

## Supplementary Materials:

# Harvesting Environmental Microalgal Blooms for Remediation and Resource Recovery: A Laboratory Scale Investigation with Economic and Microbial Community Impact Assessment

Jagroop Pandhal, Wai L. Choon, Rahul V. Kapoore, David A. Russo, James Hanotu, I. A. Grant Wilson, Pratik Desai, Malcolm Bailey, William J. Zimmerman and Andrew S. Ferguson

**Table S1:** Complete composition of artificial freshwater growth medium.

Component	Concentration (mg/L)
NaHCO <sub>3</sub>	192
MnCl <sub>2</sub> .4H <sub>2</sub> O	0.18
MgSO <sub>4</sub> .7H <sub>2</sub> O	115
KCl	0.45
H <sub>2</sub> SeO <sub>3</sub>	0.0016
Ca(NO <sub>3</sub> ) <sub>2</sub> .4H <sub>2</sub> O	0.8
NH <sub>4</sub> Cl	1
KH <sub>2</sub> PO <sub>4</sub>	0.025
K <sub>2</sub> PO <sub>4</sub>	0.025
ZnSO <sub>4</sub> .7H <sub>2</sub> O	0.022
Na <sub>2</sub> EDTA.2H <sub>2</sub> O	0.5
H <sub>3</sub> BO <sub>3</sub>	0.114
FeSO <sub>4</sub> .7H <sub>2</sub> O	0.05
CuSO <sub>4</sub> .5H <sub>2</sub> O	0.016
CoCl <sub>2</sub> .6H <sub>2</sub> O	0.016
(NH <sub>4</sub> ) <sub>6</sub> Mo <sub>7</sub> O <sub>24</sub> .4H <sub>2</sub> O	0.011

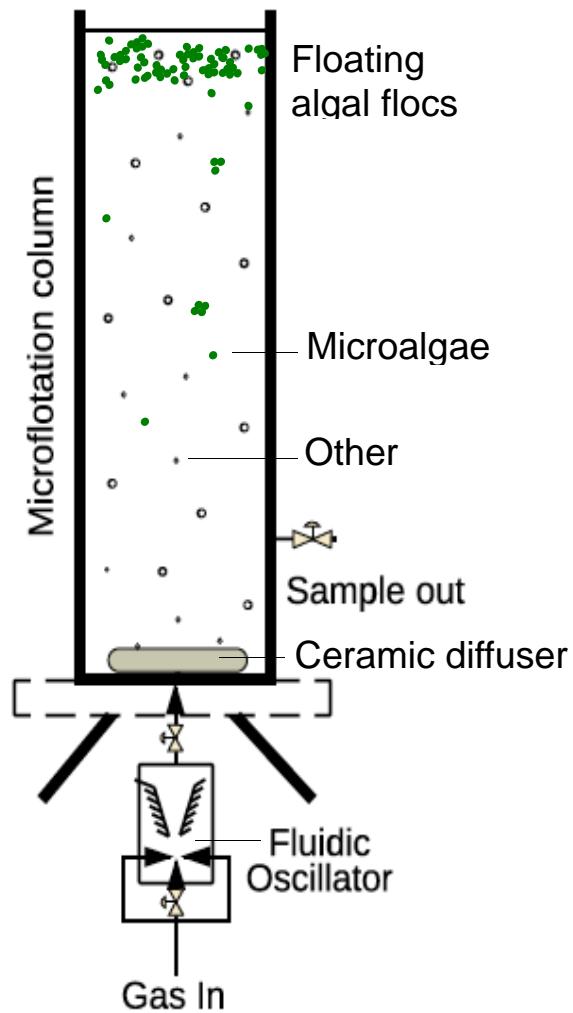
**Table S2:** A summary of FAME profiles between different microalgae species mostly focussing on recent studies. The studies were selected based on similar methodologies applied to sample processing steps, catalysts, GC columns and GC conditions. NB. Percentages will vary depending on the number of FAMEs quantified in each experiment and only the major FAMEs detected are shown for clarity in the comparison (see citation for more details of exact experimental procedures).

Species	Author	Year	C14:	C16:	C16:	C16:	C16:	C17:	C18:	C18:	C18:	C18:	C20:	C20:	C22:	Other	
			0	0	1	2	3	0	0	1	2	3	4	4	5	6	s
<b>Published results</b>																	
<i>Chlamydomonas reinhardtii</i>	Talebi et al.	2013	-	23.8	1.9	-	-	4.4	19.7	6.6	25.9	-	-	-	-	1.2	
<i>Chlorella pyrenoidosa</i>	Montes et al.	2011	0.7	17.3	0.8	7.0	9.3	1.2	3.3	18.5	41.8	-	-	-	-	-	
<i>Chlorella sp. 227</i>	Sunja et al.	2010	-	22.8	-	-	-	-	8.3	17.2	52.0	-	-	-	-	-	
<i>Chlorella vulgaris</i>	Laurens et al.	2012	0.2	18.3	12.3	6.1	0.3	1.2	18.9	15.1	24.0	-	0.0	0.0	-	0.9	
<i>Dunaliella primolecta</i>	Lang et al.	2011	0.6	26.0	0.9	-	-	1.6	16.3	7.0	38.7	0.6	-	-	-	-	
<i>Dunaliella tertiolecta</i>	Chen et al.	2011	-	28.1	0.0	2.8	1.4	0.6	19.3	14.7	33.2	-	-	-	-	-	
<i>Emiliana huxleyi</i>	Lang et al.	2011	18.8	10.3	-	-	-	10.8	42.2	-	-	8.7	-	-	-	9.2	
<i>Heterosigma akashiwo</i>	Lang et al.	2011	6.6	40.0	12.7	4.0				4.5	6.7	5.2	3.5	14.8	-	-	
<i>Isochrysis galbana</i>	Fidalgo et al.	1998	14.4	12.6	19.4	0.5	0.2	0.4	3.1	0.7	1.6	15.2	-	22.2	7.9	0.3	
<i>Nannochloris sp.</i>	Lang et al.	2011	13.3	17.8	-	-	-	23.9	10.8	28.2	6.1	-	-	-	-	-	
<i>Nannochloropsis</i>	Koberg et al.	2011	6.6	42.8	27.3	-	-	0.4	1.0	9.1	1.3	0.4	-	-	4.9	-	3.0
<i>Nannochloropsis oculata</i>	Converti et al.	2009	-	62.0	-	-	-	11.0	5.0	8.0	15.0	-	-	-	-	-	
<i>Nannochloropsis</i>	Wageningen	2012	7.0	19.0	30.0	-	-	-	8.0	-	-	-	6.0	26.0	-	-	

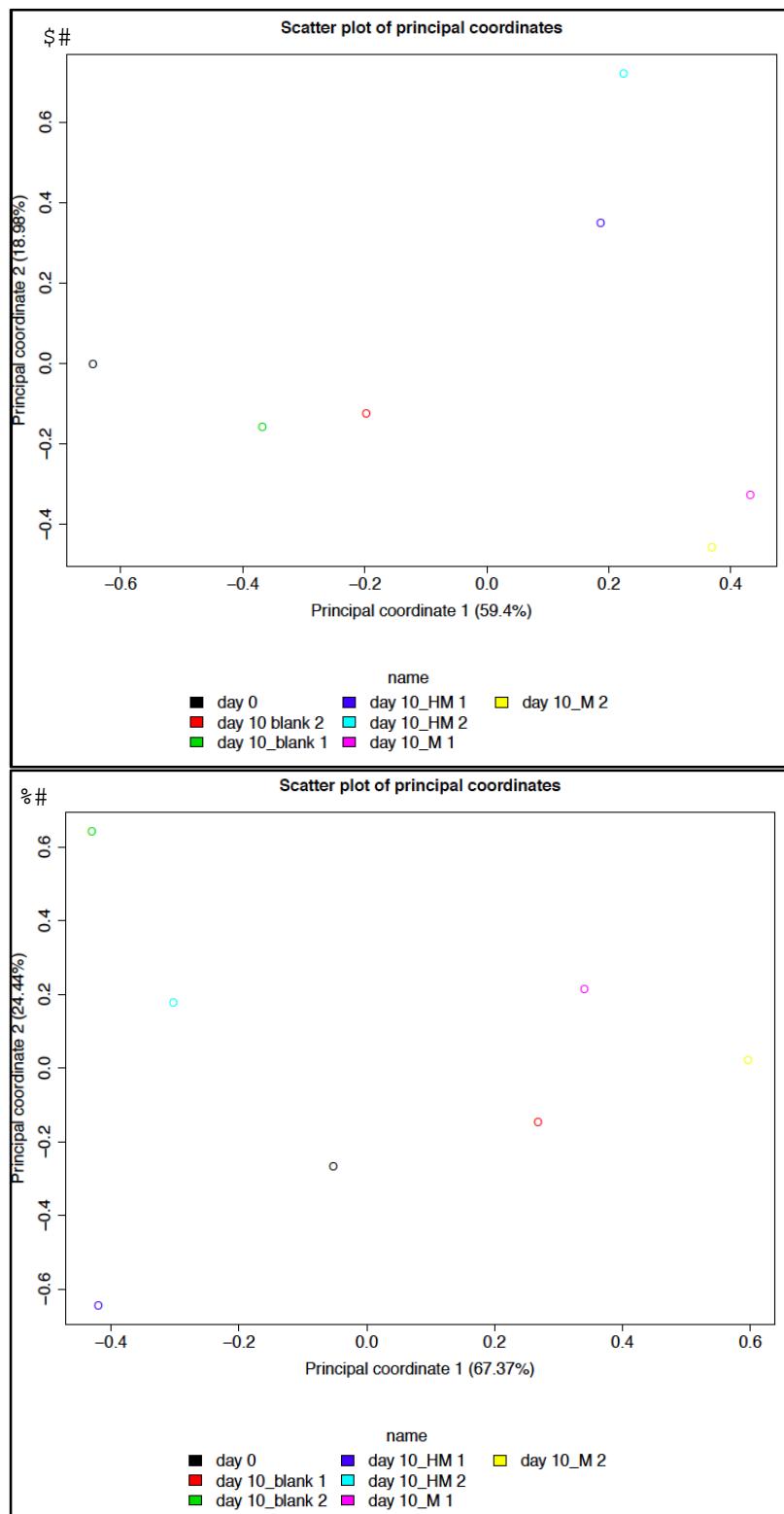
<i>Salina</i>	at al.																		
<i>Parietochloris incisa</i>	Lang et al.	2011	0.0	19.8	-	5.2	-	18.2	10.2	14.3	14.3	-	14.0	4.3	-	-	-		
<i>Pavlova lutheri</i>	Lang et al.	2011	10.1	11.1	26.3	-	-	5.2	0.6	0.5	9.1	0.3	18.0	9.7	-	-	-		
<i>Phaeodactylum tricornutum</i>	Lang et al.	2011	6.7	14.7	43.6	2.0	-	-	15.8	0.5	0.4	1.1	-	14.4	0.7	-	-		
<i>Scenedesmus obliquus</i>	Breuer et al. Talebi et al.	2013	2.0	27.0	45.0	-	1.0	2.0	3.0	1.0	<1	<1	1.0	14.0	-	2.0	-		
<i>Scenedesmus sp.</i>	al.	2013	-	15.6	4.1	-	-	3.0	15.2	7.0	23.0	-	-	-	-	-	7.5		
<i>Schizochytrium limacinum</i>	Johnson et al.	2009	5.3	56.5	-	-	-	-	-	-	-	-	-	-	29.7	5.2	-		
<i>Thalassiosira weissflogii</i>	Borges et al.	2011	6.8	17.2	24.3	-	18.1	2.5	10.0	-	-	-	-	17.3	1.6	3.7	-		
<i>Thalassiosira weissflogii</i>	Lang et al. Krohn et al.	2011	8.8	36.6	40.5	-	-	-	14.0	-	-	-	-	-	-	-	-		
<i>Wild algae</i>		2011	-	47.1	-	-	-	7.0	-	-	-	-	-	-	-	-	-		
<b>This study</b>																			
NBAS	Pandhal et al.	2015	9.8	28.1	1.6	-	-	2.9	5.0	8.9	12.2	31.4	-	-	-	-	-		
SBAS	Pandhal et al.	2015	7.2	53.7	4.4	-	-	2.5	4.0	21.5	0.2	-	-	-	0.3	6.1	-		
ESTH	Pandhal et al.	2015	8.2	30.3	0.6	-	-	0.9	9.2	2.9	7.6	16.4	-	-	-	4.0	19.8		
NLT	Pandhal et al.	2015	0.5	52.9	0.9	-	-	0.3	3.0	3.6	12.5	23.8	-	-	-	-	2.5		
WRP	Pandhal et al.	2015	19.3	18.0	1.6	-	-	0.7	13.7	8.7	4.3	22.1	-	-	0.3	11.1	-		

**Table S3:** Chlorophyll a measurements during eFLOAT long-term experiment.

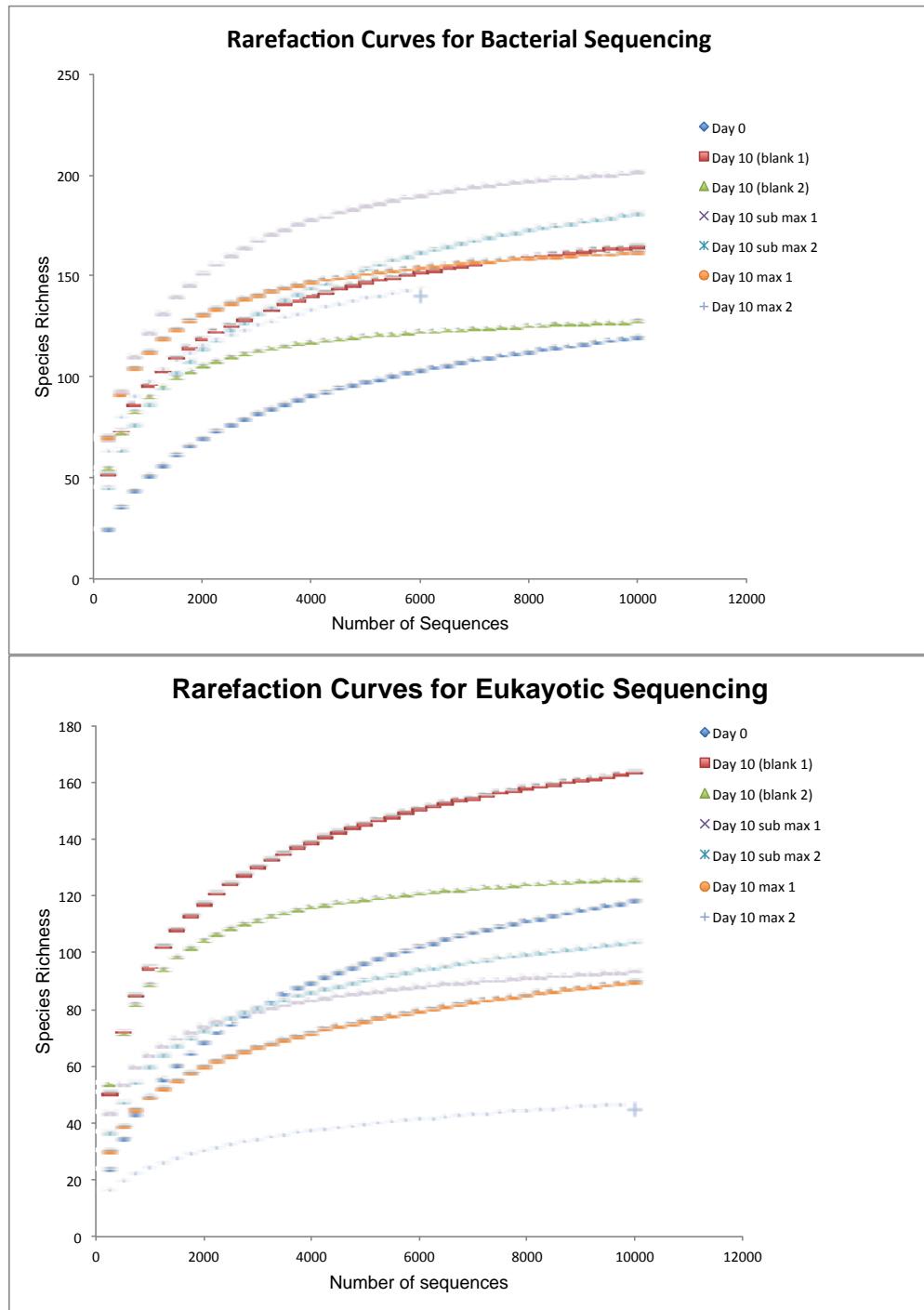
Sample	Time post harvest (days)	Chl a ( $\mu\text{g L}^{-1}$ )	Average Chl <i>a</i> ( $\mu\text{g L}^{-1}$ )	Variation ( $\mu\text{g L}^{-1}$ )	Decrease %	Average %	Variation %
WRP	0	443.23					
Blank 1	10	404.37	395.7	12.2	8.8	10.7	2.8
Blank 2					12.7		
Sub-max 1	10	145.37	154.1	12.4	67.2	65.2	2.8
Sub-max 2					63.2		
Max 1	10	95.82	79.0	23.8	78.4	82.2	5.4
Max 2					86.0		



**Figure S1:** 1 litre eFLOAT Rig using during laboratory tests



**Figure S2:** Scatter plot of coordinates (PCoA) to show relative abundance (%) variation between identified genus/class in biological replicate samples (A) Bacterial (16S rDNA sequencing) diversity (B) Eukaryotic (18S rDNA sequencing) diversity. HM: half max harvesting, M: max harvesting.



**Figure S3:** Rarefaction plots for bacterial primers (28F-519R) and eukaryotic primers (565-981).