

Table S1. Sample and clone codes of plant accessions that were used in the various photosynthesis fluorescence experiments

Sample	Plant ID	ϕ_{PSII}	LC	KC	OJIP	Sample	Plant ID	ϕ_{PSII}	LC	KC	OJIP
2x, 10.0 h						S4-16	LH1406030B5-05	x			x
C2-12	J10xJ30/03	x				S4-2	LH1406030B1-04	x		x	
C2-13	J10xJ30/11	x	x	x	x	S4-20	LH1406030B5-13	x			x
C2-15	J20xJ2/16	x	x	x	x	S4-21	LH1406030B5-18	x	x	x	
C2-19	J24xJ22/12	x	x	x	x	S4-22	LH1406030B5-20	x			
C2-21	F3xJ6/25	x	x	x	x	S4-23	LH1406030B4-10	x	x	x	x
C2-24	J10xJ14/18	x		x	x	S4-24	LH1406030G1-8	x			x
C2-25	J6xF3/19		x	x	x	S4-25	LH1406030G1-16		x	x	x
C2-26	J10xJ30/05	x	x			S4-26	LH1406030G1-18		x	x	
C2-27	J6xF3/14		x	x	x	S4-27	LH1406030B5-04	x			
C2-4	F3xJ6/28	x				S4-4	LH1406030B2-01	x			x
C2-7	J6xF3/14	x	x	x	x	S4-5	LH1406030B2-07	x	x	x	
C2-8	J6xF7/12	x	x	x	x	S4-6	LH1406030B4-01		x	x	x
C2-9	J6xF7/14	x	x	x	x	S4-8	LH1406030B4-11	x	x	x	
2x, 16.5 h						6x_29, 10.0 h					
S2-1	F3xJ6/01	x	x	x	x	C6-1	29/15-3N/02	x	x		
S2-18	J24xJ22/03	x	x	x	x	C6-12	29/15-5K/21	x	x	x	x
S2-2	F3xJ6/04	x	x	x	x	C6-5	29/15-5K/02	x	x	x	x
S2-21	F3xJ6/19	x	x	x	x	C6-6	29/15-5K/05	x	x	x	x
S2-23	F10xJ3/03	x	x	x	x	C6-7	29/15-5K/09	x	x	x	x
S2-24	J6xF3/06	x	x	x	x	6x_29, 16.5 h					
S2-25	J10xJ14/09	x				S6-39	29/15-5K/20	x	x	x	x
S2-27	J24xJ22/09	x				S6-9	29/15-5K/06	x	x	x	x
S2-3	F3xJ6/05	x	x	x	x	S6-1	29/15-5K/03	x	x	x	x
S2-6	J6xF3/02	x	x	x	x	6x_35, 10.0 h					
S2-7	J6xF3/05	x	x	x	x	C6-15	35/28-4*/26	x	x	x	x
S2-9	J6xF7/08	x	x	x	x	C6-16	35/28-4*/28	x	x	x	x
4x, 10.0 h						C6-22	35/28-4a/16	x			x
C4-11	LH1406030B4-08	x	x	x	x	C6-23	35/28-4*/27		x	x	x
C4-13	LH1406030B4-16	x	x	x	x	C6-25	35/28-4a/22	x	x	x	x
C4-15	LH1406030B4-18	x	x	x	x	C6-33	35/28-4Q/82	x	x		x
C4-19	LH1406030B5-07	x	x	x	x	6x_35, 16.5 h					
C4-21	LH1406030B5-16	x	x	x	x	S6-17	35/28-4*/03	x	x	x	x
C4-22	LH1406030B5-17	x	x	x	x	S6-19	35/28-4*/18	x	x	x	x
C4-23	LH1406030B5-19	x	x	x	x	S6-21	35/28-4*/24		x	x	x
C4-26	LH4B005	x				S6-22	35/28-4*/22			x	
C4-5	LH1406030B2-04	x	x	x	x	S6-24	35/28-4*/26		x	x	x
C4-8	LH1406030B4-02	x	x	x	x	S6-25	35/28-4*/40		x	x	x
C4-9	LH1406030B4-05	x	x	x	x	S6-29	35/28-4Q/27	x		x	
4x, 16.5 h						S6-33	35/28-4Q/28	x		x	
S4-11	LH1406030B4-19	x	x	x	x	S6-37	35/28-4a/40			x	x
S4-14	LH1406030B2-02	x	x	x	x						

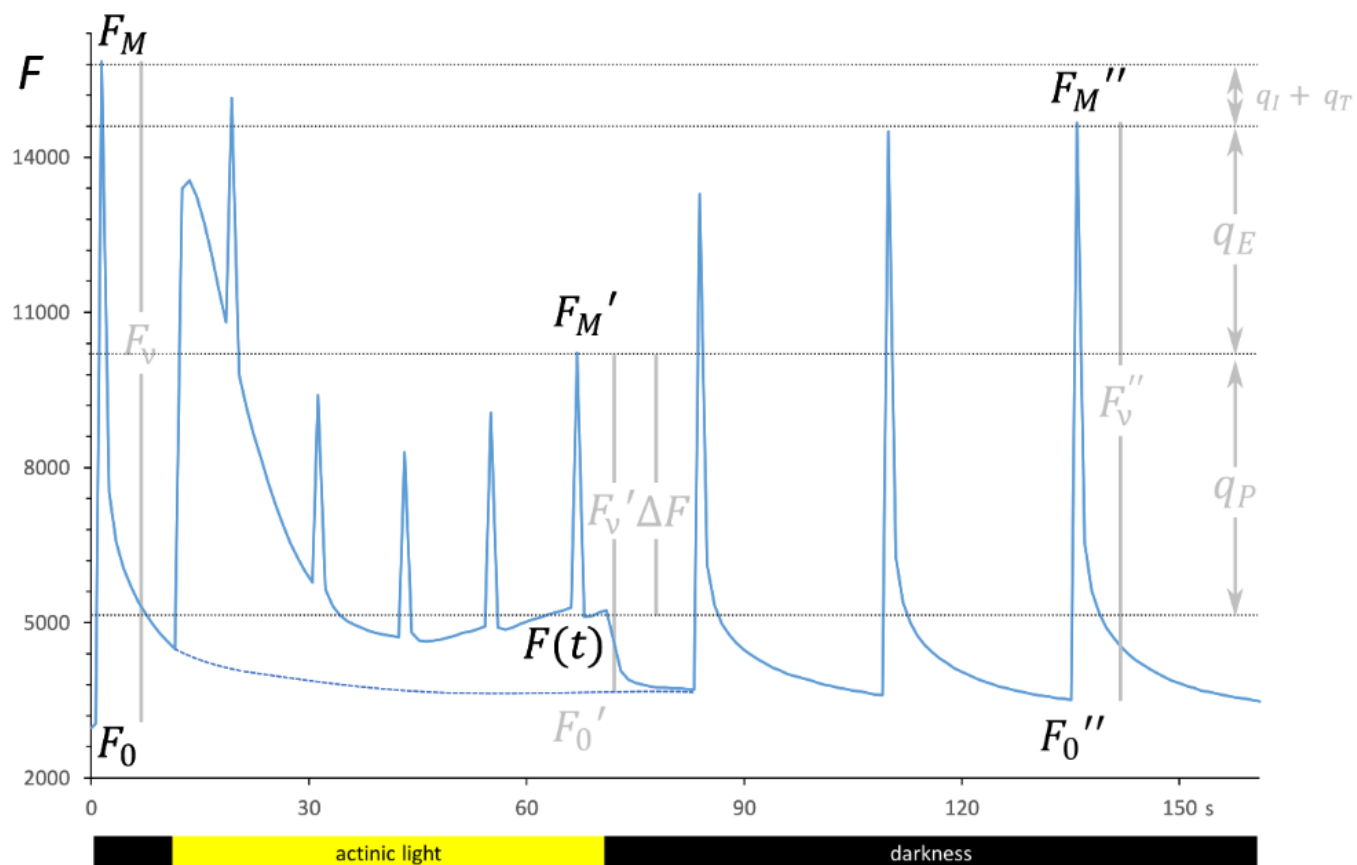


Figure S1: Exemplary Kautsky curve of KC experiments, indication of fluorescence parameters and coefficients, for details see Materials and Methods and Table S2.

Table S2. Calculation and definition of photosynthesis coefficients (Strasser and Govindjee, 1992; Strasser et al., 2004; Baker, 2008; Tsimilli-Michael and Strasser, 2013; Lazár, 2015; Rusaczonek et al., 2015)

Quenching of induced fluorescence analysis	
F_0	Saturation pulse, baseline fluorescence
F_0'	Measuring pulse baseline fluorescence during actinic light phase
F_0''	Baseline fluorescence during subsequent dark phase
F_M	Saturation pulse, maximum fluorescence
F_M'	Saturation pulse, maximum fluorescence during actinic light phase
F_M''	Saturation pulse, maximum F during subsequent dark light phase
$F(t)$	Baseline F during actinic light phase
$F_V = F_M - F_0$	Variable F after adapted dark phase
$F_V' = F_M' - F_0'$	Variable F in actinic light phase
$F_V'' = F_M'' - F_0''$	Variable F in subsequent dark phase
$\Delta F = F_M' - F(t)$	Difference between F_V' and $F(t)$
$\phi_{max} = F_V / F_M$	PSII maximum quantum yield after dark adaptation
$\phi_{PSII} = F_V' / F_M'$	PSII potential quantum yield
$rETR = \phi_{PSII} \times PPFD \times 0.5$	Relative electron transport rate; PPFD: Photosynthetic photon flux density
$PQ = (F_M / F(t)) - (F_M / F_M')$	Photochemical quenching
$NPQ = (F_M / F_M') / F_M'$	Non-photochemical quenching
$q_E = (F_M'' - F_V') / F_M''$	Energy-dependent non-photochemical quenching
$q_I = 1 - (F_V'' / F_V)$	Photoinhibitory non-photochemical quenching coefficient
$q_P = \Delta F / (F_M' - F_0')$	Photochemical quenching coefficient, correlates nonlinearly to the fraction of “open” PSII centers
$q_L = ((F_M' - F(t)) / (F_M' - F_0')) \times (F_0' / F(t))$	Photochemical quenching coefficient, correlates linearly to the fraction of “open” PSII centers (lake model)
OJIP transient analysis	
$F_O (= F_0)$	F intensity at 50 μs
F_I	F intensity at J-step (at 2ms)
F_L	F intensity at I-step (at 30 ms)
$F_P (= F_m)$	Maximum F intensity
$F_v = F_P - F_O$	Maximal variable F
F_P / F_O	F ratio
$Area$	Area between fluorescence curve and F_P
$M_0 = 4 (F_{300\mu s} - F_O) / (F_M - F_O)$	Approximated initial slope of the O–J fluorescence transient (in ms ⁻¹)
$S_M = Area / (F_P - F_O)$	The normalized area below the OJIP curve until F_P
$V_I = (F_I - F_O) / (F_P - F_O)$	Relative fluorescence during the I-step
$V_J = (F_J - F_O) / (F_P - F_O)$	Relative fluorescence during the J-step
$N = S_M \times M_0 \times (1 / V_J)$	Turn-over number of Q_A
$\phi_{P_0} = F_V / F_P = \phi_{max}$	Maximum quantum yield of PSII
$V_I = (F_I - F_O) / (F_M - F_O)$	Relative variable fluorescence at the I-step
$V_J = (F_J - F_O) / (F_M - F_O)$	Relative variable fluorescence at the J-step
$\psi_0 = 1 - V_J$	Probability at t = 0 that an electron moves into the electron transport chain beyond Q_A^-
Specific energy fluxes	
$ABS/RC = (M_0 / V_J) / \phi_{P_0}$	Absorption flux per reaction center (RC) (apparent antenna size of an active PSII)
$TR_0/RC = M_0 / V_J$	Trapped energy flux per RC
$ET_0/RC = (M_0 / V_J) \times \psi_0$	Electron transport flux per RC
$DI_0/RC = ABS/RC - TR_0/RC$	Dissipated energy flux per RC
Performance index	
$PI_{ABS} = RC/ABS \times (\phi_{P_0} / 1 - \phi_{P_0}) \times (\psi_0/1 - \psi_0)$	Performance index on absorption basis related to the overall photosynthetic activity of PSII

References:

Baker, N.R. (2008). Chlorophyll fluorescence: a probe of photosynthesis in vivo. *Annu. Rev. Plant Biol.* 59, 89-113.

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Table S3. Summary statistics and 95% Tukey multiple range test of ϕ_{PSII} and ϕ_{MAX} among cytotypes exposed to different photoperiods (SE, standard error; CL, 95% confidential limits).

Parameter	Cytotype	Photoperiod (h)	Mean	SE	<i>n</i>	Lower CL	upper CL	Group
ϕ_{PSII}	2x	10.0	0.720	0.006	665	0.707	0.731	<i>b</i>
	2x	16.5	0.710	0.006	665	0.698	0.722	<i>b</i>
	4x	10.0	0.707	0.006	665	0.694	0.719	<i>b</i>
	4x	16.5	0.697	0.006	665	0.685	0.709	<i>ab</i>
	6x_29	10.0	0.720	0.010	665	0.699	0.739	<i>b</i>
	6x_29	16.5	0.722	0.012	665	0.698	0.745	<i>b</i>
	6x_35	10.0	0.617	0.010	665	0.597	0.636	<i>a</i>
	6x_35	16.5	0.676	0.009	665	0.659	0.693	<i>a</i>
ϕ_{MAX}	2x	10.0	0.821	0.008	170	0.804	0.837	<i>b</i>
	2x	16.5	0.815	0.009	170	0.797	0.832	<i>b</i>
	4x	10.0	0.823	0.009	170	0.805	0.839	<i>b</i>
	4x	16.5	0.818	0.008	170	0.802	0.833	<i>b</i>
	6x_29	10.0	0.826	0.012	170	0.800	0.849	<i>b</i>
	6x_29	16.5	0.808	0.017	170	0.774	0.839	<i>b</i>
	6x_35	10.0	0.705	0.015	170	0.674	0.735	<i>a</i>
	6x_35	16.5	0.671	0.014	170	0.643	0.697	<i>a</i>

Table S4. Summary statistic and 95% Duncan multiple range test of coefficients from transient fluorescence analyses (OJIP) of cytotypes that were exposed to two photoperiods (SD, standard deviation).

Coeff.	Cytotype	Photoper. (h)	Mean	SD	<i>n</i>	Min	Max	Group
<i>ABS/RC</i>	2x	10.0	1.902	0.181	10	1.662	2.264	<i>c</i>
	2x	16.5	2.090	0.154	10	1.926	2.400	<i>bc</i>
	4x	10.0	1.987	0.162	10	1.827	2.376	<i>c</i>
	4x	16.5	1.885	0.290	11	1.383	2.380	<i>c</i>
	6x_29	10.0	2.017	0.099	4	1.869	2.083	<i>c</i>
	6x_29	16.5	1.931	0.295	3	1.596	2.154	<i>c</i>
	6x_35	10.0	2.714	0.756	7	2.000	4.000	<i>b</i>
	6x_35	16.5	3.866	1.399	7	2.000	5.870	<i>a</i>
<i>DI₀/RC</i>	2x	10.0	0.336	0.052	10	0.268	0.412	<i>c</i>
	2x	16.5	0.403	0.054	10	0.350	0.498	<i>c</i>
	4x	10.0	0.357	0.045	10	0.313	0.460	<i>c</i>
	4x	16.5	0.354	0.087	11	0.224	0.485	<i>c</i>
	6x_29	10.0	0.357	0.028	4	0.319	0.384	<i>c</i>
	6x_29	16.5	0.350	0.078	3	0.260	0.395	<i>c</i>
	6x_35	10.0	0.867	0.551	7	0.348	2.000	<i>b</i>
	6x_35	16.5	1.545	0.905	7	0.504	2.974	<i>a</i>
<i>TR₀/RC</i>	2x	10.0	1.566	0.134	10	1.393	1.853	<i>c</i>
	2x	16.5	1.688	0.102	10	1.576	1.902	<i>c</i>
	4x	10.0	1.631	0.120	10	1.507	1.917	<i>c</i>
	4x	16.5	1.572	0.251	11	1.159	2.000	<i>c</i>
	6x_29	10.0	1.659	0.073	4	1.550	1.699	<i>c</i>
	6x_29	16.5	1.582	0.219	3	1.337	1.759	<i>c</i>
	6x_35	10.0	2.000	0.000	7	2.000	2.000	<i>b</i>
	6x_35	16.5	2.393	0.434	7	2.000	2.985	<i>a</i>
<i>ET₀/RC</i>	2x	10.0	0.964	0.067	10	0.866	1.097	<i>b</i>
	2x	16.5	0.993	0.047	10	0.917	1.072	<i>b</i>
	4x	10.0	0.981	0.057	10	0.887	1.047	<i>b</i>
	4x	16.5	0.886	0.081	11	0.699	0.985	<i>b</i>
	6x_29	10.0	0.965	0.071	4	0.910	1.067	<i>b</i>
	6x_29	16.5	0.952	0.079	3	0.862	1.011	<i>b</i>
	6x_35	10.0	0.987	0.023	7	0.942	1.000	<i>b</i>
	6x_35	16.5	1.301	0.331	7	0.949	1.794	<i>a</i>
<i>PI_{ABS}</i>	2x	10.0	4.121	1.014	10	2.666	5.928	<i>a</i>
	2x	16.5	3.005	0.786	10	1.986	3.997	<i>a</i>
	4x	10.0	3.696	1.044	10	2.073	4.965	<i>a</i>
	4x	16.5	3.990	2.502	11	1.000	9.670	<i>a</i>
	6x_29	10.0	3.267	0.650	4	2.547	3.919	<i>a</i>
	6x_29	16.5	3.909	1.715	3	2.622	5.856	<i>a</i>
	6x_35	10.0	1.410	0.949	7	0.325	3.000	<i>b</i>
	6x_35	16.5	0.787	0.645	7	0.224	2.000	<i>b</i>

Table S5. Summary statistics and 95% Duncan multiple range test of relative electron transport rates (*rETR*) among cytotypes that were exposed to different photoperiods in increasing PPFD intensities (SD, standard deviation).

PPFD ($\mu\text{mol m}^{-2} \text{ sec}^{-1}$)	Cytotype	Photoperiod (h)	Mean	SD	<i>n</i>	Min	Max	Group
10	2x	10.0	5.200	0.499	10	4.200	5.900	<i>bc</i>
	2x	16.5	5.720	0.361	10	5.300	6.400	<i>a</i>
	4x	10.0	4.860	0.556	10	4.000	5.700	<i>cd</i>
	4x	16.5	5.578	0.331	9	5.200	6.100	<i>ab</i>
	6x_29	10.0	5.160	0.288	5	4.900	5.600	<i>bc</i>
	6x_29	16.5	5.167	0.306	3	4.900	5.500	<i>bc</i>
	6x_35	10.0	4.500	0.339	5	4.100	4.900	<i>d</i>
	6x_35	16.5	4.460	0.416	5	4.000	4.900	<i>d</i>
20	2x	10.0	10.860	1.108	10	9.000	12.800	<i>ab</i>
	2x	16.5	11.640	0.595	10	10.800	12.800	<i>a</i>
	4x	10.0	10.440	0.947	10	9.000	12.200	<i>b</i>
	4x	16.5	11.622	0.484	9	10.800	12.400	<i>a</i>
	6x_29	10.0	10.720	0.502	5	10.400	11.600	<i>ab</i>
	6x_29	16.5	10.800	0.346	3	10.400	11.000	<i>ab</i>
	6x_35	10.0	9.040	0.669	5	8.400	10.000	<i>c</i>
	6x_35	16.5	9.120	0.944	5	8.000	10.200	<i>c</i>
50	2x	10.0	27.950	2.852	10	22.500	32.000	<i>ab</i>
	2x	16.5	29.500	1.683	10	27.000	32.500	<i>a</i>
	4x	10.0	26.600	2.492	10	23.000	31.000	<i>b</i>
	4x	16.5	29.000	1.031	9	27.500	31.000	<i>ab</i>
	6x_29	10.0	27.000	1.225	5	26.000	29.000	<i>ab</i>
	6x_29	16.5	27.833	0.764	3	27.000	28.500	<i>ab</i>
	6x_35	10.0	22.300	1.789	5	20.500	25.000	<i>c</i>
	6x_35	16.5	23.000	2.739	5	19.500	26.000	<i>c</i>
100	2x	10.0	49.500	6.786	10	36.000	58.000	<i>a</i>
	2x	16.5	52.000	4.422	10	44.000	59.000	<i>a</i>
	4x	10.0	47.000	5.121	10	39.000	55.000	<i>a</i>
	4x	16.5	51.000	2.550	9	47.000	55.000	<i>a</i>
	6x_29	10.0	47.600	1.673	5	45.000	49.000	<i>a</i>
	6x_29	16.5	49.333	1.528	3	48.000	51.000	<i>a</i>
	6x_35	10.0	39.200	2.683	5	37.000	43.000	<i>b</i>
	6x_35	16.5	41.000	5.339	5	35.000	47.000	<i>b</i>
300	2x	10.0	88.800	21.872	10	51.000	117.000	<i>ab</i>
	2x	16.5	90.900	13.932	10	57.000	102.000	<i>ab</i>
	4x	10.0	67.500	9.618	10	51.000	81.000	<i>c</i>
	4x	16.5	75.000	13.332	9	54.000	90.000	<i>bc</i>
	6x_29	10.0	88.800	9.149	5	78.000	99.000	<i>ab</i>
	6x_29	16.5	94.000	1.732	3	93.000	96.000	<i>abc</i>
	6x_35	10.0	70.200	6.573	5	60.000	78.000	<i>c</i>
	6x_35	16.5	76.200	11.345	5	63.000	87.000	<i>a</i>
500	2x	10.0	106.000	30.074	10	60.000	145.000	<i>ab</i>
	2x	16.5	103.500	18.265	10	60.000	120.000	<i>ab</i>
	4x	10.0	71.000	11.972	10	50.000	90.000	<i>d</i>
	4x	16.5	78.889	17.989	9	50.000	100.000	<i>cd</i>
	6x_29	10.0	113.000	12.550	5	100.000	125.000	<i>ab</i>
	6x_29	16.5	116.667	2.887	3	115.000	120.000	<i>a</i>
	6x_35	10.0	91.000	7.416	5	80.000	100.000	<i>bcd</i>
	6x_35	16.5	98.000	13.038	5	80.000	110.000	<i>abc</i>

Table S6. Summary statistic and 95% Duncan multiple range test of coefficients calculated from Kautsky curve experiments for different cytotypes that were exposed to two photoperiods (SD, standard deviation).

Coeff.	Cytotype	Photoper. (h)	Mean	SD	<i>n</i>	Min	Max	Group
<i>NPQ</i>	2x	10.0	0.654	0.213	10	0.334	0.951	<i>cd</i>
	2x	16.5	0.990	0.347	10	0.606	1.729	<i>abc</i>
	4x	10.0	1.102	0.390	10	0.290	1.782	<i>ab</i>
	4x	16.5	1.241	0.376	10	0.886	2.007	<i>a</i>
	6x_29	10.0	0.451	0.088	4	0.342	0.549	<i>d</i>
	6x_29	16.5	0.753	0.173	3	0.641	0.952	<i>bcd</i>
	6x_35	10.0	1.164	0.451	4	0.631	1.699	<i>ab</i>
	6x_35	16.5	1.237	0.381	9	0.716	1.897	<i>a</i>
<i>q_E</i>	2x	10.0	0.450	0.178	10	0.161	0.762	<i>bc</i>
	2x	16.5	0.728	0.289	10	0.412	1.361	<i>ab</i>
	4x	10.0	0.698	0.244	10	0.171	1.054	<i>ab</i>
	4x	16.5	0.869	0.269	10	0.483	1.415	<i>a</i>
	6x_29	10.0	0.350	0.108	4	0.218	0.480	<i>c</i>
	6x_29	16.5	0.605	0.149	3	0.503	0.776	<i>abc</i>
	6x_35	10.0	0.801	0.225	4	0.511	1.045	<i>a</i>
	6x_35	16.5	0.889	0.180	9	0.582	1.119	<i>a</i>
<i>q_I</i>	2x	10.0	0.172	0.057	10	0.118	0.304	<i>ab</i>
	2x	16.5	0.178	0.057	10	0.111	0.312	<i>ab</i>
	4x	10.0	0.263	0.060	10	0.155	0.350	<i>a</i>
	4x	16.5	0.229	0.052	10	0.136	0.306	<i>a</i>
	6x_29	10.0	0.120	0.015	4	0.105	0.139	<i>b</i>
	6x_29	16.5	0.131	0.005	3	0.126	0.134	<i>b</i>
	6x_35	10.0	0.258	0.171	4	0.120	0.506	<i>a</i>
	6x_35	16.5	0.256	0.085	9	0.132	0.363	<i>a</i>
<i>PQ</i>	2x	10.0	1.263	0.238	10	1.008	1.753	<i>abc</i>
	2x	16.5	1.425	0.276	10	0.808	1.700	<i>a</i>
	4x	10.0	1.093	0.296	10	0.507	1.458	<i>bcd</i>
	4x	16.5	1.358	0.218	10	0.897	1.639	<i>ab</i>
	6x_29	10.0	0.789	0.248	4	0.447	0.980	<i>e</i>
	6x_29	16.5	1.004	0.147	3	0.905	1.173	<i>cde</i>
	6x_35	10.0	0.856	0.214	4	0.580	1.032	<i>de</i>
	6x_35	16.5	0.869	0.111	9	0.688	1.008	<i>de</i>
<i>q_P</i>	2x	10.0	0.601	0.065	10	0.528	0.708	<i>a</i>
	2x	16.5	0.600	0.050	10	0.483	0.656	<i>a</i>
	4x	10.0	0.486	0.075	10	0.352	0.580	<i>bc</i>
	4x	16.5	0.569	0.049	10	0.465	0.618	<i>ab</i>
	6x_29	10.0	0.447	0.095	4	0.312	0.518	<i>c</i>
	6x_29	16.5	0.509	0.023	3	0.488	0.535	<i>abc</i>
	6x_35	10.0	0.569	0.149	4	0.358	0.706	<i>ab</i>
	6x_35	16.5	0.573	0.069	9	0.414	0.638	<i>ab</i>
<i>q_L</i>	2x	10.0	0.299	0.097	10	0.198	0.523	<i>ab</i>
	2x	16.5	0.317	0.057	10	0.224	0.404	<i>ab</i>
	4x	10.0	0.226	0.069	10	0.097	0.335	<i>bc</i>
	4x	16.5	0.306	0.059	10	0.222	0.438	<i>ab</i>
	6x_29	10.0	0.160	0.057	4	0.083	0.207	<i>c</i>
	6x_29	16.5	0.230	0.023	3	0.211	0.255	<i>bc</i>
	6x_35	10.0	0.397	0.202	4	0.129	0.619	<i>a</i>
	6x_35	16.5	0.402	0.101	9	0.220	0.489	<i>a</i>