



Callyspongia spp.: Secondary Metabolites, Pharmacological Activities, and Mechanisms

Yuni Elsa Hadisaputri ^{1,*}^(D), Annida Adha Nurhaniefah ¹, Sendi Sukmara ¹^(D), Ade Zuhrotun ¹, Rini Hendriani ² and Iyan Sopyan ³^(D)

- ¹ Department of Pharmaceutical Biology, Faculty of Pharmacy, Universitas Padjadjaran, Jatinangor 45363, Indonesia
- ² Department of Pharmacology and Clinical Pharmacy, Faculty of Pharmacy, Universitas Padjadjaran, Jatinangor 45363, Indonesia
- ³ Departement of Pharmaceutics and Technology of Pharmacy, Faculty of Pharmacy, Universitas Padjadjaran, Jatinangor 45363, Indonesia
- * Correspondence: yuni.elsa@unpad.ac.id; Tel.: +62-22-842-88888

Abstract: One of the most widespread biotas in the sea is the sponge. *Callyspongia* is a sponge genus found in the seas, making it easily available. In this review, the pharmacological activity and mechanism of action of the secondary metabolites of *Callyspongia* spp. are addressed, which may lead to the development of new drugs and targeted therapeutic approaches. Several scientific databases, such as Google Scholar, PubMed, ResearchGate, Science Direct, Springer Link, and Wiley Online Library, were mined to obtain relevant information. In the 41 articles reviewed, *Callyspongia* spp. was reported to possess pharmacological activities such as cytotoxicity against cancer cell lines (36%), antifungal (10%), anti-inflammatory (10%), immunomodulatory (10%), antidiabetic and antiobesity (6%), antimicrobial (8%), antioxidant (4%), antineurodegenerative (4%), antihypercholesterolemic (2%), antihypertensive (2%), antiparasitic (2%), antiallergic (2%), antiviral (2%), antiosteoporotic (2%), and antituberculosis (2%) activities. Of these, the antioxidant, antituberculosis, and anti-inflammatory activities, particularly cytotoxicity, show promise, and the compounds responsible may be developed into new drugs.

Keywords: Callyspongia spp.; secondary metabolite; pharmacological activity; mechanism; cytotoxic

1. Introduction

The ocean, which covers 71% of the earth's surface, regulates our climate and contains abundant resources [1]. The sea encompasses a large area, but it is well connected, and the temperature is less extreme compared with that on land. Although containing more biodiversity, only 16% of all species have been identified [2].

One of the most ubiquitous sea organisms is the sponge. Sponges are often abundant in shallow water habitats, making them a unique biodiversity component [3]. They are one of the most diverse sessile organisms, with approximately 8876 valid species identified worldwide. Each has its unique characteristics, while some features are shared [4].

Callyspongia belongs to the family Callyspongidae. More than 60 species are widely distributed in the tropical sea [5]. It is also found in the Indian, Western Atlantic, and Eastern Pacific oceans, including Indonesia [6], the Red sea [7,8], Cuba [4], Barbados [9], Brazil [10,11], and Ecuador [12]. At a depth of 6–10 m below sea level, *Callyspongia* spp. can live under coral reefs, ranging from moderate to damaged conditions, or in habitats dominated by hard coral, sand, and coral rubble [13].

Sponges from the *Callyspongia* genus are formed from primary, secondary, and tertiary spongin fibers [4]. *Callyspongia* sponges are encrusting, form a single erect branch or a mass of round branches, and many are bifurcated. The longest branch that has been observed is



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Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). approximately 40 cm. The branches are approximately 100–150 mm in diameter and have oscula or excretory organs that are slightly elevated, numerous, scattered throughout, and 0.5–2 mm in diameter. When pressed or cut, *Callyspongia* spp. secrete mucus. They have a smooth surface [14]. Skeletal fractions, such as spicules and cell debris, constitute 69.8% of the biomass of *Callyspongia* spp., the spongy cells (choanosomes) comprise 18.8%, and bacterial pellets account for 11.3%. The skeleton fraction dominates the biomass, resulting in the stiffer morphology of *Callyspongia* sponges [13].

The morphology of the callyspongia species varies. For example, *Callyspongia (Cladochalina) aerizusa* (Figure 1e) and *Callyspongia (Siphonochalina) siphonella* (Figure 1a) have a tubular and clustered form, but with different colors, tubes, and oscula size. *Callyspongia aerizusa* has a green–orange color, whereas *Callyspongia siphonella* has pale lavender color. There are also species with varying forms, such as *Callyspongia (Cladochalina) diffusa* (Figure 1b), which has a dull champagne pink or purplish-pink color, long or short thick cylindrical branches that vary from fanlike to upright or irregular [15]. Other species have unlikely forms, *Callyspongia samarensis* is called a spaghetti sponge because of its forms (Figure 1d) [16]. The color of *Callyspongia* spp. also varies with bright colors, such as *Callyspongia truncate* (Figure 1c), or deep colors like *Callyspongia aerizusa*.

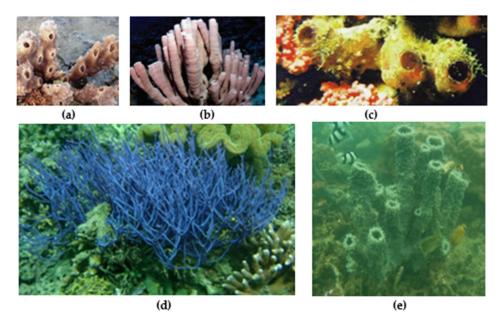


Figure 1. (a) Callyspongia diffusa, (b) Callyspongia siphonella, (c) Callyspongia truncata [15], (d) Callyspongia samarensis [3], (e) Callyspongia aerizusa.

Sponges are a potential repository of new drugs. There are several drugs originating from sponges that have entered clinical trials and approved, including cytarabine (Ara-C) for cancer treatment and vidarabine as an antiviral [17]. In addition, Eribulin mesylate (E7389) is an anticancer drug that is undergoing clinical phase 3 testing [18,19] Gemcitabine (GEM) (Gemzar) is an anticancer agent which has entered clinical phase 2 [20], whereas IPL576,092 (contignasterol derivative) is an anti-inflammatory compound that has entered clinical phase 2 testing [21]. PM-10450 (Zalypsis[®]) [22], discodermolide, HT1286 (hemi-asterlin derivative), LAF389 (bengamide B derivative), hemiasterlin (E7974), KRN7000 (agelasphin derivative) [23], PM-060184 [24], and NVP-LAQ824 (psammaplin derivative) have entered clinical phase 1 trials as anticancer drugs [25].

Other pharmacological activities of sponge compounds include antibacterial, antihyperlipidemic, antiproliferative, immunomodulatory, and anti-inflammatory effects have been reported, including *Callyspongia* spp. [26,27]. Sponges contain multiple primary and secondary metabolites, such as fatty acids, alkaloids, steroids, nucleotides, peptides, polyacetylenes, and terpenoids. A total of 212 compounds have been isolated from *Callyspongia* spp. and their structures and bioactivities have been presented [28]. This review summarizes the potential pharmacological activities exhibited by *Callyspongia* spp. compounds that may be developed into new drugs. We also discuss the related mechanisms that may contribute to targeted therapy.

2. Materials and Methods

The literature review of *Callyspongia* spp. was based on topics related to pharmacological activity and the mechanism of action of secondary metabolites contained therein. This review was conducted with a qualitative and quantitative approach to obtain information from several scientific databases, including Google Scholar, PubMed, ResearchGate, Science Direct, Springer Link, and the Wiley Online Library. Several keywords, such as *"Callyspongia* sp.", "metabolites", and "pharmacology activity", were used to procure relevant resources. The inclusion criterion for selecting articles was that they should describe the isolation and functional studies of secondary metabolites from *Callyspongia* sponges. Articles describing the isolation and activities of fungi or bacteria in *Callyspongia* species were excluded. The abstracts were carefully read to identify and select relevant articles. From 72 identified articles after screening information sources, 41 published between 1980 and 2021 were selected and reviewed (Figure 2).

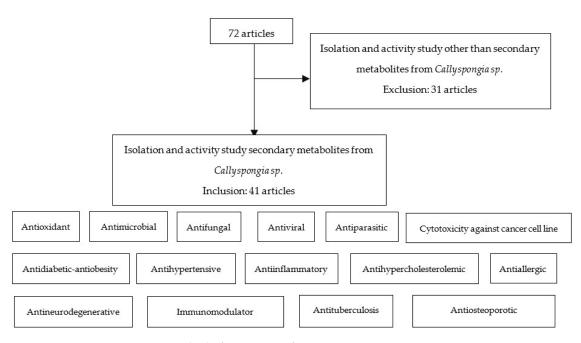


Figure 2. Method of screening information sources.

3. Results

Sixteen pharmacological activities have been reported for *Callyspongia* spp. These activities along with their descriptions are listed in Table 1.

Pharmacological Activity	Callyspongia spp.	Secondary Metabolite	Description of Activity	Ref.
	Callyspongia truncata	Callyspongynic acid	IC ₅₀ against α-glucosidase: 0.25 μg/mL	[29,30]
Antidiabetic and antiobesity	Callyspongia samarensis	-	EC ₅₀ 14.47 μg/mL (AMPK Activation)	[31]
	Callyspongia sp.	β-Sitosterol	Activation of GLUT-4 and insulin receptors	[32]

Pharmacological Activity	Callyspongia spp.	Secondary Metabolite	Description of Activity	Ref.
Antihypercholesterolemic	<i>Callyspongia</i> sp.	Callyspongiamide A	IC_{50} against SOAT-1 and SOAT-2: 0.78 ± 0.19 and $2.8\pm0.72~\mu M$	[5]
		Callyspongiamide B	IC_{50} against SOAT-1 and SOAT-2: 1.2 ± 0.31 and $2.4\pm0.96~\mu M$	
		Disamide A	IC_{50} against SOAT-1 and SOAT-2: 5.2 ± 0.93 and $4.2\pm0.76~\mu M$	
Antihypertensive	Callyspongia diffusa	Callypyrone A	IC ₅₀ against Angiotensin I-converting enzyme (ACE): 0.48 mM	[33]
		Callypyrone B	IC ₅₀ against ACE: 0.57 mM	
	Callyspongia crassa	-	61.47% inhibition of protein denaturation	[34]
	Callyspongia sp.	-	97% inhibition of hemolysis (at a dose of 3200 ppm)	[35]
		Cyclo[L-Hyp-L-Ala]	Increase secretion of IL-10 (J774A.1 cells) by 1.65-fold	
	Callyspongia sp.	Cyclo[L-Pro-Gly]	Increase secretion of IL-10 (J774A.1 cells) 1.29-fold	[36]
Anti-inflammatory		Cyclo[L-Pro-Phe]	Increase secretion of IL-10 (J774A.1 cells) 1.54-fold	
Anti-inianiniatory		Cyclo[L-Pro-Ala]	Increased secretion of IL-10 (J774A. 1 cells) 1.56-fold	
	Callyspongia sp.	β-Sitosterol	 ED₅₀ 155.6 (mg/kg/ip) on adrenal pituitary axis 54% of inflammatory effect at dose 320 mg/kg (p.o.) 	[37]
	Callyspongia siphonella	Callysterol	19.5 ± 7.3 mL (Edema volume)	[38]
	Callyspongia aff. implexa	Gelliusterol E	IC_{50} against <i>Chlamydia</i> trachomatis: $2.34 \pm 0.22 \mu M$ (No inclusion at a concentration of $40 \mu M$)	[39]
		Callyaerin A	<i>Chlamydia albican</i> inhibition with a zone of inhibition of 25–30 mm	[6]
Antifungal	Callyspongia aerizusa	Callyaerin B	<i>Chlamydia albican</i> inhibition with a zone of inhibition of 15 mm	
		Callyaerin E	<i>Chlamydia albican</i> inhibition with a zone of inhibition of 20 mm	
	Callyspongia sp.	β-Sitosterol	Average inhibition diameter against <i>Fusarium</i> spp.: 10 mm	[39,40]
	Callyspongia sp.	(-)-Siphonodiol	MIC against <i>Trichophyton</i> asteroides: 25.0 µg/mL	[41]
	Callyspongia fibrosa	4-hydroxybenzoic acid	Antifungal against Ganoderma boninense	[42,43]

Pharmacological Activity	Callyspongia spp.	Secondary Metabolite	Description of Activity	Ref.
		-	 IC₅₀ against: Caco-2 cell line: 5.57 μg/mL MCF-7 cell line: 1.39 μg/mL 	[34]
		Neviotine-C	IC ₅₀ against: PC-3 cell line: $53.6 \pm 0.17 \mu M$ A549 cell line: 87.2 ± 1.34 μM MCF-7 cell line: 45.5 ± 0.06 μM	[44]
		Neviotine A	$IC_{50} \text{ against:}$ • PC-3 cell line: 71.2 \pm 0.34 μ M • A549 cell line: 76.3 \pm 0.35 μ M • MCF-7 cell line: 46.3 \pm 0.06 μ M	
		_	IC ₅₀ against: • MCF-7 cell line: 12.3 \pm 0.7 µg/mL • HepG-2 cell line: 11.8 \pm 1.2 µg/mL	[45]
Cytotoxicity against cancer cell lines	Callyspongia siphonella	Sipholenol-A	$\begin{array}{c} \text{IC}_{50} \text{ against:} \\ \bullet \text{PC-3 cell line:} \\ 7.9 \pm 0.12 \ \mu\text{M} \\ \bullet \text{A549 cell line:} \\ 8.9 \pm 0.01 \ \mu\text{M} \\ \bullet \text{MCF-7 cell line:} \\ 56.3 \pm 0.17 \ \mu\text{M} \end{array}$	[44]
			$IC_{50} \text{ against:}$ • MCF-7 cell line: 19.2 \pm 0.6 µg/mL • HepG-2 cell line: 9.6 \pm 0.8 µg/mL	[45]
		Sipholenone A	$IC_{50} \text{ against:}$ • PC-3 cell line: 53.9 \pm 0.25 μ M • A549 cell line: 24.8 \pm 0.22 μ M • MCF-7 cell line: 36.2 \pm 0.13 μ M	[44]
		_	IC ₅₀ against: • MCF-7 cell line: $3 \pm 0.4 \ \mu g/mL$ • HepG-2 cell line: $2.8 \pm 0.4 \ \mu g/mL$	[45]

Pharmacological Activity	Callyspongia spp.	Secondary Metabolite	Description of Activity	Ref.
		Sipholenol L	$IC_{50} \text{ against:}$ • MCF-7 cell line: $4.0 \pm 0.22 \ \mu\text{g/mL}$ • HepG-2 cell line: $18.7 \pm 0.9 \ \mu\text{g/mL}$	
	Callyspongia crassa	-	 IC₅₀ against: Caco-2 cell line: 13.05 μg/mL MCF-7 cell line: 9.47 μg/mL 	[34]
	<i>Callyspongia</i> sp.	Callyspongiolide	IC ₅₀ against: L5178Y cell line: 320 nM Jurkat J16 T cell line: 70 nM Ramos B lymphocyte cell line: 60 nM	[46]
	Callyspongia sp.	Callypeptide A	 GI₅₀ against: MDA-MB-231 cell line: 29 μM HT-29 cell line: 30 μM A549 cell line: 18.5 μM 	[47]
		Callyazepin	IC ₅₀ against: • K562 cell line: 7.4 μM • A549 cell line: 3.0 μM	[48]
	Callyspongia sp.	(3R)- methylazacyclodecane	 IC₅₀ against: K562 cell line: 3.2 μM A549 cell line: 3.8 μM 	[40]
	<i>Callyspongia</i> sp. (CMB-01152)	Hymenialdisine	IC ₅₀ against: • SW620 cell line: 3.1 μM • KB-3-1: 2.0 μM	[49]
	Callyspongia schulzei	-	IC ₅₀ against: • HT-29 cell line: $35.57 \pm 0.87 \ \mu\text{g/mL}$ • T47D cell line: $37.98 \pm 2.12 \ \mu\text{g/mL}$ • Casky tumor cell line: $63.20 \pm 0.76 \ \mu\text{g/mL}$	[50]
	Callyspongia	Callyaerin E	IC ₅₀ against L5178Y cell line: 0.39 μM	[6]
_	aerizusa	Callyaerin H	IC ₅₀ against L5178Y cell line: 0.48 μM	r.1
	Callyspongia truncata	Callystatin -	IC ₅₀ against KB cell line: 0.01 μg/mL Further research: IC ₅₀ against: • KB cell line: 10 pg/mL • L1210 cell line: 20 pg/ml	[51]

Pharmacological Activity	Callyspongia spp.	Secondary Metabolite	Description of Activity	Ref.
	Callyspongia sp.	(–)-(3R,18R) alcohol	IC ₅₀ against TR-LE cell line: 0.11 μM	[52]
		(+)-(3S,18S)	IC $_{50}$ against TR-LE cell line: 0.47 μM	[02]
	Callyspongia sp.	Siphonodiol	IC ₅₀ against HL-60 cell line: 6.5 μg/mL	
		Callyspongidiol	IC ₅₀ against HL-60 cell line: 2.8 μ g/mL	[53]
		14,15- dihydrosphonodiol	IC ₅₀ against HL-60 cell line: 6.5 μg/mL	
			IC ₅₀ against:	
		Callyspongenols A	 P388 cell line: 2.2 μg/mL HeLa cell line: 4.5 μg/mL 	
			IC ₅₀ against:	
	<i>Callyspongia</i> sp.	Callyspongenols B	 P388 cell line: 10 μg/mL HeLa cell line: 10 μg/ mL 	[54]
		Callyspongenols C	IC ₅₀ against:	
			 P388 cell line: 2.2 μg/mL HeLa cell line: 3.9 μg/mL 	
			IC ₅₀ against:	
		Callyspongenols D	 P388 cell line: 0.4 µg/mL HeLa cell line: 0.066 µg/mL 	
	Callyspongia fistularis	Callyspongamide A	IC_{50} against HeLa cell line: $$4.1\ \mu g/mL$$	[55]
	Calluaranaiaara	Alkupikanye E	IC ₅₀ : 5 μg/mL	
	Callyspongia sp.	Alkupikanye F	IC ₅₀ : 10 μg/mL	[56]
		-	IC ₅₀ : 2 μg/mL against NIH3T3 cells transfected with the human EGF receptor	
	Callyspongia sp.	8-Bromooctyl tert-butyldimethylsilyl ether (fraction, $n = 3$) 9-(3-Pyridyl)nonyl alcohol (fraction, $n = 3$)	IC ₅₀ : 1.3 μg/mL against NIH3T3 cells transfected with human EGF receptor gene	[57]
	Callyspongia sp.	Akaterpin	IC ₅₀ against PI-PLC: 0.5 μg/mL	[58]
		-	IC ₅₀ against:	
	Callyspongia aerizusa		 A549 cell line: 9.38 µg/mL TE-8 cell line: 3.12 µg/mL HEP G2 cell line: 10.62 µg/mL MIA PaCa-2 cell line: 10.72 µg/mL 	[59]

Pharmacological Activity

Secondary Metabolite	Description of Activity	Ref.
	LC ₅₀ against:	
-	 Staphylococcus aureus: 215.2 ± 32.9 μg/mL Bacillus subtilis: 18.2 ± 3.56 μg/mL 	[60]
Siphonocholin	MIC against <i>Pseudomonas</i> aeruginosa: 64 μg/mL	[61]
	Inhibition against:	
Sipholenol L	 Staphylococcus aureus (Zone of inhibition: 12.3 ± 0.72 mm) Bacillus subtitus (Zone of inhibition: 14.5 ± 0.72 mm) 	
	Inhibition against:	
	• <i>Staphylococcus aureus</i> (Zone of inhibition:	

Table 1. Cont.

Callyspongia spp.

Callyspongia crassa

		Siphonocholin	MIC against Pseudomonas aeruginosa: 64 μg/mL	[61]
		Sipholenol L	 Inhibition against: Staphylococcus aureus (Zone of inhibition: 	
	– Callyspongia siphonella	Sipholenoi L	• $Bacillus subtitus$ (Zone of inhibition: 14.5 \pm 0.72 mm)	
		Neviotine A	 Inhibition against: Staphylococcus aureus (Zone of inhibition: 14.1 ± 0.72 mm) Bacillus subtitus (Zone of inhibition: 17.2 ± 0.58 mm) Escherichia coli (Zone of inhibition 12.7 ± 0.58 mm) 	[45]
Antimicrobial		Sipholenone A	 Inhibition 12.7 ± 0.38 mm) Inhibition against: Staphylococcus aureus (Zone of inhibition: 8.2 ± 0.72 mm) Bacillus subtitus (Zone of inhibition: 2.4 ± 0.58 mm) Escherichia coli (Zone of 	
		Callyaerin A	 inhibition 5.4 ± 0.58 mm) Inhibition against: Escherichia coli (moderate) with zone of inhibition: 10–15 mm Staphylococcus aureus (mild) with zone of inhibition: 9 mm 	
	Callyspongia aerizusa	Callyaerin E	 Inhibition against: Bacillus subtilis (potent) with zone of inhibition: 15–17 mm Escherichia coli (mild) with zone of inhibition: 9–11 mm Staphylococcus aureus (mild) with zone of inhibition: 9–10 mm 	[6]
	Callyspongia crassa	-	Percentage of inhibition: 58.1% at 671 μg/mL	[34]
Antioxidant	Callyspongia aerizusa	-	Percentage of inhibition 56.6% at 0.5 μg/mL, 57.2% at 0.6 μg/mL, and 58.4% at 0.7 μg/mL	[62]

Pharmacological Activity	Callyspongia spp.	Secondary Metabolite	Description of Activity	Ref.
Antiparasitic	<i>Callyspongia</i> sp.	Isoakaterpine	IC ₅₀ against adenosine phosphoribosyltransferase of <i>Leishmania</i> spp: 1.05 μM	[11]
Antiallergic	Callyspongia sp.	3-(2-(4- hydroxyphenyl)-2- oxoethyl)-5,6- dihydropyridine- 2(1H)-one	IC_{50} againts RBL-2H3: $18.7\pm6.7~\mu M$	[63,64]
Antituberculosis	Callyspongia	Callyaerin A	MIC ₉₀ against Mycobacterium tuberculosis: 2 μM	[64]
Antituderculosis	aerizusa	Callyaerin B	MIC ₉₀ against Mycobacterium tuberculosis: 5 μM	
Antivinal	Callyspongia crassa	-	85.3% against hepatitis A virus (MIC 9.765 μg/mL)	[34]
Antiviral	Callyspongia siphonella	-	83.7% against hepatitis A virus (MIC 0.625 μg/mL)	
	Callyspongia sp.	Niphatoxin C	IC ₅₀ P2X ₇ antagonism: 11.5 μM	[65]
	Callyspongia sp.	Siphonodiol	 Increases IL-12p70 secretion in LPS-primed DCs Modulates dendritic cell function for T1 cell proliferation 	[66]
Immunomodulatory	<i>Callyspongia</i> sp.	Callyspongidiol	- Modulates dendritic cell function for T1 cell proliferation	[53]
		14,15- dihydrosphonodiol		
	<i>Callyspongia</i> sp.	β-Sitosterol	Modulates dendritic cell activity and increases peripheral blood mononuclear cell viability	[67]
	Callyspongia sp.	-	Increase levels of IFN- γ and TNF- α (Wistar strain mice) at extract doses of 300 mg/kg and 400 mg/kg	[68]
Antineurodegenerative	Callyspongia samarensis	-	IC ₅₀ against β-secretase: 99.82 μg/mL	[31]
		Hymenialdisine	IC ₅₀ against:	
	Callyspongia sp.		 GSK3β: 0.07 μM CK5.p25: 0.16 μM CK1δ: 0.03 μM 	[49]
	Callyspongia	Neviotine D	IC ₅₀ against RANKL: 12.8 μM	[69]
Antiosteoporotic	orotic siphonella	Neviotine A	IC ₅₀ against RANKL: 32.8 μM	

 Table 1. Cont.

 IC_{50} : Half maximal inhibitory concentration; EC_{50} : Half maximal effective concentration; ED_{50} : Median effective dose; MIC: Minimum inhibitory concentration; GI_{50} : Half maximal growth inhibition; LC_{50} : Median Lethal Concentration.

4. Discussion

We have discussed the pharmacological activities of *Callyspongia* spp. that have been previously reported.

4.1. Antidiabetic, Antihypercholesterolemic, and Antiobesity

The active compound from *Callyspongia truncata*, callyspongynic acid (Figure 3), shows higher antidiabetic activity by inhibiting α -glucosidase with an IC₅₀ of 0.25 µg/mL [29,30] compared with acarbose (IC₅₀ 1.3 µg/mL) [70]. Inhibiting this enzyme reduces caloric intake by attenuating appetite, suppressing hunger, and increasing satiety [71,72], thereby supporting weight loss [73] to a moderate level [74]. It is also one of the targets of diabetes therapy. Compared with α -amylase, inhibiting α -glucosidase can improve hyperglycemia, especially postprandial hyperglycemia, by decreasing glucose production (Figure 3) [75].

Compounds, such as callyspongiamide A and B as well as disamide A (Figure 3), exert antihypercholesterolemic activity, which can also lead to an antiobesity effect by inhibiting sterol O-acyltransferase (SOAT), the enzyme that catalyzes the formation of cholesteryl ester [76]. In addition, other sterols may be used as activators or substrates of this enzyme [77], which implicates it as a potential drug target [61] in hypercholesterolemia; however, the underlying mechanism remains unknown.

In a cell-based testing assay, the IC₅₀ values of callyspongiamide A against SOAT 1 and SOAT 2 were 0.78 \pm 0.19 and 2.8 \pm 0.72 μ M, those of callyspongiamide B were 1.2 \pm 0.31 and 2.4 \pm 0.96 μ M, whereas those of disamide A were 5.2 \pm 0.93 and 4.2 \pm 0.76 μ M. Although each compound markedly inhibited SOAT 2, as evidenced by the lower IC₅₀ compared with the control beauveruolide III (IC₅₀ > 20 μ M), only callyspongiamide A significantly inhibited SOAT 1 [5].

Callyspongia sp. also contains β -Sitosterol. This compound exhibits potent antidiabetic activity related to insulin receptor activation and increased glucose transporter 4 (GLUT-4) translocation to adipose tissue [78,79]. In addition, these compounds can potentially maintain glucose homeostasis through sensitization of insulin resistance by increasing the expression of peroxisome proliferator-activated receptor and GLUT-4 (Figure 4) [80]. Another study on HFD-fed and sucrose-induced type-2 diabetic rats indicates that β -Sitosterol enhances the glycemic regulation [32,79].

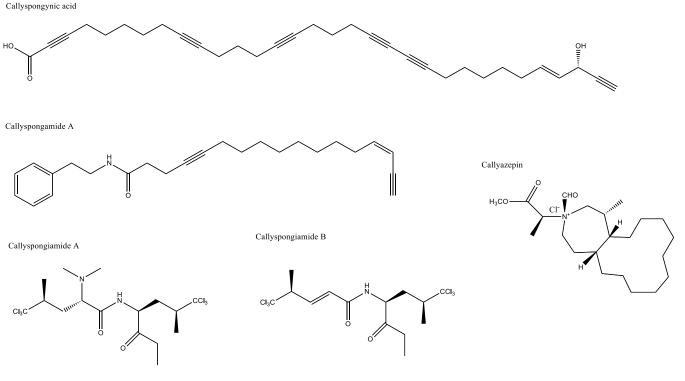
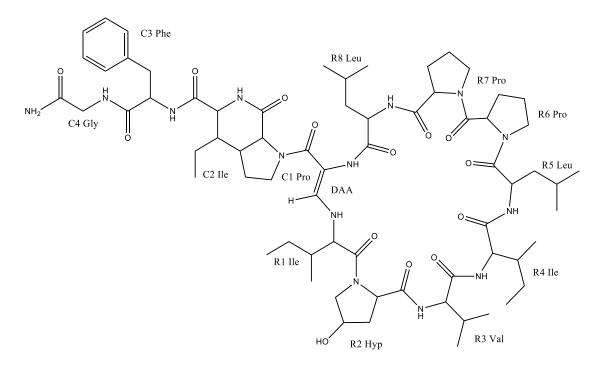


Figure 3. Cont.

Callyaerin A



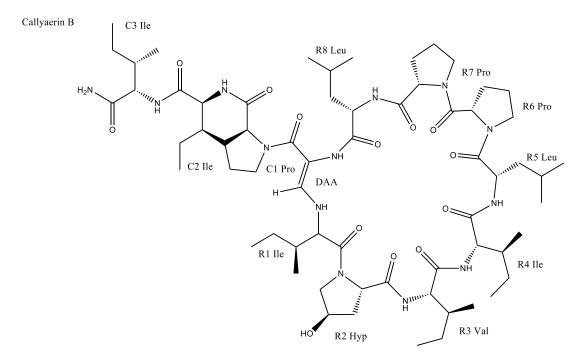
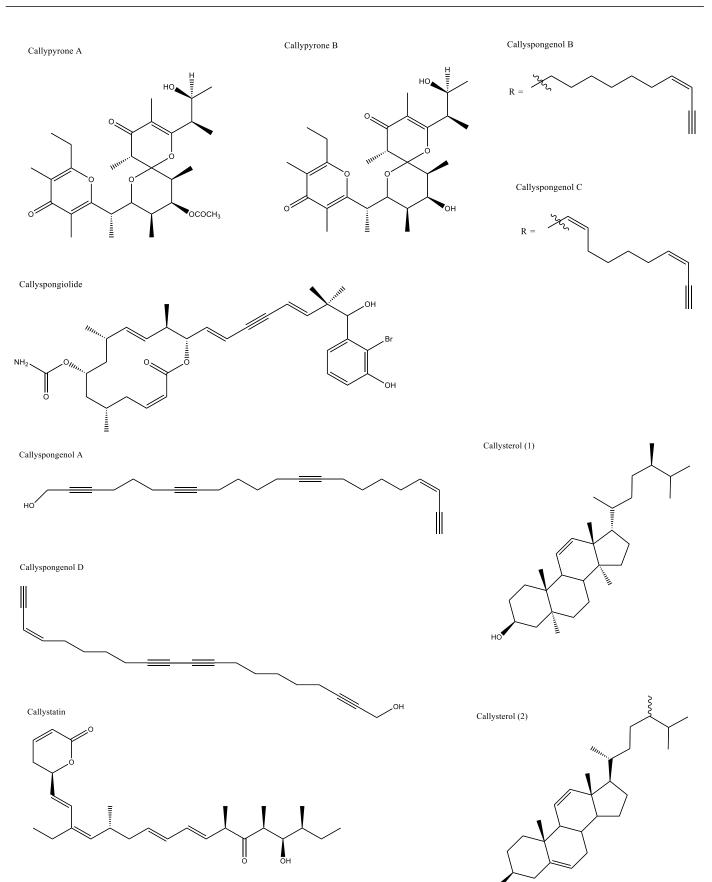


Figure 3. Cont.



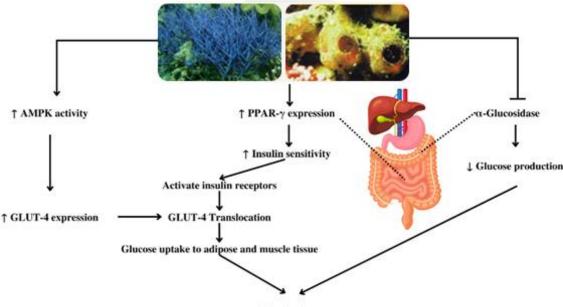
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Figure 3. Cont.

Callyaerin E

R6 Pro R7 Val C3 Ile R5 Pro 0 Н 0 NĤ N H 0= DAA NH₂ \\ \\ R4 Phe C4 Gly uШ C2 Ile ŅΗ H C1 Pro 0 ŃH 0 R1 Leu 'N H R3 Phe R2 Pro Callypeptide A R7 3-OHLeu R6 Ile OH. R5 Phe R8 Tyr 0‴ Callyaerin H R2 Ala 0 R6 Pro R7 Leu 0= R5 Pro R4 Tyr 0 R1 Thr он H_2 R3 Leu όн DAA R4 Phe C2 Ile H NΗ C1 Pro Н н, Callystatin R1 Val R3 Val R2 Pro Ē

Figure 3. Structure of secondary metabolites from the Callyspongia subgenus.



↓ Blood glucose

Figure 4. Antidiabetic mechanism of secondary metabolites from the subgenus Callyspongia.

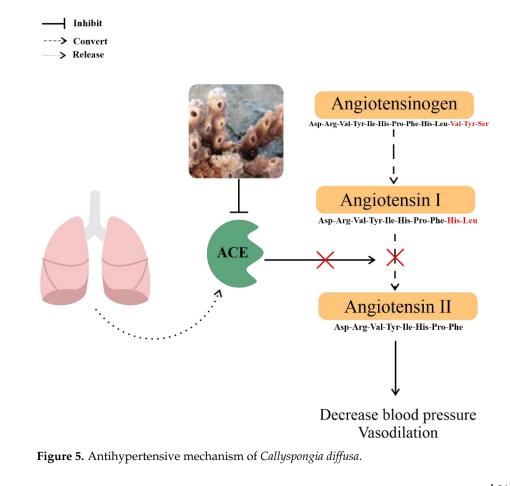
The methanolic extract of *Callyspongia samarensis* also exerts antidiabetic activity by enhancing the activity of AMP-activated protein kinase (AMPK) with an EC₅₀ of 14.47 μ g/mL, which is more potent compared with the positive control aspirin (EC₅₀ 100 μ g/mL). This activity may originate from compounds with phenolic groups in the extract [31]. AMPK is an important target for treating type-2 diabetes because its activation affects various aspects of cellular metabolism. It increases glucose metabolism, uptake in the bone and muscle, fatty acid oxidation in the bone, muscle, and liver, mitochondrial oxidative capacity, and insulin sensitivity, whereas it decreases fatty acid synthesis in the liver through GLUT-4 expression (Figure 4) [81–84].

4.2. Antihypertensive

Callypyrone A and callypyrone B (Figure 3) from *Callyspongia diffusa* exhibit antihypertensive activity by inhibiting angiotensin I-converting enzyme (ACE), which leads to a reduction in angiotensin production. Because angiotensin can constrict blood vessels and increase the heart work rate [85], ACE inhibition results in vasodilation and a decrease in blood pressure (Figure 5). The IC₅₀ values of these two compounds against ACE were 0.48 mM and 0.57 mM, respectively [33], weaker than the standard drug, captopril (IC₅₀ 0.36 mM) [33]. From the results, Callypyrone A and callypyrone B are not considered antihypertensive.

4.3. Anti-Inflammation

Diketopiperazines derived from *Callyspongia* sp., such as cyclo(L-Hyp-L-Ala), cyclo(L-Pro-Gly), cyclo(L-Pro-Phe), and cyclo(L-Pro-Ala), at a concentration of 5 μ g/mL, showed anti-inflammatory activity by increasing the secretion of the anti-inflammatory cytokine, interleukin-10 (IL-10) (Figure 6). IL10 levels were increased by 1.65-, 1.29-, 1.54, and 1.56-fold in J774A.1 cells, respectively [36].



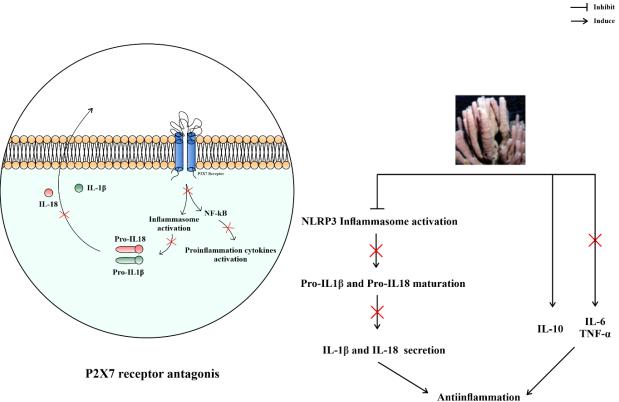


Figure 6. Prediction of anti-inflammatory mechanisms of compounds from the subgenus Callyspongia.

The anti-inflammatory activity of β -Sitosterol is independent of the adrenal pituitary axis. It inhibits the maturation of IL-1 β via the NOD-, LRR-, and pyrin domain-containing protein 3 (NLRP3) inflammasome and inhibits other inflammatory cytokines, such as IL-6 and tumor necrosis factor- α [86]. In a carrageenin-induced edema model in bilaterally adrenalectomized rats, β -Sitosterol exhibited a 54% anti-inflammatory effect at a dose of 320 mg/kg, which was weaker than the control, oxyphenbutazone, which had a 74% antiinflammatory effect at a dose of 100 mg/kg [37]. Therefore, β -Sitosterol has no potential as an anti-inflammatory agent.

Niphatoxin C significantly affects the viability of pre-monocytic THP 1 cells, which express the P2X7 receptor [65]. Activation of this receptor promotes inflammation by releasing inflammatory cytokines, such as IL-18 and IL-1 β , and by activating the NLRP3 inflammasome [87,88]. Thus, an antagonist of this receptor may inhibit the secretion of these cytokines (Figure 7). Furthermore, it can inhibit allograft rejection [87], sterile liver inflammation [88], and can potentially treat inflammatory diseases, such as osteoarthritis, rheumatoid arthritis, and chronic obstructive pulmonary disease [28,37].

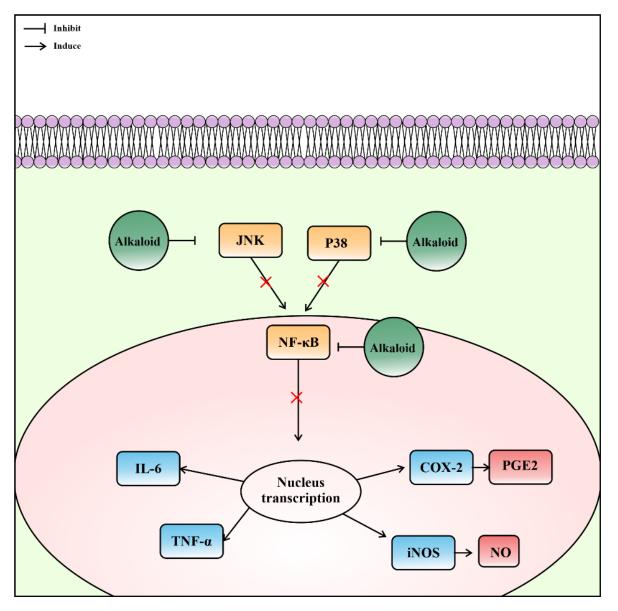


Figure 7. Prediction of anti-inflammatory mechanisms of alkaloids from the subgenus Callyspongia.

Callysterol (Figure 3) from *Callyspongia siphonella* exhibits anti-inflammatory activity against rat paw edema that was similar to the control drug, cortisone. The activity was measured by a reduction in edema volume of 19.5 ± 7.3 mL for callysterol and 17.0 ± 7.0 mL for cortisone, whereas the negative control was 61.9 ± 4.7 mL [38]. *Callyspongia crassa* extracts also showed anti-inflammatory effects with a 61.47% inhibition of protein denaturation [34]. Alkaloids are considered responsible for these anti-inflammatory mechanisms [89], which vary according to the metabolite. The alkaloid that has been identified from ethanolic extract of *Callyspongia siphonella* was 5-bromo trisindoline and 6-bromo trisindoline [7]. Although the specific mechanism of 5-bromo trisindoline and 6-bromo trisindoline is unknown, some indole alkaloids were known to interfere with the nuclear factor- κ B and c-Jun N-terminal kinase signaling pathways [90,91], preventing the synthesis or action of specific pro-inflammatory cytokines, and suppressing histamine release and nitric oxide production (Figure 7) [92,93]. Alkaloids are effective for treating inflammatory bowel disease [94–97].

4.4. Antifungal

Callyaerin A, B, and E (Figure 3) from *Callyspongia aerizusa* were shown to potently inhibit *Candida albicans*, with zones of inhibition of 25–30 mm, 15 mm, and 20 mm, respectively using the same concentration. Callyaerin A and E were more potent than callyaerin B [6].

Gelliusterol E from *Callyspongia* aff. *implexa* also exerts activity against chlamydial fungi in a dose-dependent manner by inhibiting the formation and growth of chlamydial inclusions. At the highest concentration tested (40 μ M), no inclusions were observed, similar to the effect of the control, tetracycline. Thus, this compound not only inhibits the formation of *Chlamydia*, but also affects its development cycle [39]. In addition, the structure of gelliusterol E is similar to that of cholesterol, which is needed for the growth of *Chlamydia trachomatis*. Furthermore, this compound inhibits lipid acquisition and fungal growth [98].

β-Sitosterol compounds found in *Callyspongia* sp. also exhibit antifungal activity against *Fusarium* spp, with 10 mm of average inhibition diameter [39,40]. (-)-siphonodiol from sponges display antifungal activities against *Trichophyton asteroids*, with moderately strong activity (MIC 25.0 µg/mL) [41]. Meanwhile, 4-hydroxybenzoic acid against *Ganoderma boninense* and (–)-loliolide display a broad spectrum of activity [42,43]. Active secondary metabolites that attack fungi are responsible for these antifungal activities, but their specific mechanisms of action remain unclear [99].

4.5. Cytotoxicity against Cancer Cell Lines

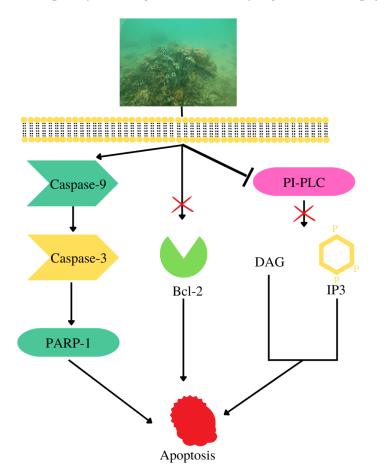
Callyspongia siphonella and *Callyspongia crassa* crude extracts were cytotoxic against a colon cancer (Caco-2) cell line with IC₅₀ values of 5.57 µg/mL and 13.05 µg/mL, respectively, and against breast cancer (MCF-7) cell line with IC₅₀ values of 1.39 µg/mL and 9.47 µg/mL [34]. Neviotine-C, neviotine A, sipholenol-A, and sipholenol from *Callyspongia siphonella* also exhibited cytotoxicity against cancer cell lines (Table 1). Sipholenol-A showed higher activity against the PC-3 and A549 cell lines (IC₅₀: $7.9 \pm 0.12 \mu$ M and $8.9 \pm 0.01 \mu$ M), sipholenol L against the HepG-2 cell line (IC₅₀: $18.7 \pm 0.9 \mu$ g/mL), and sipholenone A against the MCF-7 cell line (IC₅₀: $36.2 \pm 0.13 \mu$ M or $3 \pm 0.4 \mu$ g/mL) [44,45].

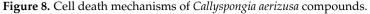
Callyspongiolide, extracted from *Callyspongia* sp., exhibited an IC₅₀ of 320 nM against a mouse lymphoma cell line (L5178Y), 70 nM against human Jurkat J16 T cells, and 60 nM against Ramos B lymphocyte cells [46]. Callypeptide A (Figure 3) inhibited the growth of human cancer cells with GI₅₀ values of 29 μ M against breast adenocarcinoma (MDA-MB-231), 30 μ M against colorectal carcinoma (HT-29), and 18.5 μ M against lung carcinoma (A549). Its activity was weaker compared with doxorubicin as a control (GI₅₀ values of 0.30, 0.40, and 0.35 μ M, respectively) [47].

Callyazepine and (3R)-methylazacyclodecane (Figure 3) exhibited IC₅₀ values of 7.4 μ M and 3.2 μ M against K562 cells, and 3.0 μ M and 3.8 μ M against A549 cells, re-

spectively [100]. Hysmenialdisine from *Callyspongia* sp. had an IC₅₀ value of 3.1 μ M against colonic adenocarcinoma cells (SW620) and 2.0 μ M against epidermoid carcinoma cells (KB-3-1) [49]. In addition, it produces akaterpine, which exhibited an IC₅₀ of 0.5 μ g/mL against phosphoinositide-specific phospholipase C [58]. Callyspongamide A (Figure 3), isolated from *Callyspongia fistularis*, had an IC₅₀ of 4.1 μ g/mL against HeLa cells [55].

Methanolic extract of *Callyspongia aerizusa* had IC₅₀ values of 9.38 µg/mL against A549 cells, 3.12 µg/mL against TE-8 cells, 10.62 µg/mL against HepG-2 cells, and 10.72 µg/mL against MIA PaCa-2 cells [59]. It also produces callyaerin E and H (Figure 6), which exhibited IC₅₀ values of 0.39 µM and 0.48 µM against L5178Y cells [6]. *Callyspongia aerizusa* extract stimulates the expression of caspase-9, which in turn activates caspase-3, and subsequently downregulates Bcl-2, a key regulator of antiapoptosis (Figure 8) [59].





Callystatin (Figure 3) from *Callyspongia truncata* exhibited IC₅₀ values of 0.01 μ g/mL against KB cells and 20 pg/mL against L1210 cells [101]. *Callyspongia* sp. also contains two unknown compounds with antiproliferative activity against TR-LE cells: (–)-(3R,18R) alcohol with an IC₅₀ of 0.11 μ M and (+)-(3S,18S) with an IC₅₀ of 0.47 μ M [52].

The US National Cancer Institute classifies the cytotoxicity of a compound as high if its $IC_{50} < 20 \ \mu\text{g/mL}$, moderate if it falls between $21-200 \ \mu\text{g/mL}$, weak if it falls between $201-500 \ \mu\text{g/mL}$, and non-cytotoxic if the $IC_{50} > 500 \ \mu\text{g/mL}$ [102]. Based on these criteria, most *Callyspongia* extracts possess high cytotoxic activity, except that of *Callyspongia schulzei*, which exhibits moderate activity, but no study was conducted against non-cancerous cell line except for *Callyspongia aerizusa* extract. Cytotoxicity of the methanol extract of *Callyspongia aerizusa* against TE-8 cells (IC₅₀ 3.12 $\mu\text{g/mL}$) was more effective compared with that of the control drug, cisplatin (IC₅₀ 8.1 $\mu\text{g/mL}/27 \ \mu\text{M}$), meanwhile this extract was non-cytotoxic for non-cancerous cell (HET-1A cell) up to 1000 $\mu\text{g/mL}$. The compounds responsible for suppressing A549 cell proliferation were identified as ergots-22-en-3-one and ergost-7-en-3-ol [59,103].

4.6. Antimicrobial and Antiparasitic

Isoakaterpine compounds from *Callyspongia* sp. exert antiparasitic activity by inhibiting adenosine phosphoribosyltransferase, one of the functional routes in *Leishmania* adenine metabolism, with an IC₅₀ of 1.05 μ M [11,104], resulting in death of the parasite (Figure 9).

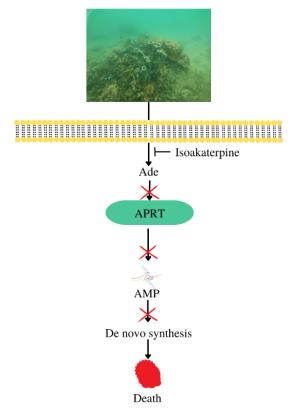


Figure 9. Antiparasitic mechanism of compounds from Callyspongia sp.

Besides antiparasitic activity, the subgenus *Callyspongia* possesses antituberculosis activity resulting from callyaerins A and B (Figure 5) isolated from *Callyspongia aerizusa*. Their MIC₉₀ values (2 μ M and 5 μ M, respectively) were less effective compared with the controls, rifampicin (<1 μ M), ethambutol (1.25 μ M), and isoniazid (0.625 μ M). Beside the weaker activity compared with the control, there is no in vivo data to support this activity. These compounds inhibited the growth of *Mycobacterium tuberculosis* as evidenced by reduced cell viability using the resazurin dye reduction method and measuring cell fluorescence [105].

Siphonocolin from *Callyspongia siphonella* exhibited antimicrobial activity against *Pseudomonas aeruginosa* with an MIC of 64 μ g/mL [61]. Moreover, neviotine A, sipholenol L, and sipholenone A from *Callyspongia siphonella* also exhibited antimicrobial activity against *Staphylococcus aureus*, *Bacillus subtilis*, and *Escherichia coli* (Table 1). Neviotine has higher antimicrobial activity with a zone of inhibition against *Staphylococcus aureus* of 14.1 \pm 0.72 mm, against *Bacillus subtilis* of 17.2 \pm 0.58 mm, and against *Escherichia coli* of 12.7 \pm 0.58 mm [45].

Callyspongia crassa extract potently inhibited *Bacillus subtilis* and *Staphylococcus aureus* with zones of inhibition of 16–25 mm (at concentration 500 μ g/mL), 9–15 mm and 16–25 mm (at concentration 250 μ g/mL) respectively, while exhibiting high activity against marine bacteria. The IC₅₀ of the extract was determined by a microdilution test and ranged from 5 μ g/mL to 500 μ g/mL. *Callyspongia crassa* is the most active among the Red sea sponges against *Bacillus subtilis*, with an LC₅₀ 18.2 \pm 3.56 μ g/mL, but was weak against

Staphylococcus aureus with an LC₅₀ 215.2 \pm 32.9 µg/mL [60]. Callyaerin A also exhibits antimicrobial activity against *Escherichia coli* and *Staphylococcus aureus*, with zones of inhibition of 10–15 mm and 9 mm, respectively, whereas callyaerin E has activity against *Escherichia coli*, *Bacillus subtilis*, and *Staphylococcus aureus*, with zones of inhibition of 9–11 mm, 15–17 mm, and 9–10 mm, respectively [6].

4.7. Antioxidant

Sponge extracts inhibited oxidative stress and carbohydrate hydrolysis enzymes linearly in a dose-dependent manner. Based on the 2,2-diphenyl-1-picryl-hydrazyl-hydrate assay, a *Callyspongia aerizusa* extract displayed an antioxidant activity of 56.6% at 0.5 μ g/mL, 57.2% at 0.6 μ g/mL, and 58.4% at 0.7 μ g/mL, indicating that it may be classified as an antioxidant (>50%). *Callyspongia crassa* extract showed an antioxidant activity of 58.1% at 671 μ g/mL, which was lower than the control, ascorbic acid (>90%), likely because *Callyspongia* was used in the form of an extract [34,62].

4.8. Antiallergic

The compound 3-(2-(4-hydroxyphenyl)-2-oxoethyl)-5,6-dihydropyridine-2(1H)-one was isolated from an ethanol extract of *Callyspongia* sp. [63]. This δ -lactam derivative was predicted to possess antiallergic activity based on its in silico inhibition of β -hexosaminidase (β -hex), which was determined using Origin 8.0. This compound inhibited β -hex activity in rat basophilic leukemia cells (RBL-2H3) with an IC₅₀ of 18.7 ± 6.7 μ M, which was weaker than the positive control, ketotifen fumarate (IC₅₀ 15.0 ± 1.3 μ M), but more potent than azelastine (IC₅₀ 32.0 μ M) [64]. β -hex is released from mast cell degranulation, thus its activity can be used as a biomarker of mast cell allergic response to quantify degranulation [106–108].

4.9. Antiviral

Callyspongia crassa and *Callyspongia siphonella* extracts exhibited cytotoxic effects on Vero cells, which were cultured for the isolation and multiplication of enterovirus and hepatitis A virus, with MICs of 9.765 μ g/mL and 0.625 μ g/mL, respectively. The maximum non-toxic concentrations of these extracts were 4.9 and 0.3 μ g/mL, respectively. *Callyspongia crassa* crude extract had an antiviral activity of 85.3%, whereas the antiviral activity of *Callyspongia siphonella* extract was 16.4% [34].

4.10. Immunomodulatory

Callyspongia extract at doses of 300 mg/kg and 400 mg/kg body weight, increased *S. aureus*-induced production of interferon- γ (IFN- γ) (455.265 pg/mL and 384.319 pg/mL) and tumor necrosis factor- α (TNF- α)(954 pg/mL and 1042 pg/mL) in male Wistar rats. It was more effective compared with 0.5% carboxymethyl cellulose sodium as negative control (160.314 pg/mL for INF- γ and 785.5 pg/mL for TNF- α) and bay leaf extract as positive control (353.486 pg/mL for INF- γ and 976 pg/mL for TNF- α) [68]. β -Sitosterol compounds from *Callyspongia* spp. modulate the activity of dendritic cells and increase the viability of peripheral blood mononuclear cells [67]. Siphonodiol, callyspongidiol, and 14,15-dihydrosphonodiol modulate the function of dendritic cells for T1 cell proliferation as well as IL-2 and IFN- γ production [66]. IL-2, along with other ILs, regulates innate and adaptive immunity by promoting an increase in the population of various immune cells [109,110]. Meanwhile, IFN- γ activates macrophages and enhances their immune response [111]. *Callyspongia* extract can stimulate the branch of the immune system involved in forming a receptor complex with gp130 to eventually inhibit the bioactivity of IL-6 (Figure 10) [112,113].

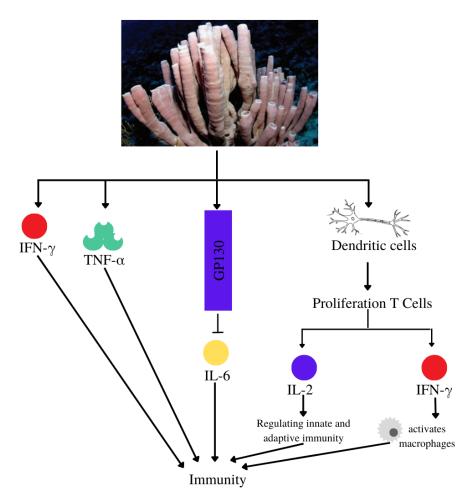


Figure 10. Immunomodulatory mechanisms of compounds from the subgenus Callyspongia.

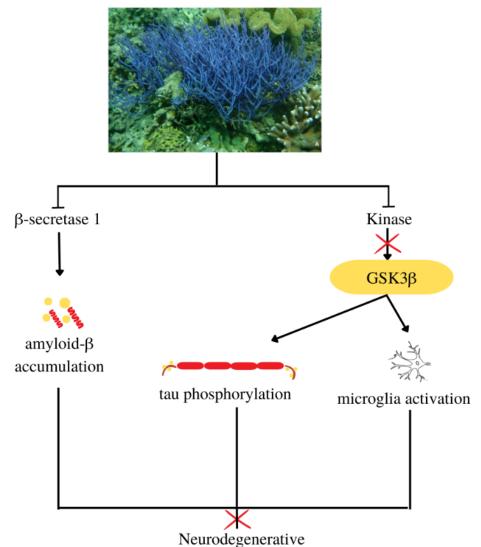
4.11. Antineurodegenerative

• β-secretase 1

Selectively inhibiting β -secretase 1 in specific subcellular compartments is an effective strategy to reduce the accumulation of neurotoxic amyloid plaques [114]. The methanol extract of *Callyspongia samarensis* significantly and non-competitively inhibited β -secretase 1 (IC₅₀ 99.82 µg/mL). An acute oral toxicity test revealed that the extract was non-toxic, with an LD₅₀ value of less than 2000 mg/kg. Moreover, an unknown compound in the extract, with a mass/charge ratio of 337.9 [M + H]+, was able to permeate the blood–brain barrier, making it a suitable candidate for developing central nervous system drugs [31].

• Kinase inhibitor

Kinases have a role in neurodevelopmental and central nervous system physiology. Activation of the glycogen synthase kinase 3β (GSK3 β) results in tau phosphorylation, amyloid- β accumulation, microglia activation, neurogenesis, and memory abnormalities [115]. This suggests that its inhibition restores and repairs pathways and neurogenesis (Figure 11) [116,117]. Hymenialdisine, isolated from *Callyspongia* sp. (CMB-01152), inhibits casein kinase 1, cyclin-dependent kinase 5, and GSK3 β with IC₅₀ values of 0.03 µg/mL, 0.16 µg/mL, and 0.07 µg/mL, respectively. They abnormal hyperphosphorylate highly soluble microtubule-associated proteins to produce neurofibrillary tangles [49].



reurodegenerative

Figure 11. Antineurodegenerative mechanisms of compounds from the subgenus Callyspongia.

4.12. Antiosteoporotic

Neviotine A and D are isolated triterpene-type compounds from *Callyspongia siphonella*. These compounds possess antiosteoporotic activity by inhibiting receptor activator of nuclear factor-kB ligand (Rankl) with IC₅₀ values of 32.8 μ M and 12.8 μ M (quercetin as positive control: 25 μ g/mL) [69]. The interaction between Rankl and Rank receptor translocate the tumor necrosis factor receptor-associated factors (TRAF6) to the RANK cytoplasmic domain, results in the activation of ERK, p38, and JNK via activation of signaling cascades and downstream targets. Thus, AP-1 and NF-kB transcription factors were activated and stimulated the formation and activity of osteoclasts, which affect resorption activity [118]. Neviotine A and D inhibit cell differentiation into multinucleated tartrate-resistant acid phosphatase (TRAP)-positive osteoclasts, which was upregulated via RANKL-induced osteoclastogenesis (Figure 12) [69,119].

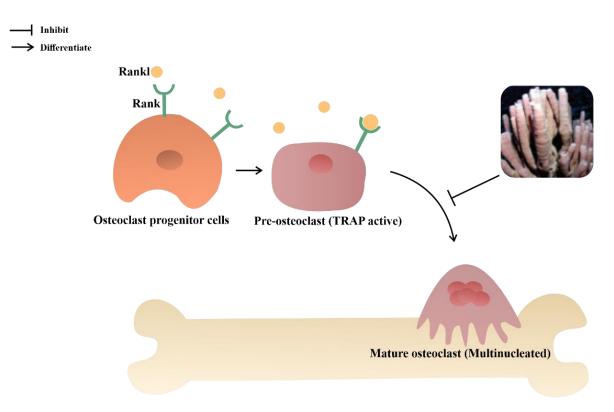


Figure 12. Antiosteoporotic mechanisms of Neviotine A and D from the Callyspongia siphonella.

5. Conclusions

In the 41 articles we reviewed, the pharmacological activities that Callyspongia spp. is reported to possess include cytotoxic against cancer cell line (36%), antifungal (10%), antiinflammatory (10%), immunomodulatory (10%), antidiabetic and antiobesity (6%), antimicrobial (8%), antioxidant (4%), antineurodegenerative (4%), antihypercholesterolemic (2%), antihypertensive (2%), antiparasitic (2%), antiallergic (2%), antiviral (2%), antiosteoporotic (2%), and antituberculosis (2%) activities (Figure 13). The most studied pharmacological activity is cytotoxicity against cancer cell lines. Most of the research was limited to in vitro testing and there is insufficient in vivo data to support such activity. In addition, not all secondary metabolites responsible for certain activities have been identified. Several activities require modification and further study because of a lack of testing or low activity. For example, the antiallergic activity of Callyspongia sp. predicted from in silico results or the antioxidant, antituberculosis, and anti-inflammatory activities of Callyspongia extract were weaker compared with those of the control drugs. Although many promising compounds with a high potential to become drugs remain to be comprehensively evaluated in vivo, Callyspongia with its known mechanisms of action, such as antidiabetic and cytotoxic effects, may be further developed for targeted therapy.

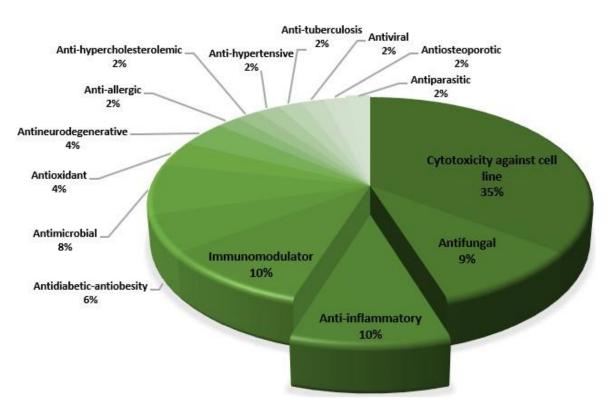


Figure 13. Distribution of the pharmacological activities reported in *Callyspongia* spp.

Author Contributions: Conceptualization, Y.E.H.; methodology, A.A.N. and S.S.; software, A.A.N. and S.S.; data curation, A.A.N. and S.S.; writing—original draft preparation, A.A.N. and S.S.; writing—review and editing, Y.E.H., A.Z., R.H. and I.S.; visualization, Y.E.H., A.Z., R.H. and I.S.; supervision, Y.E.H., A.Z., R.H. and I.S.; project administration, Y.E.H., A.Z., R.H. and I.S.; funding acquisition, Y.E.H., A.Z., R.H. and I.S. All authors have read and agreed to the published version of the manuscript.

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Conflicts of Interest: The authors declare no conflict of interest.

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