



Amino Acids Biostimulants and Protein Hydrolysates in Agricultural Sciences

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Abstract: The effects of different types of biostimulants on crops include improving the visual quality of the final products, stimulating the immune systems of plants, inducing the biosynthesis of plant defensive biomolecules, removing heavy metals from contaminated soil, improving crop performance, reducing leaching, improving root development and seed germination, inducing tolerance to abiotic and biotic stressors, promoting crop establishment and increasing nutrient-use efficiency. Protein hydrolysates are mixtures of polypeptides and free amino acids resulting from enzymatic and chemical hydrolysis of agro-industrial protein by-products obtained from animal or plant origins, and they are able to alleviate environmental stress effects, improve growth, and promote crop productivity. Amino acids involve various advantages such as increased yield and yield components, increased nutrient assimilation and stress tolerance, and improved yield components and quality characteristics. They are generally achieved through chemical or enzymatic protein hydrolysis, with significant capabilities to influence the synthesis and activity of some enzymes, gene expression, and redox-homeostasis. Increased yield, yield components, and crop quality; improved and regulated oxidation-reduction process, photosynthesis, and physiological activities; decreased negative effects of toxic components; and improved anti-fungal activities of plants are just some of the more important benefits of the application of phenols and phenolic biostimulants. The aim of this manuscript is to survey the impacts of amino acids, different types of protein hydrolysates, phenols, and phenolic biostimulants on different plants by presenting case studies and successful paradigms in several horticultural and agricultural crops.

Keywords: amino acids; biostimulants; medicinal plants; phenols; protein hydrolysates

1. Introduction

Biostimulants are considered bioactive substances that are either inorganic or organic microorganisms that can increase crop performance when utilized in small quantities [1] as they can enhance both performance and growth as well as improve nutrient- and water-use efficiencies of different crops [2–8]. Amino acids have a dual function as building blocks for proteins and as providers of organic nitrogen, which can alleviate the negative impacts of drought and salt stress [9], and promote cell growth. They are vital in metabolite synthesis, growth, and development, and appropriate in plants because of their structure as protein units [10–14]. The positive effects of the foliar application of amino acids and biostimulants based on amino acids on both the qualitative and quantitative characteristics of *Foeniculum vulgare* Mill, *Coriandrum sativum* L., *Achillea millefolium* L., *Nigella sativa* L., *Ocimum basilicum* L., *Urtica pilulifera* L., *Mentha piperita*, *Calendula officinalis* L., and *Satureja hortensis* L. plants have been reported [11–25].

Amino acids used for the production of biostimulants are obtained from the chemical synthesis of plant proteins, such as algae, soybean, and corn, as well as from animal



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Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). proteins by both chemical and enzymatic hydrolysis. Amino acids that have been used for foliar usage are the result of enzymatic hydrolysis from both animal and plant protein hydrolysates, and as it is very energy-consuming, foliar application is a common process in the agricultural industry. Protein hydrolysate is related to the product of the hydrolytic action of protease(s) on a pure protein sample, or a complicated proteinaceous sample [26,27], which is necessarily a mixture of peptides, free amino acids, and probably partially degraded proteins [28,29]. Protein hydrolysates and amino acids, which are also known as protein-based biostimulants, are usually readily available because of the abundance of raw materials and their affordable cost [30–32]. Protein-based biostimulants can usually be obtained from the hydrolysis of protein-rich agro-wastes, which includes chemical, thermal, and enzymatic processes, or a combination of them [33–36]. They are usually considered as a crude peptide mixture, and they are usually used as the initial raw material for bioactivity testing [37–39]. Fish protein hydrolysates are famous in different parts of the world for pharmaceutical, cosmetic, and nutritional usage [40–42].

Several studies have reported that biostimulants promote plant resilience, especially by improving antioxidant activity within the plant under negative environmental conditions [43,44]. It could behave directly on the plant through an adjustment of the nitrogen and carbon metabolisms and the plant hormonal profile, or indirectly through the microbiome [45]. Food-derived bioactive proteins have physiological impacts on major body systems, such as opioid agonists and opioid antagonists on the nervous system; anti-hyperlipidemic, anti-thrombotic, anti-oxidative, and anti-hypertensive effects on the cardiovascular system; cytomodulatory, immunomodulatory, and anti-microbial effects on the immune system; and mineral binding, anti-appetizing, and anti-microbial impacts on the gastrointestinal system [46-50]. Phenols have notable roles in plant development and growth [51–53], as they are products of secondary metabolic procedures and are generally converted from sugars via the pentose phosphate pathway, the manganiferous acid pathway, the glycolytic pathway, or the benzene-propane pathway [54–56]. Phenolic acids include a carboxylic acid group in addition to the basic phenolic structure and are categorized into hydroxybenzoic and hydroxycinnamic acids. Phenolic acids can be utilized to grow crops by soil and foliar application as well as seed treatment, but foliar utilization of phenolic acids is usually suggested. This research examines the scientific literature on biostimulants from 1990 to October 2022 by conducting a bibliometric analysis of the literature published on the Web of Science database, including more than one thousand articles. The goal of this review article is to survey the effects of different biostimulants, such as amino acids, protein hydrolysates, and phenols, by presenting case studies and successful paradigms in different agricultural and horticultural crops. The information provided is obtained from randomized control experiments, review articles, and analytical observations and studies that have been gathered from various literature sources such as PubMed, Science Direct, Scopus, and Google Scholar. The keywords used were the Latin and common names of different agricultural and horticultural species, amino acids, protein hydrolysates, phenols, phenolic biostimulants, and medicinal plants.

2. Amino Acids

Amino acids for the production of biostimulants are derived by chemical synthesis from plant proteins such as soybean, corn, algae, corn, etc., as well as from animal proteins by enzymatic and chemical hydrolysis [57–62]. Amino acids act as vital molecules with various physiological roles [63] and play an important function in seed germination [64,65], and under salinity stress, they can behave as osmolytes, which can promote stomatal opening control, transport regulation, enzyme activation, heavy metals detoxification, redox homeostasis maintenance, and gene expression [66–70]. Supplementing plants with environmentally friendly amino acid biostimulants can decrease the application of inorganic fertilizers [71,72].

Amino acids are also important in the agriculture industry as chelates of metal ions and microelements chelated with amino acids from very small, electrically neutral molecules

increase their transport and absorption within the plant [73–75]. Some of the most important products in the market which contain amino acids are Delfan Plus (Tradecorp, Madrid, Spain), Natural Crop SL (Natural Crop Poland Sp. Z o.o., Warsaw, Poland), Bosfoliar Activ (COMPO EXPERT, Munster, Germany), Amino Quelant Ca (Bioiberica, Barcelona, Spain), Tecamin Max, Tecamin Brix, Tecnokel Amino Mix, Terra-Sorb Foliar (Agritecno Fertilizants, Valencia, Spain), Agrocean B (Agrimer, Plouguerneau, France), Metalosate Calcium and Metalosate Fe (Albion Minerals, Layton, UT, USA) [76–79]. The usage of amino acids can increase co-enzyme formation and the photosynthesis procedure [80], and supports different plant organisms that may face environmental stresses [81]. It has been also reported that the exogenous utilization of amino acids can enhance nitrogen status, and the contents of mineral elements in plant tissues [82,83]. Depending on environmental conditions and plant species, plants reduce inorganic nitrogen to amino acids in roots, nodules, and leaves [84-86]. Many studies have reported the important and notable effects of the foliar application of concentrations with phenylalanine and tyrosine solutions on essential oil, the total amount of phenols, and their compositions in Ocimum basilicum L., Melissa officinalis L., and Coleus blumei L. plants [87–89]. Phenylalanine is an amino acid [90, 91], and its foliar application can help mustard (Brassica campestris L.) plants overcome drought stress and increase total chlorophyll contents, shoot length, and biological yield [92]. Roman et al. [93] reported that foliar application of methyl jasmonate and phenylalanine can increase the content of volatile compounds in grapes, and Portu et al. [94] introduced it as an important management tool for boosting grape quality. The impacts of different amino acids on several experimental plants are shown in Table 1. The roles of different amino acids as biostimulants are shown in Table 2. The main mechanisms of amino acids biostimulants are shown in Figure 1.

Table 1. The effects of amino acids on different plants.

Plant	Plant Family	Key Point	Reference
Chickpea (Cicer arietinum L.)	Fabaceae	The combined application of amino acids of commercial compounds with proline + valine, and proline + alanine can reduce the negative impacts of drought stress on chickpeas.	[95]
Cowpea (Vigna unguiculata)	Fabaceae	The foliar application of amino acid liquid fertilizer and liquid biological fertilizer can enhance crop yield.	[96]
Grapevine (<i>Vitis vinifera</i> L.)	Vitaceae	A biostimulant that contains amino acids can enhance the growth of the microbial community on berry skin.	[97]
Lettuce (<i>Lactuca sativa</i> L.)	Asteraceae	The foliar utilization of amino acids biostimulants (Perfectose TM , liquid) can improve the nutritive value and yield of lettuce.	[98]
		An amino-acid-based Phytostim [®] biostimulant can improve growth and yield attributes.	[99]
		The biostimulant Codasil [®] , which is composed of amino acids, can improve lettuce physiology and growth, and enhance the crop resistance to water stress.	[100]
		The application of proline and methionine increased proteinogenic amino acid expression.	[101]
		Terra Sorb [®] radicular and Terramin [®] Pro, which contain high amino-acid content, are useful biostimulants for plant development in nitrogen-limiting areas.	[102]
Mint (<i>Mentha × Piperita</i> L.)	Lamiaceae	The application of phenylalanine at 100 mg L^{-1} concentration enhanced the essential oil.	[103]
Moldavian balm (Dracocephalum moldavica L.)	Lamiaceae	Leaf spraying of biostimulants based on amino acids can notably mitigate the adverse impact of salinity stress on the growth and physiological growth of plants.	[104]
Strawberry (Fragaria × ananassa)	Rosaceae	The combined application of humic acids and amino acids can improve strawberry nutritional traits such as phenolic compounds, and commercial characteristics such as external color and firmness.	[105]

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Plant	Plant Family	Key Point	Reference
Sunflower (<i>Helianthus annuus</i> L.)	Asteraceae	Plant biostimulant Amino Expert [®] Impuls can increase sunflower plant height, head diameter, seed yield, seed oil, and absolute seed mass.	[106]
Tomato (Solanum lycopersicum L.)	Solanaceae	The combined application of amino acids and humic acids could significantly influence yield in conventional nutrition.	[107]
		The combined application of amino acids and humic acids could positively improve total antioxidant capacity, total flavonoid content, and total phenol content.	[108]
		Biostimulants that contain anino acids could increase the accumulation of plant biomass as well as improve the tolerance of plants to water deficit.	[109]
		soluble sugars.	[110]
Olive (Olea europaea L.)	Oleaceae	The complex of natural amino acids, such as hydroxyproline, proline, and glycine, can induce higher stomatal conductance and leaf photosynthetic rates.	[111]
_		The combined application of amino acids, fulvic acid, and humic acid can significantly increase the quality and the oil content of olives.	[111]
Peanut (Arachis hypogaea L.)	Fabaceae	Foliar utilization of 100 mg/L aspartic acid can increase seed and oil yield.	[112]
Pepper (Capsicum annuum L.)	Solanaceae	Actium [®] , provided by Grupo Agrotecnologia (Alicante, Spain), contains amino acids that could enhance carotenoids, total monosaccharides, and phenylalanine in plants.	[113]
		The application of biostimulants that contain amino acids can boost the activity of important enzymes such as peroxidase, phenylalanine ammonia lyase, and capsaicin synthase.	[114]
Rice (<i>Oryza sativa</i> L.)	Poaceae	Zinc-enriched amino acids (Zn-AAC) increased salt-stressed rice yield, chlorophyll content, and quality of rice.	[115]
		The combined application of potassic fertilizer with amino acids can improve both the yield and quality of rice.	[116]
Soybean (<i>Glycine max</i> L.)	Fabaceae	Amino acid application can increase plant height, the number of seeds and pods, flavonoid content, and phenolic content.	[117]
-		The application of phenylalanine and cysteine could enhance the production of soybean plants by at least 21%.	[118]
Spinach (<i>Spinacia oleracea</i> L.)	Amaranthaceae	The application of different amino acid treatments such as tyrosine, methionine, proline, and phenylalanine could increase dry and fresh weight, shoot length, root length, leaf area, and final yield.	[119]
Weeping alkaligrass (Puccinellia distans)	Poaceae	Two biostimulants, namely Bonamid [®] at 2 g/L, and Algabon [®] at 0.5 g/L, which contained amino acids could positively increase K ⁺ content, chlorophyll content, K ⁺ /Na ⁺ ratio, leaf relative water content, and biomass as well as reduce the adverse effects of NaCl-caused stress in vacuoles.	[120]
Winter wheat (<i>Triticum aestivum</i> L.)	Poaceae	INTERMAG Co. (Olkusz, Poland)—AminoHort and AminoPrim, containing 20% and 15% amino acids at 1.25 L/ha and 1.0 L/ha, could significantly increase nutrient contents such as molybdenum, calcium, sodium, and copper in grains.	[121]
		The application of amino acids together with yeast extract can significantly boost physiological yield and traits.	[122]

Table 1. Cont.

Amino Acids	Function	References
Deflan Plus	Increase co-enzyme formation; improve photosynthesis procedure; increase	[76-81]
	resistant of plants to environmental stresses	
Natural Crop SL	resistant of plants to environmental stresses	[76-81]
Tecomin Max	Increase co-enzyme formation; improve photosynthesis procedure; increase	[76 91]
recarring wax	resistant of plants to environmental stresses	[/0-01]
Tecamin Brix	Increase co-enzyme formation; improve photosynthesis procedure; increase	[76-81]
	resistant of plants to environmental stresses	[, , , ,]
Agrocean B	Increase co-enzyme formation; improve photosynthesis procedure; increase	[76-81]
Ũ	resistant of plants to environmental stresses	
Metalosate Calcium	increase co-enzyme formation; improve photosynthesis procedure; increase	[76-81]
	Increase co-enzyme formation: improve photosynthesis procedure: increase	
Metalosate Fe	resistant of plants to environmental stresses	[76-81]
Phenylalanine and tyrosine solutions	Improve essential oil, and increase the total amount of phenols	[87–89]
Phenylalanine	An important amino acid that can enhance shoot length, biological yield, and total chlorophyll contents	[90-93]
Methyl jasmonate	Enhance the content of volatile components	[93,94]
Proline, Valine, Alanine	They can reduce the adverse effects of drought stress	[95]
Perfectose TM	It can increase the yield and nutritive value of plants	[98]
Phytostim [®]	It can increase final yield and growth	[99]
Codasil [®]	It can increase resistance to drought stress	[100]
Sorb [®] , Radicular, Terramin [®]	They are appropriate to improve the yield of plants in nitrogen-limiting regions	[102]
Amino Expert [®]	It may increase yield and yield components	[106]
Actium®	It can increase carotenoids and quality parameters	[113]
Bonamid [®] , Algabon [®]	They can increase chlorophyll content, biomass, and leaf relative water content	[120]
INTERMAG Co. (Olkusz,		
Poland)—AminoHort, and	It can improve mineral components in plants	[121]
AminoPrim		

Tuble 2. The foles of anterent antino actas as biostinitation	Table 2.	The roles of	different	amino aci	ids as	biostimulan	ts
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Figure 1. The most important mechanisms of amino acids biostimulants.

3. Protein Hydrolysates

Protein hydrolysates, especially those that contain antioxidant peptides, are obtained from natural components, and many researchers and scholars consider them biostimulants because of their minimum side effects, easy absorption, low cost, high activity, and lower molecular weight [123–130]. Protein hydrolysates and peptides can be used as notable in-

gredients in the formulation of functional foods [131–141]. They can be used as foliar sprays or through drip irrigation systems, and the amino acids can be absorbed through both leaves and roots [142–144]. Their utilization can significantly affect nitrogen metabolism in plants, and boost productivity, particularly when applied as a seed pre-treatment [144]. For separating the amino acids in protein hydrolysates, a liquid chromatography process can be used [145,146]. Numerous methods have been considered to produce hydrolysates from fish and fish by-products such as thermal hydrolysis, autolysis, chemical hydrolysis, and enzymatic hydrolysis [146,147]. The basic procedures utilized following hydrolysis of protein are heat inactivation, which has a function in the inactivation of proteolytic enzymes; ultrafiltration, which is important in the removal of high molecular weight peptides and proteins; use of specific enzymes, which can reduce the content of specific amino acids; hydrolysis by exoproteases, which is active in hydrolysis and the reduction of bitterness; carob activation, which has a notable role in the reduction of bitterness; and absorption chromatography, which can decrease the content of aromatic amino acids. Microbial-based biostimulants such as Environoc 401®, Bioyield®, Rootshield Plus⁺ WP[®], Spectrum + Myco[®], Select[®], and Endomaxx[®] inconsistently increased the quality of bell pepper (*Capsicum annuum* L.) in a greenhouse experiment [148]. Ghorbel-Bellaaj et al. [149] reported that five proteolytic enzymes, namely Alcalase[®], trypsin, a crude enzyme extract from sardinelle (Sardinella aurita) viscera, and an enzyme preparation from Aspergillus clavatus ESA and Bacillus licheniformis NH1, which are protein hydrolysates, were obtained from shrimp via by-products processing, and they have revealed notable degrees of antioxidant activities, such as β -carotene bleaching, reducing power, and 1,1-diphenyl-2-picrylhydrazyl (DPPH)-scavenging activity assays, which can be a promising and helpful alternative for accessible commercial nitrogen sources from other origins. It can be a good source for microbial growth and protease production by Saccharomyces cerevisiae, Escherichia coli, Bacillus subtilis A26, and Bacillus mojavensis A21.

Some of the available plant biostimulants, their composition, and application strategies are C Fish, which contain peptides and amino acids that are used on vegetables and fruits to increase the plant's resistance to insect pressure, disease and drought or heat stress which originates from white fish/mixed fish composition autolysates and hydrolysates in fruits and vegetables; Radifarm, which contains peptides, amino acids, betaines, saponins, vitamins, polysaccharides, and microelements, has been used to promote the formation of an extensive root system by speeding up the elongation of adventitious and lateral roots of vegetables and fruits; Megafol, which contains betaines, amino acids, auxin, vitamins, proteins, cytokine, and gibberellin, can improve the balance between vegetative productivity and development as well as plant resistance to stressors such as hail, weeding, root asphyxia, and frost; Biozyme, which includes plant hormones, algae extract, and chelated micronutrients, can boost nutrient uptake, photosynthesis, and the activity of chlorophyll of legumes, vegetables and fruits; BioRoot, which contains humates, plant and mineral-derived organic acids, enhances rooting ability, protein content, and chlorophyll of fruits and vegetables; Grow-plex SP, which contain humic acids, can increase soil bacteria, shoot and root growth, and zinc and iron uptake of vegetables and fruits; Ergonfil, which has cysteine, animal protein hydrolysates, keratin derivatives, and folic acid, can promote chlorophyll synthesis and indole acetic acid, increase chelation, and improve translocation in fruits and vegetables; Benefit, which contains nucleotides, amino acids, vitamins, free enzymatic proteins, can improve cell division and increase the number of cells per fruit [150–153]. Animal-derived gelatin, which has peptides and amino acids, can improve shoot dry weight and promote root nitrogen assimilation in broccoli, arugula, tomato, pepper, cucumber, and field corn [154]. There are notable reports and evidence that the application of non-structural and structural amino acids, such as histidine, proline, taurine, and glutamate, can provide protection to the plant from environmental stresses or play an important function in metabolic signaling by regulating nitrogen acquisition by the roots [155,156]. Amino acids can act as osmoprotectants, which stabilize membranes, enzymes, and proteins against denaturing caused by high salt components and

non-physiological temperatures [157]; moreover, arginine has been proven to have an important function in nitrogen transport and storage in plants during biotic and abiotic stress conditions [158]. Amino acids can also reduce plant toxicity by heavy metals by acting as metal chelators [159,160]. Rouphael et al. [161] reported that the application of vegetal-protein hydrolysates based microgranules can increase carotenoids and total chlorophyll content. Protein hydrolysate has a positive influence on total root area and on root length, which can increase mineral-nutrient and water-use efficiency as well as promote plant productivity and resistance to harmful conditions [162–164]. It can also positively influence the leaf area and yield of horticultural plants and fruit trees [165,166]. The exogenous utilization of protein hydrolysate and isolated amino acids can promote plant antioxidant performance by improving the non-enzymatic and enzymatic antioxidant defense machinery of the cell [167]. The most important effects of different kinds of protein hydrolysates have been shown in Table 3.

Table 3. The impacts of different protein hydrolysates on various plants.

Plant	Plant Family	Protein Hydrolysate	Key Point	Reference
Apple (Malus domestica)	Rosaceae	Alfalfa protein hydrolysate	It can improve sensorial characteristics and fruit quality. Promote nutraceutical value, and decrease post-harvest disease.	[168]
Banana (Musa acuminata)	Musaceae	Chicken feathers hydrolysate	Promote chlorophyll content and increase photosynthetic. Increase fruit yield, filling, and set as well as antioxidants and decrease time to flowering	[169]
Basil (Ocimum basilicum)	Lamiaceae	Protein hydrolysate	It can decrease nitrate leaf content, and enhance basil resilience.	[170]
Castor (<i>Ricinus communis</i>)	Euphorbiaceae	Soybean protein hydrolysate (SPH)	It could lead to a significant increase in castor husks and final yield.	[171]
Celery (Apium graveolens L.)	Apiaceae	Protein hydrolysates	It can boost the total phenolic content in plants.	[172]
Chickpea (Cicer arietinum L.)	Fabaceae	Chicken feathers hydrolysate	Increases secondary roots and biomass production, and reveals phytohormone-like activities.	[173]
Common bean (Phaseolus vulgaris L.)	Fabaceae	Pumpkin seed protein hydrolysate	Application of 2000 μ L L ⁻¹ to obtain appropriate yield and growth of plants under salt stress.	[174]
Coriander (Coriandrum sativum)	Apiaceae	Commercial amino acid preparation	It has glycine. which can improve the growth of shoots and leaves and increase the micronutrient content of leaves.	[175]
Florist's daisy (Chrysanthemum morifolium)	Asteraceae	Two plant protein hydrolysates (Trainer [®] , and Vegamin©), and one animal protein hydrolysate (Hicure [®])	Plant protein hydrolysates could decrease nitrate concentration in flowers and leaves. Animal protein hydrolysate caused a faster duration of flower stems to wilt stage.	[176]
Grape tomatoes (Solanum lycopersicum var. cerasiforme)	Solanaceae	Fish-derived protein hydrolysates	Application of fish-derived protein hydrolysates could reduce the negative impacts of drought, and improve total plant biomass yield, leaf dry weight, and fruit number.	[177]
Grapevine (Vitis vinifera L.)	Vitaceae	Protein hydrolysates Trainer and Stimtide	Both of them induced alterations in leaf metabolome and proteome, which can delay physiological maturity and keep higher acidity.	[178]

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Table 3. Cont.

Plant	Plant Family	Protein Hydrolysate	Key Point	Reference
		Animal- and plant-derived protein hydrolysates, namely lupin, soybean, and casein	It can increase fruit parameters and alleviate the adverse effects of water stress.	[179]
Hemp (Cannabis sativa L.)	Cannabaceae	A commercial legume-derived protein hydrolysate	It can increase seed yield and improve fiber production.	[180]
		Fish hydrolysate, <i>Aloe</i> <i>vera,</i> and Kelp	It can increase branching, root growth, and propagation effectiveness as well as improve potassium and phosphorous uptake.	[181]
Hibiscus (<i>Hibiscus moscheutos</i> L. subsp. <i>palustris</i>)	Malvaceae	Protein hydrolysates from biowaste as biostimulants	It could improve leaf gaseous exchanges, biometric parameters, nitrogen-use efficiency, and biomass accumulation.	[182]
Kiwifruit (Actinidia deliciosa)	Actinidiaceae	Gelatin hydrolysate	Increase root and shoot biomass. Boost metabolism and assimilation of nitrogen.	[183]
Lettuce (Lactuca sativa L.)		Fish-derived protein hydrolysate	It contains amino acids and peptides, which can improve root biomass and leaf number and enhance photosynthetic rate and chlorophyll content.	[184]
Lettuce (<i>Lactuca sativa</i> L.)	Asteraceae	Commercial amino acids preparation	It has glutamine and glycine, which can enhance vitamin C content, leaf chlorophyll, and yield.	[185]
		Protein hydrolysates	Application of Molybdenum dosage together with protein hydrolysates can increase yield, nutritional, morphology, and functional features.	[186]
		Soy protein hydrolysate	Application of 0.01 mg/mL protein hydrolysate can promote lettuce weight and length.	[187]
		A <i>Graminaceae</i> -derived protein hydrolysate	It can improve the growth and yield of plants and improve the resistance of plants under mild salinity conditions.	[188]
		Plant-derived protein hydrolysates	It can improve root dry weight and dry biomass and increase fresh yield. Its application can improve anthocyanins and	[189]
		Protein hydrolysate derived from pig blood	flavonoids as well as root and shoot fresh weight.	[190]
Maize (Zea mays L.)	Poaceae	Soybean protein hydrolysate (SPH)	Application of fertilizer with SPH can increase one thousand grain weight, the grain number per ear, and total yield.	[188,189]
		A solid biostimulant (AA309) derived through thermobaric hydrolysis applied on trimmings and shavings of bovine hides tanned with wet-blue technology	It can improve the yield of crops. It can influence plant physiology because of changes they can induce in plant-associated microbes [,] composition and activity.	[190]
		hydrolysate-based biostimulant	It can promote root and shoot growth and increase lipid peroxidation.	[191]
		Meat flour protein hydrolysate	It contains amino acids and peptides, which can improve leaf and root biomass and promote effective nutrient utilization by plants.	[192]

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Plant	Plant Family	Protein Hydrolysate	Key Point	Reference
		Chicken feather hydrolysates	Amino acids and peptides can increase macronutrient and micronutrient concentrations of leaves and grain protein content.	[193]
		A novel biostimulant (APR [®] , ILSA S.p.A., Arzigano VI, Italy)	It can influence shoot and root growth and improve the resistance of plants to various stresses.	[194]
		A commercial collagen-derived protein hydrolysate	It can stimulate lateral root growth and final yield.	[195]
Melon (Cucumis melo L.)	Cucurbitaceae	Fish protein hydrolysates	It can promote the activity of sucrose phosphate synthase. It can improve fructose and glucose contents by increasing the activity of acid invertase. It can boost the synthesis direction of sucrose synthase.	[196]
Oregano (Origanum vulgare L.)	Lamiaceae	Fish protein hydrolysates at 1000 mg/L	It can prevent vitrification in oregano shoot clones regenerated from axillary bud explants. Fish protein hydrolysate-treated shoots can decrease elongation and induce higher chlorophyll content.	[197]
Passion fruit (Passiflora Edulis)	Passifloraceae	Commercial preparation of peptides and amino acids	It has peptides and amino acids, which can increase the photosynthetic process in plants and increase transplanting success.	[198]
Pea (Pisum sativum L.)	Fabaceae	Papain and pepsin-hydrolyzed whey protein	Application of 2000 mg/L of biostimulant can increase pod length, pod growth, and the number of seeds per pod.	[199]
Peppermint (<i>Mentha × piperita</i> L.)	Lamiaceae	Amino16 [®] , a commercial protein hydrolysate	It could not impact dry or fresh weight; however, it decreased plant height. It promoted total soluble phenol and total antioxidant capacity.	[200]
Persimmon (Diospyros kaki)	Ebenaceae	Protein hydrolysate	Increases the biosynthesis of salt stress response proteins	[201]
Rapeseed (Brassica napus subsp. napus)	Brassicaceae	Soybean protein hydrolysate (SPH)	It improved yield and promote the growth of plants.	[202]
Rice (Oryza sativa L.)	Poaceae	Soybean protein hydrolysates	It can decrease long- and short-term retrogradation of gelatinized rice starch.	[203]
Sea grape (Coccoloba uvifera L.)	Polygonaceae	Jackfruit (<i>Artocarpus</i> <i>heteropyllus</i> L.) leaf protein hydrolysates	It has shown emulsifying properties, and it could be used as an alternative to conventional emulsifiers.	[204]
Snapdragon (Antirrhinum majus L.)	Plantaginaceae	Protein hydrolysates	The combined application of protein hydrolysates, humic acids, and seaweed extracts could increase the number of leaves and improve the performance of ornamental plants.	[205]
Soybean (<i>Glycine max</i> L.)	Fabaceae	Protein hydrolysates	It can improve the final yield of plants.	[206]
Spearmint (Mentha spicata L.)	Lamiaceae	Amino16 [®] , a commercial protein hydrolysate Xaell Poort a mixture of	It could increase the quality of spearmint without negative impacts on crop yield.	[207]
Spinach (Spinacia oleracea L.)	Amaranthaceae	fish protein hydrolysates and kelp extract	It is highly beneficial for promoting the tolerance of spinach to water shortage stress.	[208]

Table 3. Cont.

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Plant	Plant Family	Protein Hydrolysate	Key Point	Reference
		Trainer [®] , a plant-derived protein hydrolysates	It can increase total amino acid content, but reduce polyphenol content and increase final yield.	[209]
Sweet basıl (Ocimum basilicum L.)	Lamiaceae	Animal-derived protein hydrolysate	It can decrease plant growth, photosynthetic performance, and yield.	[210]
Sugar beet (Beta vulgaris)	Amaranthaceae	Protein-based biostimulants (PBBs)	The application of 2 g/kg soil PBBs increased protein-related characteristics in samples and induced higher photosynthesis, growth, and quality of plants.	[211]
		Hydrolyzed wheat gluten and potato protein	It can enhance final yield and plant growth.	[212]
Sweet cherry (Prunus avium L.)	Rosaceae	An organic fertilizer (Defender Ca; Kenya Biologics Ltd., Runyenjes, Kenya)	It can improve fruit yield, soluble solids content, and calcium concentration in fruits.	[213]
Sweet pepper (<i>Capsicum annuum</i> L.)	Solanaceae	Organic fertilizer based on hydrolyzed proteins	It could improve the performance in nitrogen uptake, increase resistance to tolerance, and mitigate the negative impacts of toxic elements	[214]
Sweet potato (Ipomoea batatas L.)	Convolvulaceae	Whey protein hydrolysates (WPH)	Foliar application of WPH at 0.10 and 0.20% could improve uptake of K, P, and N by shoots, shoot dry weight per plant, final yield, marketable yield, and total yield.	[215]
Tea (Camellia sinensis)	Theaceae	Chicken feather protein hydrolysate	It can be applied as a growth booster for gaining higher yields.	[216]
Tomato (Solanum lycopersicum L.)	Solanaceae	CycoFlow, Agriges, BN, Italy, a novel protein hydrolysate-based biostimulant	It can induce better pollen viability and water status as well as improve antioxidant contents in fruits and leaves.	[217]
		Soy protein hydrolysates (SPH13 and SPH18 at 10 g L^{-1})	It can notably improve plant resistance to foliar inoculation with <i>Pseudomonas syringae</i> pv. tomato DC3000.	[218]
		Protein hydrolysates	Its usage can stimulate plant growth.	[219]
		Protein hydrolysates	Its application could enhance fruit antioxidants such as ascorbic acid levels, polyphenols, and lycopene	[220]
		Plant-derived protein hydrolysates	It can enhance nitrogen use and uptake as well as tomato yield.	[221]
		Protein hydrolysates	biostimulant to improve plant resilience to abiotic stresses.	[222]
		<i>Arthrospira platensis</i> protein hydrolyzate	Its application as 68.9 mg mL ⁻ free amino acids can improve plant yield and growth.	[223]
		The pig blood-derived protein hydrolysate	It can increase salt tolerance in tomatoes and improve photosynthetic efficiency, chlorophyll levels, and plant growth.	[224]
		Pig blood-derived protein hydrolysate	impacts of drought stress by regulating chloroplast ultrastructure, antioxidant systems, stomatal aperture, and osmotic changes.	[225]
		Legume-derived protein hydrolysate	Its application at 5.0 mL L^{-1} improved mineral composition, total soluble solids, and antioxidant activities.	[226]

Table 3. Cont.

Plant	Plant Family	Protein Hydrolysate	Key Point	Reference
		An enzymatically hydrolyzed animal protein-based biostimulant (Pepton)	It can show a positive impact, increasing the lateral and primary growth of tomato plants.	[227]
Wall rocket (Diplotaxis tenuifolia (L.) DC.)	Brassicaceae	Legume-derived protein hydrolysates and <i>Trichoderma</i> <i>harzianum</i> T22; Protein hydrolysates + <i>Trichoderma harzianum</i> T22	They can boost the hydrophilic and lipophilic antioxidant activity.	[228]
Wheat (Triticum aestivum L.)	Poaceae	AGROMOREE, a biostimulant based on a protein hydrolysate rainbow trout (Oncorhynchus mykiss)	It can increase gluten content, seed protein, and final productivity, and reduce the use of nitrogen fertilizers.	[229]
		Protein hydrolysate	It can improve wheat grain seed germination and improve final production.	[230]
		Papain-produced whey protein hydrolysates	It can improve spike number, flag leaf area, and grain yield.	[231]
White mustard (<i>Sinapis alba</i> L.)	Brassicaceae	Protein hydrolysate (Hemozym)	It can significantly increase the physicochemical properties and microbial activity of the soil.	[232]

Table 3. Cont.

4. Phenols and Phenolic Biostimulants

Phenols are a major type of antioxidant phytochemical, which have significant importance because of their free radical scavenging and biological characteristics [233–236]. Phenolic compounds are the most abundant secondary metabolites in many plants which are usually discovered in the cell walls of subepidermal and in the vacuoles of epidermal cells [237,238]. Endogenous phenolic components in plants have different functions, which can be used by plants to defend themselves against pathogens, herbivores, and weeds. They are implicated in seed germination and dormancy, appropriate as screens against damaging UV radiation, and act as pigments to attract seed dispersal agents and pollinators [239–241]. The function of phenolic acids as signaling molecules in plant-microbe symbioses has been reported in previous research [242]. Some of the most important phenolic compounds with bioprotectant activities are ferulic acid, curcumin, ellagic acid, catechol, gallic acid, coumarin, caffeic acid, catechin, quercetin, sinapic acid, rutin, resveratrol, salicylic acid, and syringic acid [243,244]. The accumulation of phenolic compounds and the subsequent production of quinones in turnip (Brassica rapa L.) may happen when plants are susceptible to Boron deficiency [245]. Phenolic compound concentration can be important in the biochemical pathway of toxigenic fungal species because of the induction of stress via sublethal contents and depletion of the phenolic compounds [246]. Phenolics have meaningful functions in plant development, especially in pigment and lignin biosynthesis as well as considerable roles in plant protection against stress. It has been reported the correlation between antifungal activity and total phenolics of plants [247] and the accumulation of amino acids and phenolics may boost tolerance to both copper and cobalt stress in barley [248]. Silva et al. [249] reported that tyrosol, which is a phenolic compound from olive oil and several endophytic fungi such as Phomopsis sp., can be used as an important biostimulant in soybean seed treatment, which can alter soybean plant metabolism without meaningful impacts on crop yield. Masondo et al. [250] reported that two phenolic biostimulants, namely eckol and phloroglucinol, isolated from brown algae Ecklonia maxima can have a significant effect on the phytochemical and growth of Eucomis autumnalis. While the phenolic acid metabolism in *Kandelia obovata* may decrease the negative impacts of cadmium and zinc [251], it has been reported that the phenolic compounds of leave extracts of Calligonum

arich L. are effectual against pathogenic bacteria [252], and the phenolic compounds of apricot branches have shown antifungal activity against *Monilinia laxa* growth [253–255]. One of the notable impacts of phenolics is to improve the resistance of *Nicotiana langsdorffii* to Cr(VI) [256–259].

5. Conclusions and Future Prospects

The innovative agronomic tools of agriculture are biostimulants, which are composed of inorganic and organic substances that consist of several microorganisms and substances. Biostimulants can do various agronomic functions such as increasing the growth and development of plants during their entire life cycle; promoting the resistance of plants to abiotic stresses such as cold, heat, and lack of water; improving soil fertility, especially increasing the development of soil microorganisms; promoting the use efficiency of nutrients by plants; and finally, increasing yield and crop quality. They can also be used as the best alternative for chemical fertilizers and are the best strategy for promoting organic agriculture. Amino acids are appropriate candidates to boost stress tolerance through metal chelation, nutrient availability, osmo-protection, and reactive oxygen species (ROS), which can notably affect the synthesis and stimulation of gene expression and some enzymes. Amino acids are organic components, which contain amine and carboxyl C(=O)OH) functional groups together with a side chain (R group). They can promote and stimulate the process of protein synthesis and photosynthesis; promote nutrient assimilation, translocation, and utilization; and strengthen plant growth and yield formation. Protein hydrolysates are manufactured from plant-derived protein sources using partial thermal hydrolysis, chemical hydrolysis, and enzymatic hydrolysis. Different sources of protein hydrolysates on the basis of protein sources are animal origin, leather by-products, blood meal, fish by-products, chicken feathers, casein, plant origin, legume seeds, alfalfa hay, and vegetable by-products. The positive impacts of the utilization of amino acids have been discovered; however, there is not enough knowledge about the effects of each amino acid on both the physiological and metabolic processes of plants. A better understanding of biostimulants, such as amino acids, protein hydrolysates, phenols, and phenolic biostimulants, while considering their various effects on different functions of crops, namely crop yield and yield components, growth promotion, and nutrient availability, may help agricultural scientists and farmers to better understanding and utilization of them.

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References

- 1. La Bella, S.; Consentino, B.B.; Rouphael, Y.; Ntatsi, G.; De Pasquale, C.; Iapichino, G.; Sabatino, L. Impact of *Ecklonia maxima* seaweed extract and Mo foliar treatments on biofrotification, spinach yield, quality and NUE. *Plants* **2021**, *10*, 1139. [CrossRef]
- 2. Mashamaite, C.V.; Ngcobo, B.L.; Manyevere, A.; Bertling, I.; Fawole, O.A. Assessing the usefulness of *Moringa oleifera* leaf extract as a biostimulant to supplement synthetic fertilizers: A review. *Plants* **2022**, *11*, 2214. [CrossRef]
- 3. Sun, W.; Shahrajabian, M.H.; Lin, M. Research progress of fermented functional foods and protein factory-microbial fermentation technology. *Fermentation* 2022, *8*, 688. [CrossRef]
- 4. Sun, W.; Shahrajabian, M.H. The application of arbuscular mycorrhizal fungi as microbial biostimulant, sustainable approaches in modern agriculture. *Plants* **2023**, *12*, 3101. [CrossRef]
- 5. Sun, W.; Shahrajabian, M.H. Therapeutic potential of phenolic compounds in medicinal plants-natural health products for human health. *Molecules* **2023**, *28*, 1845. [CrossRef]
- 6. Sun, W.; Shahrajabian, M.H.; Petropoulos, S.A.; Shahrajabian, N. Developing sustainable agriculture systems in medicinal and aromatic plant production by using chitosan and chitin-based biostimulants. *Plants* **2023**, *12*, 2469. [CrossRef]

- Rodriguez-Morgado, B.; Gomez, I.; Parrado, J.; Garcia-Martinez, A.M.; Aragon, C.; Tejada, M. Obtaining edaphic biostimulants/biofertilizers from different sewage sludges. Effects on soil biological properties. *Environ. Technol.* 2015, *36*, 2217–2226. [CrossRef]
- 8. Orts, A.; Tejada, M.; Parrado, J.; Paneque, P.; Garcia, C.; Hernandez, T.; Gomez-Parrales, I. Production of biostimulants from okara through enzymatic hydrolysis and fermentation with *Bacillus licheniformis*: Comparative effect on soil biological properties. *Environ. Technol.* **2019**, *40*, 2073–2084. [CrossRef]
- 9. Abdelkader, M.M.; Gaplaev, M.S.; Terekbaev, A.A.; Puchkov, M.Y. The influence of biostimulants on tomato plants cultivated under hydroponic systems. *J. Hortic. Res.* 2021, 29, 107–116. [CrossRef]
- 10. Lonnerdal, B. Dietary factors influencing zinc absorption. J. Nutr. 2000, 130, 1378S–1383S. [CrossRef]
- 11. Alcazar, R.; Altabella, T.; Marco, F.; Bortolotti, C.; Reymond, M.; Koncz, C.; Carrasco, P.; Tiburcio, A.F. Polyamines: Molecules with regulatory functions in plant abiotic stress tolerance. *Planta* **2010**, *231*, 1237–1249. [CrossRef]
- 12. Shahrajabian, M.H.; Sun, W. Various techniques for molecular and rapid detection of infectious and epidemic diseases. *Lett. Org. Chem.* 2023, *20*, 779–801. [CrossRef]
- 13. Shahrajabian, M.H.; Sun, W. Survey on multi-omics, and multi-omics data analysis, integration and application. *Curr. Pharm. Anal.* **2023**, *19*, 267–281. [CrossRef]
- 14. Shahrajabian, M.H.; Petropoulos, S.A.; Sun, W. Survey of the influences of microbial biostimulants on horticultural crops: Case studies and successful paradigms. *Horticulturae* **2023**, *9*, 193. [CrossRef]
- 15. Aghaye Noroozlo, Y.; Souri, M.K.; Delshad, M. Stimulation effects of foliar application of glycine and glutamine amino acids on growth and quality of sweet basil. *Adv. Hortic. Sci.* **2019**, *33*, 495–501. [CrossRef]
- 16. Ayyat, A.M.; Kenawy, A.G.M.; Aboel-Ainin, M.A.; Abdel-Mola, M.A.M. Improving growth, productivity and oil yield of *Nigella sativa* L. Plants by foliar spraying with some stimulants. *J. Plant Prod.* **2021**, *12*, 339–344. [CrossRef]
- 17. Elsayed, A.A.A.; El-Gohary, A.E.; Khalid, K.A.; Ahmed, A.M.A. Changes in bitter fennel essential oils exposed to foliar spray with L-phenylalanine. *Egypt. J. Bot.* **2022**, *62*, 241–253. [CrossRef]
- Rafiee, H.; Mehrafarin, A.; Qaderi, A.; Jari, S.K.; Badi, H.N. Phytochemical, agronomical and morphological response of pot marigold (*Calendula officinalis* L.) to foliar application of biostimulators (Bioactive amino acid compounds). *J. Med. Plants* 2013, 12, 48–61.
- 19. Mehrabi, S.; Mehrafarin, A.; Badi, H.N. Clarifying the role of methanol and amino acids application on savory (*Satureja hortensis* L.). *Ann. Biol. Res.* **2013**, *4*, 190–195.
- 20. Hendawy, S.F.; Hussein, M.S.; El-Gohary, A.E.; Ibrahim, M.E. Effect of foliar organic fertilization on the growth, yield and oil content of *Mentha piperita* var. citrata. *Asian J. Agric. Res.* **2015**, *9*, 237–248. [CrossRef]
- Shafie, F.; Bayat, H.; Aminifard, M.H.; Saeid Daghighi, S. Biostimulant effects of seaweed extract and amino acids on growth, antioxidants, and nutrient content of Yarrow (*Achillea millefolium* L.) in the field and greenhouse conditions. *Commun. Soil Sci. Plant Anal.* 2021, 52, 964–975. [CrossRef]
- 22. Wafaa, H.A.A.A.; Rania, M.R.K.; El-Shafay, R.M.M. Effect of spraying with extracts of plants and amino acids on growth and productivity on *Coriandrum satioum* plants under Shalateen condition. *Plant Arch.* **2021**, *21*, 300–307. [CrossRef]
- Wahba, H.E.; Motawe, H.M.; Ibrahim, A.Y. Growth and chemical of *Urtica pilulifera* L. plant as influenced by foliar application of some amino acids. J. Mater. Environ. Sci. 2015, 6, 499–506.
- 24. Shahrajabian, M.H.; Sun, W.; Shen, H.; Cheng, Q. Chinese herbal medicine for SARS and SARS-CoV-2 treatment and prevention, encouraging using herbal medicine for COVID-19 outbreak. *Acta Agric. Scand. Sec. B. Soil Plant Sci.* 2020, 70, 437–443. [CrossRef]
- 25. Shahrajabian, M.H.; Sun, W.; Cheng, Q. Traditional herbal medicine for the prevention and treatment of cold and flu in the autumn of 2020, overlapped with COVID-19. *Nat. Prod. Commun.* **2020**, *15*, 1934578X20951431. [CrossRef]
- Islam, M.; Huang, Y.; Islam, S.; Fan, B.; Tong, L.; Wang, F. Influence of the degree of hydrolysis on functional properties and antioxidant activity of enzymatic soybean protein hydrolysates. *Molecules* 2022, 27, 6110. [CrossRef] [PubMed]
- 27. Peslerbes, M.; Fellenberg, A.; Jardin, J.; Deglaire, A.; Ibanez, R.A. Manufacture of whey protein hydrolysates using plant enzymes: Effect of processing conditions and simulated gastrointestinal digestion on angiotensin-I-converting enzyme (ACE) inhibitory activity. *Foods* **2022**, *11*, 2429. [CrossRef]
- Cosovanu, D.; Acosta, A.M.; Lopez, P.C.; Gernaey, K.V.; Li, Q.; Lametsch, R.; Canela-Garayoa, R.; Eras, J.; Villorbina, G. Renderedprotein hydrolysates as a low-cost nitrogen source for the fungal biotransformation of 5-hydroxymethylfurfural. *Catalysts* 2022, 12, 839. [CrossRef]
- 29. Aluko, R.E. Amino acids, peptides, and proteins as antioxidants for food preservation. In *Handbook of Antioxidants for Food Preservation*; Shahidi, F., Ed.; Elsevier Inc.: Cambridge, UK, 2015; pp. 105–140.
- Makhaye, G.; Aremu, A.O.; Gerrano, A.S.; Tesfay, S.; Du Plooy, C.P.; Amoo, S.O. Biopriming with seaweed extract and microbialbased commercial biostimulants influences seed germination of five *Abelmoschus esculentus* genotypes. *Plants* 2021, 10, 1327. [CrossRef] [PubMed]
- 31. Martinez-Alvarez, O.; Chamorro, S.; Brenes, A. Protein hydrolysates from animal processing by-products as a source of bioactive molecules with interest in animal feeding: A review. *Food Res. Int.* **2015**, *73*, 204–212. [CrossRef]
- Ebinezer, L.B.; Franchin, C.; Trentin, A.R.; Carletti, P.; Trevisan, S.; Agrawal, G.K.; Rakwal, R.; Quaggiotti, S.; Arriogoni, G.; Masi, A. Quantitative proteomics of maize roots treated with a protein hydrolysate: A comparative study with transcriptomics highlights the molecular mechanisms responsive to biostimulants. *J. Agric. Food Chem.* 2020, *68*, 7541–7553. [CrossRef] [PubMed]

- 33. Brown, P.; Saa, S. Biostimulants in agriculture. Front. Plant Sci. 2015, 6, 671. [CrossRef] [PubMed]
- 34. Baqer, H.A.A.-R.; Zeboon, N.; Al-Behadili, A. The tole and importance of amino acids within plants: A review. *Plant Arch.* **2019**, *19*, 1402–1410.
- 35. Calvo, P.; Nelson, L.; Kloepper, J.W. Agricultural uses of plant biostimulants. Plant Soil 2014, 383, 3–41. [CrossRef]
- Caruso, G.; De Pascale, S.; Cozzolino, E.; Giordano, M.; El-Nakhel, C.; Cuciniello, A.; Cenvinzo, V.; Colla, G.; Rouphael, Y. Protein hydrolysate or plant extract-based biostimulants enhanced yield and quality performances of greenhouse perennial wall rocket grown in different seasons. *Plants* 2019, *8*, 208. [CrossRef]
- 37. Chai, T.-T.; Law, Y.-C.; Wong, F.-C.; Kim, S.-K. Enzyme-assisted discovery of antioxidant peptides from edible marine invertebrates: A review. *Mar. Drugs* **2017**, *15*, 42. [CrossRef] [PubMed]
- Chai, T.-T.; Ee, K.-Y.; Kumar, D.T.; Manan, F.A.; Wong, F.-C. Plant bioactive peptides: Current status and prospects towards use on human health. *Protein Pept. Lett.* 2021, 28, 623–642. [CrossRef]
- Wong, F.-C.; Xiao, J.; Wang, S.; Ee, K.-Y.; Chai, T.-T. Advances on the antioxidant peptides from edible plant sources. *Trends Food Sci. Technol.* 2020, 99, 44–57. [CrossRef]
- 40. Ryu, B.; Shin, K.-H.; Kim, S.-K. Muscle protein hydrolysates and amino acid composition in fish. *Mar. Drugs* **2021**, *19*, 377. [CrossRef]
- Siddik, M.A.B.; Howieson, J.; Fotedar, R.; Partridge, G.J. Enzymatic fish protein hydrolysates in finfish aquaculture: A review. *Rev. Aquac.* 2021, 13, 406–430. [CrossRef]
- 42. Sierra Lopera, L.M.; Sepulveda Rincon, C.T.; Vasquez Mazo, P.; Figueroa Moreno, O.A.; Zapata Montoya, J.E. Byproducts of aquaculture processes: Development and prospective uses. Review. *Vitae* 2018, 25, 128–140. [CrossRef]
- 43. Colla, G.; Nardi, S.; Cardarelli, M.; Ertani, A.; Lucini, L.; Canaguier, R.; Rouphael, Y. Protein hydrolysates as biostimulants in horticulture. *Sci. Hortic.* **2015**, *196*, 28–38. [CrossRef]
- 44. Gonzalez-Morales, S.; Solis-Gaona, S.; Valdes-Caballero, M.V.; Juarez-Maldonado, A.; Loredo-Trevino, A.; Benavides-Mendoza, A. Transcriptomics of biostimulation of plants under abiotic stress. *Front. Genet.* **2021**, *12*, 36. [CrossRef] [PubMed]
- Colla, G.; Hoagland, L.; Ruzzi, M.; Cardarelli, M.; Bonini, P.; Canaguier, R.; Rouphael, Y. Biostimulant action of protein hydrolysates: Unraveling their effects on plant physiology and microbiome. *Front. Plant Sci.* 2017, *8*, 2202. [CrossRef] [PubMed]
- Bazsefidar, N.; Yazdi, A.P.G.; Karimi, A.; Yahyavi, M.; Amini, M.; Gavlighi, H.A.; Simal-Gandara, J. Brewers spent grain protein hydrolysate as a functional ingredient for muffins: Antioxidant, antidiabetic, and sensory evaluation. *Food Chem.* 2024, 435, 137565. [CrossRef] [PubMed]
- McCarthy, A.L.; O'Callaghan, Y.C.; O'Brien, N.M. Protein hydrolysates from agricultural crops-bioactivity and potential for functional food development. *Agriculture* 2013, *3*, 112–130. [CrossRef]
- Kong, Y.; Toh, N.P.; Wu, Y.; Huang, D. Trypsin-treated chickpea protein hydrolysate enhances the cytoaffinity of microbeads for cultures meat application. *Food Res. Int.* 2023, 173, 113299. [CrossRef]
- Sharma, S.; Pradhan, R.; Manickavasagan, A.; Thimmanagari, M.; Dutta, A. Exploration of corn distillers solubles from selective milling technology as a novel source of plant-based ACE inhibitory protein hydrolysates. *Food Chem.* 2022, 388, 133036. [CrossRef]
- Wang, C.; Rao, J.; Li, X.; He, D.; Zhang, T.; Xu, J.; Chen, X.; Wang, L.; Yuan, Y.; Zhu, X. Chickpea protein hydrolysate as a novel plant-based cryoprotectant in frozen surimi: Insights into protein structure integrity and gelling behaviors. *Food Res. Int.* 2023, 169, 112871. [CrossRef]
- 51. Zhang, L.; Qu, H.; Xie, M.; Shi, T.; Shi, P.; Yu, M. Effects of different cooking methods on phenol content and antioxidant activity in sprouted peanut. *Molecules* 2023, *28*, 4684. [CrossRef]
- 52. Zakraoui, M.; Hannachi, H.; Paskovic, I.; Vidovic, N.; Paskovic, M.P.; Palcic, I.; Major, N.; Ban, S.G.; Hamrouni, L. Effect of geographical location on the phenolic and mineral composition of Chetoui olive leaves. *Foods* **2023**, *12*, 2565. [CrossRef]
- 53. Horvat, D.; Simic, G.; Drezner, G.; Lalic, A.; Ledencan, T.; Tucak, M.; Pavsic, H.; Andric, L.; Zdunic, Z. Phenolic acid profiles and antioxidant activity of major cereal crops. *Antioxidants* **2020**, *9*, 527. [CrossRef] [PubMed]
- 54. Eseberri, I.; Trepiana, J.; Leniz, A.; Gomez-Garcia, I.; Carr-Ugarte, H.; Gonzalez, M.; Portillo, M.P. Variability in beneficinal effects of phenolic compounds: A review. *Nutrients* **2022**, *14*, 1925. [CrossRef] [PubMed]
- 55. Wang, Z.; Barrow, C.J.; Dunshea, F.R.; Suleria, H.A.R. A comparative investigation on phenolic composition, characterization and antioxidant potentials of five different Australian grown pear varieties. *Antioxidants* **2021**, *10*, 151. [CrossRef]
- Zhang, H.; Pu, J.; Tang, Y.; Wang, M.; Tian, K.; Wang, Y.; Luo, X.; Deng, Q. Changes in phenolic compounds and antioxidant activity during development of Qiangcuili and Cuihongli fruit. *Foods* 2022, *11*, 3198. [CrossRef] [PubMed]
- 57. Shahrajabian, M.H.; Sun, W.; Cheng, Q. Using bacteria and fungi as plant biostimulants for sustainable agricultural production systems. *Rec. Pat. Biotechnol.* 2022, 17, 206–244. [CrossRef] [PubMed]
- 58. Shahrajabian, M.H.; Sun, W.; Cheng, Q. Foliar application of nutrients on medicinal and aromatic plants, the sustainable approaches for higher and better production. *Beni-Suef Uni. J. Basic Appl. Sci.* **2022**, *11*, 26. [CrossRef]
- 59. Shahrajabian, M.H.; Sun, W. Mechanism of action of collagen and epidermal growth factor: A review on theory and research methods. *Mini Rev. Med. Chem.* 2023, 23, 453–477. [CrossRef]
- 60. Shahrajabian, M.H.; Sun, W. Five important seeds in traditional medicine, and pharmacological benefits. *Seeds* **2023**, *2*, 290–308. [CrossRef]
- 61. Shahrajabian, M.H.; Sun, W. The golden spice for life: Turmeric with the pharmacological benefits of curcuminoids components, including curcumin, bisdemethoxycurcumin, and demethoxycurcumin. *Curr. Org. Synth.* **2023**, 20. [CrossRef]

- 62. Shahrajabian, M.H.; Kuang, Y.; Cui, H.; Fu, L.; Sun, W. Metabolic changes of active components of important medicinal plants on the basis of traditional Chinese medicine under different envrionmental stresses. *Curr. Org. Chem.* 2023, 27, 782–806. [CrossRef]
- 63. Sierras, N.; Botta, A.; Staasing, L.; Martinez, M.J.; Bru, R. Understanding the effect of amino acids based biostimulant by an enantiomeric analysis of their active principles and a proteomic profiling approach. *Acta Hortic.* **2016**, *1148*, 93–100. [CrossRef]
- 64. Atilio, J.B.; Causin, H.F. The central role of amino acids on nitrogen utilization and plant growth. *J. Plant Physiol.* **1996**, 149, 358–362. [CrossRef]
- 65. Rai, V.K. Role of amino acids in plant responses to stresses. Biol. Plant. 2002, 45, 481–487. [CrossRef]
- 66. Shahrajabian, M.H.; Sun, W.; Soleymani, A.; Cheng, Q. Traditional herbal medicines to overcome stress, anxiety and improve mental health in outbreaks of human coronaviruses. *Phytother. Res.* **2020**, *1237–1247*. [CrossRef] [PubMed]
- 67. Shahrajabian, M.H.; Chaski, C.; Polyzos, N.; Tzortzakis, N.; Petropoulos, S.A. Sustainable agriculture systems in vegetable production using chitin and chitosan as plant biostimulants. *Biomolecules* **2021**, *11*, 819. [CrossRef]
- 68. Shahrajabian, M.H.; Chaski, C.; Polyzos, N.; Petropoulos, S.A. Biostimulants application: A low input cropping management tool for sustainable farming vegetables. *Biomolecules* **2021**, *11*, 698. [CrossRef]
- Shahrajabian, M.H.; Sun, W.; Cheng, Q. Different methods for molecular and rapid detection of human novel coronavirus. *Curr. Pharm. Des.* 2021, 27, 2893–2903. [CrossRef]
- Shahrajabian, M.H.; Sun, W. Sustainable approaches to boost yield and chemical constituents of aromatic and medicinal plants by application of biostimulants. *Recent Pat. Food Nutr. Agric.* 2022, 13, 72–92. [CrossRef]
- 71. Shahrajabian, M.H.; Cheng, Q.; Sun, W. The effects of amino acids, phenols and protein hydrolysates as biostimulants on sustainable crop production and alleviated stress. *Rec. Pat. Biotechnol.* **2022**, *16*, 319–328. [CrossRef]
- 72. Cheng, Y.; Tian, Q.; Zhang, W.-H. Glutamate receptors are involved in mitigating effects of amino acids on seed germination of *Arabidopsis thaliana* under salt stress. *Environ. Exp. Bot.* **2016**, *130*, 68–78. [CrossRef]
- 73. Parthasarathy, A.; Savka, M.A.; Hudson, A.O. The synthesis and the role of B-alanine in plants. *Front. Plant Sci.* 2019, 10, 921. [CrossRef] [PubMed]
- Shahrajabian, M.H.; Sun, W.; Cheng, Q. Chemical components and pharmacological benefits of basil (*Ocimum bacilicum*): A review. *Int. J. Food Prop.* 2020, 23, 1961–1970. [CrossRef]
- 75. Paleckiene, R.; Sviklas, A.; Slinksiene, R. Physicochemical properties of a microelement fertilizer with amino acids. *Russ. J. Appl. Chem.* **2007**, *80*, 352–357. [CrossRef]
- 76. Johansson, A. Conversations on chelation and mineral nutrition. Aust. J. Grape Wine Res. 2008, 583, 53–56.
- 77. Seadh, S.R.; El-Abady, M.I.; Farouk, S.; El-Saidy Amal, E.A. Effect of foliar nutrition with humic and amino acids under N-levels on wheat productivity and quality of grains and seeds. *Egypt. J. Appl. Sci.* **2008**, *23*, 543–558.
- Toscano, S.; Gomez-Bellot, M.J.; Romano, D.; Sanchez-Blanco, M.J. Physiological and biochemical changes in response to *Moringa* oleifera biostimulant in petunia plants under water deficit. *Sci. Hortic.* 2023, 319, 112187. [CrossRef]
- 79. Popko, M.; Michalak, I.; Wilk, R.; Gramza, M.; Chojnacka, K.; Gorecki, H. Effect of the new plant growth biostimulants based on amino acids on yield and grain quality of winter wheat. *Molecules* **2018**, *23*, 470. [CrossRef]
- Amin, A.A.; Gharib, F.A.; El-Awadi, M.; Rashad, E.-S.M. Physiological response of onion plants to foliar application of putrescine and glutamine. *Sci. Hortic.* 2011, 129, 353–360. [CrossRef]
- 81. Meister, A. Glutathione metabolism and its selective modification. J. Biol. Chem. 1988, 263, 17205–17208. [CrossRef]
- 82. Das, C.; Sengupta, T.; Chattopadhyay, S.; Setua, M.; Das, N.K.; Saratchandra, B. Involvement of kinetin and spermidine in controlling salinity stress in mulberry (*Morus alba* L. cv. S1). *Acta Physiol. Plant.* **2002**, *24*, 53–57. [CrossRef]
- Khan, S.; Yu, H.; Li, Q.; Gao, Y.; Sallam, B.N.; Wang, H.; Liu, P.; Jiang, W. Exogenous application of amino acids improves the growth and yield of lettuce by enhancing photosynthetic assimilation and nutrient availability. *Agronomy* 2019, 9, 266. [CrossRef]
- 84. Maeda, H.; Dudareva, N. The shikimate pathway and aromatic amino acid biosynthesis in plants. *Annu. Rev. Plant Biol.* **2012**, *63*, 73–105. [CrossRef] [PubMed]
- Tegeder, M.; Masclaux-Daubresse, C. Source and sink mechanisms of nitrogen transport and use. *New Phytol.* 2018, 217, 35–53.
 [CrossRef]
- 86. Musbah, H.M.; Ibrahim, K.M. Effects of feeding tyrosine or phenylalanine on the accumulation of polyphenols in *Coleus blumei in vivo* and *in vitro*. J. Biotechnol. Res. Cent. **2019**, 13, 35–43. [CrossRef]
- 87. Reham, M.S.; Khattab, M.E.; Ahmed, S.S.; Kandil, M.A.M. Influence of foliar spray with phenylalanine and nickel on growth, yield quality, and chemical composition of genoveser basil plant. *Afr. J. Agric. Res.* **2016**, *14*, 934–941. [CrossRef]
- Baharlou, M.J.; Pirbalouti, A.G.; Malekpoor, F. Effect of different concentrations of L-phenylalanine on chemical compositions and yield of essential oil of lemon balm (*Melissa officinalis*). J. Herb. Drugs 2019, 10, 175–183.
- Noviyanti, R.; Sari, R.L.K.; Kristanti, A.N.; Yachya, A.; Manuhara, Y.S.W. Biomass and flavonoid production of gynura procumbens adventitious roots induced by sucrose, phenylalanine, and tyrosine. *Biosci. Res.* 2017, 14, 934–941.
- Portu, J.; Santamaria, P.; Lopez, R.; Garde-Cerdan, T. Phenolic composition of Tempranillo grapes following foliar applications of phenylalanine and urea: A two-year study. *Sci. Hortic.* 2017, 219, 191–199. [CrossRef]
- 91. Feng, Z.; Xie, X.; Wu, P.; Chen, M.; Qin, Y.; Zhou, Y.; Zhu, H.; Yao, Q. Phenylalanine-mediated changes in the soil bacterial community promote nitrogen cycling and plant growth. *Microbiol. Res.* **2023**, 275, 127447. [CrossRef]

- 92. Ramzan, T.; Shahbaz, M.; Maqsood, M.F.; Zulfiqar, U.; Saman, R.U.; Lili, N.; Irshad, M.; Maqsood, S.; Haider, A.; Shahzad, B.; et al. Phenylalanine supply alleviates the drought stress in mustard (*Brassica campestris*) by modulating plant growth, photosynthesis, and antioxidant defense system. *Plant Physiol. Biochem.* **2023**, 201, 107828. [CrossRef] [PubMed]
- 93. Roman, S.M.-S.; Garde-Cerdan, T.; Baroja, E.; Rubio-Breton, P.; Perez-Alvarez, E.P. Foliar application of phenylalanine plus methyl jasmonate as a tool to improve Grenache grape aromatic composition. *Sci. Hortic.* **2020**, 272, 109515. [CrossRef]
- 94. Portu, J.; Lopez-Alfaro, I.; Gomez-Alonso, S.; Lopez, R.; Garde-Cerdan, T. Changes on grape phenolic composition induced by grapevine foliar applications of phenylalanine and urea. *Food Chem.* **2015**, *180*, 171–180. [CrossRef] [PubMed]
- 95. Khalesi, A.; Mirkalaeia, S.A.M.; Sanavy, S.A.M.M.; Eftekhari, A.; Moghadam, M.N. Effect of foliar application of amino acids on grain yield and physiological traits of chickpea under drought stress. *Gesunde Pflanz.* **2023**, *75*, 1705–1718. [CrossRef]
- 96. Wang, D.; Deng, X.; Wang, B.; Zhang, N.; Zhu, C.; Jiao, Z.; Li, R.; Shen, Q. Effects of foliar application of amino acid liquid fertilizers, with or without *Bacillus amyloliquefaciens* SQR9, on cowpea yield and leaf microbiota. *PLoS ONE* 2019, 14, e0222048. [CrossRef]
- 97. Mian, G.; Belfiore, N.; Musetti, R.; Tomasi, D.; Cantone, P.; Lovat, L.; Lupinelli, S.; Iacumin, L.; Celotti, E.; Golinelli, F. Effects of a triacontanol-rich biostimulant on the ripening dynamic and wine must technological parameters in *Vitis vinifera* cv. Ribolla Gialla. *Plant Physiol. Biochem.* **2022**, *188*, 60–69. [CrossRef]
- 98. Al-Karaki, G.N.; Othman, Y. Effect of foliar application of amino acid biostimulants on growth, macronutrient, total phenol contents and antioxidant activity of soilless grown lettuce cultivars. *S. Afr. J. Bot.* **2023**, *154*, 225–231. [CrossRef]
- Mpai, S.; Mokganya, L.M.; Raphoko, L.; Masoko, P.; Ndhlala, A.R. Untargeted metabolites and chemometric approach to elucidate the response of growth and yield attributes on different concentrations of an amino acid based biostimulant in two lettuce cultivars. *Sci. Hortic.* 2022, 306, 111478. [CrossRef]
- 100. Hidalgo-Santiago, L.; Navarro-Leon, E.; Lopez-Moreno, F.J.; Arjo, G.; Gonzalez, L.M.; Ruiz, J.M.; Blasco, B. The application of the silicon-based biostimulant Codasil®offset water deficit of lettuce plants. *Sci. Hortic.* **2021**, *285*, 110177. [CrossRef]
- Abdelkader, M.; Voronina, L.; Baratova, L.; Shelepova, O.; Zargar, M.; Puchkov, M.; Lktionova, E.; Amantayev, B.; Kipshakbaeva, A.; Arinov, B. Biostimulants-based amino acids augment physio-biochemical responses and promote salinity tolerance of lettuce plants (*Lactuca sativa* L.). *Horticulturae* 2023, *9*, 807. [CrossRef]
- 102. Navarro-Leon, E.; Lopez-Moreno, F.J.; Borda, E.; Marin, C.; Sierras, N.; Blasco, B.; Ruiz, J.M. Effect of L-amino acid-based biostimulants on nitrogen use efficiency (NUE) in lettuce plants. *J. Sci. Food Agric.* **2022**, *102*, 7098–7106. [CrossRef] [PubMed]
- Velicka, A.; Taraseviciene, Z.; Hallmann, E.; Kieltyka-Dadasiewicz, A. Impact of foliar application of amino acids on essential oil content, odor profile, and flavonoids content of different mint varieties in field conditions. *Plants* 2022, 11, 2938. [CrossRef]
- 104. Seyedi, A.; Fathi, S.; Movlodzadeh, R. The effect of biostimulants based on free amino acids on some growth and physiological parameters of *Dracocephalum moldavica* L. under salinity stress. *J. Med. Plants By-Prod.* 2023; *in press.* [CrossRef]
- 105. Soppelsa, S.; Kelderer, M.; Casera, C.; Bassi, M.; Robatscher, P.; Matteazzi, A.; Andreotti, C. Foliar applications of biostimulants promote growth, yield and fruit quality of strawberry plants grown under nutrient limitation. *Agronomy* **2019**, *9*, 483. [CrossRef]
- 106. Neshev, N.; Balabanova, D.; Yanev, M.; Mitkov, A. Is the plant biostimulant application ameliorative for herbicide-damaged sunflower hybrids? *Ind. Crops Prod.* 2022, *182*, 114926. [CrossRef]
- 107. Klokic, I.; Koleska, I.; Hasanagic, D.; Murtic, S.; Bosancic, B.; Todorovic, V. Biostimulants' influence on tomato fruit characteristics at conventional and low-input NPK regime. *Acta Agric. Scand. Sec. B Soil Plant Sci.* **2020**, *70*, 233–240. [CrossRef]
- 108. Niu, C.; Wang, G.; Sui, J.; Liu, G.; Ma, F.; Bao, Z. Biostimulants alleviate temperature stress in tomato seedlings. *Sci. Hortic.* 2022, 293, 110712. [CrossRef]
- Alfosea-Simon, M.; Zavala-Gonzalez, E.A.; Camara-Zapata, J.M.; Martinez-Nicolas, J.J.; Simon, I.; Simon-Grao, S.; Garcia-Sanchez, F. Effect of foliar application of amino acids on the salinity tolerance of tomato plants cultivated under hydroponic system. *Sci. Hortic.* 2020, 272, 109509. [CrossRef]
- 110. Almadi, L.; Paoletti, A.; Cinosi, N.; Daher, E.; Rosati, A.; Di Vaio, C.; Famiani, F. A biostimulant based on protein hydrolysates promotes the growth of young olive trees. *Agriculture* **2020**, *10*, 618. [CrossRef]
- 111. Nargesi, M.M.; Sedaghathoor, S.; Hashemabadi, D. Effect of foliar application of amino acid, humic acid, and fulvic acid on the soil content and quality of olive. *Saudi J. Biol. Sci.* 2022, 29, 3473–3481. [CrossRef]
- 112. Sadak, M.S.; Bakry, B.A.; Abdel-Razik, T.M.; Hanafy, R.S. Amino acids foliar application for maximizing growth, productivity and quality of peanut grown under sandy soil. *Braz. J. Biol.* 2023, *6*, e256338. [CrossRef]
- 113. Barrajon-Catalan, E.; Alvarez-Martinez, F.J.; Borras, F.; Perez, D.; Herrero, N.; Ruiz, J.J.; Micol, V. Metabolomic analysis of the effects of a commercial complez biostimulant on pepper crops. *Food Chem.* **2020**, *310*, 125818. [CrossRef] [PubMed]
- 114. Zamljen, T.; Medic, A.; Hudina, M.; Veberic, R.; Slatnar, A. Biostimulative effect of amino acids on the enzymatic and metabolic response of two *Capsicum annuum* L. cultivars grown under salt stress. *Sci. Hortic.* **2023**, *309*, 111713. [CrossRef]
- 115. Raza, S.; Zia-ur-Rehman, M.; Alghamdi, S.A.; Alghanem, S.M.S.; Usman, M.; Ahmed, R.; Waris, A.A.; Rizwan, M.; Abeed, A.H.A.; Al-Haithloul, H.A.S. Effects of zinc-enriched amino acids on rice plants (*Oryza sativa* L.) for adaptation in saline-sodic soil conditions: Growth, nutrient uptake and biofortification of zinc. S. Afr. J. Bot. 2023, 162, 370–380. [CrossRef]
- 116. Mirtaleb, S.H.; Niknejad, Y.; Fallah, H. Foliar spray of amino acids and potassic fertilizer improves the nutritional quality of rice. *J. Plant Nutr.* **2021**, *44*, 2029–2041. [CrossRef]
- 117. Kocira, S. Effect of amino acid biostimulant on the yield and nutraceutical potential of soybean. *Chil. J. Agric. Res.* 2019, 79, 17–25. [CrossRef]

- 118. Teixeira, W.F.; Fagan, E.B.; Soares, L.H.; Soares, J.N.; Reichardt, K.; Neto, D.D. Seed and foliar application of amino acids improve variables of nitrogen metabolism and productivity in soybean crop. *Front. Plant Sci.* **2018**, *9*, 396. [CrossRef]
- Kausar, A.; Zahra, N.; Tahir, H.; Hafeez, M.B.; Abbas, W.; Raza, A. Modulation of growth and biochemical responses in spinach (*Spinacia oleracea* L.) through foliar application of some amino acids under drought conditions. S. Afr. J. Bot. 2023, 158, 243–253. [CrossRef]
- Hosseini, S.; Shabani, L.; Sabzalian, M.R.; Gharibi, S. Foliar spray of commercial seaweed and amino-acid derived biostimulants promoted phytoremediation potential and salinity stress tolerance in halophytic grass, *Puccinellia distans*. *Int. J. Phytoremed.* 2023, 25, 415–429. [CrossRef]
- 121. Popko, M.; Wilk, R.; Gorecki, H. New amino acid biostimulators based on protein hydrolysate of keratin. *Przem. Chem.* **2014**, *93*, 1012–1015.
- 122. Hammad, S.A.R.; Ali, O.A.M. Physiological and biochemical studies on drought tolerance of wheat plants by application of amino acids and yeast extract. *Annals Agric. Sci.* 2014, *59*, 133–145. [CrossRef]
- 123. Lin, H.-C.; Alashi, A.M.; Aluko, R.E.; Pan, B.S.; Chang, Y.-W. Antihypertensive properties of tilapia (*Oreochromis* spp.) frame and skin enzymatic protein hydrolysates. *Food Nutr. Res.* **2017**, *61*, 1391666. [CrossRef]
- Czelej, M.; Czernecki, T.; Garbacz, K.; Wawrzykowski, J.; Jamiol, M.; Michalak, K.; Walczak, N.; Wilk, A.; Wasko, A. Egg yolk as a new source of peptides with antioxidant and antimicrobial properties. *Foods* 2023, 12, 3394. [CrossRef] [PubMed]
- 125. Bello, A.S.; Ben-Hamadou, R.; Hamdi, H.; Saadaoui, I.; Ahmed, T. Application of cyanobacteria (*Roholtiella* sp.) liquid extract for the alleviation of salt stress in bell pepper (*Capsiucum annuum* L.) plants grown in a soilless system. *Plants* 2022, *11*, 104. [CrossRef]
- 126. Chan, Y.-J.; Lu, W.-C.; Lin, H.-Y.; Wu, Z.-R.; Liou, C.-W.; Li, P.-H. Effect of rice protein hydrolysates as an egg replacement on the physicochemical properties of flaky egg rolls. *Foods* **2020**, *9*, 245. [CrossRef] [PubMed]
- 127. Liao, X.; Zhu, Z.; Wu, S.; Chen, M.; Huang, R.; Wang, J.; Wu, Q.; Ding, Y. Preparation of antioxidant protein hydrolysates from *Pleurotus geesterans* and their protective effects on H₂O₂ oxidative damaged PC12 cells. *Molecules* 2020, 25, 5408. [CrossRef] [PubMed]
- 128. Leni, G.; Soetemans, L.; Caligiani, A.; Sforza, S.; Bastiaens, L. Degree of hydrolysis affects the techno-functional properties of lesser mealworm protein hydrolysates. *Foods* **2020**, *9*, 381. [CrossRef]
- Kristinsson, H.G.; Rasco, B.A. Fish protein hydrolysates: Production, biochemical, and functional properties. *Crit. Rev. Food Sci.* Nutr. 2000, 40, 43–81. [CrossRef] [PubMed]
- Kanbargi, K.D.; Sonawane, S.K.; Arya, S.S. Encapsulation characteristics of protein hydrolyaste extracted from Ziziphus jujube seed. Int. J Food Prop. 2017, 20, 3215–3224. [CrossRef]
- 131. Sun, W.; Shahrajabian, M.H.; Cheng, Q. Organic waste utilization and urban food waste composting strategies in China—A review. *Not. Sci. Biol.* **2021**, *13*, 10881. [CrossRef]
- Sun, W.; Shahrajabian, M.H.; Cheng, Q. Archaea, bacteria and termite, nitrogen fixation and sustainable plants production. *Not. Bot. Horti Agrobot.* 2021, 49, 1–32. [CrossRef]
- 133. Sun, W.; Shahrajabian, M.H.; Cheng, Q. Barberry (*Berberis vulgaris*), a medicinal fruit and food with traditional and modern pharmaceutical uses. *Isr. J. Plant Sci.* 2021, *68*, 61–71. [CrossRef]
- 134. Sun, W.; Shahrajabian, M.H.; Cheng, Q. Fenugreek cultivation with emphasis on historical aspects and its uses in traditional medicine and modern pharmaceutical science. *Mini Rev. Med. Chem.* **2021**, *21*, 724–730. [CrossRef]
- 135. Sun, W.; Shahrajabian, M.H.; Cheng, Q. Natural dietary and medicinal plants with anti-obesity therapeutics activities for treatment and prevention of obesity during lock down and in post-COVID-19 era. *Appl. Sci.* 2021, *11*, 7889. [CrossRef]
- 136. Sun, W.; Shahrajabian, M.H. The effectiveness of *Rhizobium* bacteria on soil fertility and sustainable crop production under cover and catch crops management and green manuring. *Not. Bot. Horti Agrobot.* **2022**, *50*, 12560. [CrossRef]
- 137. Tchorbanov, B.; Iliev, I.; Litchev, V. Enzymic protein hydrolysates. Biotechnol. Biotechnol. Equip. 1991, 5, 32–36. [CrossRef]
- 138. Povolo, C.; Avolio, R.; Doria, E.; Marra, A. Development and validation of an analytical method to ensure quality requirements of hydrolysed proteins intended for agricultural use as biostimulants. *Talanta Open* **2022**, *5*, 100082. [CrossRef]
- 139. Jensen, C.; Dale, H.F.; Hausken, T.; Hatlebakk, J.G.; Bronstad, I.; Lied, G.A.; Hoff, D.A.L. Supplementation with low doses of a cod protein hydrolysate on glucose regulation and lipid metabolism in adults with metabolic syndrome: A randomized, double-blind study. *Nutrients* **2020**, *12*, 1991. [CrossRef] [PubMed]
- 140. Sarabandi, K.; Gharehbeglou, P.; Jafari, S.M. Spray-drying encapsulation of protein hydrolysates and bioactive peptides: Opportunities and challenges. *Dry. Technol.* **2020**, *38*, 577–595. [CrossRef]
- 141. Balan, D.; Luta, G.; Stanca, M.; Jerca, O.; Niculescu, M.; Gaidau, C.; Jurcoane, S.; Mihalcea, A. Effect of protein gel treatments on biometric and biochemical attributes of tomato seedlings in greenhouse condition. *Agriculture* **2023**, *13*, 54. [CrossRef]
- 142. Santi, C.; Zamboni, A.; Varanini, Z.; Pandolfini, T. Growth stimulatory effects and genome-wide transcriptional changes produced by protein hydrolysates in maize seedlings. *Front. Plant Sci.* **2017**, *8*, 433. [CrossRef]
- 143. Ertani, A.; Francioso, O.; Ferrari, E.; Schiavon, M.; Nardi, S. Spectroscopic-chemical fingerprint and biostimulant activity of a protein-based product in solid form. *Molecules* 2018, 23, 1031. [CrossRef]
- 144. Dash, P.; Ghosh, G. Amino acid profiling and antimicrobial activity of *Cucurbita moschata* and *Lagenaria siceraria* seed protein hydrolysates. *Nat. Prod. Res.* 2018, 32, 2050–2053. [CrossRef]
- 145. Schmidt, G.J.; Olson, D.C.; Slavin, W. Amino acid profiling of protein hydrolysates using liquid chromatography and fluorescence detection. *J. Liquid Chromatogr.* **1979**, *2*, 1031–1045. [CrossRef]

- 146. Phetchthumrongchai, T.; Tachapuripunya, V.; Chintong, S.; Roytrakul, S.; E-kobon, T.; Klaypradit, W. Properties of protein hydrolysates and bioinformatics prediction of peptides derived from thermal and enzymatic process of Skipjack tuna (*Katsuwonus pelamis*) roe. *Fishes* **2022**, *7*, 255. [CrossRef]
- 147. Chalamaiah, M.; Jyothirmayi, T.; Diwan, P.V.; Dinesh Kumar, B. Antioxidant activity and functional properties of enzymatic protein hydrolysates from common carp (*Cyprinus carpio*) roe (egg). *J. Food Sci. Technol.* **2015**, *52*, 5817–5825. [CrossRef]
- 148. Bilenky, M.; Nair, A. Biostimulants combined with water soluble fertilizer had little effect on transplant growth and pepper yield grown under greenhouse conditions. *Int. J. Veg. Sci.* 2023, *29*, 25–39. [CrossRef]
- 149. Ghorbel-Bellaaj, O.; Jellouli, K.; Maalej, H. Shrimp processing by-products protein hydrolysates: Evaluation of antioxiant activity and application in biomass and proteases production. *Biocatal. Biotransform.* **2017**, *35*, 287–297. [CrossRef]
- 150. Li, H.; Aluko, R.E. Structural modulation of calmodulin and calmodulin-dependent protein kinase II by pea protein hydrolysates. *Int. J. Food Sci. Nutr.* **2006**, *57*, 178–189. [CrossRef]
- 151. Paradikovic, N.; Vinkovic, T.; Vinkovic Vrcek, I.; Zuntar, I.; Bojic, M.; Medic-Saric, M. Effect of natural biostimulants on yield and nutritional quality: An example of sweet yellow pepper (*Capsicum annuum* L.) plants. *J. Sci. Food Agric.* **2011**, *91*, 2146–2152. [CrossRef]
- 152. Rouphael, Y.; Cardarelli, M.; Bonini, P.; Colla, G. Synergistic action of a microbial-based biostimulant and a plant derived-protein hydrolysate enhances lettuce tolerance to alkalinity and salinity. *Front. Plant Sci.* **2017**, *8*, 131. [CrossRef]
- 153. Paradikovic, N.; Teklic, T.; Zelikovic, S.; Lisjak, M.; Spoljarevic, M. Biostimulants research in some horticultural plant species- A review. *Food Energy Secur.* 2019, *8*, e00162. [CrossRef]
- 154. Wilson, H.T.; Amirkhani, M.; Taylor, A.G. Evaluation of gelatin as a biostimulant seed treatment to improve plant performance. *Front. Plant Sci.* **2018**, *9*, 1006. [CrossRef]
- 155. Liang, X.; Zhang, L.; Natarajan, S.K.; Becker, D.F. Proline mechanisms of stress survival. *Anthoxidants Redox Signal.* **2013**, *19*, 998–1011. [CrossRef]
- 156. Vranova, V.; Rejsek, K.; Skene, K.R.; Formanek, P. Non-protein amino acids: Plant, soil, and ecosystem interactions. *Plant Soil* **2011**, 342, 31–48. [CrossRef]
- 157. Ashraf, M.; Foolad, M.R. Roles of glycine betaine and proline in improving plant abiotic stress resistance. *Environ. Exp. Bot.* 2007, 59, 206–216. [CrossRef]
- Flores, T.; Todd, C.D.; Tovar-Mendez, A.; Dhanoa, P.K.; Correa-Aragunde, N.; Hoyos, M.E.; Brownfield, D.M.; Mullen, R.T.; Lamattina, L.; Polacco, J.C. Arginase-negative mutants of Arabidopsis exhibit increased nitric oxide signaling in root development. *Plant Physiol.* 2008, 147, 1936–1946. [CrossRef]
- 159. Sharma, S.S.; Dietz, K.J. The significance of amino acids and amino acid-derived molecules in plant responses and adaptation to heavy metal stress. *J. Exp. Bot.* 2006, 57, 711–726. [CrossRef]
- 160. Sytar, O.; Kumar, A.; Latowski, D.; Kuczynska, P.; Strzalka, K.; Prasad, M.N.V. Heavy metal-induced oxidative damage, defense reactions, and detoxification mechanisms in plants. *Acta Physiol. Plant* **2013**, *35*, 985–999. [CrossRef]
- 161. Rouphael, Y.; Colla, G.; Hoagland, L.; Giordano, M.; El-Nakhel, C.; Cardarelli, M. Vegetal-protein hydrolysates based microgranule enhances growth, mineral content, and quality traits of vegetable transplants. *Sci. Hortic.* **2021**, 290, 110554. [CrossRef]
- 162. Colantoni, A.; Recchia, L.; Bernaberi, G.; Cardarelli, M.; Rouphael, Y.; Colla, G. Analyzing the environmental impact of chemicallyproduced protein hydrolysate from leather waste vs. enzymatically-produced protein hydrolysate from legume grains. *Agriculture* **2017**, *7*, 62. [CrossRef]
- 163. Porterfield, D.M. Environmental sensing and directional growth of plant roots. In *Plant Roots: The Hidden Half*, 4th ed.; CRC Press: Boca Raton, FL, USA, 2002.
- 164. Trevisan, S.; Manoli, A.; Ravazzolo, L.; Franceschi, C.; Quaggiotti, S. mRNA-sequencing analysis reveals transcriptional changes in root of maize seedings treated with two increasing concentrations of a new biostimulant. *J. Agric. Food Chem.* 2017, 65, 9956–9969. [CrossRef] [PubMed]
- 165. Kisvarga, S.; Farkas, D.; Boronkay, G.; Nemenyi, A.; Orloci, L. Effects of biostimulants in horticulture, with emphasis on ornamental plant production. *Agronomy* **2022**, *12*, 1043. [CrossRef]
- 166. Tanou, G.; Ziogas, V.; Molassiotis, A. Foliar nutrition, biostimulants and prime-like dynamics in fruit tree physiology: New insights on an old topic. *Front. Plant Sci.* 2017, *8*, 75. [CrossRef]
- Malecange, M.; Sergheraert, R.; Teulat, B.; Mounier, E.; Lothier, J.; Sakr, S. Biostimulant properties of protein hydrolysates: Recent advances and future challenges. *Int. J. Mol. Sci.* 2023, 24, 9714. [CrossRef] [PubMed]
- 168. Soppela, S.; Kelderer, M.; Casera, C.; Bassi, M.; Robatscher, P.; Andreotti, C. Use of biostimulants for organic apple production: Effects on tree growth, yield, and fruit quality at harvest and during storage. *Front. Plant Sci.* **2018**, *9*, 1342. [CrossRef]
- 169. Gurav, R.; Jadhav, J. A novel source of biofertilizer from feather biomass for banana cultivation. *Environ. Sci. Pollut. Res.* 2013, 20, 4532–4539. [CrossRef] [PubMed]
- 170. Consentino, B.B.; Vultaggio, L.; Sabatino, L.; Ntatsi, G.; Rouphael, Y.; Bondi, C.; Pasquale, C.D.; Guarino, V.; Iacuzzi, N.; Capodici, G.; et al. Combined effects of biostimulants, N level and drought stress on yield, quality and physiology of greenhouse-grown basil. *Plant Stress* 2023, 10, 100268. [CrossRef]
- 171. Rosiane, L.S.L.; Liv, S.S.; Ligia, R.S.; Sofiatti, V.; Gomes, J.A.; Beltrao, N.E.M. Blends of castor meal and castor husks for optimized use as organic fertilizer. *Ind. Crops Prod.* 2011, *33*, 364–368. [CrossRef]

- 172. Consentino, B.B.; Virga, G.; Placa, G.G.L.; Sabatino, L.; Rouphael, Y.; Ntatsi, G.; Iapichino, G.; Bella, S.L.; Mauro, R.P.; Anna, F.D.; et al. Celery (*Apium graveolens* L.) performances as subjected to different sources of protein hydrolysates. *Plants* 2020, 9, 1633. [CrossRef]
- 173. Paul, T.; Halder, S.K.; Das, A.; Bera, S.; Maity, C.; Mandal, A.; Das, P.S.; Mohapatra, P.K.; Pati, B.R.; Mondal, K.C. Exploitation of chicken feather waste as a plant growth promoting agent using keratinase producing novel isolate *Paenibacillus woosongensis* TKB2. *Biocatal. Agric. Biotechnol.* 2013, 2, 50–57. [CrossRef]
- 174. Sitohy, M.Z.; Desoky, E.M.; Osman, A.; Rady, M.M. Pumpkin seed protein hydrolysate treatment alleviates salt stress effects on *Phaseolus vulgaris* by elevating antioxidant capacity and recovering ion homeostasis. *Sci. Hortic.* **2020**, *271*, 109495. [CrossRef]
- 175. Mohammadipour, N.; Souri, M.K. Effects of different levels of glycine in the nutrient solution on the growth, nutrient composition, and antioxidant activity of coriander (*Coriandrum sativum* L.). *Acta Agrobot.* **2019**, 72, 13–16. [CrossRef]
- 176. Carillo, P.; Pannico, A.; Cirillo, C.; Ciriello, M.; Colla, G.; Cardarelli, M.; Pascale, S.D.; Rouphael, Y. Protein hydrolysates from animal or vegetal sources affect morpho-physiological traits, ornamental quality, mineral composition, and shelf-life of Chrysanthemum in a distinctive manner. *Plants* 2022, *11*, 2321. [CrossRef]
- 177. Garcia-Santiago, J.C.; Cavazos, C.J.; Gonzalez-Fuentes, J.; Zermeno-Gonzalez, A.; Alvarado, E.R.; Duarte, A.R.; Preciado-Rangel, P.; Troyo-dieguez, E.; Ramos, F.M.P.; Valdez-Aguilar, L.A.; et al. Effect of fish-derived protein hydrolysate, animal-based organic fertilisers and irrigation method on the growth and quality of grape tomatoes. *Biol. Agric. Hortic.* 2021, 37, 107–124. [CrossRef]
- 178. Bavaresco, L.; Lucini, L.; Squeri, C.; Zamboni, M.; Frioni, T. Protein hydrolysates modulate leaf proteome and metabolome in water-stressed grapevines. *Sci. Hortic.* 2020, 270, 109413. [CrossRef]
- 179. Boselli, M.; Bahouaoui, M.A.; Lachhab, N.; Sanzani, S.M.; Ferrara, G.; Ippolito, A. Protein hydrolysates effects on grapevine (*Vitis vinifera* L., cv. Corvina) performance and water stress tolerance. *Sci. Hortic.* **2019**, 258, 108784. [CrossRef]
- 180. Di Mola, I.; Conti, S.; Cozzolino, E.; Melchionna, G.; Ottaiano, L.; Testa, A.; Sabatino, L.; Rouphael, Y.; Mori, M. Plant-based protein hydrolysate improves salinity tolerance in hemp: Agronomical and physiological aspects. *Agronomy* **2021**, *11*, 342. [CrossRef]
- Wise, K.; Selby-Pham, J.; Chai, X.; Simovich, T.; Gupta, S.; Gill, H. Fertiliser supplementation with a biostimulant complex of fish hydrolysate, *Aloe vera* extract, and kelp alters cannabis root architecture to enhance nutrient uptake. *Sci. Hortic.* 2023, 323, 112483. [CrossRef]
- 182. Massa, D.; Lenzi, A.; Montoneri, E.; Ginepro, M.; Prisa, D.; Burchi, G. Plant response to biowaste soluble hydrolysates in hibiscus grown under limiting nutrient availability. *J. Plant Nutr.* **2018**, *41*, 396–409. [CrossRef]
- 183. Qurartieri, M.; Lucchi, A.; Cavani, L. Effects of the rate of protein hydrolysis and spray concentration on growth of potted kiwifruit (*Actinidia deliciosa*) plants. *Acta Hortic.* **2022**, *594*, 341–347. [CrossRef]
- Xu, C.; Mou, B. Drench application of fish-derived protein hydrolysates affects lettuce growth, chlorophyll content, and gas exchange. *Hortechnology* 2017, 27, 539–543. [CrossRef]
- 185. Naroozlo, Y.A.; Souri, K.M.; Mojtaba, D. Stimulation effects of foliar applied glycine and glutamine amino acids on lettuce growth. *Open Agric.* **2019**, *4*, 164. [CrossRef]
- 186. Sabatino, L.; Consentino, B.B.; Rouphael, Y.; De Pasquale, C.; Iapichino, G.; D Anna, F.; La Bella, S. Protein hydrolysates and Mo-biofortification interactively modulate plant performance and quality of Canasta lettuce grown in a protected environment. *Agronomy* 2021, 11, 1023. [CrossRef]
- Dass, S.M.; Chai, T.-T.; Cao, H.; Ooi, A.L.; Wong, F.-C. Application of enzyme-digested soy protein hydrolysate on hydroponicplanted lettuce: Effects on phytochemical contents, biochemical profiles and physical properties. *Food Chem.* 2021, 12, 100132. [CrossRef]
- 188. El-Nakhel, C.; Cristofano, F.; Colla, G.; Pii, Y.; Lucini, L.; Rouphael, Y. A *Graminaceae*-derived protein hydrolysate and its fractions provide differential growth and modulate qualitative traits of lettuce grown under non-saline and mild salinity conditions. *Sci. Hortic.* 2023, *319*, 112130. [CrossRef]
- Lucini, L.; Rouphael, Y.; Cardarelli, M.; Canaguier, R.; Kumar, P.; Colla, G. The effect of a plant-derived biostimulant on metabolic profiling and crop performance of lettuce grown under saline conditions. *Sci. Hortic.* 2015, *182*, 124–133. [CrossRef]
- 190. Zhou, W.; Zheng, W.; Wang, W.; Lv, H.; Liang, B.; Li, J. Exogenous pig blood-derived protein hydrolysates as a promising method for alleviation of salt stress in tomato (*Solanum lycopersicum* L.). *Sci. Hortic.* **2022**, 294, 110779. [CrossRef]
- 191. Liu, N.; Qu, P.; Huang, H.; Wei, Z. Soybean protein hydrolysate-formaldehyde-urea block copolymer for controlled release fertilizer. *Environ. Pollut. Bioavailab.* 2019, 31, 94–102. [CrossRef]
- Sharma, S.; Pradhan, R.; Manickavasagan, A.; Tsopmo, A.; Thimmanagari, M.; Dutta, A. Corn distillers solubles by two-step proteolytic hydrolysis as a new source of plant-based protein hydrolysates with ACE and DPP4 inhibition activities. *Food Chem.* 2023, 401, 134120. [CrossRef]
- 193. Ertani, A.; Cavani, L.; Pizzeghello, D.; Brandellero, E.; Altissimo, A.; Ciavatta, C.; Nardi, S. Biostimulant activity of two protein hydrolyzates in the growth and nitrogen metabolism of maize seedlings. *J. Plant Nutr. Soil Sci.* 2009, 172, 237–244. [CrossRef]
- 194. Vaseva, I.I.; Simova-Stoilova, L.; Kostadinova, A.; Yuperlieva-Mateeva, B.; Karakicheva, T.; Vassileva, V. Heat-stress-mitigating effects of a protein-hydrolysate-based biostimulant are linked to changes in *Protease*, *DHN*, and *HSP* gene expression in maize. *Agronomy* **2022**, *12*, 1127. [CrossRef]
- 195. Trevisan, S.; Manoli, A.; Quaggiotti, S. A novel biostimulant, belonging to protein hydrolysates, mitigates abiotic stress effects on maize seedlings grown in hydroponics. *Agronomy* **2019**, *9*, 28. [CrossRef]

- 196. Ambrosini, S.; Prinsi, B.; Zamboni, A.; Espen, L.; Zanzoni, S.; Santi, C.; Varanini, Z.; Pandolfini, T. Chemical characterization of a collagen-derived protein hydrolysate and biostimulant activity assessment of its peptidic components. *J. Agric. Food Chem.* 2020, 70, 11201–11211. [CrossRef] [PubMed]
- 197. Lei, H.; Zhang, J.; Jia, C.; Feng, J.; Liang, L.; Chen, Q.Q.; Li, T.; Hao, J. Foliar application of fish protein peptide improved the quality of deep-netted melon. *J. Plant Nutr.* **2023**, *46*, 3683–3696. [CrossRef]
- 198. Eguchi, Y.; Milazzo, M.C.; Ueno, K.; Shetty, K. Partial improvement of vitrification and acclimation of oregano (*Origanum vulgare* L.) tissue culture by fish protein hydrolysates. J. Herb Spice Med. *Plants* **2000**, *6*, 29–38. [CrossRef]
- Morales-Payan, P.; Stall, W. Passion fruit (*Passiflora Edulis*) transplant production is affected by selected biostimulants. In Proceedings of the Florida State Horticultural Society, Fort Lauderdale, FL, USA, 6–8 June 2004; Volume 117, pp. 224–227.
- Osman, A.; Merwad, A.-R.M.; Mohamed, A.H.; Sitohy, M. Foliar spray with pepsin-and papain-whey protein hydrolysates promotes the productivity of pea plants cultivated in clay loam soil. *Molecules* 2021, 26, 2805. [CrossRef] [PubMed]
- Aktsoglou, D.-C.; Kasampalis, D.S.; Sarrou, E.; Tsouvaltzis, P.; Chatzopoulou, P.; Martens, S.; Siomos, A.S. Protein hydrolysates supplement in the nutrient solution of soilless grown fresh peppermint and spearmint as a tool for improving product quality. *Agronomy* 2021, 11, 317. [CrossRef]
- Visconti, F.; de Paz, J.M.; Bonet, L.; Jorda, M.; Quinones, A.; Intrigliolo, D.S. Effects of a commercial calcium protein hydrolysate on the salt tolerance of *Diospyros kaki* L. cv. Rojo Brillante grafted on *Diospyros lotus* L. Sci. Hortic. 2015, 185, 129–138. [CrossRef]
- 203. Wang, J.; Liu, Z.; Wang, Y.; Cheng, Q.; Mou, H. Production of a water-soluble fertilizer containing amino acids by solid-state fermentation of soybean meal and evaluation of its efficacy on the rapeseed growth. *J. Biotechnol.* **2014**, *187*, 34–42. [CrossRef]
- 204. Luo, Y.; Niu, L.; Li, D.; Xiao, J. Synergistic effects of plant protein hydrolysates and xanthan gum on the short- and long-term retrogradation of rice starch. *Int. J. Biol. Macromol.* **2020**, 144, 967–977. [CrossRef]
- 205. Calderon-chiu, C.; Calderon-santoyo, M.; Damasceno-gomes, S.; Ragazzo-Sanchez, J.A. Use of jackfruit leaf (*Artocarpus heterophyllus* L.) protein hydrolysates as a stabilizer of the nanoemulsions loaded with extract-rich pentacyclic triterpenes obtained from *Coccoloba uvifera* L. leaf. *Food Chem.* 2021, 12, 100138. [CrossRef] [PubMed]
- 206. Yuan, Y.; Dickinson, N. Nutrient interactions influence the efficacy of biostimulants. J. Plant Nutr. 2023, 46, 1616–1626. [CrossRef]
- 207. Lachhab, N.; Sanzani, S.M.; Adrian, M.; Chiltz, A.; Balacey, S.; Boselli, M.; Ippolito, A.; Pointssot, B. Soybean and casein hydrolysates induce grapevine immune responses and resistance against *Plasmopara viticola*. *Front. Plant Sci.* 2014, *5*, 716. [CrossRef] [PubMed]
- 208. Liatile, P.C.; Potgieter, G.; Moloi, M.J. A natural biostimulant consisting of a mixture of fish protein hydrolysates and kelp extract enhances the physiological, biochemical, and growth responses of Spinach under different water levels. *Plants* 2022, 11, 3374. [CrossRef] [PubMed]
- 209. Carillo, P.; Colla, G.; Fusco, G.M.; Aversana, E.D.; El-Nakhel, C.; Giordano, M.; Pannico, A.; Cozzolino, E.; Mori, M.; Reynaud, H.; et al. Morphological and physiological responses induced by protein hydrolysate-based biostimulant and nitrogen rates in greenhouse spinach. *Agronomy* 2019, *9*, 450. [CrossRef]
- Rouphael, Y.; Carillo, P.; Cristofano, F.; Cardarelli, M.; Colla, G. Effects of vegetal-versus animal-derived protein hydrolysate on sweet basil morpho-physiological and metabolic traits. *Sci. Hortic.* 2021, 284, 110123. [CrossRef]
- 211. Jolayemi, O.L.; Malik, A.H.; Vetukuri, R.R.; Saripella, G.V.; Kalyandurg, P.B.; Ekblad, T.; Yong, J.W.H.; Olsson, M.E.; Johansson, E. Metabolic processes and biological macromolecules defined the positive effects of protein-rich biostimulants on sugar beet plant development. *Int. J. Mol. Sci.* 2023, 24, 9720. [CrossRef]
- Jolayemi, O.L.; Malik, A.H.; Ekblad, T.; Fredlund, K.; Olsson, M.E.; Johansson, E. Protein-based biostimulants to enhance plant growth-state of the art and future direction with sugar beet as an example. *Agronomy* 2022, 12, 3211. [CrossRef]
- 213. Basile, B.; Brown, N.; Valdes, J.M.; Cardarelli, M.; Scognamiglio, P.; Mataffo, A.; Rouphael, Y.; Bonini, P.; Colla, G. Plant-based biostimulants as sustainable alternative to synthetic growth regulators in two sweet cherry cultivars. *Plants* 2021, 10, 619. [CrossRef]
- 214. Testani, E.; Campanelli, G.; Leteo, F.; Ciaccia, C.; Canali, S.; Tittarelli, F. Sweet pepper (*Capsicum annuum* L.) organic seedling production: The role of compost, cultivar, and protein hydrolyzate. *Compost Sci. Util.* **2017**, *25*, 112–119. [CrossRef]
- 215. Elwaziri, E.; Ismail, H.; El-Khairi, E.-S.A.; Al-Qahtani, S.M.; Al-Harbi, N.A.; El-Gawad, H.G.A.; Omar, W.A.; Abdelaal, K.; Osman, A. Biostimulant application of whey protein hydrolysates and potassium fertilization enhances the productivity and tuber quality of sweet potato. *Not. Bot. Horti Agrobot.* **2023**, *51*, 13122. [CrossRef]
- 216. Raguraj, S.; Kasim, S.; Md Jaafar, N.; Nazli, M.H. Growth of tea nursery plants as influenced by different rates of protein hydrolysate derived from chicken feathers. *Agronomy* **2022**, *12*, 299. [CrossRef]
- Francesca, S.; Cirillo, V.; Raimondi, G.; Maggio, A.; Barone, A.; Rigano, M.M. A novel protein hydrolysate-based biostimulant improves tomato performances under drought stress. *Plants* 2021, 10, 783. [CrossRef]
- 218. Barrada, A.; Delisle-Houde, M.; Nguyen, T.T.A.; Tweddell, R.J.; Dorais, M. Drench application of soy protein hydrolysates increases tomato plant fitness, fruit yield, and resistance to a Hemibiotrophic pathogen. *Agronomy* **2022**, *12*, 1761. [CrossRef]
- Ceccarelli, A.V.; Miras-Moreno, B.; Buffagni, V.; Senizza, B.; Pii, Y.; Cardarelli, M.; Rouphael, Y.; Colla, G.; Lucini, L. Foliar application of different vegetal-derived protein hydrolysates distinctively modulates tomato root development and metabolism. *Plants* 2021, *10*, 326. [CrossRef] [PubMed]

- 220. Tallarita, A.V.; Vecchietti, L.; Golubkina, N.A.; Sekara, A.; Cozzolino, E.; Mirabella, M.; Cuciniello, A.; Maiello, R.; Cenvinzo, V.; Lombardi, P.; et al. Effects of plant biostimulation time span and soil electrical conductivity on greenhouse tomato Miniplum yield and quality in diverse crop seasons. *Plants* **2023**, *12*, 1423. [CrossRef]
- 221. Choi, S.; Colla, G.; Cardarelli, M.; Kim, H.-J. Effects of plant-derived protein hydrolysates on yield, quality and nitrogen use efficiency of greenhouse grown lettuce and tomato. *Agronomy* **2022**, *12*, 1018. [CrossRef]
- Francesca, S.; Najai, S.; Zhou, R.; Decros, G.; Cassan, C.; Delmas, F.; Ottosen, C.-O.; Barone, A.; Rigano, M.M. Phenotyping to disset the biostimulant action of a protein hydrolysate in tomato plants under combined abiotic stress. *Plant Physiol. Biochem.* 2022, 179, 32–43. [CrossRef]
- 223. Munaro, D.; Mazo, C.H.; Bauer, C.M.; Gomes, L.D.S.; Teodoro, E.B.; Mazzarino, L.; Veleirinho, M.B.D.R.; Silva, S.M.; Maraschin, M. A novel biostimulant from chitosan nanoparticles and microalgae-based protein hydrolysate: Improving crop performance in tomato. *Sci. Hortic.* 2024, 323, 112491. [CrossRef]
- 224. Zhou, W.; Zheng, W.; Lv, H.; Wang, Q.; Liang, B.; Li, J. Foliar application of pig blood-derived protein hydrolysates improves antioxidant activities in lettuce by regulating phenolic biosynthesis without compromising yield production. *Sci. Hortic.* 2022, 291, 110602. [CrossRef]
- 225. Wang, W.; Zheng, W.; Lv, H.; Liang, B.; Jin, S.; Li, J.; Zhou, W. Animal-derived plant biostimulant alleviates drought stress by regulating photosynthesis, osmotic adjustment, and antioxidant systems in tomato plants. *Sci. Hortic.* **2022**, *305*, 111365. [CrossRef]
- 226. Rouphael, Y.; Colla, G.; Giordano, M.; El-Nakhel, C.; Kyriacou, M.C.; Pascale, S.D. Foliar applications of a legume-derived protein hydrolysate elicit dose-dependent increases of growth, leaf mineral composition, yield and fruit quality in two greenhouse tomato cultivars. *Sci. Hortic.* 2017, 226, 353–360. [CrossRef]
- 227. Casadesus, A.; Perez-Llorca, M.; Munne-Bosch, S.; Polo, J. An enzymatically hydrolyzed animal protein-based biostimulant (Pepton) increases salicylic acid and promotes growth of tomato roots under temperature and nutrient stress. *Front. Plant Sci.* 2020, 11, 953. [CrossRef] [PubMed]
- 228. Caruso, G.; El-Nakhel, C.; Rouphael, Y.; Comite, E.; Lombardi, N.; Cuciniello, A.; Woo, S.L. *Diplotaxis tenuifolia* (L.) DC. yield and quality as influenced by cropping season, protein hydrolysates and *Trichoderma* applications. *Plants* **2020**, *9*, 697. [CrossRef]
- 229. Mironenko, G.A.; Zagorskii, I.A.; Bystrova, N.A.; Kochetkov, K.A. The effect of a biostimulant based on a protein hydrolysate of a rainbow trout (*Oncorhynchus mykiss*) on the growth and yield of wheat (*Triticum aestivum* L.). *Molecules* 2022, 27, 6663. [CrossRef]
- 230. Trakselyte-Rupsiene, K.; Juodeikiene, G.; Cernauskas, D.; Bartkiene, E.; Klupsaite, D.; Zadeike, D.; Bendoraitiene, J.; Damasius, J.; Ignatavicius, J.; Sikorskaite-Gudziuniene, S. Integration of ultrasound into the development of plant-based protein hydrolysate and its bio-stimulatory effect for growth of wheat grain seedlings in vivo. *Plants* 2021, 10, 1319. [CrossRef] [PubMed]
- 231. El-Sanatawy, A.M.; Ash-Shormillesy, S.M.A.I.; El-Yazied, A.A.; El-Gawad, H.G.A.; Azab, E.; Gobouri, A.A.; Sitohy, M.; Osman, A. Enhancing grain yield and nitrogen accumulation in wheat plants grown under a Mediterranean arid environment by foliar spray with papin-released whey peptides. *Agronomy* 2021, *11*, 1913. [CrossRef]
- 232. Chwil, S.; Matraszek, R.; Kozlowska-Strawska, J.; Chwil, M.; Zapalski, P. Effects of protein hydrolysate on soil fertility and heavy-metal accumulation in *Sinapis alba* L. *Commun. Soil Sci. Plant Anal.* **2016**, *47*, 298–304. [CrossRef]
- 233. Bajpai, M.; Pande, A.; Tewari, S.K.; Prakash, D. Phenolic contents and antioxidant activity of some food and medicinal plants. *Int. J. Food Sci. Nutr.* **2005**, *56*, 287–291. [CrossRef]
- 234. Owen-Going, T.N.; Beninger, C.W.; Sutton, J.C.; Hall, J.C. Accumulation of phenolic compounds in plants and nutrient solution of hydroponically grown peppers inoculated with *Pythium aphanidermatum*. *Can. J. Plant Pathol.* 2008, 30, 214–225. [CrossRef]
- Prakash, D.; Suri, S.; Upadhyay, G.; Singh, B.N. Total phenol, antioxidant and free radical scavenging activities of some medicinal plants. *Int. J. Food Sci. Nutr.* 2007, 58, 18–28. [CrossRef]
- Wong, J.Y.; Matanjun, P.; Ooi, Y.B.H.; Chia, K.F. Characterization of phenolic compounds, carotenoids, vitamins and antioxidant activities of selected Malaysian wild edible plants. *Int. J. Food Sci. Nutr.* 2013, *5*, 621–631. [CrossRef] [PubMed]
- 237. Bhattacharya, A.; Sood, P.; Citovsky, V. The roles of plant phenolics in defence and communications during Agrobacterium and Rhizobium infection. *Mol. Plant Pathol.* **2010**, *11*, 705–719. [CrossRef] [PubMed]
- Lattanzio, V.; Lattanzio, V.M.T.; Cardinali, A. Role of phenolics in the resistance mechanisms of plants against fungal pathogens and insects. In *Phytochemistry: Advances in Research*; Imperato, F., Ed.; Research Signpost: Kerala, India, 2006; Volume 661, pp. 23–67.
- Sharma, A.; Shahzad, B.; Rehman, A.; Bhardwaj, R.; Landi, M.; Zheng, B. Response of phenylpropanoid pathway and the role of polyphenols in plants under abiotic stress. *Molecules* 2019, 24, 3242. [CrossRef] [PubMed]
- 240. Jain, C.; Khatana, S.; Vijayvergia, R. Bioactivity of secondary metabolites of various plants: A review. *Int. J. Pharm. Sci. Res.* 2019, 10, 494–504.
- Takshak, S.; Agrawal, S.B. Defense potential of secondary metabolites in medicinal plants under UV-B stress. J. Photochem. Photobiol. B Biol. 2019, 193, 51–88. [CrossRef] [PubMed]
- Mandal, S.M.; Chakraborty, D.; Dey, S. Phenolic acids act as signaling molecules in plant-microbe symbioses. *Plant Signal. Behav.* 2010, *5*, 359–368. [CrossRef]
- 243. Kumar, K.; Debnath, P.; Singh, S.; Kumar, N. An overview of plant phenolics and their involvement in abiotic stress tolerance. *Stresses* 2023, *3*, 570–585. [CrossRef]

- 244. Kisiriko, M.; Anastasiadi, M.; Terry, L.A.; Yasri, A.; Beale, M.H.; Ward, J.L. Phenolics from medicinal and aromatic plants: Characterisation and potential as biostimulants and bioprotectants. *Molecules* **2021**, *26*, 6343. [CrossRef]
- 245. Hajiboland, R.; Farhanghi, F. Remobilization of boron, photosynthesis, phenolic metabolism and anti-oxidant defense capacity in boron-deficient turnip (*Brassica rapa* L.) plants. *Soil Sci. Plant Nutr.* **2010**, *56*, 427–437. [CrossRef]
- 246. Schoneberg, T.; Kibler, K.; Sulyok, M.; Musa, T.; Bucheli, T.D.; Mascher, F.; Bertossa, M.; Voegele, R.T.; Vogelgsang, S. Can plant phenolic compounds reduce *Fusarium* growth and mycotoxin production in cereals? *Food Addit. Contamin.* 2018, 35, 2455–2470. [CrossRef] [PubMed]
- 247. Mohamed, M.S.M.; Saleh, A.M.; Abdel-Farid, I.B.; El-Naggar, S.A. Growth, hydrolases and ultrastructure of *Fusarium oxysporum* as affected by phenolic rich extracts from several xerophytic plants. *Pestic. Biochem. Physiol.* **2017**, *141*, 57–64. [CrossRef]
- 248. Lwalaba, J.L.W.; Zvobgo, G.; Mwamba, T.M.; Louis, L.T.; Fu, L.; Kirika, B.A.; Tshibangu, A.K.; Adil, M.F.; Sehar, S.; Mukobo, R.P. High accumulation of phenolics and amino acids confers tolerance to the combined stress of cobalt and copper in barley (*Hordeum vulgare*). *Plant Physiol. Biochem.* **2020**, 155, 927–937. [CrossRef] [PubMed]
- Silva, G.H.; Souza, J.A.R.D.; Macedo, W.R.; Pinto, F.G. Tyrosol, a phenolic compound from *Phomopsis* sp., is a potential biostimulant in soybean seed treatment. *Phytochem. Lett.* 2021, 43, 40–44. [CrossRef]
- Masondo, N.A.; Aremu, A.O.; Rengasamy, K.R.R.; Amoo, S.O.; Gruz, J.; Subrtova, M.; Dolezal, K.; Van Staden, J. Growth and phytochemical response in *Eucomis autumnalis* (Mill.) Chitt. Treated with phenolic biostimulants from brown alga, *Ecklonia* maxima. S. Afr. J. Bot. 2015, 98, 211. [CrossRef]
- 251. Chen, S.; Lin, R.; Lu, H.; Wang, Q.; Yang, J.; Liu, J.; Yan, C. Effects of phenolic acids on free radical scavenging and heavy metal bioavailability in *Kandelia obovata* under cadmium and zinc stress. *Chemosphere* **2020**, 249, 126341. [CrossRef]
- 252. Yahia, Y.; Bagues, M.; Zaghdoud, C.; Al-Amri, S.M.; Nagaz, K.; Guerfel, M. Phenolic profile, antioxidant capacity and antimicrobial activity of *Calligonum arich* L., desert endemic plant in Tunisia. *S Afr. J. Bot.* **2019**, 124, 414–419. [CrossRef]
- Cueto, J.D.; Kosinska-Cagnazzo, A.; Stefani, P.; Heritier, J.; Roch, G.; Oberhansli, T.; Audergon, J.-M.; Christen, D. Phenolic compounds identified in apricot branch tissues and their role in the control of *Monilinia laxa* growth. *Sci. Hortic.* 2021, 275, 109707. [CrossRef]
- 254. Jakubke, H.D.; Jeschkeit, H. Aminokwasy, Peptydy, Bialka, 2nd ed.; Polskie Wydawnictwo Naukowe: Warsaw, Poland, 1989.
- 255. Shahrajabian, M.H.; Khoshkharam, M.; Sun, W.; Cheng, Q. Germination and seedlings growth of corn (*Zea mays* L.) to allelopathic effects of rice (*Oryza sativa* L.). *Trop. Plant Res.* **2019**, *6*, 152–156. [CrossRef]
- Del Bubba, M.; Ancillotti, C.; Checchini, L.; Ciofi, L.; Fibbi, D.; Gonnelli, C.; Mosti, S. Chromium accumulation and changes in plant growth, selected phenolics and sugars of wild type and genetically modified *Nicotiana langsdorfii*. *J. Hazard. Mater.* 2013, 262, 394–403. [CrossRef]
- 257. Kucinska, J.K.; Magnucka, E.G.; Oksinska, M.P.; Pietr, S.J. Bioefficacy of hen feather keratin hydrolysate and compost on vegetable plant growth. *Compost Sci. Utiliz.* 2014, 22, 179–187. [CrossRef]
- Tejada, M.; Rodriguez-Morgado, B.; Paneque, P.; Parrado, J. Effects of foliar fertilization of a biostimulant obtained from chicken feathers on maize yield. *Eur. J. Agron.* 2018, *96*, 54–59. [CrossRef]
- Wong, F.-C.; Chai, T.-T. Bioactive peptides and protein hydrolysates as lipoxygenase inhibitors. *Biology* 2023, 12, 917. [CrossRef]
 [PubMed]

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