) -induced lipopolysaccharide sepsis-like syndrome (LPS), bacterial cell wall products ultimately bind to the toll-like receptor-4 (TLR-4) on the endothelial cell wall, to induce the intracellular response of EC of cytokines, adhesion molecules, and reactive oxygen species (ROS), and similarly amplify the immune response. Elevated biomarkers of endothelial activation/dysfunction in the systemic inflammatory response of critical illnesses and sepsis have been shown to be associated with a higher risk of developing respiratory failure, multiple-organ dysfunction, and death [18,70–72]. The decrease in NO bioavailability also plays a role in sepsis. Systemic NO release during sepsis has been shown and later thought to be responsible for hemodynamic and vascular instability, prompting the use of a non-selective inhibitor of NO (L-NAME) as a clinical therapeutic intervention in sepsis, which failed [73,74]. In animal models, an increase in NO bioavailability using NO donors or arginine administration, or a decrease in asymmetric dimethylarginine (ADMA, an endogenous inhibitor of nitric oxide synthase) appears to show some promise [75]. It is important to note that the reduced bioavailability of NO at the microvascular level comes primarily from the reduction in eNOS produced by the EC and that produces small amounts of NO, in contrast to iNOS which produces large amounts of NO, primarily by neutrophils and macrophages. NO derived from eNOS has been shown to be protective in sepsis [76,77]. Thus, increasing the bioavailability of NO through eNOS would provide a new avenue of therapeutics [46,48,78].

4. Exercise

4.1. Exercise for Pulsatile Shear Stress

Exercise is defined as “a subset of physical activity that is planned, structured, and repetitive and has as a final or intermediate objective the improvement or maintenance of physical fitness” [79,80]. For this narrative review, the exercise strategies considered involve walking, jogging, or running. PSS and/or circumferential wall stress or stretch

(which arises from the effect of blood pressure on the vascular wall and is applied to all layers of the arterial wall) are the primary signals produced by EXER that are mechanically transduced by the EC [8,29,34,81–86].

During walking, jogging, or running, pulses are added to the circulation as a result of the frequency of cadence of the foot. The frequency of steps for both men and women who are recreational runners has been estimated to be between 163 and 169 steps per min [87]. This frequency, added to a baseline pulsatility of 60–100 beats/min, generates total pulsation close to 220–240 beats per min (3–4 Hz) during running. Since running is not timed for the cardiac cycle, the expected pulsatility frequencies are in the range of 1.3 to 4 Hz.

This narrative review will not discuss the various exercise strategies or mechanisms of exercise-induced cardioprotection that others have thoroughly reviewed [88–98]. Exercise reduces cardiovascular morbidity and mortality and is positively correlated with beneficial health outcomes but requires subject cooperation and thus may prevent patients from participating and remaining in an exercise program, particularly those in an intensive care setting or those with physical and cognitive limitations.

4.2. Exercise (EXER) and Ischemia Reperfusion Injury

Exercise is a well-known cardioprotective strategy. Physical activity, exercise, and a healthy diet are the pillars of cardiovascular health [99]. Exercise induces a variety of cardioprotective signals, including a decrease in the inflammatory phenotype [98]. Regular exercise induces interleukin-6 (IL-6) produced by muscle fibers, which stimulates anti-inflammatory cytokines (IL-1ra and IL-10) and inhibits tumor necrosis alpha (TNF- α) [100]. In a recent systematic review and meta-analysis, regular exercise decreased aging-induced inflammasome activation related to inflammatory cytokines (IL-1 β and IL-18) [101]. In exercised (voluntary free-wheel running) mice fed a high-fat diet, exercise suppressed the pyrin domain of the NOD-like receptor family containing the 3 (NLRP3) inflammasome, improved nitric oxide production, and reduced oxidative stress [102].

Two specific periods have been explored concerning the role of EXER in cardiovascular protection against IRI or MI: EXER as a preconditioning (pretreatment) strategy (EXER performed prior to the onset of IRI) and a postcondition (post-treatment) strategy (EXER, performed after IRI). Both strategies aim to induce cardioprotection through various pathways, which ultimately increase myocardial tolerance, reduce the size of the infarct, and IRI-induced arrhythmias. The effects of exercise preconditioning against IRI have been well-established in animal models and human epidemiological studies summarized by Borges et al. [103]. Additionally, the beneficial effects of exercise after MI have also been well established, with exercise being an important component of the post-MI rehabilitation program [104]. However, a link between cardioprotection induced by exercise and decreased endothelial activation remains to be established. Exercise has been shown to increase the bioavailability of NO, specifically eNO and IL-6, both of which play an anti-inflammatory role [105–107]. It is important to note that excessive, prolonged, and strenuous overtraining can lead to damaging oxidative stress, with an attendant decrease in NO bioavailability. The concept of redox and exercise-induced hormesis has been advanced and previously reviewed; therefore, it appears that too much of a good thing may not necessarily be effective when it comes to EXER [80,108–112].

Data on the beneficial effects of EXER as a pre- or post-conditioning strategy for cardiac arrest and resuscitation are scarce. There is little doubt about the cardioprotective role of EXER and physical activity, on overall cardiovascular health suggesting a beneficial effect of EXER, and the benefits of post-MI EXER rehabilitation [113–115]. Recent data from the Korean National Outpatient CA registry showed that patients with higher intensity physical activity during exercise before and index CA had better survival outcomes and a successful percutaneous coronary intervention [116] suggesting a protective role for EXER specifically in CA.

4.3. Exercise and Sepsis

The effects of exercise on survival in animal models of sepsis and LPS have been well-documented, showing a favorable survival response to various EXER interventions [117–124].

Gholamnezhad et al. recently performed a systematic review of the modulatory effects of EXER on LPS-induced lung inflammation. The results showed that aerobic exercise (prior to LPS) in rodents reduced LPS-induced oxidative stress, inflammation, protein leakage, levels of IL-6, IL-1 β , IL-17, TNF- α , granulocyte–macrophage colony stimulating factor, and improved IL-10 and IL-1Ra, and a change in the balance between pro-inflammatory and anti-inflammatory phenotype, thus supporting the role of exercise in LPS-induced lung injury [125].

In human studies, the effects of exercise on sepsis have also been reported in a limited number of studies. Low rates of physical exercise and high rates of watching television (physical inactivity) are associated with higher morbidity and mortality from community-acquired sepsis [109], and physical rehabilitation in septic patients was shown to improve physical function and reduce the inflammatory response [126,127]. A review on the effects of exercise in the treatment of sepsis in animal models and patients has recently been published [128].

5. Enhanced External Counterpulsation

5.1. Enhance External Counterpulsation (EECP) for Pulsatile Shear Stress

Enhanced external counterpulsation (EECP) involves compression of the legs to buttocks using pneumatic cuffs, timed to early diastole [129,130]. EECP induces pulsations and imparts a circumferential stretch that doubles the heart rate (2–3.3 Hz). The beneficial clinical effects of EECP have been reported for angina, peripheral artery disease, diabetes, erectile dysfunction, and possibly Alzheimer’s disease. Similarly, to exercise, EECP via PSS induces NO production [131], improves endothelial function [132,133], and attenuates pro-inflammatory signaling pathways [134,135]. The risks and guidelines of EECP have been published by Lin et al. [136].

5.2. Enhanced External Counterpulsation (EECP) and Ischemia Reperfusion Injury

The use of EECP on acute and chronic MI-induced IRI has been shown in both animal models and human studies and has been reviewed [129], and its use as a myocardial conditioning strategy has also been reviewed [137].

In nonischemic hypercholesterolemic patients, seven weeks of EECP was compared with the control. The EECP group had a significant reduction in atherosclerosis lesion, and a reduction in C-reactive protein, vascular cell adhesion molecule-1 (VCAM-1), iNOS, mitogen-activated protein kinase phosphorylation (MAPK-p38), and activation of NF κ - β [134].

In a dog model of MI by coronary artery occlusion, EECP use significantly improved myocardial perfusion and function after 4 and 6 weeks of EECP compared to controls, along with increased expression of vascular endothelial growth factor (VEGF) and increased microvascular density [138]. EECP has also been studied in a dog model of CA. Post-CA EECP (4 h of use after CA) increased cerebral blood flow, improved microcirculation recovery, and improved neurological outcomes from 24 to 96 h compared to control animals [139]. In a similar dog model of CA with also 4 h of EECP post-CA, others have also shown improved survival and myocardial function [140].

Casey et al. and Braith et al. studied the effects of EECP in patients with chronic angina and symptomatic coronary artery disease, respectively. Both investigators showed a significant reduction in TNF- α , monocyte chemoattractant protein-1 (MCP-1), and soluble vascular adhesion molecule (sVCAM-1) compared to controls [133,141]. Yang et al. have summarized the additional benefits of EECP beyond hemodynamics [142].

5.3. Enhanced External Counter Pulsation and Sepsis

The effects of EECP on sepsis or LPS animal models have not been published. A single case study has documented the use of EECP in a female patient diagnosed with

coronavirus disease 2019 (COVID-19) and treated at home with a cocktail of vitamins and hydroxychloroquine for 6 days. Most of her symptoms resolved but remained with fatigue, headaches, and shortness of breath at rest and during activities, and “brain fog” (subjective lack of clarity) for months. She was treated with 35 sessions of EECF, and the aforementioned symptoms resolved. The resolution of symptoms was subjectively measured, and this resolution was attributed to the use of EECF [143].

6. Whole-Body Vibration

6.1. Whole-Body Vibration (WBV) for Pulsatile Shear Stress

The effects of vibration on the entire body were first described in the 1900s and began to appear in the scientific literature in the 1960s describing the effects of whole-body vibration (WBV) on ventilation, behaviors, and central hemodynamics [144–147].

The mechanical oscillations imparted by WBV are performed using a platform, moving in a linear or pivotal motion with a standing or seated subject at frequencies from 12–60 Hz and displacements from 1 to 10 mm producing accelerations of $+1.5 \text{ m/s}^2$ [148–152]. Most studies exploring the effects of WBV use a structured exercise program performed on the WBV platform. WBV has been shown to increase skin blood flow [153–155], improve endothelial function in an elderly population [156], and its effect is summarized by others [157–159].

6.2. Whole-Body Vibration and Ischemia Reperfusion Injury

WBV has been used as a pretreatment strategy in a rodent model of acute MI without reperfusion. WBV was carried out for 1 and 3 weeks (30 min per day for 6 days) versus a control group. Myocardial infarct size and severity of ventricular fibrillation were significantly lower in WBV at 1 and 3 weeks [160]. WBV has also been used as an adjunct to a cardiac rehabilitation program over a 24-days, both standard exercise rehabilitation and those who received adjunct WBV had improvement in exercise tolerance and left ventricular ejection fraction, and both groups obtained similar effects [161]. There are no published results on the use of WBV post CA, likely due to the need for subject cooperation and the critical nature of these patients.

6.3. Whole-Body Vibration and Sepsis

Similarly, to EXER, data on WBV applied in the setting of sepsis or LPS are scarce. A single study explored the effects of WBV on LPS-induced inflammatory bone loss. The report focused on trabecular bone loss, which decreased with WBV. Furthermore, in the same study using in vitro stimulation of human mesenchymal stromal cells with WBV, WBV reduced the increase in LPS-induced IL-1 β and TNF- α induced by LPS [162]. Two recent reviews have addressed the potential for WBV to improve acute and long-term clinical conditions associated with COVID-19 and provide a framework for the use of WBV in the acute care setting [163,164].

Sanni et al. investigated the acute effects of WBV (HI = 88.7 ms^2 and LO = 44.4 ms^2 , both at 30 Hz) in healthy volunteers. They report a higher muscle oxygen consumption in the LO compared to the HI group and an increase in IL-6 in both groups but a higher in the LO [165]. Rodriguez-Miguel studied healthy elderly volunteers (70 years) who performed an 8-week training protocol on WBV. WBV produced a significant decrease in TNF- α , and an increase in IL-10, and a significant decrease in the mediator myeloid differentiation response gene 88 (MyDD88, an essential protein in the production of inflammatory cytokines) and transcription factor p65 (also known as the nuclear factor NF- β p65 subunit) [166]. In contrast, Jawed et al. showed in healthy male volunteers trained on WBV (35 Hz, eight 60 s sets, with 2 min between sets) an increase in IL-10 and an increase in TNF- α [167]. Similarly, Neves et al., in adult patients with chronic obstructive pulmonary disease (45–80 years) enrolled in a 12-week WBV protocol, and Cristi et al., in a 9-week WBV protocol in elderly volunteers (80 years), failed to show changes in IL-6 or soluble receptors of TNF, TNF- α ,

IL-10, and IL-1 β [168,169]. Therefore, the effects of WBV on the parameters of inflammation and endothelial activation markers remain inconsistent.

7. Whole-Body Periodic Acceleration (WBPA aka pGz)

7.1. Whole-Body Periodic Acceleration (WBPA, aka pGz) and Pulsatile Shear Stress

WBPA is the sinusoidal motion of the body on a platform in a headward to footward direction, which via inertial forces introduces small pulsations to the vasculature inducing PSS. [170,171]. The pulsations produced by WBPA are not synchronized with the cardiac cycle. The WBPA motion frequency in humans is between 100 and 150 cycles per min (1.6 to 2.5 Hz) with acceleration forces in the z plane $Gz \pm 0.3 \text{ mt/sec}^2$. A more detailed description of the platform has previously been reported [147,170–172]

WBPA has been shown to produce endothelial-derived NO release in human subjects [170,173,174], and animal models [171,175,176] and genomic upregulation of eNOS occurring over a relatively short time period [175,177], along with increased expression of antioxidant capacity [177], and endothelial-derived anticoagulant, vasoactive proteins, and adrenomedullin [172,178].

7.2. Whole-Body Periodic Acceleration (WBPA, aka pGz) and Ischemia Reperfusion Injury

The use of WBPA in CA models was first evaluated nearly 20 years ago, when laboratory observations on its use as a cardiopulmonary resuscitation (CPR) method were made [179]. In an adult porcine model of CA (15 min of ventricular fibrillation), WBPA used as a CPR method allowed for successful resuscitation and return of circulation in 100% of animals with normal neurological evaluation 24 h later [179]. This observation was also replicated in a newborn porcine model of CA induced by asphyxia [180]. The use of WBPA as a CPR method can be considered a method of preconditioning (or conditioning), which is an intervention performed during the ischemic period designed to improve injury produced by ischemia and reperfusion [137,181].

WBPA used before CA (preconditioning) has also been used in CA porcine models. In addition to a decrease in post-resuscitation myocardial stunning and improved regional blood flow to vital organs, WBPA increases eNOS expression in the myocardium and decreases tissue damage [182]. WBPA used as a preconditioning strategy in a rat model of MI improved survival, decreased infarct size, and increased eNOS expression [183].

WBPA has also been used after CA (postconditioning) in the same porcine model of ventricular fibrillation of CA, showing decreased post-arrest myocardial stunning, improved regional microvascular blood flow to vital organs, and increased expression of eNOS [184]. Chronic use of WBPA (1 h daily for 4 weeks) has also been used in a rat MI survival model as a chronic postconditioning strategy. WBPA improved survival and myocardial contractility and decreased myocardial fibrosis compared to controls. Furthermore, WBPA restored the expression of eNO, decreased the expression of IL-6 and TNF- α , and increased the expression of IL-10 [185]. The role of endothelium and nitric oxide synthases in CA and resuscitation was previously reviewed, showing the importance of eNO [169]. Together, these studies suggest that PSS induced by WBPA is cardioprotective, reduces the inflammatory phenotype induced by CA and focal MI, and improves microvascular flow to vital organs.

Currently, there are no human studies on the use of WBPA in patients with CA. In patients with angina, who are not candidates for percutaneous coronary intervention and/or coronary artery bypass surgery, WBPA used for 20 days improved exercise capacity and improved myocardial perfusion and function [186]. In patients with severe leg ischemia, WBPA performed for 10 days significantly increased laser Doppler blood flow to the ischemic limb, and these findings were also replicated in a mouse model of severe leg ischemia, which also confirmed upregulation of eNOS and VEGF expression [187].

7.3. The Effects of WBPA/pGz on Sepsis

WBPA was used as a pre- and post-treatment strategy in a murine lethal dose endotoxemia (LPS) model. Both WBPA strategies have been shown to increase survival, decrease microvascular leakage, and restore the expression of the tunica interna endothelial cell kinase-2 (TIE2) receptor enriched with tyrosine kinase and its phosphorylation (important for maintaining tight junctions of the endothelium and decreasing vascular permeability) [188]. Furthermore, WBPA restored the expression of eNOS and decreased the proinflammatory cytokines; TNF- α , NF κ - β p65, IL-1 β , and IL-6, and increased the anti-inflammatory cytokine Il-10 [189]. These studies suggest that WBPA through PSS has a protective effect (preconditioning and postconditioning) effect on LPS-induced endothelial activation and improves the pro-inflammatory environment induced by LPS. The effect of WBPA on human subjects with critical illness or sepsis has not been explored. It should be noted that PSS (due to the antiviral properties of NO) has been theorized to potentially be therapeutic adjuncts in the fight against COVID-19 [190].

The WBPA human platform is large and heavy; therefore, a smaller and simpler device (a predicate device for WBPA) was designed. The details of this passive simulated jogging device (JD, (Gentle Jogger, Movewell Technologies LLC, Hollywood, Florida, USA)) have been previously published [191–194]. The JD passively moves the feet using a motor, in alternating motion simulating walking or jogging, and with each downward stroke of the fore foot, a pulsation is added to the circulation, thus inducing PSS. This device has been shown to increase NO bioavailability in humans [195].

The characteristics of the four described interventions to produce PSS have been previously summarized [147] (Figure 2). These vary with respect to frequency, accelerations, and known effects on endothelial activation. All have been shown to increase the bioavailability of eNOS and NO, improve endothelial function, and induce an anti-inflammatory and fibrinolytic effect. They are also varied in their subject–device interphase and portability, and thus the limitation of usage particularly in critically ill patients, and those with physical or cognitive limitations.

	 Exercise	 EECP	 WBV	 WBPA/pGz
IRI	<ul style="list-style-type: none"> ↑ IL-6, IL-1ra, IL-10, Inhibits TNF-α (Peterson2005) ↓ IL-1β, IL-18 (Ding and Xu 2021) ↓ NLRP3 (Lee,Hong et al 2020) <p>Pre-Treatment of MI – A & H Beneficial (Borges and da Silva Verdoorn 2017)</p> <p>Post-Treatment of MI –H Beneficial (Borges and Lessa 2015, Fegers-Wustrow et al 2022)</p>	<ul style="list-style-type: none"> ↓ CRP, VCAM-1, iNOS, MAPK-p38 and NFκ-β (Zhang, He et al 2010) <p>Post- Treatment of MI-A ↑ VEGF ↑ myocardial perfusion, function, and microvascular density (Wu,Du et al 2006)</p> <p>Post- Treatment of MI-H ↓ TNF-α, MCP-1 sVCAM-1 (Casey,Conti et al2008,Brath Conti et al 2010) Improved Angina, ↑ LVEF,Myocardial Perfusion, Intracoronary Flow(Raza, Steinberg et al 2017)</p>	<ul style="list-style-type: none"> ↓ TNF-α, myDD88, NFκ-β-p65, ↑ IL-10 (Rodriguez-Miguelez,Fernandez-Gonzalo et al 2015) <p>Pre- Treatment of MI-A ↓ Infarct Size, and severity of VF (Shekarforoush and Noghri 2019)</p> <p>Post-Treatment of MI-H ↑ Exercise tolerance, EF (Nowak-Lis, Nowak et al 2022)</p>	<ul style="list-style-type: none"> ↑ eNOS, SOD, Adrenomedullin(Adams, bassuk 2005,Martinez, Arias et al 2008, Uryash et al 2012) <p>Pre-Treatment of MI-A ↓ Infarct Size, and severity of VF ↑ Survival and ↓ arrhythmias ↑ eNOS (Uryash, Bassuk et al 2015, Adams, Uryash et al 2011)</p> <p>Post-Treatment of MI-A ↑ Survival, and eNOS ↓ IL-6, TNF-α ↑ IL-10(Uryash, Bassuk et al 2015)</p> <p>Post-Treatment of MI-H ↑ Myocardial Perfusion and Function(Myamoto, Fujita et al 2011)</p>
CA	<p>Pre-Treatment of CA-H Better Survival Outcomes (Kim,Park et al 2022)</p>	<p>Post-Treatment of CA-A Improved Neurological Outcome and microcirculation(Hu,Liu et al 2013)</p> <p>↑ Survival, myocardial function(Xiang, Zhang et al 2020)</p>	<p>No Data on CA</p>	<p>Treatment of CA-A Improved Survival , Neurological Outcome and microcirculation(Adams, Bassuk et al 2003)</p> <p>Pre-Treatment of CA-A ↑ Survival, regional microvascular flow improved myocardial function, ↑ eNOS (Adams , Wu et al 2010)</p> <p>Post-Treatment of CA-A ↑ microvascular flow, cardiac function and eNOS (Adams, Uryash et al 2013)</p>
Sepsis LPS	<p>Pre-Treatment-LPS-A ↑ Survival ↓ Oxidative Stress and Protein leak (de Araujo,Silva et al 2012,Ihara,Sato et al 2016,Kimand Kang,2019) ↓ IL-1β, IL-6,IL-17,TNFα ↑ IL-10, IL1-ra(Gholamzhad,Safarian et al 2022)</p> <p>Post- Treatment of Sepsis-H Improves Physical Function(Ahn, Song et al 2018) ↓ Inflammatory Response (Wu,Guo et al 2022)</p>	<p>Treatment-H ↓ “brain fog” after COVID-19 (Doayrik, Verdugo-Gutierrez et al 2022)</p>	<p>Treatment-LPS- Cells ↓ IL-β, TNF-α (Kim, Lee et al 2014)</p>	<p>Pre-Treatment & Post-Treatment-LPS- A ↑ Survival ↑ TIE2 ↓ Microvascular leak ↑ eNOS and IL-10 ↓ TNF-α,NF$\kappa$$\beta$-p65,IL-6 (Adams Uryash et al 2019, Adams Lopez et al 2021)</p>

Figure 2. Summary of the Effects of Exercise, Enhanced External Counterpulsation (EECP), Whole-Body Vibration (WBV), and Whole-Body Periodic Acceleration (WBPA also known as pGz) on Ischemia Reperfusion Injury (IRI), Cardiac Arrest (CA), and E. Coli endotoxin induced Sepsis-like syndrome (LPS).

Summary table of the effects of four noninvasive methods to induce pulsatile shear stress (PSS), and conditions, that increase endothelial activation and decrease microvascular flow; Ischemia Reperfusion Injury (IRI), cardiac arrest (CA), and sepsis/E. Coli endotoxin induced sepsis like syndrome (LPS). For each intervention, a summary of the changes is provided, when the method to induce PSS is performed as a pretreatment (preconditioning) or posttreatment (post-conditioning) strategy, in **A** (Animals), **H** (Humans). Abbreviations: \uparrow = increase, \downarrow = decrease, MI = Myocardial Infarction, VEGF = vascular endothelial growth factor, TNF- α = tumor necrosis factor alpha, IL-1 β = interleukin1-beta, IL-6 = interleukin 6, IL-17 = interleukin 17, IL-10 = interleukin 10, IL1-ra = interleukin 1 receptor antagonist, MCP-1 = Monocyte Chemoattractant Protein-1, sVCAM-1 = soluble Vascular Cell Adhesion Molecule-1, TIE2 = endothelial-specific receptor tyrosine kinase family, eNOS = endothelial derived nitric oxide synthase, NF κ B-p65 = nuclear factor kappa beta p65 subunit, VF = ventricular fibrillation, LVEF = left ventricular ejection fraction, EF = Ejection Fraction. [100–103,114–117,121,122,125,126,128,129,133,134,138–141,143,160–162,166,172,175,177–179,182–186,188,189].

8. Limitations

The review has focused on four of the most published methods for PSS; however, there are other methods. Vibroacoustic Therapy (VAT) is a non-invasive delivery of a sound frequency of 50–100 Hz and a sound pressure of 0.5–20 dyn/cm² by a sound speaker that delivers stimulation directed toward the vascular wall. Sound-distributed stimulation to the endothelial cell matrix can trigger mechanosensors in cells affected by PSS and has been shown to increase eNO, can induce the expression of the mechanosensory protein Syndecan-4 (Syn4) and VEGF, as well as increase blood flow and clot dissolution [196–200]. This method may also be a plausible therapeutic strategy for MI [201,202] and sepsis [203].

Additional methods of delivering PSS also include passive cycling or limb movement [204,205], as well as a recently published method of 160-degree V-shaped WBPA in supine posture [206]. The latter methods may also have potential for clinical therapeutics, but studies on their use in MI, CA, or sepsis are not available. The study of endothelial activation markers in the context of these PSS interventions is an open area of investigation that may provide fruitful areas of mechanistic research and therapeutic interventions in a variety of diseases.

9. Conclusions

The effects of PSS on the vascular endothelium provide a protective and potential therapeutic intervention for the management of diseases with acute or chronic endothelial activation, endothelial dysfunction, and reduced microvascular flow. This narrative review has shown that externally applied PSS, as provided by EXER, EECP, WBV, and WBPA/pGz, positively modifies endothelial activation, improves microvascular dysfunction, and can be a viable adjunct as a clinical intervention. More studies are needed to improve the subject device interface, provide guidance on the optimal duration of use of these devices, and elucidate additional mechanistic insights.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/biomedicines10123050/s1>, The supplemental file contains the schematic figures of literature search strategy. The search terms and number of citations ($n =$). Figure S1. Schematic of Exercise or Physical Activity Search Strategy. Figure S2. Schematic of Enhanced External Counterpulsation or External Counterpulsation or EECP Search Strategy. Figure S3. Schematic of Whole-Body Vibration or Whole-Body Vibration Therapy Search Strategy. Figure S4. Schematic of Whole-Body Periodic Acceleration or WBPA or pGz Search Strategy.

Author Contributions: J.A.A. conceived and wrote initial manuscript, A.U. performed laboratory experiments and edited manuscript, J.R.L. performed experiments and co-wrote manuscript. All authors have read and agreed to the published version of the manuscript.

Funding: Grant funding was received by J.A.A. from the Florida Heart Research Institute (Miami, FL, USA).

Data Availability Statement: Not applicable.

Conflicts of Interest: J.A.A. is a scientific advisor and draws no salary from Movewell Technologies L.L.C., a company that owns the passive jogging device patent. There are no conflicts of interest to declare for all other authors. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript; or in the decision to publish the results.

References

1. Aird, W.C. Endothelial cell heterogeneity. *Crit. Care Med.* **2003**, *31*, S221–S230. [[CrossRef](#)]
2. Davies, P.F. Flow-mediated endothelial mechanotransduction. *Physiol. Rev.* **1995**, *75*, 519–560. [[CrossRef](#)]
3. Ohura, N.; Yamamoto, K.; Ichioka, S.; Sokabe, T.; Nakatsuka, H.; Baba, A.; Shibata, M.; Nakatsuka, T.; Harii, K.; Wada, Y.; et al. Global Analysis of Shear Stress-Responsive Genes in Vascular Endothelial Cells. *J. Atheroscler. Thromb.* **2003**, *10*, 304–313. [[CrossRef](#)]
4. Bryan, M.T.; Duckles, H.; Feng, S.; Hsiao, S.T.; Kim, H.R.; Serbanovic-Canic, J.; Evans, P.C. Mechanoresponsive Networks Controlling Vascular Inflammation. *Arterioscler. Thromb. Vasc. Biol.* **2014**, *34*, 2199–2205. [[CrossRef](#)]
5. Firasat, S.; Hecker, M.; Binder, L.; Asif, A.R. Advances in endothelial shear stress proteomics. *Expert Rev. Proteomics* **2014**, *11*, 611–619. [[CrossRef](#)]
6. He, M.; Martin, M.; Marin, T.; Chen, Z.; Gongol, B. Endothelial mechanobiology. *APL Bioeng.* **2020**, *4*, 10904. [[CrossRef](#)] [[PubMed](#)]
7. Zhang, J.; Friedman, M.H. Adaptive response of vascular endothelial cells to an acute increase in shear stress magnitude. *Am. J. Physiol. Circ. Physiol.* **2012**, *302*, H983–H991. [[CrossRef](#)] [[PubMed](#)]
8. Zhang, J.; Friedman, M.H. Adaptive response of vascular endothelial cells to an acute increase in shear stress frequency. *Am. J. Physiol. Circ. Physiol.* **2013**, *305*, H894–H902. [[CrossRef](#)] [[PubMed](#)]
9. Himburg, H.A.; Dowd, S.E.; Friedman, M.H. Frequency-dependent response of the vascular endothelium to pulsatile shear stress. *Am. J. Physiol. Circ. Physiol.* **2007**, *293*, H645–H653. [[CrossRef](#)]
10. Resnick, N. Fluid shear stress and the vascular endothelium: For better and for worse. *Prog. Biophys. Mol. Biol.* **2003**, *81*, 177–199. [[CrossRef](#)]
11. Chien, S. Mechanotransduction and endothelial cell homeostasis: The wisdom of the cell. *Am. J. Physiol. Circ. Physiol.* **2007**, *292*, H1209–H1224. [[CrossRef](#)] [[PubMed](#)]
12. Campinho, P.; Vilfan, A.; Vermot, J. Blood Flow Forces in Shaping the Vascular System: A Focus on Endothelial Cell Behavior. *Front. Physiol.* **2020**, *11*, 552. [[CrossRef](#)]
13. Triggle, C.R.; Dinga, H.; Mareia, I.; Andersone, T.; Hollenber, M. Why the endothelium? The endothelium as a target to reduce diabetes-associated vascular disease. *Can. J. Physiol. Pharmacol.* **2020**, *12*, 638491. [[CrossRef](#)] [[PubMed](#)]
14. Theofilis, P.; Sigris, M.; Oikonomou, E.; Antonopoulos, A.; Siasos, G.; Tsioufis, C.; Tousoulis, D. Inflammatory Mechanisms Contributing to Endothelial Dysfunction. *Biomedicines* **2021**, *9*, 781. [[CrossRef](#)] [[PubMed](#)]
15. Gaudette, S.; Hughes, D.; Boller, M. The endothelial glycocalyx: Structure and function in health and critical illness. *J. Veter.-Emerg. Crit. Care* **2020**, *30*, 117–134. [[CrossRef](#)]
16. van Ierssel, S.H.; Jorens, P.G.; Van Craenenbroeck, E.M.; Conraads, V.M. The endothelium, a protagonist in the pathophysiology of critical illness: Focus on cellular markers. *BioMed Res. Int.* **2014**, *2014*, 985813. [[CrossRef](#)] [[PubMed](#)]
17. Lee, W.L.; Liles, W.C. Endothelial activation, dysfunction and permeability during severe infections. *Curr. Opin. Hematol.* **2011**, *18*, 191–196. [[CrossRef](#)]
18. Page, A.V.; Liles, W.C. Biomarkers of endothelial activation/dysfunction in infectious diseases. *Virulence* **2013**, *4*, 507–516. [[CrossRef](#)]
19. Neubauer, K.; Zieger, B. Endothelial cells and coagulation. *Cell Tissue Res.* **2022**, *387*, 391–398. [[CrossRef](#)]
20. Kruger-Genge, A.; Blocki, A.; Franke, R.P.; Jung, F. Vascular Endothelial Cell Biology: An Update. *Int. J. Mol. Sci.* **2019**, *20*, 4411. [[CrossRef](#)]
21. Pober, J.S.; Sessa, W.C. Evolving functions of endothelial cells in inflammation. *Nat. Rev. Immunol.* **2007**, *7*, 803–815. [[CrossRef](#)]
22. Jin, Y.; Ji, W.; Yang, H.; Chen, S.; Zhang, W.; Duan, G. Endothelial activation and dysfunction in COVID-19: From basic mechanisms to potential therapeutic approaches. *Signal Transduct. Target. Ther.* **2020**, *5*, 293. [[CrossRef](#)] [[PubMed](#)]
23. Roux, E.; Bougaran, P.; Dufourcq, P.; Couffignal, T. Fluid Shear Stress Sensing by the Endothelial Layer. *Front. Physiol.* **2020**, *11*, 861. [[CrossRef](#)] [[PubMed](#)]
24. Gordon, E.; Schimmel, L.; Frye, M. The Importance of Mechanical Forces for in vitro Endothelial Cell Biology. *Front. Physiol.* **2020**, *11*, 684. [[CrossRef](#)] [[PubMed](#)]
25. Chiu, J.-J.; Chien, S. Effects of Disturbed Flow on Vascular Endothelium: Pathophysiological Basis and Clinical Perspectives. *Physiol. Rev.* **2011**, *91*, 327–387. [[CrossRef](#)] [[PubMed](#)]
26. Busse, R.; Fleming, I. Pulsatile Stretch and Shear Stress: Physical Stimuli Determining the Production of Endothelium-Derived Relaxing Factors. *J. Vasc. Res.* **1998**, *35*, 73–84. [[CrossRef](#)]

27. Malek, A.M.; Alper, S.L.; Izumo, S. Hemodynamic Shear Stress and Its Role in Atherosclerosis. *JAMA* **1999**, *282*, 2035–2042. [[CrossRef](#)]
28. Zhou, J.; Li, Y.-S.; Chien, S. Shear Stress–Initiated Signaling and Its Regulation of Endothelial Function. *Arterioscler. Thromb. Vasc. Biol.* **2014**, *34*, 2191–2198. [[CrossRef](#)]
29. Laughlin, M.H.; Newcomer, S.C.; Bender, S.B. Importance of hemodynamic forces as signals for exercise-induced changes in endothelial cell phenotype. *J. Appl. Physiol.* **2008**, *104*, 588–600. [[CrossRef](#)]
30. Souilhol, C.; Serbanovic-Canic, J.; Fragiadaki, M.; Chico, T.J.; Ridger, V.; Roddie, H.; Evans, P.C. Endothelial responses to shear stress in atherosclerosis: A novel role for developmental genes. *Nat. Rev. Cardiol.* **2020**, *17*, 52–63. [[CrossRef](#)]
31. Ajami, N.E.; Gupta, S.; Maurya, M.R.; Nguyen, P.; Li, J.Y.-S.; Shyy, J.Y.-J.; Chen, Z.; Chien, S.; Subramaniam, S. Systems biology analysis of longitudinal functional response of endothelial cells to shear stress. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, 10990–10995. [[CrossRef](#)]
32. Kadohama, T.; Nishimura, K.; Hoshino, Y.; Sasajima, T.; Sumpio, B.E. Effects of different types of fluid shear stress on endothelial cell proliferation and survival. *J. Cell Physiol.* **2007**, *212*, 244–251. [[CrossRef](#)] [[PubMed](#)]
33. Qiu, Y.; Tarbell, J.M. Interaction between Wall Shear Stress and Circumferential Strain Affects Endothelial Cell Biochemical Production. *J. Vasc. Res.* **2000**, *37*, 147–157. [[CrossRef](#)] [[PubMed](#)]
34. Wang, Y.-X.; Liu, H.-B.; Li, P.-S.; Yuan, W.-X.; Liu, B.; Liu, S.-T.; Qin, K.-R. ROS and NO Dynamics in Endothelial Cells Exposed to Exercise-Induced Wall Shear Stress. *Cell Mol. Bioeng.* **2019**, *12*, 107–120. [[CrossRef](#)]
35. Sumpio, B.; Chin, J. Vessel Wall Biology. In *Rutherford’s Vascular Surgery and Endovascular Therapy*; Elsevier Inc.: Amsterdam, The Netherlands, 2019; pp. 30–43.
36. Duran, W.N.; Beuve, A.V.; Sanchez, F.A. Nitric oxide, S-Nitrosation, and endothelial permeability. *IUBMB Life* **2013**, *65*, 819–826. [[CrossRef](#)]
37. Förstermann, U.; Sessa, W.C. Nitric oxide synthases: Regulation and function. *Eur. Heart J.* **2012**, *33*, 829–837. [[CrossRef](#)]
38. Farah, C.; Michel, L.Y.M.; Balligand, J.-L. Nitric oxide signalling in cardiovascular health and disease. *Nat. Rev. Cardiol.* **2018**, *15*, 292–316. [[CrossRef](#)]
39. Ghimire, K.; Altmann, H.M.; Straub, A.C.; Isenberg, J.S. Nitric oxide: What’s new to NO? *Am. J. Physiol. Cell Physiol.* **2017**, *312*, C254–C262. [[CrossRef](#)]
40. Ziolo, M.T.; Kohr, M.J.; Wang, H. Nitric oxide signaling and the regulation of myocardial function. *J. Mol. Cell Cardiol.* **2008**, *45*, 625–632. [[CrossRef](#)]
41. Hutcheson, I.R.; Griffith, T.M. Release of endothelium-derived relaxing factor is modulated both by frequency and amplitude of pulsatile flow. *Am. J. Physiol. Circ. Physiol.* **1991**, *261*, H257–H262. [[CrossRef](#)]
42. Cirino, G.; Fiorucci, S.; Sessa, W.C. Endothelial nitric oxide synthase: The Cinderella of inflammation? *Trends Pharmacol. Sci.* **2003**, *24*, 91–95. [[CrossRef](#)]
43. Albrecht, E.W.J.A.; Stegeman, C.A.; Heeringa, P.; Henning, R.; Van Goor, H. Protective role of endothelial nitric oxide synthase. *J. Pathol.* **2003**, *199*, 8–17. [[CrossRef](#)]
44. Kolluru, G.K.; Siamwala, J.H.; Chatterjee, S. eNOS phosphorylation in health and disease. *Biochimie* **2010**, *92*, 1186–1198. [[CrossRef](#)]
45. Lupu, F.; Kinasevitz, G.; Dormer, K. The role of endothelial shear stress on haemodynamics, inflammation, coagulation and glycocalyx during sepsis. *J. Cell Mol. Med.* **2020**, *24*, 12258–12271. [[CrossRef](#)]
46. Lambden, S. Bench to bedside review: Therapeutic modulation of nitric oxide in sepsis—An update. *Intensive Care Med. Exp.* **2019**, *7*, 64. [[CrossRef](#)]
47. Cyr, A.R.; Huckaby, L.V.; Shiva, S.S.; Zuckerbraun, B.S. Nitric Oxide and Endothelial Dysfunction. *Crit. Care Clin.* **2020**, *36*, 307–321. [[CrossRef](#)] [[PubMed](#)]
48. Lundberg, J.O.; Weitzberg, E. Nitric oxide signaling in health and disease. *Cell* **2022**, *185*, 2853–2878. [[CrossRef](#)]
49. Weaver, S.R.C.; Rendeiro, C.; Lucas, R.A.I.; Cable, N.T.; Nightingale, T.E.; McGettrick, H.M.; Lucas, S.J.E. Non-pharmacological interventions for vascular health and the role of the endothelium. *Eur. J. Appl. Physiol.* **2022**, *122*, 2493–2514. [[CrossRef](#)]
50. Kuhlencordt, P.J.; Rosel, E.; Gerszten, R.E.; Morales-Ruiz, M.; Dombkowski, D.; Atkinson, W.J.; Han, F.; Preffer, F.; Rosenzweig, A.; Sessa, W.C.; et al. Role of endothelial nitric oxide synthase in endothelial activation—insights from eNOS knockout endothelial. *Am. J. Physiol. Cell Physiol.* **2004**, *286*, C1195–C1202. [[CrossRef](#)]
51. Stefano, G.; Prevot, V.; Cadet, P.; Dardik, I. Vascular pulsations stimulating nitric oxide release during cyclic exercise may benefit health: A molecular approach (Review). *Int. J. Mol. Med.* **2001**, *7*, 205–213. [[CrossRef](#)]
52. Virani, S.S.; Alonso, A.; Aparicio, H.J.; Benjamin, E.J.; Bittencourt, M.S.; Callaway, C.W.; Carson, A.P.; Chamberlain, A.M.; Cheng, S.; Delling, F.N.; et al. Prevention Statistics C, and Stroke Statistics S. Heart Disease and Stroke Statistics-2021 Update: A Report From the American Heart Association. *Circulation* **2021**, *143*, e254–e743. [[CrossRef](#)] [[PubMed](#)]
53. Jou, C.; Shah, R.; Figueroa, A.; Patel, J.K. The Role of Inflammatory Cytokines in Cardiac Arrest. *J. Intensive Care Med.* **2020**, *35*, 219–224. [[CrossRef](#)]
54. Adrie, C.; Adib-Conquy, M.; Laurent, I.; Monchi, M.; Vinsonneau, C.; Fitting, C.; Fraisse, F.; Dinh-Xuan, A.T.; Carli, P.; Spaulding, C.; et al. Successful Cardiopulmonary Resuscitation after Cardiac Arrest as a “Sepsis-Like” Syndrome. *Circulation* **2002**, *106*, 562–568. [[CrossRef](#)] [[PubMed](#)]

55. Bro-Jeppesen, J.; Johansson, P.I.; Hassager, C.; Wanscher, M.; Ostrowski, S.R.; Bjerre, M.; Kjaergaard, J. Endothelial activation/injury and associations with severity of post-cardiac arrest syndrome and mortality after out-of-hospital cardiac arrest. *Resuscitation* **2016**, *107*, 71–79. [[CrossRef](#)] [[PubMed](#)]
56. Cunningham, C.A.; Coppler, P.J.; Skolnik, A.B. The immunology of the post-cardiac arrest syndrome. *Resuscitation* **2022**, *179*, 116–123. [[CrossRef](#)]
57. Gando, S.; Nanzaki, S.; Morimoto, Y.; Kobayashi, S.; Kemmotsu, O. Out-of-hospital cardiac arrest increases soluble vascular endothelial adhesion molecules and neutrophil elastase associated with endothelial injury. *Intensive Care Med.* **2000**, *26*, 38–44. [[CrossRef](#)]
58. Chaban, V.; Nakstad, E.R.; Stær-Jensen, H.; Schjalm, C.; Seljeflot, I.; Vaage, J.; Lundqvist, C.; Benth, J.; Sunde, K.; Mollnes, T.E.; et al. Complement activation is associated with poor outcome after out-of-hospital cardiac arrest. *Resuscitation* **2021**, *166*, 129–136. [[CrossRef](#)]
59. Fink, K.; Schwarz, M.; Feldbrügge, L.; Sunkomat, J.N.; Schwab, T.; Bourgeois, N.; Olschewski, M.; Mühlen, C.V.Z.; Bode, C.; Busch, H.-J. Severe endothelial injury and subsequent repair in patients after successful cardiopulmonary resuscitation. *Crit. Care* **2010**, *14*, R104. [[CrossRef](#)]
60. Langeland, H.; Damås, J.K.; Mollnes, T.E.; Ludviksen, J.K.; Ueland, T.; Michelsen, A.E.; Løberg, M.; Bergum, D.; Nordseth, T.; Skjærvold, N.K.; et al. The inflammatory response is related to circulatory failure after out-of-hospital cardiac arrest: A prospective cohort study. *Resuscitation* **2022**, *170*, 115–125. [[CrossRef](#)] [[PubMed](#)]
61. Kofler, S.; Nickel, T.; Weis, M. Role of cytokines in cardiovascular diseases: A focus on endothelial responses to inflammation. *Clin. Sci.* **2005**, *108*, 205–213. [[CrossRef](#)] [[PubMed](#)]
62. Krishnaswamy, G.; Kelley, J.L.; Yerra, L.; Smith, J.K.; Chi, D.S. Human Endothelium as a Source of Multifunctional Cytokines: Molecular Regulation and Possible Role in Human Disease. *J. Interferon Cytokine Res.* **1999**, *19*, 91–104. [[CrossRef](#)]
63. Mai, J.; Virtue, A.; Shen, J.; Wang, H.; Yang, X.-F. An evolving new paradigm: Endothelial cells—Conditional innate immune cells. *J. Hematol. Oncol.* **2013**, *6*, 61. [[CrossRef](#)] [[PubMed](#)]
64. Moccetti, F.; Brown, E.; Xie, A.; Packwood, W.; Qi, Y.; Ruggeri, Z.; Shentu, W.; Chen, J.; López, J.A.; Lindner, J.R. Myocardial Infarction Produces Sustained Proinflammatory Endothelial Activation in Remote Arteries. *J. Am. Coll. Cardiol.* **2018**, *72*, 1015–1026. [[CrossRef](#)]
65. Toldo, S.; Mauro, A.G.; Cutter, Z.S.; Abbate, A. Inflammasome, pyroptosis, and cytokines in myocardial ischemia-reperfusion injury. *Am. J. Physiol. Circ. Physiol.* **2018**, *315*, H1553–H1568. [[CrossRef](#)]
66. Zindel, J.; Kubers, P. DAMPs, PAMPs, and LAMPs in Immunity and Sterile Inflammation. *Annu. Rev. Pathol. Mech. Dis.* **2020**, *15*, 493–518. [[CrossRef](#)]
67. Rudd, K.E.; Johnson, S.C.; Agesa, K.M.; Shackelford, K.A.; Tsoi, D.; Kievlan, D.R.; Colombara, D.V.; Ikuta, K.S.; Kisson, N.; Finfer, S.; et al. Global, regional, and national sepsis incidence and mortality, 1990–2017: Analysis for the Global Burden of Disease Study. *Lancet* **2020**, *395*, 200–211. [[CrossRef](#)] [[PubMed](#)]
68. Dolmatova, E.V.; Wang, K.; Mandavilli, R.; Griendling, K.K. The effects of sepsis on endothelium and clinical implications. *Cardiovasc. Res.* **2020**, *117*, 60–73. [[CrossRef](#)]
69. Raia, L.; Zafrani, L. Endothelial Activation and Microcirculatory Disorders in Sepsis. *Front. Med.* **2022**, *9*, 907992. [[CrossRef](#)]
70. Fernández-Sarmiento, J.; Molina, C.F.; Salazar-Pelaez, L.M.; Flórez, S.; Alarcón-Forero, L.C.; Sarta, M.; Hernández-Sarmiento, R.; Villar, J.C. Biomarkers of Glycocalyx Injury and Endothelial Activation are Associated with Clinical Outcomes in Patients with Sepsis: A Systematic Review and Meta-Analysis. *J. Intensive Care Med.* **2022**, *38*, 95–105. [[CrossRef](#)]
71. Mikacenic, C.; Hahn, W.; Price, B.L.; Harju-Baker, S.; Katz, R.L.; Kain, K.; Himmelfarb, J.; Liles, W.C.; Wurfel, M.M. Biomarkers of Endothelial Activation Are Associated with Poor Outcome in Critical Illness. *PLoS ONE* **2015**, *10*, e0141251. [[CrossRef](#)]
72. Juffermans, N.P.; van den Brom, C.E.; Kleinveld, D.J.B. Targeting Endothelial Dysfunction in Acute Critical Illness to Reduce Organ Failure. *Anesth. Analg.* **2020**, *131*, 1708–1720. [[CrossRef](#)]
73. López, A.; Lorente, J.A.; Steingrub, J.; Bakker, J.; McLuckie, A.; Willatts, S.; Brockway, M.; Anzueto, A.; Holzapfel, L.; Breen, D.; et al. Multiple-center, randomized, placebo-controlled, double-blind study of the nitric oxide synthase inhibitor 546C88: Effect on survival in patients with septic shock. *Crit. Care Med.* **2004**, *32*, 21–30. [[CrossRef](#)] [[PubMed](#)]
74. Bakker, J.; Grover, R.; McLuckie, A.; Holzapfel, L.; Andersson, J.; Lodato, R.; Watson, D.; Grossman, S.; Donaldson, J.; Takala, J.; et al. Administration of the nitric oxide synthase inhibitor NG-methyl-L-arginine hydrochloride (546C88) by intravenous infusion for up to 72 hours can promote the resolution of shock in patients with severe sepsis: Results of a randomized, double-blind, placebo-controlled multicenter study (study no. 144-002). *Crit. Care Med.* **2004**, *32*, 1–12. [[CrossRef](#)] [[PubMed](#)]
75. Singh, J.; Lee, Y.; Kellum, J.A. A new perspective on NO pathway in sepsis and ADMA lowering as a potential therapeutic approach. *Crit. Care* **2022**, *26*, 246. [[CrossRef](#)] [[PubMed](#)]
76. Yamashita, T.; Kawashima, S.; Ohashi, Y.; Ozaki, M.; Ueyama, T.; Ishida, T.; Inoue, N.; Hirata, K.-I.; Akita, H.; Yokoyama, M. Resistance to endotoxin shock in transgenic mice overexpressing endothelial nitric oxide synthase. *Circulation* **2000**, *101*, 931–937. [[CrossRef](#)]
77. Ichinose, F.; Buys, E.S.; Neilan, T.G.; Furutani, E.M.; Morgan, J.G.; Jassal, D.S.; Graveline, A.R.; Searles, R.J.; Lim, C.C.; Kaneki, M.; et al. Cardiomyocyte-Specific Overexpression of Nitric Oxide Synthase 3 Prevents Myocardial Dysfunction in Murine Models of Septic Shock. *Circ. Res.* **2007**, *100*, 130–139. [[CrossRef](#)] [[PubMed](#)]

78. Handa, O.; Stephen, J.; Cepinskas, G. Role of endothelial nitric oxide synthase-derived nitric oxide in activation and dysfunction of cerebrovascular endothelial cells during early onsets of sepsis. *Am. J. Physiol. Circ. Physiol.* **2008**, *295*, H1712–H1719. [[CrossRef](#)]
79. Piercy, K.L.; Troiano, R.P.; Ballard, R.M.; Carlson, S.A.; Fulton, J.E.; Galuska, D.A.; George, S.M.; Olson, R.D. The Physical Activity Guidelines for Americans. *JAMA* **2018**, *320*, 2020–2028. [[CrossRef](#)]
80. Liguori, G. (Ed.) Benefits and Risks Associated with Physical Activity. In *ACSMs Guidelines for Exercise Testing and Prescription*; Wolters Kluwer: Alphen aan den Rijn, The Netherlands, 2021.
81. Bender, S.B.; Laughlin, M.H. Modulation of endothelial cell phenotype by physical activity: Impact on obesity-related endothelial dysfunction. *Am. J. Physiol. Circ. Physiol.* **2015**, *309*, H1–H8. [[CrossRef](#)]
82. Padilla, J.; Simmons, G.H.; Bender, S.B.; Esquivel, A.A.; Whyte, J.J.; Laughlin, M.H. Vascular Effects of Exercise: Endothelial Adaptations Beyond Active Muscle Beds. *Physiology* **2011**, *26*, 132–145. [[CrossRef](#)] [[PubMed](#)]
83. Green, D.J.; Smith, K.J. Effects of Exercise on Vascular Function, Structure, and Health in Humans. *Cold Spring Harb. Perspect. Med.* **2017**, *8*, a029819. [[CrossRef](#)] [[PubMed](#)]
84. Green, D.J.; Hopman, M.T.E.; Padilla, J.; Laughlin, M.H.; Thijssen, D.H.J. Vascular Adaptation to Exercise in Humans: Role of Hemodynamic Stimuli. *Physiol. Rev.* **2017**, *97*, 495–528. [[CrossRef](#)]
85. Du, H.; Bartleson, J.M.; Butenko, S.; Alonso, V.; Liu, W.F.; Winer, D.A.; Butte, M.J. Tuning immunity through tissue mechanotransduction. *Nat. Rev. Immunol.* **2022**, *16*, 1–15. [[CrossRef](#)]
86. Genkel, V.V.; Kuznetcova, A.S.; Shaposhnik, I.I. Biomechanical Forces and Atherosclerosis: From Mechanism to Diagnosis and Treatment. *Curr. Cardiol. Rev.* **2020**, *16*, 187–197. [[CrossRef](#)]
87. Clermont, C.A.; Benson, L.C.; Osis, S.T.; Kobsar, D.; Ferber, R. Running patterns for male and female competitive and recreational runners based on accelerometer data. *J. Sports Sci.* **2019**, *37*, 204–211. [[CrossRef](#)]
88. Adams, V.; Reich, B.; Uhlemann, M.; Niebauer, J. Molecular effects of exercise training in patients with cardiovascular disease: Focus on skeletal muscle, endothelium, and myocardium. *Am. J. Physiol. Circ. Physiol.* **2017**, *313*, H72–H88. [[CrossRef](#)]
89. Kojda, G.; Hambrecht, R. Molecular mechanisms of vascular adaptations to exercise. Physical activity as an effective antioxidant therapy? *Cardiovasc. Res.* **2005**, *67*, 187–197. [[CrossRef](#)] [[PubMed](#)]
90. Ploughman, M. Exercise is brain food: The effects of physical activity on cognitive function. *Dev. Neurorehabil.* **2008**, *11*, 236–240. [[CrossRef](#)] [[PubMed](#)]
91. Trigiani, L.J.; Hamel, E. An endothelial link between the benefits of physical exercise in dementia. *J. Cereb. Blood Flow Metab.* **2017**, *37*, 2649–2664. [[CrossRef](#)]
92. Trinity, J.D.; Richardson, R.S. Physiological Impact and Clinical Relevance of Passive Exercise/Movement. *Sports Med.* **2019**, *49*, 1365–1381. [[CrossRef](#)] [[PubMed](#)]
93. Pinckard, K.; Baskin, K.K.; Stanford, K.I. Effects of Exercise to Improve Cardiovascular Health. *Front. Cardiovasc. Med.* **2019**, *6*, 69. [[CrossRef](#)] [[PubMed](#)]
94. Fletcher, G.F.; Landolfo, C.; Niebauer, J.; Ozemek, C.; Arena, R.; Lavie, C.J. Reprint of: Promoting Physical Activity and Exercise: JACC Health Promotion Series. *J. Am. Coll. Cardiol.* **2018**, *72*, 3053–3070. [[CrossRef](#)] [[PubMed](#)]
95. Wilson, M.; Ellison, G.; Cable, N. Basic science behind the cardiovascular benefits of exercise. *Br. J. Sports Med.* **2016**, *50*, 93–99. [[CrossRef](#)] [[PubMed](#)]
96. Seo, D.Y.; Ko, J.R.; Jang, J.E.; Kim, T.N.; Youm, J.B.; Kwak, H.B.; Bae, J.H.; Kim, A.H.; Ko, K.S.; Rhee, B.D.; et al. Exercise as A Potential Therapeutic Target for Diabetic Cardiomyopathy: Insight into the Underlying Mechanisms. *Int. J. Mol. Sci.* **2019**, *20*, 6284. [[CrossRef](#)] [[PubMed](#)]
97. Yu, M.; Tsai, S.-F.; Kuo, Y.-M. The Therapeutic Potential of Anti-Inflammatory Exerkines in the Treatment of Atherosclerosis. *Int. J. Mol. Sci.* **2017**, *18*, 1260. [[CrossRef](#)]
98. Bernardo, B.C.; Ooi, J.Y.Y.; Weeks, K.L.; Patterson, N.L.; McMullen, J.R. Understanding Key Mechanisms of Exercise-Induced Cardiac Protection to Mitigate Disease: Current Knowledge and Emerging Concepts. *Physiol. Rev.* **2018**, *98*, 419–475. [[CrossRef](#)]
99. Arnett, D.K.; Blumenthal, R.S.; Albert, M.A.; Buroker, A.B.; Goldberger, Z.D.; Hahn, E.J.; Himmelfarb, C.D.; Khera, A.; Lloyd-Jones, D.; McEvoy, J.W.; et al. 2019 ACC/AHA Guideline on the Primary Prevention of Cardiovascular Disease: A Report of the American College of Cardiology/American Heart Association Task Force on Clinical Practice Guidelines. *Circulation* **2019**, *140*, e596–e646. [[CrossRef](#)] [[PubMed](#)]
100. Petersen, A.M.W. The anti-inflammatory effect of exercise. *J. Appl. Physiol.* **2005**, *98*, 1154–1162. [[CrossRef](#)] [[PubMed](#)]
101. Ding, Y.; Xu, X. Effects of regular exercise on inflammasome activation-related inflammatory cytokine levels in older adults: A systematic review and meta-analysis. *J. Sports Sci.* **2021**, *39*, 2338–2352. [[CrossRef](#)]
102. Lee, J.; Hong, J.; Umetani, M.; Lavoy, E.C.; Kim, J.-H.; Park, Y. Vascular Protection by Exercise in Obesity: Inflammasome-associated Mechanisms. *Med. Sci. Sports Exerc.* **2020**, *52*, 2538–2545. [[CrossRef](#)]
103. Borges, J.P.; Verdoorn, K.D.S. Cardiac Ischemia/Reperfusion Injury: The Beneficial Effects of Exercise. *Adv. Exp. Med. Biol.* **2017**, *999*, 155–179. [[CrossRef](#)]
104. Dibben, G.; Faulkner, J.; Oldridge, N.; Rees, K.; Thompson, D.R.; Zwisler, A.D.; Taylor, R.S. Exercise-based cardiac rehabilitation for coronary heart disease. *Cochrane Database Syst. Rev.* **2021**, *11*, CD001800. [[PubMed](#)]
105. Couto, G.K.; Paula, S.M.; Gomes-Santos, I.L.; Negrão, C.E.; Rossoni, L.V. Exercise training induces eNOS coupling and restores relaxation in coronary arteries of heart failure rats. *Am. J. Physiol. Circ. Physiol.* **2018**, *314*, H878–H887. [[CrossRef](#)] [[PubMed](#)]

106. Pedersen, B.K. Anti-inflammatory effects of exercise: Role in diabetes and cardiovascular disease. *Eur. J. Clin. Investig.* **2017**, *47*, 600–611. [[CrossRef](#)]
107. Zhao, H.; He, Z.; Yun, H.; Wang, R.; Liu, C. A Meta-Analysis of the Effects of Different Exercise Modes on Inflammatory Response in the Elderly. *Int. J. Environ. Res. Public Health* **2022**, *19*, 10451. [[CrossRef](#)] [[PubMed](#)]
108. Caldwell, R.W.; Rodriguez, P.C.; Toque, H.A.; Narayanan, S.P. Arginase: A Multifaceted Enzyme Important in Health and Disease. *Physiol. Rev.* **2018**, *98*, 641–665. [[CrossRef](#)] [[PubMed](#)]
109. Pope, A.J.; Karuppiyah, K.; Cardounel, A.J. Role of the PRMT-DDAH-ADMA axis in the regulation of endothelial nitric oxide production. *Pharmacol. Res.* **2009**, *60*, 461–465. [[CrossRef](#)]
110. Powers, S.K.; Deminice, R.; Ozdemir, M.; Yoshihara, T.; Bomkamp, M.P.; Hyatt, H. Exercise-induced oxidative stress: Friend or foe? *J. Sport Health Sci.* **2020**, *9*, 415–425. [[CrossRef](#)] [[PubMed](#)]
111. Radak, Z.; Chung, H.Y.; Goto, S. Exercise and hormesis: Oxidative stress-related adaptation for successful aging. *Biogerontology* **2005**, *6*, 71–75. [[CrossRef](#)] [[PubMed](#)]
112. Radak, Z.; Chung, H.Y.; Koltai, E.; Taylor, A.W.; Goto, S. Exercise, oxidative stress and hormesis. *Ageing Res. Rev.* **2008**, *7*, 34–42. [[CrossRef](#)]
113. Anderson, L.; Oldridge, N.; Thompson, D.R.; Zwisler, A.D.; Rees, K.; Martin, N.; Taylor, R.S. Exercise-Based Cardiac Rehabilitation for Coronary Heart Disease: Cochrane Systematic Review and Meta-Analysis. *J. Am. Coll. Cardiol.* **2016**, *67*, 1–12. [[CrossRef](#)]
114. Borges, J.P.; Lessa, M.A. Mechanisms Involved in Exercise-Induced Cardioprotection: A Systematic Review. *Arq. Bras. Cardiol.* **2015**, *105*, 71–81. [[CrossRef](#)]
115. Tucker, W.J.; Fegers-Wustrow, I.; Halle, M.; Haykowsky, M.J.; Chung, E.H.; Kovacic, J.C. Exercise for Primary and Secondary Prevention of Cardiovascular Disease: JACC Focus Seminar 1/4. *J. Am. Coll. Cardiol.* **2022**, *80*, 1091–1106. [[CrossRef](#)]
116. Kim, W.H.; Park, J.H.; Jeong, J.; Ro, Y.S.; Hong, K.J.; Song, K.J.; Shin, S.D.; Hwang, S. Intensity of physical activity for out-of-hospital cardiac arrests during exercise and survival outcomes. *Am. J. Emerg. Med.* **2022**, *55*, 221–223. [[CrossRef](#)]
117. Irahara, T.; Sato, N.; Inoue, K.; Otake, K.; Ohtsuru, S.; Koike, K.; Fushiki, T.; Yokota, H. Low-intensity exercise in the acute phase of lipopolysaccharide-induced sepsis improves lipid metabolism and survival in mice by stimulating PGC-1 α expression. *J. Trauma Acute Care Surg.* **2016**, *80*, 933–940. [[CrossRef](#)]
118. Yamada, M.; Hokazono, C.; Okutsu, M. Maternal exercise training attenuates endotoxin-induced sepsis in mice offspring. *Biochem. Biophys. Rep.* **2018**, *15*, 19–24. [[CrossRef](#)]
119. Tysl, K.; Swarbreck, S.; Pape, C.; Secor, D.; Koropatnick, J.; Feng, Q.; Veldhuizen, R.A.W.; Gill, S.E. Voluntary running exercise protects against sepsis-induced early inflammatory and pro-coagulant responses in aged mice. *Crit. Care* **2017**, *21*, 210. [[CrossRef](#)]
120. Williams, P.T. Inadequate Exercise as a Risk Factor for Sepsis Mortality. *PLoS ONE* **2013**, *8*, e79344. [[CrossRef](#)]
121. Kim, D.; Kang, H. Exercise training modifies gut microbiota with attenuated host responses to sepsis in wild-type mice. *FASEB J.* **2019**, *33*, 5772–5781. [[CrossRef](#)]
122. de Araujo, C.C.; Silva, J.D.; Samary, C.S.; Guimaraes, I.H.; Marques, P.S.; Oliveira, G.P.; do Carmo, L.G.; Goldenberg, R.C.; Bakker-Abreu, I.; Diaz, B.L.; et al. Regular and moderate exercise before experimental sepsis reduces the risk of lung and distal organ injury. *J. Appl. Physiol.* **2012**, *112*, 1206–1214. [[CrossRef](#)]
123. Sossdorf, M.; Fischer, J.; Meyer, S.; Dahlke, K.; Wissuwa, B.; Seidel, C.; Schreppler, A.; Bockmeyer, C.L.; Lupp, A.; Neugebauer, S.; et al. Physical Exercise Induces Specific Adaptations Resulting in Reduced Organ Injury and Mortality During Severe Polymicrobial Sepsis. *Crit. Care Med.* **2013**, *41*, e246–e255. [[CrossRef](#)]
124. Chen, H.I.; Hsieh, S.-Y.; Yang, F.-L.; Hsu, Y.H.; Lin, C.-C. Exercise Training Attenuates Septic Responses in Conscious Rats. *Med. Sci. Sports Exerc.* **2007**, *39*, 435–442. [[CrossRef](#)]
125. Gholamnezhad, Z.; Safarian, B.; Esparham, A.; Mirzaei, M.; Esmaeilzadeh, M.; Boskabady, M.H. The modulatory effects of exercise on lipopolysaccharide-induced lung inflammation and injury: A systemic review. *Life Sci.* **2022**, *293*, 120306. [[CrossRef](#)]
126. Ahn, J.Y.; Song, J.E.; Ann, H.W.; Jeon, Y.; Ahn, M.Y.; Jung, I.Y.; Kim, M.H.; Jeong, W.; Jeong, S.J.; Ku, N.S.; et al. Effects of Early Exercise Rehabilitation on Functional Recovery in Patients with Severe Sepsis. *Yonsei Med. J.* **2018**, *59*, 843–851. [[CrossRef](#)]
127. Kayambu, G.; Boots, R.; Paratz, J. Early physical rehabilitation in intensive care patients with sepsis syndromes: A pilot randomised controlled trial. *Intensive Care Med.* **2015**, *41*, 865–874. [[CrossRef](#)]
128. Wu, Y.; Guo, X.; Peng, Y.; Fang, Z.; Zhang, X. Roles and Molecular Mechanisms of Physical Exercise in Sepsis Treatment. *Front. Physiol.* **2022**, *13*, 879430. [[CrossRef](#)]
129. Raza, A.; Steinberg, K.; Tartaglia, J.; Frishman, W.H.; Gupta, T. Enhanced External Counterpulsation Therapy: Past, Present, and Future. *Cardiol. Rev.* **2017**, *25*, 59–67. [[CrossRef](#)]
130. Zheng, Z.S.; Yu, L.Q.; Cai, S.R.; Kambic, H.; Li, T.M.; Ma, H.; Chen, P.Z.; Huang, B.J.; Nose, Y. New sequential external counterpulsation for the treatment of acute myocardial infarction. *Artif. Organs* **1984**, *8*, 470–477. [[CrossRef](#)]
131. Akhtar, M.; Wu, G.-F.; Du, Z.-M.; Zheng, Z.-S.; Michaels, A.D. Effect of External Counterpulsation on Plasma Nitric Oxide and Endothelin-1 Levels. *Am. J. Cardiol.* **2006**, *98*, 28–30. [[CrossRef](#)]
132. Shechter, M.; Matetzky, S.; Feinberg, M.S.; Chouraqui, P.; Rotstein, Z.; Hod, H. External counterpulsation therapy improves endothelial function in patients with refractory angina pectoris. *J. Am. Coll. Cardiol.* **2003**, *42*, 2090–2095. [[CrossRef](#)]
133. Braith, R.W.; Conti, C.R.; Nichols, W.W.; Choi, C.Y.; Khuddus, M.A.; Beck, D.T.; Casey, D.P. Enhanced external counterpulsation improves peripheral artery flow-mediated dilation in patients with chronic angina: A randomized sham-controlled study. *Circulation* **2010**, *122*, 1612–1620. [[CrossRef](#)]

134. Zhang, Y.; He, X.; Liu, D.; Wu, G.; Chen, X.; Ma, H.; Du, Z.; Dong, Y.; Jin, Y.; He, W.; et al. Enhanced External Counterpulsation Attenuates Atherosclerosis Progression Through Modulation of Proinflammatory Signal Pathway. *Arterioscler. Thromb. Vasc. Biol.* **2010**, *30*, 773–780. [[CrossRef](#)]
135. Martin, J.S.; Braith, R.W. Anti-inflammatory effects of enhanced external counterpulsation in subjects with abnormal glucose tolerance. *Appl. Physiol. Nutr. Metab.* **2012**, *37*, 1251–1255. [[CrossRef](#)]
136. Lin, S.; Xiao-Ming, W.; Gui-Fu, W. Expert consensus on the clinical application of enhanced external counterpulsation in elderly people (2019). *Aging Med.* **2020**, *3*, 19–27. [[CrossRef](#)]
137. Bøtker, H.E.; Lassen, T.R.; Jespersen, N.R. Clinical translation of myocardial conditioning. *Am. J. Physiol. Circ. Physiol.* **2018**, *314*, H1225–H1252. [[CrossRef](#)]
138. Wu, G.; Du, Z.; Hu, C.; Zheng, Z.; Zhan, C.; Ma, H.; Fang, D.; Ahmed, K.T.; Laham, R.J.; Hui, J.C.K.; et al. Angiogenic effects of long-term enhanced external counterpulsation in a dog model of myocardial infarction. *Am. J. Physiol. Circ. Physiol.* **2006**, *290*, H248–H254. [[CrossRef](#)]
139. Hu, C.L.; Liu, R.; Liao, X.X.; Wei, H.Y.; Li, X.; Zhan, H.; Jing, X.L.; Xiong, Y.; Huang, G.Q.; Wu, G.F. Early enhanced external counter pulsation improves neurological recovery after the return of spontaneous circulation in a mongrel dog cardiac arrest model. *Crit. Care Med.* **2013**, *41*, e62–e73. [[CrossRef](#)]
140. Xiong, J.; Zhang, W.; Wei, H.; Li, X.; Dai, G.; Hu, C. Enhanced external counterpulsation improves cardiac function in Beagles after cardiopulmonary resuscitation. *Braz. J. Med. Biol. Res.* **2020**, *53*, e9136. [[CrossRef](#)]
141. Casey, D.P.; Conti, C.R.; Nichols, W.W.; Choi, C.Y.; Khuddus, M.A.; Braith, R.W. Effect of Enhanced External Counterpulsation on Inflammatory Cytokines and Adhesion Molecules in Patients with Angina Pectoris and Angiographic Coronary Artery Disease. *Am. J. Cardiol.* **2008**, *101*, 300–302. [[CrossRef](#)]
142. Yang, D.-Y.; Wu, G.-F. Vasculoprotective properties of enhanced external counterpulsation for coronary artery disease: Beyond the hemodynamics. *Int. J. Cardiol.* **2013**, *166*, 38–43. [[CrossRef](#)]
143. Dayrit, J.K.; Verduzco-Gutierrez, M.; Teal, A.; Shah, S.A. Enhanced External Counterpulsation as a Novel Treatment for Post-acute COVID-19 Sequelae. *Cureus* **2021**, *13*, 14358. [[CrossRef](#)]
144. Hoover, G.N.; Ashe, W.F.; Dines, J.H.; Fraser, T.M. Vibration studies. III. Blood pressure responses to whole-body vibration in anesthetized dogs. *Arch. Environ. Health* **1961**, *3*, 426–432. [[CrossRef](#)]
145. Young, W.A.; Shaw, D.B.; Navach, J.; Shizgal, H.; Kowalsky, N. Effect of CO₂ and whole-body vibration on ventilation. *J. Appl. Physiol.* **1965**, *20*, 844–848. [[CrossRef](#)]
146. Shoenberger, R.W. Human Response to Whole-Body Vibration. *Percept. Mot. Ski.* **1972**, *34*, 127–160. [[CrossRef](#)]
147. Adams, J.A.; Uryash, A.; Lopez, J.R.; Sackner, M.A. The Endothelium as a Therapeutic Target in Diabetes: A Narrative Review and Perspective. *Front. Physiol.* **2021**, *12*, 638491. [[CrossRef](#)]
148. Rittweger, J. Vibration as an exercise modality: How it may work, and what its potential might be. *Eur. J. Appl. Physiol.* **2010**, *108*, 877–904. [[CrossRef](#)]
149. Cardinale, M.; Wakeling, J. Whole body vibration exercise: Are vibrations good for you? *Br. J. Sports Med.* **2005**, *39*, 585–589. [[CrossRef](#)]
150. Huang, Y.; Zhang, P. Subjective discomfort caused by vertical whole-body vibration in the frequency range 2–100 Hz. *Ergonomics* **2019**, *62*, 420–430. [[CrossRef](#)]
151. Zhou, Z.; Griffin, M.J. Response of the seated human body to whole-body vertical vibration: Discomfort caused by sinusoidal vibration. *Ergonomics* **2014**, *57*, 714–732. [[CrossRef](#)]
152. Pel, J.J.M.; Bagheri, J.; van Dam, L.M.; van den Berg-Emons, H.J.G.; Horemans, H.L.D.; Stam, H.J.; van der Steen, J. Platform accelerations of three different whole-body vibration devices and the transmission of vertical vibrations to the lower limbs. *Med. Eng. Phys.* **2009**, *31*, 937–944. [[CrossRef](#)]
153. Johnson, P.K.; Feland, J.B.; Johnson, A.W.; Mack, G.W.; Mitchell, U.H. Effect of Whole Body Vibration on Skin Blood Flow and Nitric Oxide Production. *J. Diabetes Sci. Technol.* **2014**, *8*, 889–894. [[CrossRef](#)] [[PubMed](#)]
154. Games, K.E.; Sefton, J.M.; Wilson, A.E. Whole-Body Vibration and Blood Flow and Muscle Oxygenation: A Meta-Analysis. *J. Athl. Train.* **2015**, *50*, 542–549. [[CrossRef](#)]
155. Robbins, D.; Yoganathan, P.; Goss-Sampson, M. The influence of whole body vibration on the central and peripheral cardiovascular system. *Clin. Physiol. Funct. Imaging* **2014**, *34*, 364–369. [[CrossRef](#)] [[PubMed](#)]
156. Aoyama, A.; Yamaoka-Tojo, M.; Obara, S.; Shimizu, E.; Fujiyoshi, K.; Noda, C.; Matsunaga, A.; Ako, J. Acute Effects of Whole-Body Vibration Training on Endothelial Function and Cardiovascular Response in Elderly Patients with Cardiovascular Disease. *Int. Heart J.* **2019**, *60*, 854–861. [[CrossRef](#)] [[PubMed](#)]
157. Robinson, C.C.; Barreto, R.; Sbruzzi, G.; Plentz, R.D.M. The effects of whole body vibration in patients with type 2 diabetes: A systematic review and meta-analysis of randomized controlled trials. *Braz. J. Phys. Ther.* **2016**, *20*, 4–14. [[CrossRef](#)]
158. Fischer, M.; Vialleron, T.; Laffaye, G.; Fourcade, P.; Hussein, T.; Cheze, L.; Deleu, P.-A.; Honeine, J.-L.; Yiou, E.; Delafontaine, A. Long-Term Effects of Whole-Body Vibration on Human Gait: A Systematic Review and Meta-Analysis. *Front. Neurol.* **2019**, *10*, 627. [[CrossRef](#)] [[PubMed](#)]
159. Saquetto, M.B.; Pereira, F.F.; Queiroz, R.S.; da Silva, C.M.; Conceição, C.S.; Neto, M.G. Effects of whole-body vibration on muscle strength, bone mineral content and density, and balance and body composition of children and adolescents with Down syndrome: A systematic review. *Osteoporos. Int.* **2018**, *29*, 527–533. [[CrossRef](#)]

160. Shekarforoush, S.; Naghii, M.R. Whole-Body Vibration Training Increases Myocardial Salvage Against Acute Ischemia in Adult Male Rats. *Arq. Bras. Cardiol.* **2019**, *112*, 32–37. [[CrossRef](#)]
161. Nowak-Lis, A.; Nowak, Z.; Gabrys, T.; Szmatlan-Gabrys, U.; Batalik, L.; Knappova, V. The Use of Vibration Training in Men after Myocardial Infarction. *Int. J. Environ. Res. Public Health* **2022**, *19*, 3326. [[CrossRef](#)]
162. Kim, I.; Lee, B.; Yoo, S.; Hwang, S. Whole Body Vibration Reduces Inflammatory Bone Loss in a Lipopolysaccharide Murine Model. *J. Dent. Res.* **2014**, *93*, 704–710. [[CrossRef](#)] [[PubMed](#)]
163. Sañudo, B.; Seixas, A.; Gloeckl, R.; Rittweger, J.; Rawer, R.; Taiar, R.; Van Der Zee, E.A.; Van Heuvelen, M.J.; Lacerda, A.C.; Sartorio, A.; et al. Potential Application of Whole Body Vibration Exercise for Improving the Clinical Conditions of COVID-19 Infected Individuals: A Narrative Review from the World Association of Vibration Exercise Experts (WAVex) Panel. *Int. J. Environ. Res. Public Health* **2020**, *17*, 3650. [[CrossRef](#)] [[PubMed](#)]
164. Sá-Caputo, D.C.; Coelho-Oliveira, A.C.; Pessanha-Freitas, J.; Paineiras-Domingos, L.L.; Lacerda, A.C.R.; Mendonça, V.A.; Souza, A.; Taiar, R.; Sartorio, A.; Seixas, A.; et al. Whole-Body Vibration Exercise: A Possible Intervention in the Management of Post COVID-19 Complications? *Appl. Sci.* **2021**, *11*, 5733. [[CrossRef](#)]
165. Sanni, A.A.; Blanks, A.M.; Derella, C.C.; Horsager, C.; Crandall, R.H.; Looney, J.; Sanchez, S.; Norland, K.; Ye, B.; Thomas, J.; et al. The effects of whole-body vibration amplitude on glucose metabolism, inflammation, and skeletal muscle oxygenation. *Physiol. Rep.* **2022**, *10*, e15208. [[CrossRef](#)] [[PubMed](#)]
166. Rodriguez-Miguel, P.; Fernandez-Gonzalo, R.; Collado, P.S.; Almar, M.; Martinez-Florez, S.; de Paz, J.A.; González-Gallego, J.; Cuevas, M.J. Whole-body vibration improves the anti-inflammatory status in elderly subjects through toll-like receptor 2 and 4 signaling pathways. *Mech. Ageing Dev.* **2015**, *150*, 12–19. [[CrossRef](#)]
167. Jawed, Y.; Beli, E.; March, K.; Kaleth, A.; Loghmani, M.T. Whole-Body Vibration Training Increases Stem/Progenitor Cell Circulation Levels and May Attenuate Inflammation. *Mil. Med.* **2020**, *185*, 404–412. [[CrossRef](#)] [[PubMed](#)]
168. Neves, C.D.C.; Lacerda, A.C.R.; Lage, V.; Soares, A.A.; Chaves, M.G.A.; Lima, L.P.; Silva, T.J.; Vieira, L.M.; Teixeira, A.L.; Leite, H.R.; et al. Whole body vibration training increases physical measures and quality of life without altering inflammatory-oxidative biomarkers in patients with moderate COPD. *J. Appl. Physiol.* **2018**, *125*, 520–528. [[CrossRef](#)] [[PubMed](#)]
169. Cristi-Montero, C.; Collado, P.S.; Márquez, S.; Garatachea, N.; Cuevas, M.J. Whole-body vibration training increases physical fitness measures without alteration of inflammatory markers in older adults. *Eur. J. Sport Sci.* **2014**, *14*, 611–619. [[CrossRef](#)] [[PubMed](#)]
170. Sackner, M.A.; Gummels, E.; Adams, J.A. Nitric Oxide Is Released into Circulation with Whole-Body, Periodic Acceleration. *Chest* **2005**, *127*, 30–39. [[CrossRef](#)] [[PubMed](#)]
171. Uryash, A.; Wu, H.; Bassuk, J.; Kurlansky, P.; Sackner, M.A.; Adams, J.A. Low-amplitude pulses to the circulation through periodic acceleration induces endothelial-dependent vasodilatation. *J. Appl. Physiol.* **2009**, *106*, 1840–1847. [[CrossRef](#)] [[PubMed](#)]
172. Adams, J.A.; Bassuk, J.; Wu, N.; Graña, M.; Kurlansky, P.; Sackner, M.A. Periodic acceleration: Effects on vasoactive, fibrinolytic, and coagulation factors. *J. Appl. Physiol.* **2005**, *98*, 1083–1090. [[CrossRef](#)]
173. Sackner, M.A.; Gummels, E.; Adams, J.A. Effect of Moderate-Intensity Exercise, Whole-Body Periodic Acceleration, and Passive Cycling on Nitric Oxide Release into Circulation. *Chest* **2005**, *128*, 2794–2803. [[CrossRef](#)]
174. Fujita, M.; Tambara, K.; Ikemoto, M.; Sakamoto, S.; Ogai, A.; Kitakaze, M.; Sackner, M. Periodic Acceleration Enhances Release of Nitric Oxide in Healthy Adults. *Int. J. Angiol.* **2005**, *14*, 11–14. [[CrossRef](#)]
175. Wu, H.; Jin, Y.; Arias, J.; Bassuk, J.; Uryash, A.; Kurlansky, P.; Webster, K.; Adams, J.A. In vivo upregulation of nitric oxide synthases in healthy rats. *Nitric Oxide* **2009**, *21*, 63–68. [[CrossRef](#)]
176. Wu, H.; Uryash, A.; Bassuk, J.; Kurlansky, P.; Giridharan, G.A.; Shakeri, M.; Estrada, R.; Sethu, P.; Adams, J.A. Mechanisms of Periodic Acceleration Induced Endothelial Nitric Oxide Synthase (eNOS) Expression and Upregulation Using an In Vitro Human Aortic Endothelial Cell Model. *Cardiovasc. Eng. Technol.* **2012**, *3*, 292–301. [[CrossRef](#)]
177. Uryash, A.; Bassuk, J.; Kurlansky, P.; Altamirano, F.; Lopez, J.R.; Adams, J.A. Antioxidant Properties of Whole Body Periodic Acceleration (pGz). *PLoS ONE* **2015**, *10*, e0131392. [[CrossRef](#)] [[PubMed](#)]
178. Martínez, A.; Arias, J.; Bassuk, J.A.; Wu, H.; Kurlansky, P.; Adams, J.A. Adrenomedullin is increased by pulsatile shear stress on the vascular endothelium via periodic acceleration (pGz). *Peptides* **2008**, *29*, 73–78. [[CrossRef](#)] [[PubMed](#)]
179. Adams, J.A.; Bassuk, J.; Wu, D.; Kurlansky, P. Survival and normal neurological outcome after CPR with periodic Gz acceleration and vasopressin. *Resuscitation* **2003**, *56*, 215–221. [[CrossRef](#)] [[PubMed](#)]
180. Adams, J.A.; Bassuk, J.A.; Arias, J.; Wu, H.; Jorapur, V.; Lamas, G.A.; Kurlansky, P. Periodic acceleration (pGz) CPR in a swine model of asphyxia induced cardiac arrest: Short-term hemodynamic comparisons. *Resuscitation* **2008**, *77*, 132–138. [[CrossRef](#)] [[PubMed](#)]
181. Vinten-Johansen, J.; Shi, W. Preconditioning and postconditioning: Current knowledge, knowledge gaps, barriers to adoption, and future directions. *J. Cardiovasc. Pharmacol. Ther.* **2011**, *16*, 260–266. [[CrossRef](#)] [[PubMed](#)]
182. Adams, J.A.; Wu, H.; Bassuk, J.A.; Arias, J.; Uryash, A.; Jorapur, V.; Lamas, G.A.; Kurlansky, P. Periodic acceleration (pGz) prior to whole body Ischemia reperfusion injury provides early cardioprotective preconditioning. *Life Sci.* **2010**, *86*, 707–715. [[CrossRef](#)] [[PubMed](#)]
183. Uryash, A.; Wu, H.; Bassuk, J.; Kurlansky, P.; Adams, J.A. Preconditioning with periodic acceleration (pGz) provides second window of cardioprotection. *Life Sci.* **2012**, *91*, 178–185. [[CrossRef](#)] [[PubMed](#)]

184. Adams, J.A.; Uryash, A.; Wu, H.; Bassuk, J.A.; Nadkarni, V.; Berg, R.; Jorapur, V.; Kurlansky, P. Microcirculatory and therapeutic effects of whole body periodic acceleration (pGz) applied after cardiac arrest in pigs. *Resuscitation* **2011**, *82*, 767–775. [[CrossRef](#)] [[PubMed](#)]
185. Uryash, A.; Bassuk, J.; Kurlansky, P.; Altamirano, F.; Lopez, J.R.; Adams, J.A. Non-invasive technology that improves cardiac function after experimental myocardial infarction: Whole Body Periodic Acceleration (pGz). *PLoS ONE* **2015**, *10*, e0121069. [[CrossRef](#)] [[PubMed](#)]
186. Miyamoto, S.; Fujita, M.; Inoko, M.; Oba, M.; Hosokawa, R.; Haruna, T.; Izumi, T.; Saji, Y.; Nakane, E.; Abe, T.; et al. Effect on Treadmill Exercise Capacity, Myocardial Ischemia, and Left Ventricular Function as a Result of Repeated Whole-Body Periodic Acceleration with Heparin Pretreatment in Patients with Angina Pectoris and Mild Left Ventricular Dysfunction. *Am. J. Cardiol.* **2011**, *107*, 168–174. [[CrossRef](#)]
187. Rokutanda, T.; Izumiya, Y.; Miura, M.; Fukuda, S.; Shimada, K.; Izumi, Y.; Nakamura, Y.; Araki, S.; Hanatani, S.; Matsubara, J.; et al. Passive Exercise Using Whole-Body Periodic Acceleration Enhances Blood Supply to Ischemic Hindlimb. *Arterioscler. Thromb. Vasc. Biol.* **2011**, *31*, 2872–2880. [[CrossRef](#)]
188. Adams, J.A.; Uryash, A.; Lopez, J.R.; Sackner, M.A. Whole body periodic acceleration improves survival and microvascular leak in a murine endotoxin model. *PLoS ONE* **2019**, *14*, e0208681. [[CrossRef](#)]
189. Adams, J.A.; Lopez, J.R.; Uryash, A.; Sackner, M.A. Whole body periodic acceleration (pGz) improves endotoxin induced cardiomyocyte contractile dysfunction and attenuates the inflammatory response in mice. *Heliyon* **2021**, *7*, e06444. [[CrossRef](#)] [[PubMed](#)]
190. Sackner, M.A.; Adams, J.A. Endothelial pulsatile shear stress is a backstop for COVID-19. *Emerg. Top. Life Sci.* **2020**, *4*, 391–399. [[CrossRef](#)]
191. Adams, J.A.; Patel, S.; Lopez, J.R.; Sackner, M.A. The Effects of Passive Simulated Jogging on Short-Term Heart Rate Variability in a Heterogeneous Group of Human Subjects. *J. Sports Med.* **2018**, *2018*, 4340925. [[CrossRef](#)] [[PubMed](#)]
192. Sackner, M.A.; Patel, S.; Adams, J.A. Changes of blood pressure following initiation of physical inactivity and after external addition of pulses to circulation. *Eur. J. Appl. Physiol.* **2019**, *119*, 201–211. [[CrossRef](#)]
193. Adams, J.A.; Banderas, V.; Lopez, J.R.; Sackner, M.A. Portable Gentle Jogger Improves Glycemic Indices in Type 2 Diabetic and Healthy Subjects Living at Home: A Pilot Study. *J. Diabetes Res.* **2020**, *2020*, 8317973. [[CrossRef](#)] [[PubMed](#)]
194. Sackner, M.A.; Lopez, J.R.; Banderas, V.; Adams, J.A. Can Physical Activity While Sedentary Produce Health Benefits? A Single-Arm Randomized Trial. *Sports Med. Open* **2020**, *6*, 47. [[CrossRef](#)] [[PubMed](#)]
195. Adams, J.A.; Lopez, J.R.; Nadkarni, V.; Zolkipli-Cunningham, Z.; Ischiropoulos, H.; Sackner, M.A. The effects of a motorized passive simulated jogging device on descent of the arterial pulse waveform diastolic notch: A single arm placebo-controlled cross-over trial. *Physiol. Rep.* **2022**, *10*, e15418. [[CrossRef](#)] [[PubMed](#)]
196. Hoffmann, A.K.; Gill, H. A study to determine chest wall vibratory attachment interface locations for a low frequency sonic vibrator in treatment of acute coronary thrombosis. *J. Thromb. Thrombolysis* **2011**, *32*, 167–176. [[CrossRef](#)] [[PubMed](#)]
197. Hoffmann, A.; Gill, H. Diastolic timed Vibro-Perfusion at 50 Hz delivered across a chest wall sized meat barrier enhances clot dissolution and remotely administered Streptokinase effectiveness in an in-vitro model of acute coronary thrombosis. *Thromb. J.* **2012**, *10*, 23. [[CrossRef](#)] [[PubMed](#)]
198. Nechypurenko, O.; Kaliuzhka, A.; Lutsenko, O. The effect of vibro-acoustic stimulation on cell dynamics of inflammation in rats and the possibilities for vibroacoustic sound therapy in complex treatment of bronchitis in children. *Georgian Med. News* **2018**, *285*, 107–111.
199. Uryash, A.; Adams, J.A. Wearable Vibroacoustic Transthoracic Stimulation (VATS) Provides Cardioprotection by Syndecan-4 Mechanosensor Regulation of NFAT, JNK/ERK in Rats After Myocardial Infarction. *Circulation* **2017**, *136*, A17906.
200. Skille, O.; Wigram, T.; Weekes, L. Vibroacoustic Therapy: The Therapeutic Effect of Low Frequency Sound on Specific Physical Disorders and Disabilities. *J. Br. Music Ther.* **1989**, *3*, 6–10. [[CrossRef](#)]
201. Uryash, A.; Gill, H.; Hoffmann, A. Can Upper Torso Vibro Acoustic Stimulation Treat No Reflow Following STEMI-Directed PPCI? Rationale and Literature Review. *CathLab Digest* **2016**, *29*.
202. Yohannes, F.G.; Hoffmann, A.K. Non-invasive low frequency vibration as a potential emergency adjunctive treatment for heart attack and stroke. An in vitro flow model. *J. Thromb. Thrombolysis* **2007**, *25*, 251–258. [[CrossRef](#)] [[PubMed](#)]
203. Bekniyazova, A.Z.; Kadralinova, A.; Konkayeva, M.E.; Yeltayeva, A.A.; Konkayev, A.K. Case Report: Complex Treatment Using Vibroacoustic Therapy in a Patient with Co-Infection and COVID-19. *Front. Med.* **2022**, *9*, 893306. [[CrossRef](#)] [[PubMed](#)]
204. Trinity, J.D.; Groot, H.J.; Layec, G.; Rossman, M.J.; Ives, S.J.; Morgan, D.E.; Gmelch, B.S.; Bledsoe, A.; Richardson, R.S. Passive leg movement and nitric oxide-mediated vascular function: The impact of age. *Am. J. Physiol. Circ. Physiol.* **2015**, *308*, H672–H679. [[CrossRef](#)] [[PubMed](#)]
205. Hosseini, Z.-S.; Peyrovi, H.; Gohari, M. The Effect of Early Passive Range of Motion Exercise on Motor Function of People with Stroke: A Randomized Controlled Trial. *J. Caring Sci.* **2019**, *8*, 39–44. [[CrossRef](#)] [[PubMed](#)]
206. Wong, J.K.S.; Wu, C.-J.; Lin, Y.-Y.; Lee, S.-D. Acute Effects of 160-Degree V-Shape Whole-Body Periodic Acceleration (WBPA) on Blood Pressure and Cardiovascular Hemodynamics. *Appl. Sci.* **2022**, *12*, 9116. [[CrossRef](#)]