

SUPPLEMENTAL MATERIALS

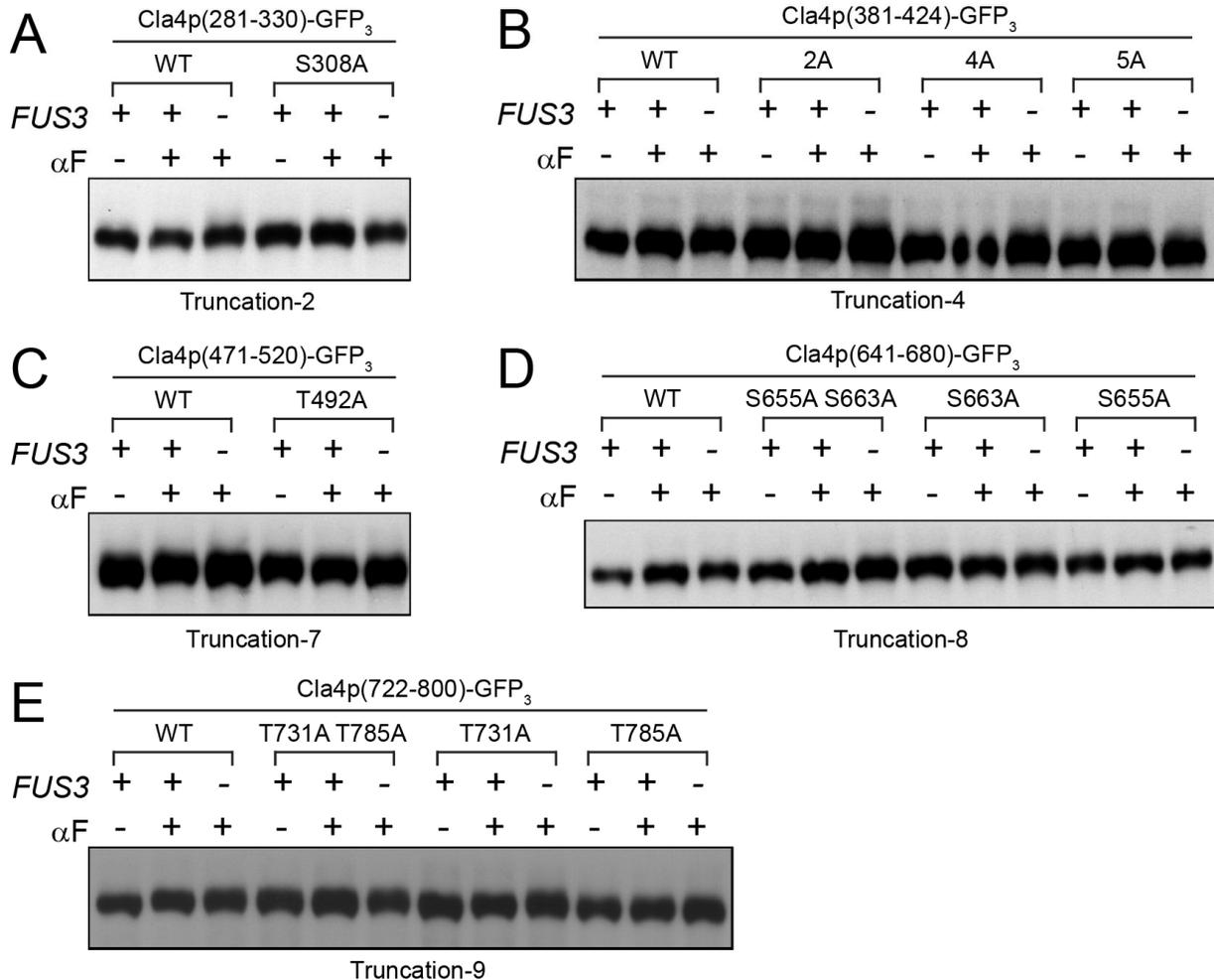


Figure S1. Screening for mating specific and Fus3p-dependent phosphorylation sites of Cla4p. Indicated fragments of Cla4p were expressed in *FUS3* (MY11198) or *fus3Δ cln3Δ* (MY10273) cells under the *GAL1* promoter for 90 min during mitosis or after α -factor arrest for 2 h as described in Fig. 2, C to F. Samples were run on 50 μ M Phos-tag gels and detected by anti-GFP immunoblotting. (A) Indicated Cla4p fragments (MR6412 and 6413) were expressed. (B) Indicated Cla4p fragments (MR6428, 6429, 6430 and 6431) were expressed. (C) Indicated Cla4p fragments (MR6424 and 6425) were expressed. (D) Indicated Cla4p fragments (MR6416, 6417, 6418 and 6419) were expressed. (E) Indicated Cla4p fragments (MR6420, 6421, 6422 and 6423) were expressed.

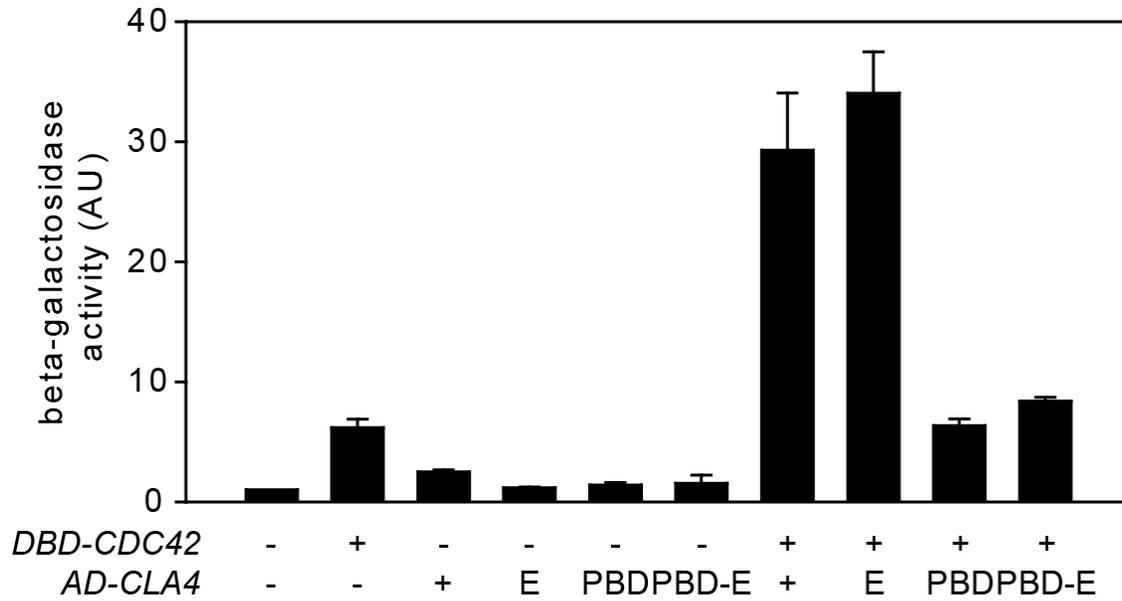


Figure S2. Interaction between Cla4p mutants and Cdc42p by yeast two-hybrid. The effect of Cla4-S186 phosphorylation on the interaction with Cdc42p was analyzed by yeast two-hybrid. The plasmids used in yeast two-hybrid are listed in Table S2. Cdc42p was fused to the DNA-binding domain (DBD) in pEG202. All Cdc42 hybrid proteins carry the C188S mutation to prevent localization to the plasma membrane. Cla4p was fused to the activation domain (AD) in pJG4-5. Full-length wild-type Cla4p, Cla4p-S186E, Cla4p-PBD domain (184-242), and Cla4p-PBD-S186E were fused to the activation domain (AD) in pJG4-5. The reporter strain EGY48-p1840 (MY15371) was co-transformed with pEG202 and pJG4-5 constructs. Transformants grown to mid-log in 2% raffinose were transferred to media with 2% galactose for 4h and then β -galactosidase activity was measured. All assays were performed in triplicate on three independent transformants for each combination of plasmids and plotted, mean \pm SD. E, full-length Cla4p-S186E. PBD, Cla4p-PBD domain. PBD-E, Cla4p-PBD-S186E.

Supplemental Table S1. Yeast strains used in this study.

Strain	Genotype	Source*
BY4741 derivatives		
BY4741	<i>MATa ura3-Δ0 his3-Δ200 leu2-Δ0 met15-Δ0</i>	
MY10273	<i>fus2Δ::HIS3 fus3Δ::natMX cln3Δ::LEU2 CDC3-mCherry::kanMX</i>	
MY11198	<i>cdc28-as1</i>	
MY12282	<i>fus2Δ::HIS3 fus3Δ::natMX cln3Δ::LEU2 CDC3-mCherry::kanMX</i>	
MR6152	<i>[P_{GALI}-fus2(54-83)-GFP₃]</i>	
MY12292	<i>fus2Δ::HIS3 fus3Δ::natMX cln3Δ::LEU2 CDC3-mCherry::kanMX</i>	
MR6289	<i>[P_{GALI}-cla4(315-364)-GFP₃]</i>	
MY12300	<i>fus2Δ::HIS3 CDC3-mCherry::kanMX MR6152 [P_{GALI}-fus2(54-83)-GFP₃]</i>	
MY12553	<i>fus2Δ::HIS3 CDC3-mCherry::kanMX MR6153 [P_{GALI}-fus2(54-83)-S67A-GFP₃]</i>	
MY13001	<i>cla4Δ::natMX MR6194 [P_{GALI}-myc-CLAA]</i>	
MY13003	<i>rim11Δ::kanMX MR6289 [P_{GALI}-cla4(315-364)-GFP₃]</i>	
MY13004	<i>kss1Δ::kanMX MR6289 [P_{GALI}-cla4(315-364)-GFP₃]</i>	
MY13005	<i>smk1Δ::kanMX MR6289 [P_{GALI}-cla4(315-364)-GFP₃]</i>	
MY13006	<i>pcl1Δ::kanMX MR6289 [P_{GALI}-cla4(315-364)-GFP₃]</i>	
MY13007	<i>pcl2Δ::kanMX MR6289 [P_{GALI}-cla4(315-364)-GFP₃]</i>	
MY13010	<i>hog1Δ::kanMX MR6289 [P_{GALI}-cla4(315-364)-GFP₃]</i>	
MY13011	<i>slt2Δ::kanMX MR6289 [P_{GALI}-cla4(315-364)-GFP₃]</i>	
MY13012	<i>yak1Δ::kanMX MR6289 [P_{GALI}-cla4(315-364)-GFP₃]</i>	
MY13013	<i>ykl161cΔ::kanMX MR6289 [P_{GALI}-cla4(315-364)-GFP₃]</i>	
MY13018	<i>rim11Δ::kanMX MR6291 [P_{GALI}-cla4(433-482)- GFP₃]</i>	
MY13019	<i>kss1Δ::kanMX MR6291 [P_{GALI}-cla4(433-482)- GFP₃]</i>	
MY13020	<i>smk1Δ::kanMX MR6291 [P_{GALI}-cla4(433-482)- GFP₃]</i>	
MY13021	<i>pcl1Δ::kanMX MR6291 [P_{GALI}-cla4(433-482)- GFP₃]</i>	
MY13022	<i>pcl2Δ::kanMX MR6291 [P_{GALI}-cla4(433-482)- GFP₃]</i>	
MY13025	<i>hog1Δ::kanMX MR6291 [P_{GALI}-cla4(433-482)- GFP₃]</i>	
MY13026	<i>slt2Δ::kanMX MR6291 [P_{GALI}-cla4(433-482)- GFP₃]</i>	
MY13027	<i>yak1Δ::kanMX MR6291 [P_{GALI}-cla4(433-482)- GFP₃]</i>	
MY13028	<i>ykl161cΔ::kanMX MR6291 [P_{GALI}-cla4(433-482)- GFP₃]</i>	
MY15123	<i>fus2Δ::kanMX fus3Δ::hphMX rvs161Δ::natMX</i> <i>MR6962 [fus2-S67A-GFP] MR4938 [FLAG-fus3-as1] MR6955[RVS161-mCherry]</i>	
MY15137	<i>fus2Δ::kanMX fus3Δ::hphMX cla4Δ::natMX</i>	
MY15305	<i>fus2Δ::kanMX fus3Δ::hphMX cla4Δ::natMX</i> <i>MR6998 [FUS2::GFP₁₀₄] MR4938 [FLAG-fus3-as1]</i>	
MY15308	<i>fus2Δ::kanMX fus3Δ::hphMX cla4Δ::natMX</i> <i>MR6998 [FUS2::GFP₁₀₄] MR4937 [FLAG-FUS3] MR6266[CLA4-HA₃]</i>	
MY15309	<i>fus2Δ::kanMX fus3Δ::hphMX cla4Δ::natMX</i> <i>MR6998 [FUS2::GFP₁₀₄] MR4937 [FLAG-FUS3] MR6275[cla4-KD-HA₃]</i>	
MY15310	<i>fus2Δ::kanMX fus3Δ::hphMX cla4Δ::natMX</i> <i>MR6998 [FUS2::GFP₁₀₄] MR1868 [pRS416] MR6266[CLA4-HA₃]</i>	

MY15311 *fus2Δ::kanMX fus3Δ::hphMX cla4Δ::natMX*
 MR6998 [*FUS2::GFP₁₀₄*] MR4937 [*FLAG-FUS3*] MR1867[*pRS415*]
 MY15342 *fus2Δ::kanMX fus3Δ::hphMX cla4Δ::natMX*
 MR6998 [*FUS2::GFP₁₀₄*] MR4938 [*FLAG-fus3-as1*] MR6266 [*CLA4-HA₃*]
 MY15343 *fus2Δ::kanMX fus3Δ::hphMX cla4Δ::natMX*
 MR6998 [*FUS2::GFP₁₀₄*] MR4938 [*FLAG-fus3-as1*] MR6275 [*cla4-KD-HA₃*]
 MY15344 *fus2Δ::kanMX fus3Δ::hphMX cla4Δ::natMX*
 MR6998 [*FUS2::GFP₁₀₄*] MR4938 [*FLAG-fus3-as1*] MR6343[*cla4-S186A-HA₃*]
 MY15345 *fus2Δ::kanMX fus3Δ::hphMX cla4Δ::natMX*
 MR6998 [*FUS2::GFP₁₀₄*] MR4938 [*FLAG-fus3-as1*] MR6991 [*cla4-S186E-HA₃*]
 MY15345 *fus2Δ::kanMX fus3Δ::hphMX cla4Δ::natMX*
 MR6998 [*FUS2::GFP₁₀₄*] MR4938 [*FLAG-fus3-as1*] MR6991[*cla4-S186E-HA₃*]
 MY15349 *fus2Δ::kanMX fus3Δ::hphMX cla4Δ::natMX*
 MR6998 [*FUS2::GFP₁₀₄*] MR4938 [*FLAG-fus3-as1*] MR6992 [*cla4-S425E-HA₃*]
 MY15351 *fus2Δ::kanMX fus3Δ::hphMX cla4Δ::natMX*
 MR6998 [*FUS2::GFP₁₀₄*] MR4938 [*FLAG-fus3-as1*] MR6994 [*cla4-EE-HA₃*]
 MY15355 *cdc28-as1 fus2Δ::HIS3 cla4Δ::kanMX*
 MR6160 [*P_{GALI}-fus2(54-99)-GFP₃*] MR6266 [*CLA4-HA₃*]
 MY15356 *cdc28-as1 fus2Δ::HIS3 cla4Δ::kanMX*
 MR6160 [*P_{GALI}-fus2(54-99)-GFP₃*] MR6991 [*cla4-S186E- HA₃*]
 MY15363 *cdc28-as1 fus2Δ::HIS3 cla4Δ::kanMX*
 MR6009 [*P_{GALI}-fus2(54-104)-GFP₃*] MR6266 [*CLA4-HA₃*]
 MY15364 *cdc28-as1 fus2Δ::HIS3 cla4Δ::kanMX*
 MR6166 [*P_{GALI}-fus2(54-104)-S67A-GFP₃*] MR6266 [*CLA4- HA₃*]
 MY15365 *cdc28-as1 fus2Δ::HIS3 cla4Δ::kanMX*
 MR6009 [*P_{GALI}-fus2(54-104)-GFP₃*] MR6991 [*cla4-S186E- HA₃*]
 MY15366 *cdc28-as1 fus2Δ::HIS3 cla4Δ::kanMX*
 MR6009 [*P_{GALI}-fus2(54-104)-GFP₃*] MR6275 [*cla4-KD- HA₃*]
 MY15647 *fus2Δ::kanMX fus3Δ::hphMX cla4Δ::natMX*
 MR6998 [*FUS2::GFP₁₀₄*] MR4938 [*FLAG-fus3-as1*] MR7080 [*cla4-AA- HA₃*]
 MY15951 *cla4Δ::kanMX* MR6343 [*cla4-S186A-HA₃*] MR5142 [*CDC3-GFP*]
 MY15953 *cla4Δ::kanMX* MR6266 [*CLA-HA₃*] MR5142 [*CDC3-GFP*]

Yeast two-hybrid reporter strain

MY15371 EGY48-p1840 *MATα ura3 his3 trp1 leu2 lexAop-lacZ-URA3*

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*All of cells without source were constructed in this study.

Supplemental Table S2: Plasmids used in this study

Plasmid	Description	Source*
pMR1867	<i>pRS415 CEN LEU2</i>	1
pMR1868	<i>pRS416 CEN URA3</i>	1
pMR4937	<i>CEN URA3 FLAG-FUS3</i>	2
pMR4938	<i>CEN URA3 FLAG-fus3-Q93G (fus3-as1)</i>	2
pMR5142	<i>CEN URA3 CDC3-GFP</i>	
pMR6009	<i>CEN URA3 P_{GAL}-fus2(54-104)-GFP₃</i>	3
pMR6152	<i>CEN URA3 P_{GAL}-fus2(54-83)-GFP₃</i>	
pMR6153	<i>CEN URA3 P_{GAL}-fus2(54-83)-S67A-GFP₃</i>	
pMR6160	<i>CEN URA3 P_{GAL}-fus2(54-99)-S84E-GFP₃</i>	
pMR6166	<i>CEN URA3 P_{GAL}-fus2(54-104)-S67A-GFP₃</i>	
pMR6194	<i>CEN URA3 P_{GAL}-myc-CLA4</i>	4
pMR6266	<i>CEN LEU2 CLA4-HA₃</i>	
pMR6275	<i>CEN LEU2 cla4-K594A (cla4-KD)-HA₃</i>	
pMR6343	<i>CEN LEU2 cla4-S186A-HA₃</i>	
pMR6955	<i>CEN HIS3 RVS161-mCherry</i>	
pMR6962	<i>CEN LEU2 fus2-S67A::GFP₁₀₄</i>	
pMR6991	<i>CEN LEU2 cla4-S186E-HA₃</i>	
pMR6992	<i>CEN LEU2 cla4-S425E-HA₃</i>	
pMR6994	<i>CEN LEU2 cla4-EE (S186E S425E)-HA₃</i>	
pMR6998	<i>CEN HIS3 FUS2::GFP₁₀₄</i>	
pMR7080	<i>CEN LEU2 cla4-AA (S186A S425A)-HA₃</i>	
Screening for pheromone and Fus3p-specific phosphorylation sites in Cla4p		# of Truncation
pMR6415	<i>CEN URA3 P_{GAL}-cla4(161-210)-GFP₃</i>	Truncation-1
pMR6414	<i>CEN URA3 P_{GAL}-cla4(161-210)-S186A-GFP₃</i>	
pMR6413	<i>CEN URA3 P_{GAL}-cla4(281-330)-GFP₃</i>	Truncation-2
pMR6412	<i>CEN URA3 P_{GAL}-cla4(281-330)-S308A-GFP₃</i>	
pMR6289	<i>CEN URA3 P_{GAL}-cla4(315-364)-GFP₃</i>	Truncation-3
pMR6290	<i>CEN URA3 P_{GAL}-cla4(315-364)-S351A-GFP₃</i>	
pMR6429	<i>CEN URA3 P_{GAL}-cla4(381-424)-GFP₃</i>	Truncation-4
pMR6230	<i>CEN URA3 P_{GAL}-cla4(381-424)-2A(S396A S398A)-GFP₃</i>	
pMR6428	<i>CEN URA3 P_{GAL}-cla4(381-424)-4A(S396A S398A S400A S402A)-GFP₃</i>	
pMR6431	<i>CEN URA3 P_{GAL}-cla4(381-424)-5A(S396A S398A S400A S402A S404A)-GFP₃</i>	
pMR6427	<i>CEN URA3 P_{GAL}-cla4(406-460)-S445A-GFP₃</i>	Truncation-5

pMR6426	<i>CEN URA3 P_{GAL}-cla4(406-460)-S425A S445A-GFP₃</i>	
pMR6291	<i>CEN URA3 P_{GAL}-cla4(433-482)-GFP₃</i>	Truncation-6
pMR6292	<i>CEN URA3 P_{GAL}-cla4(433-482)-S445A-GFP₃</i>	
pMR6425	<i>CEN URA3 P_{GAL}-cla4(471-520)-GFP₃</i>	Truncation-7
pMR6424	<i>CEN URA3 P_{GAL}-cla4(471-520)-T492A-GFP₃</i>	
pMR6419	<i>CEN URA3 P_{GAL}-cla4(641-680)-GFP₃</i>	Truncation-8
pMR6417	<i>CEN URA3 P_{GAL}-cla4(641-680)-S663A-GFP₃</i>	
pMR6416	<i>CEN URA3 P_{GAL}-cla4(641-680)-S655A-GFP₃</i>	
pMR6418	<i>CEN URA3 P_{GAL}-cla4(641-680)-S663A S655A-GFP₃</i>	
pMR6423	<i>CEN URA3 P_{GAL}-cla4(722-800)-GFP₃</i>	Truncation-9
pMR6420	<i>CEN URA3 P_{GAL}-cla4(722-800)-T731A-GFP₃</i>	
pMR6421	<i>CEN URA3 P_{GAL}-cla4(722-800)-T785A-GFP₃</i>	
pMR6422	<i>CEN URA3 P_{GAL}-cla4(722-800)-T731A T785A-GFP₃</i>	
Yeast two-hybrid plasmids		Source*
pMR6995	<i>pEG202 for construction of LexA fusion protein, HIS3, 2μm</i>	5
pMR6996	<i>pJG4-5 for construction of B42AD fusion proteins, TRP1, 2μm</i>	5
pMR7002	<i>B42AD-CLA4 in pJG4-5</i>	
pMR7004	<i>B42AD-cla4(S186E) in pJG4-5</i>	
pMR7012	<i>LexA-cla4(C188S) in pEG202</i>	5
pMR7016	<i>B42AD-cla4-PBD (184-242) in pJG4-5</i>	
pMR7018	<i>B42AD-cla4-PBD-S186E (184-242) in pJG4-5</i>	

*All plasmids without a source indicated were constructed in this study.

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4. Versele M, Thorner J (2004) Septin collar formation in budding yeast requires GTP binding and direct phosphorylation by the PAK, Cla4. *J Cell Biol* 164(5): 701-715.
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