

Review

Circulating Biomarkers Involved in the Development of and Progression to Chronic Pancreatitis—A Literature Review

Valborg Vang Poulsen ¹, Amer Hadi ¹, Mikkel Parsberg Werge ¹, John Gásdal Karstensen ^{1,2} and Srđan Novovic ^{1,2,*}

¹ Pancreatitis Center East, Gastrounit, Copenhagen University Hospital—Amager and Hvidovre, 2000 Copenhagen, Denmark; valborg.vang.poulsen.02@regionh.dk (V.V.P.); amer.hadi@regionh.dk (A.H.); mikkel.parsberg.werge@regionh.dk (M.P.W.); john.gasdal.karstensen@regionh.dk (J.G.K.)
² Department of Clinical Medicine, University of Copenhagen, 2000 Copenhagen, Denmark
 * Correspondence: srđan.novovic@regionh.dk; Tel.: +45-38621350

Abstract: Chronic pancreatitis (CP) is the end-stage of continuous inflammation and fibrosis in the pancreas evolving from acute- to recurrent acute-, early, and, finally, end-stage CP. Currently, prevention is the only way to reduce disease burden. In this setting, early detection is of great importance. Due to the anatomy and risks associated with direct sampling from pancreatic tissue, most of our information on the human pancreas arises from circulating biomarkers thought to be involved in pancreatic pathophysiology or injury. The present review provides the status of circulating biomarkers involved in the development of and progression to CP.

Keywords: acute pancreatitis; chronic pancreatitis; fibrosis; inflammation; oxidative stress



Citation: Poulsen, V.V.; Hadi, A.; Werge, M.P.; Karstensen, J.G.; Novovic, S. Circulating Biomarkers Involved in the Development of and Progression to Chronic Pancreatitis—A Literature Review. *Biomolecules* **2024**, *14*, 239. <https://doi.org/10.3390/biom14020239>

Academic Editors: Paulina Dumnicka and Beata Kusnierz-Cabala

Received: 29 December 2023

Revised: 13 February 2024

Accepted: 16 February 2024

Published: 18 February 2024



Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Background

Acute pancreatitis (AP), chronic pancreatitis (CP), and pancreatic ductal adenocarcinoma (PDAC) place a significant burden on healthcare systems worldwide. AP is among the three most common benign gastrointestinal diseases, with a mortality rate of 0.9% and an estimated economic burden of USD2.6 billion per year in the US [1]. CP is characterized by gradual irreversible damage to the endocrine and exocrine parenchyma caused by inflammation and subsequent replacement of these tissues with fibrotic tissue and atrophy [2]. Over the last two decades, the incidence of CP has increased by 50%, and there are currently no treatments available to alter this disease's course, resulting in significantly reduced life expectancy and quality of life. Prevention is the only way to reduce the disease burden, as serious complications including exocrine pancreatic insufficiency, malabsorption, diabetes mellitus, and PDAC may evolve as this disease progresses [3].

Approximately 50% of patients with CP have a history of AP [3]. There is continual replacement of the pancreatic tissue with fibrosis. Individuals who experience first-time AP have a 22% chance of developing recurrent acute pancreatitis (RAP) [4], and patients who experience three episodes of RAP have a 16% chance of developing CP. In addition, patients with four or more episodes of RAP have a much higher risk, around 50%, of developing CP [5].

Thus, the continuum from the first episode of AP to the manifestation of CP provides a framework for epidemiologic studies and the time-dependent evolution of circulating biomarkers involved in the progression of this disease.

This review aims to compile and synthesize findings on existing human studies on biomarkers thought to be involved in the development and progression of CP.

2. Materials and Methods

The literature search was conducted using PubMed. The search was performed to a cut-off date of 1 September 2023 to ensure the inclusion of the most relevant and up-to-date

studies. The main search included a combination of text words and MeSH terms: (inflammation, oxidative stress, and fibrosis), combined with chronic pancreatitis and (serum, plasma, or biomarker). Additionally, cross-references were identified manually through the citation list of selected studies to capture additional sources. To ensure completeness, a search was conducted on PubMed for each biomarker mentioned in the review.

We included human studies that compared blood/serum/plasma biomarkers of inflammation, fibrosis, and oxidative stress in patients with CP compared to healthy controls. Animal studies, studies evaluating tissue biopsy biomarkers, and studies evaluating cancer biomarkers were excluded. All the figures included were generated using BioRender.com.

3. Results

We identified 96 studies spanning from 1981 to 2022, examining 126 different biomarkers, with 59 being examined multiple times. Tables 1–3 provide an overview of the biomarkers examined at least twice, with brief additional information, their potential role in pancreatitis, and their levels in patients with CP. Table 4 provides an overview of additional biomarkers only examined once.

3.1. Inflammation

From 1994 to 2022, 55 articles examined 64 inflammatory biomarkers, of which 23 were examined multiple times. In addition, five of these studies included patients with AP [6–10]. The key findings are summarized in Table 1. Of the 23 biomarkers, 12 were either not elevated in CP or the findings were inconclusive. Several pro-inflammatory interleukins (IL-6, IL-8, and IL-12) were found to be elevated in patients with CP compared to healthy controls [11–13], along with vascular endothelial growth factor (VEGF), intercellular adhesion molecule (ICAM), chemerin, fractalkine, resistin, osteopontin, and neopterin [7,10,14–18]. In contrast, leptin was found to be reduced in patients with CP [16]. IL-1 β , IL-6, IL-10, tumor necrosis factor α (TNF- α), adiponectin, and leptin were the most studied inflammatory biomarkers in CP. However, the findings were cohesive only for IL-6, TNF- α , and leptin. IL-10, IL-12, TNF- α , and INF- γ were elevated in patients with AP [6,8]. Figure 1 demonstrates a schematic overview of the inflammatory biomarkers involved in the progression to CP.

3.1.1. Interleukin 6

IL-6 induces the synthesis of acute-phase proteins and the production of other cytokines, including C-reactive protein (CRP) [19]. Sixteen studies measured IL-6, with twelve observing higher levels in patients with CP compared to healthy controls [6,11,12,20–28], although the difference was not significant in three studies [20,25,28]. Four studies found no difference in the IL-6 levels [27,29–31]. In one study, a surge in IL-6 serum levels was observed in patients with alcoholic CP after the consumption of alcohol, with a decrease to the pre-stimulatory levels after 4–24 h, suggesting a correlation between alcohol consumption and IL-6 levels [24]. Elevated IL-6 levels were also evident in AP [6]. IL-6 rises 1–2 days before CRP, making it suitable for an early distinction between severe and mild AP [32,33]. Higher concentrations of IL-6 are linked to the increased risk of complications and death in severe AP [34–37].

3.1.2. Tumor Necrosis Factor α

TNF- α is a cytokine that facilitates both inflammation and fibrosis formation. It plays a key role in regulating other cytokines towards inflammation and activating pancreatic stellate cells (PSCs). TNF- α triggers the activation of PSCs, which, in turn, start producing extracellular matrix (ECM). This disorganization of the ECM leads to fibrosis formation and chronic inflammation of the pancreas [38,39]. The levels of TNF- α in patients with CP were investigated in 12 studies from 1999 to 2022. Elevated levels were found in six studies [9,11,22,31,39–41], one found lower levels [9], while the remaining five found no significant differences [8,20,25,29,42]. Kiyici et al. discovered significantly higher serum

levels of TNF- α in AP compared to CP, indicating TNF- α 's potential role in the progression of the disease. However, it is worth noting that this study included only 13 patients with AP, 36 patients with CP, and 14 controls [8].

3.1.3. Leptin

Leptin, an adipokine with a crucial role in metabolism, obesity, and cardiovascular diseases, has also been found to activate macrophages and T-lymphocytes, stimulating their cytokine secretion [43]. Moreover, it has been demonstrated to induce fibrosis in the liver by inhibiting hepatic stellate cell apoptosis [16,44,45]. Five studies found reduced levels of leptin in patients with CP [16,46–49], while one study found elevated leptin levels compared to healthy controls [41]. Because leptin is secreted by adipocytes, a higher fat percentage results in a higher amount of circulating leptin. Patients with CP had lower BMI across the involved studies, making it difficult to determine if the reduced levels were due to pancreatitis or to a lower fat mass. Additionally, patients with CP with diabetes mellitus (DM) were found to have higher levels of leptin than patients with CP without DM [41]. Lower levels of leptin may play a protective role in the development of CP by increasing apoptosis of the PSCs.

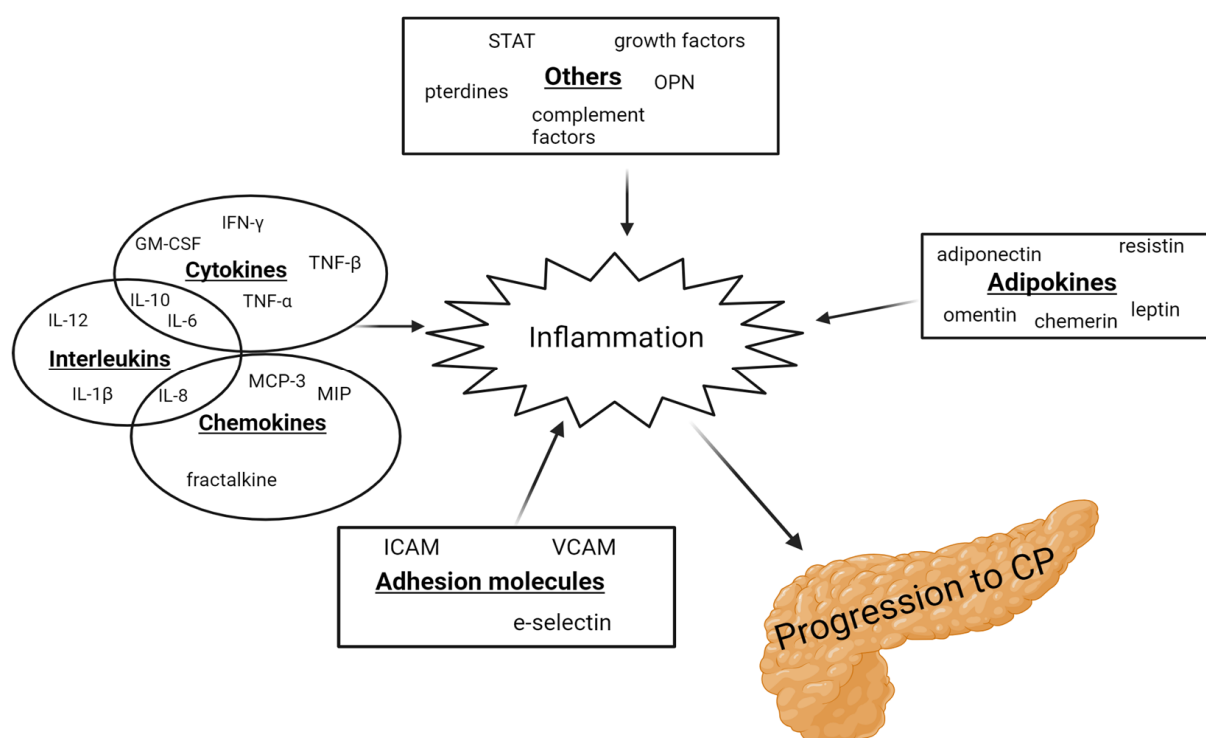


Figure 1. Schematic illustration of the inflammatory biomarkers involved in the progression to CP. GM-CSF: granulocyte-macrophage colony-stimulating factor; ICAM: intracellular adhesion molecule; IFN: interferon; IL: interleukins; MCP: monocyte chemotactic protein; MIP: macrophage inflammatory protein; OPN: osteopontin; STAT: signal transducer and activator of transcription; TNF: tumor necrosis factor; and VCAM: vascular cell adhesion molecule.

Table 1. An overview of the inflammatory biomarkers involved in the development and progression of chronic pancreatitis. (Symbols in the column “Blood Levels in Patients with CP” correspond to the reference in the column “Biomarkers”; =: no difference; ↓: reduced levels; and ↑: elevated levels).

Biomarkers	Mechanism	Pancreas-Specific Effects	Blood Levels in Patients with CP	Comment
IL-1β [12,22,29–31,50]	A pro-inflammatory cytokine that activates several intracellular responses, e.g., stimulation of IL-6, IL-8, and TNF-α [51,52].	Excessive or prolonged IL-1β activation can lead to CP [52,53]. Activates proliferation and collagen secretion in fibroblasts [54].	= = ↓ ↓ ↑ =	Overexpression of IL-1β in murine pancreas results in CP [53]. Increases protease inhibitors having a protective effect in CP [30].
IL-1α [9,29,50]	A pro-inflammatory cytokine that induces inflammation via activation of, e.g., COX2, IL-6, and TNF-α [55].	Not typically associated with the pancreas but can indirectly be involved in pancreatic diseases.	↓ ↓ =	
IL-1Ra [12,29]	An IL-1 receptor antagonist. Anti-inflammatory cytokine with protumor activity [12].	Has a protective effect on both AP and CP [56,57].	↑ =	Higher levels in PDAC compared to CP [29].
IL-2 [6,29,58]	A potent Th1-related cytokine that acts on NK cells and T-cells [59].	Increases T-cells in the pancreas and induces expression of T-cell-associated proteins [60].	↑ ↓ ↓	
IL-2R [20,61–63]	IL-2 receptor		↑ = = =	
IL-4 [6,9,29]	Modulates the differentiation of precursor Th cells to Th2 cells [64]; inhibition of pro-inflammatory cytokine synthesis [65].	Secreted by PSCs, mediates macrophage activation by participating in the promotion of pancreatic fibrosis [66].	↓ ↑ ↓	Potentially, levels of IL-4 in patients with CP depend on whether inflammation or fibrosis is the dominant process.
IL-6 [6,11,12,20–31,67]	A pro-inflammatory cytokine that causes cell proliferation, differentiation, and inflammatory responses and triggers the synthesis of acute-phase proteins [19,30].	Promotes PSCs activation and collagen synthesis through the upregulation of TGF-β1 [68].	↑ ↑ ↑ = ↑ ↑ ↑ ↑ = ↑ = = = ↑	Levels are closely linked to the quantity of alcohol consumed by patients with alcoholic CP [24]. Elevated in AP and reflects the severity and prognosis of the pancreatitis [37].
IL-8 [9,11,12,31]	A chemoattractant that acts as a neutrophil activator and a pro-angiogenic factor [9,69].	Circulating neutrophils from patients with CP express mRNA for IL-8 [69]. High levels of IL-8 are found in CP tissue [69–71].	↑ ↑ ↑ ↑	Depending on the etiology, the amount of IL-8 correlates with the severity of the pancreatitis [69–71].
IL-10 [6,23,29,31,58]	An anti-inflammatory cytokine that inhibits cytokine release from lymphocytes, e.g., IL-2, IL-6, and TNF-α [13,72].	Has a protective effect on the pancreas during inflammation. The absence of IL-10 prevents the downregulation of inflammation [72,73].	↓ ↑ ↑ = =	Is seen to have a protective effect on the pancreas in mice [74].
IL-12 [6,9,13,29]	Activates Th1-cells and induces the secretion of cytokines, e.g., INF-γ, IL-2, and TNF-α [6].	The level escalates during the transition from AP to CP. Increased levels in both conditions [6].	↑ ↑ ↑ ↓	Potential role in the progression of the disease [6].
IL-17 [23,75]	A pro-inflammatory cytokine with a key role in the initial immune response [76].	Triggers damage to pancreatic acinar cells by producing and releasing cytokines/chemokines recruiting immune cells [76].	= ↑	Valuable severity and prognostic factor in AP progression [77].
GM-CSF [46,78]	A growth and differentiation factor for granulocytes and macrophages [79].	Regulates cancer-associated inflammation in PDAC [80].	↓ ↑	
IFN-γ [6,13,29]	A pro-inflammatory cytokine produced by activated T-cells and NK-cells, with chemotactic abilities [81].	Stimulated by upregulated IL-18 and IL-12 in CP [6]. Elevated levels were found in CP tissue [82–84].	↑ = ↓	Potential role in the progression of pancreatitis [6].

Table 1. Cont.

Biomarkers	Mechanism	Pancreas-Specific Effects	Blood Levels in Patients with CP	Comment
TNF- α [8,9,11,20,22,25,29,31,39–42]	Regulates cytokines and adhesion molecules; also, a priming activator of inflammatory cells and PSCs [38].	Induces PSC activation and collagen synthesis leading to fibrosis and inflammation in the pancreas [40].	= \downarrow \uparrow = \uparrow = = \uparrow \uparrow \uparrow \uparrow =	Elevated levels are also seen in patients with AP [33,36].
ICAM [15,85–87]	An adhesion molecule that serves to mediate the adhesion of immune cells to endo-/epithelial cells [88].	Overexpression of ICAM-1 in pancreatic endothelial cells leads to inflammatory cell infiltration in the pancreatic parenchyma [88].	\uparrow = \uparrow \uparrow	Elevated levels in AP correlate with higher mortality rates and necrosis development [88]
VEGF [14,89]	A pro-angiogenic mediator that enhances vascular permeability and stimulates immune cell migration [90].	Not typically associated with the pancreas, it can indirectly be involved in pancreas diseases.	\uparrow =	
Fractalkine [7,91,92]	Adhesion molecule that can be cleaved and functions as a chemoattractant [93].	Expressed on the cell membranes of PSCs, it induces monocyte recruitment in the inflamed pancreas [7].	\uparrow \uparrow \uparrow	Alcohol consumption influences the levels of fractalkine. One study only found elevation in mild and severe CP [91].
Chemerin [10,94,95]	An adipokine with chemoattractant properties, promotes the differentiation of adipocytes [96].	Promotes the recruitment of macrophages to the inflamed pancreas [96].	\uparrow \uparrow \uparrow	No correlation between chemerin levels and alcohol intake or diabetes [94].
Adiponectin [16,22,41,47,48,97]	An adipokine with anti-inflammatory properties. Reduces the levels of circulating fatty acids, activates their oxidation, and prevents lipid accumulation in cells [98,99].	A lack of adiponectin accelerates the progression of CP in mice [100].	= \uparrow \downarrow = = \uparrow	Levels are inversely proportional to fat percentage.
Leptin [16,41,46–49]	An adipokine with pro-inflammatory and pro-fibrogenic properties [44,45].	Inhibits SC apoptosis; therefore, lower levels are thought to induce SC apoptosis and thereby inhibit fibrosis [16].	\downarrow \uparrow \downarrow \downarrow \downarrow \downarrow	Higher levels were found among patients with CP with DM [41].
Resistin [16,46,101]	An adipokine that acts in a pro-inflammatory manner by upregulating IL-6 and TNF- α [102].	Increases the concentration of TNF- α , which, in turn, activates PSCs [16].	\uparrow \uparrow \uparrow	Higher levels were found among patients with CP with DM [41].
Osteopontin [17,103–106]	A glycoposphoprotein produced and secreted by osteoblasts, activated T cells, macrophages, and others. Functions as a chemoattractant in sites of inflammation [17,107].	May play a part in the calcification and the formation of pancreas calculi [108].	\uparrow \uparrow \uparrow = =	
Neopterin [18,20,62]	A compound secreted by activated macrophages stimulated by INF- γ [20].	A marker of the cellular immunity mediated by the lymphocyte–macrophage axis [20].	= \uparrow \downarrow	Elevated in patients with AP and can reflect the severity and prognosis of AP [109,110].

DM: diabetes mellitus; GM-CSF: granulocyte-macrophage colony-stimulating factor; ICAM: intracellular adhesion molecule; IFN- γ : interferon- γ ; NK: natural killer; PDAC: pancreatic ductal adenocarcinoma; and VEGF: vascular endothelial growth factor.

3.2. Fibrosis

A total of 46 studies spanning from 1995 to 2022 examined 28 potential biomarkers of fibrosis in patients with CP. Of these, 13 were studied multiple times. Three studies also included patients with AP [7,10,111]. Table 2 provides an overview of the examined biomarkers. The biomarkers mainly consist of PSCs activators, with the most extensively studied being TGF- β , PDGF, and MIC-1, and components of the ECM, with TIMP-1 and MMP-9 being studied the most. Figure 2 demonstrates a schematic overview of the fibrotic biomarkers associated with the development of CP.

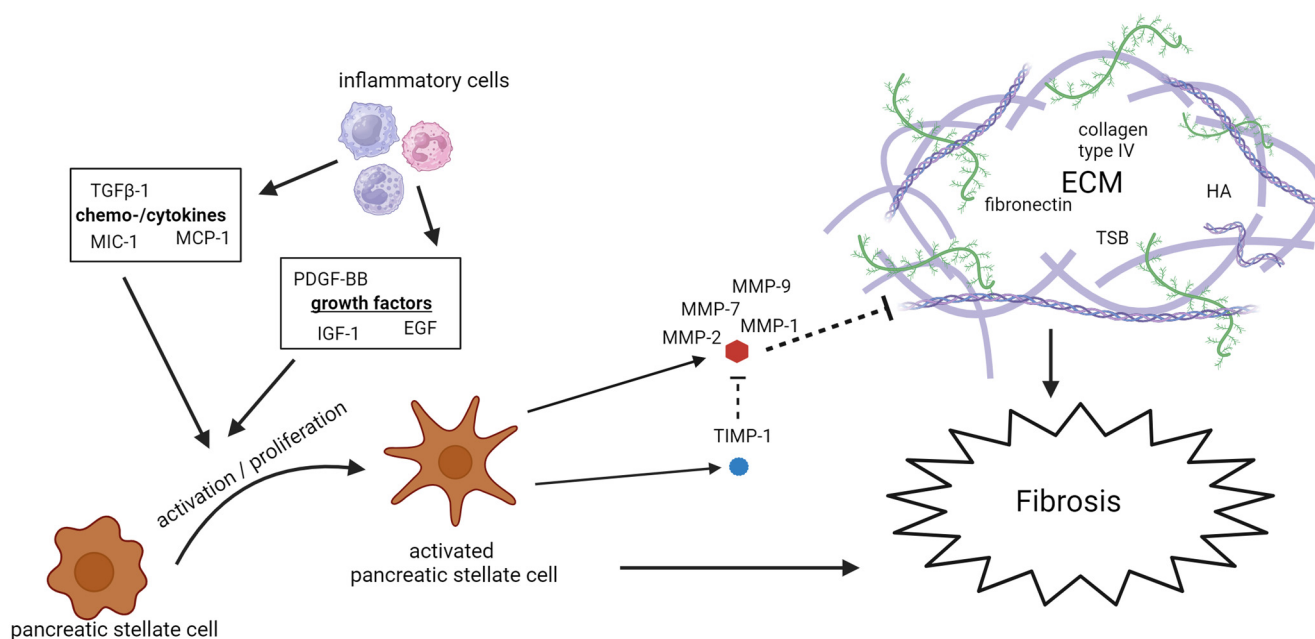


Figure 2. Schematic overview of the fibrotic biomarkers associated with the development of CP. ECM: extracellular matrix; EGF: epidermal growth factor; HA: hyaluronic acid; IGF: insulin-like growth factor; MCP: monocyte chemotactic protein; MIC: macrophage inhibitory cytokine; MMP: matrix metalloproteinase; PDGF: platelet-derived growth factor; TGF: tumor growth factor; TIMP: tissue inhibitors of metalloproteinases; and TSP: tissue polypeptide specific antigen.

3.2.1. Extracellular Matrix Remodeling

Continuous modulation of the ECM leads to fibrosis. Matrix metalloproteinases (MMPs) degrade the ECM, while tissue inhibitors of matrix metalloproteinases (TIMPs) inhibit MMPs. Numerous studies have measured the concentration of these biomarkers in patients with CP. MMPs are included in seven of the studies we reviewed [39,47,48,87,112–114]. Elevated levels of MMP-1, MMP-2, MMP-7, and MMP-9 were found in patients with CP compared to the control group, while MMP-3 was not seen to be elevated in patients with CP.

TIMP-1 concentrations in patients with CP were studied in nine of the studies we reviewed [15,48,85,87,105,106,113,115,116], all showing elevated concentrations in patients with CP, although three did not reach significance [48,85,105]. Hyaluronic acid (HA), laminin, and fibronectin are also important components of the ECM and are directly associated with the potential role of the ECM in the context of CP, see Figure 2. Elevated levels of all these components of the ECM were found in patients with CP. Four studies found elevated levels of HA [92,101,117,118], a fundamental component of the ECM in the pancreas. The Mac-2-binding protein (M2BP), a ligand which binds to ECM proteins and a novel biomarker of liver fibrosis, has also been found to be elevated in patients with CP. Additional biomarkers of the ECM and their potential role in CP are shown in Table 4.

3.2.2. Activation of PSCs

The activation and proliferation of PSCs influence the development of pancreatic fibrosis by the synthesis and remodeling of the ECM. The remodeling of the ECM is primarily mediated through the PSCs' secretion of MMP and TIMP [119].

The cytokine transforming growth factor β 1 (TGF- β 1), the growth factor platelet-derived growth factor (PDGF), and the chemokine monocyte chemoattractant protein 1 (MCP-1) are among the most important mediators involved in the activation of PSCs. With few exceptions, these biomarkers are all found to be elevated in patients with CP compared to the controls. Elevated levels of TGF- β , PDGF, and MCP-1 were also found in patients with AP [7,10,111]. Macrophage inhibitory cytokine 1 (MIC-1), a cytokine part of the TGF- β family, has also been found to be elevated in patients with CP in five different studies. Its specific role in the pancreas is not extensively studied, but, as a part of the TGF- β family, it can be presumed that it has a role in the activation of PSCs.

Some of the inflammatory cytokines listed in Table 1, especially IL-4, IL-6, and TNF- α , also play a major role in pancreatic fibrosis formation. Additional activators and proliferators of PSCs are listed in Tables 2 and 4.

Table 2. An overview of the fibrotic biomarkers involved in the development and progression of chronic pancreatitis. (Symbols in the column “Blood Levels in Patients with CP” correspond to the reference in the column “Biomarkers”; =: no difference; ↓: reduced levels; and ↑: elevated levels).

Biomarkers	Mechanism	Pancreas-Specific Effects	Blood Levels in Patients with CP	Comment
MMP-7 [48,87] MMP-9 [39,112–114]	Enzymes secreted by activated PSCs that degrade the ECM [120].	Degradation of basement collagen (type IV) [39,112].	= ↑ ↑↑ = ↑	One study found elevated levels in the plasma and not in the serum [114].
TIMP-1 [15,48,85,87,105,106,113,115,116]	Enzymes secreted by activated PSCs that inhibit MMPs [120].	Inhibits the proteolytic activity of MMPs. An imbalance between MMP And TIMPs supports the abnormal formation of the ECM [120].	↑↑ = ↑ = ↑↑ =	mRNA expression in the pancreas increases with disease progression [121].
HA [92,101,117,118]	A protein component of the ECM [101].	Marker of ECM proliferation.	↑↑↑ =	
TGF-β [7,10,24,91,92,94,101,117,122,123]	A multipotent growth factor, with various functions, e.g., cell differentiation, proliferation, matrix production, and apoptosis. Promotes the recruitment of inflammatory cells and contributes to fibrosis [124].	Activates PSCs leading to fibrosis formation in CP [124].	↑↑ = ↑↑↑↑↑↑↑ =	Higher in patients with pancreatic atrophy than in patients with a non-atrophic pancreas [7]. Correlates with the severity of alcoholic CP [91].
PDGF [10,12,46,89,94,117,122]	A growth factor and mitogen acting on fibroblasts and promoting cell proliferation and migration [125].	Acts as a growth factor on PSCs leading to ECM formation and, consequently, fibrosis [101].	↑ = ↓ = ↑↑ =	One paper studied PDGF-AA [122]. No correlation between PDGF-BB and alcohol intake [117].
MCP-1 [7,9,12,24,25,29,48,91,92,101,111]	A chemoattractant that recruits an inflammatory infiltrate and initiates inflammation [24,126].	Activates PSCs via TNF-β and promotes pancreatic fibrosis [101].	= ↓ = = ↑ = = ↑↑↑	Negative association with alcohol [24]. Treatment with MCP-1 antagonist in rats inhibits pancreatic fibrosis [7].
MIC-1 [17,106,127–129]	Part of the TGF-β family. An autocrine regulator of macrophage activation [130].	The specific mechanism in the pancreas is not clear [106,129].	↑↑↑↑↑ =	Further elevated in patients with PDAC, making it a potential biomarker [106].
M2BP [131,132]	A ligand that binds to extracellular proteins such as integrins, collagens, and fibronectin [133].	Suggested to be associated with cell-to-cell and cell-to-ECM adhesion and plays a role in the facilitation of fibrosis [134].	↑↑	A novel biomarker of liver fibrosis [135,136].
ET-1 [22,137]	A mediator with vasoconstrictive and pro-inflammatory properties, secreted by damaged endothelial cells [137].	Affects the activation of PSCs and stimulates the migration of PSCs [138].	= =	Elevated levels seen in smokers [137].
EGF [29,89,139]	A growth factor that stimulates the proliferation of, e.g., fibroblasts and epithelial cells [140].	Regulates both chemoattraction and stimulation of the proliferation of PSCs [141].	↑↓↑	
IGF-1 [24,48,50,139,142–144]	A growth factor that plays an important role in many bioactivities such as cell proliferation, differentiation, and survival [145].	Stimulates migration and proliferation of PSCs [146].	= = = = ↑ = =	One study found reduced levels of IGF-1R [144].
IGFBP-2 [48,142]	Insulin growth factor-binding protein 2		↑↑	

ECM: extracellular matrix; EGF: epidermal growth factor; ET-1: endothelin; HA: hyaluronic acid; IGF: insulin-like growth factor; IGFBP: insulin-like growth factor-binding protein; MCP: monocyte chemoattractant protein; MIC: macrophage inhibitory cytokine; MMP: matrix metalloproteinase; M2BP: mac-2-binding protein; PDAC: pancreatic adenocarcinoma; PDGF: platelet-derived growth factor; TGF: transforming growth factor; and TIMP: tissue inhibitor of metalloproteinases.

3.3. Oxidative Stress

Twenty-three studies from 1981 to 2022 examined 34 biomarkers of oxidative stress, of which 23 were studied multiple times. Four articles also included patients with AP [147–150]. A potential relationship between oxidative stress and pancreatic inflammation has been extensively studied. Research indicates an early occurrence of pancreatic oxidative stress in AP. Free oxygen radicals play a crucial role in regulating the extent of necrosis in acinar cells, the development of pancreatic edema, the sequestration of inflammatory cells within the pancreas, and the release of inflammatory mediators [151]. Additionally, there is growing evidence connecting oxidative stress and CP. The use of antioxidant therapy has been shown to reduce the severity of CP, resulting in less fibrosis in murine models [152], as well as improve the well-being, decrease pain, and improve the overall functioning of patients with CP [153,154]. The biomarkers of oxidative stress are challenging to evaluate, primarily due to their complex metabolism and high turnover, making them difficult to measure in systemic circulation. Most of the biomarkers included in the present review are, therefore, indirect markers of oxidative stress. The low blood antioxidant levels could be attributed to poor nutritional status due to malabsorption, maldigestion, and reduced food intake, often observed in patients with CP. Figure 3 gives a schematic overview of oxidative stress biomarkers associated with CP development.

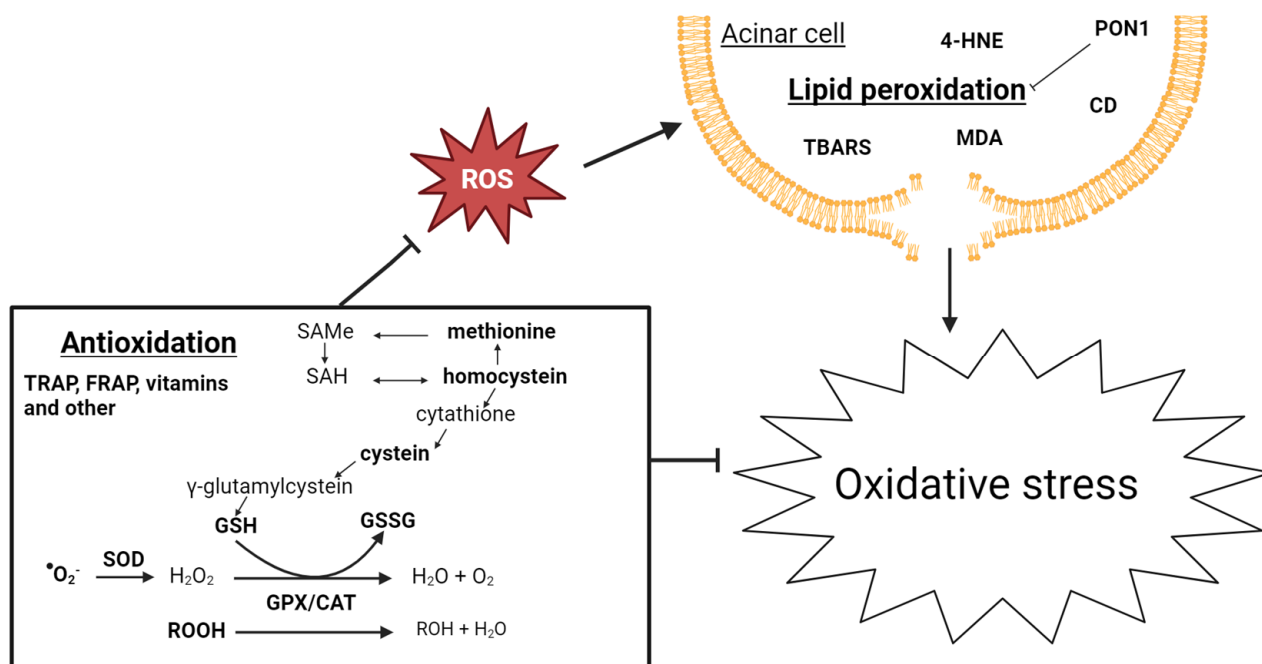


Figure 3. Schematic overview of biomarkers of oxidative stress associated with CP development. The biomarkers included in this review are marked with a bold font. CAT: catalase; GPX: glutathione peroxidase; GSH: glutathione; GSSG: glutathione disulfide; ROOH: hydroperoxides; SAH: S-adenosyl homocysteine; S-AdoMet: S-adenosyl methionine; and SOD: superoxide dismutase.

3.3.1. Lipid Peroxidation

Prolonged exposure to oxygen radicals results in lipid peroxidation and the oxidation of fatty acids in cell membranes. Lipid peroxidation has been the focus of several studies investigating oxidative stress in CP [155]. Many of the biomarkers in Table 3 are primarily byproducts of lipid peroxide; these include TBARS, 4-HNE, MDA, and CD. Fourteen studies have included one of these biomarkers, and, apart from a few non-significant results, all these biomarkers are found to be elevated in patients with CP. Few studies include oxygen radicals in patients with CP. Superoxides, main reactive oxygen species (ROS) in cells, and ROS production in cells after phorbol myristate acetate (PMA) stimulation have all been investigated in patients with CP. These biomarkers are difficult to measure in the

blood as they have a very short half-life. Elevated levels were found in all four studies; however, in two of them, the elevation was not significant. On the other hand, PON1 is a free radical-scavenging molecule contributing to the detoxification of free radicals involved in lipid peroxidation [156], and consistently reduced levels of PON1 were found in patients with CP in the studies we analyzed, indicating elevated lipid peroxidation in patients with CP. MDA, superoxide anion, and CAT were also found to be elevated in AP [147–150].

3.3.2. Antioxidation

The reactive superoxide is catalyzed to hydrogen peroxide by superoxide dismutase (SOD). The main ROS scavenger molecule is glutathione (GSH), which is used by glutathione peroxidase (GPX) and catalase (CAT) to reduce/neutralize ROS [157]; see Figure 3. GSH, GPX, CAT, and SOD have been measured in patients with CP in, respectively six, eight, four, and six studies. GSH, GPX, and SOD levels were reduced in patients with CP, while the results on CAT were contradictory, as two studies found no difference, one found lower levels, and one study found elevated levels. The antioxidant capacity in the blood is measured by the ferrin-reducing ability of the plasma (FRAP) and the total peroxyl radical-trapping antioxidant parameter (TRAP). While FRAP is reduced in patients with CP, the TRAP concentrations are no different from the controls. Reduced levels of GSH and elevated levels of CAT and TRAP were found in patients with AP [147–149].

Table 3. An overview of the oxidative stress biomarkers involved in the development and progression of CP. (Symbols in the column “Blood Levels in Patients with CP” correspond to the reference in the column “Biomarkers”; =: no difference; ↓: reduced levels; and ↑: elevated levels).

Biomarkers	Mechanism	Blood Levels in Patients with CP	Comment
TBARS [122,147,158–162]	A byproduct of the lipid peroxidation process [163].	↑↑↑↑↑↑↑ =	TBARS are higher in patients with TCP than in patients with ACP [160].
4-HNE [164,165]	A byproduct of the lipid peroxidation process [166].	↑↑	Also elevated in RAP, especially during attacks on AP [164].
MDA [149,164,165]	One of the final products of lipid peroxidation [164].	↑ = ↑	Elevated levels are also found in pancreatic tissue samples [149].
CD [155,157]	Primary products in lipid peroxidation in cells [157].	= ↑	Elevated levels are also found in pancreatic tissue samples [149].
ROS [158,167]	Reactive oxygen species	= ↑	Difficult to measure in the blood due to a short half-life.
O ₂ [−] [148,167]	Reactive oxygen species molecule [148].	↑↑	Elevated in both PMA-stimulated and resting neutrophils [167].
GSH [157,159–161,168]	The main ROS scavenger. Used by GPX to metabolize H ₂ O ₂ and lipid hydroperoxides to water/alcohols [157].	= ↓↓↓↓	
GPX [157,159,160,162,164,167,169,170]	Catalyzes hydrogen peroxide to oxygen and water and, therefore, has an important function in the protection against oxidative stress [167].	↓↓↓↓↓ = ↓	
CAT [150,157,167,169]		= = ↑↓	
SOD [157,159–161,164,167,169]	Catalyzes the dismutation of superoxide anions to hydrogen peroxide [157].	= ↓↓ = ↑ =	One study found elevated serum SOD and lower levels of erythrocyte SOD in patients with CP [161].
PON1 [156,157]	An HDL-associated enzyme. Plays a role in the hydrolyzation of active oxidized phospholipids and in the destruction of lipid hydroperoxides and H ₂ O ₂ and prevents oxidation of LDL [156].	↓↓	
TRAP [147,165]	Total peroxyl radical-trapping antioxidant parameter.	= =	
FRAP [122,158,161,164]	Ferrin-reducing ability of the plasma. A measurement of the non-enzymatic antioxidant capacity of the plasma [158].	↓↓↓↓	Lower levels are also observed in patients with RAP [164].
Vitamin A [42,161,162,169,171]	Blood antioxidant	↓↓↓↓↓	Dietary-dependent
Vitamin C [147,159–161,164,169]	Blood antioxidant	↓↓↓ = = =	Dietary-dependent
Vitamin E [42,161,162,169–171]	Blood antioxidant	↓↓↓↓↓	Dietary-dependent
Zink [162,169,171]	Blood antioxidant	= = =	Elevated levels in patients with RAP [171].
Copper [162,169,171]	Induces oxidative stress by increasing ROS [172].	↑↑ =	Reduced levels in patients with RAP [171].
Selenium [162,169,171]	Blood antioxidant	↓↓↓	
Homocysteine [158,168]	Amino acid mediator in the synthesis of GSH.	= ↑	
Cysteine [158,168]	Essential amino acid necessary for the formation of GSH.	↓	
Methionine [49,168,173]	Essential amino acid necessary for the formation of GSH.	= ↓↓	One study only found elevated levels in TCP [168].
β-carotene [156,169,171]	Blood antioxidant	↓↓↓	

ACP: alcoholic chronic pancreatitis; CAT: catalase; CD: conjugated dienes; FRAP: ferrin-reducing ability of the plasma; GPX: glutathione peroxidase; GSH: glutathione; MDA: malondialdehyde; O₂[−]: ion oxide; PON1: paraoxonase; PMA: phorbol myristate acetate; RAP: recurrent acute pancreatitis; ROS: reactive oxygen species; SOD: superoxide dismutase; TBARS: Thiobarbituric acid-reactive substances; TCP: tropical chronic pancreatitis; TRAP: total peroxyl radical-trapping antioxidant parameter; and 4-HNE: 4-hydroxynonenal.

Table 4. An overview of the biomarkers examined once.

Inflammation						Fibrosis			Oxidative stress						
Group	Biomarker	Expression Changes in CP	Group	Biomarker	Expression Changes in CP	Group	Biomarker	Expression Changes in CP	Group	Biomarker	Expression Changes in CP				
Interleukins	IL-5 [9]	ns	Adhesion molecules	CD44 [62]	↓	Components of the ECM	Collagen IV [87]	↑	Antioxidants	GR [157]	ns				
	IL-7 [9]	ns		e-selectin [22]	ns		Fibronectin [87]	↑		Xanthine [171]	↓				
	IL-13 [9]	ns		VCAM [22]	ns		Laminin [117]	↑		B-cyproxanthine [171]	↓				
	IL-15 [29]	↓	C1q [9]	↓	MMP-1 [112]		↑	Lycopene [171]		↓					
	IL-16 [9]	ns	C3 [9]	ns	MMP-2 [47]		↑	SH groups [147]	↓						
	IL-18 [13]	↑	Complement factors	C4 [9]	↑		MMP-3 [112]	ns	Lipid peroxidation	Ox-LDL/LDL [157]	↑				
	IL-23 [75]	↑		C4BPA [174]	↑		PICP [87]	ns		ROOH [170]	↑				
Cytokines	TNF-β [9]	↓	Adipokines	C5 [9]	↑	Others	PINP [87]	ns		Protein damage	Lipid peroxide [170]	↑			
	GCSF [78]	ns		pro-C3 [118]	ns		THBS1 [85]	ns	3-NT [157]		↑				
	MCSF [78]	ns		Pro-C5 [175]	ns		TPS [18]	↑	Carbonyls [158]	↑					
	IFN-α [29]	ns		Properdin [9]	↓		TSP-2 [87]	↑	Others	Nitrites [165]	↑				
Chemokines	CCL5 [85]	ns	Adipokines	Omentin [95]	↑	Others	AZGP1 [85]	↑							
	CXCL16 [176]	ns		ANG-1 [89]	↓		CCN1 [87]	ns							
	IP10 [12]	ns		HMGB1 [10]	ns		CCN2 [87]	↑							
	MCP-3 [9]	ns		LBP [85]	ns		PLG [87]	ns							
	MIP-1β [12]	ns	Others	LTF [85]	↑										
	MIP-3α [11]	↑		RORγT [75]	↑										
	PPBP [85]	ns		STAT3 [75]	↑										
	RBP-4 [143]	ns		YKL-40 [26]	ns										
Growth factors	IGF-2 [48]	ns		CD40L [29]	↑										
	IGFBP1,3 [48]	↑													

ns: non-significant; ANG-1: angiopoietin; AZGP: zinc-α2-glycoprotein 1; CCL: C-C motif chemokine ligand; CCN: cellular communication network; CXCL: C-X-C motif ligand; G-CSF: granulocyte colony-stimulating factor; GR: glutathione reductase; HGMB: high-mobility group box; IGF: insulin-like growth factor; IGFBP: insulin-like growth factor-binding protein; IL: interleukin; INF: interferon; IP: induced protein; LBP: lipopolysaccharide-binding protein; LTF: lactoferrin; MCP: monocyte chemotactic protein; MCSF: macrophage colony-stimulating factor; MIP: macrophage inflammatory protein; MMP: matrix metalloproteinase; PLG: plasminogen; PICP/PICP: procollagen type I C-terminal propeptide/N-terminal propeptide; PPBP: pro-platelet basic protein; RBP: retinol-binding protein; ROOH: lipid hydroperoxide; RORγ: retinoic acid receptor-related orphan receptor gamma; SH groups: thiols; STAT: signal transducer and activator of transcription; THBS: thrombospondin; TNF: tumor necrosis factor; TSP-2: tissue polypeptide specific antigen; TSP: thrombospondin; VCAM: vascular cell adhesion protein; and 3-NT: 3-nitrotyrosine.

4. Discussion

The identification of a biomarker for the early diagnosis of CP is of great importance, as it has the potential to make a significant impact on disease prevention and intervention. Moreover, an early-stage CP biomarker could facilitate clinical trials with anti-fibrotic, anti-inflammatory, and antioxidative therapies. Several biomarkers of inflammation, fibrosis, and oxidative stress have been studied so far to improve our understanding of the transition from AP to CP. Herein, we present a general overview of established inflammatory, fibrotic, and oxidative stress biomarkers associated with the progression to CP as well as a brief presentation of the most promising biomarkers. It is important to note that 50% of patients with CP have not had an episode of AP prior to their diagnosis; therefore, some of the biomarkers associated with CP might not be associated with AP [3].

Studies on CP biomarkers present varying results. Some inflammatory biomarkers have been linked to CP development, mainly IL-6, IL-8, IL-10, IL-12, TNF- α , ICAM, fractalkine, and some adipokines. All the findings are not coherent; therefore, we found no conclusive inflammatory pattern of CP. Several biomarkers of fibrosis are used to determine fibrotic development in patients with CP. The main biomarkers of circulating ECM components are MMPs, TIMP-1, and HA, and the main stimulators of PSCs are TGF- β , PDGF-BB, and MCP-1. In addition, oxidative stress is increased, and antioxidant capacity is lowered in CP. The exact impact of oxidative stress on disease progression and development is still not thoroughly understood.

CP is histologically characterized by the loss of acinar cells, irregular interlobular fibrosis, infiltration of inflammatory cells, and, eventually, also by the loss of intralobular ducts and islets [177]. The pattern of fibrosis varies depending on the type of cell primarily affected by the injury. Different causes of CP result in different fibrotic patterns. For instance, alcoholic CP is more likely to exhibit inter(perilobular) fibrosis, whereas nonalcoholic CP more frequently shows periductal or intralobular fibrosis [119]. After exposure to risk factors, such as smoking, alcohol, intraductal obstruction, or other injuries, oxidative stress occurs, leading to the necrosis and apoptosis of pancreatic cells. This triggers the activation and infiltration of inflammatory cells. Once the inflammatory cells are activated, they secrete cytokines, growth factors, and other molecules that promote the differentiation and activation of the PSCs. Activated PSCs excessively secrete and synthesize extracellular matrix (ECM). PSCs cause ECM to be continuously remodeled, resulting in fibrosis. Activated PSCs secrete matrix metalloproteinases (MMPs), enzymes which degrade the ECM, and tissue inhibitors of MMPs (TIMPs), enzymes which inhibit MMPs. An imbalance between MMPs and TIMPs can lead to the abnormal formation of the ECM [119]. Oxidative stress occurs when ROS damage cell lipids and proteins. Lipid peroxidation is the process by which ROS interact with polyunsaturated fatty acids. Lipid peroxidation in cell membranes causes cell damage [158]. Antioxidants neutralize ROS molecules, having a protective effect on CP. The processes of fibro-inflammation and oxidative stress are intertwined, as oxidative stress causes inflammation but also directly activates PSCs [178], while inflammation causes the formation of ROS and activates PSCs; see Figure 4.

This review presents promising CP biomarkers. The three most promising biomarkers of inflammation seem to be IL-6, IL-8, and fractalkine, meanwhile, those for fibrosis are TGF- β 1, HA, and MIC-1 and, for oxidative stress, TBARS, FRAP, and GPX. However, these candidates require further investigation before they can be implemented in clinical practice.

Several methodological limitations must be taken into consideration when interpreting the findings of current research efforts. The studies included in this review have marked variability in their design, population, and CP diagnosis criteria. The lack of standardization and proper characterization of patients included in studies can lead to biased results and inaccurate conclusions. Disease stage standardization is particularly crucial, as biomarker patterns may differ depending on whether samples were collected in the early or later stages of the disease or during or between flare-ups. Due to the fact that CP can be difficult to identify in its early stage, most clinical studies included patients with advanced CP or

a flare-up, as these patients attend the sites of research for treatment. One study found elevated levels of TGF- β 1 in only mild and moderate CP, the assumed reason for this is the replacement of pancreatic tissue with fibrotic tissue. Advanced CP is characterized by fibrosis formation, which might play a role in the levels of biomarkers, as less pancreatic tissue is present to produce cytokines and other factors. Disease severity should, therefore, be a target for standardization and should at least be characterized in participating patients.

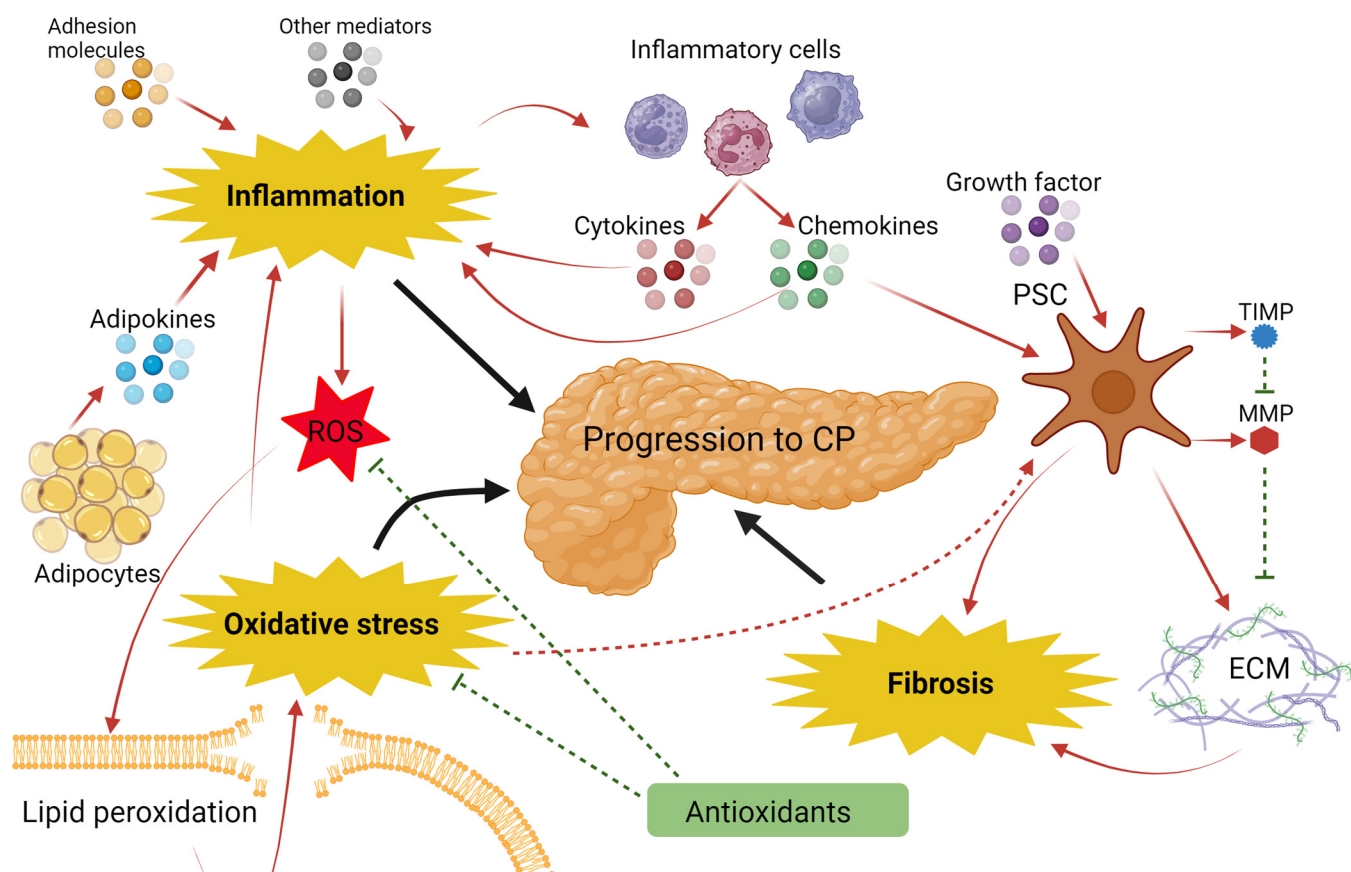


Figure 4. Schematic overview of the fibro-inflammatory and oxidative stress process in the development of CP. Created with BioRender.com. ECM: extracellular matrix; MMP: matrix metalloproteinase; PSC: pancreatic stellate cells; ROS: reactive oxygen species; and TIMP: tissue inhibitor of metalloproteinase.

How biomarkers are measured can also affect the accuracy of the outcome. Measuring biomarkers can be challenging at times, especially when certain markers have a short half-life and are difficult to measure correctly in the plasma or the serum. This can lead to inaccurate results. The studies included in this review have used different methods, which might explain the varying results which the same biomarker has in different studies.

Additionally, CP is a complex disease with different etiologies that may have different sets of biomarker patterns, and factors such as smoking, alcohol, diabetes, age, gender, and BMI are not matched in all the studies, which may lead to confounding effects, as inflammation and oxidative stress may be present due to other causes. It is also important to note that the interaction between cytokines and their specific inhibitors in the plasma makes the accurate analysis and interpretation of cytokine levels and activity difficult.

Many cytokines and MMPs primarily exert paracrine effects, which are not necessarily expressed in the systemic circulation. Oxidative stress is typically localized and organ-specific. By implementing the notion that the levels of certain molecules in the bloodstream may not accurately reflect the specific processes occurring in the organs, it becomes clear that many of the biomarkers included in this review may also be elevated in other diseases

and conditions. Therefore, it is important to consider factors such as smoking, alcohol use, and diabetes when comparing biomarker levels to those of healthy individuals, as these typically are present in some form in patients with CP.

The development of CP can be associated with disturbances in the flow of pancreatic fluid from the acinar cells through the side branches and ducts in the pancreas. The disturbances can be caused by strictures, the formation of intraductal stones, or intraductal hypertension caused by (extra)pancreatic collections. Pancreatic obstruction is linked to enhanced levels of inflammatory, fibrogenic, and oxidative stress markers in the pancreas. Interventional procedures like endoscopic retrograde cholangiopancreatography or extracorporeal shock-wave lithotripsy are often used to reduce pancreatic duct obstruction, thereby reducing intraductal pressure and, presumably, decreasing the levels of the measured biomarkers. Their effect on biochemical patterns in CP has not been extensively studied and remains a topic of interest.

Some of the studies included in the present review also examined biomarkers in patients with AP and RAP. These studies indicate that some of the biomarkers elevated in CP are also elevated in patients with AP (e.g., IL-6, IL-2, MCP-1) and in patients with RAP (e.g., MCP-1, 4-HNE, MDA). This overlap in biomarkers makes it challenging to use them as early predictors of CP. Further prospective evaluation of a large and well-characterized cohort of patients with AP, RAP, and CP may clarify the differences in the expression levels of biomarkers within these separate entities.

The current literature review mainly includes cross-sectional studies on patients with advanced CP, which limits the ability of this work to accurately assess disease progression. Future longitudinal studies including patients with AP, RAP, and early CP are essential in order to provide valuable insights into the time-dependent interplay between inflammatory, fibrogenic, and oxidative stress biomarkers and the development of CP. Such a prospective and prolonged follow-up may give us valuable insight and a rationale for interventional studies [179,180].

CP is a rather heterogenous disease, and, within this diagnosis, several phenotypes as well as clinical challenges exist. Future research should be aimed at associating different phenotypes (obstructive, inflammatory, painful, etc.) with different biomarker profiles.

5. Conclusions

The fibro-inflammatory process involved in the progression from acute to recurrent acute and, ultimately, chronic pancreatitis is epidemiologically and clinically evident. Several biomarkers have been investigated in an attempt to unravel this process, but their exact role in the continuous process of inflammation, fibrosis, and oxidative stress towards CP is not yet fully established. Currently, there is no reliable biomarker specifically indicative of chronic pancreatitis. The development of chronic pancreatitis remains poorly understood, and a better understanding of it may help researchers identify new targets for intervention.

Author Contributions: Conceptualization, S.N.; methodology, V.V.P.; investigation, V.V.P.; writing—original draft preparation, V.V.P.; writing—review and editing, S.N., A.H., J.G.K. and M.P.W.; visualization, V.V.P.; supervision, S.N.; project administration, S.N. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable as no humans or animals were included in this study.

Informed Consent Statement: Not applicable. Patient consent was waived as this is a narrative review of present literature.

Data Availability Statement: No new data were created or analyzed in this study. Data sharing is not applicable to this article.

Conflicts of Interest: The authors declare no conflicts of interest.

References

1. Peery, A.F.; Crockett, S.D.; Murphy, C.C.; Lund, J.L.; Dellon, E.S.; Williams, J.L.; Jensen, E.T.; Shaheen, N.J.; Barritt, A.S.; Lieber, S.R.; et al. Burden and Cost of Gastrointestinal, Liver, and Pancreatic Diseases in the United States: Update 2018. *Gastroenterology* **2019**, *156*, 254–272.e11. [\[CrossRef\]](#)
2. Schneider, A.; Löhr, J.M.; Singer, M.V. The M-ANNHEIM Classification of Chronic Pancreatitis: Introduction of a Unifying Classification System Based on a Review of Previous Classifications of the Disease. *J. Gastroenterol.* **2007**, *42*, 101–119. [\[CrossRef\]](#)
3. Cai, Q.Y.; Tan, K.; Zhang, X.L.; Han, X.; Pan, J.P.; Huang, Z.Y.; Tang, C.W.; Li, J. Incidence, Prevalence, and Comorbidities of Chronic Pancreatitis: A 7-Year Population-Based Study. *World J. Gastroenterol.* **2023**, *29*, 4671–4684. [\[CrossRef\]](#)
4. Sankaran, S.J.; Xiao, A.Y.; Wu, L.M.; Windsor, J.A.; Forsmark, C.E.; Petrov, M.S. Frequency of Progression from Acute to Chronic Pancreatitis and Risk Factors: A Meta-Analysis. *Gastroenterology* **2015**, *149*, 1490–1500.e1. [\[CrossRef\]](#) [\[PubMed\]](#)
5. Hegyi, P.J.; Soós, A.; Tóth, E.; Ébert, A.; Venglovecz, V.; Márta, K.; Mátrai, P.; Mikó, A.; Bajor, J.; Sarlós, P.; et al. Evidence for Diagnosis of Early Chronic Pancreatitis after Three Episodes of Acute Pancreatitis: A Cross-Sectional Multicentre International Study with Experimental Animal Model. *Sci. Rep.* **2021**, *11*, 1367. [\[CrossRef\]](#) [\[PubMed\]](#)
6. Bhatnagar, A.; Wig, J.D.; Majumdar, S. Immunological Findings in Acute and Chronic Pancreatitis. *ANZ J. Surg.* **2003**, *73*, 59–64. [\[CrossRef\]](#) [\[PubMed\]](#)
7. Ito, T. Can Measurement of Chemokines Become Useful Biological and Functional Markers of Early-Stage Chronic Pancreatitis? *J. Gastroenterol.* **2007**, *42*, 72–77. [\[CrossRef\]](#)
8. Kıyıcı, A.; İbiş, M.; Akbulut, Ş.; Köklü, S.; Uçar, E.; Ünlü, A. Serum TNF-Alpha Levels in Acute and Chronic Pancreatitis. *Eur. J. Gen. Med.* **2009**, *6*, 103–107. [\[CrossRef\]](#) [\[PubMed\]](#)
9. Sandström, A.; Andersson, R.; Segersvärd, R.; Löhr, M.; Borrebaeck, C.A.K.; Wingren, C. Serum Proteome Profiling of Pancreatitis Using Recombinant Antibody Microarrays Reveals Disease-Associated Biomarker Signatures. *Proteom. Clin. Appl.* **2012**, *6*, 486–496. [\[CrossRef\]](#)
10. Stojek, M.; Adrych, K.; Rojek, L.; Smoczynski, M.; Sledzinski, T.; Szrok, S.; Swierczynski, J. Decreased Serum Platelet Derived Growth Factor BB Levels in Acute and Increased in Chronic Pancreatitis. *World J. Gastroenterol.* **2014**, *20*, 13127–13132. [\[CrossRef\]](#)
11. Miron, N.; Miron, M.-M.; Milea, V.G.I.; Cristea, V. Proinflammatory Cytokines: An Insight into Pancreatic Oncogenesis. *Rom. Arch. Microbiol. Immunol.* **2010**, *69*, 183–189.
12. Shaw, V.E.; Lane, B.; Jenkinson, C.; Cox, T.; Greenhalf, W.; Halloran, C.M.; Tang, J.; Sutton, R.; Neoptolemos, J.P.; Costello, E. Serum Cytokine Biomarker Panels for Discriminating Pancreatic Cancer from Benign Pancreatic Disease. *Mol. Cancer* **2014**, *13*, 114. [\[CrossRef\]](#) [\[PubMed\]](#)
13. Schneider, A.; Haas, S.L.; Hildenbrand, R.; Siegmund, S.; Reinhard, I.; Nakovics, H.; Singer, M.V.; Feick, P. Enhanced Expression of Interleukin-18 in Serum and pancreas of Patients with Chronic Pancreatitis. *World J. Gastroenterol.* **2006**, *12*, 6507. [\[CrossRef\]](#) [\[PubMed\]](#)
14. Berindan-Neagoe, I.; Burz, C.; Balacescu, O.; Balacescu, L.; Seicean, A.; Cristea, V.; Irimie, A. Molecular Angiogenesis Profile as a Tool to Discriminate Chronic Pancreatitis (CP) from Pancreatic Cancer (PC). *CA Cancer J. Clin.* **2011**, *40*, 482–483. [\[CrossRef\]](#) [\[PubMed\]](#)
15. Jenkinson, C.; Elliott, V.; Menon, U.; Apostolidou, S.; Fourkala, O.E.; Gentry-Maharaj, A.; Pereira, S.P.; Jacobs, I.; Cox, T.F.; Greenhalf, W.; et al. Evaluation in Pre-Diagnosis Samples Discounts ICAM-1 and TIMP-1 as Biomarkers for Earlier Diagnosis of Pancreatic Cancer. *J. Proteom.* **2015**, *113*, 400–402. [\[CrossRef\]](#)
16. Adrych, K.; Smoczynski, M.; Sledzinski, T.; Dettlaff-Pokora, A.; Goyke, E.; Swierczynski, J. Increased Serum Resistin Concentration in Patients With Chronic Pancreatitis Possible Cause of Pancreatic Fibrosis. *J. Clin. Gastroenterol.* **2008**, *43*, 63–68. [\[CrossRef\]](#) [\[PubMed\]](#)
17. Song, J.; Sokoll, L.J.; Pasay, J.J.; Rubin, A.L.; Li, H.; Bach, D.M.; Chan, D.W.; Zhang, Z. Identification of Serum Biomarker Panels for the Early Detection of Pancreatic Cancer. *Cancer Epidemiol. Biomark. Prev.* **2019**, *28*, 174–182. [\[CrossRef\]](#)
18. Talar-Wojnarowska, R.; Gasiorowska, A.; Olakowski, M.; Lekstan, A.; Lampe, P.; Malecka-Panas, E. Clinical Value of Serum Neopterin, Tissue Polypeptide-Specific Antigen and CA19-9 Levels in Differential Diagnosis between Pancreatic Cancer and Chronic Pancreatitis. *Pancreatol.* **2011**, *10*, 689–694. [\[CrossRef\]](#)
19. Manohar, M.; Verma, A.K.; Venkateshaiah, S.U.; Sanders, N.L.; Mishra, A. Pathogenic Mechanisms of Pancreatitis. *World J. Gastrointest. Pharmacol. Ther.* **2017**, *8*, 10–25. [\[CrossRef\]](#)
20. Manes, G.; Spada, O.A.; Rabitti, P.G.; Feola, B.; Misso, S.; Minerva, A.; Uomo, G. Neopterin Serum Levels in Pancreatic Adenocarcinoma. *Int. J. Pancreatol.* **1999**, *25*, 31–37. [\[CrossRef\]](#) [\[PubMed\]](#)
21. Talar-Wojnarowska, R.; Gasiorowska, A.; Smolarz, B.; Romanowicz-Makowska, H.; Kulig, A.; Malecka-Panas, E. Clinical Significance of Interleukin-6 (IL-6) Gene Polymorphism and IL-6 Serum Level in Pancreatic Adenocarcinoma and Chronic Pancreatitis. *Dig. Dis. Sci.* **2009**, *54*, 683–689. [\[CrossRef\]](#) [\[PubMed\]](#)
22. Gasiorowska, A.; Talar-Wojnarowska, R.; Kaczka, A.; Borkowska, A.; Czupryniak, L.; Malecka-Panas, E. Subclinical Inflammation and Endothelial Dysfunction in Patients with Chronic Pancreatitis and Newly Diagnosed Pancreatic Cancer. *Dig. Dis. Sci.* **2016**, *61*, 1121–1129. [\[CrossRef\]](#)
23. Tanțău, A.; Leucuța, D.C.; Tanțău, M.; Boțan, E.; Zaharie, R.; Mândruțu, A.; Tomuleasa, I.C. Inflammation, Tumoral Markers and Interleukin-17, -10, and -6 Profiles in Pancreatic Adenocarcinoma and Chronic Pancreatitis. *Dig. Dis. Sci.* **2021**, *66*, 3427–3438. [\[CrossRef\]](#) [\[PubMed\]](#)

24. Pedersen, N.; Larsen, S.; Seidelin, J.B.; Nielsen, O.H. Alcohol Modulates Circulating Levels of Interleukin-6 and Monocyte Chemoattractant Protein-1 in Chronic Pancreatitis. *Scand. J. Gastroenterol.* **2004**, *39*, 277–282. [\[CrossRef\]](#)
25. Zhang, J.; Fan, H.; Gross, M.; Liu, N.; Carlson, H.; Wood, A.; Hoffman, K.; Petrosino, J.; Pankratz, N.; Thyagarajan, B.; et al. Progressive Reduction in Circulating Levels of Carotenoids and Other Micronutrients in Patients with Chronic Pancreatitis. *Pancreatol.* **2022**, *22*, 1126–1133. [\[CrossRef\]](#)
26. Hansen, M.; Rinnov Nielsen, A.; Vilsbøll, T.; Lund, A.; Krarup, T.; Knop, F.K.; Vestergaard, H. Increased Levels of YKL-40 and Interleukin 6 in Patients With Chronic Pancreatitis and Secondary Diabetes. *Pancreas* **2012**, *41*, 1316–1318. [\[CrossRef\]](#) [\[PubMed\]](#)
27. Mroczko, B.; Groblewska, M.; Gryko, M.; Kędra, B.; Szmitkowski, M. Diagnostic Usefulness of Serum Interleukin 6 (IL-6) and C-Reactive Protein (CRP) in the Differentiation between Pancreatic Cancer and Chronic Pancreatitis. *J. Clin. Lab. Anal.* **2010**, *24*, 256–261. [\[CrossRef\]](#)
28. Singh, N.; Gupta, S.; Rashid, S.; Saraya, A. Association of Inflammatory Markers with the Disease & Mutation Status in Pancreatic Cancer. *Indian J. Med. Res.* **2022**, *155*, 49–55. [\[CrossRef\]](#)
29. Chung, H.W.; Jang, S.; Lim, J.B. Clinical Implications and Diagnostic Usefulness of Correlation between Soluble Major Histocompatibility Complex Class I Chain-Related Molecule a and Protumorigenic Cytokines in Pancreatic Ductal Adenocarcinoma. *Cancer* **2013**, *119*, 233–244. [\[CrossRef\]](#)
30. Bamba, T.; Yoshioka, U.; Hosoda, S. Serum Levels of Interleukin-Lp and Interleukin-6 in Patients with Chronic Pancreatitis. *J. Gastroenterol.* **1994**, *29*, 314–319. [\[CrossRef\]](#)
31. Dima, S.O.; Tanase, C.; Albulescu, R.; Herlea, V.; Chivu-Economescu, M.; Purnichescu-Purtan, R.; Dumitrascu, T.; Duda, D.G.; Popescu, I. An Exploratory Study of Inflammatory Cytokines as Prognostic Biomarkers in Patients With Ductal Pancreatic Adenocarcinoma. *Pancreas* **2012**, *41*, 1001–1007. [\[CrossRef\]](#)
32. Cho, I.R.; Do, M.Y.; Han, S.Y.; Jang, S.I.; Cho, J.H. Comparison of Interleukin-6, C-Reactive Protein, Procalcitonin, and the Computed Tomography Severity Index for Early Prediction of Severity of Acute Pancreatitis. *Gut Liver* **2023**, *17*, 629–637. [\[CrossRef\]](#) [\[PubMed\]](#)
33. Heresbach, D.; Letourneur, J.P.; Bahun, I.; Pagenault, M.; Guillou, Y.M.; Dyard, F.; Fauchet, R.; Mallédant, Y.; Bretagne, J.F.; Gosselin, M. Value of Early Blood Th-1 Cytokine Determination in Predicting Severity of Acute Pancreatitis. *Scand. J. Gastroenterol.* **1998**, *33*, 554–560. [\[CrossRef\]](#) [\[PubMed\]](#)
34. Berney, T.; Gasche, Y.; Robert, J.; Jenny, A.; Mensi, N.; Grau, G.; Vermeulen, B.; Morel, P. Serum Profiles of Interleukin-6, Interleukin-8, and Interleukin-10 in Patients with Severe and Mild Acute Pancreatitis. *Pancreas* **1999**, *18*, 37–38. [\[CrossRef\]](#) [\[PubMed\]](#)
35. Inagaki, T.; Hoshino, M.; Hayakawa, T.; Ohara, H.; Yamada, H.; Iida, M.; Nakazawa, T.; Ogasawara, T.; Uchida, A.; Hasegawa, C.; et al. Interleukin-6 Is a Useful Marker for Early Prediction of the Severity of Acute Pancreatitis. *Pancreas* **1997**, *14*, 1–8. [\[CrossRef\]](#) [\[PubMed\]](#)
36. Brivet, F.G.; Emilie, D.; Galanaud, P. Pro- and Anti-Inflammatory Cytokines during Acute Severe Pancreatitis: An Early and Sustained Response, Although Unpredictable of Death. *Crit. Care Med.* **1999**, *27*, 749–755. [\[CrossRef\]](#) [\[PubMed\]](#)
37. Leser, H.G.; Gross, V.; Scheibenbogen, C.; Heinisch, A.; Salm, R.; Lausen, M.; Rückauer, K.; Andreesen, R.; Farthmann, E.H.; Schölmerich, J. Elevation of Serum Interleukin-6 Concentration Precedes Acute-Phase Response and Reflects Severity in Acute Pancreatitis. *Gastroenterology* **1991**, *101*, 782–785. [\[CrossRef\]](#) [\[PubMed\]](#)
38. Norman, J. The Role of Cytokines in the Pathogenesis of Acute Pancreatitis. *Am. J. Surg.* **1998**, *175*, 76–83. [\[CrossRef\]](#)
39. Manjari, K.S.; Jyothy, A.; Vidyasagar, A.; Prabhakar, B.; Nallari, P.; Venkateshwari, A. Matrix Metalloproteinase-9, Transforming Growth Factor-B1, and Tumor Necrosis Factor- α Plasma Levels in Chronic Pancreatitis. *Indian J. Gastroenterol.* **2013**, *32*, 103–107. [\[CrossRef\]](#)
40. Sri Manjari, K.; Jyothy, A.; Shravan Kumar, P.; Prabhakar, B.; Uma Devi, M.; Ramanna, M.; Nallari, P.; Venkateshwari, A. A Single-Nucleotide Polymorphism in Tumor Necrosis Factor- α (-308 G/A) as a Biomarker in Chronic Pancreatitis. *Gene* **2014**, *539*, 186–189. [\[CrossRef\]](#)
41. Hontsariuk, D.O.; Ferfetska, K.V.; Khrystych, T.M.; Fediv, O.I.; Temerivska, T.G.; Jiguleva, E.O.; Honcharuk, L.M.; Olinik, O.Y. Incides of C-Reactive Protein, Tumor Necrosis Factor- α , Adiponectin, Leptin and Resistin in the Blood of Patients Suffering from Chronic Pancreatitis and Type 2 Diabetes Mellitus. *J. Med. Life* **2020**, *13*, 568–571. [\[CrossRef\]](#)
42. Greer, J.B.; Greer, P.; Sandhu, B.S.; Alkaade, S.; Wilcox, C.M.; Anderson, M.A.; Sherman, S.; Gardner, T.B.; Lewis, M.D.; Guda, N.M.; et al. Nutrition and Inflammatory Biomarkers in Chronic Pancreatitis Patients. *Nutr. Clin. Pract.* **2019**, *34*, 387–399. [\[CrossRef\]](#)
43. Zhao, S.; Kusminski, C.M.; Scherer, P.E. Adiponectin, Leptin and Cardiovascular Disorders. *Circ. Res.* **2021**, *128*, 136–149. [\[CrossRef\]](#)
44. Saxena, N.K.; Titus, M.A.; Ding, X.; Floyd, J.; Srinivasan, S.; Sitaraman, S.V.; Anania, F.A. Leptin as a Novel Profibrogenic Cytokine in Hepatic Stellate Cells: Mitogenesis and Inhibition of Apoptosis Mediated by Extracellular Regulated Kinase (Erk) and Akt Phosphorylation. *FASEB J.* **2004**, *18*, 1612–1614. [\[CrossRef\]](#) [\[PubMed\]](#)
45. Petrescu, A.D.; Grant, S.; Williams, E.; An, S.Y.; Seth, N.; Shell, M.; Amundsen, T.; Tan, C.; Nadeem, Y.; Tjahja, M.; et al. Leptin Enhances Hepatic Fibrosis and Inflammation in a Mouse Model of Cholestasis. *Am. J. Pathol.* **2022**, *192*, 484–502. [\[CrossRef\]](#) [\[PubMed\]](#)

46. Park, W.G.; Li, L.; Appana, S.; Wei, W.; Stello, K.; Andersen, D.K.; Hughes, S.J.; Whitcomb, D.C.; Brand, R.E.; Yadav, D.; et al. Unique Circulating Immune Signatures for Recurrent Acute Pancreatitis, Chronic Pancreatitis and Pancreatic Cancer: A Pilot Study of These Conditions with and without Diabetes: Immune Profiling of Pancreatic Disorders. *Pancreatology* **2020**, *20*, 51–59. [[CrossRef](#)] [[PubMed](#)]
47. Dranka-Bojarowska, D.; Lekstan, A.; Olakowski, M.; Jablonska, B.; Lewinski, A.; Musialski, P.; Sobczyk, W.; Kapalka, A.; Lampe, P. The Assessment of Serum Concentration of Adiponectin, Leptin and Serum Carbohydrate Antigen-19.9 in Patients with Pancreatic Cancer and Chronic Pancreatitis. *J. Physiol. Pharmacol.* **2015**, *66*, 653–663. [[PubMed](#)]
48. Hrabák, P.; Šoupal, J.; Kalousová, M.; Krechler, T.; Vočka, M.; Hanuš, T.; Petruželka, L.; Svačina, Š.; Žák, A.; Zima, T. Novel Biochemical Markers for Non-Invasive Detection of Pancreatic Cancer. *Neoplasma* **2022**, *69*, 474–483. [[CrossRef](#)] [[PubMed](#)]
49. Adrych, K.; Smoczynski, M.; Stojek, M.; Sledzinski, T.; Slominska, E.; Goyke, E.; Smolenski, R.T.; Swierczynski, J. Decreased Serum Essential and Aromatic Amino Acids in Patients with Chronic Pancreatitis. *World J. Gastroenterol.* **2010**, *16*, 4422–4427. [[CrossRef](#)]
50. Basso, D.; Plebani, M.; Panozzo, M.; Meggiato, T.; De Paoli, M.; Del Favero, G. Insulin-like Growth Factor-I, Interleukin-1 g and in Pancreatic Cancer: Role in Tumor Invasiveness and Associated Diabetes. *Int. J. Clin. Lab. Res.* **1995**, *25*, 40–43. [[CrossRef](#)]
51. Voronov, E.; Dotan, S.; Krelin, Y.; Song, X.; Elkabets, M.; Carmi, Y.; Rider, P.; Cohen, I.; Romzova, M.; Kaplanov, I.; et al. Unique versus Redundant Functions of IL-1 α and IL-1 β in the Tumor Microenvironment. *Front. Immunol.* **2013**, *4*, 177. [[CrossRef](#)]
52. Tu, S.; Bhagat, G.; Cui, G.; Takaishi, S.; Kurt-Jones, E.A.; Rickman, B.; Betz, K.S.; Penz-Oesterreicher, M.; Bjorkdahl, O.; Fox, J.G.; et al. Overexpression of Interleukin-1 β Induces Gastric Inflammation and Cancer and Mobilizes Myeloid-Derived Suppressor Cells in Mice. *Cancer Cell* **2008**, *14*, 408–419. [[CrossRef](#)] [[PubMed](#)]
53. Marrache, F.; Tu, S.P.; Bhagat, G.; Pendyala, S.; Österreicher, C.H.; Gordon, S.; Ramanathan, V.; Penz-Österreicher, M.; Betz, K.S.; Song, Z.; et al. Overexpression of Interleukin-1 β in the Murine Pancreas Results in Chronic Pancreatitis. *Gastroenterology* **2008**, *135*, 1277–1287. [[CrossRef](#)]
54. Schmidt, J.A.; Oliver, C.N.; Lepe-Zuniga, J.L.; Green, I.; Gery, I. Silica-Stimulated Monocytes Release Fibroblast Proliferation Factors Identical to Interleukin 1. A Potential Role for Interleukin 1 in the Pathogenesis of Silicosis. *J. Clin. Investig.* **1984**, *73*, 1462–1472. [[CrossRef](#)] [[PubMed](#)]
55. Malik, A.; Kanneganti, T.D. Function and Regulation of IL-1 α in Inflammatory Diseases and Cancer. *Immunol. Rev.* **2018**, *281*, 124–137. [[CrossRef](#)]
56. Fink, G.; Yang, J.; Carter, G.; Norman, J. Acute Pancreatitis-Induced Enzyme Release and Necrosis Are Attenuated by IL-1 Antagonism through an Indirect Mechanism. *J. Surg. Res.* **1997**, *67*, 94–97. [[CrossRef](#)]
57. Shen, J.; Gao, J.; Zhang, J.; Xiang, D.; Wang, X.; Qian, L.; Shen, J.; Yang, L.; Zhu, S.; Wu, M.; et al. Recombinant Human Interleukin-1 Receptor Antagonist (RhIL-1Ra) Attenuates Caerulein-Induced Chronic Pancreatitis in Mice. *Biomed. Pharmacother.* **2012**, *66*, 83–88. [[CrossRef](#)] [[PubMed](#)]
58. Chung, H.W.; Lim, J.B. Clinical Significance of Serum Levels of Immune-Associated Molecules, Uric Acid and Soluble MHC Class I Chain-Related Molecules A and B, as Diagnostic Tumor Markers for Pancreatic Ductal Adenocarcinoma. *Cancer Sci.* **2011**, *102*, 1673–1679. [[CrossRef](#)]
59. Thornton, A.M.; Donovan, E.E.; Piccirillo, C.A.; Shevach, E.M. Cutting Edge: IL-2 Is Critically Required for the in Vitro Activation of CD4+CD25+ T Cell Suppressor Function. *J. Immunol.* **2004**, *172*, 6519–6523. [[CrossRef](#)]
60. Grinberg-Bleyer, Y.; Baeyens, A.; You, S.; Elhage, R.; Fourcade, G.; Gregoire, S.; Cagnard, N.; Carpentier, W.; Tang, Q.; Bluestone, J.; et al. IL-2 Reverses Established Type 1 Diabetes in NOD Mice by a Local Effect on Pancreatic Regulatory T Cells. *J. Exp. Med.* **2010**, *207*, 1871–1878. [[CrossRef](#)]
61. Kayhan, B.; Kayhan, M.; Akdogan, M. Can IL-2R Alpha Be a Valuable Marker along with CA 19-9 in the Diagnosis of Chronic Pancreatitis and Pancreatic Cancer? *Int. J. Biol. Markers* **2004**, *19*, 196–202. [[CrossRef](#)]
62. Schlosser, W.; Gansauge, F.; Schlosser, S.; Gansauge, S.; Beger, H.G. Low Serum Levels of CD44, CD44v6, and Neopterin Indicate Immune Dysfunction in Chronic Pancreatitis. *Pancreas* **2001**, *23*, 335–340. [[CrossRef](#)]
63. Gansauge, F.; Steinbach, G.; Gansauge, S.; Ko Enig, H.-H.; Èrg, J.; Èller, M.; Ènert, A.G.; Beger, H.G. Prognostic Significance of Soluble Interleukin-2 Receptor- α in Adenocarcinoma of the Pancreas. *Cancer Lett.* **1998**, *134*, 193–199. [[CrossRef](#)]
64. Bluestone, J.A.; MacKay, C.R.; O'Shea, J.J.; Stockinger, B. The Functional Plasticity of T Cell Subsets. *Nat. Rev. Immunol.* **2009**, *9*, 811–816. [[CrossRef](#)]
65. Opal, S.M.; Depalo, V.A. Anti-Inflammatory Cytokines. *Chest* **2000**, *117*, 1162–1172. [[CrossRef](#)]
66. Xue, J.; Sharma, V.; Hsieh, M.H.; Chawla, A.; Murali, R.; Pandol, S.J.; Habtezion, A. Alternatively Activated Macrophages Promote Pancreatic Fibrosis in Chronic Pancreatitis. *Nat. Commun.* **2015**, *6*, 7158. [[CrossRef](#)] [[PubMed](#)]
67. Duggan, S.N.; Purcell, C.; Kilbane, M.; O'Keane, M.; McKenna, M.; Gaffney, P.; Ridgway, P.F.; Boran, G.; Conlon, K.C. An Association between Abnormal Bone Turnover, Systemic Inflammation, and Osteoporosis in Patients with Chronic Pancreatitis: A Case-Matched Study. *Am. J. Gastroenterol.* **2015**, *110*, 336–345. [[CrossRef](#)] [[PubMed](#)]
68. Zheng, M.; Li, H.; Sun, L.; Brigstock, D.R.; Gao, R. NC-ND License Interleukin-6 Participates in Human Pancreatic Stellate Cell Activation and Collagen I Production via TGF-B1/Smad Pathway. *Cytokine* **2021**, *143*, 1043–4666. [[CrossRef](#)] [[PubMed](#)]
69. Saurer, L.; Reber, P.; Schaffner, T.; Buchlerbubuchler, M.W.; Buri, C.; Kappeler, A.; Walz, A.; Friess, H.; Mueller, C. Differential Expression of Chemokines in Normal Pancreas and in Chronic Pancreatitis. *Gastroenterology* **2000**, *118*, 356–367. [[CrossRef](#)] [[PubMed](#)]

70. Di Sebastiano, P.; Di Mola, F.F.; Di Febbo, C.; Baccante, G.; Porreca, E.; Innocenti, P.; Friess, H.; Büchler, M.W. Expression of Interleukin 8 (IL-8) and Substance P in Human Chronic Pancreatitis. *Gut* **2000**, *47*, 423–428. [\[CrossRef\]](#)
71. Motoo, Y.; Xie, M.-J.; Mouri, H.; Sawabu, N. Expression of Interleukin-8 in Human Obstructive Pancreatitis. *JOP* **2004**, *5*, 138–144.
72. Demols, A.; Van Laethem, J.-L.; Quertinmont, E.; Degraef, C.; Delhaye, M.; Geerts, A.; Deviere, J. Endogenous Interleukin-10 Modulates Fibrosis and Regeneration in Experimental Chronic Pancreatitis. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2002**, *282*, G1105–G1112. [\[CrossRef\]](#)
73. Pezzilli, R.; Billi, P.; Miniero, R.; Barakat, B. Serum Interleukin-10 in Human Acute Pancreatitis. *Dig. Dis. Sci.* **1997**, *42*, 1469–1472. [\[CrossRef\]](#)
74. Van Laethem, J.L.; Eskinazi, R.; Louis, H.; Rickaert, F.; Robberecht, P.; Devière, J. Multisystemic Production of Interleukin 10 Limits the Severity of Acute Pancreatitis in Mice. *Gut* **1998**, *43*, 408–413. [\[CrossRef\]](#) [\[PubMed\]](#)
75. Khan, I.A.; Singh, N.; Gunjan, D.; Gopi, S.; Dash, N.R.; Gupta, S.; Saraya, A. Increased Circulating Th17 Cell Populations in Patients with Pancreatic Ductal Adenocarcinoma. *Immunogenetics* **2023**, *75*, 433–443. [\[CrossRef\]](#) [\[PubMed\]](#)
76. Li, G.; Chen, H.; Liu, L.; Xiao, P.; Xie, Y.; Geng, X.; Zhang, T.; Zhang, Y.; Lu, T.; Tan, H.; et al. Role of Interleukin-17 in Acute Pancreatitis. *Front. Immunol.* **2021**, *12*, 674803. [\[CrossRef\]](#) [\[PubMed\]](#)
77. Jia, R.; Tang, M.; Qiu, L.; Sun, R.; Cheng, L.; Ma, X.; Yin, G.; Hu, G.; Wang, X.; Zhao, Y. Increased Interleukin-23/17 Axis and C-Reactive Protein Are Associated with Severity of Acute Pancreatitis in Patients. *Pancreas* **2015**, *44*, 321–325. [\[CrossRef\]](#) [\[PubMed\]](#)
78. Mroczko, B.; Szmitkowski, M.; Wereszczyńska-Siemiatkowska, U.; Jurkowska, G. Hematopoietic Cytokines in the Sera of Patients with Pancreatic Cancer. *Clin. Chem. Lab. Med.* **2005**, *43*, 146–150. [\[CrossRef\]](#) [\[PubMed\]](#)
79. Bhattacharya, P.; Budnick, I.; Singh, M.; Thiruppathi, M.; Alharshawy, K.; Elshabrawy, H.; Holterman, M.J.; Prabhakar, B.S. Dual Role of GM-CSF as a Pro-Inflammatory and a Regulatory Cytokine: Implications for Immune Therapy. *J. Interferon Cytokine Res.* **2015**, *35*, 585–599. [\[CrossRef\]](#) [\[PubMed\]](#)
80. Bayne, L.J.; Beatty, G.L.; Jhala, N.; Clark, C.E.; Rhim, A.D.; Stanger, B.Z.; Vonderheide, R.H. Tumor-Derived Granulocyte-Macrophage Colony Stimulating Factor Regulates Myeloid Inflammation and T Cell Immunity in Pancreatic Cancer. *Cancer Cell* **2012**, *21*, 822–835. [\[CrossRef\]](#) [\[PubMed\]](#)
81. Halloran, P.F.; Autenried, P.; Ramassar, V.; Urmson, J.; Cockfield, S. Local T Cell Responses Induce Widespread MHC Expression. Evidence That IFN-Gamma Induces Its Own Expression in Remote Sites. *J. Immunol.* **1992**, *148*, 3837–3846. [\[CrossRef\]](#)
82. Xie, M.J.; Motoo, Y.; Su, S.B.; Sawabu, N. Expression of Tumor Necrosis Factor-Alpha, Interleukin-6, and Interferon-Gamma in Spontaneous Chronic Pancreatitis in the WBN/Kob Rat. *Pancreas* **2001**, *22*, 400–408. [\[CrossRef\]](#) [\[PubMed\]](#)
83. Sparmann, G.; Behrend, S.; Merkord, J.; Kleine, H.D.; Graser, E.; Ritter, T.; Liebe, S.; Emmrich, J. Cytokine mRNA Levels and Lymphocyte Infiltration in Pancreatic Tissue during Experimental Chronic Pancreatitis Induced by Dibutyltin Dichloride. *Dig. Dis. Sci.* **2001**, *46*, 1647–1656. [\[CrossRef\]](#) [\[PubMed\]](#)
84. Hasel, C.; Rau, B.; Perner, S.; Sträter, J.; Möller, P. Differential and Mutually Exclusive Expression of CD95 and CD95 Ligand in Epithelia of Normal Pancreas and Chronic Pancreatitis. *Lab. Investig.* **2001**, *81*, 317–326. [\[CrossRef\]](#) [\[PubMed\]](#)
85. Pan, S.; Chen, R.; Crispin, D.A.; May, D.; Stevens, T.; McIntosh, M.W.; Bronner, M.P.; Ziogas, A.; Anton-Culver, H.; Brentnall, T.A. Protein Alterations Associated with Pancreatic Cancer and Chronic Pancreatitis Found in Human Plasma Using Global Quantitative Proteomics Profiling. *J. Proteome Res.* **2011**, *10*, 2359–2376. [\[CrossRef\]](#)
86. Mohamed, A.; Saad, Y.; Saleh, D.; Elawady, R.; Eletreby, R.; Kharalla, A.S.; Badr, E. Can Serum ICAM 1 Distinguish Pancreatic Cancer from Chronic Pancreatitis? *Asian Pac. J. Cancer Prev.* **2016**, *17*, 4671–4675. [\[CrossRef\]](#)
87. Resovi, A.; Bani, M.R.; Porcu, L.; Anastasia, A.; Minoli, L.; Allavena, P.; Cappello, P.; Novelli, F.; Scarpa, A.; Morandi, E.; et al. Soluble Stroma-related Biomarkers of Pancreatic Cancer. *EMBO Mol. Med.* **2018**, *10*, e8741. [\[CrossRef\]](#)
88. Sato, T.; Shibata, W.; Maeda, S. Adhesion Molecules and Pancreatitis. *J. Gastroenterol.* **2019**, *54*, 99–107. [\[CrossRef\]](#)
89. Rahbari, N.N.; Schmidt, T.; Falk, C.S.; Hinz, U.; Herber, M.; Bork, U.; Büchler, M.W.; Weitz, J.; Koch, M. Expression and Prognostic Value of Circulating Angiogenic Cytokines in Pancreatic Cancer. *BMC Cancer* **2011**, *11*, 286. [\[CrossRef\]](#)
90. Shibuya, M. Vascular Endothelial Growth Factor (VEGF) and Its Receptor (VEGFR) Signaling in Angiogenesis: A Crucial Target for Anti- and Pro-Angiogenic Therapies. *Genes Cancer* **2011**, *2*, 1097–1105. [\[CrossRef\]](#)
91. Yasuda, M.; Ito, T.; Oono, T.; Kawabe, K.; Kaku, T.; Igarashi, H.; Nakamura, T.; Takayanagi, R. Fractalkine and TGF-β1 Levels Reflect the Severity of Chronic Pancreatitis in Humans. *World J. Gastroenterol.* **2008**, *14*, 6488–6495. [\[CrossRef\]](#) [\[PubMed\]](#)
92. Kozak, A.; Talar-Wojnarowska, R.; Kaczka, A.; Borkowska, A.; Czupryniak, L.; Małecka-Panas, E.; Gąsiorowska, A. Utility of Different Serum Fibrosis Markers in Diagnosing Patients with Chronic Pancreatitis and Pancreatic Adenocarcinoma. *World J. Gastrointest. Oncol.* **2016**, *8*, 635–641. [\[CrossRef\]](#) [\[PubMed\]](#)
93. Bazan, J.F.; Bacon, K.B.; Hardiman, G.; Wang, W.; Soo, K.; Rossi, D.; Greaves, D.R.; Zlotnik, A.; Schall, T.J. A New Class of Membrane-Bound Chemokine with a CX3C Motif. *Nature* **1997**, *385*, 640–644. [\[CrossRef\]](#) [\[PubMed\]](#)
94. Adrych, K.; Stojek, M.; Smoczynski, M.; Sledzinski, T.; Sylwia, S.W.; Swierczynski, J. Increased Serum Chemerin Concentration in Patients with Chronic Pancreatitis. *Dig. Liver Dis.* **2012**, *44*, 393–397. [\[CrossRef\]](#) [\[PubMed\]](#)
95. Kiczmer, P.; Szydło, B.; Prawdzic Seńkowska, A.; Jopek, J.; Wiewióra, M.; Piecuch, J.; Ostrowska, Z.; Świętochowska, E. Serum Omentin-1 and Chemerin Concentrations in Pancreatic Cancer and Chronic Pancreatitis. *Folia Medica Cracoviensia* **2018**, *LVIII*, 77–87. [\[CrossRef\]](#)
96. Hart, R.; Greaves, D.R. Chemerin Contributes to Inflammation by Promoting Macrophage Adhesion to VCAM-1 and Fibronectin through Clustering of VLA-4 and VLA-5. *J. Immunol.* **2010**, *185*, 3728–3739. [\[CrossRef\]](#)

97. Chang, M.-C.; Chang, Y.-T.; Su, T.-C.; Yang, W.-S.; Chen, C.-L.; Tien, Y.-W.; Liang, P.-C.; Wei, S.-C.; Wong, J.-M. Adiponectin as a Potential Differential Marker to Distinguish Pancreatic Cancer and Chronic Pancreatitis. *Pancreas* **2007**, *35*, 16–21. [\[CrossRef\]](#)
98. Trayhurn, P.; Wood, I.S. Adipokines: Inflammation and the Pleiotropic Role of White Adipose Tissue. *Br. J. Nutr.* **2004**, *92*, 347–355. [\[CrossRef\]](#)
99. Fang, H.; Judd, R.L. Adiponectin Regulation and Function. *Compr. Physiol.* **2018**, *8*, 1031–1063. [\[CrossRef\]](#)
100. Yamada, T.; Araki, H.; Watabe, K.; Kamada, Y.; Kiso, S.; Ogiyama, H.; Nishihara, T.; Kihara, S.; Funahashi, T.; Shimomura, I.; et al. Adiponectin Deficiency Enhanced the Severity of Cerulein-Induced Chronic Pancreatitis in Mice. *J. Gastroenterol.* **2010**, *45*, 742–749. [\[CrossRef\]](#) [\[PubMed\]](#)
101. Kamath, M.G.; Pai, C.G.; Kamath, A.; Kurien, A. Monocyte Chemoattractant Protein-1, Transforming Growth Factor-B1, Nerve Growth Factor, Resistin and Hyaluronic Acid as Serum Markers: Comparison between Recurrent Acute and Chronic Pancreatitis. *Hepatobiliary Pancreat. Dis. Int.* **2016**, *15*, 209–215. [\[CrossRef\]](#)
102. Silswal, N.; Singh, A.K.; Aruna, B.; Mukhopadhyay, S.; Ghosh, S.; Ehtesham, N.Z. Human Resistin Stimulates the Pro-Inflammatory Cytokines TNF-Alpha and IL-12 in Macrophages by NF-KappaB-Dependent Pathway. *Biochem. Biophys. Res. Commun.* **2005**, *334*, 1092–1101. [\[CrossRef\]](#)
103. Rychlíková, J.; Vecka, M.; Jáchymová, M.; Macásek, J.; Hrabák, P.; Zeman, M.; Vávrová, L.; Řoupal, J.; Krechler, T.; Ák, A. Osteopontin as a Discriminating Marker for Pancreatic Cancer and Chronic Pancreatitis. *Cancer Biomark.* **2016**, *17*, 55–65. [\[CrossRef\]](#) [\[PubMed\]](#)
104. Kolb, A.; Kleeff, J.; Guweidhi, A.; Esposito, I.; Giese, N.A.; Adwan, H.; Giese, T.; Büchler, M.W.; Berger, M.R.; Friess, H. Osteopontin Influences the Invasiveness of Pancreatic Cancer Cells and Is Increased in Neoplastic and Inflammatory Conditions. *Cancer Biol. Ther.* **2005**, *4*, 740–746. [\[CrossRef\]](#) [\[PubMed\]](#)
105. Poruk, K.E.; Firpo, M.A.; Scaife, C.L.; Adler, D.G.; Emerson, L.L.; Boucher, K.M.; Mulvihill, S.J. Serum Osteopontin and Tissue Inhibitor of Metalloproteinase 1 as Diagnostic and Prognostic Biomarkers for Pancreatic Adenocarcinoma. *Pancreas* **2013**, *42*, 193–197. [\[CrossRef\]](#) [\[PubMed\]](#)
106. Koopmann, J.; Rosenzweig, C.N.W.; Zhang, Z.; Canto, M.I.; Brown, D.A.; Hunter, M.; Yeo, C.; Chan, D.W.; Breit, S.N.; Goggins, M. Serum Markers in Patients with Resectable Pancreatic Adenocarcinoma: Macrophage Inhibitory Cytokine 1 versus CA19-9. *Clin. Cancer Res.* **2006**, *12*, 442–446. [\[CrossRef\]](#)
107. Lamort, A.-S.; Giopanou, I.; Psallidas, I.; Stathopoulos, G.T. Cells Osteopontin as a Link between Inflammation and Cancer: The Thorax in the Spotlight. *Cells* **2019**, *8*, 815. [\[CrossRef\]](#) [\[PubMed\]](#)
108. Nakamura, M.; Oka, M.; Iizuka, N.; Kawauchi, S.; Gondo, T.; Ueno, T.; Tangoku, A. Osteopontin Expression in Chronic Pancreatitis. *Pancreas* **2002**, *25*, 182–187. [\[CrossRef\]](#) [\[PubMed\]](#)
109. Xiang, Z.; Gu, Y.; Huang, Y.; Zhang, L.; Zhang, X.; Xu, H.; Liu, H.; Zhong, Y. Elevated Serum Neopterin Concentration Increases Mortality Risk in Patients with Acute Pancreatitis. *Pteridines* **2019**, *30*, 16–20. [\[CrossRef\]](#)
110. Uomo, G.; Spada, O.A.; Manes, G.; Feola, B.; Misso, S.; Cavallera, A.; Rabitti, P.G. Neopterin in Acute Pancreatitis. *Scand. J. Gastroenterol.* **1996**, *31*, 1032–1036. [\[CrossRef\]](#)
111. Cavestro, G.M.; Zuppardo, R.A.; Bertolini, S.; Sereni, G.; Frulloni, L.; Okolicsanyi, S.; Calzolari, C.; Singh, S.K.; Sianesi, M.; Del Rio, P.; et al. Connections between Genetics and Clinical Data: Role of Mcp-1, Cfr, and Spink-1 in the Setting of Acute, Acute Recurrent, and Chronic Pancreatitis. *Am. J. Gastroenterol.* **2010**, *105*, 199–206. [\[CrossRef\]](#)
112. Venkateshwari, A.; Sri Manjari, K.; Krishnaveni, D.; Nallari, P.; Vidyasagar, A.; Jyothy, A. Role of Plasma MMP 9 Levels in the Pathogenesis of Chronic Pancreatitis. *Indian J. Clin. Biochem.* **2011**, *26*, 136–139. [\[CrossRef\]](#) [\[PubMed\]](#)
113. Mroczko, B.; Lukaszewicz-Zajac, M.; Wereszczynska-Siemiatkowska, U.; Groblewska, M.; Gryko, M.; Kedra, B.; Jurkowska, G.; Szmikowski, M. Clinical Significance of the Measurements of Serum Matrix Metalloproteinase-9 and Its Inhibitor (Tissue Inhibitor of Metalloproteinase-1) in Patients With Pancreatic Cancer Metalloproteinase-9 as an Independent Prognostic Factor. *Pancreas* **2009**, *38*, 613–618. [\[CrossRef\]](#) [\[PubMed\]](#)
114. Dranka-Bojarowska, D.; Lewinski, A.; Lekstan, A.; Gajda, M.; Ciosek, J.; Mrowiec, S. The Assessment of Serum and Diagnostic Peritoneal Lavage Concentration of Matrix Metalloproteinase-2, Matrix Metalloproteinase-9, Carbohydrate Antigen 19-9, and Carcinoembryonic Antigen in Patients with Pancreatic Cancer and Chronic Pancreatitis. *J. Physiol. Pharmacol.* **2020**, *71*, 689–704. [\[CrossRef\]](#)
115. Grünwald, B.; Harant, V.; Schaten, S.; Frühschütz, M.; Spallek, R.; Höchst, B.; Stutzer, K.; Berchtold, S.; Erkan, M.; Prokopchuk, O.; et al. Pancreatic Premalignant Lesions Secrete Tissue Inhibitor of Metalloproteinases-1, Which Activates Hepatic Stellate Cells Via CD63 Signaling to Create a Premetastatic Niche in the Liver. *Gastroenterology* **2016**, *151*, 1011–1024.e7. [\[CrossRef\]](#) [\[PubMed\]](#)
116. Prokopchuk, O.; Grünwald, B.; Nitsche, U.; Jäger, C.; Prokopchuk, O.L.; Schubert, E.C.; Friess, H.; Martignoni, M.E.; Krüger, A. Elevated Systemic Levels of the Matrix Metalloproteinase Inhibitor TIMP-1 Correlate with Clinical Markers of Cachexia in Patients with Chronic Pancreatitis and Pancreatic Cancer. *BMC Cancer* **2018**, *18*, 128. [\[CrossRef\]](#)
117. Adrych, K.; Smoczynski, M.; Stojek, M.; Sledzinski, T.; Korczynska, J.; Goyke, E.; Swierczynski, J. Coordinated Increase in Serum Platelet-Derived Growth Factor-BB and Transforming Growth Factor-B1 in Patients with Chronic Pancreatitis. *Pancreatol.* **2011**, *11*, 434–440. [\[CrossRef\]](#)
118. Chen, I.M.; Willumsen, N.; Dehlendorff, C.; Johansen, A.Z.; Jensen, B.V.; Hansen, C.P.; Hasselby, J.P.; Bojesen, S.E.; Pfeiffer, P.; Nielsen, S.E.; et al. Clinical Value of Serum Hyaluronan and Propeptide of Type III Collagen in Patients with Pancreatic Cancer. *Int. J. Cancer* **2020**, *146*, 2913–2922. [\[CrossRef\]](#)

119. Suda, K. Distribution, Pathogenesis and Progression of Human Pancreatic Fibrosis. *Gastroenterology* **2007**, *33*, 67–79. [\[CrossRef\]](#)
120. Patel, M.; Fine, D.R. Fibrogenesis in the Pancreas after Acinar Cell Injury. *Scand. J. Surg.* **2005**, *94*, 108–111. [\[CrossRef\]](#)
121. Zhang, H.; Liu, B.; Xu, X.-F.; Jiang, T.-T.; Zhang, X.-Q.; Shi, Y.-L.; Chen, Y.; Liu, F.; Gu, J.; Zhu, L.-J.; et al. Pathophysiology of Chronic Pancreatitis Induced by Dibutyltin Dichloride Joint Ethanol in Mice. *World J. Gastroenterol.* **2016**, *22*, 2960–2970. [\[CrossRef\]](#)
122. Dhingra, R.; Singh, N.; Sachdev, V.; Ashish Datt Upadhyay, P.; Saraya, A. Effect of Antioxidant Supplementation on Surrogate Markers of Fibrosis in Chronic Pancreatitis A Randomized, Placebo-Controlled Trial. *Pancreas* **2013**, *42*, 589–595. [\[CrossRef\]](#) [\[PubMed\]](#)
123. Manjari, K.S.; Nallari, P.; Vidyasagar, A.; Jyothy, A.; Venkateshwari, A. Plasma TGF-B1, MMP-1 and MMP-3 Levels in Chronic Pancreatitis. *Indian J. Clin. Biochem.* **2012**, *27*, 152–156. [\[CrossRef\]](#) [\[PubMed\]](#)
124. Li, Z.; Yu, X.; Werner, J.; Bazhin, A.V.; D’Haese, J.G. The Role of Interleukin-18 in Pancreatitis and Pancreatic Cancer. *Cytokine Growth Factor Rev.* **2019**, *50*, 1–12. [\[CrossRef\]](#) [\[PubMed\]](#)
125. Heldin, C.-H. Platelet-Derived Growth Factor (PDGF). In *Encyclopedia of Hormones*; Henry, H.L., Norman, A.W., Eds.; Academic Press: Cambridge, MA, USA, 2003; pp. 231–237.
126. Singh, S.; Anshita, D.; Ravichandiran, V. MCP-1: Function, Regulation, and Involvement in Disease. *Int. Immunopharmacol.* **2021**, *101*, 107598. [\[CrossRef\]](#)
127. Wang, X.; Li, Y.; Tian, H.; Qi, J.; Li, M.; Fu, C.; Wu, F.; Wang, Y.; Cheng, D.; Zhao, W.; et al. Macrophage Inhibitory Cytokine 1 (MIC-1/GDF15) as a Novel Diagnostic Serum Biomarker in Pancreatic Ductal Adenocarcinoma. *BMC Cancer* **2014**, *14*, 578. [\[CrossRef\]](#) [\[PubMed\]](#)
128. Zhou, Y.F.; Xu, L.X.; Huang, L.Y.; Guo, F.; Zhang, F.; He, X.Y.; Yuan, Y.Z.; Yao, W.Y. Combined Detection of Serum U16-Binding Protein 2 and Macrophage Inhibitory Cytokine-1 Improves Early Diagnosis and Prognostic Prediction of Pancreatic Cancer. *Oncol. Lett.* **2014**, *8*, 2096–2102. [\[CrossRef\]](#)
129. Kaur, S.; Chakraborty, S.; Baine, M.J.; Mallya, K.; Smith, L.M.; Sasson, A.; Brand, R.; Guha, S.; Jain, M.; Wittel, U.; et al. Potentials of Plasma NGAL and MIC-1 as Biomarker(s) in the Diagnosis of Lethal Pancreatic Cancer. *PLoS ONE* **2013**, *8*, e55171. [\[CrossRef\]](#)
130. Bootcov, M.R.; Bauskin, A.R.; Valenzuela, S.M.; Moore, A.G.; Bansal, M.; He, X.Y.; Zhang, H.P.; Donnellan, M.; Mahler, S.; Pryor, K.; et al. MIC-1, a Novel Macrophage Inhibitory Cytokine, Is a Divergent Member of the TGF-Superfamily. *Cell Biol. South Wales* **1997**, *94*, 11514–11519. [\[CrossRef\]](#)
131. Fujiyama, T.; Ito, T.; Ueda, K.; Tachibana, Y.; Yasunaga, K.; Miki, M.; Takaoka, T.; Lee, L.; Kawabe, K.; Ogawa, Y. Serum Levels of Wisteria Floribunda Agglutinin-Positive Mac-2 Binding Protein Reflect the Severity of Chronic Pancreatitis. *J. Dig. Dis.* **2017**, *18*, 302–308. [\[CrossRef\]](#)
132. Maekawa, T.; Kamada, Y.; Ebisutani, Y.; Ueda, M.; Hata, T.; Kawamoto, K.; Takamatsu, S.; Mizutani, K.; Shimomura, M.; Sobajima, T.; et al. Serum Mac-2 Binding Protein Is a Novel Biomarker for Chronic Pancreatitis. *World J. Gastroenterol.* **2016**, *22*, 4403–4410. [\[CrossRef\]](#)
133. Sasaki, T.; Brakebusch, C.; Engel, J.; Timpl, R. Mac-2 Binding Protein Is a Cell-Adhesive Protein of the Extracellular Matrix Which Self-Assembles into Ring-like Structures and Binds Beta1 Integrins, Collagens and Fibronectin. *EMBO J.* **1998**, *17*, 1606–1613. [\[CrossRef\]](#)
134. Inohara, H.; Akahani, S.; Kohts, K.; Raz, A. Interactions between Galectin-3 and Mac-2-Binding Protein Mediate Cell-Cell Adhesion. *Cancer Res.* **1996**, *56*, 4530–4534. [\[PubMed\]](#)
135. Fujiyoshi, M.; Kuno, A.; Gotoh, M.; Fukai, M.; Yokoo, H.; Kamachi, H.; Kamiyama, T.; Korenaga, M.; Mizokami, M.; Narimatsu, H.; et al. Clinicopathological Characteristics and Diagnostic Performance of Wisteria Floribunda Agglutinin Positive Mac-2-Binding Protein as a Preoperative Serum Marker of Liver Fibrosis in Hepatocellular Carcinoma. *J. Gastroenterol.* **2015**, *50*, 1134–1144. [\[CrossRef\]](#) [\[PubMed\]](#)
136. Kamada, Y.; Fujii, H.; Fujii, H.; Sawai, Y.; Doi, Y.; Uozumi, N.; Mizutani, K.; Akita, M.; Sato, M.; Kida, S.; et al. Serum Mac-2 Binding Protein Levels as a Novel Diagnostic Biomarker for Prediction of Disease Severity and Nonalcoholic Steatohepatitis. *Proteom. Clin. Appl.* **2013**, *7*, 648–656. [\[CrossRef\]](#)
137. Śliwińska-Mosson, M.; Milnerowicz, S.; Nabzdyk, S.; Kokot, I.; Nowak, M.; Milnerowicz, H. The Effect of Smoking on Endothelin-1 in Patients With Chronic Pancreatitis. *Appl. Immunohistochem. Mol. Morphol.* **2014**, *23*, 288–296. [\[CrossRef\]](#)
138. Jonitz, A.; Fitzner, B.; Jaster, R. Molecular Determinants of the Profibrogenic Effects of Endothelin-1 in Pancreatic Stellate Cells. *World J. Gastroenterol.* **2009**, *15*, 4143–4149. [\[CrossRef\]](#)
139. Meggiato, T.; Plebani, M.; Basso, D.; Panozzo, M.P.; Del Favero, G. Serum Growth Factors in Patients with Pancreatic Cancer. *Tumor Biol.* **1999**, *20*, 65–71. [\[CrossRef\]](#)
140. Wong, R.W.C.; Guillaud, L. The Role of Epidermal Growth Factor and Its Receptors in Mammalian CNS. *Cytokine Growth Factor Rev.* **2004**, *15*, 147–156. [\[CrossRef\]](#)
141. Blaine, S.A.; Ray, K.C.; Branch, K.M.; Robinson, P.S.; Whitehead, R.H.; Means, A.L. Epidermal Growth Factor Receptor Regulates Pancreatic Fibrosis. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2009**, *297*, 434–441. [\[CrossRef\]](#)
142. Włodarczyk, B.; Borkowska, A.; Włodarczyk, P.; Malecka-Panas, E.; Gasiorowska, A. Insulin-like Growth Factor 1 and Insulin-like Growth Factor Binding Protein 2 Serum Levels as Potential Biomarkers in Differential Diagnosis between Chronic Pancreatitis and Pancreatic Adenocarcinoma in Reference to Pancreatic Diabetes. *Przegląd Gastroenterol.* **2021**, *16*, 36–72. [\[CrossRef\]](#)

143. Włodarczyk, B.; Gasiorowska, A.; Borkowska, A.; Malecka-Panas, E. Evaluation of Insulin-like Growth Factor (IGF-1) and Retinol Binding Protein (RBP-4) Levels in Patients with Newly Diagnosed Pancreatic Adenocarcinoma (PDAC). *Pancreatology* **2017**, *17*, 623–628. [\[CrossRef\]](#)
144. Xu, J.W.; Wang, T.X.; You, L.; Zheng, L.F.; Shu, H.; Zhang, T.P.; Zhao, Y.P. Insulin-like Growth Factor 1 Receptor (IGF-1R) as a Target of MiR-497 and Plasma IGF-1R Levels Associated with TNM Stage of Pancreatic Cancer. *PLoS ONE* **2014**, *9*, e92847. [\[CrossRef\]](#) [\[PubMed\]](#)
145. Al-Samerria, S.; Radovick, S. The Role of Insulin-like Growth Factor-1 (Igf-1) in the Control of Neuroendocrine Regulation of Growth. *Cells* **2021**, *10*, 2664. [\[CrossRef\]](#) [\[PubMed\]](#)
146. Rosendahl, A.H.; Gundewar, C.; Hilmersson, K.S.; Ni, L.; Saleem, M.A.; Andersson, R. Conditionally Immortalized Human Pancreatic Stellate Cell Lines Demonstrate Enhanced Proliferation and Migration in Response to IGF-I. *Exp. Cell Res.* **2014**, *330*, 300–310. [\[CrossRef\]](#) [\[PubMed\]](#)
147. Sajewicz, W.; Milnerowicz, S.; Nabzdyk, S. Blood Plasma Antioxidant Defense in Patients With Pancreatitis. *Pancreas* **2006**, *32*, 139–144. [\[CrossRef\]](#) [\[PubMed\]](#)
148. Tsuji, N.; Watanabe, N.; Okamoto, T.; Niitsu, Y. Specific Interaction of Pancreatic Elastase and Leucocytes to Produce Oxygen Radicals and Its Implication in Pancreatitis. *Gut* **1994**, *35*, 1659–1664. [\[CrossRef\]](#) [\[PubMed\]](#)
149. Schoenberg, M.H.; Buchler, M.; Pietrzyk, C.; Uhl, W.; Birk, D.; Eisele, S.; Marzinzig, M.; Beger, H.G. Lipid Peroxidation and Glutathione Metabolism in Chronic Pancreatitis. *Pancreas* **1995**, *10*, 36–43. [\[CrossRef\]](#) [\[PubMed\]](#)
150. Fukui, M.; Kanoh, M.; Takamatsu, Y.; Arakawa, Y. Analysis of Serum Catalase Activities in Pancreatic Diseases. *J. Gastroenterol.* **2004**, *39*, 469–474. [\[CrossRef\]](#)
151. López, M.A.; Alcaraz, C.A. Oxidative Stress and Acute Pancreatitis. *World J. Gastroenterol.* **2011**, *103*, 559–562. [\[CrossRef\]](#)
152. Tasci, I.; Deveci, S.; Isik, A.T.; Comert, B.; Akay, C.; Mas, N.; Inal, V.; Yamanel, L.; Mas, M.R. Allopurinol in Rat Chronic Pancreatitis: Effects on Pancreatic Stellate Cell Activation. *Pancreas* **2007**, *35*, 366–371. [\[CrossRef\]](#)
153. Kirk, G.R.; White, J.S.; McKie, L.; Stevenson, M.; Young, I.; Clements, W.D.B.; Rowlands, B.J. Combined Antioxidant Therapy Reduces Pain and Improves Quality of Life in Chronic Pancreatitis. *J. Gastrointest. Surg.* **2006**, *10*, 499–503. [\[CrossRef\]](#) [\[PubMed\]](#)
154. Salim, A.S. Role of Oxygen-Derived Free Radical Scavengers in the Treatment of Recurrent Pain Produced by Chronic Pancreatitis. A New Approach. *Arch. Surg.* **1991**, *126*, 1109–1114. [\[CrossRef\]](#)
155. Santini, S.A.; Spada, C.; Bononi, F.; Foschia, F.; Mutignani, M.; Perri, V.; Giardina, B.; Gentiloni Silveri, N.; Costamagna, G. Enhanced Lipoperoxidation Products in Pure Pancreatic Juice: Evidence for Organ-Specific Oxidative Stress in Chronic Pancreatitis. *Dig. Liver Dis.* **2003**, *35*, 888–892. [\[CrossRef\]](#) [\[PubMed\]](#)
156. Zhang, L.; Lin, B. Decreased Serum Paraoxonase Activity in Patients With Chronic Pancreatitis. *Am. J. Med. Sci.* **2013**, *346*, 363–365. [\[CrossRef\]](#)
157. Kodydkova, J.; Vavrova, L.; Stankova, B.; Macasek, J.; Krechler, T.; Zak, A. Antioxidant Status and Oxidative Stress Markers in Pancreatic Cancer and Chronic Pancreatitis. *Pancreas* **2013**, *42*, 614–621. [\[CrossRef\]](#) [\[PubMed\]](#)
158. Verlaan, M.; Roelofs, H.M.; van Schaik, A.; Wanten, G.J.; Jansen, J.B.; Peters, W.H.; Drenth, J.P. Assessment of Oxidative Stress in Chronic Pancreatitis Patients. *World J. Gastroenterol.* **2006**, *12*, 5705. [\[CrossRef\]](#)
159. Girish, B.N.; Rajesh, G.; Vaidyanathan, K. Deficiency of Folate and Vitamin B12 Increases Oxidative Stress in Chronic Pancreatitis Patients. *Indian J. Gastroenterol.* **2022**, *41*, 77–83. [\[CrossRef\]](#)
160. Girish, B.; Rajesh, G.; Vaidyanathan, K.; Balakrishnan, V. Assessment of Oxidative Status in Chronic pancreatitis and Its Relation with Zinc Status. *Indian J. Gastroenterol.* **2011**, *30*, 63–65. [\[CrossRef\]](#)
161. Bhardwaj, P.; Garg, P.K.; Maulik, S.K.; Saraya, A.; Tandon, R.K.; Acharya, S.K. A Randomized Controlled Trial of Antioxidant Supplementation for Pain Relief in Patients With Chronic Pancreatitis. *Gastroenterology* **2009**, *136*, 149–159. [\[CrossRef\]](#)
162. Van Gossum, A.; Closset, P.; Noel, E.; Cremer, M.; Neve, J. Deficiency in Antioxidant Factors in Patients with Alcohol-Related Chronic Pancreatitis. *Dig. Dis. Sci.* **1996**, *41*, 1225–1231. [\[CrossRef\]](#)
163. Aguilar Diaz De Leon, J.; Borges, C.R. Evaluation of Oxidative Stress in Biological Samples Using the Thiobarbituric Acid Reactive Substances Assay. *J. Vis. Exp.* **2020**, *159*, e61122. [\[CrossRef\]](#)
164. Bopanna, S.; Nayak, B.; Prakash, S.; Shalimar; Mahapatra, S.J.; Garg, P.K. Increased Oxidative Stress and Deficient Antioxidant Levels May Be Involved in the Pathogenesis of Idiopathic Recurrent Acute Pancreatitis. *Pancreatology* **2017**, *17*, 529–533. [\[CrossRef\]](#) [\[PubMed\]](#)
165. Podborska, M.; Sevcikova, A.; Trna, J.; Dite, P.; Lojek, A.; Kubala, L. Increased Markers of Oxidative Stress in Plasma of Patients with Chronic Pancreatitis. *Neuroendocrinol. Lett.* **2009**, *30*, 300709–300728.
166. Trevisani, M.; Siemens, J.; Materazzi, S.; Bautista, D.M.; Nassini, R.; Campi, B.; Imamachi, N.; Andrè, E.; Patacchini, R.; Cottrell, G.S.; et al. 4-Hydroxynonenal, an Endogenous Aldehyde, Causes Pain and Neurogenic Inflammation through Activation of the Irritant Receptor TRPA1. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 13519–13524. [\[CrossRef\]](#) [\[PubMed\]](#)
167. Szuster-Ciesielska, A.; Daniluk, J.; Kandefer-Szerszeń, M. Oxidative Stress in Blood of Patients with Alcohol-Related Pancreatitis. *Pancreas* **2001**, *22*, 261–266. [\[CrossRef\]](#) [\[PubMed\]](#)
168. Girish, B.N.; Vaidyanathan, K.; Nanjundarao, P.; Rao, A.; Rajesh, G.; Reshmi, S.; Balakrishnan, V. Chronic Pancreatitis Is Associated With Hyperhomocysteinemia and Derangements in Transsulfuration and Transmethylation Pathways. *Pancreas* **2009**, *39*, e11–e16. [\[CrossRef\]](#)

169. Quilliot, D.; Walters, E.; Bonte, J.-P.; Fruchart, J.-C.; Duriez, P.; Ziegler, O. Diabetes Mellitus Worsens Antioxidant Status in Patients with Chronic Pancreatitis. *Am. J. Clin. Nutr.* **2005**, *81*, 1117–1125. [[CrossRef](#)]
170. Matsumoto, M.; Wakasugi, H.; Ibayashi, H. Serum Vitamine E, Lipid Peroxide and Glutathione Peroxidase in Patients with Chronic Pancreatitis. *Clinica Chimica Acta* **1981**, *110*, 121–125. [[CrossRef](#)]
171. Morris-Stiff, G.J.; Bowrey, D.J.; Oleesky, D.; Davies, M.; B Clark, G.W.; A Puntis, M.C. The Antioxidant Profiles of Patients With Recurrent Acute and Chronic Pancreatitis. *Am J. Gastroenterol.* **1999**, *94*, 2135–2140. [[CrossRef](#)]
172. Liu, H.; Guo, H.; Jian, Z.; Cui, H.; Fang, J.; Zuo, Z.; Deng, J.; Li, Y.; Wang, X.; Zhao, L. Copper Induces Oxidative Stress and Apoptosis in the Mouse Liver. *Oxidative Med. Cell. Longev.* **2020**, *2020*, 1359164. [[CrossRef](#)]
173. Kawaguchi, Y.; Ogawa, M.; Ito, H.; Mine, T. Alterations in Plasma Amino Acid Levels in Alcoholic Chronic Pancreatitis in Japanese. *Digestion* **2012**, *86*, 155–160. [[CrossRef](#)]
174. Sogawa, K.; Yamanaka, S.; Takano, S.; Sasaki, K.; Miyahara, Y.; Furukawa, K.; Takayashiki, T.; Kuboki, S.; Takizawa, H.; Nomura, F.; et al. Fucosylated C4b-Binding Protein α -Chain, a Novel Serum Biomarker That Predicts Lymph Node Metastasis in Pancreatic Ductal Adenocarcinoma. *Oncol. Lett.* **2021**, *21*, 127. [[CrossRef](#)]
175. Nissen, N.I.; Johansen, A.Z.; Chen, I.M.; Jensen, C.; Madsen, E.A.; Hansen, C.P.; Thorlacius-Ussing, J.; Karsdal, M.; Johansen, J.S.; Diab, H.M.H.; et al. High Serum Levels of the C-Propeptide of Type V Collagen (PRO-C5) Are Prognostic for Short Overall Survival in Patients with Pancreatic Ductal Adenocarcinoma. *Front Mol Biosci* **2023**, *10*, 1158058. [[CrossRef](#)]
176. Slater, E.P.; Fendrich, V.; Strauch, K.; Rospleszcz, S.; Ramaswamy, A.; Matthäi, E.; Chaloupka, B.; Gress, T.M.; Langer, P.; Bartsch, D.K. LCN2 and TIMP1 as Potential Serum Markers for the Early Detection of Familial Pancreatic Cancer. *Transl. Oncol.* **2013**, *6*, 99–103. [[CrossRef](#)] [[PubMed](#)]
177. Iglesias-García, J.; Abdulkader, I.; Lariño-Noia, J.; Forteza, J.; Dominguez-Muñoz, J.E. Histological Evaluation of Chronic Pancreatitis by Endoscopic Ultrasound-Guided Fine Needle Biopsy. *Gut* **2006**, *55*, 1661–1662. [[CrossRef](#)] [[PubMed](#)]
178. Kikuta, K.; Masamune, A.; Satoh, M.; Suzuki, N.; Satoh, K.; Shimosegawa, T. Hydrogen Peroxide Activates Activator Protein-1 and Mitogen-Activated Protein Kinases in Pancreatic Stellate Cells. *Mol. Cell. Biochem.* **2006**, *291*, 11–20. [[CrossRef](#)]
179. Lee, B.; Jones, E.K.; Manohar, M.; Li, L.; Yadav, D.; Conwell, D.L.; Hart, P.A.; Vege, S.S.; Fogel, E.L.; Serrano, J.; et al. Distinct Serum Immune Profiles Define the Spectrum of Acute and Chronic Pancreatitis From the Multicenter Prospective Evaluation of Chronic Pancreatitis for Epidemiologic and Translational Studies (PROCEED) Study. *Gastroenterology* **2023**, *165*, 173–186. [[CrossRef](#)]
180. Novovic, S.; Borch, A.; Werge, M.; Karran, D.; Gluud, L.; Schmidt, P.N.; Hansen, E.F.; Nøjgaard, C.; Jensen, A.B.; Jensen, F.K.; et al. Characterisation of the Fibroinflammatory Process Involved in Progression from Acute to Chronic Pancreatitis: Study Protocol for a Multicentre, Prospective Cohort Study. *BMJ Open* **2019**, *9*, e028999. [[CrossRef](#)] [[PubMed](#)]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.