

Supporting Information

Role of Estradiol Hormone in Human Life and Electrochemical Aptasensing of 17 β -Estradiol: A Review

P. P. Waifalkar ^{1,2}, Daegwon Noh ^{1,2}, Poorva Derashri ³, Sagar Barage ^{3,4} and Eunsoon Oh ^{1,2,*}

¹ Department of Physics, Chungnam National University, Daejeon 34134, Republic of Korea

² Institute of QuanTable 34134, Republic of Korea

³ Amity Institute of Biotechnology, Amity University, Maharashtra, Mumbai-Pune Expressway, Panvel, Maharashtra 410206, India

⁴ Centre for Computational Biology and Translational Research, Amity University, Maharashtra, Mumbai-Pune Expressway, Panvel, Maharashtra 410206, India

* Correspondence: esoh@cnu.ac.kr; Tel.: +82-10-6349-5453

Table S1. Electrode materials, aptamer sequences, selectivity in different matrix and environmental sample measurements.

Electrode	Estradiol Specific Aptamer Sequence	Selectivity in a Matrix (Interference Effects from Various Materials)	Environmental Sample Measurements	References
Au/streptavidin/biotin	5'-Biotin-GCTTCCAGCTTATTGAATTACACCGCAGAGG-GTAGCGGCTCTGCGCATTCAATTGCTGCGCGCTGAAG-CGCAGAAGC-3'	2-methoxynaphthalene, 1-aminoanthraquinone	N/A	(Kim et al., 2007) [34]

Electrode	Estradiol Specific Aptamer Sequence	Selectivity in a Matrix (Interference Effects from Various Materials)	Environmental Sample Measurements	References
PEDOT/AuNP /streptavidin	5'-Biotin- GCTTCCAGCTTATTGAATTACACGCAGAGGGTAGCGGCTCT GCGC ATTCAATTGCTGCCGCTGAAGCGCGGAAGC-3'	17 α -ethynestradiol, estrone, naphthalene	N/A	(Olowu et al., 2010) [46]
Au	5'-SH- GCTTCCAGCTTATTGAATTACACGCAGAGGGTAGCGGCTCT GCGC ATTCAATTGCTGCCGCTGAAGCGCG GAAGC-3'	2-methoxynaphthalene, 1-aminoanthraquinone	Urine	(Lin et al., 2012) [37]
GCE/VS2/Au	5'-SH-(CH ₂) ₆ -TTT TTT TTT T GCT TCC AGC TTA TTG AAT TAC ACG CAG AGG GTA GCG GCT CTG CGC ATT CAA TTG CTG CGC GCT GAA GCG CGG AAG C-3'	Naphthalene, 1-aminoanthraquinone	Urine	(Huang et al., 2014a) [50]
BDD/Au	5'- SHGCTTCCAGCTTATTGAATTACACGCAGAGGGTAGCGGCT CT- GCGC ATTCAATTGCTGCCGCTGAAGCGCGGAAGC-3'	Estriol, bisphenol A, nonyl phenol, diethyl phthalate, resorcinol, and atrazine	Wastewater	(Ke et al., 2014) [36]
GCE/WS2 /Au	5'-SH-(CH ₂) ₆ -TTT TTT TTT T GCT TCC AGC TTA TTG AAT TAC ACG CAG AGG GTA GCG GCT CTG CGC ATT CAA TTG CTG CGC GCT GAA GCG CGG AAG C-3'	Naphthalene, 1-aminoanthraquinone	Serum and Water	(Huang et al., 2014) [51]
GCE/CuS/Au/ GO/Au	5'-SH-(CH ₂) ₆ -TTT TTT TTT T GCT TCC AGC TTA TTG AAT TAC ACG CAG AGG GTA GCG GCT CTG CGC ATT CAATTG CTG CGC GCT GAA GCG CGG AAG C-3'	Bisphenol A, 1-aminoanthraquinone and naphthalene	Urine	(Huang et al., 2015a) [52]

Electrode	Estradiol Specific Aptamer Sequence	Selectivity in a Matrix (Interference Effects from Various Materials)	Environmental Sample Measurements	References
GCE/AuNp/ CoS	5'-SH-(CH ₂) ₆ -TTT TTT TTT T GCT TCC AGC TTA TTG AAT TAC ACG CAG AGG GTA GCG GCT CTG CGC ATT CAA TTG CTG CGC GCT GAA GCG CGG AAG C-3'	1-aminoanthraquinone, naphthalene, polychlorinated biphenyl, bisphenol A, phthalic acid ester, testosterone, and cholesterol	Urine	(Huang et al., 2015b) [42]
	cDNA: 5'-GGA GGA G G CTT CCG CGC TTC AGC GCG CAG CAA-3'			
Au/NiHCF/A _u	5'-SH-(CH ₂) ₆ -GCT-TCC-AGC-TTA-TTG-AAT-TAC-ACG-CAG-AGG-GTA-GCG-GCT-CTG-CGC-ATT-CAA-TTG-CTG-CGC-GCT-GAA-GCG-CGG-AAG-C-3' ; Random DNA sequence: 5'-(SH)-(CH ₂) ₆ -CTG-ACA-CCA-TAT-TAT-GAA-GA-3'	Ethinylestradiol, bisphenol A, estriol, polychlorinated biphenyl (PCB 101), diethyl phthalate, 4-nonyl phenol, and atrazine	Municipal wastewater	(Fan et al., 2015) [48]
GCE/poly(Py-co-PAA)	75-mer E2 aptamer: NH ₂ (CH ₂) ₆ - ATACGAGCTTGTCAATACGAAGGGATGCCGTTGGGCCA AGTTCCGGCATAGTGTGGTATAGTAAGAGCAATC	Bisphenol-A, progesterone	Urine and tap water	(Zhu et al., 2015) [39]
	35-mer E2 aptamer: NH ₂ (CH ₂) ₆ - AAGGGATGCCGTTGGGCCAAGTTCGGCATAGTG			
	Adenosine aptamer: NH ₂ (CH ₂) ₆ -ACCTGGGGAGTATTGCGGAGGAAGGT			

Electrode	Estradiol Specific Aptamer Sequence	Selectivity in a Matrix (Interference Effects from Various Materials)	Environmental Sample Measurements	References
	Non-specific ssDNA as control in urine detection: NH ₂ (CH ₂) ₆ - AGGTATTGATCCGCCGGCTAGCATTGTGGTCTCCAGGGG TACTCCCC			
GCE/ERGO	38-mer amine-functionalized ssDNA (NH ₂ -APT)	Estrogen and testosterone	Wastewater and pharmaceutical sample	(Rather et al., 2018) [13]
Au/MCH/Graphene	5'- GCTTCCAGCTTATTGAATTACACCGCAGAGGGTAGCGGCTCT GCGCAT TCAATTGCTGCCGCTGAAGCGCGGAAGC-3'	Estriol, bisphenol A, nolyl phenol, polychlorinated biphenyl and atrazine	Tap water	(Liu et al., 2019) [44]
Screen printed gold electrode	5'-Thiol- TTTTTTTTTTGCTTCCAGCTTATTGAATTACACGCAGAG GGTA-3' (split1); 5'- GCGGCTCTGCCATTCAATTGCTGCCGCTGAAGCGCGAA GCTTTTTTTTT-Thiol-3' (split2)	Estriol, progesterone, dibutyl phthalate, bovine serum albumin, testosterone, atrazine, and bisphenol A	Tap water and milk	(Nameghi et al., 2019) [49]
Carbon/NH ₂ -SWCNT/NMB /AuNP	5'-SH-(CH ₂) ₆ -GCT-TCC-AGC-TTA-TTG-AAT-TAC-ACG-CAG- AGG-GTA-GCG-GCT-CTG-CGC- ATT-CAA-TTG-CTG-CGC-GCT- GAA-GCG-CGGAAG-C-3'	Follicle-stimulating hormone, luteinizing hormone, glutamic acid, ascorbic acid, uric acid, neuron-specific	Serum	(Ming et al., 2019) [54]

Electrode	Estradiol Specific Aptamer Sequence	Selectivity in a Matrix (Interference Effects from Various Materials)	Environmental Sample Measurements	References
		enolase, or carcinoembryonic antigen		
AuNP/thionine/MCNTs	5'-SH-(CH ₂) ₆ -GCTTCCAG CTTATTGAATTACACGCAGAGGGTA GCGGCTCTGCGCATTCAATTGCTGCGCGCTGAAGCGCGAA GC- 3'	Ascorbic acid, dopamine, uric acid, L-lysine, L-cysteine, L-tyrosine, hypoxanthine, progesterone, and cholesterol	Serum	(Liu et al., 2019) [55]
Screen-printed carbon electrode/Carbon nanodot	NH ₂ -5'-GCTTCCAGCTTATTGAATTACACGCAGAGGGTA GCGGCTCTGCGCATTCAATTGCT GCGCGCTGAAGCGCG GAAGC-3'	Estriol, progesterone, and bisphenol A	River water	(Mat Zaid et al., 2020) [40]
GCE and laser-scribed graphene /poly(β-CD)/AF1-ADA/ON1/AF2-Au	The sequence of 5'-amine modified AF 1 was 5'-NH ₂ -AAG GGA TGC CGT TTG GG-3'; the 3' - thiol modified AF2 sequence was 5'-CCC AAG TTC GGC ATA GTG-SH-3'; the ON 1 was 5'-AAG CTT GGG CCA TGC CCA GGA AGG ACC CAA ACG G-3' and ON 2 :5'-CCG TTT GGG TCC TTC CTG GGC ATG GCC CAA GCT T-3'.	Glucose, bisphenol A, progesterone, and bovine serum albumin	Milk	(Chang et al., 2021) [43]

Electrode	Estradiol Specific Aptamer Sequence	Selectivity in a Matrix (Interference Effects from Various Materials)	Environmental Sample Measurements	References
SPCE/poly(pyrrrole-co-pyrrole-3-carboxylic acid)	NH ₂ -5'-ATA CGA GCT TGT TCA ATA CGA AGG GAT GCC GTT TGG GCC CAA GTT CGG CAT AGT GTG GTG ATA GTA AGA GCA ATC-3'	N/A	N/A	(Rozi et al., 2021) [53]
PEDOT-GO/Au@Pt/Ap t/MCH/GCE	5-SH-(CH ₂) ₆ -GCT-TCC-AGC-TTA-TTG-AAT-TAC-ACG-CAG-AGG-GTA-GCG-GCT-CTG-CGC-ATT-CAA-TTG-CTG-CGC-GCT-GAA-GCG-CGG-AAG-C-3'	Bisphenol A, estrone, estriol, and ethinylestradiol	Serum	(Zhao et al., 2022) [76]

References

13. Rather, J.A.; Khudaish, E.A.; Kannan, P. Graphene-Amplified Femtosensitive Aptasensing of Estradiol, an Endocrine Disruptor. *Analyst* **2018**, *143*, 1835–1845. <https://doi.org/10.1039/C7AN02092A>.
35. Kim, Y.S.; Jung, H.S.; Matsuura, T.; Lee, H.Y.; Kawai, T.; Gu, M.B. Electrochemical Detection of 17 β -Estradiol Using DNA Aptamer Immobilized Gold Electrode Chip. *Biosens. Bioelectron.* **2007**, *22*, 2525–2531. <https://doi.org/10.1016/j.bios.2006.10.004>.
37. Ke, H.; Liu, M.; Zhuang, L.; Li, Z.; Fan, L.; Zhao, G. A Femtomolar Level 17 β -Estradiol Electrochemical Aptasensor Constructed On Hierachical Dendritic Gold Modified Boron-Doped Diamond Electrode. *Electrochim. Acta* **2014**, *137*, 146–153. <https://doi.org/10.1016/j.electacta.2014.06.014>.
38. Lin, Z.; Chen, L.; Zhang, G.; Liu, Q.; Qiu, B.; Cai, Z.; Chen, G. Label-Free Aptamer-Based Electrochemical Impedance Biosensor for 17 β -Estradiol. *Analyst* **2012**, *137*, 819–822. <https://doi.org/10.1039/C1AN15856B>.
39. Zhu, B.; Alsager, O.A.; Kumar, S.; Hodgkiss, J.M.; Travas-Sejdic, J. Label-Free Electrochemical Aptasensor for Femtomolar Detection of 17 β -Estradiol. *Biosens. Bioelectron.* **2015**, *70*, 398–403. <https://doi.org/10.1016/j.bios.2015.03.050>.
40. Mat Zaid, M.H.; Abdullah, J.; Rozi, N.; Mohamad Rozlan, A.A.; Abu Hanifah, S. A Sensitive Impedimetric Aptasensor Based on Carbon Nanodots Modified Electrode for Detection of 17 β -Estradiol. *Nanomaterials* **2020**, *10*, 1346. <https://doi.org/10.3390/nano10071346>.
42. Huang, K.-J.; Liu, Y.-J.; Zhang, J.-Z.; Cao, J.-T.; Liu, Y.-M. Aptamer/Au Nanoparticles/Cobalt Sulfide Nanosheets Biosensor for 17 β -Estradiol Detection Using a Guanine-Rich Complementary DNA Sequence for Signal Amplification. *Biosens. Bioelectron.* **2015**, *67*, 184–191. <https://doi.org/10.1016/j.bios.2014.08.010>.
43. Chang, Z.; Zhu, B.; Liu, J.; Zhu, X.; Xu, M.; Travas-Sejdic, J. Electrochemical Aptasensor for 17 β -Estradiol Using Disposable Laser Scribed Graphene Electrodes. *Biosens. Bioelectron.* **2021**, *185*, 113247. <https://doi.org/10.1016/j.bios.2021.113247>.
47. Olowu, R.A.; Arotiba, O.; Mailu, S.N.; Waryo, T.T.; Baker, P.; Iwuoha, E. Electrochemical Aptasensor for Endocrine Disrupting 17 β -Estradiol Based on a Poly(3,4-Ethylenedioxylthiopene)-Gold Nanocomposite Platform. *Sensors* **2010**, *10*, 9872–9890. <https://doi.org/10.3390/s101109872>.

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48. Fan, L.; Zhao, G.; Shi, H.; Liu, M. A Simple and Label-Free Aptasensor Based on Nickel Hexacyanoferrate Nanoparticles as Signal Probe for Highly Sensitive Detection of 17 β -Estradiol. *Biosens. Bioelectron.* **2015**, *68*, 303–309. <https://doi.org/10.1016/j.bios.2015.01.015>.
49. Nameghi, M.A.; Danesh, N.M.; Ramezani, M.; Aliboland, M.; Abnous, K.; Taghdisi, S.M. An Ultrasensitive Electrochemical Sensor for 17 β -Estradiol Using Split Aptamers. *Anal. Chim. Acta* **2019**, *1065*, 107–112. <https://doi.org/10.1016/j.aca.2019.02.062>.
50. Huang, K.-J.; Liu, Y.-J.; Shi, G.-W.; Yang, X.-R.; Liu, Y.-M. Label-Free Aptamer Sensor for 17 β -Estradiol Based on Vanadium Disulfide Nanoflowers and Au Nanoparticles. *Sens. Actuators B Chem.* **2014**, *201*, 579–585. <https://doi.org/10.1016/j.snb.2014.05.055>.
51. Huang, K.-J.; Liu, Y.-J.; Zhang, J.-Z.; Liu, Y.-M. A Novel Aptamer Sensor Based on Layered Tungsten Disulfide Nanosheets and Au Nanoparticles Amplification for 17 β -Estradiol Detection. *Anal. Methods* **2014**, *6*, 8011–8017. <https://doi.org/10.1039/C4AY01478B>.
52. Huang, K.-J.; Liu, Y.-J.; Zhang, J.-Z. Aptamer-Based Electrochemical Assay of 17 β -Estradiol Using a Glassy Carbon Electrode Modified with Copper Sulfide Nanosheets and Gold Nanoparticles, and Applying Enzyme-Based Signal Amplification. *Microchim. Acta* **2015**, *182*, 409–417. <https://doi.org/10.1007/s00604-014-1352-0>.
53. Rozi, N.; Hanifah, S.A.; Zaid, M.H.M.; Abd Karim, N.H.; Ikeda, M. Feasible Study on Poly(Pyrrole-Co-Pyrrole-3-Carboxylic Acid)-Modified Electrode for Detection of 17 β -Estradiol. *Chem. Pap.* **2021**, *75*, 3493–3503. <https://doi.org/10.1007/s11696-021-01597-9>.
54. Ming, T.; Wang, Y.; Luo, J.; Liu, J.; Sun, S.; Xing, Y.; Xiao, G.; Jin, H.; Cai, X. Folding Paper-Based Aptasensor Platform Coated with Novel Nanoassemblies for Instant and Highly Sensitive Detection of 17 β -Estradiol. *ACS Sens.* **2019**, *4*, 3186–3194. <https://doi.org/10.1021/acssensors.9b01633>.
55. Liu, X.; Deng, K.; Wang, H.; Li, C.; Zhang, S.; Huang, H. Aptamer Based Ratiometric Electrochemical Sensing of 17 β -Estradiol Using an Electrode Modified with Gold Nanoparticles, Thionine, and Multiwalled Carbon Nanotubes. *Microchim. Acta* **2019**, *186*, 347. <https://doi.org/10.1007/s00604-019-3465-y>.
76. Zhao, Z.; Chen, H.; Cheng, Y.; Huang, Z.; Wei, X.; Feng, J.; Cheng, J.; Mugo, S.M.; Jaffrezic-Renault, N.; Guo, Z. Electrochemical Aptasensor Based on Electrodeposited Poly(3,4-Ethylenedioxythiophene)-Graphene Oxide Coupled with Au@Pt Nanocrystals for the Detection of 17 β -Estradiol. *Microchim. Acta* **2022**, *189*, 178. <https://doi.org/10.1007/s00604-022-05274-w>.