

**Table S1.** Bacterial strains and plasmids used in the study

Bacterial Strains/ Plasmids	Genotype/ relevant characteristics	Reference
<b><i>Pseudomonas aeruginosa</i> reference strains</b>		
PAO1	Laboratory reference strain	[1]
<b>PAO1 mutants</b>		
$\Delta mexZ$	Deletion of <i>mexZ</i> gene in PAO1	[2]
$\Delta mexXY$	Deletion of <i>mexXY</i> genes in PAO1	[2]
<i>mexY</i> <sub>G287S</sub>	Engineered mutation in the genome of PAO1 that replaced amino acid G with S at <i>mexY</i> codon 287	This study
<i>fusAI</i> <sub>R680C</sub>	Engineered mutation in the genome of PAO1 that replaced amino acid R with C at codon 680 of <i>fusAI</i>	This study
<i>fusAI</i> <sub>Y683D</sub>	PAO1 with spontaneous mutation that replaced amino acid Y with D at codon 683 of <i>fusAI</i>	[3]
<i>fusAI</i> <sub>D327S</sub>	PAO1 with spontaneous mutation that replaced amino acid D with S at codon 327 of <i>fusAI</i>	[3]
<i>amgS</i> <sub>V121G</sub>	Engineered mutation in the genome of PAO1 that replaced amino acid V with G at codon 121 of <i>amgS</i>	This study
$\Delta mexZ$ <i>mexY</i> <sub>G287S</sub>	$\Delta mexZ$ <i>mexY</i> <sub>G287S</sub> double mutant	This study
$\Delta mexZ$ <i>fusAI</i> <sub>R680C</sub>	$\Delta mexZ$ <i>fusAI</i> <sub>R680C</sub> double mutant	This study
$\Delta mexXY$ <i>fusAI</i> <sub>R680C</sub>	$\Delta mexXY$ <i>fusAI</i> <sub>R680C</sub> double mutant	This study
$\Delta mexZ$ <i>fusAI</i> <sub>Y683D</sub>	$\Delta mexZ$ <i>fusAI</i> <sub>Y683D</sub> double mutant	This study
$\Delta mexZ$ <i>fusAI</i> <sub>D327S</sub>	$\Delta mexZ$ <i>fusAI</i> <sub>D327S</sub> double mutant	This study
$\Delta mexZ$ <i>amgS</i> <sub>V121G</sub>	$\Delta mexZ$ <i>amgS</i> <sub>V121G</sub> double mutant	This study
$\Delta mexXY$ <i>amgS</i> <sub>V121G</sub>	$\Delta mexXY$ <i>amgS</i> <sub>V121G</sub> double mutant	This study
<i>mexY</i> <sub>G287S</sub> <i>amgS</i> <sub>V121G</sub>	<i>mexY</i> <sub>G287S</sub> <i>amgS</i> <sub>V121G</sub> double mutant	This study
<i>mexY</i> <sub>G287S</sub> <i>fusAI</i> <sub>R680C</sub>	<i>mexY</i> <sub>G287S</sub> <i>fusAI</i> <sub>R680C</sub> double mutant	This study
<i>fusAI</i> <sub>R680C</sub> <i>amgS</i> <sub>V121G</sub>	<i>fusAI</i> <sub>R680C</sub> <i>amgS</i> <sub>V121G</sub> double mutant	This study
$\Delta mexZ$ <i>amgS</i> <sub>V121G</sub> <i>fusAI</i> <sub>R680C</sub>	$\Delta mexZ$ <i>amgS</i> <sub>V121G</sub> <i>fusAI</i> <sub>R680C</sub> triple mutant	This study
$\Delta mexXY$ <i>amgS</i> <sub>V121G</sub> <i>fusAI</i> <sub>R680C</sub>	$\Delta mexXY$ <i>amgS</i> <sub>V121G</sub> <i>fusAI</i> <sub>R680C</sub> triple mutant	This study
$\Delta mexZ$ <i>fusAI</i> <sub>R680C</sub> <i>mexY</i> <sub>G287S</sub>	$\Delta mexZ$ <i>fusAI</i> <sub>R680C</sub> <i>mexY</i> <sub>G287S</sub> triple mutant	This study
$\Delta mexZ$ <i>amgS</i> <sub>V121G</sub> <i>mexY</i> <sub>G287S</sub>	$\Delta mexZ$ <i>amgS</i> <sub>V121G</sub> <i>mexY</i> <sub>G287S</sub> triple mutant	This study

<i>fusAI</i> <sub>R680C</sub> <i>amgS</i> <sub>V121G</sub> <i>mexY</i> <sub>G287S</sub>	<i>fusAI</i> <sub>R680C</sub> <i>amgS</i> <sub>V121G</sub> <i>mexY</i> <sub>G287S</sub> triple mutant	This study
<i>ΔmexZ fusAI</i> <sub>R680C</sub> <i>amgS</i> <sub>V121G</sub> <i>mexY</i> <sub>G287S</sub>	<i>ΔmexZ fusAI</i> <sub>R680C</sub> <i>amgS</i> <sub>V121G</sub> <i>mexY</i> <sub>G287S</sub> quadruple mutant	This study

#### Strains of *Escherichia coli*

JM83	<i>ara</i> , <i>Δ(lac-proAB) rpsL(strA)</i> , <i>φ80</i> , <i>lacZΔM15</i>	[4]
ST18	S 17 <i>λpirΔhemA</i>	[5]

#### Plasmids used in the study

pEX18Tc	pMB1 replicon, <i>oriT</i> , <i>sacB</i> , Tc <sup>R</sup> , allelic exchange vector	[6]
pEX18Tc:: <i>ΔmexXY</i>	pEX18Tc derivative carrying <i>mexXY</i> ( <i>PA2018-PA2019</i> ) flanking regions	[2]
pEX18Tc:: <i>ΔmexZ</i>	pEX18Tc derivative carrying <i>mexZ</i> ( <i>PA2020</i> ) flanking regions	[2]
pEX18Tc:: <i>mexY</i> <sub>G287S</sub>	pEX18Tc containing DNA fragment with a mutation that encodes amino acid S instead of G at residue 287 in MexY	This study
pEX18Tc:: <i>fusAI</i> <sub>R680C</sub>	pEX18Tc containing DNA fragment with a mutation that encodes amino acid C instead of R at residue 680 in FusA1	This study
pEX18Tc:: <i>amgS</i> <sub>V121G</sub>	pEX18Tc containing DNA fragment with a mutation that encodes amino acid G instead of V at residue 121 in AmgS	This study

**Table S2.** Primers used in the study.

Primers	Sequences	Reference
<b><i>mexXY</i> deletion Primers</b>		
<i>mexXY_F1</i>	5' CCCCC <u>GGTACC</u> GAGTCGGCTGATG ACCTACA 3'	[2]
<i>mexXY_R1</i>	5'GGGGG <u>TCTAGA</u> GTACCGCTGTTCTTCCTGGT 3'	[2]
<i>mexXY_F2</i>	5' CCCCC <u>TCTAGAT</u> GTCCCTCGATTC GTGAACT 3'	[2]
<i>mexXY_R2</i>	5' GGGGG <u>AAGCTT</u> GCTCTACATCGAC GGCAAG 3'	[2]
<b><i>mexXY</i> deletion screening Primers</b>		
<i>PA2017_F</i>	5' GCA GCC TGT ACG TGG TCA 3'	[2]
<i>mexZ_R</i>	5' GGG TTT TCT GGG ATT CCT CT 3'	[2]
<b><i>mexZ</i> deletion Primers</b>		
<i>mexX_F</i>	5' CCCCC <u>GAATTC</u> GTTCTCGACGATCACCCACT 3'	[2]
<i>mexX_R</i>	5' GGGGG <u>TCTAGA</u> GGGTTTTCTGGGATTCCTCT 3'	[2]
<i>PA2022_F</i>	5' CCCCC <u>TCTAGA</u> CGCAGTTCTCCCTACCTGTT 3'	[2]
<i>PA2022_R</i>	5' GGGGG <u>AAGCTT</u> CGCAGTATCTGGCTGTCGTA 3'	[2]
<b><i>mexZ</i> deletion screening Primers</b>		
<i>mexZ_SR_F</i>	5' GTGTCCCTCGATTCGTGAAC 3'	[2]
<i>mexZ_SR_R</i>	5' CGTGAAGCTACCGTGACAGA 3'	[2]
<b><i>mexY</i><sub>G287S</sub> mutation engineering primers</b>		
<i>mexY_F</i>	5' GGGGA <u>AATTC</u> CCTGTTCCGCAATCCGCATC 3'	This study
<i>mexY_R</i>	5'GGGGG <u>ATCCAGT</u> CCTTCAGGGTGGCGAAG 3'	This study
<b><i>mexY</i><sub>G287S</sub> mutation screening primers</b>		
<i>mexY_scr_F</i>	5' CTCGTCCAACGTGTTGCAGG 3'	This study
<i>mexY_scr_R</i>	5' AACGCCGAGGTGTCATAGGG 3'	This study
<b><i>fusA1</i><sub>R680C</sub> mutation engineering primers</b>		
<i>fusA1_F</i>	5' GGGGG <u>TCTAGA</u> TTACTCGATGATCTTGGAAC 3'	This study
<i>fusA1_R</i>	5' CCCCC <u>AAGCTT</u> GAAGCCGAGATCAAGGAAGG 3'	This study
<b><i>fusA1</i><sub>R680C</sub> mutation screening primers</b>		
<i>fusA1_scr_F</i>	5' GTATTCAACGTGCGAGGTGT 3'	This study

<i>fusAI_scr_R</i>	5' GTTCAAGATCGCTGCTTCCA 3'	This study
<b><i>amgS</i><sub>V121G</sub> mutation engineering primers</b>		
<i>amgS_F</i>	5'CCCCC <u>GAATTC</u> TTCTCCGTAAGCACGAGAGG 3'	This study
<i>amgS_R</i>	5' CCCCC <u>TCTAGA</u> AGCAGATGGATCGCCTTCTA 3'	This study
<b><i>amgS</i><sub>V121G</sub> mutation screening primers</b>		
<i>amgS_scr_F</i>	5' ATACACCTCGGCCATTTCAC 3'	This study
<i>amgS_scr_R</i>	5' CTGATGAACGAGGACGTGAT 3'	This study
<b>RT-qPCR Primers</b>		
<i>mexX_RT_F</i>	5' GGC CCT GGT CGC CCT ATT C 3'	[7]
<i>mexX_RT_R</i>	5' TCC TCG TAC AGG CGA CGG 3'	[7]
<i>clpX_F</i>	5' GTG GGC GAG GAT GTC GAG AAC 3'	[7]
<i>clpX_R</i>	5' CGG TAC CCT CGA TGA GCT TCA G 3'	[7]
<i>oprL_F</i>	5' CCA ACA GCG GTG CCG TTG A 3'	[7]
<i>oprL_R</i>	5' GCC ATA TTG TAC TCG CGG GT 3'	[7]
<b>Universal primers</b>		
M13_F	GTA AAA CGA CGG CCA GT	Universal
M13_R	CAG GAA ACA GCT ATG AC	Universal

<sup>a</sup>Introduced restriction sites are underlined.

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