



# **Assessment of Nanoparticle-Mediated Tumor Oxygen Modulation by Photoacoustic Imaging**

Maharajan Sivasubramanian and Leu-Wei Lo\*

Department of Biomedical Engineering and Nanomedicine, National Health Research Institutes, Zhunan 350, Taiwan; 020609@nhri.edu.tw

\* Correspondence: lwlo@nhri.edu.tw

Abstract: Photoacoustic imaging (PAI) is an invaluable tool in biomedical imaging, as it provides anatomical and functional information in real time. Its ability to image at clinically relevant depths with high spatial resolution using endogenous tissues as contrast agents constitutes its major advantage. One of the most important applications of PAI is to quantify tissue oxygen saturation by measuring the differential absorption characteristics of oxy and deoxy Hb. Consequently, PAI can be utilized to monitor tumor-related hypoxia, which is a crucial factor in tumor microenvironments that has a strong influence on tumor invasiveness. Reactive oxygen species (ROS)-based therapies, such as photodynamic therapy, radiotherapy, and sonodynamic therapy, are oxygen-consuming, and tumor hypoxia is detrimental to their efficacy. Therefore, a persistent demand exists for agents that can supply oxygen to tumors for better ROS-based therapeutic outcomes. Among the various strategies, NP-mediated supplemental tumor oxygenation is especially encouraging due to its physio-chemical, tumor targeting, and theranostic properties. Here, we focus on NP-based tumor oxygenation, which includes NP as oxygen carriers and oxygen-generating strategies to alleviate hypoxia monitored by PAI. The information obtained from quantitative tumor oxygenation by PAI not only supports optimal therapeutic design but also serves as a highly effective tool to predict therapeutic outcomes.

check for **updates** 

Citation: Sivasubramanian, M.; Lo, L.-W. Assessment of Nanoparticle-Mediated Tumor Oxygen Modulation by Photoacoustic Imaging. *Biosensors* 2022, *12*, 336. https://doi.org/ 10.3390/bios12050336

Received: 14 April 2022 Accepted: 11 May 2022 Published: 13 May 2022

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). **Keywords:** nanoparticles; photoacoustic imaging; oxygen carriers; oxygen generators; tumor oxygen saturation

## 1. Introduction

Photoacoustic imaging (PAI) is an emerging biomedical imaging technique that involves optical excitation of intrinsic contrast agents by near-infrared (NIR) light pulsed lasers, causing thermoelastic effects of photoacoustic signals. The transient expansion and contraction resultant from the absorption of NIR laser energy induce the generation of pressure waves that can be detected, received by ultrasound (US) transducers, and constructed into images. The co-operative use of intrinsic optical contrast agents and US detection endows them with both high spatial resolution and deep tissue penetration [1–4]. The three main advantages of PAI over traditional imaging techniques are: (1) the use of intrinsic contrast agents (hemoglobin, melanin, or lipids); (2) high spatial resolution and imaging at clinically relevant depths; and (3) ability to gather anatomical and functional information in real time. Due to these features, it is possible to obtain biologically relevant information, such as angiogenesis, tumor hypoxia, changes in tumor oxygen saturation (sO<sub>2</sub>), and total Hb concentration [5–9]. Several exogenous contrast agents for PAI are synthesized and currently being used [10–15].

Hypoxia is a prominent feature of solid tumors caused by uncontrollable proliferation and defective vasculature. This leads to an insufficient blood supply, producing a reduction in oxygen tension in tumor tissues ( $pO_2 \leq 2.5 \text{ mmHg}$ ), then promoting the invasion and metastasis of tumor cells [16–18]. In addition, hypoxia also increases tumor resistance to reactive oxygen species (ROS)-based therapies, such as photodynamic therapy (PDT, combination of light and photosensitizer (PS) to generate ROS), radiotherapy (RT, combination of radiosensitzer and X-ray to generate ROS), and sonodynamic therapy (SDT, combination of ultrasound and sonosensitizer to generate ROS). The therapeutic efficacy of PDT depends highly on oxygen, and oxygen consumption further aggravates hypoxia [19–21].

As a consequence, the development of strategies that can alleviate hypoxia by supplying oxygen to tumors for better therapeutic outcomes is crucial. For example, hyperbaric oxygen therapy (HBOT) involves breathing pure oxygen in a pressurized chamber [22–24]. A study demonstrated that HBOT improved tissue angiogenesis and tumor hypoxia, and it increased apoptosis [25]. Although encouraging, its broad application is constrained by certain disadvantages, such as the overproduction of ROS in healthy tissues, which causes hyperoxic seizures and barotrauma [26]. As an alternative, uses of nanomaterials for oxygen-releasing or oxygen-generation strategies are gaining significance. Due to their physio-chemical properties, on the one hand, nanomaterials can be tailor-made to accommodate oxygen-carrying materials and assist them to reach the targeted site of action and deliver oxygen [27–30]. For instance, perfluorocarbons (PFCs), due to chemical and biological inertness, as well as their ability to dissolve a large amount of oxygen, have been widely employed as artificial blood substitutes [31–33]. In a study, hollow PEG-Bi<sub>2</sub>Se<sub>3</sub> NPs effectively accommodated oxygen-loaded PFC as an oxygen reservoir. When exposed to NIR light, a burst release of oxygen occurred and subsequently increased tumor oxygenation, which overcame hypoxia-associated radiotherapy resistance [34]. On the other hand, NPs could also increase oxygen saturation by the decomposition of endogenous hydrogen peroxide, which is overproduced by hypoxic tumors [35,36]. For example, albumin bound-MnO2 NPs increased tumor oxygenation by 45% by converting endogenous H2O2 in the tumor to oxygen. When combined with radiotherapy, it resulted in increased DNA double-strand breaks, which significantly inhibited tumor growth [37].

## 2. Biomedical Applications of PAI

PAI is a combination of optical and ultrasound imaging. When illuminated with a non-ionizing pulsed laser in the visible or near infrared wavelength, endogenous molecules absorb energy (Figure 1). Subsequently, the molecules in the ground state are elevated to an excited state, and they release energy in the form of light or heat when they relax back to the ground state. Through non-radiative relaxation, the absorbed photon is converted into heat. The induced heat mediates thermoelastic expansion, and thereby, pressure waves are generated. The pressure waves then propagate through tissues as ultrasound, which can be picked up and received by a US transducer to form an image [38,39]. Since sound waves are less scattered than photons, PAI can produce images with better optical contrast with high spatial resolution in deep tissues. In addition, since the image generated is based entirely on the optical absorption of endogenous molecules, PAI can deliver multi-contrast images of molecules based on their chemical composition. Owing to these features, PAI is successfully used to image endogenous molecules, such as oxy-Hb, deoxy-Hb, lipids, melanin, cytochrome, DNA/RNA, bilirubin, and water [40–44].

Breast cancer is the second-leading cause of cancer-related death in women, and the breast cancer mortality rate is continuing to increase [45,46]. Mammography is widely utilized as a main clinical diagnostic imaging technique for breast cancer [47–49]. However, high false-negative rates and decreased sensitivity with dense breast tissues remain challenging [50,51]. Breast tissues with high vascular density and increased oxygen saturation are potential markers. The vast potential of PAI, however, can easily overcome the limitations of mammography. An advanced SBH-PACT was developed that could image patients' breast in 15 s and accurately detect tumors based on angiographic anatomy. The results obtained were verified with US-guided biopsy. Without the use of an external contrast agent, this breast imaging technique could monitor patients' response to therapy [52].



Figure 1. Schematic illustration of PAI.

Toi et al., precisely characterized the breast tumor microenvironment by visualizing oxygen saturation status using a new PAI system with a hemispherical detector array, which is otherwise not visible on standard contrast-enhanced magnetic resonance imaging (MRI). Interestingly, this system was also able to monitor anti-cancer treatment-driven changes in tumor vasculature, such as improved intratumoral blood perfusion and functional changes in intravascular Hb saturation of oxygen [53]. Dogra et al., performed ex vivo PAI analysis using endogenous biomolecules (for example: oxy-Hb, deoxy-Hb, lipids, and water) as chromophores to distinguish prostrate tumors from benign prostatic hyperplasia and normal prostrate. The authors discussed that the presence of deoxy-Hb as a main constituent in hypoxic prostrate tumors enabled them to detect malignant prostrate tissue [54]. Wang et al., developed a dual US/PAI system and tested its ability to quantify tissue oxygenation in vivo. The results showed that the established PAI system could readily image the vasculature in rat skin and quantify tissue oxygenation in an in vivo hypoxia model. Finally, in an orthotiopic pancreatic tumor mice model, real-time hypoxia dynamics was successfully assessed by measuring oxy-Hb and deoxy-Hb [55].

The potential of PAI for label-free, non-invasive quantification of tumor oxygenation as a biomarker of radiation response was tested in human papilloma virus-positive (HPV+) and -negative (HPV-) patient-derived xenograft (PDX) models of squamous cell carcinoma head and neck cancer. For this purpose, assessment of tumor oxygen saturation and Hb concentration before, during, and after fractionated radiation therapy (fRT) was performed. The results showed that HPV+ and HPV- xenografts exhibited a differential response to fRT. Modulation in oxygen saturation was observed within days after the initiation of fRT prior to palpable change in tumor volume [56]. In a PDX model of head and neck cancer, Seshadri et al., monitored the  $sO_2$ % 24 h post RT and chemo-RT using combined PAI, MRI, and histopathology. An early increase in %sO<sub>2</sub> was associated with significant tumor growth inhibition and, 24 h after RT, radiation-induced vascular damage was detected by PAI due to the loss of hemodynamic response to gustatory stimulation in murine salivary gland. This study established the utility of PAI in assessing both tumor and normal tissue changes in oxygen saturation and Hb concentration to radiation in head and neck cancers [57].

#### 3. NPs as Oxygen Carriers or Generators

Hypoxia is a major factor in the tumor microenvironment, and it is characterized by depressed oxygen tension that encourages tumor cells to migrate, invade, and metastasize to distant organs. The partial oxygen pressure  $(pO_2)$  of tumor hypoxia can be as low

as 2.5 mm Hg compared to 40 mm Hg in normal cells, affecting cellular functions and causing improper functioning of organs. Hypoxia is either perfusion-related (temporary impairment in the delivery of blood) or diffusion-related (hampered movement of gas from blood capillaries to cancer cells) [58,59]. Furthermore, hypoxia severely diminishes the therapeutic performance of PDT, RT, and SDT, which strongly rely on oxygen to generate copious ROS to kill cancer cells [58]. The consumption of oxygen from tumors exacerbates hypoxia, raising the need for supplemental oxygen. In addition to several strategies, NP-mediated tumor reoxygenation is emerging and promising to alleviate hypoxia. This is because, by tailoring physiochemical properties, NPs can be made either to carry oxygen-philic materials or exhibit intrinsic enzyme-like properties to decompose endogenous hydrogen peroxides or self-decompose to generate oxygen in the tumor [30].

#### 3.1. NPs with PFCs as Oxygen Carriers

PFCs are hydrocarbons, in which hydrogen atoms are completely or mostly replaced with fluorine atoms. The highly hydrophobic property of PFCs contributes to their biological inertness, which has the ability to dissolve oxygen. The interaction between PFCs and oxygen is not chemical but rather loosely bound to PFC macromers through van der Waals interaction [60,61]. Due to this feature, PFCs were used in MR and US imaging as a diagnostic imaging agent [62]. The dissolution equilibrium of oxygen in PFCs directly depends on oxygen partial pressure, which governs their loading and the release of oxygen at biological environments. Moreover, the oxygen-loading capacity is not influenced by temperature, ionic strength, surfactant, storage, etc. [63,64]. Attributable to the immiscibility of PFCs in water, PFCs for oxygen-carrying applications are usually emulsified with suitable surfactants as NPs for intravenous (i.v.) administration. For instance, Fluosol-DA emulsion in albumin was the first PFC system to be approved by the FDA. Due to its shortcomings, however, such as low oxygen transport capacity, premature oxygen release, short shelf-life, etc., the product has been withdrawn [65]. To overcome these limitations, by combining three different PFCs along with egg-yolk phospholipids, Oxygent was developed. This developed oxygen carrier displayed a diameter of 160–180 nm in diameter with reduced macrophage activation, and an ability to circulate in micro-capillaries to deliver oxygen. However, advanced clinical trials of the product in patients were terminated because of an increased risk of stroke and other adverse events [66].

## 3.2. NPs as Carriers for Oxygenphilic Materials

In order to improve oxygen level and enhance PDT efficacy, Cheng et al., developed lipid NPs loaded with a photosensitizer and perfluorohexane [67]. After i.v. injection into tumor-bearing mice, the oxy-PDT selectively accumulated in the tumor and maintained a high level of oxygen. When illuminated with laser light, the oxy-PDT generated abundant ROS in oxygen-enriched tumors and exhibited superior PDT effects compared to conventional PDT. It was also found that the lifetime of  ${}^{1}O_{2}$  was prolonged from  $5 \times 10^{-6}$  s in water to  $5 \times 10^{-2}$  s in perfluorohexane (PFH). This work demonstrated the effect of oxygen-rich tumors on PDT effects. Jiang et al., designed a hierarchical nanodroplet system (Au + perfluorooctylbromide (PFOB) + O<sub>2</sub>) to amplify DNA damage and inhibit the DNA repair mechanism in RT. PFOB, as an oxygen source, alleviated hypoxia and favored RT by elevated ROS production. Ultrasmall Au NPs, as radiosensitizers, confine X-ray energy to induce DNA damage. In vivo, the nanodroplets demonstrated multimodal imaging capability, through which real-time image-guided precision RT was realized. This strategy was not only able to reoxygenate the tumor but also inhibited DNA repair [68].

Zhao et al., integrated PS, oxygen reservoir, and tumor-penetrating peptide in a single nanoplatform (CNPs/IP). The results showed that CRGDK peptide with tumor-penetration property guided the tumor-specific accumulation and penetration of PS and PFOB into both tumor periphery and hypoxic regions. PFOB, as a reservoir, released oxygen in the tumor to alleviate hypoxia to enable enhanced PDT. In an MDA-MB-231 tumor model, CNP/IP demonstrated elevated PDT effects, as evidenced by a reduction in hypoxic regions in tumor tissues. The combined effects of improved intratumoral distribution of PS and adequate oxygen supply significantly improved PDT efficacy [69]. Similarly, Song et al., presented PEG-PFC nanodroplets decorated with tantalum oxide (TaOx) NPs via a simple emulsion method TaOx@PFC-PEG@O<sub>2</sub>. The high oxygen affinity of PFC allowed TaOx@PFC-PEG to be loaded with oxygen. In addition, TaOx NPs, as a radiosensitizer, was able to absorb X-ray and amplify DNA damage. In vivo, TaOx@PFC-PEG@O<sub>2</sub> demonstrated the capability to increase oxygenation levels, which was followed by enhanced RT efficacy, by overcoming hypoxia-associated radio-resistance [70].

Zhou et al., developed a two-stage oxygen-delivery nanoplatform that consists of perfluorotributylamine (PFTA) as an oxygen reservoir loaded in albumin NPs. Among the various PFCs, the authors found that PFTA could increase RBCs infiltration and selected it as an oxygen carrier. In vivo, oxygen-saturated albumin NP selectively accumulated in tumor and increased oxygenation. Simultaneously, PFTA effectively increased RBCs infiltration and oxygen delivery by inhibiting platelet activation in tumor blood vessels. In vivo RT revealed a significant decrease in breast cancer tumor growth rate from 40% to 14% compared to control. In a colon cancer model, which is more hypoxic, the tumor growth rate decreased from 30% to 15% compared to control [71]. Zhang et al., constructed an "all-in-one theranostic amphiphile NPs featuring PEG-boron dipyrromethene amphiphile (PEG-F54-BODIPY)" to emulsify PFH into a theranostic nanoemulsion. The as-prepared nanoemulsion exhibited enhanced tumor accumulation, as evidenced by multimodal imaging ability and long tumor retention time. Due to these features, in a melanoma cancer xenograft model, the nanoemulsion demonstrated potential as an oxygen carrier and PS-quenching reliever to achieve highly efficient PDT [72].

To eliminate hypoxia-induced drug resistance, Li et al., developed an EGFR-targeted liposome for the co-delivery of PFOB as an oxygen generator and erlotinib against hypoxic lung cancer. The targeted liposome selectively accumulated in EGFR-overexpressing cells and co-delivered oxygen and erlotinib, which induced apoptosis and down-regulated the expression of EGFR, p-EGFR, and HIF-1 $\alpha$ . In lung tumor-bearing mice, the targeted liposome showed preferential accumulation in tumors, which was followed by the codelivery of oxygen and erlotinib. Tumors were relieved of hypoxia-induced drug resistance, and a strong antitumor effect was consequently observed [73]. Xing et al., designed a multifunctional nanoplatform that employed fluorinated polymer NPs encapsulating Ce6 and an indoleamine 2,3-dioxygenase (IDO) inhibitor (NLG919). The fluorinated polymer NPs loaded with oxygen in advance released oxygen in the tumor, thereby diminishing hypoxia levels. The incorporation of IDO inhibitor along with PS greatly improved PDT efficacy by inhibiting the growth of primary and abscopal tumors via enhanced T cell infiltration (Figure 2) [74]. Yu et al., presented nano red blood cell (nnRBC) by replacing heme with perfluorodecalin (FDC). The developed formulation overcomes the autoxidative cytotoxicity and renal toxicity of heme, which demonstrated long-circulation, low immunogenicity, relieved hypoxia, and enhanced RT efficacy [75]. Chen et al., successfully developed an oxygen-generating SDT nanoplatform that encompasses FC-functionalized hollow mesoporous organosilica NPs for improving SDT performance. The well-defined mesoporous structure allowed high loading of a sonosensitizer (IR780), and the FC chain delivered oxygen in the tumor to mitigate hypoxia. In the presence of US, in vivo results showed enhanced tumor accumulation of NPs in hypoxic tumor, followed by accelerated oxygen release. In hypoxic PANC-1 pancreatic cancer, elevated ROS production resulted in highly efficient SDT by overcoming hypoxia-induced resistance [76].



**Figure 2.** (a) The diagram of dosage regimen. (b) Inhibition of 4T1 tumor growth by PDT and IDO inhibitor. Tumor-bearing mice were intratumorally injected with oxygen-saturated PF-PEG@Ce6@NLG 919, and the tumor sites were illuminated (660 nm, 55 W/cm<sup>2</sup>). (c) Photograph of the tumors removed on day 14. (d) The weight of the tumor treated with different formulations on the last day. (e) TUNEL staining of tumor sections. The green light spots represent DNA damage tagged on FITC; the blue represents the nucleus labeled with DAPI. (f) The body weight of the mice. *p* values: \*\* *p* < 0.01, \*\*\* *p* < 0.001, one-way ANOVA, *n* = 5. (g) Immunofluorescence staining of tumor sections to detect whether the PF-PEG group could improve hypoxia in vivo. The nucleus and the anoxic region were stained with DAPI (blue) and SOSG (green), respectively. Reproduced with permission [74]. Copyright 2019, Elsevier B.V.

#### 3.3. NPs with Hb as Oxygen Carrier

The primary function of RBC is to transport oxygen from lungs to other tissues by binding to iron molecules containing the integral complex protein Hb. Hb is a globular protein that encompasses four polypeptide chains (two  $\alpha$  and two  $\beta$ ) folded onto itself. The three-dimensional folding pattern allows very efficient binding of a heme group. The tendency of oxygen to be either bound or released by Hb strongly depends on the partial pressure of oxygen [77–79]. Although the oxygen binding and releasing property of Hb is promising, free Hb possesses adverse side effects. For instance, after i.v. administration,

patients suffered renal toxicity and cardiovascular complications. Cell-free Hb was also determined to have a very short circulation residence time because, when cell-free, Hb is unstable and dissociates into dimeric and monomer forms, and it is engaged by hepatobiliary and renal mechanisms leading to Hb-based toxicities in these organs. Moreover, free Hb is known to sequester nitric oxide, thus causing vasoconstriction and cardiovascular complications [80]. As an alternative, cross-linked Hb products were introduced, such as HemAssist and Optro, but these were associated with an increase in mortality rates. Then, several polymerized Hb products (Hemopure, PolyHeme, and HemoLink) were introduced by precisely controlling the polymer molecular weight. These products, however, showed a high risk of several complications in clinical trials [81].

Wang et al., established Hb-linked conjugated polymer NPs that do not require an external light source and mitigate hypoxia for efficient PDT. Hb acts as the catalyst for the luminol–H<sub>2</sub>O<sub>2</sub> chemiluminescence system and supplies molecular oxygen. When luminol and  $H_2O_2$  were extraneously added, PDT was initiated. A series of chemical reactions occurred, in which luminol, in the presence of  $H_2O_2$  and Hb, radiated blue light while the polymeric NPs had an absorption in the range of 400–550 nm, which establishes a donor-acceptor for chemiluminescence energy transfer. As a consequence, Hb-bound NPs sensitize oxygen molecules by the absorption of luminol emission, producing ROS for enhanced PDT [82]. A multifunctional nanocomplex system was developed for oxygenrich two-photon PDT that comprises two-photon absorbing molecules, PS and Hb, as an oxygen donor. PDT was initiated by the indirect activation of PS by two-photon laser through intraparticle fluorescence energy transfer, while Hb increased oxygen saturation in the tumor for improved PDT effect. Indeed, the nanocomplex developed in this study achieved improved PDT depth and mitigated hypoxia. Limited drug accumulation and the hypoxic tumor environment contribute to chemoresistance and lead to poor efficacy [83]. You et al., presented multifunctional liposomes for the synchronous delivery of oxygen and chemotherapeutic drug doxorubicin (DOX). Hb was embedded on the surface of the liposomes, while DOX was loaded inside of the liposomes. When i.v. administered, Hb enabled tumor-specific accumulation, which was followed by oxygen and DOX release. The oxygen released by liposomes alleviates hypoxia-driven chemoresistance that enabled DOX uptake in cancer cells. Due to the cooperative effects of tumor oxygenation and DOX release, liposomes exhibited stronger antitumor effects compared to controls [84]. Similarly, You et al., prepared liposomes for PDT against hypoxic tumors by encapsulating indocyanine green as PS and Hb as an oxygen donor. When i.v. administered, oxygencarrying liposomes with PS demonstrated preferential accumulation in tumors. This led to an increase in tumor oxygenation levels, as evidenced by  $T_2$ -weighted magnetic resonance imaging and immunostaining, and the level of hypoxia inducible factor- $1\alpha$  (HIF- $1\alpha$ ) and vascular endothelial growth factor (VEGF) in the tumor was down-regulated. As a result, enhanced PDT efficacy against hypoxic tumor was observed compared to control groups after laser irradiation [85].

In order to maximize ROS generation efficacy by SDT, Hb was embedded in a zeolitic imidazolate framework (ZIF-8). In the as-prepared nanoplatform, Hb not only serves as an oxygen carrier but also exhibits the function of a sonosensitizer. In addition, the ZIF-8 shell in the NPs can be decomposed in the presence of low pH in tumor microenvironment to release oxygen, thereby alleviating hypoxia for enhanced SDT (Figure 3). In both subcutaneous and deep-seated tumors, NPs demonstrated improved SDT effects due to the cooperative effects of oxygen and SDT. The authors also found that this effective tumor inhibition was ascribed to activation of the mitochondrial apoptosis pathway [86]. As a strategy to diminish hypoxia and elevate the therapeutic effects of RT, Hu et al., presented Au-Hb NPs embedded with platelets. Here, platelets function as a tumor-targeting agent that could deliver radio-sensitizing Au NPs and oxygen-carrying Hb. In the presence of low-dose X-ray irradiation, Au NPs sensitized tumor cells to induce apoptosis through the formation of low-energy photons and secondary-charged particles with adequate O<sub>2</sub> delivered by Hb. Although this nanoplatform achieved an antitumor effect with minimal



side effects, clinical translation is not possible due to a high demand for platelets and translation-related challenges [87].

**Figure 3.** (a) Schematic illustration of the synthesis procedures and antitumor mechanism of OHZ NP. (b) Schematic illustration of OHZ NP for tumor treatment. (c) Relative tumor volume curves of mice treated with various treatments, (d) tumor weights of mice at the 14th day after the treatments, and (e) the photographs of mice and tumors in different groups at the end of treatments of the subcutaneous tumor treatment group. (f) Relative tumor volume curves of mice treated with various treatments, (g) tumor weights of mice at the 14th day after the treatments, and (h) the photographs of mice and tumors in different groups at the end of treatments, and (h) the photographs of mice and tumors in different groups at the end of treatments, and (h) the photographs of mice and tumors in different groups at the end of treatments of the deep-seated tumor treatment group mimicked by 2 cm chicken slice blocking. *p* values were calculated via ANOVA (\*\*\* *p* < 0.001). Fluorescence images of (i) subcutaneous tumor slices and (j) deep-seated tumor slices after being stained by H&E and TUNEL. Reproduced with permission [86]. Copyright 2021, American Chemical Society.

## 3.4. Oxygen Generation by Catalase (CAT) or CAT Mimicking NPs

CAT, an extraneous heme-containing enzyme that can rapidly decompose  $H_2O_2$ into H<sub>2</sub>O and O<sub>2</sub>, is a highly suitable candidate for increasing tumor oxygenation. The expression levels of this enzyme in tumor, however, are lower than in normal tissues. Consequently, tumor-targeted delivery of CAT is essential to lower hypoxia levels, and the proteolytic degradation of CAT presents another challenge that needs to be addressed [88]. Several designed nanomaterials that can encapsulate or CAT have been reported [89]. The purpose of nanomaterials design is to preserve the enzyme activity of CAT during in vivo circulation and deliver them in tumors to increase oxygenation levels. A multifunctional CAT-loaded liposome with cisplatin (IV)-prodrug-conjugated phospholipid was developed by Liu et al., for chemo-radiotherapy [90]. The enzyme activity of CAT loaded inside of the liposome was preserved, and it decomposed endogenous  $H_2O_2$  in tumor to mitigate hypoxia. When injected i.v. in a tumor mouse model, multifunctional liposome accumulated in the tumor and contributed to the highest level of DNA damage in cancer cells after X-ray radiation, and it further demonstrated an elevated therapeutic outcome in chemoradiotherapy synergistically. Liu et al., presented an in situ free radical polymerization to modify CAT using meso-tetra(p-hydroxyphenyl) porphine (THPP), a photosensitizer, as the crosslinker and short-chained PEG as the drafting moiety. The obtained nanocapsules

preserved the enzyme activity of CAT, as an oxygen generator can decompose  $H_2O_2$  in tumor to relieve hypoxia for enhanced PDT. THHP not only functions as a PS but also acts as a chelating agent to label <sup>99m</sup>Tc4+ for in vivo single-photon emission computed tomography (SPECT) imaging. When i.v. injected, nanocapsules passively accumulated in the tumor, as confirmed by SPECT imaging, and they generated oxygen by decomposing tumor-laden  $H_2O_2$ , which enabled remarkable PDT by destroying tumors [88].

Zhao et al., developed self-assembled NP for oxygen-boosted PDT. The nanosystem contains ß cyclodextrin and CAT-conjugated hyaluronic acid loaded with adamantanemodified Ce6. The obtained nanosystem (HA-CAT@aCe6) maintained the enzyme activity and could accumulate in CD44 receptor-overexpressing cancer cells, thus generating adequate oxygen to abrogate hypoxia to elevate PDT. In vivo, HA-CAT@Ce6 actively accumulated in MDA-MB-231 tumor-bearing mice by the cooperative effects of oxygen and PDT, and significant inhibition in tumor volume was observed [91]. Zhang et al., constructed a biomimetic core-shell nanoplatform that consists of a pH-sensitive zeolitic imidazolate framework embedded with CAT and doxorubicin as the core and murine melanoma cell membrane coating as the shell. The core acts as a reservoir for drugs and an oxygen generator, whereas the shell provides tumor-targeting ability and elicits an immune response due to an abundance of antigens. In an in vivo tumor-bearing mouse model, core-shell NP reduced hypoxia levels, enhanced chemotherapeutic effects, and simultaneously down-regulated the expression of programmed death ligand 1 (PD-L1). When combined with immune checkpoints blockade therapy, the dual inhibition of the PD-1/PD-L1 axis elicited a strong immune response, prolonged tumor recurrence, and inhibited tumor metastasis [92].

Peng et al., developed a multifunctional CAT, i.e., DOX- and lysosome-targeted NIR PS MBDP-loaded liposome (FA-L@MD@CAT). It was demonstrated that FA-L@MD@CAT accumulated in tumor by both active and passive targeting mechanisms to increase tumor oxygenation for a remarkable chemo-combined PDT. In a tumor-bearing mouse model, FA-L@MD@CAT provided adequate oxygen for PDT and reversed immunosuppressive TME by modulating immune cytokines to elicit antitumor immunities, thus enhancing tumor inhibition in vivo [93]. Van Hest et al., designed a synergistic nanoplatform for MRI-guided tumor growth inhibition by boosting PDT efficacy. The nanoplatform consists of Ce6 conjugated glycol chitosan micelles loaded with CAT-stabilized MnO<sub>2</sub> NP (CMGCC). GC provides a long in vivo half-life with a pH-stimulated charge switch for tumor accumulation, and MnO<sub>2</sub> acts as an intracellular GSH scavenger to amplify ROS levels and also serves as a T<sub>1</sub> contrast agent for MRI by releasing Mn<sup>2+</sup>. The systemic administration of CMGCC in a HeLa tumor demonstrated that it is a promising theranostic agent for PDT [94].

Qi et al., combined multimodal imaging for diagnosis, guided surgery, and effective therapy to treat glioma (Figure 4). The developed CAT-integrated albumin theranostic nanoprobe (ICG/AuNR@BCNP) penetrated the blood–brain barrier and accumulated into glioma via albumin-binding protein-mediated transportation to perform theranostic functions. The combined fluorescence, PAI, and infrared thermal imaging were able to clearly differentiate brain tumors from surrounding tissues. The nanoprobe reduced hypoxia levels in glioma by generating adequate oxygen and induced elevated local hyperthermia. Through i.v. or i.t. in several animal models, guided by external multimodal imaging, nanoprobe inhibited glioma with improved apoptosis and antiangiogenic effects by remarkable phototherapy [95].



**Figure 4.** (a) Relative tumor growth curves recorded during treatment. The dosage of ICG/AuNR@BCNP in mice was equivalent to ICG 1.0 mg/kg and Au 0.76 mg/kg. (b) Weight and photograph of isolated tumor at the end of treatment. (c) Immunofluorescence staining of tumor sections with TUNEL, anti-HIF-1 $\alpha$ , and anti-CD31 antibody. Scale bar: 50 or 100 µm. Corresponding semiquantitative analysis of (d) the apoptotic index and (e) HIF-1 $\alpha$  and (f) CD31 positive areas (mean  $\pm$  SD, n = 3-5, \* p < 0.05, \*\* p < 0.01, \*\*\* p < 0.001). (g) H&E and ki67 staining of tumor sections. Scale bars: 1 mm for low magnification, 100 µm for high magnification. Reproduced with permission [95]. Copyright 2020, American Chemical Society.

Manganese dioxide (MnO<sub>2</sub>) has a strong capacity to undergo CAT-like activity to decompose  $H_2O_2$  to generate oxygen. In the presence of a mild acidic environment, MnO<sub>2</sub> NP releases Mn<sup>2+</sup> ions to promote the decomposition of  $H_2O_2$  for oxygen generation and

anticancer therapy. In addition to oxygen generation, Mn<sup>2+</sup> ions could also be utilized as a T1 contrast agent for MRI [96]. Zhang et al., developed HA-MnO<sub>2</sub> NP as an oxygenmodulating targeted MR imaging agent for glioma. NP was synthesized by toxicity-free simple mixing of sodium permanganate and HA solution, where HA serves as a reducing agent, a dispersing agent, and a CD44-targeting agent. After i.v. in a rat intracranial glioma model, HA-MnO<sub>2</sub> NP exhibited sustained attenuation of tumor hypoxia by downregulation of VEGF and HIF-1 $\alpha$  expression. In the acidic tumor microenvironment, Mn<sup>2+</sup> was released and demonstrated imaging sensitivity for detection with MRI for a prolonged period of up to 3 days. From these results, HA-MnO<sub>2</sub> NPs has the capability for simultaneous targeted imaging, real-time monitoring, and tumor microenvironment modulation [97]. Guo et al., developed a self-assembled NP by mixing KMnO<sub>4</sub> with HA solution followed by Ce6 loading for oxygen-assisted PDT to treat bladder cancer. The prepared HSA-MnO<sub>2</sub>-Ce6 NPs demonstrated oxygen generation with  $H_2O_2$  that resulted in two-fold higher ROS production. In vivo, HSA-MnO<sub>2</sub>-Ce6 showed preferential accumulation in tumor, as confirmed by NIR and MR imaging with a  $\approx$ 3.5-fold increase in oxygen levels. In an orthotopic bladder cancer mouse model, when combined with laser irradiation, it demonstrated remarkably improved therapeutic efficacy and significantly prolonged lifetime [98].

A composite core-shell NP was presented by Gang et al., for combined ROS-mediated PDT/CDT to treat breast cancer. The core-shell NP consists of indocyanine green loaded mesoporous silica as a core and MnO<sub>2</sub> as a shell. MnO<sub>2</sub> could not only generate oxygen in the presence of endogenous  $H_2O_2$  but could also release  $Mn^{2+}$  ions to scavenge intracellular glutathione and generate ROS by Fenton-type reactions. In a breast cancer model in vivo, ICG-loaded nanozymes selectively accumulated in the tumor, and they inhibited tumor growth and metastasis by the cooperative effects of oxygen-boosted combined PDT/CDT [99]. A tumor microenvironment-responsive theranostic nanoplatform was developed by Liu et al., for combined PDT and chemotherapy favoring antitumor immunities. The nanoplatform is Ce6, DOX dual drug-loaded hollow mesoporous MnO<sub>2</sub> shells (H- $MnO_2$ -PEG/C&D). The multifunctions of hollow mesoporous  $MnO_2$  shells are their ability to load drugs, release Mn<sup>2+</sup> ions in acidic tumor microenvironment for tumor-specific MRI, and generate oxygen by decomposing endogenous H2O2 (Figure 5). Moreover, tumor oxygenation reversed the immune-suppressing mechanism by polarizing macrophages from M2 to M1 transition. Chemo/PDT combined with PD-L1 checkpoint blockade induced an abscopal effect, which not only inhibited primary tumors but also distant tumors without light exposure, likely through CTL migration as confirmed by subsequent T-cell depletion experiments [100].

Prussian blue (PB) NP has also been shown to perform CAT-like functions to decompose  $H_2O_2$  to  $O_2$ . Zhang et al., designed a mutt homologue 1 inhibitor and PS-loaded mesoporous silica-coated PB nanoplatform for oxygen-enhanced PDT. PB NP demonstrated an increase in tumor oxygenation, which could elevate ROS production to aggravate oxidative damage for cancer therapy by inhibiting the MTH1-mediated damage-repairing process [101]. In another study by Cai et al., porous hollow PB NP were embedded with glucose oxidase (GOx) and then modified with HA for active targeting to the tumor. PB NP of GOx actively accumulated in the tumor and decomposed  $H_2O_2$  to  $O_2$  to reduce hypoxia levels and elevate glucose depletion for tumor starvation therapy [102].

12 of 20



**Figure 5.** In vivo combined chemo-PDT treatment with H-MnO<sub>2</sub>-PEG/C&D. (**a**) Representative immunofluorescence images of 4T1 tumor slices collected from untreated control mice and mice 6 h and 12 h post i.v. injection with H-MnO<sub>2</sub>-PEG/C&D. The nuclei, blood vessels, and hypoxic areas were stained with DAPI (blue), anti-CD31 antibody (red), and anti-pimonidazole antibody (green), respectively (three mice per group). (**b**) Quantification of hypoxia areas in tumors at different time points post injection of our NP. (**c**) Tumor growth curves of different groups of mice after various treatments indicated. Error bars were based on standard errors of the mean (SEM) (six mice per group). (**d**) Average weight of tumors collected from mice at day 14 post initiation of various treatments. The predicted addictive effect was calculated by multiplying the tumor growth inhibition ratios of group 4 (PDT alone) and group 5 (chemotherapy alone). (**e**) H&E-stained tumor slices collected from mice post various treatments indicated. *p* values in (**c**,**d**) were calculated by Tukey's post-test (\*\*\* *p* < 0.001, \*\* *p* < 0.01, or \* *p* < 0.05). Reproduced with permission [100]. Copyright 2017, Nature publishing group.

## 4. PAI for the Assessment of NP-Mediated Tumor Oxygen Saturation

Tumor hypoxia is a detrimental factor affecting the therapeutic outcomes of PDT, SDT and RT, because these therapeutic modalities strongly depend on oxygen tensions in tumor to induce ROS-mediated cancer cell death [58]. It is critical to identify the hypoxia status by quantifying tumor oxygenation. Specifically, it will not only assist clinicians to identify moderate-to-severely hypoxic tumors to plan treatment strategies, management and patient classification but also predict treatment efficacy in a very early stage, which will vastly improve patients' quality of life. Invasive polarographic electrodes are the gold standard for the detection and characterization of tumors, but several factors have constrained their clinical application [103,104]. PAI offers non-invasive real-time monitoring and quantification of tumor oxygen saturation without the aid of exogenous contrast agents (Table 1) [8].

Design	Treatment Modality	Tumor sO <sub>2</sub> Quantification by PAI	Ref
PFC-decorated tantalum oxide NP	RT	$\approx$ 37% increase in tumor sO <sub>2</sub> post i.t. injection	[70]
PFH-incorporated theranostic nanoemulsion	PDT	${\approx}25\%$ increase in tumor $sO_2$ post i.v. injection	[72]
Hb-incorporated multifunctional nanocomplex	2 photon PDT	Strong PA signal from oxy-Hb were observed 6 h post-i.v. injection and continued to increase with time	[83]
HA-porous hollow Prussian blue NP	Tumor starvation therapy	$\approx 35\%$ increase in tumor sO <sub>2</sub> 2 h post i.t. injection	[102]
Benzoporphyrin derivative as PS	PDT	Treatment responders exhibited $\approx$ 95% and $\approx$ 85% decrease in sO <sub>2</sub> at 6- and 24-h post-PDT	[105]
Liposome-loaded DOX (HaT-DOX)	Chemotherapy	Treatment responders exhibited on average a 22% drop in sO <sub>2</sub> 2 h post-chemotherapy.	[106]
Dendritic mesoporous organosilica NP-encapsulated ICG and CAT (ICG-CAT@MONs)	PAI/US guided PDT	${\approx}27\%$ increase in tumor sO_2 6 h post injection	[107]
Manganese ferrite NP embedded in hypoxia-responsive amphiphilic polymer membranes loaded with δ-aminolevulinic acid (ALA-hMVs)	SDT	The tumor vascular sO_2 increased from 1.6 $\pm$ 0.3% (pre-injection) to 13.6 $\pm$ 0.8% at 24 h post-injection	[108]
Biosynthetic functional vesicles (BFVs) presenting PD1 and TRAIL on the surface, loading CAT in their inner core	Immunotherapy	Tumor sO <sub>2</sub> levels in the BFVs/PD1-TRAIL-CAT or free CAT-treated tumors were significantly higher than PBS group	[109]
CAT@liposome	Radio combined Immunotherapy	Tumor sO <sub>2</sub> levels increased to $\approx$ 32% at 24 h post injection of CAT@liposome combined H <sub>2</sub> O <sub>2</sub> @liposome	[110]
In situ gelation system containing PS-modified CAT together with PEG-double acrylate (PEGDA) as the polymeric matrix loading immune adiuvant NP	PDT combined immunotherapy	Tumor sO <sub>2</sub> increased to $\approx$ 30% at 48 h post local injection	[111]
Self-delivery nanomedicine	PDT	Tumor sO <sub>2</sub> increased to $\approx 45\%$ at 6 h post i.v. injection	[112]
Tirapazamine-loaded metal–organic framework	Hypoxia activated therapy	Tumor sO <sub>2</sub> decreased from $\approx$ 75% to $\approx$ 25% at 2 h post i.t. injection	[113]
Photoacoustic nanodroplets	PDT	Tumor sO <sub>2</sub> increased to $\approx$ 9% post i.v. injection	[114]
Multifunctional theranostic NP Biodegradable catalytic NP	SDT and starvation therapy Tumor catalytic therapy	Tumor sO <sub>2</sub> increased to $\approx 18\%$ at 24 h post i.v. injection Tumor sO <sub>2</sub> increased to $\approx 40\%$ post i.v. injection	[115] [116]

**Table 1.** Summary of NP-mediated tumor sO<sub>2</sub> increase quantified by PAI.

Hasan et al., used PAI and mapped changes in glioma tumor  $sO_2$  as a surrogate marker for predicting the probability of PDT success. PDT consumes oxygen in the tumor, which will cause a change in the tumor  $sO_2$ . By measuring oxy-Hb and deoxy-Hb by PAI, a 3D atlas of tumor  $sO_2$  before, during, and after the PDT can be obtained. The authors found a  $\approx$ 95% and  $\approx$ 85% decrease in  $sO_2$  at 6 and 24 h post-PDT, and they predicted that tumors were responding to the treatment, as evidenced by no tumor recurrence observed up to one-month post-PDT. In contrast, no significant changes in  $sO_2$  were observed in the non-responding tumors. Information about the possibility of tumor regrowth in 24 h is crucial and presents the possibility of an early intervention. The tumor prediction  $sO_2$ map was validated with caliper measurements and photographs of the recurred tumor [99]. Similarly, Kolios et al., monitored DOX-loaded liposome treatment-induced changes in tumor vasculature using PAI. They found that quantitative PAI analysis at 30 min posttreatment correlated with a decrease in  $sO_2$  (22%) due to changes in tumor vasculature [106].

Wu et al., investigated modulation in tumor oxygenation after i.v. administration of ICG, CAT loaded dendritic MSN. Changes in tumor sO<sub>2</sub> in the four T1 tumors were evaluated by a Vevo-LAZR PAI system by measuring oxy-Hb and deoxy-Hb before and after injection. The results showed that sO<sub>2</sub> signals in the PA image appeared at 2 h and became strong at 6 h, and signal intensity declined and became weak at 24 h. The quantitative analysis revealed an increase in average sO<sub>2</sub> total to a maximum of  $\approx$ 27% at 6 h from  $\approx$ 10% at 2 h, and it decreased to  $\approx$ 8% at 24 h. The CAT-loaded NP group increased the average sO<sub>2</sub> total by approximately 3.94 and 4.43 times compared to control groups. The data obtained from quantitative sO<sub>2</sub> analysis by PAI guided the authors to initiate PDT treatment at 6 h post-injection to increase the therapeutic performance of PDT [107]. Like PDT, SDT efficacy depends on oxygen tension in the tumor. Chen at al. developed

hypoxia-responsive nanovesicles (hMVs) to alleviate hypoxia for enhanced PDT (Figure 6). When administered i.v., hMVs accumulated in the tumor followed by disassembly to release manganese ferrite NP to catalyze  $H_2O_2$  to  $O_2$ . The tumor oxygen saturation changes were monitored by PAI. Tumor  $sO_2$  levels were evaluated using PAI by measuring oxy-Hb and deoxy-Hb. The tumor vascular  $sO_2$  increased from  $1.6 \pm 0.3\%$  (pre-injection) to  $13.6 \pm 0.8\%$  at 24 h post-injection of nanovesicles, which was several-fold higher than that of controls. This indicates efficient tumor oxygenation and hypoxia relief by the hMVs, which was also confirmed by immunofluorescence staining of HIF-1 $\alpha$  in tumor sections after 24 h post-injection. From the tumor  $sO_2$  values obtained from the quantitative PA analysis, SDT was applied at 24 h post-injection of hMVs, which resulted in significant inhibition in tumor growth [108].



**Figure 6.** (a) Representative PA images of B16 tumors on mice showing signals of oxygenated hemoglobin ( $\lambda = 850$  nm) before/after i.v. injection of various formulations. (b) The corresponding quantification of the tumor vascular saturated O<sub>2</sub> levels (sO<sub>2</sub>) calculated from (a). (c) Immunofluorescence staining of tumor sections showing the expression of HIF-1 $\alpha$  after i.v. injection of various formulations (d) Quantitative analysis of HIF-1 $\alpha$  positive areas for each group in (c) by using the ImageJ software (n = 5). (\*\*\* p < 0.001, \*\* p < 0.01). Reproduced with permission [108]. Copyright 2021, Elsevier B.V.

Liu et al., developed biosynthetic functional vesicles (BFV) covered with PD1 and TRAIL and loaded with CAT to boost systemic antitumor immunity. After local injection of BFV, PAI was used to determine tumor  $sO_2$ , and it was found that  $sO_2$  levels of BFV-treated tumor were comparable to CAT-only treated tumor. Furthermore, a reduction in hypoxia levels infiltrated cytotoxic T cells in the tumor. Overall, the immuno-modulating ability and robust antitumor immunity of BFVs facilitated a significant regression of tumor growth, prevention of abscopal tumors, and excellent inhibition of lung metastasis [109]. Liu et al., developed CAT@liposomes for oxygen-enriched radioimmunotherapy with CTLA4 blockade. To realize this, liposomes were individually encapsulated with CAT and  $H_2O_2$ , respectively. In vivo, CAT liposomes were i.v. administered first to consume  $H_2O_2$  in TME to generate oxygen. After 4 h,  $H_2O_2$  liposomes were injected, which amplified the tumor oxygenation confirmed by PAI analysis. The well-oxygenated tumors not only improved RT efficacy but also transformed cold immune suppressive-type TME to hot

immune responsive type. When combined with immune checkpoint blockade, robust antitumor immune response was induced to destroy tumors [110].

Liu et al., developed an in situ gelation system by mixing immune adjuvant NP and PS-modified CAT together with PEG-diacrylate to induce immune responses after PDT. The mixed precursor solutions were locally injected followed by irradiation to induce gelation. The tumor-resident hybrid gel changed the tumor  $sO_2$  by decomposing  $H_2O_2$  to  $O_2$ . The changes were quantitatively measured using PAI. Tumors on mice treated with precursor materials without irradiation showed increased  $sO_2$  levels in the first 2 h. However,  $sO_2$  levels decreased at later time points due to rapid tumor clearance. When they irradiated the tumor with precursor materials, in situ gelation occurred, and their  $sO_2$  levels exhibited no significant increase in the first 2 h due to oxygen consumption by PDT. After 2 h, due to long-term tumor retention, tumors in this group showed greatly increased levels of  $sO_2$  even 48 h post-injection. In order to confirm tumor  $sO_2$  analysis by PAI, immunofluorescence staining of hypoxia was performed, which correlated with PAI results. In addition, multi-round PDT combined with a-CTLA4 inhibited metastasis offered long-term immune memory protection from tumor rechallenge [111].

#### **5.** Conclusions and Future Perspectives

Attributes of tumor hypoxia pose a significant risk and limitations to anticancer therapies, such as PDT, RT, and SDT. Reduced oxygen tensions in tumor hypoxia not only impair the production of ROS to kill cancer cells but also exacerbate the condition by consuming oxygen. As a consequence, it is quite evident that the detection and quantitative assessment of oxygen levels in the tumor are crucial. Specifically, it will assist to classify patients based on their hypoxic tumor status and guide optimal decision making in the therapy management of patients. Vast advances in interdisciplinary sciences, such as materials, physics and chemistry, have resulted in the development of sophisticated nanomaterials to increase tumor oxygenation levels by either releasing or generating oxygen. In this review, we summarized the following: (1) the key nanomaterial-based strategies to increase tumor oxygenation, which include (a) oxygen-releasing strategies, in which nanomaterials act as a host to oxygenphilic materials, such as PFCs and Hb, and (b) oxygen is generated by in situ reactions of CAT or CAT-like nanomaterials with endogenous  $H_2O_2$ , and (2) non-invasive PAI to monitor tumor hypoxia and quantify nanomaterials-mediated increase in tumor oxygenation in a real-time manner.

Although the results are encouraging, four main concerns must be addressed prior to transition into clinics: (1) the feasibility of large-scale industrial production of these functional nanomaterials with quality control; (2) proper understanding of the in vivo fate of nanomaterials, such as their biodistribution and excretion; (3) since the premature release of oxygen can cause toxicity to normal tissues, controlled release of oxygen from nanomaterials is desired; and (4) due to dissipation of photons in the tissues, the penetration depth of PAI is currently restricted to  $\approx 10$  cm. Due to rapid advances in nanotechnology and PAI, we believe that the above challenges could be fully addressed to effectively detect, monitor, and overcome tumor hypoxia.

**Author Contributions:** L.-W.L. conceptualized the idea, supervised the manuscript and M.S. wrote the manuscript. All authors have read and agreed to the published version of the manuscript.

**Funding:** This publication was supported by the National Health Research Institutes of Taiwan under award No. BN-110-PP04.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Data are contained within the article.

Conflicts of Interest: The authors declare no conflict of interest.

## Abbreviations

CDT	Chemodynamic therapy
CAT	Catalase
CTLA4	Cytotoxic T-lymphocyte-associated protein 4
DOX	Doxorubicin
FDC	Perfluorodecalvn
GOx	Glucoseoxidase
HIF-1α	Hypoxia-inducible factor- $1\alpha$
Hb	Hemoglobin
HBOT	Hyperbaric oxygen therapy
HPV	Human papilloma virus
IDO	Indoleamine 2,3-dioxygenase
i.v.	Intravenous
i.t.	Intratumor
ICG	Indocyanine green
MRI	Magnetic resonance imaging
NP	Nanoparticles
NIR	Near infrared
pO <sub>2</sub>	Partial pressure of oxygen
PDT	Photodynamic therapy
PD-L1	Programmed death-ligand 1
PD-1	Programmed cell death protein 1
PS	Photosensitizer
PFC	Perfluorocarbon
PFOB	Perfluorooctyl bromide
PFH	Perfluorohexane
PFTA	Perfluorotributylamine
RT	Radiotherapy
ROS	Reactive oxygen species
$sO_2$	Oxygen saturation
SDT	Sonodynamic therapy
SBH-PACT	Single breath hold-photoacoustic computed tomography
US	Ultrasound

#### References

- 1. Attia, A.B.E.; Balasundaram, G.; Moothanchery, M.; Dinish, U.S.; Bi, R.; Ntziachristos, V.; Olivo, M. A review of clinical photoacoustic imaging: Current and future trends. *Photoacoustics* **2019**, *16*, 100144. [CrossRef] [PubMed]
- Gargiulo, S.; Albanese, S.; Mancini, M. State-of-the-Art Preclinical Photoacoustic Imaging in Oncology: Recent Advances in Cancer Theranostics. *Contrast Media Mol. Imaging* 2019, 2019, 5080267. [CrossRef] [PubMed]
- Wang, L.V.; Gao, L. Photoacoustic Microscopy and Computed Tomography: From Bench to Bedside. Ann. Rev. Biomed. Eng. 2014, 16, 155–185. [CrossRef] [PubMed]
- Zackrisson, S.; van de Ven, S.M.W.Y.; Gambhir, S.S. Light In and Sound Out: Emerging Translational Strategies for Photoacoustic Imaging. *Cancer Res.* 2014, 74, 979–1004. [CrossRef]
- 5. Mohammad, M.; Soon Joon, Y.; Douglas, Y.; Stanislav, Y.E. Photoacoustic Imaging for Cancer Detection and Staging. *Curr. Mol. Imaging* **2013**, *2*, 89–105. [CrossRef]
- 6. Yao, J.; Wang, L.V. Recent progress in photoacoustic molecular imaging. Curr. Opin. Chem. Biol. 2018, 45, 104–112. [CrossRef]
- Taruttis, A.; van Dam, G.M.; Ntziachristos, V. Mesoscopic and Macroscopic Optoacoustic Imaging of Cancer. *Cancer Res.* 2015, 75, 1548–1559. [CrossRef]
- Needles, A.; Heinmiller, A.; Ephrat, P.; Bilan-Tracey, C.; Trujillo, A.; Theodoropoulos, C.; Hirson, D.; Foster, F.S. Development of a Combined Photoacoustic Micro-Ultrasound System for Estimating Blood Oxygenation. In Proceedings of the 2010 IEEE International Ultrasonics Symposium, San Diego, CA, USA, 11–14 October 2010; pp. 390–393.
- 9. Li, L.; Wang, L.V. Recent Advances in Photoacoustic Tomography. BME Front. 2021, 2021, 9823268. [CrossRef]
- 10. Sheng, Y.; Liao, L.-D.; Bandla, A.; Liu, Y.-H.; Yuan, J.; Thakor, N.; Tan, M.C. Enhanced near-infrared photoacoustic imaging of silica-coated rare-earth doped nanoparticles. *Mater. Sci. Eng. C Mater. Biol. Appl.* **2017**, *70*, 340–346. [CrossRef]
- 11. Kong, K.V.; Liao, L.-D.; Lam, Z.; Leong, W.K.T.; Olivo, M. Organometallic carbonyl clusters: A new class of contrast agents for photoacoustic cerebral vascular imaging. *Chem. Commun.* **2014**, *50*, 2601–2603. [CrossRef]

- 12. Zhao, X.; Sheng, Y.; Liao, L.-D.; Thakor, N.; Tan, M.C. Rare-Earth Doped CaF<sub>2</sub> Nanocrystals for Dual-Modal Short-Wavelength Infrared Fluorescence and Photoacoustic Imaging. *Nanosci. Nanotechnol. Lett.* **2017**, *9*, 481–488. [CrossRef]
- Cai, X.; Zhang, C.-J.; Lim, F.T.W.; Chan, S.J.; Bandla, A.; Chuan, C.K.; Hu, F.; Xu, S.; Thakor, N.V.; Liao, L.-D.; et al. Organic Nanoparticles with Aggregation-Induced Emission for Bone Marrow Stromal Cell Tracking in a Rat PTI Model. *Small* 2016, 12, 6576–6585. [CrossRef]
- Liu, J.; Cai, X.; Pan, H.-C.; Bandla, A.; Chuan, C.K.; Wang, S.; Thakor, N.; Liao, L.-D.; Liu, B. Molecular Engineering of Photoacoustic Performance by Chalcogenide Variation in Conjugated Polymer Nanoparticles for Brain Vascular Imaging. *Small* 2018, 14, e1703732. [CrossRef]
- Geng, J.; Liao, L.-D.; Qin, W.; Tang, B.Z.; Thakor, N.; Liu, B. Fluorogens with Aggregation Induced Emission: Ideal Photoacoustic Contrast Reagents Due to Intramolecular Rotation. *J. Nanosci. Nanotechnol.* 2015, 15, 1864–1868. [CrossRef]
- 16. Petrova, V.; Annicchiarico-Petruzzelli, M.; Melino, G.; Amelio, I. The hypoxic tumour microenvironment. *Oncogenesis* **2018**, 7, 10. [CrossRef]
- 17. Denko, N.C. Hypoxia, HIF1 and glucose metabolism in the solid tumour. Nat. Rev. Cancer 2008, 8, 705–713. [CrossRef]
- Walsh, J.C.; Lebedev, A.; Aten, E.; Madsen, K.; Marciano, L.; Kolb, H.C. The Clinical Importance of Assessing Tumor Hypoxia: Relationship of Tumor Hypoxia to Prognosis and Therapeutic Opportunities. *Antioxid. Redox. Signal.* 2014, 21, 1516–1554. [CrossRef]
- 19. Sørensen, B.S.; Horsman, M.R. Tumor Hypoxia: Impact on Radiation Therapy and Molecular Pathways. *Front. Oncol.* 2020, 10, 562. [CrossRef]
- Hompland, T.; Fjeldbo, C.S.; Lyng, H. Tumor Hypoxia as a Barrier in Cancer Therapy: Why Levels Matter. *Cancers* 2021, 13, 499. [CrossRef]
- 21. Jing, X.; Yang, F.; Shao, C.; Wei, K.; Xie, M.; Shen, H.; Shu, Y. Role of hypoxia in cancer therapy by regulating the tumor microenvironment. *Mol. Cancer* **2019**, *18*, 157. [CrossRef]
- 22. Moen, I.; Stuhr, L.E.B. Hyperbaric oxygen therapy and cancer—A review. Target. Oncol. 2012, 7, 233–242. [CrossRef]
- 23. Tibbles, P.M.; Edelsberg, J.S. Hyperbaric-Oxygen Therapy. N. Engl. J. Med. 1996, 334, 1642–1648. [CrossRef]
- Ortega, M.A.; Fraile-Martinez, O.; García-Montero, C.; Callejón-Peláez, E.; Sáez, M.A.; Álvarez-Mon, M.A.; García-Honduvilla, N.; Monserrat, J.; Álvarez-Mon, M.; Bujan, J.; et al. A General Overview on the Hyperbaric Oxygen Therapy: Applications, Mechanisms and Translational Opportunities. *Medicina* 2021, 57, 864. [CrossRef]
- Chen, S.-Y.; Tsuneyama, K.; Yen, M.-H.; Lee, J.-T.; Chen, J.-L.; Huang, S.-M. Hyperbaric oxygen suppressed tumor progression through the improvement of tumor hypoxia and induction of tumor apoptosis in A549-cell-transferred lung cancer. *Sci. Rep.* 2021, 11, 12033. [CrossRef]
- 26. Weaver, L.K.; Hopkins, R.O.; Chan, K.J.; Churchill, S.; Elliott, C.G.; Clemmer, T.P.; Orme, J.F.; Thomas, F.O.; Morris, A.H. Hyperbaric Oxygen for Acute Carbon Monoxide Poisoning. *N. Engl. J. Med.* **2002**, *347*, 1057–1067. [CrossRef]
- 27. Li, X.; Wu, Y.; Zhang, R.; Bai, W.; Ye, T.; Wang, S. Oxygen-Based Nanocarriers to Modulate Tumor Hypoxia for Ameliorated Anti-Tumor Therapy: Fabrications, Properties, and Future Directions. *Front. Mol. Biosci.* **2021**, *8*, 683519. [CrossRef]
- Zou, M.-Z.; Liu, W.-L.; Chen, H.-S.; Bai, X.-F.; Gao, F.; Ye, J.-J.; Cheng, H.; Zhang, X.-Z. Advances in nanomaterials for treatment of hypoxic tumor. *Nat. Sci. Rev.* 2020, *8*, nwaa160. [CrossRef]
- 29. Jahanban-Esfahlan, R.; de la Guardia, M.; Ahmadi, D.; Yousefi, B. Modulating tumor hypoxia by nanomedicine for effective cancer therapy. *J. Cell Physiol.* **2018**, 233, 2019–2031. [CrossRef]
- Wang, J.; Zhang, B.; Sun, J.; Wang, Y.; Wang, H. Nanomedicine-Enabled Modulation of Tumor Hypoxic Microenvironment for Enhanced Cancer Therapy. *Adv. Ther.* 2020, *3*, 1900083. [CrossRef]
- Jägers, J.; Wrobeln, A.; Ferenz, K.B. Perfluorocarbon-based oxygen carriers: From physics to physiology. *Pflügers Arch. Eur. J. Phys.* 2021, 473, 139–150. [CrossRef]
- Castro, C.I.; Briceno, J.C. Perfluorocarbon-Based Oxygen Carriers: Review of Products and Trials. Artif. Organs 2010, 34, 622–634. [CrossRef] [PubMed]
- Hu, H.; Yan, X.; Wang, H.; Tanaka, J.; Wang, M.; You, W.; Li, Z. Perfluorocarbon-based O2 nanocarrier for efficient photodynamic therapy. J. Mater. Chem. B 2019, 7, 1116–1123. [CrossRef] [PubMed]
- Song, G.; Liang, C.; Yi, X.; Zhao, Q.; Cheng, L.; Yang, K.; Liu, Z. Perfluorocarbon-Loaded Hollow Bi2Se3 Nanoparticles for Timely Supply of Oxygen under Near-Infrared Light to Enhance the Radiotherapy of Cancer. *Adv. Mater.* 2016, 28, 2716–2723. [CrossRef] [PubMed]
- 35. Hu, D.; Pan, M.; Yu, Y.; Sun, A.; Shi, K.; Qu, Y.; Qian, Z. Application of nanotechnology for enhancing photodynamic therapy via ameliorating, neglecting, or exploiting tumor hypoxia. *VIEW* **2020**, *1*, e6. [CrossRef]
- Zhang, Z.; Ji, Y. Nanostructured manganese dioxide for anticancer applications: Preparation, diagnosis, and therapy. *Nanoscale* 2020, 12, 17982–18003. [CrossRef]
- Prasad, P.; Gordijo, C.R.; Abbasi, A.Z.; Maeda, A.; Ip, A.; Rauth, A.M.; DaCosta, R.S.; Wu, X.Y. Multifunctional Albumin–MnO<sub>2</sub> Nanoparticles Modulate Solid Tumor Microenvironment by Attenuating Hypoxia, Acidosis, Vascular Endothelial Growth Factor and Enhance Radiation Response. ACS Nano 2014, 8, 3202–3212. [CrossRef]
- Zhang, J.; Duan, F.; Liu, Y.; Nie, L. High-Resolution Photoacoustic Tomography for Early-Stage Cancer Detection and Its Clinical Translation. *Radiol. Imaging Cancer* 2020, 2, e190030. [CrossRef]

- 39. Wang, L.V.; Hu, S. Photoacoustic Tomography: In Vivo Imaging from Organelles to Organs. *Science* **2012**, *335*, 1458–1462. [CrossRef]
- 40. Cao, Y.; Kole, A.; Hui, J.; Zhang, Y.; Mai, J.; Alloosh, M.; Sturek, M.; Cheng, J.-X. Fast assessment of lipid content in arteries in vivo by intravascular photoacoustic tomography. *Sci. Rep.* **2018**, *8*, 2400. [CrossRef]
- 41. Zhou, Y.; Zhang, C.; Yao, D.-K.; Wang, L. Photoacoustic microscopy of bilirubin in tissue phantoms. *J. Biomed. Opt.* **2012**, 17, 126019. [CrossRef]
- 42. Zhang, C.; Zhang, Y.S.; Yao, D.-K.; Xia, Y.; Wang, L. Label-free photoacoustic microscopy of cytochromes. *J. Biomed. Opt.* **2013**, 18, 20504. [CrossRef]
- Yao, D.-K.; Maslov, K.; Shung, K.K.; Zhou, Q.; Wang, L.V. In vivo label-free photoacoustic microscopy of cell nuclei by excitation of DNA and RNA. *Opt. Lett.* 2010, 35, 4139–4141. [CrossRef]
- 44. Xu, Z.; Li, C.; Wang, L. Photoacoustic tomography of water in phantoms and tissue. J. Biomed. Opt. 2010, 15, 036019. [CrossRef]
- 45. Harbeck, N.; Penault-Llorca, F.; Cortes, J.; Gnant, M.; Houssami, N.; Poortmans, P.; Ruddy, K.; Tsang, J.; Cardoso, F. Breast cancer. *Nat. Rev. Dis. Primers* **2019**, *5*, 66. [CrossRef]
- 46. Waks, A.G.; Winer, E.P. Breast Cancer Treatment: A Review. JAMA 2019, 321, 288–300. [CrossRef]
- 47. Lewin, J.M.; Patel, B.K.; Tanna, A. Contrast-Enhanced Mammography: A Scientific Review. J. Breast Imaging 2019, 2, 7–15. [CrossRef]
- Keating, N.L.; Pace, L.E. Breast Cancer Screening in 2018: Time for Shared Decision Making. JAMA 2018, 319, 1814–1815. [CrossRef]
- Gardezi, S.J.S.; Elazab, A.; Lei, B.; Wang, T. Breast Cancer Detection and Diagnosis Using Mammographic Data: Systematic Review. J. Med. Internet Res. 2019, 21, e14464. [CrossRef]
- 50. Løberg, M.; Lousdal, M.L.; Bretthauer, M.; Kalager, M. Benefits and harms of mammography screening. *Breast Cancer Res.* 2015, 17, 63. [CrossRef]
- Nelson, H.D.; O'Meara, E.S.; Kerlikowske, K.; Balch, S.; Miglioretti, D. Factors Associated with Rates of False-Positive and False-Negative Results From Digital Mammography Screening: An Analysis of Registry Data. Ann. Intern. Med. 2016, 164, 226–235. [CrossRef]
- 52. Lin, L.; Hu, P.; Shi, J.; Appleton, C.M.; Maslov, K.; Li, L.; Zhang, R.; Wang, L.V. Single-breath-hold photoacoustic computed tomography of the breast. *Nat. Commun.* **2018**, *9*, 2352. [CrossRef]
- 53. Toi, M.; Asao, Y.; Matsumoto, Y.; Sekiguchi, H.; Yoshikawa, A.; Takada, M.; Kataoka, M.; Endo, T.; Kawaguchi-Sakita, N.; Kawashima, M.; et al. Visualization of tumor-related blood vessels in human breast by photoacoustic imaging system with a hemispherical detector array. *Sci. Rep.* 2017, *7*, 41970. [CrossRef]
- 54. Dogra, V.S.; Chinni, B.K.; Valluru, K.S.; Joseph, J.V.; Ghazi, A.; Yao, J.L.; Evans, K.; Messing, E.M.; Rao, N.A. Multispectral Photoacoustic Imaging of Prostate Cancer: Preliminary Ex-vivo Results. *J. Clin. Imaging Sci.* **2013**, *3*, 41. [CrossRef]
- Wang, Y.; Jhang, D.-F.; Tsai, C.-H.; Chiang, N.-J.; Tsao, C.-H.; Chuang, C.-C.; Chen, L.-T.; Chang, W.-S.W.; Liao, L.-D. In Vivo Assessment of Hypoxia Levels in Pancreatic Tumors Using a Dual-Modality Ultrasound/Photoacoustic Imaging System. *Micromachines* 2021, 12, 668. [CrossRef]
- 56. Rich, L.J.; Miller, A.; Singh, A.K.; Seshadri, M. Photoacoustic Imaging as an Early Biomarker of Radio Therapeutic Efficacy in Head and Neck Cancer. *Theranostics* **2018**, *8*, 2064–2078. [CrossRef]
- 57. Rich, L.J.; Seshadri, M. Photoacoustic monitoring of tumor and normal tissue response to radiation. *Sci. Rep.* **2016**, *6*, 21237. [CrossRef]
- 58. Wilson, W.R.; Hay, M.P. Targeting hypoxia in cancer therapy. Nat. Rev. Cancer 2011, 11, 393–410. [CrossRef]
- 59. Vaupel, P.; Mayer, A. Hypoxia in cancer: Significance and impact on clinical outcome. *Cancer Metastasis Rev.* 2007, *26*, 225–239. [CrossRef]
- 60. Ferenz, K.B.; Steinbicker, A.U. Artificial Oxygen Carriers—Past, Present, and Future—A Review of the Most Innovative and Clinically Relevant Concepts. J. Pharmacol. Exp. Ther. 2019, 369, 300–310. [CrossRef]
- 61. Gupta, A.S. Bio-inspired nanomedicine strategies for artificial blood components. *WIREs Nanomed. Nanobiotechnol.* **2017**, *9*, e1464. [CrossRef]
- 62. Tran, T.D.; Caruthers, S.D.; Hughes, M.; Marsh, J.N.; Cyrus, T.; Winter, P.M.; Neubauer, A.M.; Wickline, S.A.; Lanza, G.M. Clinical applications of perfluorocarbon nanoparticles for molecular imaging and targeted therapeutics. *Int. J. Nanomed.* **2007**, *2*, 515–526.
- 63. Winter, P.M.; Cai, K.; Caruthers, S.D.; Wickline, S.A.; Lanza, G.M. Emerging nanomedicine opportunities with perfluorocarbon nanoparticles. *Expert Rev. Med. Devices* **2007**, *4*, 137–145. [CrossRef] [PubMed]
- 64. Riess, J.G. Perfluorocarbon-based Oxygen Delivery. Artif. Cells Blood Substit. Biotechnol. 2006, 34, 567–580. [CrossRef] [PubMed]
- 65. Gould, S.A.; Rosen, A.L.; Sehgal, L.R.; Sehgal, H.L.; Langdale, L.A.; Krause, L.M.; Rice, C.L.; Chamberlin, W.H.; Moss, G.S. Fluosol-DA as a Red-Cell Substitute in Acute Anemia. *N. Engl. J. Med.* **1986**, *314*, 1653–1656. [CrossRef]
- 66. Riess, J.G.; Keipert, P.E. Chapter 7—Update on Perfluorocarbon-Based Oxygen Delivery Systems. In *Blood Substitutes, Present and Future Perspectives*; Tsuchida, E., Ed.; Elsevier Science: Lausanne, Switzerland, 1998; pp. 91–102. [CrossRef]
- 67. Cheng, Y.; Cheng, H.; Jiang, C.; Qiu, X.; Wang, K.; Huan, W.; Yuan, A.; Wu, J.; Hu, Y. Perfluorocarbon nanoparticles enhance reactive oxygen levels and tumour growth inhibition in photodynamic therapy. *Nat. Commun.* **2015**, *6*, 8785. [CrossRef]

- Jiang, W.; Li, Q.; Xiao, L.; Dou, J.; Liu, Y.; Yu, W.; Ma, Y.; Li, X.; You, Y.-Z.; Tong, Z.; et al. Hierarchical Multiplexing Nanodroplets for Imaging-Guided Cancer Radiotherapy via DNA Damage Enhancement and Concomitant DNA Repair Prevention. ACS Nano 2018, 12, 5684–5698. [CrossRef]
- Zhao, C.; Tong, Y.; Li, X.; Shao, L.; Chen, L.; Lu, J.; Deng, X.; Wang, X.; Wu, Y. Photosensitive Nanoparticles Combining Vascular-Independent Intratumor Distribution and On-Demand Oxygen-Depot Delivery for Enhanced Cancer Photodynamic Therapy. *Small* 2018, 14, 1703045. [CrossRef]
- 70. Song, G.; Ji, C.; Liang, C.; Song, X.; Yi, X.; Dong, Z.; Yang, K.; Liu, Z. TaOx decorated perfluorocarbon nanodroplets as oxygen reservoirs to overcome tumor hypoxia and enhance cancer radiotherapy. *Biomaterials* **2017**, *112*, 257–263. [CrossRef]
- 71. Zhou, Z.; Zhang, B.; Wang, H.; Yuan, A.; Hu, Y.; Wu, J. Two-stage oxygen delivery for enhanced radiotherapy by perfluorocarbon nanoparticles. *Theranostics* **2018**, *8*, 4898–4911. [CrossRef]
- Zhang, Y.; Bo, S.; Feng, T.; Qin, X.; Wan, Y.; Jiang, S.; Li, C.; Lin, J.; Wang, T.; Zhou, X.; et al. A Versatile Theranostic Nanoemulsion for Architecture-Dependent Multimodal Imaging and Dually Augmented Photodynamic Therapy. *Adv. Mater.* 2019, *31*, e1806444. [CrossRef]
- 73. Li, F.; Mei, H.; Gao, Y.; Xie, X.; Nie, H.; Li, T.; Zhang, H.; Jia, L. Co-delivery of oxygen and erlotinib by aptamer-modified liposomal complexes to reverse hypoxia-induced drug resistance in lung cancer. *Biomaterials* **2017**, *145*, 56–71. [CrossRef]
- Xing, L.; Gong, J.-H.; Wang, Y.; Zhu, Y.; Huang, Z.-J.; Zhao, J.; Li, F.; Wang, J.-H.; Wen, H.; Jiang, H.-L. Hypoxia alleviationtriggered enhanced photodynamic therapy in combination with IDO inhibitor for preferable cancer therapy. *Biomaterials* 2019, 206, 170–182. [CrossRef]
- 75. Yu, P.; Han, X.; Yin, L.; Hui, K.; Guo, Y.; Yuan, A.; Hu, Y.; Wu, J. Artificial Red Blood Cells Constructed by Replacing Heme with Perfluorodecalin for Hypoxia-Induced Radioresistance. *Adv. Ther.* **2019**, *2*, 1900031. [CrossRef]
- Chen, J.; Luo, H.; Liu, Y.; Zhang, W.; Li, H.; Luo, T.; Zhang, K.; Zhao, Y.; Liu, J. Oxygen-Self-Produced Nanoplatform for Relieving Hypoxia and Breaking Resistance to Sonodynamic Treatment of Pancreatic Cancer. ACS Nano 2017, 11, 12849–12862. [CrossRef]
- 77. Vandegriff, K.D. Haemoglobin-based oxygen carriers. Expert Opin. Investig. Drugs 2000, 9, 1967–1984. [CrossRef]
- Riess, J.G. Oxygen Carriers ("Blood Substitutes") Raison d'Etre, Chemistry, and Some Physiology Blut ist ein ganz besondrer Saft. Chem. Rev. 2001, 101, 2797–2920. [CrossRef]
- 79. Palmer, A.F.; Intaglietta, M. Blood Substitutes. Annu. Rev. Biomed. Eng. 2014, 16, 77–101. [CrossRef]
- 80. Buehler, P.W.; D'Agnillo, F.; Schaer, D.J. Hemoglobin-based oxygen carriers: From mechanisms of toxicity and clearance to rational drug design. *Trends Mol. Med.* 2010, *16*, 447–457. [CrossRef]
- Standl, T. Artificial Oxygen Carriers: Hemoglobin-Based Oxygen Carriers—Current Status 2004. Transfus. Med. Hemother. 2004, 31, 262–268. [CrossRef]
- Jiang, L.; Bai, H.; Liu, L.; Lv, F.; Ren, X.; Wang, S. Luminescent, Oxygen-Supplying, Hemoglobin-Linked Conjugated Polymer Nanoparticles for Photodynamic Therapy. *Angew. Chem. Int. Ed.* 2019, 58, 10660–10665. [CrossRef]
- 83. Cao, H.; Wang, L.; Yang, Y.; Li, J.; Qi, Y.; Li, Y.; Li, Y.; Wang, H.; Li, J. An Assembled Nanocomplex for Improving both Therapeutic Efficiency and Treatment Depth in Photodynamic Therapy. *Angew. Chem. Int. Ed.* **2018**, *57*, 7759–7763. [CrossRef] [PubMed]
- Yang, J.; Li, W.; Luo, L.; Jiang, M.; Zhu, C.; Qin, B.; Yin, H.; Yuan, X.; Yin, X.; Zhang, J.; et al. Hypoxic tumor therapy by hemoglobin-mediated drug delivery and reversal of hypoxia-induced chemoresistance. *Biomaterials* 2018, 182, 145–156. [CrossRef] [PubMed]
- Guo, X.; Qu, J.; Zhu, C.; Li, W.; Luo, L.; Yang, J.; Yin, X.; Li, Q.; Du, Y.; Chen, D.; et al. Synchronous delivery of oxygen and photosensitizer for alleviation of hypoxia tumor microenvironment and dramatically enhanced photodynamic therapy. *Drug Deliv.* 2018, 25, 585–599. [CrossRef] [PubMed]
- 86. Yuan, M.; Liang, S.; Zhou, Y.; Xiao, X.; Liu, B.; Yang, C.; Ma, P.; Cheng, Z.; Lin, J. A Robust Oxygen-Carrying Hemoglobin-Based Natural Sonosensitizer for Sonodynamic Cancer Therapy. *Nano Lett.* **2021**, *21*, 6042–6050. [CrossRef] [PubMed]
- Xia, D.; Hang, D.; Li, Y.; Jiang, W.; Zhu, J.; Ding, Y.; Gu, H.; Hu, Y. Au–Hemoglobin Loaded Platelet Alleviating Tumor Hypoxia and Enhancing the Radiotherapy Effect with Low-Dose X-ray. ACS Nano 2020, 14, 15654–15668. [CrossRef] [PubMed]
- Wang, H.; Chao, Y.; Liu, J.; Zhu, W.; Wang, G.; Xu, L.; Liu, Z. Photosensitizer-crosslinked in-situ polymerization on catalase for tumor hypoxia modulation & enhanced photodynamic therapy. *Biomaterials* 2018, 181, 310–317. [CrossRef] [PubMed]
- Cormode, D.P.; Gao, L.; Koo, H. Emerging Biomedical Applications of Enzyme-Like Catalytic Nanomaterials. *Trends Biotechnol.* 2018, 36, 15–29. [CrossRef]
- Zhang, R.; Song, X.; Liang, C.; Yi, X.; Song, G.; Chao, Y.; Yang, Y.; Yang, K.; Feng, L.; Liu, Z. Catalase-loaded cisplatin-prodrugconstructed liposomes to overcome tumor hypoxia for enhanced chemo-radiotherapy of cancer. *Biomaterials* 2017, 138, 13–21. [CrossRef]
- 91. Phua, S.Z.F.; Yang, G.; Lim, W.Q.; Verma, A.; Chen, H.; Thanabalu, T.; Zhao, Y. Catalase-Integrated Hyaluronic Acid as Nanocarriers for Enhanced Photodynamic Therapy in Solid Tumor. *ACS Nano* 2019, *13*, 4742–4751. [CrossRef]
- Zou, M.-Z.; Liu, W.-L.; Li, C.-X.; Zheng, D.-W.; Zeng, J.-Y.; Gao, F.; Ye, J.-J.; Zhang, X.-Z. A Multifunctional Biomimetic Nanoplatform for Relieving Hypoxia to Enhance Chemotherapy and Inhibit the PD-1/PD-L1 Axis. *Small* 2018, 14, e1801120. [CrossRef]
- Shi, C.; Li, M.; Zhang, Z.; Yao, Q.; Shao, K.; Xu, F.; Xu, N.; Li, H.; Fan, J.; Sun, W.; et al. Catalase-based liposomal for reversing immunosuppressive tumor microenvironment and enhanced cancer chemo-photodynamic therapy. *Biomaterials* 2020, 233, 119755. [CrossRef]

- Zhu, J.; Xiao, T.; Zhang, J.; Che, H.; Shi, Y.; Shi, X.; van Hest, J.C.M. Surface-Charge-Switchable Nanoclusters for Magnetic Resonance Imaging-Guided and Glutathione Depletion-Enhanced Photodynamic Therapy. ACS Nano 2020, 14, 11225–11237. [CrossRef]
- Yang, Z.; Du, Y.; Sun, Q.; Peng, Y.; Wang, R.; Zhou, Y.; Wang, Y.; Zhang, C.; Qi, X. Albumin-Based Nanotheranostic Probe with Hypoxia Alleviating Potentiates Synchronous Multimodal Imaging and Phototherapy for Glioma. ACS Nano 2020, 14, 6191–6212. [CrossRef]
- 96. Ding, B.; Zheng, P.; Ma, P.A.; Lin, J. Manganese Oxide Nanomaterials: Synthesis, Properties, and Theranostic Applications. *Adv. Mater.* **2020**, *32*, e1905823. [CrossRef]
- Fu, C.; Duan, X.; Cao, M.; Jiang, S.; Ban, X.; Guo, N.; Zhang, F.; Mao, J.; Huyan, T.; Shen, J.; et al. Targeted Magnetic Resonance Imaging and Modulation of Hypoxia with Multifunctional Hyaluronic Acid-MnO<sub>2</sub> Nanoparticles in Glioma. *Adv. Healthc. Mater.* 2019, *8*, e1900047. [CrossRef]
- 98. Lin, T.; Zhao, X.; Zhao, S.; Yu, H.; Cao, W.; Chen, W.; Wei, H.; Guo, H. O(2)-generating MnO(2) nanoparticles for enhanced photodynamic therapy of bladder cancer by ameliorating hypoxia. *Theranostics* **2018**, *8*, 990–1004. [CrossRef]
- 99. Zhu, X.; Liu, Y.; Yuan, G.; Guo, X.; Cen, J.; Gong, Y.; Liu, J.; Gang, Y. In situ fabrication of MS@MnO2 hybrid as nanozymes for enhancing ROS-mediated breast cancer therapy. *Nanoscale* **2020**, *12*, 22317–22329. [CrossRef]
- Yang, G.; Xu, L.; Chao, Y.; Xu, J.; Sun, X.; Wu, Y.; Peng, R.; Liu, Z. Hollow MnO<sub>2</sub> as a tumor-microenvironment-responsive biodegradable nano-platform for combination therapy favoring antitumor immune responses. *Nat. Commun.* 2017, *8*, 902. [CrossRef]
- 101. Hu, J.-J.; Chen, Y.; Li, Z.-H.; Peng, S.-Y.; Sun, Y.; Zhang, X.-Z. Augment of Oxidative Damage with Enhanced Photodynamic Process and MTH1 Inhibition for Tumor Therapy. *Nano Lett.* **2019**, *19*, 5568–5576. [CrossRef]
- 102. Zhou, J.; Li, M.; Hou, Y.; Luo, Z.; Chen, Q.; Cao, H.; Huo, R.; Xue, C.; Sutrisno, L.; Hao, L.; et al. Engineering of a Nanosized Biocatalyst for Combined Tumor Starvation and Low-Temperature Photothermal Therapy. ACS Nano 2018, 12, 2858–2872. [CrossRef]
- Nordsmark, M.; Loncaster, J.; Aquino-Parsons, C.; Chou, S.-C.; Ladekarl, M.; Havsteen, H.; Lindegaard, J.C.; Davidson, S.E.; Varia, M.; West, C.; et al. Measurements of hypoxia using pimonidazole and polarographic oxygen-sensitive electrodes in human cervix carcinomas. *Radiother. Oncol.* 2003, 67, 35–44. [CrossRef]
- Bentzen, L.; Keiding, S.; Nordsmark, M.; Falborg, L.; Hansen, S.B.; Keller, J.; Nielsen, O.S.; Overgaard, J. Tumour oxygenation assessed by 18F-fluoromisonidazole PET and polarographic needle electrodes in human soft tissue tumours. *Radiother. Oncol.* 2003, 67, 339–344. [CrossRef]
- 105. Mallidi, S.; Watanabe, K.; Timerman, D.; Schoenfeld, D.; Hasan, T. Prediction of Tumor Recurrence and Therapy Monitoring Using Ultrasound-Guided Photoacoustic Imaging. *Theranostics* 2015, *5*, 289–301. [CrossRef] [PubMed]
- 106. Hysi, E.; Wirtzfeld, L.A.; May, J.P.; Undzys, E.; Li, S.-D.; Kolios, M.C. Photoacoustic signal characterization of cancer treatment response: Correlation with changes in tumor oxygenation. *Photoacoustics* **2017**, *5*, 25–35. [CrossRef] [PubMed]
- 107. Huang, Y.; Shen, K.; Si, Y.; Shan, C.; Guo, H.; Chen, M.; Wu, L. Dendritic organosilica nanospheres with large mesopores as multi-guests vehicle for photoacoustic/ultrasound imaging-guided photodynamic therapy. J. Colloid Interface Sci. 2021, 583, 166–177. [CrossRef] [PubMed]
- 108. Yang, K.; Yue, L.; Yu, G.; Rao, L.; Tian, R.; Wei, J.; Yang, Z.; Sun, C.; Zhang, X.; Xu, M.; et al. A hypoxia responsive nanoassembly for tumor specific oxygenation and enhanced sonodynamic therapy. *Biomaterials* **2021**, 275, 120822. [CrossRef]
- 109. Wu, M.; Zheng, D.; Zhang, D.; Yu, P.; Peng, L.; Chen, F.; Lin, Z.; Cai, Z.; Li, J.; Wei, Z.; et al. Converting Immune Cold into Hot by Biosynthetic Functional Vesicles to Boost Systematic Antitumor Immunity. *iScience* **2020**, 23, 101341. [CrossRef]
- Song, X.; Xu, J.; Liang, C.; Chao, Y.; Jin, Q.; Wang, C.; Chen, M.; Liu, Z. Self-Supplied Tumor Oxygenation through Separated Liposomal Delivery of H2O2 and Catalase for Enhanced Radio-Immunotherapy of Cancer. *Nano Lett.* 2018, 18, 6360–6368. [CrossRef]
- 111. Meng, Z.; Zhou, X.; Xu, J.; Han, X.; Dong, Z.; Wang, H.; Zhang, Y.; She, J.; Xu, L.; Wang, C.; et al. Light-Triggered In Situ Gelation to Enable Robust Photodynamic-Immunotherapy by Repeated Stimulations. *Adv. Mater.* **2019**, *31*, e1900927. [CrossRef]
- 112. Zhao, L.-P.; Zheng, R.-R.; Chen, H.-Q.; Liu, L.-S.; Zhao, X.-Y.; Liu, H.-H.; Qiu, X.-Z.; Yu, X.-Y.; Cheng, H.; Li, S.-Y. Self-Delivery Nanomedicine for O2-Economized Photodynamic Tumor Therapy. *Nano Lett.* 2020, 20, 2062–2071. [CrossRef]
- 113. Chen, H.; Fu, Y.; Feng, K.; Zhou, Y.; Wang, X.; Huang, H.; Chen, Y.; Wang, W.; Xu, Y.; Tian, H.; et al. Polydopamine-coated UiO-66 nanoparticles loaded with perfluorotributylamine/tirapazamine for hypoxia-activated osteosarcoma therapy. *J. Nanobiotechnol.* 2021, 19, 298. [CrossRef]
- 114. Xavierselvan, M.; Cook, J.; Duong, J.; Diaz, N.; Homan, K.; Mallidi, S. Photoacoustic nanodroplets for oxygen enhanced photodynamic therapy of cancer. *Photoacoustics* **2022**, *25*, 100306. [CrossRef]
- 115. Wang, J.; Huang, J.; Zhou, W.; Zhao, J.; Peng, Q.; Zhang, L.; Wang, Z.; Li, P.; Li, R. Hypoxia modulation by dual-drug nanoparticles for enhanced synergistic sonodynamic and starvation therapy. *J. Nanobiotechnol.* **2021**, *19*, 87. [CrossRef]
- 116. Lei, S.; Zhang, J.; Blum, N.T.; Li, M.; Zhang, D.-Y.; Yin, W.; Zhao, F.; Lin, J.; Huang, P. In vivo three-dimensional multispectral photoacoustic imaging of dual enzyme-driven cyclic cascade reaction for tumor catalytic therapy. *Nat. Commun.* 2022, 13, 1298. [CrossRef]