



Article Pulmonary Pharmacokinetic and Pharmacodynamic Evaluation of Ampicillin/Sulbactam Regimens for Pneumonia Caused by Various Bacteria, including Acinetobacter baumannii

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Abstract: This study aimed to assess the dosing regimens of ampicillin/sulbactam for pneumonia based on pulmonary pharmacokinetic (PK)/pharmacodynamic (PD) target attainment. Using the literature data, we developed pulmonary PK models and estimated the probabilities of attaining PK/PD targets in lung tissue. Against bacteria other than A. baumannii (the general treatment), the PK/PD target was set as both 50% time above the minimum inhibitory concentration (T > MIC) for ampicillin and 50% T > 0.5 MIC for sulbactam. For the A. baumannii treatment, the PK/PD target was set as 60% T > MIC for sulbactam. The pulmonary PK/PD breakpoint was defined as the highest minimum inhibitory concentration (MIC) at which the target attainment probability in the lung tissue was \geq 90%. The lung tissue/serum area under the drug concentration–time curve from 0 to 3 h (AUC_{0-3h}) ratios for ampicillin and sulbactam were 0.881 and 0.368, respectively. The ampicillin/sulbactam AUC_{0-3h} ratio in the lung tissue was 3.89. For the general treatment, the pulmonary PK/PD breakpoint for ampicillin/sulbactam at 3 g four times daily in typical patients with creatinine clearance (CL_{cr}) of 60 mL/min was 2 μ g/mL, which covered the MIC_{90s} (the MICs that inhibited the growth of 90% of the strains) of most gram-positive and gram-negative bacteria. For the A. baumannii treatment, the pulmonary PK/PD breakpoint for ampicillin/sulbactam at 9 g 4-h infusion three times daily (27 g/day) in patients with a CL_{cr} of 60 mL/min was 4 µg/mL, which covered the MIC₉₀ of A. baumannii. A PK/PD evaluation for pneumonia should be performed in the lung tissue (the target site) rather than in the blood because sulbactam concentrations are lower in lung tissue. These findings should facilitate the selection of ampicillin/sulbactam regimens for pneumonia caused by various bacteria, including A. baumannii.

Keywords: ampicillin; sulbactam; pharmacokinetics; pharmacodynamics; pulmonary

1. Introduction

Ampicillin/sulbactam, containing the β -lactam antimicrobial agent, ampicillin, and the β -lactamase inhibitor, sulbactam, has been widely used at a dose ratio of 2:1. Both ampicillin and sulbactam are water-soluble drugs and mainly excreted by the kidneys [1]. Ampicillin/sulbactam has been used as a first-line treatment for pneumonia and preoperative prophylaxis for pneumonectomy [1–4].

Bacterial pneumonia, which is caused by a wide variety of bacteria and is broadly classified into community-acquired, hospital-acquired, aspiration pneumonia, etc., is one of the most common infections globally. Community-acquired pneumonia is caused by bacteria such as *Streptococcus pneumoniae*, *Haemophilus influenzae*, and *Moraxella catarrhalis*. Ampicillin/sulbactam is frequently prescribed for community-acquired pneumonia because of its antibacterial activity against β -lactamase-producing pathogens [5]. Furthermore, ampicillin/sulbactam has been used to treat aspiration pneumonia mainly caused by oral



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Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). bacteria such as the *Streptococcus anginosus* group, *Peptostreptococcus* species, *Prevotella* species, and *Fusobacterium* species [6,7]. Meanwhile, *Acinetobacter baumannii* is a globally important pathogen that causes infectious diseases such as ventilator-related pneumonia [8,9]. Although ampicillin/sulbactam is prescribed to treat *A. baumannii* infection, the strain is resistant to ampicillin, but sulbactam is known to exhibit antibacterial activity against *A. baumannii*, independently of ampicillin [10,11]. Therefore, ampicillin/sulbactam has been widely used for the treatment of pneumonia caused by various bacteria.

However, ampicillin/sulbactam at 12 g/day, as compared with 6 g/day, increased the incidence of hepatobiliary enzyme elevation [12]. Therefore, dose-dependent side effects cannot be excluded, and dosage adjustment considering each patient's characteristics (e.g., renal function) should be required. Furthermore, according to the pharmacokinetic (PK)/ pharmacodynamic (PD) theory, the exposure time during which the drug concentrations remain above the minimum inhibitory concentration (MIC) for bacteria (T > MIC) is correlated with the antibacterial activity of β -lactams such as ampicillin [13,14]. The efficacy should be improved with appropriately adjusted regimens, according to the MIC for the bacteria.

In blood, optimization of the ampicillin/sulbactam dosage using population PK/PD analysis has been reported [15]. However, for pneumonia, antimicrobial agents act in the lung tissues, such as the alveoli, in which the main causative pathogens are present, rather than in the blood. Therefore, dose optimization using pulmonary PK/PD analysis might be useful. Several previous reports described the penetration of ampicillin and sulbactam into lung tissue [16] and the alveolar lining fluid [17]. However, they did not fully describe PK in the lungs, and no accurate pulmonary PK/PD evaluation using a mathematical model has been performed.

Therefore, this study characterized pulmonary PK models of ampicillin and sulbactam using previously reported data, such as PK and physiological parameters, and evaluated ampicillin/sulbactam pulmonary PK/PD target attainment considering both ampicillin and sulbactam concentrations in lung tissue. Moreover, we optimized dosing regimens of ampicillin/sulbactam for pneumonia caused by various bacteria, including *A. baumannii*.

2. Results

2.1. Lung Tissue/Serum Ratio and Ampicillin/Sulbactam Ratio in Serum and Lung Tissue

The lung tissue/serum ratio and ampicillin/sulbactam ratio in the serum and lung tissue are presented in Table 1.

Specimen and Parameter	Value		
	Ampicillin 2.0 g (15 subjects)	Sulbactam 1.0 g (15 subjects)	Ampicillin/ Sulbactam Ratio (2.0 g/1.0 g)
Serum			
C _{max} (µg/mL)	40.8	25.3	1.61
AUC _{0-3h} (µg·h/mL)	83.5	51.2	1.63
Lung tissue			
C_{max} (µg/g)	35.6	8.6	4.14
AUC_{0-3h} (µg·h/g)	73.6	18.9	3.89
Lung tissue/serum ratio			
C _{max}	0.873	0.339	
AUC _{0-3h}	0.881	0.368	

Table 1. Calculation of lung tissue/serum ratio and ampicillin/sulbactam ratio in the lung tissue.

 AUC_{0-3h} , area under the drug concentration–time curve from 0 to 3 h, calculated based on the trapezoidal rule; C_{max} , observed maximum concentration. Data are provided as the mean derived from [16].

The C_{max} values for ampicillin and sulbactam from the literature were 40.8 and 25.3 µg/mL in the serum and 35.6 and 8.6 µg/g in the lung tissue, respectively. The area under the drug concentration–time curve from the 0 to 3 h (AUC_{0–3h}) values for ampicillin and sulbactam, calculated from the literature, were 83.5 and 51.2 µg·h/mL in the serum and 73.6 and 18.9 µg·h/g in the lung tissue, respectively. For ampicillin, the lung tissue/serum ratios were 0.873 for the

 C_{max} and 0.881 for the AUC_{0-3h}. For sulbactam, the lung tissue/serum ratios were 0.339 for the C_{max} and 0.368 for the AUC_{0-3h}. For the pulmonary PK modeling, the KP_{lung} for ampicillin/sulbactam was fixed as 0.881/0.368 for the AUC_{0-3h}. The ampicillin/sulbactam ratio of the C_{max} and AUC_{0-3h} in the lung tissue was approximately 4.

2.2. Model Validation

Visual predictive checks were also performed for the observed and predicted lung tissue concentration vs. the time curves of ampicillin and sulbactam (Figure 1). The mean \pm standard deviation of the observed lung tissue concentrations was almost within the predicted 95% confidence intervals for the 2.5th, 50th, and 97.5th percentiles.

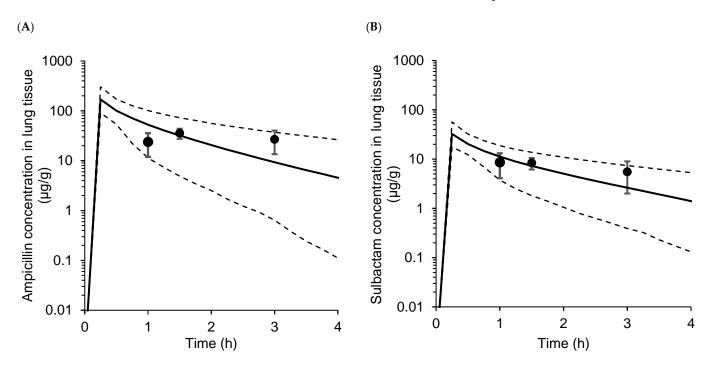


Figure 1. Visual predictive check plots of (**A**) ampicillin and (**B**) sublactam, representing the observed lung tissue concentrations (mean \pm standard deviation) after a 15-min of infusion of ampicillin/sublactam 3 g (ampicillin 2 g and sublactam 1 g) derived from the literature data [16]. The heavy line and dotted line denote the median and the 95% predicted interval, calculated from 1000 replicates, respectively.

2.3. PK and PD Evaluation

The probabilities of target attainment in the lung tissue using different ampicillin/sulbactam regimens at specific MICs are presented in Figure 2 for bacteria other than *A. baumannii* (the general treatment). Regarding the probability–MIC curve in the lung tissue for ampicillin, only the MICs with a target attainment probability of more than 90% for sulbactam (50% T > 0.5 MIC for sulbactam) are represented by red symbols, indicating that the ampicillin/sulbactam combination is effective. The results of Figure 2 demonstrate that the number of effective regimens for the ampicillin/sulbactam combination decreases with an improving renal function or as the MIC of the causative pathogen increases. Furthermore, the pulmonary PK/PD breakpoints (the highest MIC at which the target attainment probability in the lung tissue was \geq 90%) are presented in Table 2. The pulmonary PK/PD breakpoints of 3.0 g four times daily were 1 µg/mL for CL_{cr} = 90 mL/min, 2 µg/mL for CL_{cr} = 60 mL/min, and 8 µg/mL for CL_{cr} = 30 mL/min, respectively.



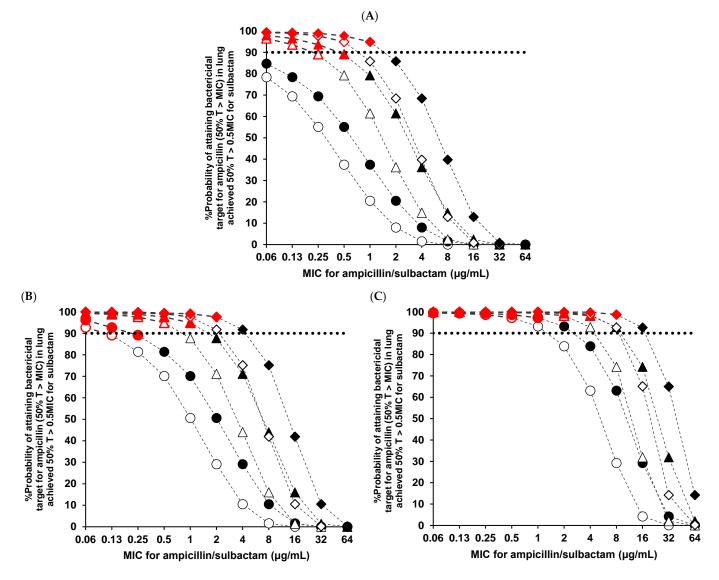


Figure 2. Probabilities of attaining bactericidal (50% T > MIC for ampicillin) targets in lung tissue at specific MICs for typical patients of (**A**) $CL_{cr} = 90 \text{ mL/min}$, (**B**) $CL_{cr} = 60 \text{ mL/min}$, and (**C**) $CL_{cr} = 30 \text{ mL/min}$, respectively. Regarding the probability–MIC curve for ampicillin in lung tissue, MICs with a probability of target attainment for sulbactam of more than 90% (50% T > 0.5 MIC for sulbactam), indicating that the ampicillin/sulbactam combination is effective, are represented by red symbols. The lines represent the probability of target attainment for each dosing regimen of ampicillin/sulbactam (\bigcirc 1.5 g twice daily, 0.5-h infusion; \bullet 3.0 g twice daily, 0.5-h infusion; \Diamond 1.5 g four times daily, 0.5-h infusion; \diamond 3.0 g four times daily, 0.5-h infusion; \diamond 1.5 g four times daily, 0.5-h infusion; \diamond 3.0 g four times daily, 0.5-h infusion). The dotted black line represents a 90% probability. MIC, minimum inhibitory concentration.

Ampicillin/Sulbactam Regimen	Bactericidal Target (50% T > MIC for Ampicillin) and 50% T > 0.5 MIC for Sulbactam	
$CL_{cr} = 90 \text{ mL/min}$		
1.5 g twice daily, 0.5-h infusion (total 3 g/day)	-	
3.0 g twice daily, 0.5-h infusion (total 6 g/day)	-	
1.5 g three times daily, 0.5-h infusion (total 4.5 g/day)	0.13	
3.0 g three times daily, 0.5-h infusion (total 9 g/day)	0.25	
1.5 g four times daily, 0.5-h infusion (total 6 g/day)	0.5	
3.0 g four times daily, 0.5-h infusion (total 12 g/day)	1	
$CL_{cr} = 60 \text{ mL/min}$		
1.5 g twice daily, 0.5-h infusion (total 3 g/day)	0.06	
3.0 g twice daily, 0.5-h infusion (total 6 g/day)	0.13	
1.5 g three times daily, 0.5-h infusion (total 4.5 g/day)	0.5	
3.0 g three times daily, 0.5-h infusion (total 9 g/day)	1	
1.5 g four times daily, 0.5-h infusion (total 6 g/day)	1	
3.0 g four times daily, 0.5-h infusion (total 12 g/day)	2	
$CL_{cr} = 30 \text{ mL/min}$		
1.5 g twice daily, 0.5-h infusion (total 3 g/day)	0.5	
3.0 g twice daily, 0.5-h infusion (total 6 g/day)	1	
1.5 g three times daily, 0.5-h infusion (total 4.5 g/day)	2	
3.0 g three times daily, 0.5-h infusion (total 9 g/day)	4	
1.5 g four times daily, 0.5-h infusion (total 6 g/day)	4	
3.0 g four times daily, 0.5-h infusion (total 12 g/day)	8	

Table 2. Pulmonary PK/PD breakpoints of ampicillin/sulbactam for general treatment with both activities of bactericidal ampicillin and β -lactamase-inhibiting sulbactam.

Note: Pulmonary PK/PD breakpoints are defined as the highest MIC attaining more than 90% probabilities in lung tissue.

Moreover, the probabilities of target attainment in the lung tissue are presented in Figure 3 for the *A. baumannii* treatment with sulbactam activity. The pulmonary PK/PD breakpoints are presented in Table 3. The pulmonary PK/PD breakpoints of 9.0 g three times daily with 4-h infusion in the lung tissue were 2 µg/mL for $CL_{cr} = 90$ mL/min, 4 µg/mL for $CL_{cr} = 60$ mL/min, and 16 µg/mL for $CL_{cr} = 30$ mL/min, respectively.

Table 3. Pulmonary PK/PD breakpoints of ampicillin/sulbactam for *A. baumannii* treatment with sulbactam activity.

Ampicillin/Sulbactam Regimen	Bactericidal Target 60% T > MIC for Sulbactam
CL _{cr} = 90 mL/min	
3.0 g twice daily, 0.5-h infusion (total 6 g/day)	-
6.0 g twice daily, 0.5-h infusion (total 12 g/day)	-
3.0 g three times daily, 0.5-h infusion (total 9 g/day)	0.13
6.0 g three times daily, 0.5-h infusion (total 18 g/day)	0.25
3.0 g four times daily, 0.5-h infusion (total 12 g/day)	0.25
6.0 g four times daily, 0.5-h infusion (total 24 g/day)	0.5
9.0 g three times daily, 4-h infusion (total 27 g/day)	2
$CL_{cr} = 60 \text{ mL/min}$	
3.0 g twice daily, 0.5-h infusion (total 6 g/day)	-
6.0 g twice daily, 0.5-h infusion (total 12 g/day)	0.06
3.0 g three times daily, 0.5-h infusion (total 9 g/day)	0.25
6.0 g three times daily, 0.5-h infusion (total 18 g/day)	0.5
3.0 g four times daily, 0.5-h infusion (total 12 g/day)	0.5
6.0 g four times daily, 0.5-h infusion (total 24 g/day)	1
9.0 g three times daily, 4-h infusion (total 27 g/day)	4

Ampicillin/Sulbactam Regimen	Bactericidal Target 60% T > MIC for Sulbactam
$CL_{cr} = 30 \text{ mL/min}$	
3.0 g twice daily, 0.5-h infusion (total 6 g/day)	0.5
6.0 g twice daily, 0.5-h infusion (total 12 g/day)	1
3.0 g three times daily, 0.5-h infusion (total 9 g/day)	1
6.0 g three times daily, 0.5-h infusion (total 18 g/day)	2
3.0 g four times daily, 0.5-h infusion (total 12 g/day)	2
6.0 g four times daily, 0.5-h infusion (total 24 g/day)	4
9.0 g three times daily, 4-h infusion (total 27 g/day)	16

Note: Pulmonary PK/PD breakpoints are defined as the highest MIC attaining more than 90% probabilities in lung tissue.

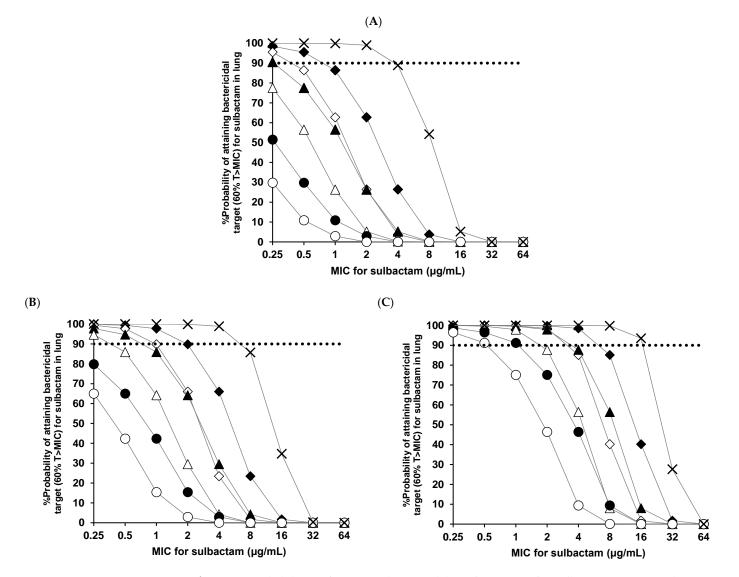


Figure 3. Probabilities of attaining bactericidal (60% T > MIC for subactam) targets in lung tissue at specific MICs for typical patients of (**A**) $CL_{cr} = 90 \text{ mL/min}$, (**B**) $CL_{cr} = 60 \text{ mL/min}$, and (**C**) $CL_{cr} = 30 \text{ mL/min}$, respectively. The lines represent the probability of target attainment for each dosing regimen of ampicillin/subactam ($\bigcirc 3.0 \text{ g}$ twice daily, 0.5-h infusion; \bullet 6.0 g twice daily, 0.5-h infusion; \triangle 3.0 g three times daily, 0.5-h infusion; \triangle 3.0 g four times daily, 0.5-h infusion; \times 9.0 g three times daily, 0.5-h infusion; \times 9.0 g three times daily, 4-h infusion). The dotted black line represents a 90% probability. MIC, minimum inhibitory concentration.

3. Discussion

No pulmonary PK/PD evaluation with a mathematical method has been reported for ampicillin/sulbactam. This study developed site-specific PK models, which are important for pneumonia.

For both the C_{max} and AUC, the lung tissue/plasma ratios of sulbactam were < half that for ampicillin. From these results, the penetration of sulbactam from the systemic circulation into the lung tissue was lower than that of ampicillin. The ampicillin/sulbactam ratio resulting in the best antibacterial activity ranged from 1.0 to 2.0 [18]. Therefore, for general treatment, if the PK/PD target in the lung tissue is attained for ampicillin but not for sulbactam, the combination treatment (ampicillin/sulbactam) will not be effective. Thus, for general treatment, it is necessary to analyze the pulmonary pharmacokinetics/pharmacodynamics of both drugs for the prediction of the efficacy of the combination regimen.

Pulmonary PK models for ampicillin and sulbactam were described using hybrid modeling, which can incorporate previously reported PK and physiological parameters. Since a specific organ clearance depends mainly on its physiological organ blood flow, both the system-to-lung clearance and lung-to-system clearance were assumed to be the same and set as the lung blood flow in the hybrid modeling. Independently from the conventional PK model, the mass balance in the lung compartment was assumed not to affect the mass balance in the central and peripheral compartments. The visual predictive check plots (Figure 1) indicated that the most observed lung concentrations were within model-predicted ranges. However, the observed lung concentrations currently available (mean \pm standard deviation at three-time points) were too few to assess the model performance. Furthermore, model validation by goodness-of-fitness plots was not possible due to a lack of individual raw concentration data in the literature [16]. The model in this study, thus, needs to be further validated in the future.

Next, the PK/PD target attainment in the lung tissue for different dosing regimens was estimated. For general treatment, the probability of target attainment for ampicillin in the lung tissue (the pulmonary PK/PD breakpoint of 3 g four times daily was 2 μ g/mL for CL_{cr} = 60 mL/min, Table 2) was in line with the previously published data in the plasma (the PK/PD breakpoint of 3 g four times daily in plasma was 2 μ g/mL for CL_{cr} = 60 mL/min, [19]). However, because the sulbactam concentration in the lung tissue and the probability of target attainment for sulbactam are lower, some regimens are considered to be poorly active as combination treatments, even though ampicillin had a probability of target attainment exceeding 90% (Figure 2). Furthermore, the probability of target attainment for sulbactam in the lung tissue (the pulmonary PK/PD breakpoint of 6 g four times daily was 1 μ g/mL for CL_{cr} = 60 mL/min, Table 3) was lower than that reported in the plasma (the PK/PD breakpoint of 6 g four times daily in plasma was 4 μ g/mL for CL_{cr} = 60 mL/min [20]). Therefore, considering the pharmacokinetics/pharmacodynamics of ampicillin/sulbactam in lung tissue, the efficacy of this combination might depend on the probability of target attainment for sulbactam rather than ampicillin.

Regarding the general treatment, the pulmonary PK/PD breakpoints of ampicillin/ sulbactam 3 g four times daily (12 g/day: the approved maximum dosage) were 1 µg/mL (the MIC₉₀ [the MIC that inhibited the growth of 90% of the strains] of the MSSA) for $CL_{cr} = 90$ mL/min, 2 µg/mL (the MIC₉₀ of the *S. pneumoniae* and *Prevotella* species) for $CL_{cr} = 60$ mL/min, and 8 µg/mL for $CL_{cr} = 30$ mL/min. Thus, the probability of attaining the pulmonary PK/PD target decreased as renal function improved. For $CL_{cr} = 30$ mL/min, the PK/PD breakpoints of the twice daily dosing regimen were lower in the lung tissue (0.5 µg/mL for 1.5 g twice daily and 1 µg/mL for 3.0 g twice daily) than in the plasma (1 µg/mL for 1.5 g twice daily and 2 µg/mL for 3.0 g twice daily) [19] because of the poor penetration of sulbactam into the lung tissue. Suzuki et al. reported that ampicillin/sulbactam treatment for elderly patients with pneumonia and renal dysfunction (10 mL/min $\leq CL_{cr} < 50$ mL/min) was more effective using a four-times daily regimen than using a twice-daily regimen [21]. Similarly, from our findings that the pulmonary PK/PD breakpoints of the four times daily regimen were higher than those of the twice-daily regimen, we recommend four times daily regimens for patients with renal dysfunction; 3 g four times daily (12 g/day) as the maximum dose for pneumonia is recommended by the Japanese Association for Infectious Diseases/Japanese Society of Chemotherapy guidelines for the clinical management of infectious diseases [6]. In this study, the pulmonary PK/PD breakpoint of 3 g four times daily was 2 µg/mL for $CL_{cr} = 60 \text{ mL/min}$, and it covered the MIC_{90s} of the MSSA, *S. pneumoniae*, *M. catarrhalis*, the *S. anginosus* group, the *Peptostreptococcus* species, the *Prevotella* species, and the *Fusobacterium* species. Therefore, this guideline dose might be valid as empiric therapy for community-acquired and aspiration pneumonia. However, because the MIC for β -lactamase–nonproducing ampicillin-resistant *H. influenzae* is high (MIC₉₀ = 8 µg/mL), the use of other antimicrobial agents might be required for typical patients with normal renal function ($CL_{cr} \ge 60 \text{ mL/min}$).

Regarding the *A. baumannii* treatment, the pulmonary PK/PD breakpoints of ampicillin/ sulbactam 3 g four times daily (12 g/day: the approved maximum dosage) were 0.25 μ g/mL for CL_{cr} = 90 mL/min, 0.5 μ g/mL for CL_{cr} = 60 mL/min, and 2 μ g/mL for CL_{cr} = 30 mL/min. From these results, the maximum approved dose does not appear to achieve 4 μ g/mL (the MIC₉₀ of *A. baumannii*) because of the low penetration of sulbactam into the lung tissue. This suggests that dosing regimens for ampicillin/sulbactam exceeding the maximum approved dose are required. A clinical report also found that a high-dose regimen was effective in patients with ventilator-associated pneumonia caused by multidrug-resistant *A. baumannii*. [22]. Furthermore, the Sanford guideline recommends a 4-h infusion of ampicillin/sulbactam at 9 g three times daily (27 g/day) for ventilator-related pneumonia caused by *A. baumannii* [23]. Our results illustrated that this dosing regimen achieved an MIC₉₀ of 4 μ g/mL against *A. baumannii* in typical patients with CL_{cr} = 60 mL/min. Thus, from the perspective of the pulmonary penetration of sulbactam, it was confirmed that high-dose regimens are necessary for the treatment of *A. baumannii*.

Concerning the limitations of this study, the PK and physiological parameters derived from previous reports [16,19,24,25] were estimated from uninfected patients or healthy volunteers. The inflammation of lung tissue caused by pneumonia can increase vascular permeability in the lungs. Therefore, the population parameters used in this study might underestimate the pulmonary penetration of ampicillin and sulbactam. Second, because our findings are only predictions of efficacy based on PK/PD simulations, it is necessary to perform clinical studies in infected patients to identify the appropriateness of the dosing regimens. The hybrid model used in our study enables the analysis of pharmacokinetics/pharmacodynamics at various target sites using the literature data, such as the organ blood flow, organ volume, and tissue/blood drug concentration ratios. This method is practical and versatile in clinical situations involving difficult tissue sampling.

4. Materials and Methods

4.1. Pulmonary PK Modeling for Ampicillin and Sulbactam

The pulmonary pharmacokinetics of ampicillin and sulbactam were separately described for each drug using the following hybrid model (Figure 4). The hybrid model is a model in which physiological parameters such as organ blood flow and volume are partially connected to the conventional PK model. Based on blood concentrations, this model has been used for the target site PK/PD analysis [19,26–29]. In this study, a lung compartment was connected to a two-compartment PK model using blood concentrations [19]; thus, the hybrid model consisted of three compartments.

$$dX(central)/dt = R_{inf} - (CL/V_{central} + Q/V_{central}) \times X(central) + Q \times X(peripheral)/V_{peripheral}$$

dX(peripheral)/dt = $Q \times X(central)/V_{central} - Q \times X(peripheral)/V_{peripheral}$

 $dX(lung)/dt = Q_{lung} \times X(central)/V_{central} - Q_{lung} \times X(lung)/V_{lung}/KP_{lung}$

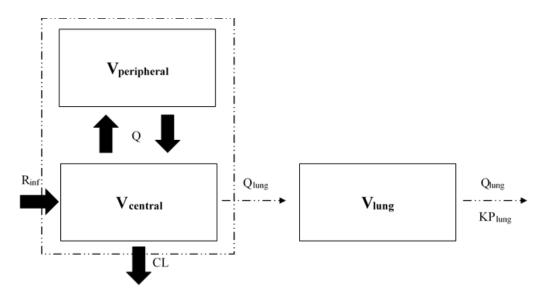


Figure 4. Pulmonary PK modeling of ampicillin and sulbactam.Model parameters: $V_{central}$ and $V_{peripheral}$, volumes of distribution of the central and peripheral compartments (L), respectively; V_{lung} , lung volume (kg); CL, clearance (L/h); Q, central–peripheral intercompartmental clearance (L/h); Q_{lung} , lung plasma flow (L/h); KP_{lung} , lung-to-plasma partition coefficient; R_{inf} , rate of infusion (mg/h).

In the formulas, X(central), X(peripheral), and X(lung) are the amounts of the drug (mg) in the central, peripheral, and lung compartments, respectively; Rinf is the rate of infusion (mg/h); CL is the clearance (L/h) from the central compartment; V_{central} and V_{peripheral} are the volumes of distribution (L) of the central and peripheral compartments, respectively; and Q is the central–peripheral intercompartmental clearance (L/h). The model parameters of ampicillin and sulbactam are listed in Table 4. In the analysis of blood concentrations, the fixed-effects parameters (θ CL, θ V_{central}, θ Q, and θ V_{peripheral}) and the interindividual variability (η CL, η V_{central}, η Q, and η V_{peripheral}) were derived from previously reported population PK parameters of ampicillin and sulbactam [19]. The fixed-effects parameters, θi (CL, V_{central}, Q, and V_{peripheral}), of ampicillin and sulbactam in blood concentrations, were fixed as follows: CL = 11.03 and 10.50 (L/h), $V_{central}$ = 7.80 and 8.96 (L), Q = 7.07 and 7.29 (L/h), and $V_{peripheral}$ = 3.98 and 4.93 (L). In addition, CL_{cr}, which was calculated by the Cockcroft–Gault formula, was incorporated as a covariate of the CL. We also used Qlung (lung plasma flow in L/h) and V_{lung} (lung volume in kg) as physiological parameters. Q_{lung} and V_{lung} were derived from reference values [24,25]. For both drugs, the physiological fixed-effects parameters were fixed as follows: $Q_{lung} = 207$ (L/h), and $V_{lung} = 0.47$ (L). Since the drug is present only in the plasma portion, lung plasma flow was calculated by multiplying the lung blood flow (360 L/h) [24,25] and human hematocrit value (approximately 42.5%), as follows: Q_{lung} = lung blood flow (360 L/h) * (1–0.425) = 207 (L/h). The result of the lung tissue/serum AUC_{0-3h} ratio calculated in the results section was used as the KP_{lung} (lung-to-plasma partition coefficient). The demographic information of the literature data used in this study is represented in the supplementary material (Table S1). PK modeling predicting the drug concentrations in lung tissue was performed using the NONMEM program (version 7.4; ICON Public Limited Company, Dublin, Ireland).

Parameter	Ampicillin	Sulbactam	
	Value (RSE%)	Value (RSE%)	
Fixed-effects parameter			
$CL (L/h) = \theta_{CL} \times (CL_{cr}/68.3)^{\theta CLcr on}$	CL		
$\theta_{\rm CL}$ (L/h) ^a	11.03 (5.1)	10.50 (5.0)	
$\theta_{\text{CLcr on CL}}^{a}$	0.831 (14.1)	0.774 (18.6)	
$V_{central}$ (L) = $\theta_{Vcentral}$ ^a	7.80 (5.9)	8.96 (9.6)	
$Q(L/h) = \theta_Q^a$	7.07 (14.3)	7.29 (21.4)	
$V_{peripheral}$ (L) = $\theta_{Vperipheral}^{a}$	3.98 (12.3)	4.93 (13.4)	
$KP_{lung} = \theta_{KPlung} b^{b}$	0.881 Fixed	0.368 Fixed	
$Q_{\text{lung}}(L/h) = \theta_{Q\text{lung}}^{c}$	207 Fixed	207 Fixed	
$V_{lung}(kg) = \theta_{Vlung}c$	0.47 Fixed	0.47 Fixed	
Interindividual variability (exponen	tial error model)		
ηCL ^a	0.0985 (26.1)	0.0626 (26.8)	
ηVcentral ^a	0.160 (21.3)	0.147 (27.5)	
ηQ^{a}	0.588 (44.2)	0.399 (48.4)	
ηVperipheral ^a	0.298 (37.2)	0.177 (37.9)	
Residual variability (additive error i	model)		
ε ^a	2.70 (26.2)	1.22 (38.8)	

Table 4. PK parameters predicting lung tissue concentration.

^a, Parameters derived from [19]; ^b, parameter derived from the lung tissue/serum AUC_{0-3h} ratio (Table 1); ^c, parameter derived from [24,25]. RSE, relative standard error; θ , population mean value; η , random variable, which is normally distributed with a mean of zero and variance; ε , random error, which is normally distributed with a mean of zero and variance. CL_{cr} was calculated by the Cockcroft–Gault formula.

4.2. Calculation of the Lung Tissue/Serum Ratio and Ampicillin/Sulbactam Ratio in Serum and Lung Tissue

The mean values of lung tissue and serum concentrations (1, 1.5, and 2–4 h each) in patients reported by Frank et al. [16] were used because individual raw data were not described. For each drug, the C_{max} was defined as the highest value in the mean concentrations described in the literature. The AUC_{0–3h}, based on the mean concentrations (three-time points), was estimated according to the trapezoidal rule. The AUC_{0–inf} was not estimated because of insufficient time points for appropriate extrapolation to infinity. The lung tissue/serum ratio and ampicillin/sulbactam ratio in the serum and lung tissue were calculated from the C_{max} or AUC_{0–3h} ratio. The specific gravity of lung tissue was assumed to be 1 (g = mL).

4.3. Model Validation

Visual predictive checks were performed to validate the models. One thousand datasets were simulated using the model parameters, including the interindividual and residual variabilities. The observed mean \pm standard deviation values in the previous report [16] were confirmed whether they were within the 95% confidential interval of the predicted concentration in lung tissue.

4.4. PK/PD Simulation

A set of fixed-effects parameters, θ i (CL, V_{central}, Q, V_{peripheral}, KP_{lung}, Q_{lung}, and V_{lung}), of 1000 virtual subjects for each renal function (three typical CL_{cr} = 90, 60, and 30 mL/min) were randomly generated using the \$SIMULATION command in NONMEM based on each mean value and interindividual variability. By describing each dosing information in the dataset, ampicillin and sulbactam concentrations in lung tissue were calculated by each dosing regimen. The time at which the drug concentration coincided with a specific MIC (0.06–64 µg/mL) was determined, and T > MIC was calculated as the cumulative percentage of time over a 24-h period for different renal functions and different dosing intervals in lung tissue. The total concentration was not able to be corrected for the free fraction because the tissue protein-binding of both drugs in lung tissue is currently unknown. For general treatment, the probability of target attainment (%) at a specific MIC in lung tissue was defined as the proportion of

1000 estimates that achieved the bactericidal target (both 50% T > MIC for ampicillin [30,31] and 50% T > 0.5 MIC for sulbactam). This target was chosen because the MIC evaluation in vitro for ampicillin/sulbactam is 2:1. [32] For the *A. baumannii* treatment, the probability of target attainment (%) at a specific MIC in lung tissue was defined as the proportion of 1000 estimates that achieved the bactericidal target (60% T > MIC for sulbactam) [33].

The MIC distribution data for ampicillin/sulbactam were obtained from the Japanese surveillance of antimicrobial susceptibilities [20,34–36]. Nine common types of pathogens were selected for pneumonia. These included methicillin-susceptible *Staphylococcus aureus* (MSSA; n = 676, the MIC for the 90th percentile of the clinical strains [MIC₉₀] = 1 µg/mL), *S. pneumoniae* (n = 565, MIC₉₀ = 2 µg/mL), *H. influenzae* (all strains: n = 544, MIC₉₀ = 4 µg/mL; β-lactamase-nonproducing ampicillin-resistant *H. influenzae*: n = 70, MIC₉₀ = 8 µg/mL), *M. catarrhalis* (n = 491, MIC₉₀ = 0.25 µg/mL), the *S. anginosus* group (n = 100, MIC₉₀ = 0.25 µg/mL), the *Peptostreptococcus* species (n = 100, MIC₉₀ = 0.03 µg/mL), the *Prevotella* species (n = 100, MIC₉₀ = 2 µg/mL), the *Fusobacterium* species (n = 50, MIC₉₀ = 0.06 µg/mL) for ampicillin/sulbactam, and *A. baumannii* (n = 27, MIC₉₀ = 4 µg/mL) for sulbactam.

5. Conclusions

This study focused on lung tissue as the target site of pneumonia, and we performed a pulmonary PK/PD evaluation. For pneumonia, pharmacokinetics/pharmacodynamics should be evaluated in the lungs rather than in the blood because the sulbactam concentration in lung tissue is low. We provided practical ampicillin/sulbactam dosing regimens for pneumonia caused by various pathogens, considering the susceptibility of pathogens and renal function.

Supplementary Materials: The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/antibiotics12020303/s1, Table S1: Demographic information of literature data used in this study.

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References

- USP, UNASYN (Ampicillin Sodium/Sulbactam Sodium). 2017. Available online: https://www.accessdata.fda.gov/drugsatfda_docs/ label/2017/050608s044lbl.pdf (accessed on 16 June 2021).
- Mandell, L.A.; Wunderink, R.G.; Anzueto, A.; Bartlett, J.G.; Campbell, G.D.; Dean, N.C.; Dowell, S.F.; File, T.M., Jr.; Musher, D.M.; Niederman, M.S.; et al. Infectious Diseases Society of America; American Thoracic Society Infectious Diseases Society of America/American Thoracic Society consensus guidelines on the management of community-acquired pneumonia in adults. *Clin. Infect. Dis.* 2007, 44, S27–S72. [CrossRef] [PubMed]
- Postma, D.F.; van Werkhoven, C.H.; van Elden, L.J.; Thijsen, S.F.; Hoepelman, A.I.; Kluytmans, J.A.; Boersma, W.G.; Compaijen, C.J.; van der Wall, E.; Prins, J.M.; et al. CAP-START Study Group Antibiotic treatment strategies for community-acquired pneumonia in adults. N. Engl. J. Med. 2015, 372, 1312–1323. [CrossRef] [PubMed]
- Wertzel, H.; Swoboda, L.; Joos-Würtemberger, A.; Frank, U.; Hasse, J. Perioperative antibiotic prophylaxis in general thoracic surgery. *Thorac. Cardiovasc. Surg.* 1992, 40, 326–329. [CrossRef]
- Yamazaki, S.; Yamagishi, K.; Murata, S.; Yokoyama, I.; Yahaba, M.; Takayanagi, S.; Kawasaki, Y.; Taniguchi, T.; Ishii, I.; Igari, H. Antibiotics prescriptions for pneumonia analyzed by claim information in Japan. *Int. J. Clin. Pharmacol. Ther.* 2021, 59, 289–297. [CrossRef] [PubMed]
- Mikasa, K.; Aoki, N.; Aoki, Y.; Abe, S.; Iwata, S.; Ouchi, K.; Kasahara, K.; Kadota, J.; Kishida, N.; Kobayashi, O.; et al. JAID/JSC Guidelines for the Treatment of Respiratory Infectious Diseases: The Japanese Association for Infectious Diseases/Japanese Society

of Chemotherapy-The JAID/JSC Guide to Clinical Management of Infectious Disease/Guideline-preparing Committee Respiratory Infectious Disease WG. J. Infect. Chemother. 2021, 22, S1–S65.

- Hasegawa, S.; Shiraishi, A.; Yaegashi, M.; Hosokawa, N.; Morimoto, K.; Mori, T. Ceftriaxone versus ampicillin/sulbactam for the treatment of aspiration-associated pneumonia in adults. J. Comp. Eff. Res. 2019, 8, 1275–1284. [CrossRef]
- Song, J.Y.; Cheong, H.J.; Choi, W.S.; Heo, J.Y.; Noh, J.Y.; Kim, W.J. Clinical and micro-biological characterization of carbapenemresistant *Acinetobacter baumannii* bloodstream infections. *J. Med. Microbiol.* 2011, 60, 605–611. [CrossRef]
- Gaynes, R.; Edwards, J.R. National Nosocomial Infections Surveillance System Overview of nosocomial infections caused by Gramnegative bacilli. *Clin. Infect. Dis.* 2005, 41, 848–854.
- 10. Peleg, A.Y.; Seifert, H.; Paterson, D.L. Acinetobacter baumannii emergence of a successful pathogen. Clin. Microbiol. Rev. 2008, 21, 538–582. [CrossRef]
- Corbella, X.; Ariza, J.; Ardanuy, C.; Vuelta, M.; Tubau, F.; Sora, M.; Pujol, M.; Gudiol, F. Efficacy of sulbactam alone and in combination with ampicillin in nosocomial infections caused by multiresistant *Acinetobacter baumannii*. J. Antimicrob. Chemother. 1998, 42, 793–802. [CrossRef]
- Pfizer, Incorporated. Unasyn-S (Ampicillin Sodium/Sulbactam Sodium for Injection) Prescribing Information. Available online: https://www. info.pmda.go.jp/go/interview/3/672212_6139504F1022_3_1F.pdf (accessed on 22 January 2023).
- Craig, W.A. Pharmacokinetic/pharmacodynamic parameters: Rationale for antibacterial dosing of mice and men. *Clin. Infect. Dis.* 1998, 26, quiz 11–12. [CrossRef]
- 14. Craig, W.A. Does the dose matter? *Clin. Infect. Dis.* **2001**, *33*, S233–S237. [CrossRef] [PubMed]
- Soto, E.; Shoji, S.; Muto, C.; Tomono, Y.; Marshall, S. Population pharmacokinetics of ampicillin and sulbactam in patients with community-acquired pneumonia: Evaluation of the impact of renal impairment. *Br. J. Clin. Pharmacol.* 2014, 77, 509–521. [CrossRef] [PubMed]
- Frank, U.; Schmidt-Eisenlohr, E.; Joos-Württemberger, A.; Hasse, J.; Daschner, F. Concentrations of sulbactam/ampicillin in serum and lung tissue. *Infection* 1990, 18, 307–309. [CrossRef] [PubMed]
- Valcke, Y.J.; Rosseel, M.T.; Pauwels, R.A.; Bogaert, M.G.; Van der Straeten, M.E. Penetration of ampicillin and sulbactam in the lower airways during respiratory infections. *Antimicrob. Agents. Chemother.* 1990, 34, 958–962. [CrossRef]
- 18. Kawasaki, K.; Niimi, H.; Ushirosako, T.; Matsunaga, T. Antibacterial activity of sulbactam ampicillin. Chemotherapy 1988, 36, 34–57.
- Onita, T.; Ikawa, K.; Nakamura, K.; Nishikawa, G.; Kobayashi, I.; Ishihara, N.; Tamaki, H.; Yano, T.; Naora, K.; Morikawa, N. Prostatic pharmacokinetic/pharmacodynamic evaluation of ampicillin-sulbactam for bacterial prostatitis and preoperative prophylaxis. *J. Clin. Pharmacol.* 2021, *61*, 820–831. [CrossRef]
- Yokoyama, Y.; Matsumoto, K.; Ikawa, K.; Watanabe, E.; Morikawa, N.; Takeda, Y. Population pharmacokinetic-pharmacodynamic target attainment analysis of sulbactam in patients with impaired renal function: Dosing considerations for *Acinetobacter baumannii* infections. J. Infect. Chemother. 2015, 21, 284–289. [CrossRef]
- Suzuki, T.; Sugiyama, E.; Nozawa, K.; Tajima, M.; Takahashi, K.; Yoshii, M.; Suzuki, H.; Sato, V.H.; Sato, H. Effects of dosing frequency on the clinical efficacy of ampicillin/sulbactam in Japanese elderly patients with pneumonia: A single- center retrospective observational study. *Pharmacol. Res. Perspect.* 2021, 9, e00746. [CrossRef]
- 22. Betrosian, A.P.; Frantzeskaki, F.; Xanthaki, A.; Georgiadis, G. High-dose ampicillin-sulbactam as an alternative treatment of late-onset VAP from multidrug-resistant *Acinetobacter baumannii*. *Scand. J. Infect. Dis.* **2007**, *39*, 38–43. [CrossRef]
- Gilbert, D.N.; Chambers, H.F.; Saag, M.S.; Pavia, A.T.; Boucher, H.W. The Sanford Guide to Antimicrobial Therapy 51th ed. Sanford Guide; Life Science Publishing: Tokyo, Japan, 2021; pp. 70–71.
- Langdon, G.; Gueorguieva, I.; Aarons, L.; Karlsson, M. Linking preclinical and clinical whole-body physiologically based pharmacokinetic models with prior distributions in NONMEM. *Eur. J. Clin. Pharmacol.* 2007, 63, 485–498. [CrossRef]
- Brown, R.P.; Delp, M.D.; Lindstedt, S.L.; Rhomberg, L.R.; Beliles, R.P. Physiological parameter values for physiologically based pharmacokinetic models. *Toxicol. Ind. Health.* 1997, 13, 407–484. [CrossRef]
- Marchand, S.; Chenel, M.; Lamarche, I.; Couet, W. Pharmacokinetic modeling of free amoxicillin concentrations in rat muscle extracellular fluids determined by microdialysis. *Antimicrob Agents Chemother*. 2005, 49, 3702–3706. [CrossRef]
- Germovsek, E.; Lutsar, I.; Kipper, K.; Karlsson, M.O.; Planche, T.; Chazallon, C.; Meyer, L.; Trafojer, U.M.T.; Metsvaht, T.; Fournier, I.; et al. Plasma and CSF pharmacokinetics of meropenem in neonates and young infants: Results from the NeoMero studies. J. Antimicrob. Chemother. 2018, 73, 1908–1916. [CrossRef]
- Sasongko, L.; Williams, K.M.; Day, R.O.; McLachlan, A.J. Human subcutaneous tissue distribution of fluconazole: Comparison of microdialysis and suction blister techniques. *Br. J. Clin. Pharmacol.* 2003, *56*, 551–561. [CrossRef]
- Ohata, Y.; Tomita, Y.; Sunakawa, K.; Drusano, G.L.; Tanigawara, Y. Cerebrospinal pharmacokinetic and pharmacodynamic analysis of efficacy of meropenem in paediatric patients with bacterial meningitis. *Int. J. Antimicrob. Agents* 2019, 54, 292–300. [CrossRef]
- Jacobs, M.R. Optimisation of antimicrobial therapy using pharmacokinetic and pharmacodynamic parameters. *Clin. Microbiol. Infect.* 2001, 7, 589–596. [CrossRef]
- 31. Drusano, G.L. Prevention of resistance: A goal for dose selection for antimicrobial agents. Clin. Infect. Dis. 2003, 36, S42–S50. [CrossRef]
- Clinical and Laboratory Standards Institute-CLSI. M100-ED31:2021 Performance Standards for Antimicrobial Susceptibility Testing, 31st Edition. Available online: http://em100.edaptivedocs.net/dashboard.aspx (accessed on 16 June 2021).

- Yokoyama, Y.; Matsumoto, K.; Ikawa, K.; Watanabe, E.; Shigemi, A.; Umezaki, Y.; Nakamura, K.; Ueno, K.; Morikawa, N.; Takeda, Y. Pharmacokinetic/pharmacodynamic evaluation of sulbactam against *Acinetobacter baumannii* in in vitro and murine thigh and lung infection models. *Int. J. Antimicrob. Agents* 2014, 43, 547–552. [CrossRef]
- Tateda, K.; Ohno, A.; Ishii, Y.; Murakami, H.; Yamaguchi, K. Surveillance of in vitro susceptibilities to levofloxacin and various antibacterial agents for 11,705 clinical isolates obtained from 65 centers in 2016. *Jpn. J. Antibiotics.* 2018, 298, 71–76.
- 35. Yanagihara, K.; Matsumoto, T.; Tokimatsu, I.; Tsukada, H.; Fujikura, Y.; Miki, M.; Morinaga, Y.; Sato, J.; Wakamura, T.; Kiyota, H.; et al. Nationwide surveillance of bacterial respiratory pathogens conducted by the surveillance committee of japanese society of chemotherapy, the japanese association for infectious diseases, and the Japanese society for clinical microbiology in 2016: General view of the pathogens' antibacterial susceptibility. *J. Infect. Chemother.* 2020, *26*, 873–881. [PubMed]
- Kaneko, A.; Yamane, N.; Watanabe, D.; Mizusawa, N.; Matsuzaki, K.; Hasegawa, M.; Sato, Y.; Kobayashi, I. Treatment of aspiration pneumonia based on the antimicrobial susceptibility pattern of oral bacterial pathogens. *Jpn. J. Chemother.* 2007, 55, 378–381.

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