



Review

Hypertension Induces Pro-arrhythmic Cardiac Connexome Disorders: Protective Effects of Treatment

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Abstract: Prolonged population aging and unhealthy lifestyles contribute to the progressive prevalence of arterial hypertension. This is accompanied by low-grade inflammation and over time results in heart dysfunction and failure. Hypertension-induced myocardial structural and ion channel remodeling facilitates the development of both atrial and ventricular fibrillation, and these increase the risk of stroke and sudden death. Herein, we elucidate hypertension-induced impairment of “connexome” cardiomyocyte junctions. This complex ensures cell-to-cell adhesion and coupling for electrical and molecular signal propagation. Connexome dysfunction can be a key factor in promoting the occurrence of both cardiac arrhythmias and heart failure. However, the available literature indicates that arterial hypertension treatment can hamper myocardial structural remodeling, hypertrophy and/or fibrosis, and preserve connexome function. This suggests the pleiotropic effects of antihypertensive agents, including anti-inflammatory. Therefore, further research is required to identify specific molecular targets and pathways that will protect connexomes, and it is also necessary to develop new approaches to maintain heart function in patients suffering from primary or pulmonary arterial hypertension.



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1. Introduction

Herein, we focus on pro-arrhythmic disorders elicited by primary hypertension (HTN). This is a prevalent risk factor for cardiovascular disease in the general population that includes younger people. HTN is defined as systolic blood pressure above 140 mmHg or diastolic blood pressure above 80 mmHg. These can result in hypertensive heart disease from pressure overload. HTN-induced compensated myocardial hypertrophy can aggravate left heart dysfunction over an extended period. This is mainly due to fibrosis, and it results in heart failure (HF). HTN also promotes the occurrence of cardiac arrhythmias, including ventricular fibrillation (VF) and atrial fibrillation (AF). In addition to HTN, there is also an increased incidence of pulmonary arterial hypertension (PAH) which affects right heart function. PAH is caused by pulmonary vasculopathy, and it results in elevated pulmonary arterial pressure above 25 mmHg at rest. This is a most serious clinical problem and has adverse prognosis due to heart disease progression and the accompanying myocardial fibrosis contributing to HF. There is also a further risk of VF and AF. Further research is also essential to differentiate gender differences in both primary HTN and PAH for efficient treatment.

It has been established that myocardial connexin-43 (Cx43) channels ensure electrical coupling between cardiomyocytes and are crucial in the development of malignant cardiac arrhythmias [1]. The available literature indicates that HTN and PAH deteriorate Cx43 channels and mediate communication at the heart’s gap junctions (GJs), as well as the function of adhesive junctions [2–6]. Cx43 GJs are essential for electrical coupling and the propagation

of action potentials between cardiomyocytes [7]. Desmosomes (D) and adherens junctions (AJs) are junctions responsible for cardiac myocyte adhesion and mechanical force transduction via actin filaments [8] located in the intercalated discs (ID), and their impairment or dysfunction in heart disease can facilitate VF or AF and contribute to HF [9–11].

Research suggests that it is important to consider connexomes and “area composite”, complex proteins which underly the interaction of Cx43-formed GJs, D, and AJs at the intercalated discs. This is highlighted in Figure 1, and Figure 2 shows their localization on the cardiomyocyte lateral aspect. Connexome defects are essential in arrhythmogenic cardiomyopathy [12–17], and are most likely implicated in both AF and VF promoted by HTN or PAH [11,18,19].

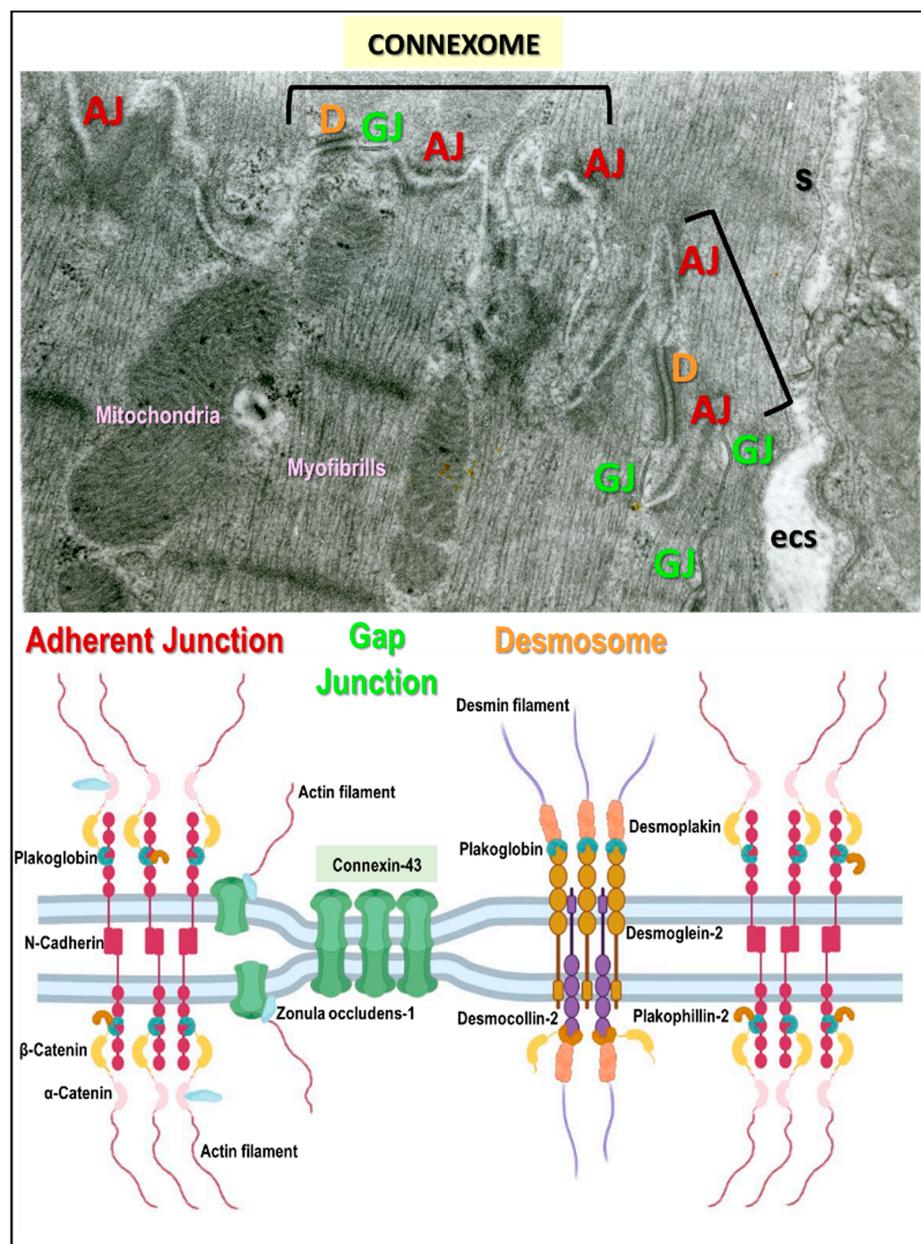


Figure 1. Intercalated discs comprise three distinct junctional complexes, the “connexome”: adherens junction (AJ), desmosome (D), and gap junction (GJ). These work together to mediate cardiomyocyte mechanical and electrical coupling. There are also various interacting proteins that can modulate connexome function [10,12–18]. Disorders of this complex structure appear to be pro-arrhythmic and promote mechanical heart dysfunction. Electron microscopic images adapted from [11].

It is most important that the connexomes affect both cardiomyocyte electrical coupling through Cx43 channels and mechanical force transduction by adhesive junctions. It is *conditio sine qua non* for synchronized myocardial contraction, and this is impaired in heart disease [8,9,20]. In addition, ion channels and Cx43 hemichannels form part of the connexome, and these affect its function [7]. Therefore, connexome's structural and functional preservation presents a challenging target for pharmacological and non-pharmacological approaches [10]. These will enhance the fight against life-threatening cardiac arrhythmias in general, and in HTN and PAH in particular.

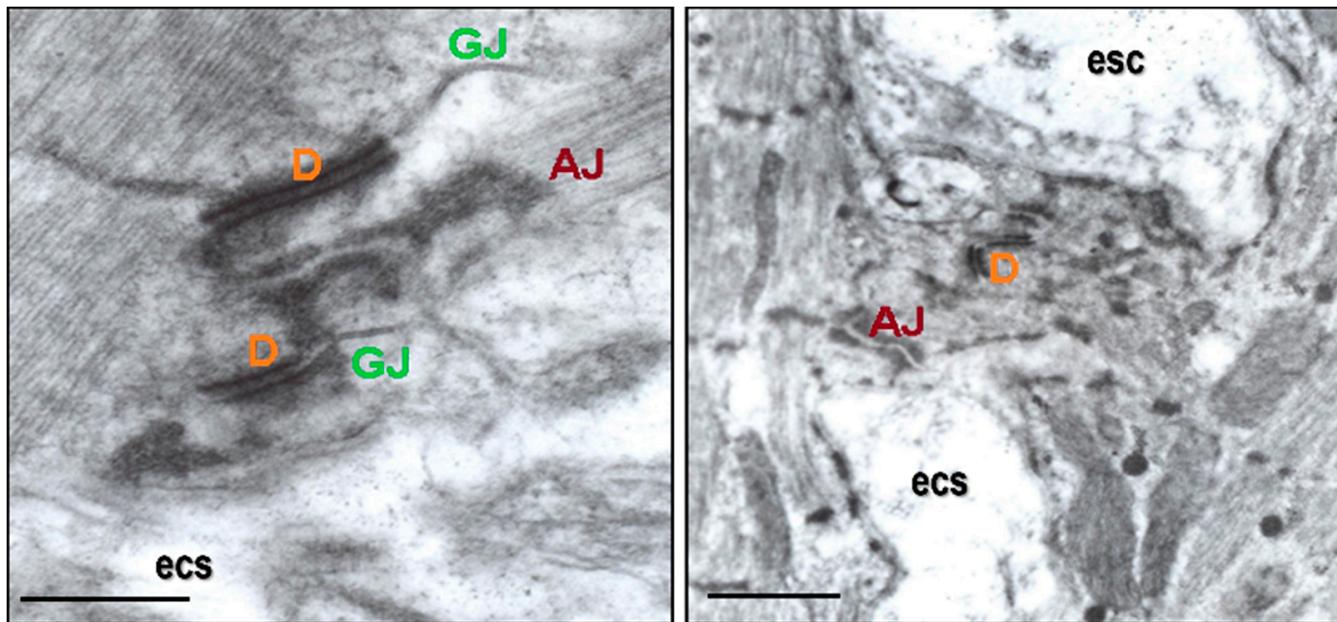


Figure 2. Connexomes are identified on the lateral sides of the hypertrophied guinea pig cardiomyocytes. Their AJ, D, and GJ components are destroyed by progressive extracellular collagen deposition [21]. Scale bar represents 1 micrometer.

2. Factors Involved in the Development of Re-Entrant Cardiac Arrhythmias, VF, and AF

The heart can “die” due to the following three major events: electromechanical dissociation, asystole, and VF, which is the most frequent case. In addition, AF is the most frequent arrhythmia in the population. AF deteriorates heart function and can cause stroke. The basic electrophysiological mechanisms of cardiac arrhythmias include the following: (1) abnormal electrical impulse generation through ectopic pacemaker-like activity or triggered activity and (2) abnormal electrical impulse propagation, due to blocks of conduction and re-entrant excitation. Simultaneous operation of abnormalities 1 and 2 may occur [7,22–25].

Figure 3 highlights that arrhythmogenic substrates, electrical triggers, and modulations are the three major factors in AF and VF development. Heart disease-related myocardial structural and ion channel remodeling are the established arrhythmogenic substrates. This includes hypertrophy, fibrosis, D and AJ impairment, and altered Cx43 topology and its downregulation. These changes can influence anisotropic conduction and promote discontinuous and re-entrant action potential propagation [26,27]. Fibrosis causes extreme disturbance of Cx43 GJ distribution at the myocardial interface, and defines the location of re-entry circuits that cause ventricular arrhythmia [2]. In addition, heterogeneous Cx43 GJ expression adversely affects the normal pattern of coordinated myocardial excitation, and this can directly depress cardiac performance [28].

ARRHYTHMOGENIC SUBSTRATE	ELECTRICAL TRIGGERS	MODULATING FACTORS
Structural and Ion Channels Remodelling Hypertrophy, Necrosis, Fibrosis Impairment of Adhesive Junctions Down-regulation of Cx43 Altered Topology of Cx43 Dysfunction of Ion Channels	Early or Delayed After-Depolarization Ectopic Pacemaker-like Activity Cx43-channels Uncoupling Ca ²⁺ - overload Ion Current Abnormalities Dispersion of Repolarisation	Humoral Disorders, Inflammation Misbalance of Autonomic Tone Redox Dysregulation, Stressors <u>Activation of Cx43 HC</u> <u>Inhibition of Cx43 GJch</u>

Figure 3. Tentative connexome implications in pro-arrhythmic disorders induced by HTN and PAH.

Research indicates that abnormal Ca²⁺ handling, Ca²⁺-overload, and ischemia-related acidosis induce Cx43-GJ uncoupling. When this is combined with ion current abnormalities, it can trigger the electrical disorders that initiate AF or VF occurrence [4,6,19,24,29,30].

Modulating factors, such as humoral and autonomic tone misbalance [31,32], inflammation, and redox dysregulation [22,33], as well as various stressors, such as stretch [34,35], impact the susceptibility of the heart to re-entrant arrhythmias. Finally, Cx43 hemichannel activation is fundamental to this process, because it can promote pro-arrhythmic signalling [7,24] and Cx43 GJ redistribution or inhibition.

Although the basic mechanisms that can cause cardiac arrhythmias are known, there is little understanding of the changes in cardiac electrical properties in heart disease, and this includes those in HTN and PAH. These properties appear to be the immediate cause of the operation of the arrhythmogenic mechanisms which occur in cardiac conditions and disease. Both AF and VF are assumed to occur due to ectopic impulse initiation, blocking conduction, and circuit re-entry. Although, AF can self-terminate and VF can be transient in the heart, avoiding arrhythmogenic substrates under the control of modulating factors. However, sustained AF or VF can aggravate the myocardial injury and connexome function (Figure 4). Therefore, protection of the connexome, GJ, and adhesion of the perinexus to preserve sodium channels and cardiac conduction present promising anti-arrhythmic research targets [36].

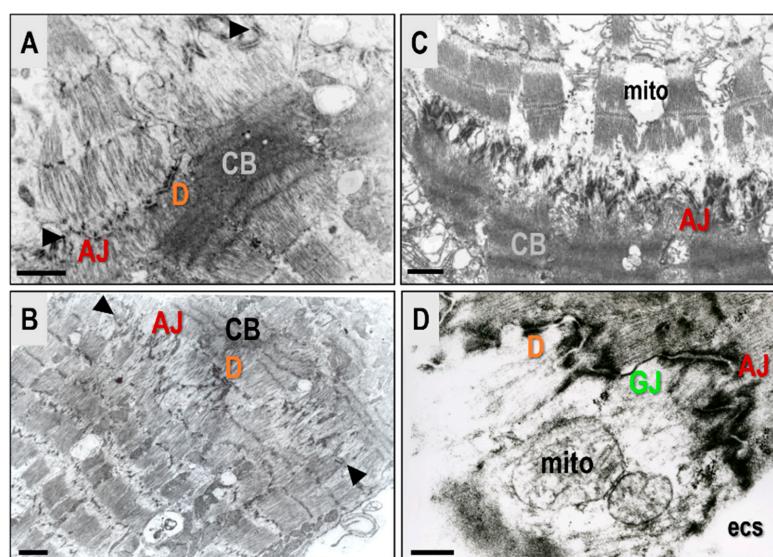


Figure 4. AF and VF aggravate connexome impairment. This includes AJ, D, and GJ due to Ca²⁺- overload, ischemia, and other factors. The subcellular rat heart alterations shown in the (A–C) images indicate desynchronized contraction of neighboring cardiomyocytes, and image (D) shows varying degrees of injury attributed to connexome impairment. Scale bar represents 0.1 micrometer. Adapted from [19].

3. Connexome Impairment Promoting AF or VF Development in Primary HTN

Primary or essential HTN is a prevalent risk factor jeopardizing cardiovascular health, and its increase in younger people is very alarming [37,38]. HTN is a multifactorial disease, and its etiopathogenesis includes the interplay between the genetic, epigenetic, environmental, and lifestyle factors which lead to meta-inflammation, oxidative stress, and auto-antibody production [23,39–52]. In addition, insufficient myocardial perfusion to match overall metabolic demand has been identified as an elevated risk of heart failure (HF) in symptomatic patients with HTN [53]. Adverse HTN consequences promote stroke, as well as heart electro-mechanical dysfunction, and this can lead to the development of AF or VF and subsequent HF [54–57]. Therefore, primary or essential HTN has become a target of considerable global public health concern [38,58].

Numerous animal and human heart studies indicate that HTN-induced structural remodeling involves hypertrophy, a shift in myosin heavy chains, cytoskeletal proteins, fibrosis, channelopathy, altered Ca^{2+} handling, and Cx43 disorders of the left ventricle. These are crucial in impaired conduction, electrical instability, heart mechanical dysfunction, and cardiac arrhythmias [47,54,55,57,59–70].

In addition, the progression of left ventricular alterations to HF is associated with right ventricular dysfunction and electrical instability [71]. This contributes to an adverse prognosis. Moreover, changes in membrane lipid composition have been reported in cardiac hypertrophy, and these can also affect GJ coupling and conduction [72].

Available literature indicates that HTN results in altered connexomes on the cardiomyocyte lateral sides, in addition to its multiple locations in ID. The subcellular alterations indicate connexome abnormalities. These include the AJ dehiscence, internalization, and GJ degradation, as demonstrated in Figures 5 and 6.

HTN-induced structural remodeling causes Cx43 redistribution from GJs to the cardiomyocyte lateral sides [61,62,67,74], Cx43 downregulation [73,75–77], and D and AJs dehiscence [4,19]. Delocalization of the Cx43 from GJs is a major pathologic-mechanism in hypertensive electrochemical remodeling [74,78]. The N-cadherin/catenin complex of AJs is a master regulator of ID function [18], and N-cadherin loss leads to altered Cx43 with reduced conduction velocity (CV) and arrhythmogenesis [13,39].

It is important that Cx43 cardiomyocyte lateralization is accompanied by the remodeling of D and microtubule-associated proteins [79]. This remodeling can affect electrical synchrony under conditions of disrupted ID integrity. Cadherin dysregulation has been demonstrated in IDs of spontaneously hypertensive stroke-prone rats [80]. This can contribute to altered heart function. In addition, reduced Cx43 expression triggers increased fibrosis due to enhanced fibroblast activity [81]. Therefore, the implications of arrhythmic fibroblast–cardiomyocyte interactions should be considered [82].

LVH is associated with increased intracellular resistivity which can be solely attributed to increased junctional resistance between adjacent cells [83]. Cardiac hypertrophy is known to be regulated by micro RNAs [84], and the upregulation of muscle-specific miR-1 noted in hypertrophy can affect cardiac arrhythmogenicity by targeting the GJA1 gene which encodes Cx43 [85]. Gender differences in miR-1 [86] can partly explain higher Cx43 protein levels in females [87] and their lower cardiac arrhythmia susceptibility [55]. It is also important that chronic distress promotes miR-1 expression [35], and this increases Cx43 displacement and induces ventricular tachyarrhythmias in hypertrophic rat hearts [88]. In addition, there is dysregulated miR-1 processing in the SHR heart associated with aging [89]. The implication of miRs in the development of cardiac arrhythmias has also been comprehensively reviewed [90].

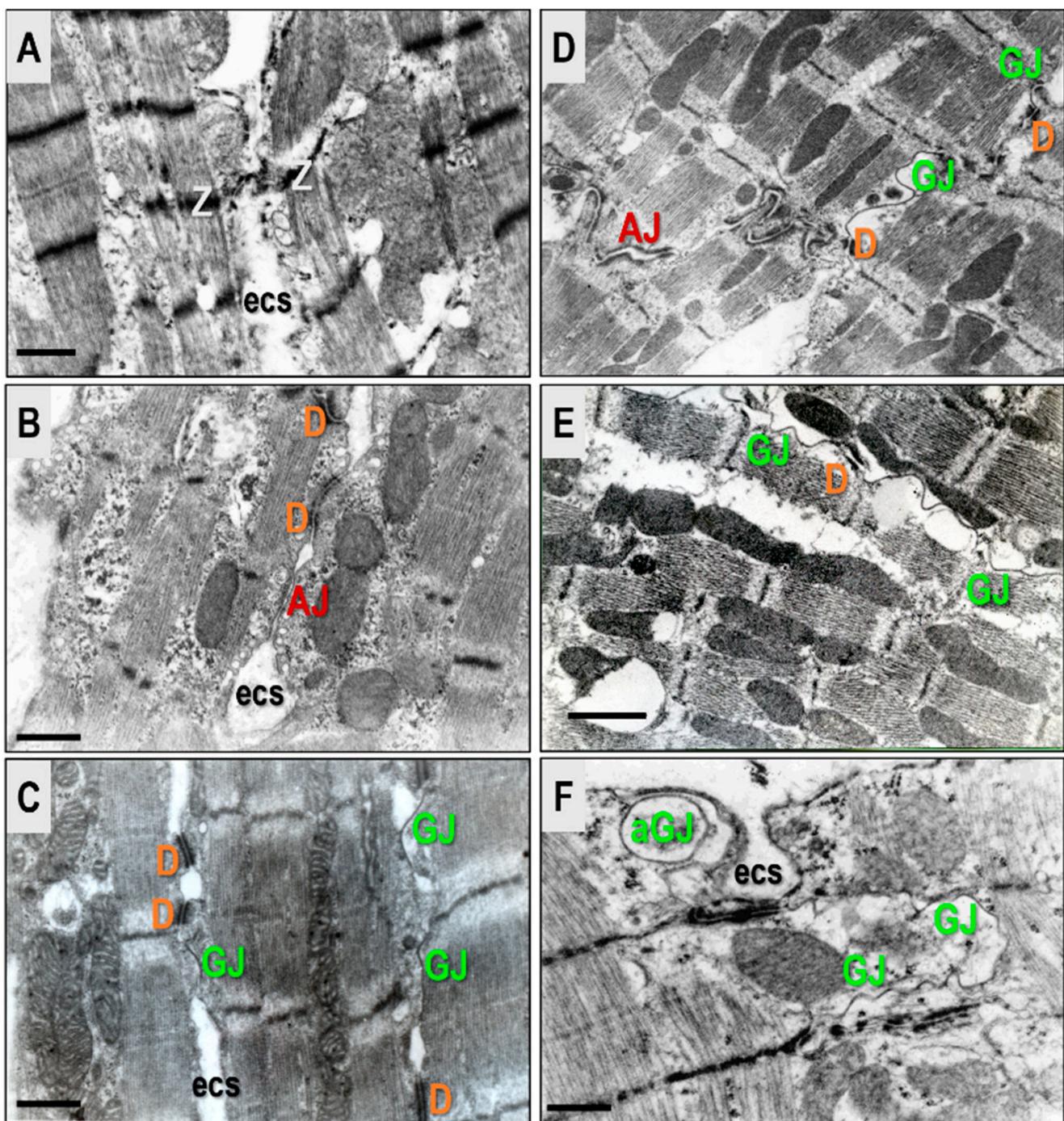


Figure 5. Electron microscope images demonstrate connexome occurrence in SHR hearts. There is formation of the connexome lateral junctions in hypertrophied cardiomyocytes (A–C), and their presence on the lateral sides with those at the intercalated disc (D). A long lateral GJ is obvious in the SHR heart (E), and its degradation and the formation of annular profile (aGJ) occur with HTN progression (F). Scale bar represents 1 micrometer. Adapted from [54,73].

GJ remodeling in human decompensated cardiac hypertrophy is associated with increased interaction of Cx43 with zonula occludens-1 (ZO-1) [91,92]. This could be implicated in the downregulation and decreased size of Cx43 GJs contributing to the arrhythmic substrate. In addition, ZO-1 as a connexin-interacting protein, determines AJ and GJ localization at the intercalated disc [93]. Moreover, high mechanical load induces rapid Cx43 phosphorylation loss, followed by decreased Cx43 protein levels [94]. It has been revealed

that phospho-Ser-368 Cx43 channels were segregated into the GJ center following PKC activation, and these were subsequently internalized and degraded [95]. Of note, ubiquitination is critical for GJ internalization [96], desmin mediates TNF- α -induced aggregate formation, and ID reorganization in the failing heart [97].

Data in the literature indicate proposed factors and mechanisms that may be involved in pro-arrhythmic “connexome” dysfunction in chronic arterial hypertension, as outlined in Figure 7.

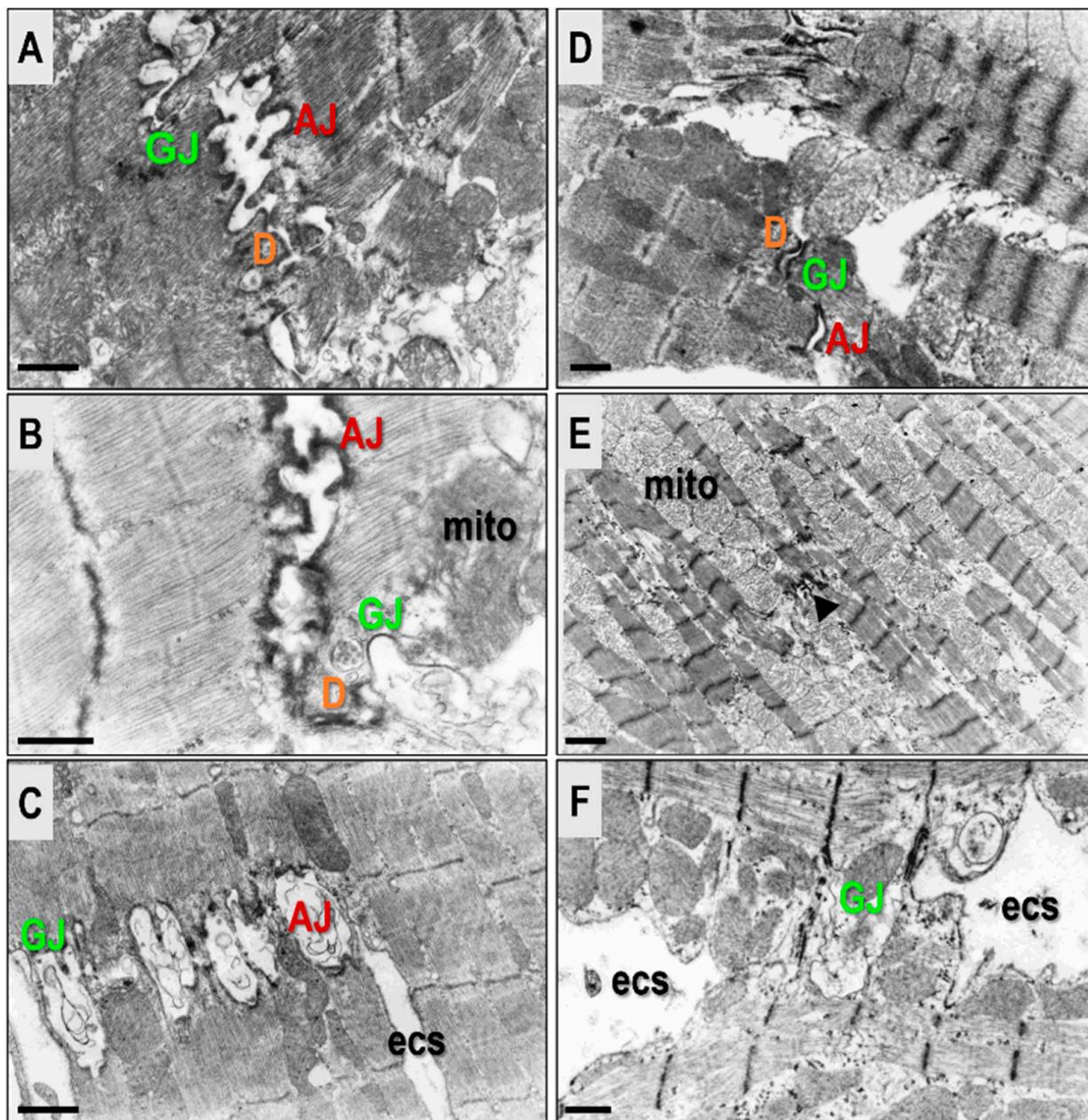


Figure 6. Electron microscope images illustrate connexome impairment in SHR hearts. Varying degrees of AJ dehiscence are apparent in images (A–C), and these are accompanied by GJs loss. The connexome impairment also results in asynchronous contractions of neighbouring cardiomyocytes (A,D). Long-lasting HTN related structural remodeling includes severely injured cardiomyocytes, with the destroyed connexome at the intercalated disc (arrowhead in (E)) and on the cardiomyocytes lateral side (F). This is accompanied with progressive fibrosis. Scale bar represents 1 micrometer. Adapted from [3].

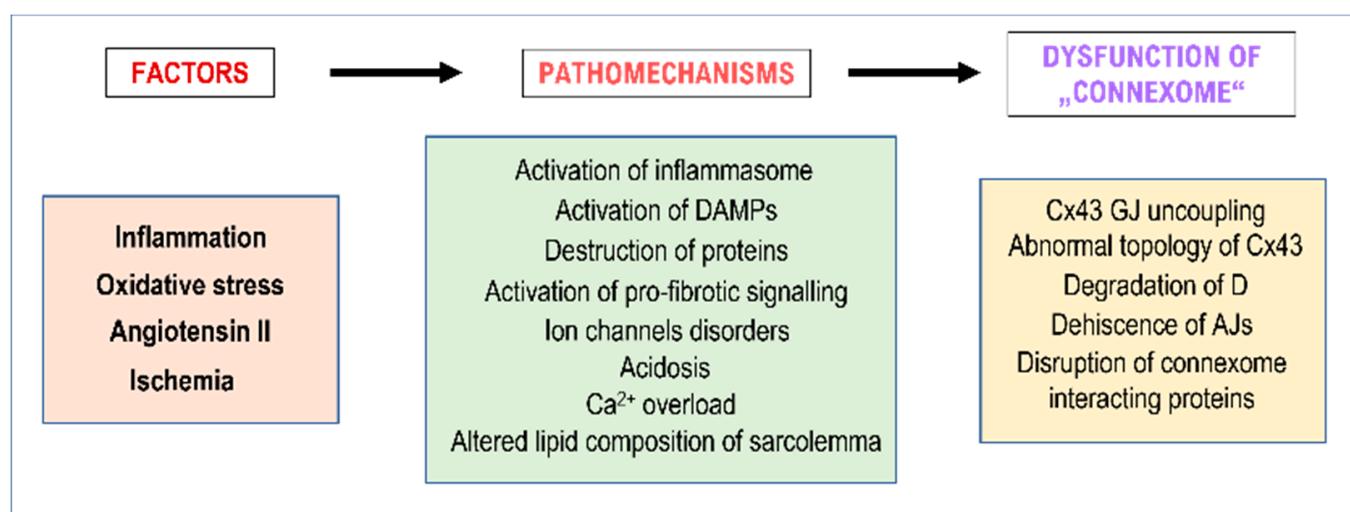


Figure 7. Proposed factors and mechanisms that may be involved in pro-arrhythmic “connexome” dysfunction in chronic arterial hypertension.

Interventions Associated with Protecting Connexome in Primary HTN

The spontaneously hypertensive rat (SHR) is the most frequent experimental model used to imitate human primary HTN. The SHR treatment with the phenylbutyrate short-chain fatty acid derivative and captopril both improved myocardial function, regressed cardiac hypertrophy, and enhanced recovery from HF [98]. It is important here that the gene set combination related to oxidative stress, growth, inflammation, protein degradation, and pro-fibrotic TGF- β signaling were downregulated; and these effects were most likely associated with improved connexome function.

The literature cites the following treatment influences on cellular communication, impulse conduction, and adhesive junctions:

(1) The distribution of Cx43 GJs became more regular and confined to the ID and attenuation of LVH in SHR after treatment with the atorvastatin 3-hydroxy-3-methylglutaryl coenzyme A inhibitor [99]. It is also noted [22] that statins have pleiotropic anti-inflammatory and anti-oxidative effects, and that they alter membrane lipid composition. This could protect connexome function.

(2) The activation of Ang II and AT1 receptors decreases GJ conductance in the failing heart, with consequent impairment of impulse propagation [100]. In addition, both Ang II and aldosterone promote inflammation and enhance collagen deposition and interstitial fibrosis with serious consequences for the spread of electrical activity through the myocardium. Ang II also induces sudden arrhythmic death and electrical remodeling in rats which harbor the human renin and angiotensin genes [68].

(3) The arrhythmias was attributed to inflammation, interstitial fibrosis, reduced transcripts of potassium channel subunit Kv4.3, and gap-junction Cx43 that was partly abolished by losartan. However, a chronic losartan blockade of the Ang II AT1 receptors increased intercellular communication, reduced fibrosis, and improved impulse propagation in the failing heart [100]. Ventricular conductance velocity was also enhanced to some extent by increased GJ conductance, decreased interstitial fibrosis, and structural remodeling.

(4) Candesartan is a further receptor blocker, but its action did not cause fibrosis regression in the SHR’s left atrium at a dose sufficient to reduce blood pressure and left ventricular hypertrophy [101]. In contrast to Ang II, angiotensin (1–7) has an opposite effect on impulse propagation, excitability, and cardiac arrhythmia [102].

(5) Gap junction A1-20k is required for Cx43 passage to the intercalated disc. This attenuates LVH by regulating GJ formation and mitochondrial function [103].

(6) Resolvin D2 protected cardiovascular function and structure when administered before and after the development of Ang II-induced HTN by attenuating inflammation

and fibrosis [104]. It is also important that hydrogen sulfide attenuated inflammation by regulating lymphocyte-confined Cx43 expression in the SHR [105]. This indicates that the resolution of inflammation could be an effective therapy against the target organ damage associated with HTN.

(7) This concept is supported by recent reviews which stressed the pleiotropic antiarrhythmic properties of cardioprotective agents affecting inflammation and oxidative stress [22,106]. These included statin sodium glucose co-transporter-2 inhibitors (SGLT2i) and omega-3 fatty acids. Consequently, these compounds attenuated the downregulation of myocardial Cx43 expression and its abnormal topology, and reduced fibrotic areas in heart ventricles in conditions, such as primary HTN.

(8) However, defective fatty acid uptake is involved in myocardial remodeling in the SHR [107]. In contrast, omega-3 fatty acids intake attenuated Cx43 GJ lateralization and ameliorated the integrity of GJs, AJs, and the sarcolemma in SHR hearts. This rendered them less susceptible to inducible VF [40,55,73,76].

(9) Omega-3 fatty acids decreased the protein kinase PKC δ involved in SHR extracellular matrix remodeling [108]. These acids also increased the PKC ϵ expression associated with Cx43 preservation at the GJs [109].

(10) Dietary omega-3 fatty acids and renin inhibition with aliskiren improved both electrical remodeling and the antiarrhythmic effects attributed to improved Cx43 expression. This also prevented Cx43 redistribution in the model of high human renin primary hypertension [69]. In addition, aliskiren ameliorates maladaptive Cx43 remodeling in the SHR [110].

(11) Finally, early primary HTN therapy can attenuate myocardial structural remodeling and suppress myocardial LVH and fibrosis in the SHR [111]. For example, relaxin can suppress AF by reversing fibrosis and LVH, and it can then increase CV and Na $^{+}$ current in this rat strain [112].

4. Connexome Impairment Promoting VF or AF in PAH

There is an increasing incidence of pulmonary arterial hypertension (PAH) in the general population. This is in addition to problems caused by primary HTN. The elevated pressure in PAH affects the pulmonary circulation and right heart function, and the attendant proliferative vasculopathy results in ongoing increased right ventricular afterload, structural remodeling, and mechanical HF. PAH is multi-factorial, and its etiopathogenesis is not completely understood because of its relationship with underlying somatic disease [49].

PAH is similar to primary HTN, as it is influenced by genetic, epigenetic, and environmental factors. However, hypertension is the most common causative factor of cardiac remodeling [113,114]. PAH also has similar involvement in the renin-angiotensin-aldosterone signaling system, and noncoding RNAs have a prevalent effect as biomarkers and therapeutic targets in preventing heart dysfunction and malignant cardiac arrhythmias [49,115].

Various authors consider that Cx43 downregulation, its dephosphorylation and internalization, and the dysregulated Cx43-mediated signaling in the right ventricle offer crucial intervention targets. [116–119]. In addition, Cx43 heterogeneous expression in the right ventricular outflow tract presents a substrate for idiopathic ventricular arrhythmias [120]. Strauss et. al. (2022) add that predominant right ventricular remodeling promotes multi-wavelet re-entry which underlies ventricular tachycardia [121].

Disorganized GJ distribution and altered anisotropic conduction predispose re-entrant arrhythmias [122]. There is also the implication of endothelin-1 in atrial arrhythmogenesis, and this presents a therapeutic target [123]. However, PAH can be associated with left heart disease, and this is an increasingly prevalent therapeutic problem associated with poor prognosis [124]. This is partly because right ventricular failure induced by blood pressure overload is associated with left ventricular electric remodeling. In addition, reduced Cx43 levels promote this remodeling through impaired cellular impulse transmission [125].

Finally, right heart disease maintains AF due to re-entrant activity, and its underlying substrate involves fibrosis and consequent conduction abnormalities [126].

Interventions Associated with Connexome Protection in PAH

PAH is characterized by reduced angiotensin-converting enzyme-2 activity (ACE2). In contrast, ACE2 augmentation improves pulmonary hemodynamics and reduces oxidant and inflammatory mediator markers [127]. Targeting this pathway most likely protects the connexome and should prove beneficial in PAH.

There were also the following therapeutic interventions:

The benefit of combined nicorandil and colchicine therapy and associated Cx43 preservation in the PAH rat model [128] and attenuated PAH was recorded in this species due to the propylthiouracil thyroid-suppressing agent's pleiotropic properties [129].

The bosentan dual endothelin receptor antagonist partly reversed Cx43 remodeling in the PAH hypertrophied right ventricle [130], and combined sildenafil and beraprost inhibited PAH arrhythmogenesis. This combination substantially suppressed hypertrophy and fibrosis and preserved Cx43 expression [131].

Finally, sildenafil confers protection in the PAH rat model by suppressing pro-fibrotic signaling and enhancing Cx43 in the right ventricle [132], and carbenoxolone decreases PAH-induced pulmonary inflammation and arteriolar remodeling in this model by decreasing T-lymphocyte connexin expression.

5. Lifestyle Recommendation as Treatment for Arterial Hypertension to Protect Connexome

In the context of the topic of this article it should be emphasized that arterial hypertension and high blood pressure-related cardiovascular disease remain global health hazards posing a major socio-economic and healthcare challenge. The global prevalence of hypertension is estimated to be in the range of 30–45% [38]. In Europe, ~25% of heart attacks have been linked with hypertension and ~40% of deaths per annum are caused by hypertension-related cardiovascular disease [133].

Lifestyle interventions are an essential and established part of hypertension management and, in combination with anti-hypertensive drug treatment, provide a most effective strategy to achieve recommended blood pressure targets and reduce cardiovascular mortality. Indeed, guidelines for the management of arterial hypertension strongly recommend lifestyle advice. Mediterranean diet, salt, sugar, and alcohol reduction, smoking cessation, elimination or overcome of stressful events and regular physical activity as well as prevention obesity are essential components not only for the management of hypertension but also for prevention of development of hypertension.

As suggested by most actual systematic reviews [133,134] non-pharmacological factors are the prerequisite for the analysis of research gaps on the way for the future generation of guideline recommendations on lifestyle treatment in patients with hypertension. Education and routine blood pressure screening should be part of the new perspectives in prevention of hypertension in general population including young. Thus, multi-level approach is a future warranty to reducing the public health burden from increased blood pressure [135] and to protect connexome.

6. Conclusions and Perspectives

Research confirms that oxidative stress, a low-grade inflammatory state, and ischemia are involved in hypertension-induced cardiomyocyte junction impairment. Herein we introduce the “connexome” term which combines desmosomes, gap junctions, adherens junctions, and ion channels; and the term is thus defined. The connexome is most important in cardiomyocyte adhesion and the propagation of contractile force, and it is essential for cardiomyocyte coupling which enables electrical and molecular signal transmission. The connexome, therefore, ensures appropriate synchronized heart function; and its impairment is a major component in life-threatening cardiac arrhythmias and heart failure in primary hypertension and pulmonary hypertension. However, there has also been the expressed opinion that preserving the connexome in arterial hypertension treatment can hinder

myocardial structural remodeling, hypertrophy, and fibrosis, as well as the occurrence of cardiac arrhythmias.

In conclusion, while major research confirms the beneficial pleiotropic effects of anti-hypertensive and anti-inflammatory agents, further research is essential to identify specific molecular targets and pathways that protect the connexome. This combination will help formulate new clinical approaches to maintain heart function in patients with arterial hypertension. Moreover, lifestyle advice remains one of the crucial pillars of both anti-hypertensive treatment and hypertension prevention.

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References

1. Danik, S.B.; Liu, F.; Zhang, J.; Suk, H.J.; Morley, G.E.; Fishman, G.I.; Gutstein, D.E. Modulation of cardiac gap junction expression and arrhythmic susceptibility. *Circ. Res.* **2004**, *95*, 1035–1041. [[CrossRef](#)] [[PubMed](#)]
2. Peters, N.S. New insights into myocardial arrhythmogenesis: Distribution of gap-junctional coupling in normal, ischaemic and hypertrophied human hearts. *Clin. Sci.* **1996**, *90*, 447–452. [[CrossRef](#)]
3. Tribulova, N.; Okruhlicova, L.; Novakova, S.; Pancza, D.; Bernatova, I.; Pechanova, O.; Weismann, P.; Manoach, M.; Seki, S.; Mochizuki, S. Hypertension-related intermyocyte junction remodelling is associated with a higher incidence of low-K⁺-induced lethal arrhythmias in isolated rat heart. *Exp. Physiol.* **2002**, *87*, 195–205. [[CrossRef](#)] [[PubMed](#)]
4. Tribulova, N.; Okruhlicova, L.; Imanaga, I.; Hirosawa, N.; Ogawa, K.; Weismann, P. Factors involved in the susceptibility of spontaneously hypertensive rats to low K⁺-induced arrhythmias. *Gen. Physiol. Biophys.* **2003**, *22*, 369–382. [[PubMed](#)]
5. Tribulova, N.; Knezl, V.; Shainberg, A.; Seki, S.; Soukup, T. Thyroid hormones and cardiac arrhythmias. *Vascul. Pharmacol.* **2010**, *52*, 102–112. [[CrossRef](#)]
6. Tribulova, N.; Seki, S.; Radosinska, J.; Kaplan, P.; Babusikova, E.; Knezl, V.; Mochizuki, S. Myocardial Ca²⁺ handling and cell-to-cell coupling, key factors in prevention of sudden cardiac death1. *Can. J. Physiol. Pharmacol.* **2009**, *87*, 1120–1129. [[CrossRef](#)]
7. Andelova, K.; Benova, T.E.; Bacova, B.S.; Sykora, M.; Prado, N.J.; Diez, E.R.; Hlivak, P.; Tribulova, N. Cardiac connexin-43 hemichannels and pannexin1 channels: Provocative antiarrhythmic targets. *Int. J. Mol. Sci.* **2021**, *22*, 260. [[CrossRef](#)] [[PubMed](#)]
8. Saffitz, J.E. Dependence of electrical coupling on mechanical coupling in cardiac myocytes: Insights gained from cardiomyopathies caused by defects in cell-cell connections. *Ann. N. Y. Acad. Sci.* **2005**, *1047*, 336–344. [[CrossRef](#)]
9. Saffitz, J.E.; Hames, K.Y.; Kanno, S. Remodeling of Gap Junctions in Ischemic and Nonischemic Forms of Heart Disease. *J. Membr. Biol.* **2007**, *218*, 65–71. [[CrossRef](#)] [[PubMed](#)]
10. Tribulova, N.; Szeiffova Bacova, B.; Benova, T.; Viczenczova, C. Can we protect from malignant arrhythmias by modulation of cardiac cell-to-cell coupling? *J. Electrocardiol.* **2015**, *48*, 434–440. [[CrossRef](#)]
11. Tribulova, N.; Egan Benova, T.; Szeiffova Bacova, B.; Viczenczova, C.; Barancik, M. New aspects of pathogenesis of atrial fibrillation: Remodeling of intercalated discs. *J. Physiol. Pharmacol.* **2015**, *66*, 625–634. [[PubMed](#)]
12. Sheikh, F.; Ross, R.S.; Chen, J. Cell-Cell Connection to Cardiac Disease. *Trends Cardiovasc. Med.* **2009**, *19*, 182–190. [[CrossRef](#)] [[PubMed](#)]
13. Li, J.; Radice, G.L. A new perspective on intercalated disc organization: Implications for heart disease. *Dermatol. Res. Pract.* **2010**, *2010*, 207835. [[CrossRef](#)]
14. Agullo-Pascual, E.; Cerrone, M.; Delmar, M. Arrhythmogenic cardiomyopathy and Brugada syndrome: Diseases of the connexome. *FEBS Lett.* **2014**, *588*, 1322–1330. [[CrossRef](#)]
15. Moncayo-Arlandi, J.; Brugada, R. Unmasking the molecular link between arrhythmogenic cardiomyopathy and Brugada syndrome. *Nat. Rev. Cardiol.* **2017**, *14*, 744–756. [[CrossRef](#)]
16. Ben-Haim, Y.; Asimaki, A.; Behr, E.R. Brugada syndrome and arrhythmogenic cardiomyopathy: Overlapping disorders of the connexome? *Europace* **2021**, *23*, 653–664. [[CrossRef](#)]

17. Stevens, T.L.; Wallace, M.J.; El Refaey, M.; Roberts, J.D.; Koenig, S.N.; Mohler, P.J. Arrhythmogenic cardiomyopathy: Molecular insights for improved therapeutic design. *J. Cardiovasc. Dev. Dis.* **2020**, *7*, 21. [[CrossRef](#)]
18. Vite, A.; Radice, G.L. N-Cadherin/Catenin Complex as a Master Regulator of Intercalated Disc Function. *Cell Commun. Adhes.* **2014**, *21*, 169–179. [[CrossRef](#)] [[PubMed](#)]
19. Tribulova, N.; Knezl, V.; Szeiffova Bacova, B.; Egan Benova, T.; Viczenczova, C.; Gonçalvesova, E.; Slezak, J. Disordered myocardial Ca^{2+} homeostasis results in substructural alterations that may promote occurrence of malignant arrhythmias. *Physiol. Res.* **2016**, *65*, S139–S148. [[CrossRef](#)]
20. Meens, M.J.; Pfenniger, A.; Kwak, B.R.; Delmar, M. Regulation of cardiovascular connexins by mechanical forces and junctions. *Cardiovasc. Res.* **2013**, *99*, 304–314. [[CrossRef](#)] [[PubMed](#)]
21. Nagibin, V.; Egan Benova, T.; Viczenczova, C.; Szeiffova Bacova, B.; Dovinova, I.; Barancik, M.; Tribulova, N. Ageing related down-regulation of myocardial connexin-43 and up-regulation of MMP-2 may predict propensity to atrial fibrillation in experimental animals. *Physiol. Res.* **2016**, *65*, S91–S100. [[CrossRef](#)]
22. Andelova, K.; Bacova, B.S.; Sykora, M.; Hlivak, P.; Barancik, M.; Tribulova, N. Mechanisms Underlying Antiarrhythmic Properties of Cardioprotective Agents Impacting Inflammation and Oxidative Stress. *Int. J. Mol. Sci.* **2022**, *23*, 1416. [[CrossRef](#)]
23. Fedele, L.; Brand, T. The intrinsic cardiac nervous system and its role in cardiac pacemaking and conduction. *J. Cardiovasc. Dev. Dis.* **2020**, *7*, 54. [[CrossRef](#)] [[PubMed](#)]
24. Nattel, S.; Heijman, J.; Zhou, L.; Dobrev, D. Molecular Basis of Atrial Fibrillation Pathophysiology and Therapy: A Translational Perspective. *Circ. Res.* **2020**, *127*, 51–72. [[CrossRef](#)] [[PubMed](#)]
25. Tribulova, N.; Bacova, B.S.; Benova, T.E.; Knezl, V.; Barancik, M.; Slezak, J. Omega-3 index and anti-arrhythmic potential of omega-3 PUFAs. *Nutrients* **2017**, *9*, 1191. [[CrossRef](#)]
26. Spach, M.S.; Heidlage, J.F.; Dolber, P.C.; Barr, R.C. Electrophysiological effects of remodeling cardiac gap junctions and cell size. Experimental and model studies of normal cardiac growth. *Circ. Res.* **2000**, *86*, 302–311. [[CrossRef](#)]
27. Valderrábano, M. Influence of anisotropic conduction properties in the propagation of the cardiac action potential. *Prog. Biophys. Mol. Biol.* **2008**, *94*, 144–168. [[CrossRef](#)]
28. Gutstein, D.E.; Morley, G.E.; Vaidya, D.; Liu, F.; Chen, F.L.; Stuhlmann, H.; Fishman, G.I. Heterogeneous expression of gap junction channels in the heart leads to conduction defects and ventricular dysfunction. *Circulation* **2001**, *104*, 1194–1199. [[CrossRef](#)] [[PubMed](#)]
29. Gazdag, P.; Oravecz, K.; Acsai, K.; Demeter-Haludka, V.; Ördög, B.; Szlovák, J.; Kohajda, Z.; Polyák, A.; Barta, B.A.; Oláh, A.; et al. Increased Ca^{2+} content of the sarcoplasmic reticulum provides arrhythmogenic trigger source in swimming-induced rat athlete's heart model. *Sci. Rep.* **2020**, *10*, 19596. [[CrossRef](#)]
30. Wu, J.; Liu, T.; Shi, S.; Fan, Z.; Hiram, R.; Xiong, F.; Cui, B.; Su, X.; Chang, R.; Zhang, W.; et al. Dapagliflozin reduces the vulnerability of rats with pulmonary arterial hypertension-induced right heart failure to ventricular arrhythmia by restoring calcium handling. *Cardiovasc. Diabetol.* **2022**, *21*, 197. [[CrossRef](#)]
31. Tribulova, N.; Kurahara, L.H.; Hlivak, P.; Hirano, K.; Bacova, B.S. Pro-arrhythmic signaling of thyroid hormones and its relevance in subclinical hyperthyroidism. *Int. J. Mol. Sci.* **2020**, *21*, 2844. [[CrossRef](#)]
32. Patel, S.; Rauf, A.; Khan, H.; Abu-Izneid, T. Renin-angiotensin-aldosterone (RAAS): The ubiquitous system for homeostasis and pathologies. *Biomed. Pharmacother.* **2017**, *94*, 317–325. [[CrossRef](#)] [[PubMed](#)]
33. Li, J.; Wang, S.; Bai, J.; Yang, X.L.; Zhang, Y.L.; Che, Y.L.; Li, H.H.; Yang, Y.Z. Novel role for the immunoproteasome subunit PSMB10 in angiotensin II-induced atrial fibrillation in Mice. *Hypertension* **2018**, *71*, 866–876. [[CrossRef](#)] [[PubMed](#)]
34. Stambler, B.S.; Ryu, K.P. Atrial natriuretic peptide accelerates onset and dynamics of ventricular fibrillation during hypokalemia in isolated rabbit hearts. *J. Electrocardiol.* **2020**, *62*, 184–189. [[CrossRef](#)] [[PubMed](#)]
35. Wahl, C.M.; Schmidt, C.; Hecker, M.; Ullrich, N.D. Distress-Mediated Remodeling of Cardiac Connexin-43 in a Novel Cell Model for Arrhythmogenic Heart Diseases. *Int. J. Mol. Sci.* **2022**, *23*, 10174. [[CrossRef](#)]
36. Hoagland, D.T.; Santos, W.; Poelzing, S.; Gourdie, R.G. The role of the gap junction perinexus in cardiac conduction: Potential as a novel anti-arrhythmic drug target. *Prog. Biophys. Mol. Biol.* **2019**, *144*, 41–50. [[CrossRef](#)]
37. Suvila, K.; Niiranen, T.J. Age of Hypertension Onset: Potential for Improving Risk Estimation and Hypertension Management? *Hypertension* **2021**, *78*, 1475–1477. [[CrossRef](#)]
38. De Simone, G.; Mancusi, C.; Hanssen, H.; Genovesi, S.; Lurbe, E.; Parati, G.; Sendzikaitė, S.; Valerio, G.; Di Bonito, P.; Di Salvo, G.; et al. Hypertension in children and adolescents. *Eur. Heart J.* **2022**, *43*, 3290–3301. [[CrossRef](#)]
39. Li, L.; Yi-Ming, W.; Li, Z.Z.; Zhao, L.; Yu, Y.S.; Li, D.J.; Xia, C.Y.; Liu, J.G.; Su, D.F. Local RAS and inflammatory factors are involved in cardiovascular hypertrophy in spontaneously hypertensive rats. *Pharmacol. Res.* **2008**, *58*, 196–201. [[CrossRef](#)]
40. Bacova, B.S.; Radosinska, J.; Wallukat, G.; Barancik, M.; Wallukat, A.; Knezl, V.; Sykora, M.; Paulis, L.; Tribulova, N. Suppression of $\beta 1$ -adrenoceptor autoantibodies is involved in the antiarrhythmic effects of omega-3 fatty acids in male and female hypertensive rats. *Int. J. Mol. Sci.* **2020**, *21*, 526. [[CrossRef](#)]
41. Bai, L.; Qu, C.; Feng, Y.; Liu, G.; Li, X.; Li, W.; Yu, S. Evidence of a causal relationship between vitamin D deficiency and hypertension: A family-based study. *Hypertens. Res.* **2022**, *45*, 1814–1822. [[CrossRef](#)]
42. Bayés-Genís, A.; Díez, J. Transition to heart failure in hypertension: Going to the heart of the matter. *Eur. Heart J.* **2022**, *43*, 3332–3334. [[CrossRef](#)] [[PubMed](#)]

43. Wang, M.; Brage, S.; Sharp, S.J.; Luo, S.; Au Yeung, S.L.; Kim, Y. Associations of genetic susceptibility and healthy lifestyle with incidence of coronary heart disease and stroke in individuals with hypertension. *Eur. J. Prev. Cardiol.* **2022**, *29*, 2101–2110. [[CrossRef](#)] [[PubMed](#)]
44. Vaura, F.; Kim, H.; Udler, M.S.; Salomaa, V.; Lahti, L.; Niiranen, T. Multi-Trait Genetic Analysis Reveals Clinically Interpretable Hypertension Subtypes. *Circ. Genom. Precis. Med.* **2022**, *15*, e003583. [[CrossRef](#)]
45. Takahashi, H.; Yoshika, M.; Komiyama, Y.; Nishimura, M. The central mechanism underlying hypertension: A review of the roles of sodium ions, epithelial sodium channels, the renin-angiotensin-aldosterone system, oxidative stress and endogenous digitalis in the brain. *Hypertens. Res.* **2011**, *34*, 1147–1160. [[CrossRef](#)] [[PubMed](#)]
46. Zhang, H.C.; Zhang, Z.S.; Zhang, L.; Wang, A.; Zhu, H.; Li, L.; Si, J.Q.; Li, X.Z.; Ma, K.T. Connexin 43 in splenic lymphocytes is involved in the regulation of CD4+CD25+ T lymphocyte proliferation and cytokine production in hypertensive inflammation. *Int. J. Mol. Med.* **2018**, *41*, 13–24. [[CrossRef](#)]
47. Wang, L.J.; Zhang, W.W.; Zhang, L.; Shi, W.Y.; Wang, Y.Z.; Ma, K.T.; Liu, W.D.; Zhao, L.; Li, L.; Si, J.Q. Association of connexin gene polymorphism with essential hypertension in Kazak and Han Chinese in Xinjiang, China. *J. Huazhong Univ. Sci. Technol.—Med. Sci.* **2017**, *37*, 197–203. [[CrossRef](#)]
48. Arif, M.; Sadayappan, S.; Becker, R.C.; Martin, L.J.; Urbina, E.M. Epigenetic modification: A regulatory mechanism in essential hypertension. *Hypertens. Res.* **2019**, *42*, 1099–1113. [[CrossRef](#)]
49. Jusic, A.; Devaux, Y. Noncoding RNAs in Hypertension. *Hypertension* **2019**, *74*, 477–492. [[CrossRef](#)]
50. Rodrigo, R.; Prat, H.; Passalacqua, W.; Araya, J.; Bächler, J.P. Decrease in oxidative stress through supplementation of vitamins C and E is associated with a reduction in blood pressure in patients with essential hypertension. *Clin. Sci.* **2008**, *114*, 625–634. [[CrossRef](#)]
51. Rodrigo, R.; González, J.; Paoletto, F. The role of oxidative stress in the pathophysiology of hypertension. *Hypertens. Res.* **2011**, *34*, 431–440. [[CrossRef](#)]
52. Rodriguez-Iturbe, B.; Johnson, R.J. Genetic Polymorphisms in Hypertension: Are We Missing the Immune Connection? *Am. J. Hypertens.* **2019**, *32*, 113–122. [[CrossRef](#)] [[PubMed](#)]
53. Brown, J.M.; Zhou, W.; Weber, B.; Divakaran, S.; Barrett, L.; Bibbo, C.F.; Hainer, J.; Taqueti, V.R.; Dorbala, S.; Blankstein, R.; et al. Low coronary flow relative to myocardial mass predicts heart failure in symptomatic hypertensive patients with no obstructive coronary artery disease. *Eur. Heart J.* **2022**, *43*, 3323–3331. [[CrossRef](#)] [[PubMed](#)]
54. Egan Benova, T.; Szeiffova Bacova, B.; Viczenczova, C.; Diez, E.; Barancik, M.; Tribulova, N. Protection of cardiac cell-to-cell coupling attenuate myocardial remodeling and proarrhythmia induced by hypertension. *Physiol. Res.* **2016**, *65*, S29–S42. [[CrossRef](#)]
55. Benova, T.E.; Viczenczova, C.; Bacova, B.S.; Zurmanova, J.; Knezl, V.; Andelova, K.; Tribulova, N. Omacor protects normotensive and hypertensive rats exposed to continuous light from increased risk to malignant cardiac arrhythmias. *Mar. Drugs* **2021**, *19*, 659. [[CrossRef](#)] [[PubMed](#)]
56. Dzau, V.J.; Balatbat, C.A. Future of hypertension: The need for transformation. *Hypertension* **2019**, *74*, 450–457. [[CrossRef](#)]
57. Marazzato, J.; Blasi, F.; Golino, M.; Verdecchia, P.; Angeli, F.; De Ponti, R. Hypertension and Arrhythmias: A Clinical Overview of the Pathophysiology-Driven Management of Cardiac Arrhythmias in Hypertensive Patients. *J. Cardiovasc. Dev. Dis.* **2022**, *9*, 110. [[CrossRef](#)] [[PubMed](#)]
58. Crea, F. Hypertension in children, adolescents, and pregnant women: Challenges and opportunities. *Eur. Heart J.* **2022**, *43*, 3275–3278. [[CrossRef](#)]
59. Bacharova, L.; Plandorova, J.; Klimas, J.; Krenek, P.; Kyselovic, J. Discrepancy between increased left ventricular mass and “normal” QRS voltage is associated with decreased connexin 43 expression in early stage of left ventricular hypertrophy in spontaneously hypertensive rats. *J. Electrocardiol.* **2008**, *41*, 730–734. [[CrossRef](#)] [[PubMed](#)]
60. Hussain, W.; Patel, P.M.; Chowdhury, R.A.; Cabo, C.; Ciaccio, E.J.; Lab, M.J.; Duffy, H.S.; Wit, A.L.; Peters, N.S. The Renin-Angiotensin System Mediates the Effects of Stretch on Conduction Velocity, Connexin43 Expression, and Redistribution in Intact Ventricle. *J. Cardiovasc. Electrophysiol.* **2010**, *21*, 1276–1283. [[CrossRef](#)]
61. Teunissen, B.E.J.; Jongsma, H.J.; Bierhuizen, M.F.A. Regulation of myocardial connexins during hypertrophic remodelling. *Eur. Heart J.* **2004**, *25*, 1979–1989. [[CrossRef](#)] [[PubMed](#)]
62. Bačová, B.; Radošinská, J.; Viczenczová, C.; Knezl, V.; Dosenko, V.; Beňova, T.; Navarová, J.; Gonçalvesová, E.; van Rooyen, J.; Weismann, P.; et al. Up-regulation of myocardial connexin-43 in spontaneously hypertensive rats fed red palm oil is most likely implicated in its anti-arrhythmic effects. *Can. J. Physiol. Pharmacol.* **2012**, *90*, 1235–1245. [[CrossRef](#)] [[PubMed](#)]
63. Strauss, B.; Sassi, Y.; Bueno-Beti, C.; Ilkan, Z.; Raad, N.; Cacheux, M.; Bisserier, M.; Turnbull, I.C.; Kohlbrenner, E.; Hajjar, R.J.; et al. Intra-tracheal gene delivery of aerosolized SERCA2a to the lung suppresses ventricular arrhythmias in a model of pulmonary arterial hypertension. *J. Mol. Cell. Cardiol.* **2019**, *127*, 20–30. [[CrossRef](#)]
64. Bacova, B.S.; Viczenczova, C.; Andelova, K.; Sykora, M.; Chaudagar, K.; Barancik, M.; Adamcova, M.; Knezl, V.; Benova, T.E.; Weismann, P.; et al. Antiarrhythmic effects of melatonin and omega-3 are linked with protection of myocardial cx43 topology and suppression of fibrosis in catecholamine stressed normotensive and hypertensive rats. *Antioxidants* **2020**, *9*, 546. [[CrossRef](#)]
65. Hein, S.; Elsässer, A.; Kostin, S.; Zimmermann, R.; Schaper, J. Functional disturbances due to structural remodeling in the failing human heart. *Arch. Mal. Coeur Vaiss.* **2002**, *95*, 815–820.

66. Hein, S.; Arnon, E.; Kostin, S.; Schönburg, M.; Elsässer, A.; Polyakova, V.; Bauer, E.P.; Klövekorn, W.P.; Schaper, J. Progression from compensated hypertrophy to failure in the pressure-overloaded human: Heart structural deterioration and compensatory mechanisms. *Circulation* **2003**, *107*, 984–991. [[CrossRef](#)] [[PubMed](#)]
67. Kostin, S.; Dammer, S.; Hein, S.; Klocekorn, W.P.; Bauer, E.P.; Schaper, J. Connexin 43 expression and distribution in compensated and decompensated cardiac hypertrophy in patients with aortic stenosis. *Cardiovasc. Res.* **2004**, *62*, 426–436. [[CrossRef](#)]
68. Fischer, R.; Dechend, R.; Gapelyuk, A.; Shagdarsuren, E.; Gruner, K.; Gruner, A.; Gratze, P.; Qadri, F.; Wellner, M.; Fiebeler, A.; et al. Angiotensin II-induced sudden arrhythmic death and electrical remodeling. *Am. J. Physiol.—Heart Circ. Physiol.* **2007**, *293*, 1242–1253. [[CrossRef](#)] [[PubMed](#)]
69. Fischer, R.; Dechend, R.; Qadri, F.; Markovic, M.; Feldt, S.; Herse, F.; Park, J.K.; Gapelyuk, A.; Schwarz, I.; Zacharzowsky, U.B.; et al. Dietary n-3 polyunsaturated fatty acids and direct renin inhibition improve electrical remodeling in a model of high human renin hypertension. *Hypertension* **2008**, *51*, 540–546. [[CrossRef](#)] [[PubMed](#)]
70. Kasi, V.S.; Xiao, H.D.; Shang, L.L.; Iravani, S.; Langberg, J.; Witham, E.A.; Jiao, Z.; Gallego, C.J.; Bernstein, K.E.; Dudley, S.C. Cardiac-restricted angiotensin-converting enzyme overexpression causes conduction defects and connexin dysregulation. *Am. J. Physiol.—Heart Circ. Physiol.* **2007**, *293*, H182–H192. [[CrossRef](#)]
71. Monitillo, F.; Di Terlizzi, V.; Gioia, M.I.; Barone, R.; Grande, D.; Parisi, G.; Brunetti, N.D.; Iacoviello, M. Right ventricular function in chronic heart failure: From the diagnosis to the therapeutic approach. *J. Cardiovasc. Dev. Dis.* **2020**, *7*, 12. [[CrossRef](#)] [[PubMed](#)]
72. Hofgaard, J.P.; Banach, K.; Mollerup, S.; Jørgensen, H.K.; Olesen, S.P.; Holstein-Rathlou, N.-H.; Nielsen, M.S. Phosphatidylinositol-bisphosphate regulates intercellular coupling in cardiac myocytes. *Pflügers Arch.—Eur. J. Physiol.* **2008**, *457*, 303–313. [[CrossRef](#)]
73. Radosinska, J.; Bacova, B.; Knezl, V.; Benova, T.; Zurmanova, J.; Soukup, T.; Arnostova, P.; Slezak, J.; Goncalvesova, E.; Tribulova, N. Dietary omega-3 fatty acids attenuate myocardial arrhythmogenic factors and propensity of the heart to lethal arrhythmias in a rodent model of human essential hypertension. *J. Hypertens.* **2013**, *31*, 1876–1885. [[CrossRef](#)]
74. Kvakan, H.; Kleinewietfeld, M.; Qadri, F.; Park, J.K.; Fischer, R.; Schwarz, I.; Rahn, H.P.; Plehm, R.; Wellner, M.; Elitok, S.; et al. Regulatory T cells ameliorate angiotensin II-induced cardiac damage. *Circulation* **2009**, *119*, 2904–2912. [[CrossRef](#)] [[PubMed](#)]
75. Hesketh, G.G.; Shah, M.H.; Halperin, V.L.; Cooke, C.A.; Akar, F.G.; Yen, T.E.; Kass, D.A.; MacHamer, C.E.; Van Eyk, J.E.; Tomaselli, G.F. Ultrastructure and regulation of lateralized connexin43 in the failing heart. *Circ. Res.* **2010**, *106*, 1153–1163. [[CrossRef](#)] [[PubMed](#)]
76. Benova, T.; Viczenczova, C.; Radosinska, J.; Bacova, B.; Knezl, V.; Dosenko, V.; Weismann, P.; Zeman, M.; Navarova, J.; Tribulova, N. Melatonin attenuates hypertension-related proarrhythmic myocardial maladaptation of connexin-43 and propensity of the heart to lethal arrhythmias. *Can. J. Physiol. Pharmacol.* **2013**, *91*, 633–639. [[CrossRef](#)]
77. Andelova, K.; Szeiffova Bacova, B.; Sykora, M.; Pavelka, S.; Rauchova, H.; Tribulova, N. Cardiac Cx43 Signaling Is Enhanced and TGF- β 1/SMAD2/3 Suppressed in Response to Cold Acclimation and Modulated by Thyroid Status in Hairless SHRM. *Biomedicines* **2022**, *10*, 1707. [[CrossRef](#)]
78. Seidel, T.; Salameh, A.; Dhein, S. A simulation study of cellular hypertrophy and connexin lateralization in cardiac tissue. *Biophys. J.* **2010**, *99*, 2821–2830. [[CrossRef](#)]
79. Chkourko, H.S.; Guerrero-Serna, G.; Lin, X.; Darwish, N.; Pohlmann, J.R.; Cook, K.E.; Martens, J.R.; Rothenberg, E.; Musa, H.; Delmar, M. Remodeling of mechanical junctions and of microtubule-associated proteins accompany cardiac connexin43 lateralization. *Heart Rhythm* **2012**, *9*, 1133–1140.e6. [[CrossRef](#)]
80. Craig, M.A.; McBride, M.W.; Smith, G.; George, S.J.; Baker, A. Dysregulation of cadherins in the intercalated disc of the spontaneously hypertensive stroke-prone rat. *J. Mol. Cell. Cardiol.* **2010**, *48*, 1121–1128. [[CrossRef](#)]
81. Jansen, J.A.; Van Veen, T.A.B.; De Jong, S.; Van Der Nagel, R.; Van Stuijvenberg, L.; Driessen, H.; Labzowski, R.; Oefner, C.M.; Bosch, A.A.; Nguyen, T.Q.; et al. Reduced Cx43 expression triggers increased fibrosis due to enhanced fibroblast activity. *Circ. Arrhythmia Electrophysiol.* **2012**, *5*, 380–390. [[CrossRef](#)]
82. Rohr, S. Arrhythmogenic implications of fibroblast-myocyte interactions. *Circ. Arrhythmia Electrophysiol.* **2012**, *5*, 442–452. [[CrossRef](#)] [[PubMed](#)]
83. Cooklin, M.; Wallis, W.R.J.; Sheridan, D.J.; Fry, C.H. Changes in Cell-to-Cell Electrical Coupling Associated With Left Ventricular Hypertrophy. *Circ. Res.* **1997**, *80*, 765–771. [[CrossRef](#)] [[PubMed](#)]
84. Wehbe, N.; Nasser, S.A.; Pintus, G.; Badran, A.; Eid, A.H.; Baydoun, E. MicroRNAs in Cardiac Hypertrophy. *Int. J. Mol. Sci.* **2019**, *20*, 4714. [[CrossRef](#)] [[PubMed](#)]
85. Yang, B.; Lin, H.; Xiao, J.; Lu, Y.; Luo, X.; Li, B.; Zhang, Y.; Xu, C.; Bai, Y.; Wang, H.; et al. The muscle-specific microRNA miR-1 regulates cardiac arrhythmogenic potential by targeting GJA1 and KCNJ2. *Nat. Med.* **2007**, *13*, 486–491. [[CrossRef](#)] [[PubMed](#)]
86. Stauffer, B.L.; Sobus, R.D.; Sucharov, C.C. Sex differences in cardiomyocyte connexin43 expression. *J. Cardiovasc. Pharmacol.* **2011**, *58*, 32–39. [[CrossRef](#)]
87. Tribulová, N.; Dupont, E.; Soukup, T.; Okruhlicová, L.; Severs, N.J. Sex differences in connexin-43 expression in left ventricles of aging rats. *Physiol. Res.* **2005**, *54*, 705–708. [[CrossRef](#)]
88. Curcio, A.; Torella, D.; Iaconetti, C.; Pasceri, E.; Sabatino, J.; Sorrentino, S.; Giampà, S.; Micieli, M.; Polimeni, A.; Henning, B.J.; et al. MicroRNA-1 Downregulation Increases Connexin 43 Displacement and Induces Ventricular Tachyarrhythmias in Rodent Hypertrophic Hearts. *PLoS ONE* **2013**, *8*, e0070158. [[CrossRef](#)]

89. Lapikova-Bryhinska, T.; Zhukovska, A.; Nagibin, V.; Tumanovska, L.; Portnichenko, G.; Goncharov, S.; Portnychenko, A.; Dosenko, V. Altered biogenesis of microRNA-1 is associated with cardiac dysfunction in aging of spontaneously hypertensive rats. *Mol. Cell. Biochem.* **2019**, *459*, 73–82. [CrossRef]
90. Luo, X.; Yang, B.; Nattel, S. MicroRNAs and atrial fibrillation: Mechanisms and translational potential. *Nat. Rev. Cardiol.* **2015**, *12*, 80–90. [CrossRef]
91. Bruce, A.F.; Rothery, S.; Dupont, E.; Severs, N.J. Gap junction remodelling in human heart failure is associated with increased interaction of connexin43 with ZO-1. *Cardiovasc. Res.* **2008**, *77*, 757–765. [CrossRef] [PubMed]
92. Kostin, S. Zonula occludens-1 and connexin 43 expression in the failing human heart. *J. Cell. Mol. Med.* **2007**, *11*, 892–895. [CrossRef] [PubMed]
93. Palatinus, J.A.; Rhett, J.M.; Gourdie, R.G. Enhanced PKC ϵ mediated phosphorylation of connexin43 at serine 368 by a carboxyl-terminal mimetic peptide is dependent on injury. *Channels* **2011**, *5*, 236–240. [CrossRef] [PubMed]
94. Bupha-Intr, T.; Haizlip, K.M.; Janssen, P.M.L. Temporal changes in expression of connexin 43 after load-induced hypertrophy in vitro. *Am. J. Physiol.—Heart Circ. Physiol.* **2009**, *296*, 806–814. [CrossRef]
95. Cone, A.C.; Cavin, G.; Ambrosi, C.; Hakozaki, H.; Wu-Zhang, A.X.; Kunkel, M.T.; Newton, A.C.; Sosinsky, G.E. Protein kinase C δ -mediated phosphorylation of connexin43 gap junction channels causes movement within gap junctions followed by vesicle internalization and protein degradation. *J. Biol. Chem.* **2014**, *289*, 8781–8798. [CrossRef]
96. Kells-Andrews, R.M.; Margraf, R.A.; Fisher, C.G.; Falk, M.M. Connexin 43 K63-polyubiquitylation on lysines 264 and 303 regulates gap junction internalization. *J. Cell Sci.* **2018**, *131*, jcs204321. [CrossRef] [PubMed]
97. Panagopoulou, P.; Davos, C.H.; Milner, D.J.; Varela, E.; Cameron, J.A.; Mann, D.L.; Capetanaki, Y. Desmin mediates TNF- α -induced aggregate formation and intercalated disk reorganization in heart failure. *J. Cell Biol.* **2008**, *181*, 761–775. [CrossRef]
98. Brooks, W.W.; Shen, S.; Conrad, C.H.; Goldstein, R.H.; Deng, L.L.; Bing, O.H.L. Transcriptional changes associated with recovery from heart failure in the SHR. *J. Mol. Cell. Cardiol.* **2010**, *49*, 390–401. [CrossRef]
99. Chen, H.J.; Yao, L.; Chen, T.G.; Yu, M.; Wang, L.H.; Chen, J.Z. Atorvastatin prevents connexin43 remodeling in hypertrophied left ventricular myocardium of spontaneously hypertensive rats. *Chin. Med. J.* **2007**, *120*, 1902–1907. [CrossRef]
100. De Mello, W.C.; Specht, P. Chronic blockade of angiotensin II AT1-receptors increased cell-to-cell communication, reduced fibrosis and improved impulse propagation in the failing heart. *JRAAS—J. Renin-Angiotensin-Aldosterone Syst.* **2006**, *7*, 201–205. [CrossRef]
101. Choisy, S.C.; Kim, S.J.; Hancox, J.C.; Jones, S.A.; James, A.F. Effects of candesartan, an angiotensin II receptor type I blocker, on atrial remodeling in spontaneously hypertensive rats. *Physiol. Rep.* **2015**, *3*, e12274. [CrossRef] [PubMed]
102. De Mello, W.C. Opposite effects of angiotensin II and angiotensin (1-7) on impulse propagation, excitability and cardiac arrhythmias. Is the overexpression of ACE2 arrhythmicogenic? *Regul. Pept.* **2009**, *153*, 7–10. [CrossRef] [PubMed]
103. Fu, Y.L.; Tao, L.; Peng, F.H.; Zheng, N.Z.; Lin, Q.; Cai, S.Y.; Wang, Q. GJA1-20k attenuates Ang II-induced pathological cardiac hypertrophy by regulating gap junction formation and mitochondrial function. *Acta Pharmacol. Sin.* **2021**, *42*, 536–549. [CrossRef] [PubMed]
104. Díaz del Campo, L.S.; García-Redondo, A.B.; Rodriguez, C.; Zaragoza, C.; Duro-Sánchez, S.; Palmas, F.; de Benito-Bueno, A.; Socuéllamos, P.G.; Peraza, D.A.; Rodrigues-Diez, R.; et al. Resolvin D2 Attenuates Cardiovascular Damage in Angiotensin II-Induced Hypertension. *Hypertension* **2023**, *80*, 84–96. [CrossRef] [PubMed]
105. Ni, X.; Zhang, L.; Peng, M.; Shen, T.W.; Yu, X.S.; Shan, L.Y.; Li, L.; Si, J.Q.; Li, X.Z.; Ma, K.T. Hydrogen sulfide attenuates hypertensive inflammation via regulating connexin expression in spontaneously hypertensive rats. *Med. Sci. Monit.* **2018**, *24*, 1205–1218. [CrossRef] [PubMed]
106. Szeiffova Bacova, B.; Andelova, K.; Sykora, M.; Egan Benova, T.; Barancik, M.; Kurahara, L.H.; Tribulova, N. Does myocardial atrophy represent anti-arrhythmic phenotype? *Biomedicines* **2022**, *10*, 2819. [CrossRef]
107. Hajri, T.; Ibrahimi, A.; Coburn, C.T.; Knapp, F.F.; Kurtz, T.; Pravenec, M.; Abumrad, N.A. Defective Fatty Acid Uptake in the Spontaneously Hypertensive Rat Is a Primary Determinant of Altered Glucose Metabolism, Hyperinsulinemia, and Myocardial Hypertrophy. *J. Biol. Chem.* **2001**, *276*, 23661–23666. [CrossRef]
108. STEINBERG, S.F. Distinctive activation mechanisms and functions for protein kinase C δ . *Biochem. J.* **2004**, *384*, 449–459. [CrossRef]
109. Kohutova, J.; Elsnicova, B.; Holzerova, K.; Neckar, J.; Sebesta, O.; Jezkova, J.; Vecka, M.; Vebr, P.; Hornikova, D.; Bacova, B.S.; et al. Anti-arrhythmic cardiac phenotype elicited by chronic intermittent hypoxia is associated with alterations in connexin-43 expression, phosphorylation, and distribution. *Front. Endocrinol.* **2019**, *10*, 1–10. [CrossRef]
110. Zhang, W.; Zhao, G.; Hu, X.; Wang, M.; Li, H.; Ye, Y.; Du, Q.; Yao, J.; Bao, Z.; Hong, W.; et al. Aliskiren-attenuated myocardium apoptosis via regulation of autophagy and connexin-43 in aged spontaneously hypertensive rats. *J. Cell. Mol. Med.* **2014**, *18*, 1247–1256. [CrossRef]
111. Rassler, B.; Hawlitschek, C.; Brendel, J.; Zimmer, H.-G. How Do Young and Old Spontaneously Hypertensive Rats Respond to Antihypertensive Therapy? Comparative Studies on the Effects of Combined Captopril and Nifedipine Treatment. *Biomedicines* **2022**, *10*, 3059. [CrossRef]
112. Parikh, A.; Patel, D.; McTiernan, C.F.; Xiang, W.; Haney, J.; Yang, L.; Lin, B.; Kaplan, A.D.; Bett, G.C.L.; Rasmusson, R.L.; et al. Relaxin suppresses atrial fibrillation by reversing fibrosis and myocyte hypertrophy and increasing conduction velocity and sodium current in spontaneously hypertensive rat hearts. *Circ. Res.* **2013**, *113*, 313–321. [CrossRef] [PubMed]
113. Mavrogeni, S.; Piaditis, G.; Bacopoulou, F.; Chrousos, G.P. Cardiac Remodeling in Hypertension: Clinical Impact on Brain, Heart, and Kidney Function. *Horm. Metab. Res.* **2022**, *54*, 273–279. [CrossRef] [PubMed]

114. Wang, Z.; Schreier, D.A.; Hacker, T.A.; Chesler, N.C. Progressive right ventricular functional and structural changes in a mouse model of pulmonary arterial hypertension. *Physiol. Rep.* **2013**, *1*, e00184. [[CrossRef](#)] [[PubMed](#)]
115. Omura, J.; Habbout, K.; Shimauchi, T.; Wu, W.H.; Breuils-Bonnet, S.; Tremblay, E.; Martineau, S.; Nadeau, V.; Gagnon, K.; Mazoyer, F.; et al. Identification of Long Noncoding RNA H19 as a New Biomarker and Therapeutic Target in Right Ventricular Failure in Pulmonary Arterial Hypertension. *Circulation* **2020**, *142*, 1464–1484. [[CrossRef](#)] [[PubMed](#)]
116. Sasano, C.; Honjo, H.; Takagishi, Y.; Uzzaman, M.; Emdad, L.; Shimizu, A.; Murata, Y.; Kamiya, K.; Kodama, I. Internalization and dephosphorylation of Connexin43 in hypertrophied right ventricles of rats with pulmonary hypertension. *Circ. J.* **2007**, *71*, 382–389. [[CrossRef](#)]
117. Bouvard, C.; Genet, N.; Phan, C.; Rode, B.; Thuillet, R.; Tu, L.; Robillard, P.; Campagnac, M.; Soleti, R.; de la Roque, E.D.; et al. Connexin-43 is a promising target for pulmonary hypertension due to hypoxaemic lung disease. *Eur. Respir. J.* **2020**, *55*, 1900169. [[CrossRef](#)]
118. Boengler, K.; Rohrbach, S.; Weissmann, N.; Schulz, R. Importance of CX43 for right ventricular function. *Int. J. Mol. Sci.* **2021**, *22*, 987. [[CrossRef](#)]
119. Htet, M.; Nally, J.E.; Martin, P.E.; Dempsie, Y. New insights into pulmonary hypertension: A role for connexin-mediated signalling. *Int. J. Mol. Sci.* **2022**, *23*, 379. [[CrossRef](#)]
120. Ou, B.; Nakagawa, M.; Kajimoto, M.; Nobe, S.; Ooie, T.; Ichinose, M.; Yonemochi, H.; Ono, N.; Shimada, T.; Saikawa, T. Heterogeneous expression of connexin 43 in the myocardium of rabbit right ventricular outflow tract. *Life Sci.* **2005**, *77*, 52–59. [[CrossRef](#)]
121. Strauss, B.; Bisserier, M.; Obus, E.; Katz, M.G.; Farnoli, A.; Cacheux, M.; Akar, J.G.; Hummel, J.P.; Hadri, L.; Sassi, Y.; et al. Right predominant electrical remodeling in a pure model of pulmonary hypertension promotes reentrant arrhythmias. *Heart Rhythm* **2022**, *19*, 113–124. [[CrossRef](#)] [[PubMed](#)]
122. Uzzaman, M.; Honjo, H.; Takagishi, Y.; Emdad, L.; Magee, A.I.; Severs, N.J.; Kodama, I. Remodeling of gap junctional coupling in hypertrophied right ventricles of rats with monocrotaline-induced pulmonary hypertension. *Circ. Res.* **2000**, *86*, 871–878. [[CrossRef](#)] [[PubMed](#)]
123. Lu, Y.Y.; Lin, F.J.; Chen, Y.C.; Kao, Y.H.; Higa, S.; Chen, S.A.; Chen, Y.J. Role of Endothelin-1 in Right Atrial Arrhythmogenesis in Rabbits with Monocrotaline-Induced Pulmonary Arterial Hypertension. *Int. J. Mol. Sci.* **2022**, *23*, 10993. [[CrossRef](#)]
124. García-Álvarez, A.; Blanco, I.; García-Lunar, I.; Jordà, P.; Rodriguez-Arias, J.J.; Fernández-Friera, L.; Zegri, I.; Nuche, J.; Gomez-Bueno, M.; Prat, S.; et al. β 3 adrenergic agonist treatment in chronic pulmonary hypertension associated with heart failure (SPHERE-HF): A double blind, placebo-controlled, randomized clinical trial. *Eur. J. Heart Fail.* **2022**, *27*, 9400. [[CrossRef](#)]
125. Hardziyenka, M.; Campian, M.E.; Verkerk, A.O.; Surie, S.; Van Ginneken, A.C.G.; Hakim, S.; Linnenbank, A.C.; De Bruin-Bon, H.A.C.M.R.; Beekman, L.; Van Der Plas, M.N.; et al. Electrophysiologic remodeling of the left ventricle in pressure overload-induced right ventricular failure. *J. Am. Coll. Cardiol.* **2012**, *59*, 2193–2202. [[CrossRef](#)] [[PubMed](#)]
126. Hiram, R.; Naud, P.; Xiong, F.; Al-u'datt, D.; Algalarroondo, V.; Sirois, M.G.; Tanguay, J.F.; Tardif, J.C.; Nattel, S. Right Atrial Mechanisms of Atrial Fibrillation in a Rat Model of Right Heart Disease. *J. Am. Coll. Cardiol.* **2019**, *74*, 1332–1347. [[CrossRef](#)]
127. Hemnes, A.R.; Rathinasabapathy, A.; Austin, E.A.; Brittain, E.L.; Carrier, E.J.; Chen, X.; Fessel, J.P.; Fike, C.D.; Fong, P.; Fortune, N.; et al. A potential therapeutic role for angiotensin-converting enzyme 2 in human pulmonary arterial hypertension. *Eur. Respir. J.* **2018**, *51*, 1702638. [[CrossRef](#)]
128. Lee, F.Y.; Lu, H.I.; Zhen, Y.Y.; Leu, S.; Chen, Y.L.; Tsai, T.H.; Chung, S.Y.; Chua, S.; Sheu, J.J.; Hsu, S.Y.; et al. Benefit of combined therapy with nicorandil and colchicine in preventing monocrotaline-induced rat pulmonary arterial hypertension. *Eur. J. Pharm. Sci.* **2013**, *50*, 372–384. [[CrossRef](#)]
129. Sun, C.K.; Yuen, C.M.; Kao, Y.H.; Chang, L.T.; Chua, S.; Sheu, J.J.; Yen, C.H.; Ko, S.F.; Yip, H.K. Propylthiouracil attenuates monocrotaline-induced pulmonary arterial hypertension in rats. *Circ. J.* **2009**, *73*, 1722–1730. [[CrossRef](#)]
130. Tan, X.Y.; He, J.G. The remodeling of connexin in the hypertrophied right ventricular in pulmonary arterial hypertension and the effect of a dual ET receptor antagonist (bosentan). *Pathol. Res. Pract.* **2009**, *205*, 473–482. [[CrossRef](#)]
131. Tanaka, Y.; Takase, B.; Yao, T.; Ishihara, M. Right Ventricular Electrical Remodeling and Arrhythmogenic Substrate in Rat Pulmonary Hypertension. *Am. J. Respir. Cell Mol. Biol.* **2013**, *49*, 426–436. [[CrossRef](#)] [[PubMed](#)]
132. Yen, C.-H.; Tsai, T.-H.; Leu, S.; Chen, Y.-L.; Chang, L.-T.; Chai, H.-T.; Chung, S.-Y.; Chua, S.; Tsai, C.-Y.; Chang, H.-W.; et al. Sildenafil improves long-term effect of endothelial progenitor cell-based treatment for monocrotaline-induced rat pulmonary arterial hypertension. *Cytotherapy* **2013**, *15*, 209–223. [[CrossRef](#)] [[PubMed](#)]
133. Parati, G.; Goncalves, A.; Soergel, D.; Bruno, R.M.; Caiani, E.G.; Gerdts, E.; Mahfoud, F.; Mantovani, L.; McManus, R.J.; Santalucia, P.; et al. New Perspectives for Hypertension Management: Progress in Methodological and Technological Developments. *Eur. J. Prev. Cardiol.* **2022**, *30*, 48–60. [[CrossRef](#)]

134. Maniero, C.; Lopuszko, A.; Papalois, K.B.; Gupta, A.; Kapil, V.; Khanji, M.Y. Non-pharmacological factors for hypertension management: A systematic review of international guidelines. *Eur. J. Prev. Cardiol.* **2022**, *30*, 17–33. [[CrossRef](#)]
135. Pagonasa, N.; Saskoa, B.; Rittera, O. Managing hypertension in the future: A multifactorial approach. *Eur. J. Prev. Cardiol.* **2022**, *30*, 46–47. [[CrossRef](#)] [[PubMed](#)]

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