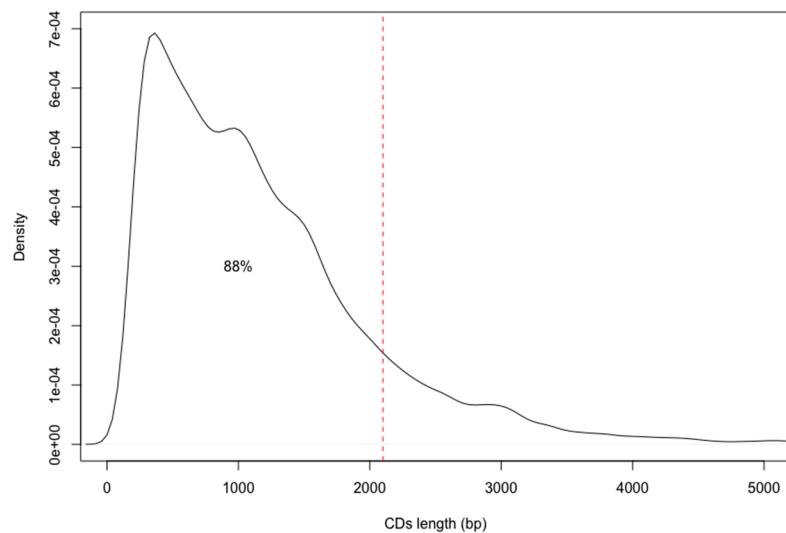
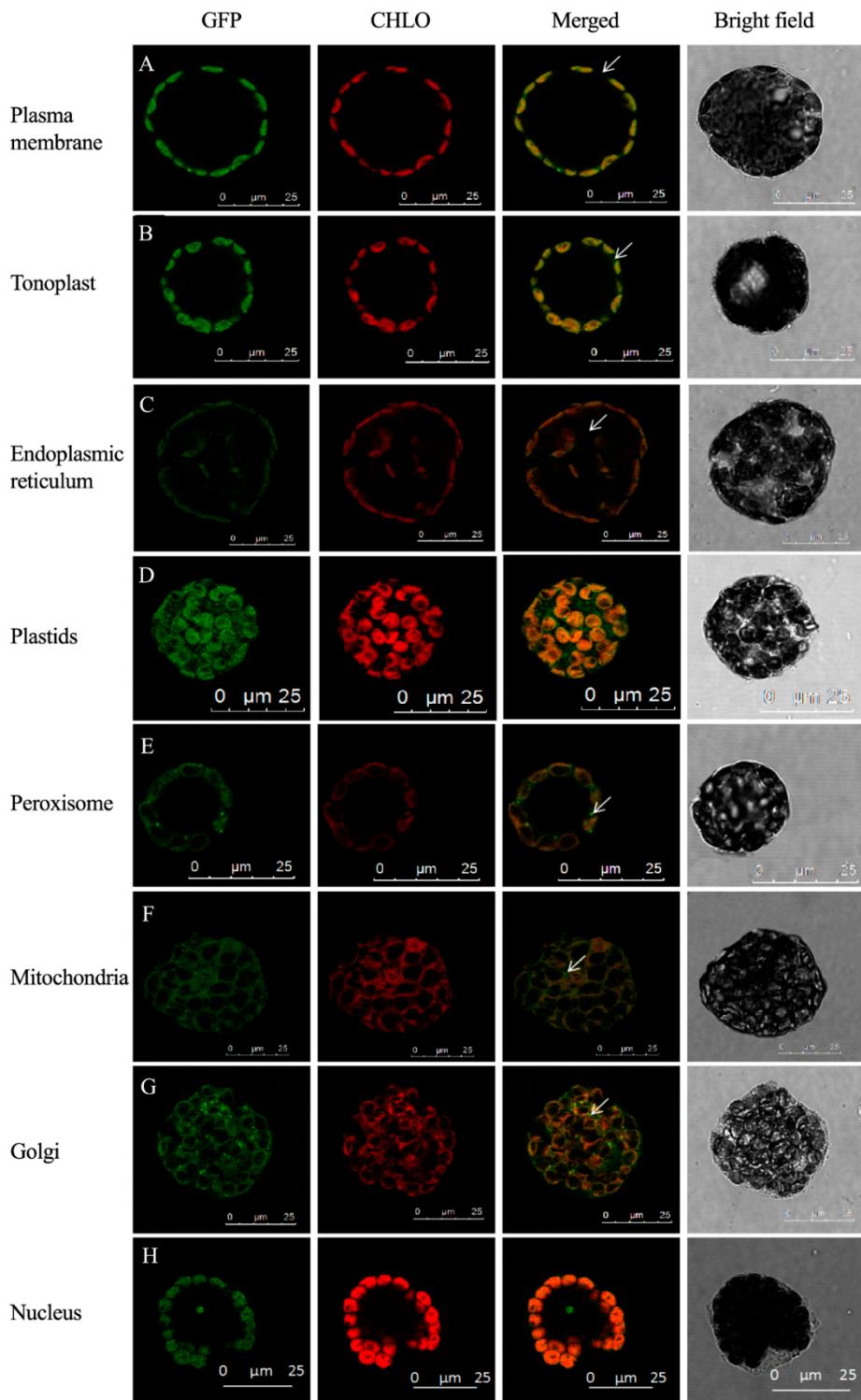


**Figure S1.** Seed imbibition method does not affect cotton growth and can stably express gene over a long period of time. A. The growth of cotton under different treatment methods, from left to right, are soil culture stage, hydroponic stage, and 4-6 true leaf stage; B. Fluorescence of real leaves (left) and cotyledons (right) of cotton in the true leaf stage under ultraviolet light.

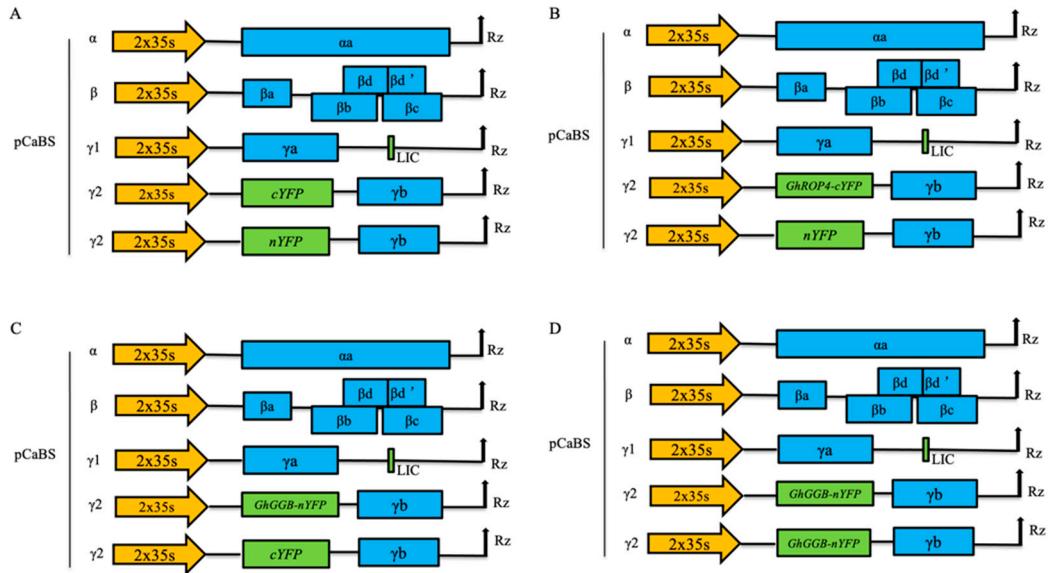


**Figure S2.** Gene length density of cotton.

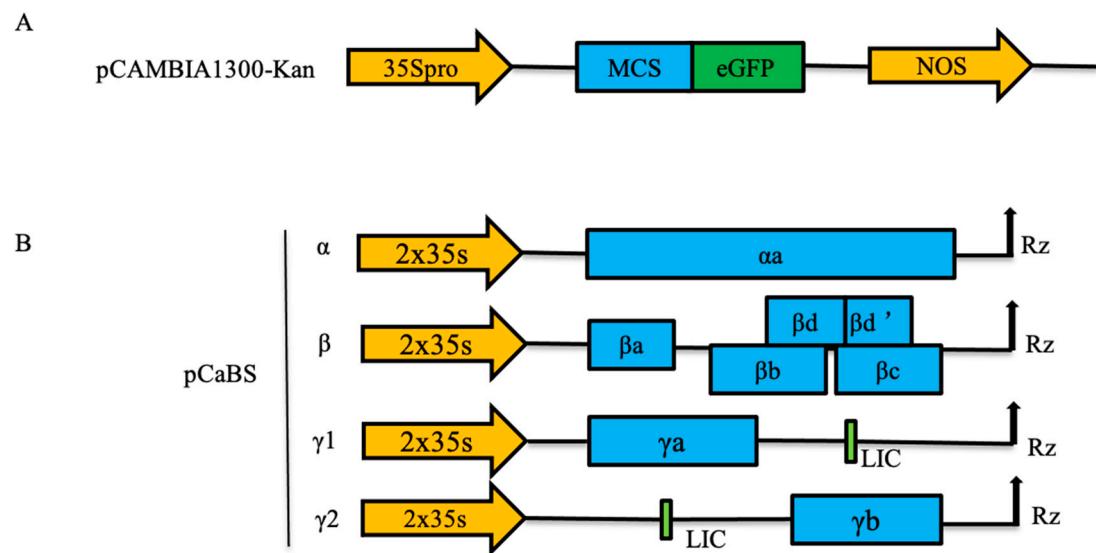


**Figure S3.** Ti vector transient transformation-mediated subcellular localization of GFP fusion-marker genes targeting in cotton protoplast. The localization constructs were transiently expressed in cotton protoplasts by PEG-mediated transformation and observed by confocal microscopy. A. The green fluorescence of *GhPIP2:GFP* was expressed transiently in the plasma membrane; B. The tonoplast marker *GhTIP2:GFP* was found in the membrane protruding into the interior of the cell; C. Fluorescence of endoplasmic reticulum marker *GhSPP:GFP* in cotton protoplasts revealed the typical network morphology of the ER; D. The plastid marker *GhClpD:GFP* in cotton protoplasts showed GFP fluorescence completely overlapped with chloroplast auto-fluorescence; E. The fluorescence of the peroxisomal marker *GhAPX3:GFP* showed as spherical spots localized near chloroplasts; F. The typical granular, thread and randomly distributed mitochondria were observed in leaf protoplasts about marker *GhALDH2:GFP*; G.

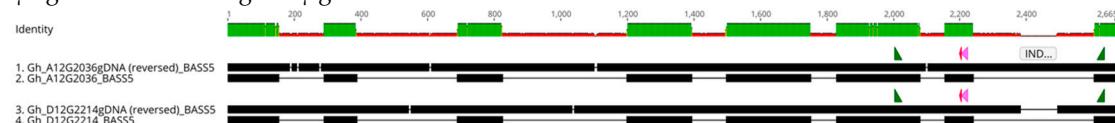
The fluorescence of the Golgi marker *GhMNS1:GFP* showed a punctate staining pattern smaller than 1  $\mu\text{m}$ ; H. The green fluorescence of the final marker *GhTAF2:GFP* was well targeted to the nucleus of the protoplasts.



**Figure S4.** Schematic representation of BSMV recombinant constructs used for BiFC assay. A. The pCaBS- $\gamma$ 2:nYFP (amino acids 1 to 156; nYFP) + pCaBS- $\gamma$ 2:cYFP (amino acids 157 to 239; cYFP); B. pCaBS- $\gamma$ 2:*GhROP4-cYFP* + pCaBS- $\gamma$ 2:nYFP; C. pCaBS- $\gamma$ 2:*GhGGB-nYFP* and pCaBS- $\gamma$ 2:cYFP; D. pCaBS- $\gamma$ 2:*GhROP4-cYFP* + pCaBS- $\gamma$ 2:*GhGGB-nYFP*.



**Figure S5.** Schematic representation of the expression construct and virus vector. A. Schematic representation of the expression vector; B. Schematic representation of the four-component BSMV system. In the pCaBS- $\alpha$ , pCaBS- $\beta$ , pCaBS- $\gamma$ 1, and pCaBS- $\gamma$ 2 vectors, the  $\alpha$ ,  $\beta$ ,  $\gamma$ 1, and  $\gamma$ 2 cDNAs were cloned between the double cauliflower mosaic virus 35S promoter and a ribozyme sequence (Rz) in the pCass4-Rz plasmid. A LIC cloning site containing an *Apal* site was inserted into the  $\gamma$ 2 genome to substitute the  $\gamma$ b genes from the original  $\gamma$  genome.



**Figure S6.** The alignment of cDNAs and genome DNA sequence between *GhBASS5A* and *GhBASS5D*. The black boxes and lines indicated exons and introns, respectively; the two green triangles showed the

position of primers for CAPs analysis; the purple arrows indicated the PAM site; the gray INDEL box represented the 109 bp indel sequence between *gGhBASS5A* and *gGhBASS5D*.

**Table S1.** Cotton organelle marker genes and their primer information (sequence underlined is the position of the introduced cleavage site).

Organelles	Cotton label	Primer sequence(5' to 3')	Length	Temperature
Nucleus (NU)	Gh_A07G0281	F: <u>CGAGGATCC</u> ATGAACCACAACCCGCAATCC R: <u>GCATCTAGA</u> ATTCCCTCTAGAACGGGATCG	411 bp	62 °C
Endoplasmic reticulum (ER)	Gh_A05G0593	F: <u>CGCGGATCC</u> ATGAAGAACACTGAAAGACTGCC' R: <u>TGCTCTAGA</u> TACATCAAATCTCAATGCTAGGGC'	765 bp	62 °C
Plasma membrane (PM)	Gh_D03G1822	F: <u>CGAGGATCC</u> ATGACTAAGGATATTGAGACCACGG R: <u>GCATCTAGAA</u> AGCATGCTCTGAAAGATCCAAGG	876 bp	61 °C
Mitochondria (MT)	Gh_A12G2471	F: <u>CGAGGATCC</u> ATGGCAGCTCGTAGAATCTCTTC R: <u>GCATCTAGACA</u> ACCATGCTGGATTCTCAAAGG	1623 bp	62 °C
Tonoplast (TP)	Gh_A04G1393	F: <u>CGATGGATCC</u> ATGCCGATCAGAACATAGCAG R: <u>GCATCTAGAA</u> ATAATCGGTGGTGGGAGCTGCTCG	756 bp	62 °C
Plastids (PL)	Gh_A09G2205	F: <u>CGCGGATCC</u> ATGGAGGTTTATCTTCTCGTCTC R: <u>TGCTCTAGA</u> TGTACCAAGATCCTATAAGTGTGTGG	1200 bp	64 °C
Golgi body (GB)	Gh_A02G0907	F: <u>CGCGGATCC</u> ATGGCGAGGAGTAGATCATCGTCAT R: <u>TGCTCTAGA</u> CAGTAGTAGTATCCCAAGCAGGTAG	628 bp	62 °C
Peroxisome (PR)	Gh_A03G1812	F: <u>CGAGGATCC</u> ATGGCITTCCAGTAGTCGATACCG R: <u>GCATCTAGAA</u> CTTCATTCTTGCGGACCTCGT	867 bp	63 °C

**Table S2.** LIC primers and qPCR primers used in this study

Primer	Primer sequence(5' to 3')	Length
LIC-NU-GFP	F:AAGGAAGTTAAATGAACCACAACCCGCAAT R:CGGGCCAGCCACCGCCACCAAGTTACTTGTACAGCTCGTCCATGC	1137 bp
LIC-MT-GFP	F:AAGGAAGTTAAATGGCAGCTCGTAGAATC R:CGGGCCAGCCACCGCCACCAAGTTACTTGTACAGCTCGTCCATGC	2340 bp
LIC-ER-GFP	F:AAGGAAGTTAAATGAAGAACACTGAAAGAC R:CGGGCCAGCCACCGCCACCAAGTTACTTGTACAGCTCGTCCATGC	1491 bp
LIC-PM-GFP	F:AAGGAAGTTAAATGGAGGGTAAAGAAGAAG R:CGGGCCAGCCACCGCCACCAAGTTACTTGTACAGCTCGTCCATGC	1593 bp
LIC-TP-GFP	F:AAGGAAGTTAAATGCCGATCAGAACATAG R:CGGGCCAGCCACCGCCACCAAGTTACTTGTACAGCTCGTCCATGC	1482 bp
LIC-PL-GFP	F:AAGGAAGTTAAATGGAGGTTTATCTTCT R:CGGGCCAGCCACCGCCACCAAGTTACTTGTACAGCTCGTCCATGC	1926 bp
LIC-GB-GFP	F:AAGGAAGTTAAATGGCAGGAGTAGATCAT R:CGGGCCAGCCACCGCCACCAAGTTACTTGTACAGCTCGTCCATGC	1354 bp
LIC-PR-GFP	F:AAGGAAGTTAAATGGCGTTCCAGTAGTC R:CGGGCCAGCCACCGCCACCAAGTTACTTGTACAGCTCGTCCATGC	1593 bp
LIC-GFP	F:AAGGAAGTTAAATGGTGAGCAAGGGCGAG R:CGGGCCAGCCACCGCCACCAAGTTACTTGTACAGCTCGTCCATGC	720 bp
LIC-GhBASS5-GFP	F:AAGGAAGTTAAATGAGTTAACCAACTGG R:CGGGCCAGCCACCGCCACCAAGTTACTTGTACAGCTCG	1950 bp
qPCR-GFP	F:GGTGATGTTAATGGGCAC R:TCAGGCATGGCACTTGA	212 bp
UBQ7	F:AGAGGTGAGTCTTCGGACA R:GCTTGATCTTCTGGGCTTG	146 bp
A-GFP	F:ATGGTGAGCAAGGGCGAG R:TTACTTGTACAGCTCGTCCATGC	720 bp

**Table S3.** Primers used for detection of gene editing

Primer	Primer sequence(5' to 3' )	Purpose of primers
<i>GhBASS5-BamHI</i>	F:GTTATTCCTCAGTGGTCACT R:AGCGAAGCCCATTCAATGACA	Amplification of DNA fragment
BSMV- $\alpha$	F:CCATCGTCGATTCCGTGGAT R:CCTCGCATTTGCATCAGCTC	RT-PCR
BSMV- $\beta$	F:GCACCCCTAGAACGTGAACGA R:AGCGAAGCAGTCTTGTCC	RT-PCR
BSMV- $\gamma 1$	F:TTCGACTTCAAGTACCCCGC R:TAGTCGAATCAGTAGCAACC F:GCTCTAGAATGGATTACAAGGACCACGAC R:CGAGCTCGCTCACCTGAGCCTTCTGGA	RT-PCR
Cas9N	F:GCTCTAGAATGGCCAGGGGGACTCGCTG R:CGAGCTCTCACCTCTTCTTCGCGCTGC	RT-PCR
Cas9C	F:AAAGGAAGTTAAATGGATTACAAGGACCACGAC R:CGGGCCAGCCACCGCCACCAAGTTCACTCACCTGAGCCTCTGGA	Clone $\gamma 2$ -Cas9N
$\gamma 2$ -Cas9N	F:AAAGGAAGTTAAATGGCCAGGGGGACTCGCTG R:CGGGCCAGCCACCGCCACCAAGTTCACCTCTTCTTCGCGCTGCC	Clone $\gamma 2$ -Cas9C
$\gamma 2$ -Cas9C		

Video S1. Time series of fluorescence signal of GhBASS5-GFP in cotton leaf.