

Supplementary Material

Development of a Redox-Polymer-Based Electrochemical Glucose Biosensor Suitable for Integration in Microfluidic 3D Cell Culture Systems

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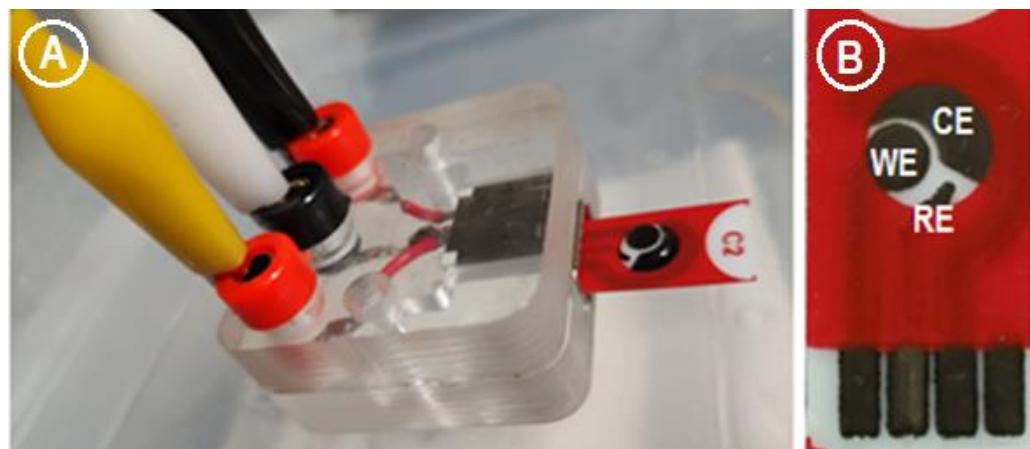


Figure S1. (A) Electrochemical cell setup with a carbon screen printed electrode (SPE.) (B) Carbon SPE with a carbon working (WE) and counter (CE) electrodes, and a C/Ag|AgCl mixture as reference electrode (RE).

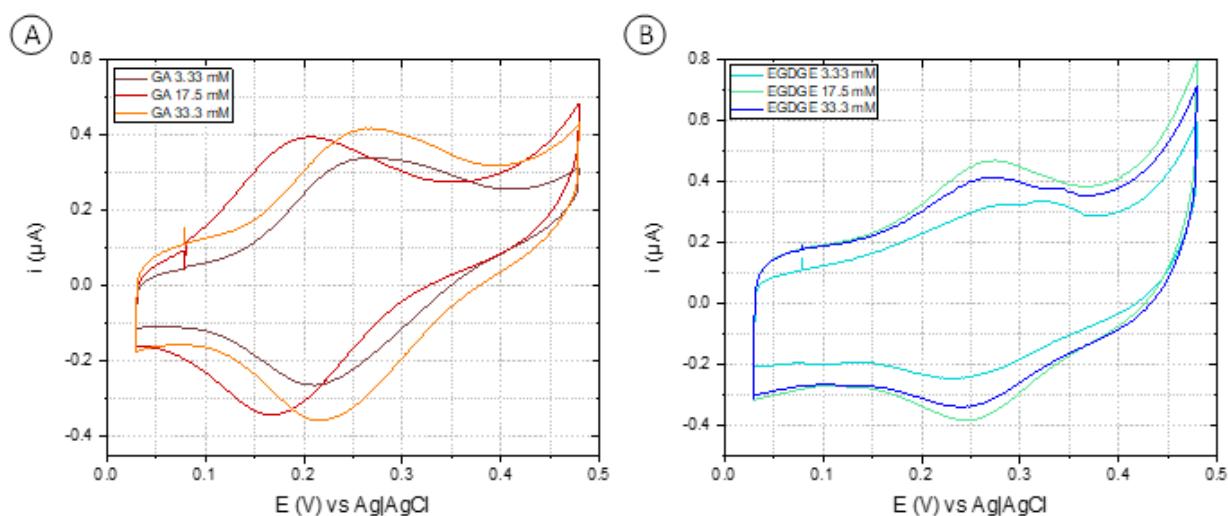


Figure S2. Cyclic voltammograms of hydrogels containing glucose oxidase (GOx) cross-linked to branched polyethylenimine modified with $\text{Os}(\text{bpy})_2\text{Cl}(\text{pyCOH})$ (OsBPEI) using either glutaraldehyde (GA) or ethylene glycol diglycidyl ether (EGDGE). (A) OsBPEI/GOx/GA and (B) OsBPEI/GOx/EGDGE hydrogels deposited on carbon SPEs in 0.1 M pH 7.4 phosphate buffer (PB) in the absence of glucose. Scan rate: 2 mV/s.

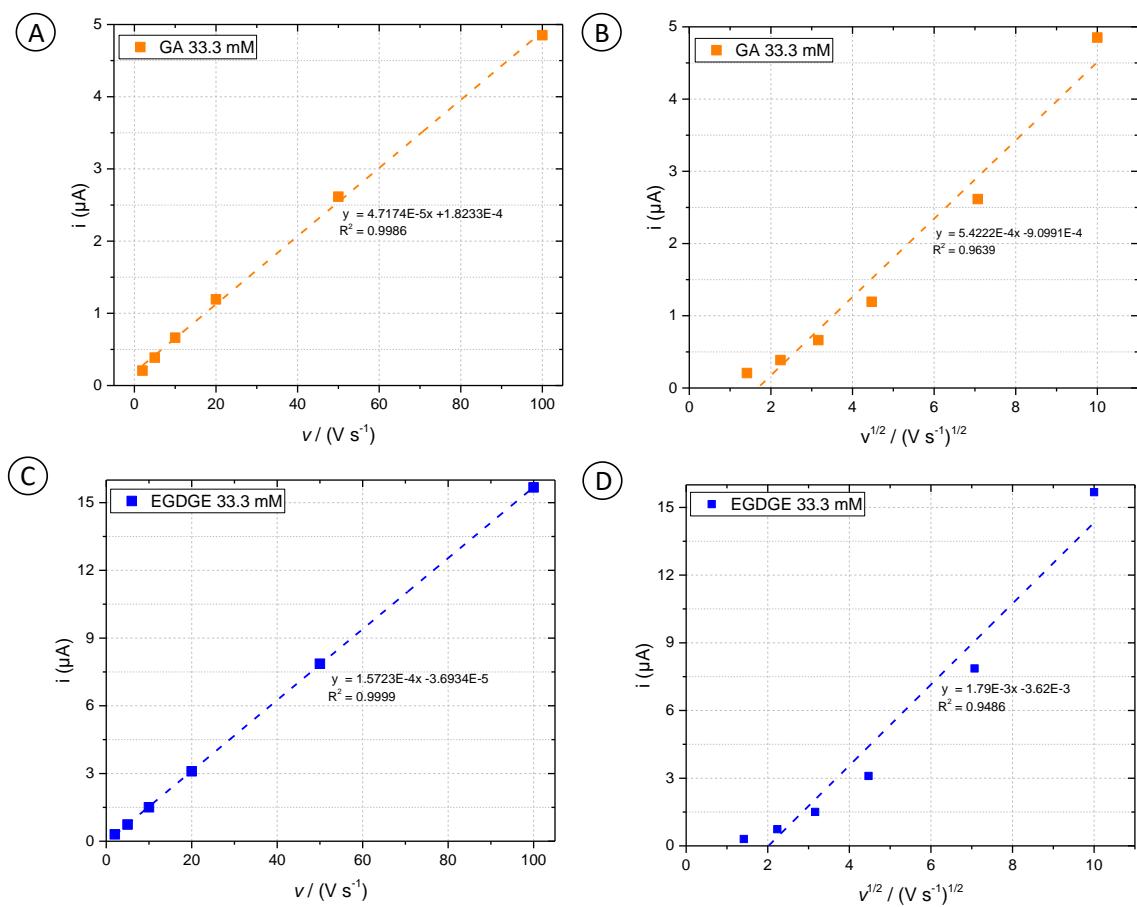


Figure S3. Dependence of the current of SPEs modified with OsBPEI/GOx/GA 33.3 mM (A,B) or OsBPEI/GOx/EGDGE composition (33.3 mM) (C,D) with respect to the potential scan rate (A,C) or square root of the scan rate (B,D) in 0.1 M pH 7.4 PB in the absence of glucose.

Table S1. Comparison of apparent Michaelis–Menten constant (K_m^{app}) and maximum current density (j_{max}) of different hydrogels compositions. Data from three independent electrodes are presented, as well as their mean value.

| | K_m^{app} (mM) | j_{max} ($\mu\text{A}/\text{cm}^2$) | R^2 |
|-----------------------------|---------------------|--|---------------------|--|---------------------|--|---------------------|--|-------|
| Mean | | | | | | | | | |
| SPEs in 0.1 M pH 7.4 PB | | | | | | | | | |
| OsBPEI/GOx/GA 3.33 mM | 19.9 | 110 | 43.1 | 298 | 30.1 | 230 | 31.8 ± 5.32 | 208 ± 13.8 | 0.987 |
| OsBPEI/GOx/GA 17.5 mM | 6.16 | 58.4 | 10.6 | 45.1 | 14.4 | 89.0 | 10.1 ± 0.75 | 63.5 ± 1.27 | 0.995 |
| OsBPEI/GOx/GA 33.3 mM | 4.68 | 39.4 | 11.4 | 75.3 | 10.0 | 69.5 | 8.76 ± 0.55 | 60.9 ± 1.04 | 0.996 |
| OsBPEI/GOx/EGDGE 3.33 mM | 14.0 | 60.9 | 13.1 | 55.5 | 14.8 | 60.6 | 14.0 ± 1.20 | 58.9 ± 1.50 | 0.994 |
| OsBPEI/GOx/EGDGE 17.5 mM | 5.53 | 17.2 | 5.53 | 16.2 | 7.20 | 35.2 | 6.31 ± 0.63 | 22.7 ± 0.58 | 0.990 |
| OsBPEI/GOx/EGDGE 33.3 mM | 14.0 | 41.6 | 8.26 | 38.7 | 7.22 | 32.4 | 10.9 ± 0.94 | 25.4 ± 0.69 | 0.993 |
| BPEI/GOx/GA 33.3 mM | 5.61 | 37.5 | 6.30 | 80.3 | 5.55 | 63.5 | 5.89 ± 0.75 | 60.4 ± 1.85 | 0.982 |
| BPEI/GOx/EGDGE | 32.9 | 190 | 22.3 | 143.9 | 22.2 | 140 | 28.4 ± 7.49 | 158 ± 15.0 | 0.968 |

| 33.3 mM | | | | | | | | | |
|---|------|------|------|------|------|------|-----------|-----------|-------|
| SPEs in Roswell Park Memorial Institute (RPMI-1640) | | | | | | | | | |
| OsBPEI/GOx/GA 33.3 mM | 4.00 | 31.0 | 4.49 | 54.0 | 5.51 | 43.9 | 4.68±0.73 | 43.0±1.62 | 0.976 |
| OsBPEI/GOx/EGDGE 33.3 mM | 3.59 | 44.3 | 13.5 | 161 | 12.7 | 151 | 10.6±0.99 | 113±3.01 | 0.992 |
| Carbon pencil leads in RPMI-1640 on-chip | | | | | | | | | |
| OsBPEI/GOx/GA 33.3 mM | 15.2 | 86.2 | 23.1 | 58.2 | 18.5 | 74.8 | 18.0±1.13 | 72.6±14.3 | 0.997 |
| OsBPEI/GOx/EGDGE 33.3 mM | 20.4 | 147 | 22.8 | 141 | 26.3 | 121 | 22.8±1.87 | 136±39.3 | 0.996 |

Table S2. Comparison of K_m^{app} and j_{max} of different hydrogels compositions. Data from three sequential calibration curves on a single electrode are presented.

| | K_m^{app} (mM) | j_{max} ($\mu\text{A}/\text{cm}^2$) | K_m^{app} (mM) | j_{max} ($\mu\text{A}/\text{cm}^2$) | K_m^{app} (mM) | j_{max} ($\mu\text{A}/\text{cm}^2$) |
|--|---------------------|--|---------------------|--|---------------------|--|
| SPEs in 0.1 M pH 7.4 PB | | | | | | |
| OsBPEI/GOx/GA 3.33 mM | 19.9 | 110 | 12.0 | 24.5 | 7.14 | 8.35 |
| OsBPEI/GOx/GA 17.5 mM | 6.16 | 58.3 | 3.50 | 19.6 | 4.38 | 5.33 |
| OsBPEI/GOx/GA 33.3 mM | 2.34 | 22.0 | 1.99 | 10.5 | 2.23 | 4.29 |
| OsBPEI/GOx/EGDGE 3.33 mM | 14.0 | 60.9 | 5.01 | 0.81 | - | - |
| OsBPEI/GOx/EGDGE 17.5 mM | 5.53 | 17.1 | 3.30 | 0.23 | - | - |
| OsBPEI/GOx/EGDGE 33.3 mM | 14.0 | 41.6 | 20.8 | 3.01 | - | - |
| BPEI/GOx/GA 33.3 mM | 5.61 | 37.4 | 3.33 | 48.6 | 4.62 | 52.3 |
| BPEI/GOx/EGDGE 33.3 mM | 32.9 | 190 | 16.4 | 187 | 11.5 | 102 |
| SPEs in RPMI-1640 | | | | | | |
| OsBPEI/GOx/GA 33.3 mM | 4.49 | 54.0 | 1.89 | 9.74 | 1.31 | 6.61 |
| OsBPEI/GOx/EGDGE 33.3 mM | 3.59 | 44.3 | 9.48 | 26.2 | 10.4 | 22.5 |
| Carbon pencil leads in RPMI-1640 on-chip | | | | | | |
| OsBPEI/GOx/GA 33.3 mM | 17.3 | 74.1 | 18.5 | 41.6 | 11.9 | 39.3 |
| OsBPEI/GOx/EGDGE 33.3 mM | 25.4 | 121 | 30.8 | 66.5 | 30.9 | 60.5 |

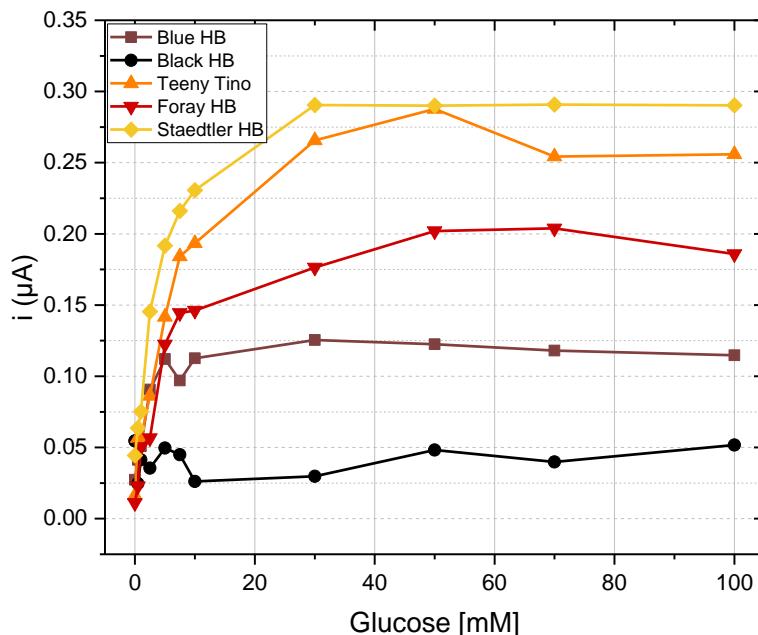


Figure S4. Electrochemical calibration curves performed with carbon pencil leads of different brands onto which hydrogels were deposited using GA as cross-linker. Chronoamperometric signals were measured in 0.1 M pH 7.4 PB and the current at $t = 2$ min taken as analytical signal.

Table S3. Comparison of analytical performance parameters of GOx based biosensors.

| Composition | Electrode | LOD (mM) | LOQ (mM) | Linear range (mM) | Sensitivity ($\mu\text{A mM}^{-1} \text{cm}^{-2}$) | Matrix | Ref. |
|---------------------------|-----------|----------|----------|-------------------|--|--------------|-----------|
| CB-PB-TEMPO-CNCs-GOx | SPE | 0.0040 | 0.015 | 0.1–2 | 5.70 | DMEM*/PB 1:4 | [1] |
| m-PPD-pHEMA-GOx | Pt | 0.0076 | - | - | - | MEBM* | [2] |
| CNT-GR-GOx | GCE | 0.0029 | 0.009 | 3–14 | 0.43 | DMEM | [3] |
| Graphene-MWCNT/GOx/nafion | PGE | 0.0149 | - | 0–39 | 35.2 | PB | [4] |
| GO-GOx | PGE | 0.0006 | - | 0.04–0.6 | 278.4 | PB | [5] |
| ZnS-CdS-Chitosan-GOx | PGE | 0.0030 | - | 0.01–1 | 11.5 | BRBS*/KCl | [6] |
| OsBPEI-GOx-GA | PGE | 1.4900 | 4.970 | 0–7.5 | 3.10 | RPMI | This work |
| OsBPEI-GOx-EGDGE | PGE | 0.5000 | 1.670 | 0–10 | 4.69 | | |

* Dulbecco's Modified Eagle Medium (DMEM)

*Mammary epithelial basal medium (MEBM)

*Britton-Robinson buffer solution (BRBS)

- Tang, Y.; Petropoulos, K.; Kurth, F.; Gao, H.; Migliorelli, D.; Guenat, O.; Generelli, S. Screen-Printed Glucose Sensors Modified with Cellulose Nanocrystals (CNCs) for Cell Culture Monitoring. *Biosensors* **2020**, *10*, 125. <https://doi.org/10.3390/bios10090125>.
- Dornhof, J.; Kieninger, J.; Muralidharan, H.; Maurer, J.; Urban, G.A.; Weltin, A. Microfluidic organ-on-chip system for multi-analyte monitoring of metabolites in 3D cell cultures. *Lab Chip* **2022**, *22*, 225–239. <https://doi.org/10.1039/d1lc00689d>.
- Madhurantakam, S.; Babu, J.K.; Balagru Rayappan, J.B.; Maheswari Krishnan, U. Fabrication of mediator-free hybrid nano-interface electrochemical biosensor for monitoring cancer cell proliferation. *Biosens. Bioelectron.* **2017**, *207*, 832–841. <https://doi.org/10.1016/j.bios.2016.09.039>.
- Torrinha, Á.; Montenegro, M.C.B.S.M.; Araújo, A.N. Microfluidic Platform with an Embedded Pencil Graphite Electrode Biosensor for the Detection of Glucose and Cadmium. *J. Electrochem. Soc.* **2019**, *166*, B155–B160. <https://doi.org/10.1149/2.1221902jes>.

5. Sehat, A.A.; Khodadadi, A.A.; Shemirani, F.; Mortazavi, Y. Fast immobilization of glucose oxidase on graphene oxide for highly sensitive glucose biosensor fabrication. *Int. J. Electrochem. Sci.* **2015**, *10*, 272–286.
6. Saglam, Ö.; Klzllkaya, B.; Uysal, H.; Dilgin, Y. Biosensing of glucose in flow injection analysis system based on glucose oxidase-quantum dot modified pencil graphite electrode. *Talanta* **2016**, *147*, 315–321. <https://doi.org/10.1016/j.talanta.2015.09.050>.