

Doxorubicin-loaded Silica Nanocomposites for Cancer Treatment

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Silica nanoparticles (SiNPs) were characterized using dynamic light scattering (DLS) on a Malvern Zetasizer Nano device (Malvern Instruments, Great Britain). For DLS studies, nanoparticles were diluted in deionized water to a concentration of 250 $\mu\text{g/mL}$.

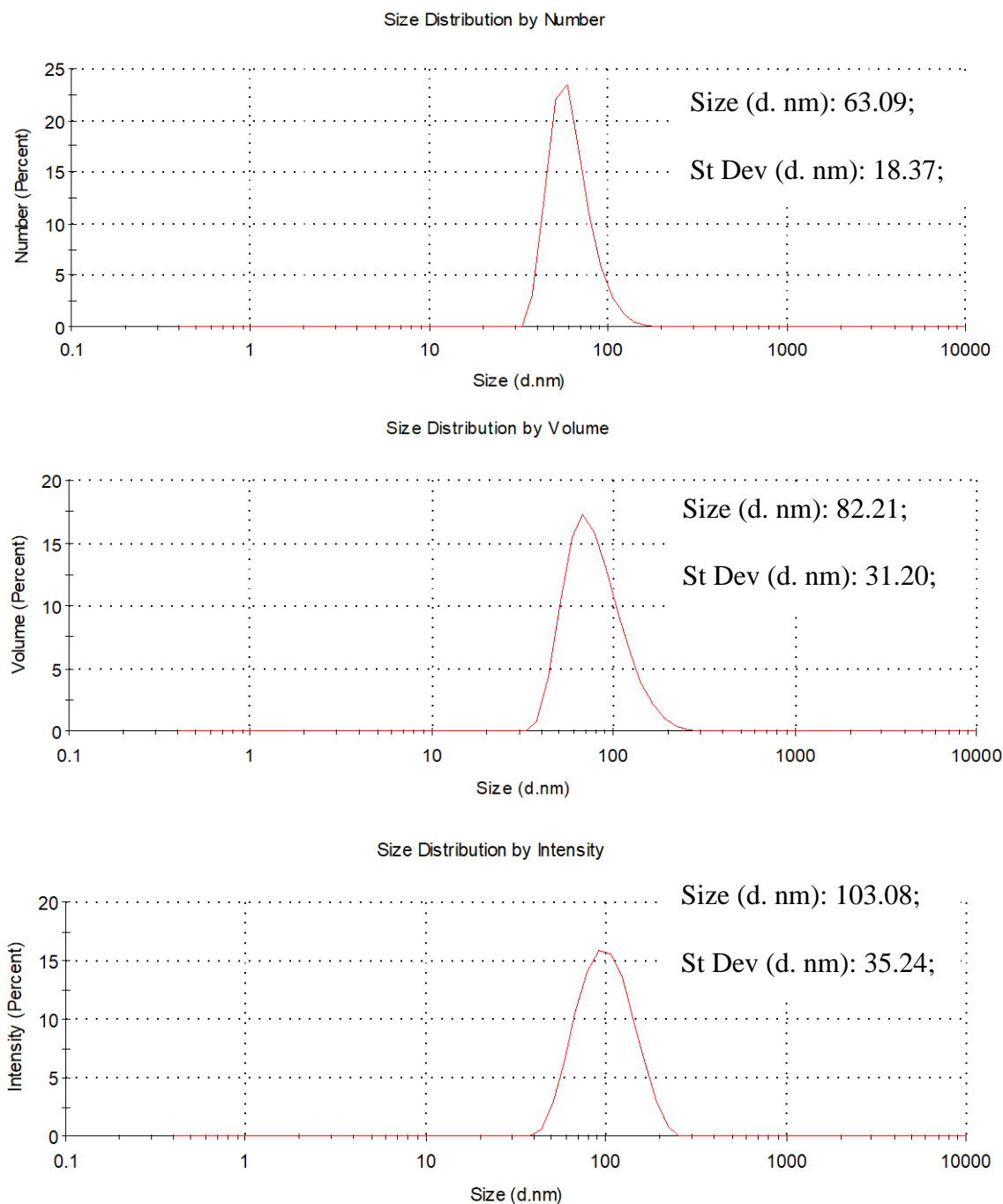


Figure S1. DLS size distribution data (Number, Volume, and Intensity) for method Rao et al. [68] with 0.018 M TEOS. The particle size by Number is 63 ± 1 nm ($\text{PDI} = 0.09 \pm 0.01$).

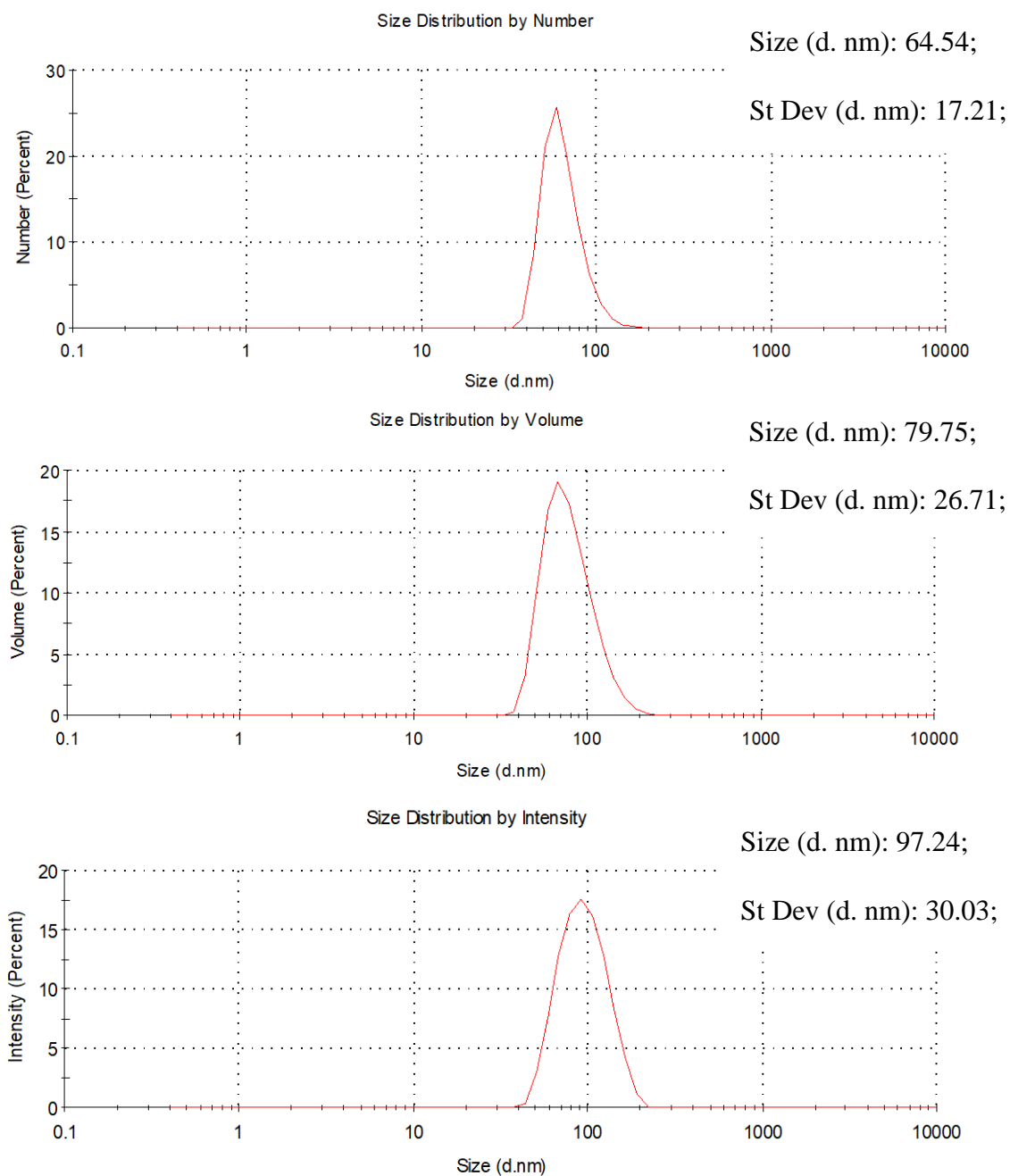


Figure S2. DLS size distribution data (Number, Volume, and Intensity) for method Rao et al. [68] with 0.018 M TEOS in 50 mL synthesis conditions. The particle size by Number is 64 ± 2 nm ($\text{PDI} = 0.10 \pm 0.01$).

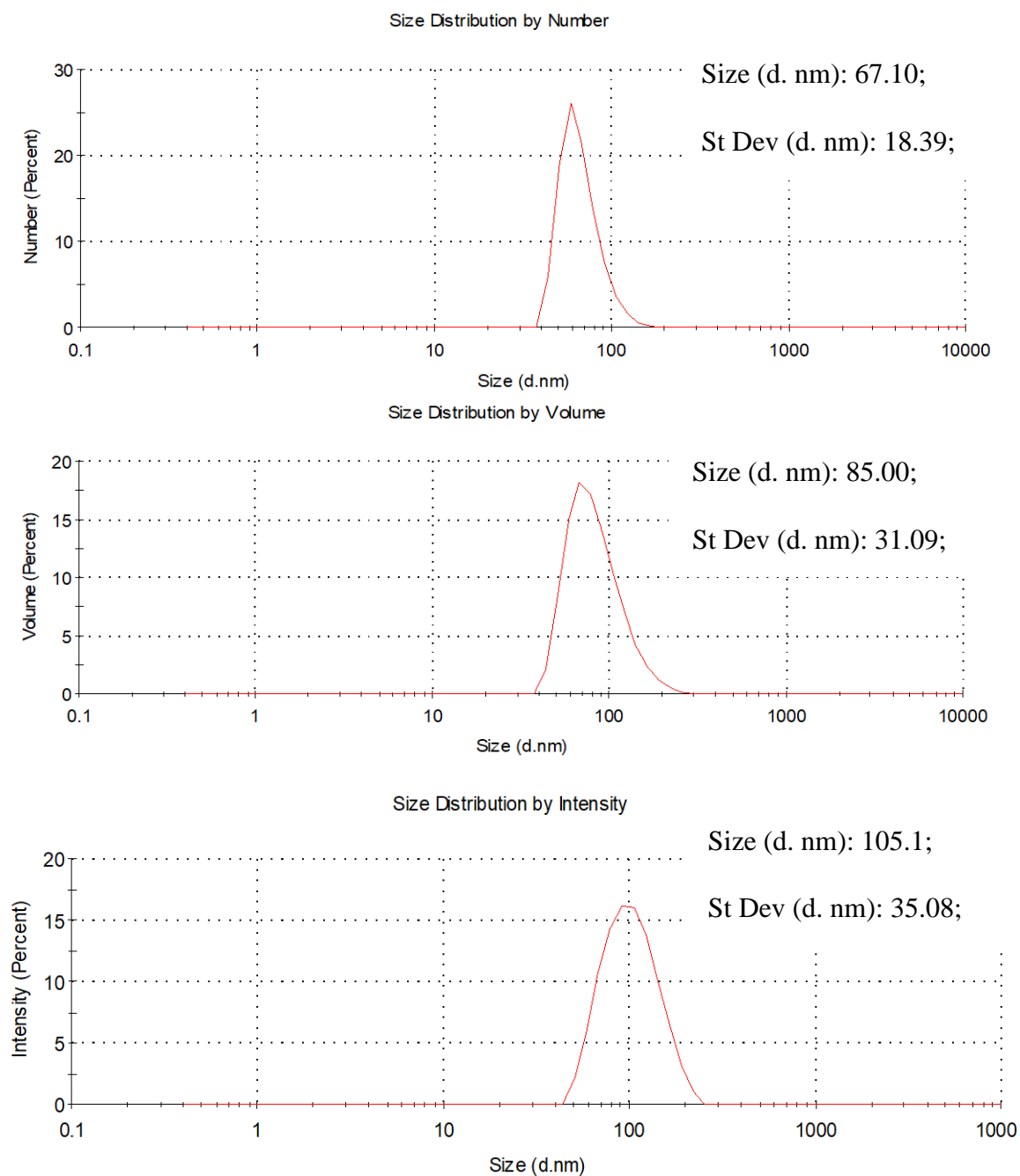


Figure S3. DLS size distribution data (Number, Volume, and Intensity) for method Rao et al. [68] with 0.018 M TEOS after one year storage. The particle size by Number is 67 ± 5 nm ($\text{PDI} = 0.10 \pm 0.01$).

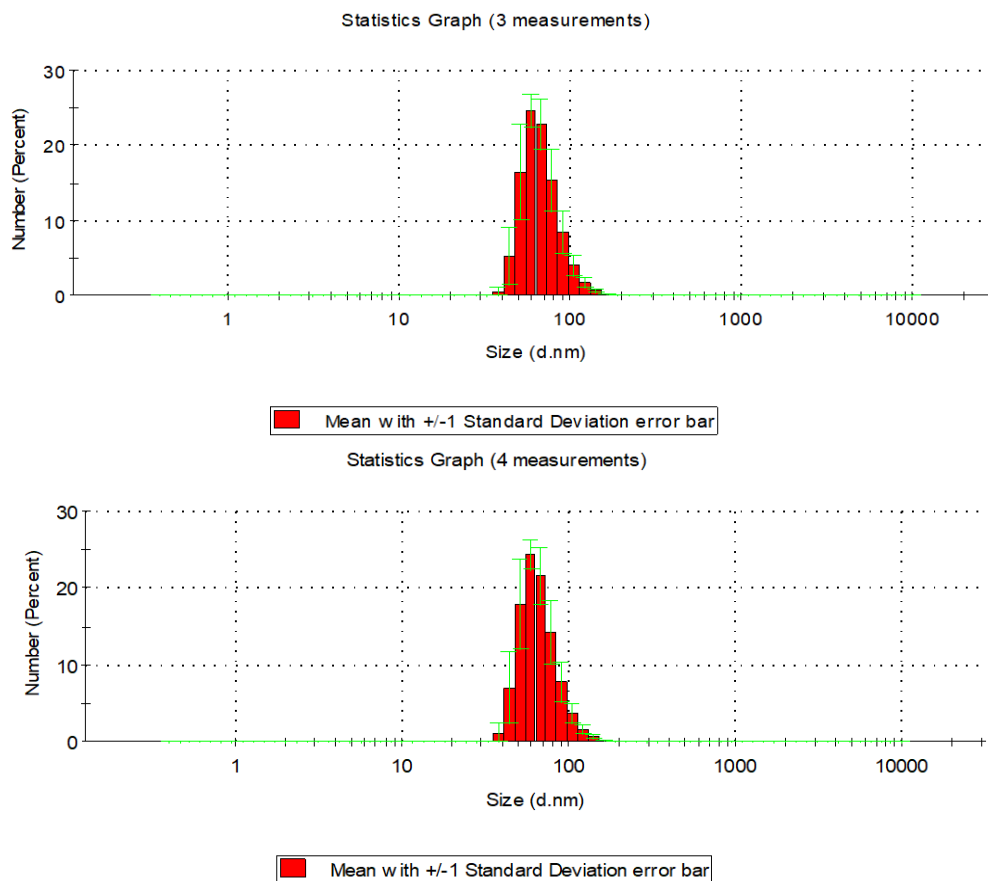


Figure S4. DLS size distribution data (Number) for method Rao et al. [68] with 0.018 M TEOS after synthesis (up) and after one year storage (bottom).

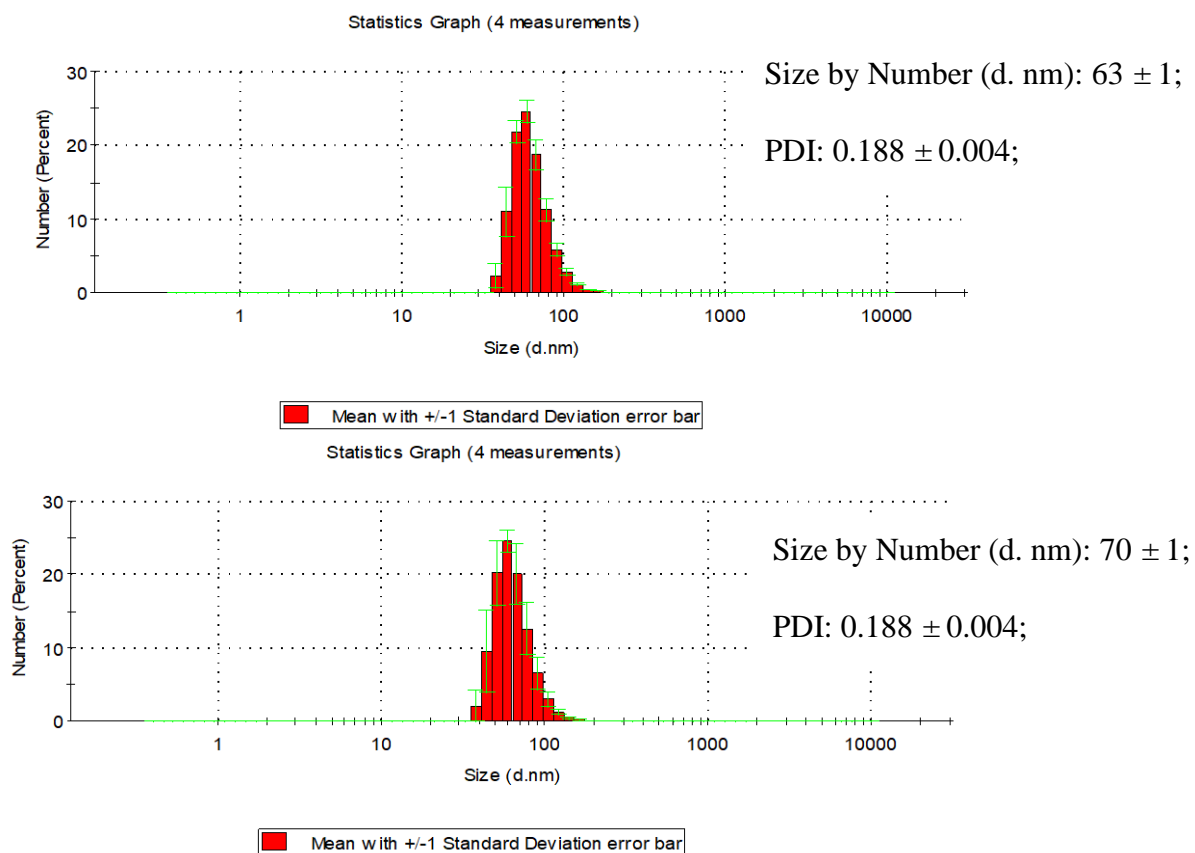


Figure S5. DLS size distribution data (Number) for method Rao et al. [68] with 0.018 M TEOS diluted in 100 mM sodium acetate buffer pH 4 (up), and after 7 days (bottom).

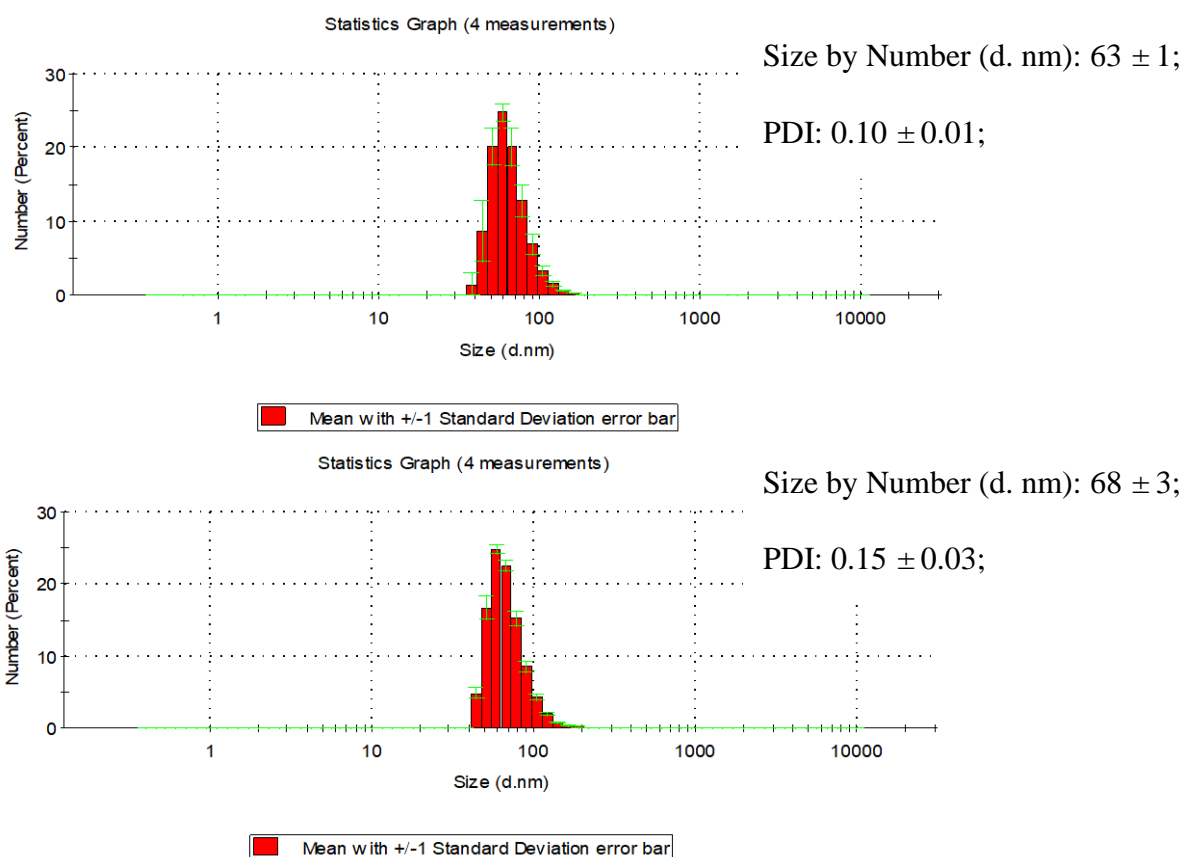


Figure S6. DLS size distribution data (Number) for method Rao et al. [68] with 0.018 M TEOS diluted in 100 mM sodium acetate buffer pH 5 (up), and after 7 days (bottom).

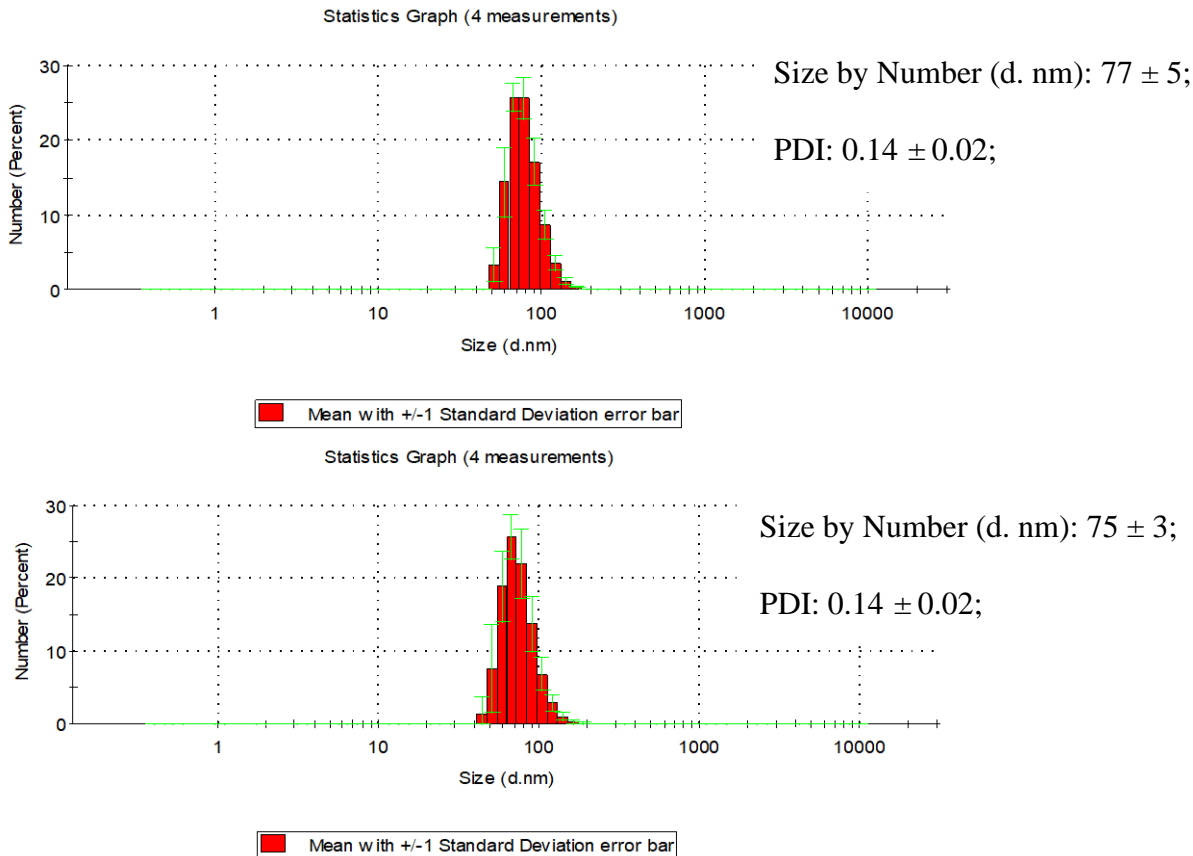


Figure S7. DLS size distribution data (Number) for method Rao et al. [68] with 0.018 M TEOS in 100 mM sodium acetate buffer pH 6 (up), and after 7 days (bottom).

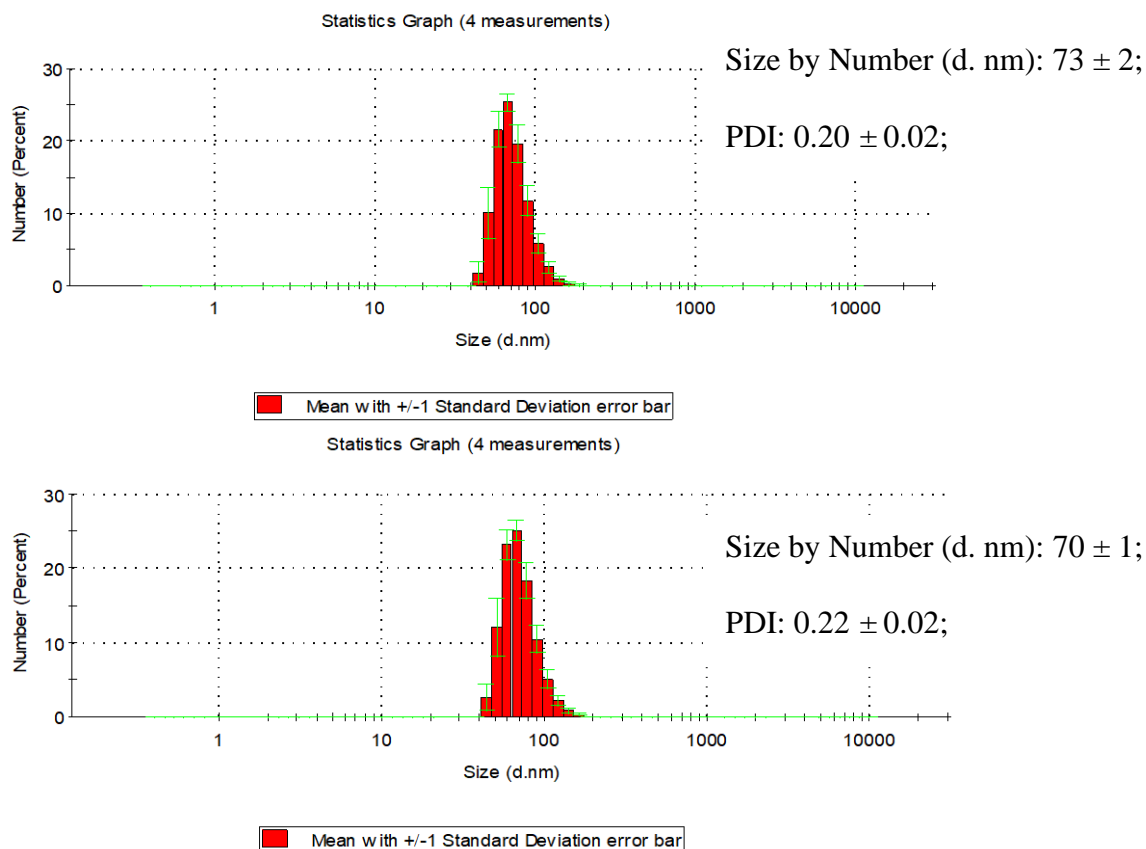


Figure S8. DLS size distribution data (Number) for method Rao et al. [68] with 0.018 M TEOS diluted in 10 mM phosphate-buffered saline pH 7.4 (up), and after 7 days (bottom).

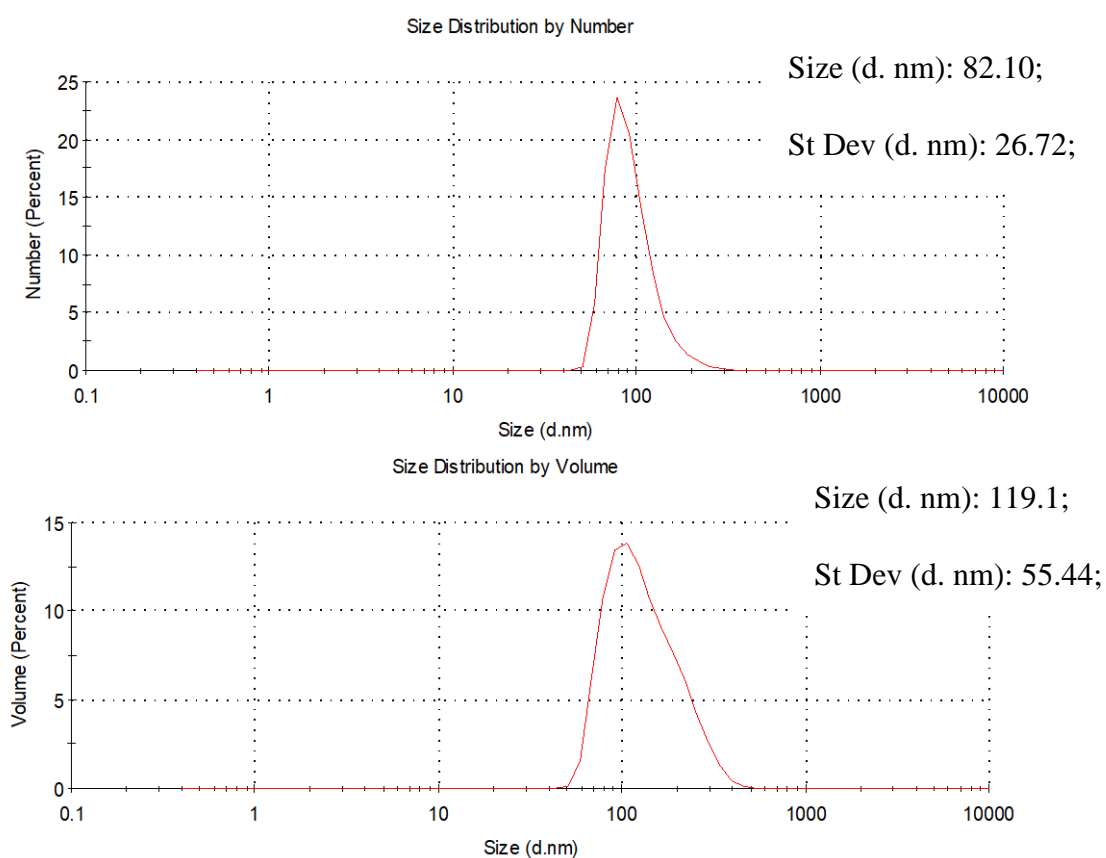


Figure S9. DLS size distribution data (Number, Volume) for method Rao et al. [68] with 0.018 M TEOS diluted in fetal bovine serum. The particle size was determined by the DLS method to be 82 ± 3 nm, with a PDI of 0.30 ± 0.01 .

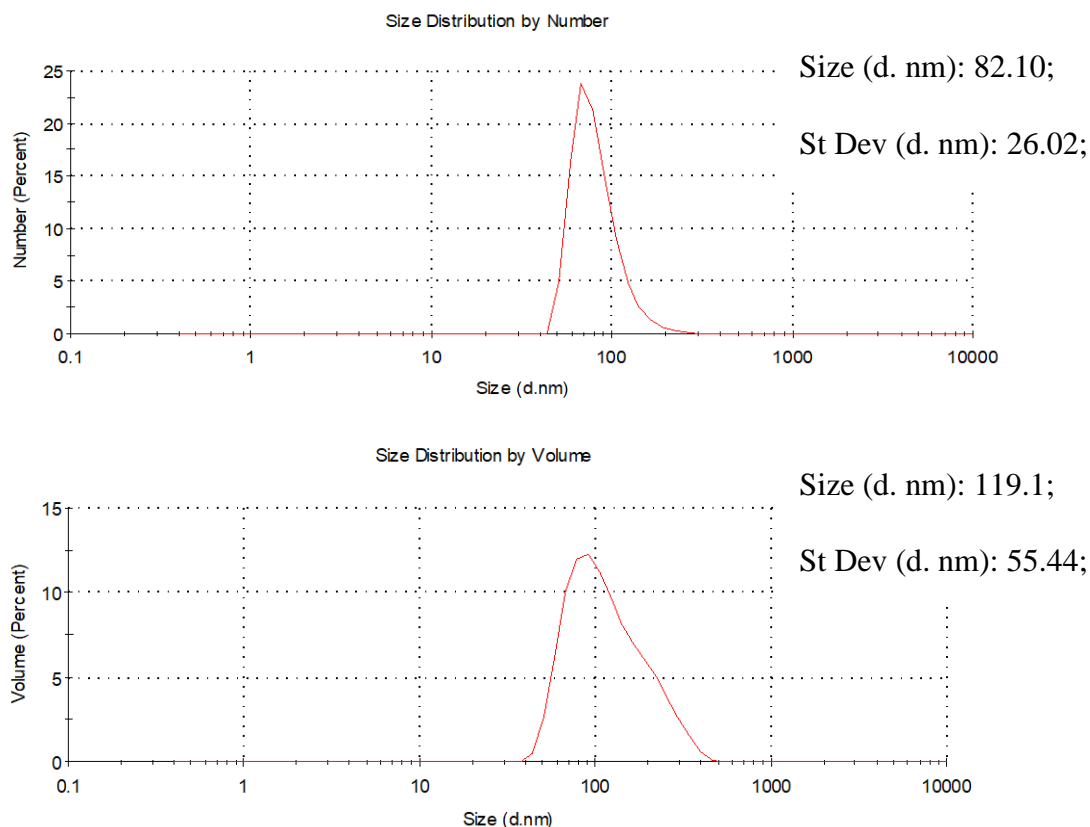


Figure S10. DLS size distribution data (Number, Volume) for method Rao et al. [68] with 0.018 M TEOS diluted in fetal bovine serum (after 10 days). The particle size was determined by the DLS method to be 80 ± 5 nm, with a PDI of 0.20 ± 0.01 .

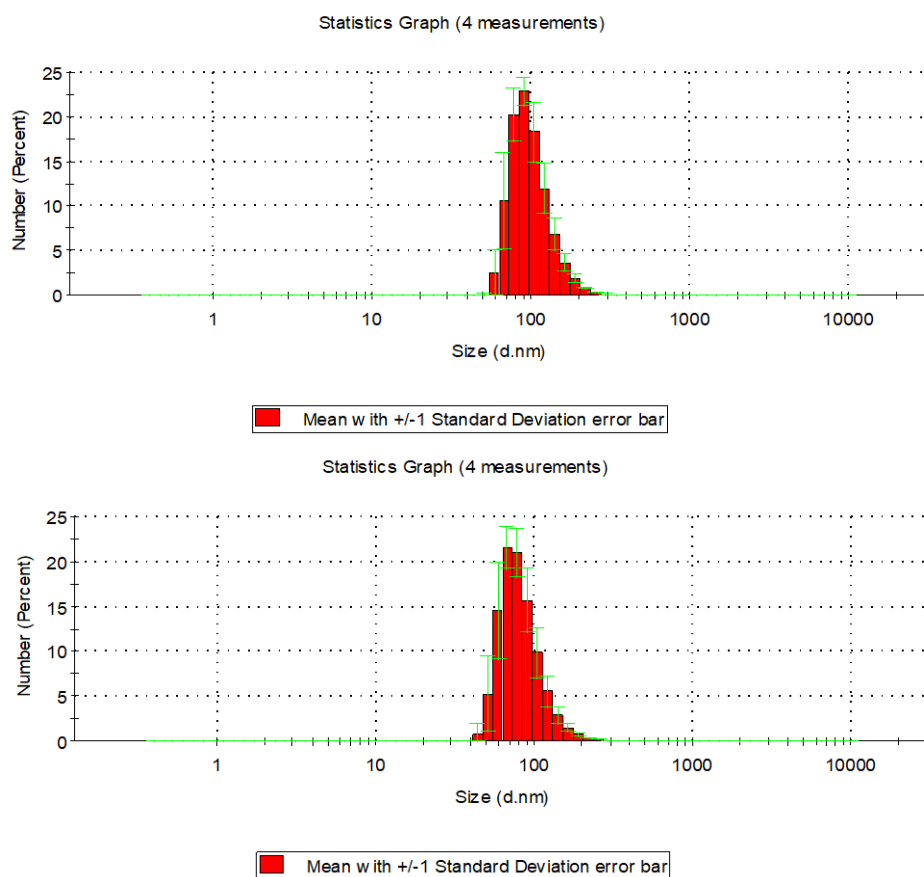


Figure S11. DLS size distribution data (Number) for method Rao et al. [68] with 0.018 M TEOS diluted in fetal bovine serum (up), and after 10 days (bottom).

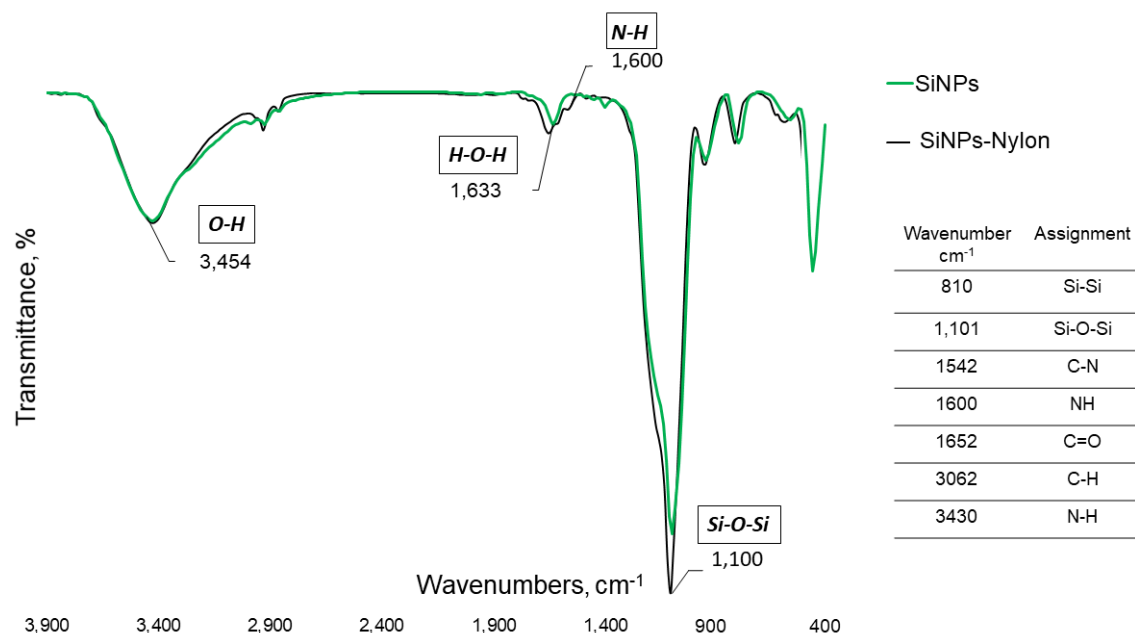


Figure S12. The FT-IR spectra of SiNPs and SiNPs-Nylon in KBr pellets using a Varian 640-IR-spectrometer (Varian, Louisiana, USA) in the range 400–4000 cm⁻¹.



Figure S13. Scheme of the qualitative reaction of the nylon determination in SiNPs and SiNPs-Nylon with N-(2-hydroxyethyl)-Phenazinium ion. The reaction product is colored blue violet in the presence of amino groups. The initial SiNPs was used as a control.

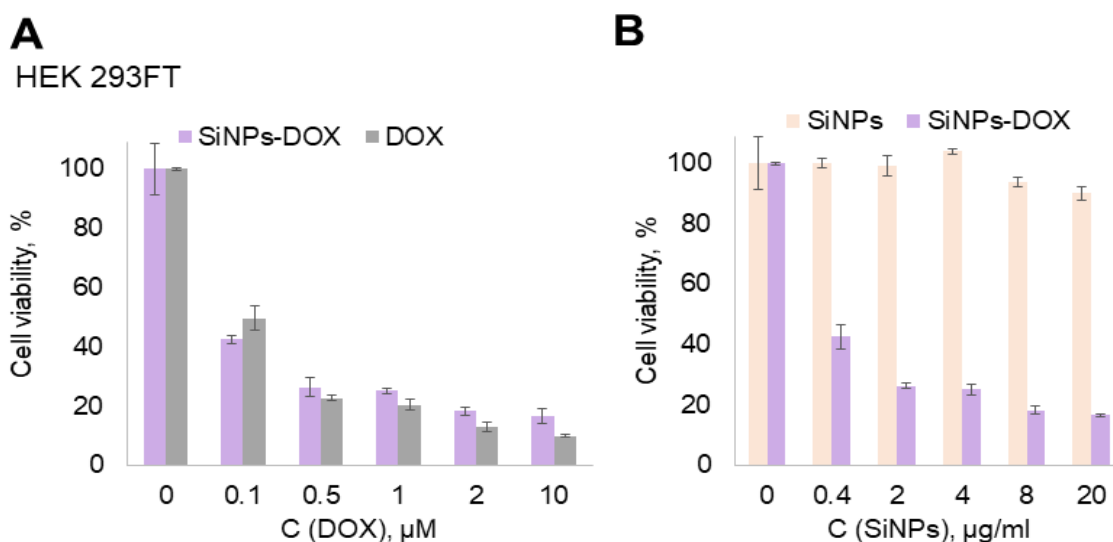


Figure S14. Cell viability assay. HEK 293FT cell line was incubated for 48 h with SiNPs, SiNPs–DOX, and DOX. Cells treated with PBS buffer were used as a 100% viability control. Cells were incubated with the same amount of free DOX or DOX loaded on SiNPs (A), or with the same mass of SiNPs and SiNPs–DOX (B). The IC₅₀ values for SiNPs-DOX and DOX are 0.08 ± 0.01 μM and 0.10 ± 0.02 μM, respectively.