

Article

Bacillamide F, Extracted from Marine *Bacillus atrophaeus* C89, Preliminary Effects on Leukemia Cell Lines

Shengnan Zhang ^{1,†}, Giorgia Croppi ^{2,†}, Heng Hu ¹, Yingxin Li ¹, Chunmiao Zhu ¹, Fang Wu ^{2,*} , Fengli Zhang ^{1,*} and Zhiyong Li ¹ 

¹ Marine Biotechnology Laboratory, State Key Laboratory of Microbial Metabolism, School of Life Sciences & Biotechnology, Shanghai Jiao Tong University, Shanghai 200240, China

² Key Laboratory of Systems Biomedicine (Ministry of Education), Shanghai Center for Systems Biomedicine, Shanghai Jiao Tong University, Shanghai 200240, China

* Correspondence: fang.wu@sjtu.edu.cn (F.W.); zhangfengli@sjtu.edu.cn (F.Z.)

† These authors contributed equally to this work.

Simple Summary: Bacillamides are important bioactive natural compounds. This paper depicts for the first time a study of a novel thiazole alkaloid compound bacillamide F that was isolated from the marine *Bacillus atrophaeus* C89. Bacillamide F inhibits the leukemia cell lines HL60 and Jurkat. The work provides a new indication of the pharmacological activity of bacillamides.

Abstract: Developing new treatments for leukemia is essential since current therapies often suffer from drug resistance and toxicity. Bacillamides are very promising, naturally occurring compounds with various bioactivities. In the present study, we investigated the use of bacillamide analogues, a new thiazole alkaloid bacillamide F that was isolated from marine *Bacillus atrophaeus* C89 associated with sponge *Dysidea avara*. The structure of the new compound bacillamide F with indolyl–thiazolyl–pyrrolidine ring was determined by high resolution mass spectrometry, secondary mass spectrometry, and nuclear magnetic resonance analyses. Intriguingly, bacillamide F is able to inhibit the proliferation of an acute myeloid leukemia cell line HL60 (IC₅₀ (24 h) 21.82 μM), and an acute T-cell leukemia Jurkat (IC₅₀ (24 h) 46.90 μM), rather than inhibit the proliferation of the acute histiocytic lymphoma U-937 cell line, human fetal lung fibroblast MRC-5 cell line, and some solid tumor cell lines (IC₅₀ (24 h) > 100 μM). The study provides a new indication of the pharmacological activity of natural product bacillamides.

Keywords: bacillamide; *Bacillus atrophaeus*; leukemia; antiproliferation effect



Citation: Zhang, S.; Croppi, G.; Hu, H.; Li, Y.; Zhu, C.; Wu, F.; Zhang, F.; Li, Z. Bacillamide F, Extracted from Marine *Bacillus atrophaeus* C89, Preliminary Effects on Leukemia Cell Lines. *Biology* **2022**, *11*, 1712. <https://doi.org/10.3390/biology11121712>

Academic Editor: Maria Rosaria Ricciardi

Received: 15 October 2022

Accepted: 23 November 2022

Published: 25 November 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Leukemia is a disease that develops from bone marrow, and can be classified into acute leukemia and chronic leukemia [1]. In regard to the type of cells involved in the disease, leukemia is further divided into acute myelogenous leukemia (AML) [2], chronic myeloid leukemia (CML) [3], acute lymphoblastic leukemia (ALL) [4], and chronic lymphoblastic leukemia (CLL) [5]. Moreover, each class of leukemia contains several subtypes [6]. For instance, AML can be subdivided into more than ten different subtypes according to the defining genetic abnormalities in the new WHO (World Health Organization) classification scheme. Due to the many different types of leukemia and fast progress of resistance of certain subtypes for current therapy, new treatments are becoming necessary [7,8].

Bacillamides, which are non-ribosomal peptides thiazole alkaloids, can be categorized as bacillamide A, B, C, D and E (Figure 1a) [9–14]. Bacillamide A was isolated from the marine bacterium *Bacillus* sp. SY-1 [9]. Bacillamide B was isolated from *Bacillus endophyticus* [10] and *Microbispora aerate* IMBAS-11A [11]. Bacillamide C was isolated from *Bacillus endophyticus* [10] and *Bacillus atrophaeus* C89 associated with marine sponge *Dysidea avara* [12]. Bacillamide D was isolated from *Thermoactinomyces* sp. strain TM-64 [13].

Bacillamide E was synthesized *in vitro* by Bloudoff et al. [14]. Bacillamides and their derivatives show significant algicidal activities against a wide range of dinoflagellates, raphidophytes, and particular species of cyanobacteria [9,15], and bacillamide analogues show cytotoxic and anti-inflammatory activities [16]. Moreover, thiazole derivatives exhibit good anticancer activity with influence on cell cycle, DNA fragmentation and mitochondrial depolarization [17].

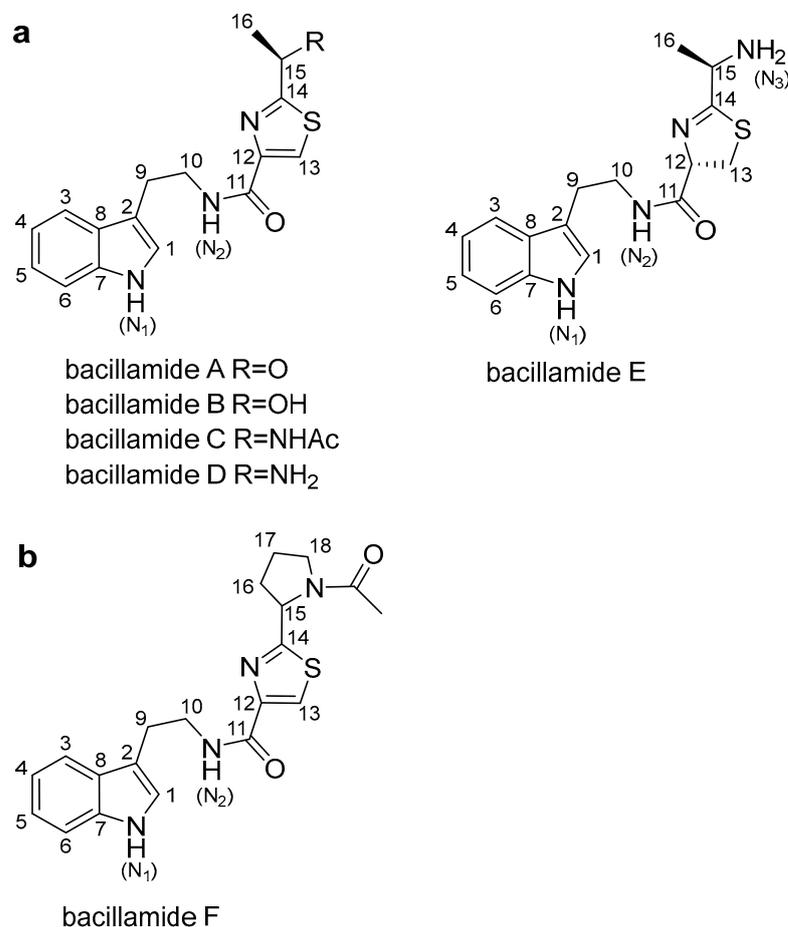


Figure 1. The bacillamide family structures of compounds: (a) This represents the known bacillamide compounds (bacillamide A–E); (b) This represents the new compound, bacillamide F.

Precursor-directed biosynthesis (PDB) was an effective method used to generate novel NRPS products analogues [18]. In our previous study, substrate selection of adenylation domains for non-ribosomal peptide synthetase (NRPS) in bacillamide C biosynthesis by *B. atrophaeus* C89 was investigated, and the results proved the A1 and A2 domains can catalyze a variety of substrate amino acids, including proline [19]. Here, we isolated a novel thiazole alkaloid bacillamide F and found that the production of bacillamide F was increased through the addition of precursor L-proline into a culture medium of marine *B. atrophaeus* C89. We elucidated the structure and inhibition activity of bacillamide F in this research. To our knowledge, this is the first report of bacillamide analogue bearing a 2-(thiazole-2-yl)pyrrolidine, and a biosynthetic pathway of bacillamide F was predicted. In this study, the antiproliferation effect on leukemia of bacillamide F was investigated, and the results showed that bacillamide F inhibited the proliferation of leukemia cell lines Jurkat and HL60.

2. Materials and Methods

2.1. Biosynthesis and Extraction of Bacillamide F

B. atrophaeus C89 (CCTCC AB 2016282, Genome Genebank No, AJRJ00000000.1) was isolated from the sponge *D. avara* in the South China Sea [12]. *B. atrophaeus* C89 was incubated at 28 °C in a solid medium containing 5 g of beef extract, 10 g of peptone, and 20 g of agar in every 1000 mL of artificial seawater (ASW) with pH 7.0–7.2. After 24 h cultivation, the strain was cultured in a modified medium (consisting of 3.64 g/L corn starch, 6.29 g/L CaCO₃, 4.00 g/L soy peptone, and 6.00 g/L peptone with ASW) at 28 °C for 96 h; and 0.10 g/L cysteine, 0.02 g/L tryptophan, and 0.02 g/L proline were added to the culture medium when the strain was cultured at 40 h.

After 96 h cultivation, the medium (18 L) was extracted three times with ethyl acetate (EtOAc). Evaporation of EtOAc extracts gave the yellow oil (3.40 g) which was subjected to Sephadex LH-20 columns with methanol (MeOH). Consequently, 2 fractions (Fr. 1–2) were obtained according to Thin Layer Chromatography (TLC) monitoring. Fr. 2 (2.40 g) was further purified on Medium Pressure Preparative Liquid Chromatography (MPLC). As a result, 8 fractions (Fr. 3–10), according to reversed-phase high-performance liquid chromatography (HPLC) monitoring (1 mL/min, detector UV λ_{\max} 220 nm, acetonitrile/H₂O 20: 80–45: 55, 15 min), were eluted with increasing proportions, respectively, of MeOH (H₂O/MeOH (*v/v*, 90:10, 70:30, 50:50, 30:70, 10:90, 0:100)). Finally, Fr. 8 (136 mg) was further purified on semi-preparative HPLC (1 mL/min, detector UV λ_{\max} 220 nm, acetonitrile/H₂O 30: 70) to deliver bacillamide F (34.80 mg).

2.2. General Experimental Procedures

Column chromatography (CC): Sephadex LH-20 (Amersham Biosciences, Amersham, UK). MPLC: Cheetah Fs-9200t (Bonna-Agela, Tianjin, China). TLC: precoated silica gel plates (Qingdao Marine Chemical Company, Qingdao, China). Semi-preparative HPLC: Agilent 1200 series liquid chromatograph Durashell C18-AM column (4.60 × 250 mm; i.d. 5 μ m). Reversed-phase HPLC: Agilent 1200 series liquid chromatograph ZORBAX Eclipse XDB-C18 column (4.60 × 150 mm; i.d. 5 μ m). Mass spectra: ACQUITY™ UPLC & Q-TOF MS Premier instrument equipped with an electrospray ionization (ESI) probe operating in positive-ion mode with direct infusion. ¹H and ¹³C- Nuclear Magnetic Resonance (NMR) spectra: Avance III 600 MHz spectrometer; chemical shifts δ in ppm, with residual methanol-d₄ (δ_{H} 3.31, δ_{C} 49.15) as internal standard, coupling constant *J* in Hz. ¹H- and ¹³C-NMR assignments were supported by ¹H, ¹H COSY, Heteronuclear multiple quantum coherence (HMQC), heteronuclear multiple bond correlation (HMBC) experiments. All chemicals used in the study were analytical grade and HPLC grade.

2.3. Cell Culture

The acute T-cell leukemia cell line (Jurkat cells) was a generous gift from Prof. H. Gehring (Department of Biochemistry, University of Zürich, Switzerland). The histiocytic lymphoma cell line (U-937 cells) and acute promyelocytic leukemia cell line (HL60 cells) were generous gifts from Dr. H. Le (Shanghai Center for Systems Biomedicine, Shanghai Jiao Tong University). The cells were cultured in RPMI-1640 medium (Invitrogen, Waltham, MA, USA) supplemented with 10% of fetal bovine serum (Lonza, Basel, Switzerland) and 1% of penicillin-streptomycin antibiotics. The cells were grown at 37 °C in a humidifier incubator under an atmosphere of 5% CO₂ and used for assay when they reached 80% of confluence in the dish.

2.4. MTS Assay

The viability of the cells was measured by MTS, a colorimetric assay which involves the biological reduction of the MTS reagent tetrazolium (3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxy-phenyl)-2-(4-sulfophenyl)-2H-tetrazolium). Cells were seeded in a 96-well plate at 5000 cells/well density. Bacillamide F was added at different concentrations (6.25–100 μ M) and the cells were treated for 24, 48 and 72 h. Absorbance was recorded at

490 nm in a plate reader (BioTek & Synergy2, Singapore). The number of cells in experiment is the means from three wells cells (biological replicates). All the experiments were repeated at least twice.

3. Results

3.1. Structural Identification of New Compound

The EtOAc extract of the bacterium was subjected to repeated Sephadex LH-20; the pure compound 1 (Figure 1b) was obtained by MPLC and semipreparative HPLC.

Compound 1 was obtained as yellow oil. It showed a molecular ion peak at m/z 383.1549 $[M + H]^+$ (calcd. for $C_{20}H_{22}N_4O_2S$, 383.1549) in the positive HR-ESI-MS spectrum, indicating twelve degrees of unsaturation. In the 1H -NMR spectrum (Table S1), five signals for H-atoms between δ_H 6.97 and 7.58 indicated the presence of one indole ring. Furthermore, one aromatic H-atom at δ_H 8.03 (s, 1 H), one CH group at δ_H 5.32 (dd, $J = 2.80, 7.60$, 1 H), five CH_2 groups at δ_H 3.05 (t, $J = 7.20$, 2 H), 3.68 (t, $J = 7.20$, 2 H), 2.23 (m, 2H), 2.03 (m, 2H) and 3.71~3.58 (m, 2H), and one Me group at 2.09 (s, 3 H) were observed. The ^{13}C -NMR and Distortionless Enhancement by Polarization Transfer (DEPT) spectra exhibited 20 C-atom signals ($1 \times CH_3$, $5 \times CH_2$, $7 \times CH$, $5 \times C$, and $2 \times C=O$), whose chemical-shift values and multiplicities confirmed the presence of an indole ring (δ_C 123.57, 113.04, 119.37, 119.65, 122.38, 112.24, 138.20, 128.77). The 1H - 1H COSY spectrum of H-9 (δ_H 3.05)/H-10 (δ_H 3.68), H-15 (δ_H 5.32)/H-16 (δ_H 2.23)/H-17 (δ_H 2.03)/H-18 (δ_H 3.71~3.58) revealed that two spin systems were present in compound 1 (Figures 2 and S1–S8 and Table S1).

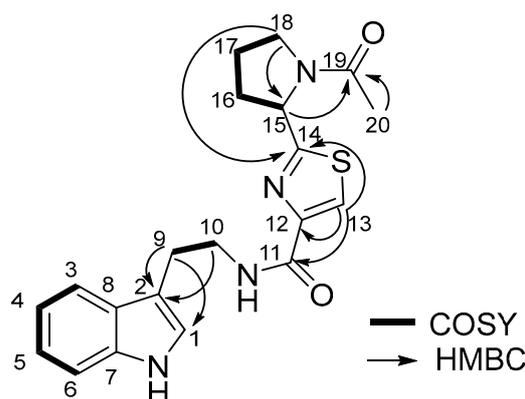


Figure 2. The key 2D NMR correlations of bacillamide F.

HMBC correlations from H-9 and H-10 to C-2 suggested that C-9 was connected to the indole ring. The other significant HMBC cross-peaks from H-13 to C-11, C-12, and C-14, from H-18 to C-14 and C-15, and from Me-20 to C-19 confirmed the connectivity of the pyrrolidine and thiazole rings as shown in Figure 2. Considering the molecular formula, the presence of a disubstituted thiazole ring was suspected and C-18 was connected to C-15 through a N-atom, while an acetyl group was anchored at the N-atom, further verified by HMBC correlation from H-15 to C-19. The compound 1 was identified as bacillamide F by analysis of its NMR spectral data as well as by comparison with the reported data [11]. Thus, the planar structure of compound 1 was established.

3.2. Plausible Biosynthesis Pathway of Bacillamide F

According to genome information, bacillamide F structure, and predicted biosynthetic pathway of bacillamide C and intermediate AlaCystthiazole [19,20], biosynthesis of bacillamide F has been predicted (Figure 3). NRPS domains, Oxidase domain, and AADC (aromatic L-amino acid decarboxylase) domain have been involved in the synthesis of bacillamide F. The enzyme-bound amino acids are combined through a nucleophilic attack of either the cysteine amino group or the heteroatom of the side chain onto the carbonyl C of proline, and a proton is abstracted through base catalysis, thereby enabling the at-

tack of either the cysteine side chain or the amino group onto the amide bond carbonyl. Intermediate hydroxylated thiazolidine has been synthesized, and then hydrated and dehydrogenated to produce thiazole-containing product ProCys^{thiazole}. Bacillamide F precursor has been synthesized by the C domain catalyzed peptide bond formation between the ProCys^{thiazole} and L-tryptamine decarboxylated by the AADC enzyme; subsequently, bacillamide F is synthesized after acetyl modifications.

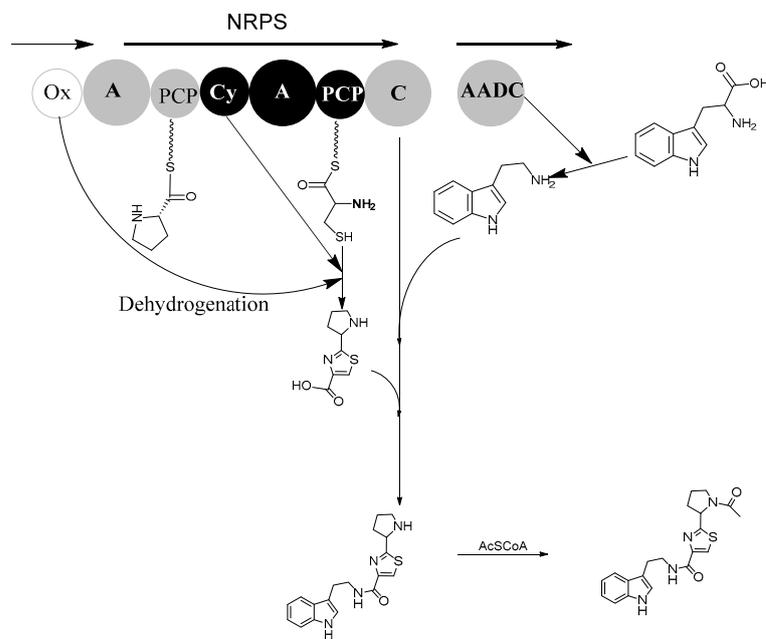


Figure 3. The plausible biosynthesis pathway of bacillamide F. The biosynthetic process of NRPS (EIM09914.1, bacitracin synthetase) domains, Ox (oxidase domain, EIM09913.1), and AADC (aromatic L-amino acid decarboxylase) domain were involved in the synthesis of bacillamide F. Each circle represents an NRPS enzymatic domain, from left to right: A, adenylation domain, Cy, cyclization domain, C, condensation domain, and PCP, peptidyl carrier protein domain.

In our previous study, a possible biosynthesis pathway for bacillamide C suggested that bacillamide C could be derived from alanine, cysteine, and tryptophan [19,21]. Consistent with this hypothesis, and followed by the optimization of the culture medium, experimental results showed that the independent addition of alanine, cysteine, and tryptophan had a positive effect on production of bacillamide C [22]. Based on the predicted biosynthetic pathway, it is possible that the pyrrolidine in compound 1 derived from L-proline (Figure 3). Therefore, the prediction of biosynthesis pathway for bacillamide F benefits the production of bacillamide F through optimized medium. In our other experiment, the increase of biosynthesis of bacillamide F was detected and quantified when the *B. atrophaeus* fed on L-proline.

3.3. Toxicity of Bacillamide F on Leukemia Cells

To determine the potential effect of the new compound on cancer cells, we performed a toxicity assay (MTS) on three different cell lines of leukemia. Bacillamide F inhibited the growth of Jurkat in a dose-dependent manner (Figure 4), showing cell survival less than 20% at 50 μM after 72 h of treatment. HL-60, acute promyelocytic leukemia, the subtype of AML, exhibited growth suppression in a dose- and time-dependent manner with a survival rate less than 30% at 12.50 μM after 72 h, while bacillamide F did not show any suppression on U937 as well as fibroblast MRC-5 or other solid tumor cells, such as pancreatic cancer cell lines PATU8988 and SW1990. Cytotoxicity of the inhibitor on Jurkat and HL60 was found to be decreased time-dependently with an IC₅₀ of 33.09 μM and 9.44 μM after the treatment of 72 h, an IC₅₀ of 37.16 μM and 14.13 μM after 48 h, and an IC₅₀ of 46.90 μM

and 21.82 μM after 24 h, respectively (Table S2). Intriguingly, bacillamide F showed a much less inhibitory effect on the proliferation of Jurkat than on HL60, e.g., at the concentration of 25 μM (Figure 4), an effect that we cannot currently explain.

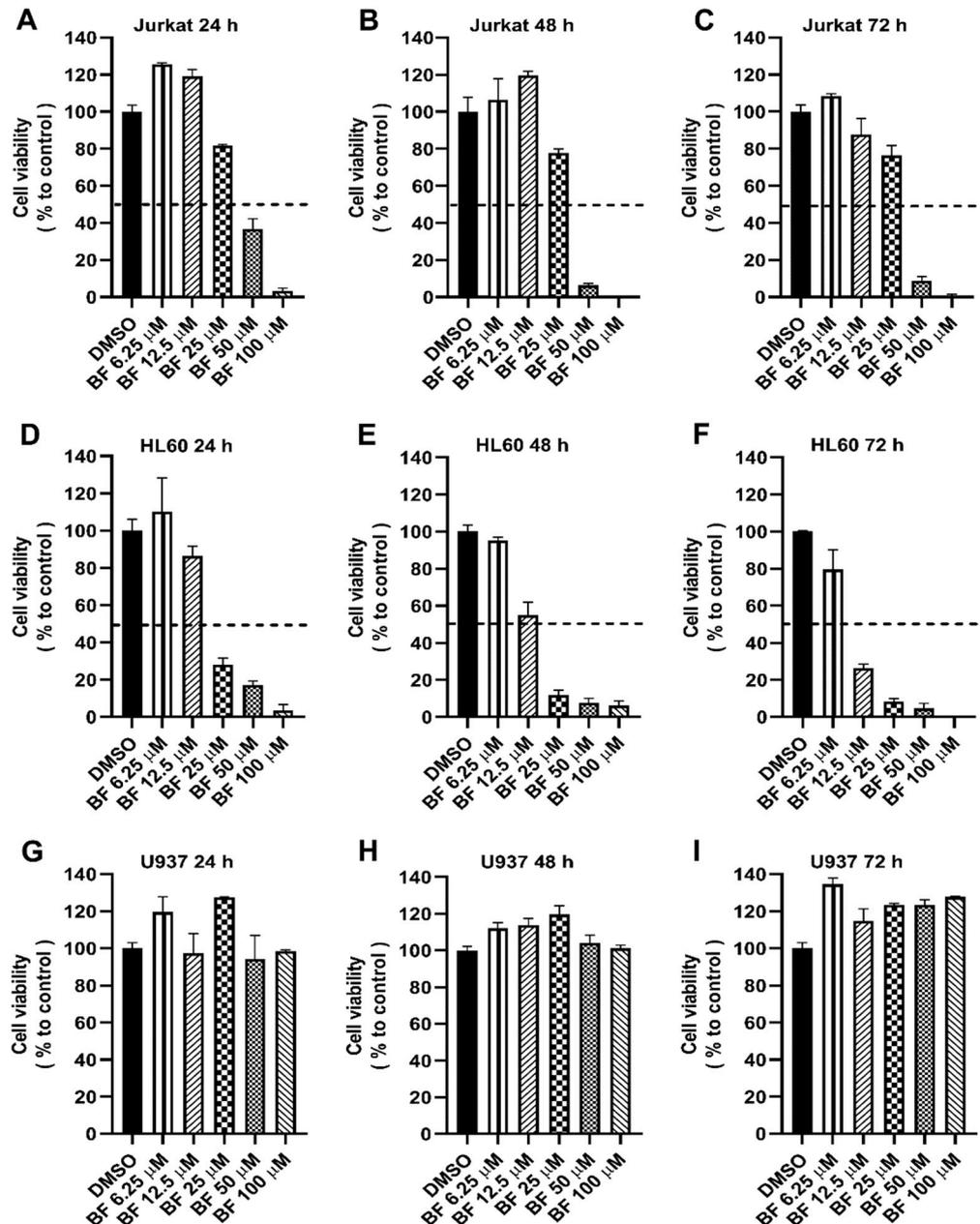


Figure 4. MTS assay on three different leukemia cell lines: Jurkat (A–C), HL60 (D–F), and U937 (G–I). Cell availability has been tested for 24, 48 and 72 h by incubating the cells in a 96-well plate with different concentrations of BF (Bacillamide F) (6.25–100 μM). The percentage of control samples (DMSO, 100%) is shown as means \pm SDs ($n = 3$, biological replicates). The experiments were repeated at least twice independently with similar results, and the representative data from one experiment are shown ($n = 3$, biological replicates).

Previous results have revealed that synthetic analogues of bacillamide type compounds with structural modifications on the amino or aminophenyl group of position 2 of the thiazole ring could suppress the proliferations of both solid tumor and leukemia cell lines [16]. In the present study, we show that a newly identified natural product bacillamide F, which has a substitution of a methyl-amino group on the same position of the

thiazole ring, can inhibit the proliferation of ALL and acute promyelocytic leukemia (APL) cell lines Jurkat and HL60 without affecting the proliferation of the solid tumor cell line (Table S2). The attractive indication of bacillamide F provides a pharmacological activity for this type of natural compound and offers a new drug lead for the treatment of ALL and APL leukemias.

4. Discussion

Bacillamides and analogues are natural products with various bioactivities, including algicidal activity [9,15,23], cytotoxic activity against three cancer cell lines (HCT-116, MDA-MD-231, and Jurkat cell lines), and anti-inflammatory activity [16]. Indole alkaloid derivatives, as the building blocks of bacillamides, have exhibited antibacterial and antifungal activity [24]. Thiazole amides, the component of bacillamides, have exhibited potential algicidal activity against harmful cyanobacterial algae [23]. Bacillamides, as a building block, have been found in cyclic peptides antibiotic zerkovamycin and argyrian A and B [25,26]. In this study, bacillamide F controlled the growth of Jurkat ALL and AML HL60 cell lines in a dose-dependent manner, rather than the acute histiocytic lymphoma U937 cell line (Figure 4 and Table S2), which is also considered to be a different subtype of AML [27]. Bacillamide F has a pyrrole structure (Figure 1b). Pyrrole is a simple heterocyclic and important building block for biologically active compounds [28]. The anticancer activity of bacillamide F has broadened the active range of the bacillamides family. Although bacillamide F shows a promising growth-inhibitory effect in two types of leukemia cell lines, further work needs to be performed in vitro and in vivo models to investigate its function and molecular mechanism in these cells.

The biosynthetic pathway of bacillamide has been predicted in our previous study [19–21]. The biosynthesis of bacillamide C has been hypothesized that decarboxylation from L-tryptophan to tryptamine could be performed before amidation by the downstream aromatic AADC to nrps gene cluster [21]. We have predicted that Oxidase encoded by oxidase gene located upstream of nrps gene cluster could catalyze AlaCys^{thiazoline} to AlaCys^{thiazole} [19]. The dehydrogenation activity of Oxidase has been further confirmed in the peptide intermediate while it is covalently attached to the NRPS [29]. In this study, addition of L-proline into a culture medium of *B. atrophaeus* C89 increased the production of bacillamide F (Figure 2). These results implied that broad substrate selectivity of NRPS A domain probably contributes to the biosynthesis of bacillamide analogues. In biosynthesis of bacillamide F as predicted, alanine, cysteine, tryptophan, and proline are precursors of bacillamide F (Figure 3). In future work, we will investigate how the concentration and proportion of various amino acid precursors affect the production of bacillamides and analogues.

5. Conclusions

In this study, a new member of the bacillamide family, bacillamide F, was obtained by a series of separation and purification methods. NMR data showed that the compound contained a novel thiazole alkaloid structure. The study also revealed that bacillamide F inhibited the growth of leukemia cell lines HL60 and Jurkat.

Supplementary Materials: The following supporting information can be downloaded at <https://www.mdpi.com/article/10.3390/biology11121712/s1>. Figure S1: HR-ESI-MS spectrum of bacillamide F; Figure S2: HR-ESI-MS-MS spectrum of bacillamide F; Figure S3: ¹H-NMR spectrum of bacillamide F; Figure S4: ¹³C-NMR spectrum of bacillamide F; Figure S5: DEPT spectrum of bacillamide F; Figure S6: COSY spectrum of bacillamide F; Figure S7: HMBC spectrum of bacillamide F; Figure S8: HSQC spectrum of bacillamide F; Table S1: ¹H and ¹³C NMR (600 and 150 MHz in methanol-d₄) data for compound 1; Table S2: IC₅₀ values of bacillamide F on different cancer cell lines.

Author Contributions: S.Z., Y.L., H.H. and C.Z. performed biosynthesis, isolation and identification experiment of the compound. G.C. characterized the anti-proliferation property of the compound on leukemia. F.Z. proposed the biosynthesis route of the compound. S.Z., G.C., F.Z., F.W. and Z.L. wrote the manuscript. F.Z. and F.W. designed the experiment. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by National Key Research and Development Program of China (2018YFA0901901), National Natural Science Foundation of China (81973230) and National Key Research and Development Program of China (2018YFC0310901, 2018YFC0310900).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: We express appreciation to Lei Feng and Jieli Wu from the Instrumental Analysis Center of Shanghai Jiao Tong University and Wei Zhang from Biocore of Shanghai Jiao Tong University for assistance in LC-MS, LCMS-MS, and NMR analysis. We thank Linquan Bai and Qianjin Kang for discussion of compound biosynthesis. We thank Heinz Gehring from Department of Biochemistry of University of Zürich, Switzerland and Huangying Le from Shanghai Center for Systems Biomedicine of Shanghai Jiao Tong University for Jurkat, HL60 and U-937 cell lines.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Arumuggam, N.; Melong, N.; Too, C.K.; Berman, J.N.; Rupasinghe, H.V. Phloridzin docosahexaenoate, a novel flavonoid derivative, suppresses growth and induces apoptosis in T-cell acute lymphoblastic leukemia cells. *Am. J. Cancer Res.* **2017**, *7*, 2452–2464. [[PubMed](#)]
2. Döhner, H.; Weisdorf, D.J.; Bloomfield, C.D. Acute myeloid leukemia. *N. Engl. J. Med.* **2015**, *373*, 1136–1152. [[CrossRef](#)] [[PubMed](#)]
3. Faderl, S.; Kantarjian, H.M.; Talpaz, M. Chronic myelogenous leukemia: Update on biology and treatment. *Oncology* **1999**, *13*, 169–180. [[PubMed](#)]
4. Rose-Inman, H.; Kuehl, D. Acute leukemia. *Hematol. Oncol. Clin. N. Am.* **2017**, *31*, 1011–1028. [[CrossRef](#)]
5. Chiorazzi, N.; Rai, K.R.; Ferrarini, M. Chronic lymphocytic leukemia. *N. Engl. J. Med.* **2005**, *352*, 804–815. [[CrossRef](#)]
6. Khoury, J.D.; Solary, E.; Abla, O.; Akkari, Y.; Alaggio, R.; Apperley, J.F.; Bejar, R.; Berti, E.; Busque, L.; Chen, J.K.C.; et al. The 5th edition of the World Health Organization classification of Haematolymphoid tumours: Myeloid and histiocytic/dendritic neoplasms. *Leuk* **2022**, *36*, 1703–1719. [[CrossRef](#)]
7. Abuelgasim, K.A.; Rehan, H.; Alsubaie, M.; Atwi, N.A.; Balwi, M.A.; Alshieban, S.; Almughairi, A. Coexistence of chronic myeloid leukemia and diffuse large b-cell lymphoma with antecedent chronic lymphocytic leukemia: A case report and review of the literature. *J. Med. Case. Rep.* **2018**, *12*, 64. [[CrossRef](#)]
8. Haybar, H.; Shahrabi, S.; Rezaeeyan, H.; Jodat, H.; Saki, N. Strategies to inhibit arsenic trioxide-induced cardiotoxicity in acute promyelocytic leukemia. *J. Cell. Physiol.* **2019**, *234*, 14500–14506. [[CrossRef](#)]
9. Jeong, S.Y.; Ishida, K.; Ito, Y.; Okada, S.; Murakami, M. Bacillamide, a novel algicide from the marine bacterium, *Bacillus* sp. SY-1, against the harmful dinoflagellate, *Cochlodinium Polykrikoides*. *Tetrahedron Lett* **2003**, *44*, 8005–8007. [[CrossRef](#)]
10. Socha, A.M.; Long, R.A.; Rowley, D.C. Bacillamides from a hypersaline microbial mat bacterium. *J. Nat. Prod.* **2007**, *70*, 1793–1795. [[CrossRef](#)]
11. Ivanova, V.; Kolarova, M.; Aleksieva, K.; Gräfe, U.; Dahse, H.M.; Laatsch, H. Microbiaeratin, a new natural indole alkaloid from a *Microbispora aerata* strain, isolated from Livingston Island, Antarctica. *Prep. Biochem. Biotechnol.* **2007**, *37*, 161–168. [[CrossRef](#)] [[PubMed](#)]
12. Yu, L.L.; Li, Z.Y.; Peng, C.S.; Li, Z.Y.; Guo, Y.W. Neobacillamide A, a novel thiazole-containing alkaloid from the marine bacterium *Bacillus vallismortis* C89, associated with South China Sea sponge *Dysidea avara*. *Helv. Chim. Acta* **2009**, *92*, 607–612. [[CrossRef](#)]
13. Omura, S.; Suzuki, Y.; Kitao, C.; Takahashi, Y.; Konda, Y. Isolation of a new sulfur-containing basic substance from a *Thermoactinomyces* species. *J. Antibiot.* **1975**, *28*, 609–610. [[CrossRef](#)] [[PubMed](#)]
14. Bloudoff, K.; Fage, C.D.; Marahiel, M.A.; Schmeing, T.M. Structural and mutational analysis of the nonribosomal peptide synthetase heterocyclization domain provides insight into catalysis. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, 95–100. [[CrossRef](#)] [[PubMed](#)]
15. Churro, C.; Alverca, E.; Sam-Bento, F.; Paulino, S.; Figueira, V.C.; Bento, A.J.; Prabhakar, S.; Lobo, A.M.; Calado, A.J.; Pereira, P. Effects of bacillamide and newly synthesized derivatives on the growth of cyanobacteria and microalgae cultures. *J. Appl. Phycol.* **2009**, *21*, 429–442. [[CrossRef](#)]
16. Kumar, S.; Aggarwal, R.; Kumar, V.; Sadana, R.; Patel, B.; Kaushik, P.; Kaushik, D. Solvent-free synthesis of bacillamide analogues as novel cytotoxic and anti-inflammatory agents. *Eur. J. Med. Chem.* **2016**, *123*, 718–726. [[CrossRef](#)]

17. de Santana, T.I.; Barbosa, M.d.O.; Gomes, P.A.T.d.M.; da Cruz, A.C.N.; da Silva, T.G.; Leite, A.C.L. Synthesis, anticancer activity and mechanism of action of new thiazole derivatives. *Eur. J. Med. Chem.* **2018**, *144*, 874–886. [[CrossRef](#)]
18. Winn, M.; Fyans, J.K.; Zhuo, Y.; Micklefield, J. Recent advances in engineering nonribosomal peptide assembly lines. *J. Nat. Prod. Rep.* **2016**, *33*, 317–347. [[CrossRef](#)]
19. Zhang, F.; Wang, Y.; Jiang, Q.; Chen, Q.; Karthik, L.; Zhao, Y.-L.; Li, Z. Substrate selection of adenylation domains for nonribosomal peptide synthetase (NRPS) in bacillamide C biosynthesis by marine *Bacillus atrophaeus* C89. *J. Ind. Microbiol. Biotechnol.* **2018**, *45*, 335–344. [[CrossRef](#)]
20. Zhang, F.; Mulati, N.; Wang, Y.; Li, Y.; Gong, S.; Karthik, L.; Sun, W.; Li, Z. Biosynthesis in vitro of bacillamide intermediate-heterocyclic AlaCysthiazole by heterologous expression of nonribosomal peptide synthetase (NRPS). *J. Biotechnol.* **2019**, *292*, 5–11. [[CrossRef](#)]
21. Yuwen, L.; Zhang, F.L.; Chen, Q.H.; Lin, S.J.; Zhao, Y.L.; Li, Z.Y. The role of aromatic L-amino acid decarboxylase in bacillamide C biosynthesis by *Bacillus atrophaeus* C89. *Sci. Rep.* **2013**, *3*, 1753. [[CrossRef](#)] [[PubMed](#)]
22. Jin, L.; Ma, W.; Peng, C.; Yin, Y.; Xu, B.; Zhang, F.; Guo, Y.; Li, Z. Bacillamide C production by the optimized cultivation of the *Bacillus atrophaeus* strain C89 associated with the South China Sea sponge *Dysidea avara*. *Process Biochem.* **2011**, *46*, 1153–1159. [[CrossRef](#)]
23. Wang, Y.; Liu, Q.; Wei, Z.; Liu, N.; Li, Y.; Li, D.; Jin, Z.; Xu, X. Thiazole amides, a novel class of algacides against freshwater harmful algae. *Sci. Rep.* **2018**, *8*, 8555. [[CrossRef](#)]
24. Vaca, J.; Salazar, F.; Ortiz, A.; Sansinenea, E. Indole alkaloid derivatives as building blocks of natural products from *Bacillus thuringiensis* and *Bacillus velezensis* and their antibacterial and antifungal activity study. *J. Antibiot.* **2020**, *73*, 798–802. [[CrossRef](#)]
25. Zhang, H.; Tomoda, H.; Tabata, N.; Oohori, M.; Shinose, M.; Takahashi, Y.; Omur, S. Isolation and structure of a new antibiotic viridomycin F produced by *Streptomyces* sp. K96-0188. *J. Antibiot.* **1999**, *52*, 61–64. [[CrossRef](#)]
26. Ferrari, P.; Vekey, K.; Galimberti, M.; Gallo, G.G.; Selva, E.; Zerilli, L.F. Antibiotics A21459 A and B, new inhibitors of bacterial protein synthesis. II. Structure elucidation. *J. Antibiot.* **1996**, *49*, 150–154. [[CrossRef](#)]
27. Bohlander, S.K.; Muschinsky, V.; Schrader, K.; Siebert, R.; Schlegelberger, B.; Harder, L.; Schemmel, V.; Fonatsch, C.; Ludwig, W.D.; Hiddemann, W.; et al. Molecular analysis of the CALM/AF10 fusion: Identical rearrangements in acute myeloid leukemia, acute lymphoblastic leukemia and malignant lymphoma patients. *Leukemia* **2000**, *14*, 93–99. [[CrossRef](#)]
28. Gholap, S.S. Pyrrole: An emerging scaffold for construction of valuable therapeutic agents. *Eur. J. Med. Chem.* **2016**, *110*, 13–31. [[CrossRef](#)]
29. Fortinez, C.M.; Bloudoff, K.; Harrigan, C.; Sharon, I.; Strauss, M.; Schmeing, T.M. Structures and function of a tailoring oxidase in complex with a nonribosomal peptide synthetase module. *Nat. Commun.* **2022**, *13*, 548. [[CrossRef](#)]