

SspH, a novel HATPase family regulator, controls antibiotic biosynthesis in *Streptomyces*

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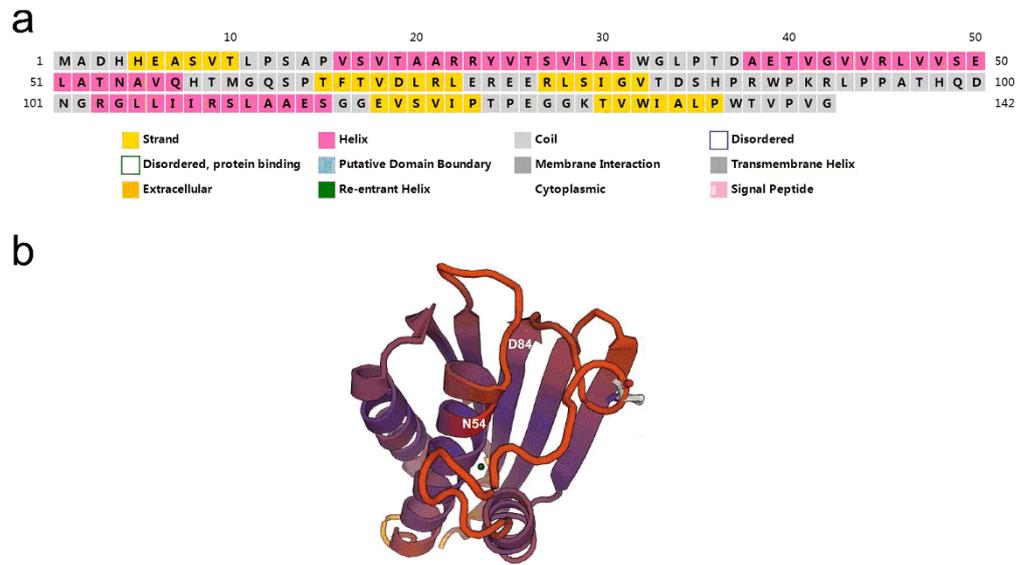


Figure S1. Secondary and three-dimensional (3D) structure of SspH. **(a)** Secondary structure of SspH. **(b)** Putative 3D structure. The homologous three - dimensional (3D) structure model for SspH was constructed based on the *Bacillus* anti - sigma factor SpoIIAB.

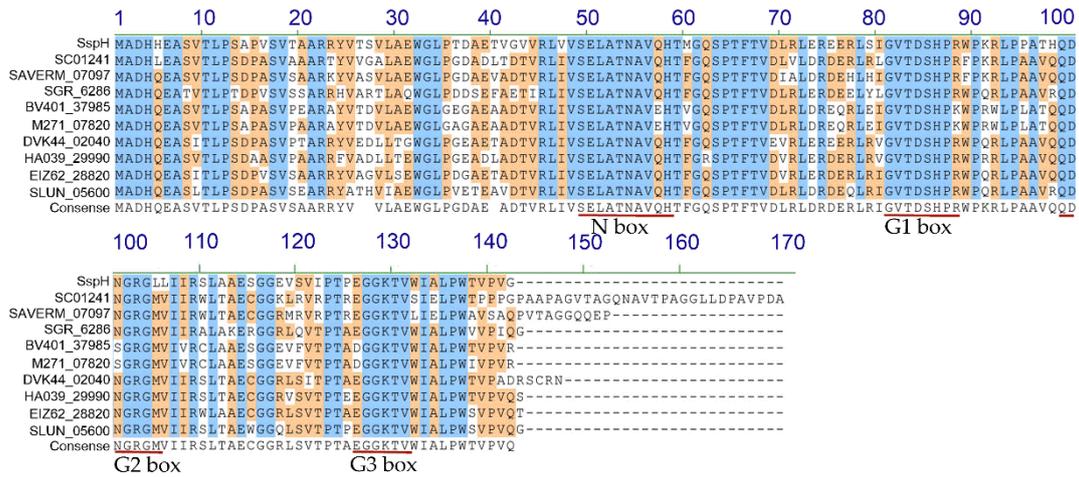


Figure S2. Amino acid sequence alignment of SspH and its orthologs. Identical or similar residues in all sequences are highlighted in blue and orange, respectively. The corresponding assumed conserved motifs are marked in red line. SCO1241, SspH ortholog from *S. coelicolor*; SAVERM_7097, SspH ortholog from *S. avermitilis*; SGR_6286, SspH ortholog from *S. griseus*, BV401_37985, SspH ortholog from *S. autolyticus*; M271_07820, SspH ortholog from *S. rapamycinicus*; DVK44_02040, SspH ortholog from *S. paludis*, HA039_29990, SspH ortholog from *S. liangshanensis*; EIZ62_28820, SspH ortholog from *S. ficellus*; SLUN_05600, SspH ortholog from *S. lunaelactis*.

Table S1. Genes whose products contain only the HATPase domain in several *Streptomyces* species

Strain	Gene ID
<i>S. coelicolor</i>	<i>sco0673, sco0702, sco0868, sco1241, sco2864, sco3066, sco3423, sco3724, sco3930, sco4412, sco4677, sco4941, sco5103, sco5460, sco5915, sco5978, sco6130, sco6156, sco6237, sco6749, sco6949, sco7313, sco7328, sco7446, sco7328, sco7614</i>
<i>S. avermitilis</i>	<i>saverm_0234, saverm_1437, saverm_1664, saverm_2076, saverm_2174, saverm_2785, saverm_2797, saverm_3186, saverm_3424, saverm_3488, saverm_3738, saverm_3761, saverm_3991, saverm_4086, saverm_4225, saverm_4398, saverm_4615, saverm_4783, saverm_5014, saverm_5201, saverm_5409, saverm_5755, saverm_6252, saverm_7097</i>
<i>S. bingchenggensis</i>	<i>sbi_02080, sbi_02469, sbi_02795, sbi_03226, sbi_03585, sbi_03656, sbi_03703, sbi_03978, sbi_04142, sbi_04846, sbi_05023, sbi_05275, sbi_05655, sbi_05899, sbi_06283, sbi_06400, sbi_06722, sbi_07227, sbi_08076, sbi_08867, sbi_09291</i>

Table S2. Amino acid sequences alignment of homologous proteins from *S. bingchenggensis* and other *Streptomyces* species

Proteins	Coverage/identity(%) to genes of
SBI_05734	<i>S. coelicolor</i> ; BldC (SCO4091): 100/100
SBI_03959	<i>S. avermitilis</i> ; WbIE (SAVERM_3016): 78/100
SBI_05953	<i>S. venezuelae</i> ; Lsr2 (SVEN_3225): 99/66
SBI_03799	<i>S. coelicolor</i> ; SCO5351: 84/81
SBI_06268	<i>S. coelicolor</i> ; BldM (SCO4768): 100/100
SBI_05811	<i>S. coelicolor</i> ; WblA (SCO3579): 100/89
SBI_04164	<i>S. coelicolor</i> ; WblI (SCO5046): 76/86

Table S3. Number of hits of SspH homologous sequence in different bacteria after PSI-BLAST search

Organism	Number of hits
<i>Streptomyces</i>	1379
<i>Kitasatospora</i>	3
<i>Streptosporangium nondiastaticum</i>	2
<i>Actinospica acidiphila</i>	6
<i>unclassified Actinomycetia</i>	4
<i>Mesorhizobium</i> sp. B2-3-3 (non-Actinomycetia)	1

Table S4. Three milbemycin biosynthetic genes (*milR*, *milF* and *milA1*) and their functions

Gene ID	Type of product	Function	Reference
<i>sbi_00734</i> (<i>milR</i>)	MilR	LAL family regulator; a cluster-situated key activator of milbemycin biosynthesis; activating the transcription of the <i>milA4-E</i> operon and <i>milF</i> directly and activating the transcription of the other milbemycin biosynthetic structural genes indirectly.	[1]
<i>sbi_00731</i> (<i>milF</i>)	MilF	The tailoring enzyme gene encoding C5-ketoreductase responsible for the ketonization of milbemycins.	[2]
<i>sbi_00789</i> (<i>milA1</i>)	MilA1	The milbemycin polyketide synthase (PKS) gene; containing the loading module and two extension modules responsible for polyketide chain extension.	[3]

Table S5. 31 antibiotic biosynthetic core genes and their putative functions

Biosynthetic core gene	Putative function	Predicted product type
<i>sbi_00140</i>	amino acid adenylation domain-containing protein	PKS-NRPS hybrid for moenomycin
<i>sbi_00220</i>	putative squalene-hopene cyclase	Terpene
<i>sbi_00319</i>	modular polyketide synthase	PKS-NRPS hybrid for leinamycin
<i>sbi_00522</i>	modular polyketide synthase	PKS-NRPS hybrid for BE-14106
<i>sbi_00625</i>	amino acid adenylation domain-containing protein	NRPS
<i>sbi_00655</i>	modular polyketide synthase	Type I PKS for pimaricin
<i>sbi_00671</i>	putative mixed polyketide synthase/non-ribosomal peptide synthetase	Type I PKS for meridamycin
<i>sbi_00822</i>	non-ribosomal peptide synthetase	NRPS
<i>sbi_00956</i>	non-ribosomal peptide synthetase	PKS-NRPS hybrid
<i>sbi_01029</i>	putative polyketide synthase	Type I PKS
<i>sbi_01080</i>	pentalenene synthase	Terpene
<i>sbi_01365</i>	amino acid adenylation domain-containing protein	PKS-NRPS hybrid for lasalocid
<i>sbi_01540</i>	putative polyketide synthase	Type I PKS for borrelidin
<i>sbi_01983</i>	modular polyketide synthase	Type I PKS for mycinamicin
<i>sbi_02068</i>	putative germacradienol synthase	Terpene
<i>sbi_02232</i>	group-specific protein	Cyclothiazomycin
<i>sbi_02590</i>	squalene/phytoene synthase	Terpene for hopanoid
<i>sbi_02764</i>	3-oxoacyl-ACP synthase I	Type II PKS for grey spore pigment
<i>sbi_02988</i>	amino acid adenylation domain protein	PKS-NRPS for calcium-dep antibiotic

Biosynthetic core gene	Putative functions	Predicted product type
<i>sbi_03179</i>	type I modular polyketide synthase	PKS-NRPS hybrid for borrelidin
<i>sbi_06052</i>	amino acid adenylation domain-containing protein	PKS-NRPS hybrid
<i>sbi_06449</i>	non-ribosomal peptide synthetase	Desferrioxamin
<i>sbi_06799</i>	non-ribosomal peptide synthetase	NRPS for melanin
<i>sbi_06873</i>	glycosyl transferase	Type II PKS for Polyketomycin
<i>sbi_08414</i>	modular polyketide synthase	Nanchangmycin (by type I PKS)
<i>sbi_08957</i>	2-methylisoborneol synthase	Terpene
<i>sbi_09057</i>	putative non-ribosomal peptide synthase	Melanin
<i>sbi_09195</i>	modular polyketide synthase	PKS-NRPS hybrid for lymphostin
<i>sbi_09249</i>	non-ribosomal peptide synthetase	NRPS for moenomycin
<i>sbi_09652</i>	non-ribosomal peptide synthetase	Oxazolomycin (by PKS-NRPS hybrid)
<i>sbi_10015</i>	putative type I polyketide synthase WcbR	Type I PKS

Table S6. Strains used in this work

Strain	Characteristics	Source
<i>Escherichia coli</i>		
<i>E. coli</i> JM109	Host for all plasmids cloning procedures	Novagen
ET12567 (pUZ8002)	Donor strain for conjugation between <i>E. coli</i> and <i>Streptomyces</i>	[1]
<i>Streptomyces bingchenggensis</i>		
BC04	Milbemycin high-yield strain obtained by random mutagenesis	[4]
BC04-C1	BC04 carrying pSET152	This work
BC04-C2	BC04 carrying pSETddCpf1	This work
BC04/OsspH	BC04 carrying pSET152::P _{hrdB} ::sspH	This work
BC04/RsspH	BC04 carrying pSETddCpf1::sspH	This work
BC04/O1241	BC04 carrying pSET152::P _{hrdB} ::1241	This work
BC04/O7097	BC04 carrying pSET152::P _{hrdB} ::7097	This work
<i>Streptomyces coelicolor</i>		
M145	Actinorhodin producer	[1]
M145/OsspH	M145 carrying pSET152::P _{hrdB} ::sspH	This work
M145/O1241	M145 carrying pSET152::P _{hrdB} ::1241	This work
M145/O7097	M145 carrying pSET152::P _{hrdB} ::7097	This work
<i>Streptomyces avermitilis</i>		
NEAU12	Avermectin high-yield producer	Laboratory storage
NEAU12/OsspH	NEAU12 carrying pSET152::P _{hrdB} ::sspH	This work
NEAU12/O1241	NEAU12 carrying pSET152::P _{hrdB} ::1241	This work
NEAU12/O7097	NEAU12 carrying pSET152::P _{hrdB} ::7097	This work

Table S7. Plasmids used in this work

Names	Characteristics	Source
pSET152	Apr ^R , integrative <i>E. coli-Streptomyces</i> shuttle vector	[1]
pSET152::P _{hrdB}	pSET152 containing P _{hrdB}	This work
pSET152::P _{hrdB} ::sspH	pSET152::P _{hrdB} containing <i>sspH</i>	This work
pSET152::P _{hrdB} ::1241	pSET152::P _{hrdB} containing <i>sco1241</i>	This work
pSET152::P _{hrdB} ::7097	pSET152::P _{hrdB} containing <i>saverm_7097</i>	This work
pSETddCpf1	pSET152 with the <i>scocpf1</i> (E1006A) gene under the control of the <i>ermE</i> * <i>p</i> promoter	[5]
pSETddCpf1::sspH	pSETddCpf1 containing the crRNA transcription cassette for editing <i>sspH</i>	This work
pBluescript II KS (+)	Amp ^R , <i>E. coli</i> cloning vector	Stratagene
pBluescript II KS(+):P _{hrdB} ::sspH	pBluescript II KS(+) containing <i>sspH</i> under the control of P _{hrdB}	This work
pBluescript II KS(+):P _{hrdB} ::N54A	pBluescript II KS(+) containing mutagenized <i>sspH</i> (mutation of Asn54 to Ala) under the control of P _{hrdB}	This work
pBluescript II KS(+):P _{hrdB} ::D84A	pBluescript II KS(+) containing mutagenized <i>sspH</i> (mutation of Asp84 to Ala) under the control of P _{hrdB}	This work
pSET152::P _{hrdB} ::N54A	pSET152 containing mutagenized <i>sspH</i> (mutation of Asn54 to Ala) under the control of P _{hrdB}	This work
pSET152::P _{hrdB} ::D84A	pSET152 containing mutagenized <i>sspH</i> (mutation of Asp84 to Ala) under the control of P _{hrdB}	This work

Table S8. Primers used in this work

Name	Sequence (5'-3')	Usage
sspH-F	GCCGTTTTTCAACGTTCCGAGAGGTTGTTCATGGCTGACCACCACGAAGCA	
sspH-R	T CGACTCTAGAGGATCCGCGGCCGCGCGGATCACCACGGGGCACGTCAAG C	Construction of pSET152::P _{hrdB} ::sspH
ddCpf1-sspH-F	GACTAGTCGTTCCAGCCGAGATCCACCGTATCTACAACAGTAGAAATTTG	
ddCpf1-R	G	Construction of pSETddCpf1::sspH
sco1241-F	GGAATTCATATGTGGATCCTACCAACCGGCACGATT	
sco1241-R	GCCGTTTTTCAACGTTCCGAGAGGTTGTTCATGGCAGACCATCTGGAAGCA TCC CGACTCTAGAGGATCCGCGGCCGCGCGGATAAGGGCCGGATCTGGCAGC C	Construction of pSET152::P _{hrdB} ::1241
saverm_7097-F	GCCGTTTTTCAACGTTCCGAGAGGTTGTTCATGGCAGATCACCAGGAAGC ATCCG	
saverm_7097-R	CGACTCTAGAGGATCCGCGGCCGCGCGGATTACCCGGACGGCCGAACA CC	Construction of pSET152::P _{hrdB} ::7097
N54A-F	GCCGCCGTCCAGCACACGATGGGCCAGTCGCC	
N54A-R	GGTGGCGAGTTCGGAGACGACGAGCCGTACGA	Construction of pSET152::P _{hrdB} ::D54A
D84A-F	GCCAGCCACCCCAGATGGCCCAAACGCCTGCC	
D84A-R	GGTGACGCCGATGCTCAGCCGCTCTTCGCGTT	Construction of pSET152::P _{hrdB} ::D84A

Name	Sequence (5'-3')	Usage
Q16S-F	TGTCGTGAGATGTTGGGTTAAG	qRT-PCR analysis of 16S rRNA
Q16S-R	TCATTGTACCGGCCATTGTAG	
QmilA1-F	GCCGTATCGTCTTCGTCTTC	qRT-PCR analysis of <i>milA1</i>
QmilA1-R	TGGAGGATGTCCGTGAGT	
QmilR-F	GCAAGAGCGAATTCCTGAAG	qRT-PCR analysis of <i>milR</i>
QmilR-R	CTGTGACGGAGCGAGTC	
QmilF-F	AGGTCACTCAGGAGGAGTT	qRT-PCR analysis of <i>milF</i>
QmilF-R	GAGGAGATGTTGACGATCCG	
QsspH-F	GGCTGGAACGCGAAGAG	qRT-PCR analysis of <i>sspH</i>
QsspH-R	CCGACCGTTGTCCTGATG	
Qsbi_00140-F	GGGAGCGTGTCGAAGAAG	qRT-PCR analysis of <i>sbi_00140</i>
Qsbi_00140-R	GCACACCGAGTGATGTATGT	
Qsbi_00220-F	CTGTCCGTCTTCTGCTATGTC	qRT-PCR analysis of <i>sbi_00220</i>
Qsbi_00220-R	CGAGCATGGTCGGATTGT	
Qsbi_00319-F	TGTACGTCAACACCACCAC	qRT-PCR analysis of <i>sbi_00319</i>
Qsbi_00319-R	AGTACGGCGTGCACATT	
Qsbi_00522-F	AAAGGCACGATCCGTACAC	qRT-PCR analysis of <i>sbi_00522</i>
Qsbi_00522-R	GGCCTCTTTCGGCGATATAC	
Qsbi_00625-F	CTTCGCCAACTACTTCCACTC	qRT-PCR analysis of <i>sbi_00625</i>
Qsbi_00625-R	CCATGAACATGTCGAAGGAGAA	

Name	Sequence (5'-3')	Usage
Qsbi_00655-F	AGCTACCAGGACTACACCA	qRT-PCR analysis of <i>sbi_00655</i>
Qsbi_00655-R	CTCGAGCCCGAACACATAG	
Qsbi_00671-F	GGCGTGATGAGCAACGATTA	qRT-PCR analysis of <i>sbi_00671</i>
Qsbi_00671-R	CCGGTGAAGTCGAAGAAGTG	
Qsbi_00822-F	CTGCCGCTGTTCCCTATATC	qRT-PCR analysis of <i>sbi_00822</i>
Qsbi_00822-R	AATTCATCGGGCCGGTTC	
Qsbi_00956-F	CACCGTACGTAACGCGAAAT	qRT-PCR analysis of <i>sbi_00956</i>
Qsbi_00956-R	CGTAGAGGTCTGTTGCGTATTC	
Qsbi_01029-F	CACCTGGTGTACGTCGAAAT	qRT-PCR analysis of <i>sbi_01029</i>
Qsbi_01029-R	AGGCCGTCCATGCTTTC	
Qsbi_01080-F	ACGGAGTATCTGCACTCCTA	qRT-PCR analysis of <i>sbi_01080</i>
Qsbi_01080-R	CGAACCGCTCGTTGAGAT	
Qsbi_01365-F	GAGAGATCGAGGAGTGGGAA	qRT-PCR analysis of <i>sbi_01365</i>
Qsbi_01365-R	GGTCGAAAGCGGTCTTGT	
Qsbi_01540-F	CTGTGGAGTTGCTGGATTCT	qRT-PCR analysis of <i>sbi_01540</i>
Qsbi_01540-R	CGTCCTCCAACGACCAATC	
Qsbi_01983-F	GACTACTGGTACCGCAATCTG	qRT-PCR analysis of <i>sbi_01983</i>
Qsbi_01983-R	CGAGCTGACCTCGATGAAC	
Qsbi_02068-F	TTCTTCTTCGACGACCACTTC	qRT-PCR analysis of <i>sbi_02068</i>
Qsbi_02068-R	GGGAGCACCGGCATAAA	

Name	Sequence (5'-3')	Usage
Qsbi_02232-F	CCGCTTCGAGCGCTATC	qRT-PCR analysis of <i>sbi_02232</i>
Qsbi_02232-R	GTGAGCACCTGTACCTGAC	
Qsbi_02590-F	CATGGCGATCTACGGCTAC	qRT-PCR analysis of <i>sbi_02590</i>
Qsbi_02590-R	AGCATCATGAGCGGATCG	
Qsbi_02764-F	TGGAGCAGGACTACGTCAA	qRT-PCR analysis of <i>sbi_02764</i>
Qsbi_02764-R	GAAGGCGTGGTGCAGATAC	
Qsbi_02988-F	CATGAGCTGCTGGCATATCT	qRT-PCR analysis of <i>sbi_02988</i>
Qsbi_02988-R	ATGTACAGCGGCAGTGATG	
Qsbi_03179-F	CCATTCGTCCTTCTGGGATATT	qRT-PCR analysis of <i>sbi_03179</i>
Qsbi_03179-R	GGACTTGGTGTCCGTCTC	
Qsbi_06052-F	TCCGGCGTATCTCATCTACA	qRT-PCR analysis of <i>sbi_06052</i>
Qsbi_06052-R	GTGACATCAGGAGGAAGGTC	
Qsbi_06449-F	GTCCTTTCGGGCGGATTT	qRT-PCR analysis of <i>sbi_06449</i>
Qsbi_06449-R	GCTGAAGCGTCAGAGAGTAAG	
Qsbi_06799-F	TCCTGGTCTTCGAGAACTATCC	qRT-PCR analysis of <i>sbi_06799</i>
Qsbi_06799-R	TAGTGGGTGGCGTCCTT	
Qsbi_06873-F	CTGGACCGGCAGGATCT	qRT-PCR analysis of <i>sbi_06873</i>
Qsbi_06873-R	CACAGGTCGGGCTCGTA	
Qsbi_08414-F	ACGCCTCCTTCTTCAACATC	qRT-PCR analysis of <i>sbi_08414</i>
Qsbi_08414-R	GGTGAGGGTGCCTTTGAG	

Name	Sequence (5'-3')	Usage
Qsbi_08957-F	CTGTACTCGTACACCAAGGAAC	qRT-PCR analysis of <i>sbi_08957</i>
Qsbi_08957-R	TCGACCGCCTTCAGGTA	
Qsbi_09057-F	GCGTACTGAACACCGAGAAG	qRT-PCR analysis of <i>sbi_09057</i>
Qsbi_09057-R	GATCTGAGTCTCCAGTTTGAGC	
Qsbi_09195-F	GCCTGTACGAGAGTGGTTATC	qRT-PCR analysis of <i>sbi_09195</i>
Qsbi_09195-R	CGCAGATCCAGTGTGTAGG	
Qsbi_09249-F	CACCTTCTACCACCGGAAAG	qRT-PCR analysis of <i>sbi_09249</i>
Qsbi_09249-R	AAAGTCGGAGAACGGTGTG	
Qsbi_09652-F	CGTCATGAACAACGACTACGA	qRT-PCR analysis of <i>sbi_09652</i>
Qsbi_09652-R	AATCGAGCACATACGACACC	
Qsbi_10015-F	GACTATGCCTTCCACAGCAA	qRT-PCR analysis of <i>sbi_10015</i>
Qsbi_10015-R	CCTGTGACGGTGGAGTAGA	

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