



Review

Aggregation-Induced Emission Luminogens for Enhanced Photodynamic Therapy: From Organelle Targeting to Tumor Targeting

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Abstract: Photodynamic therapy (PDT) has attracted much attention in the field of anticancer treatment. However, PDT has to face challenges, such as aggregation caused by quenching of reactive oxygen species (ROS), and short ¹O₂ lifetime, which lead to unsatisfactory therapeutic effect. Aggregation-induced emission luminogen (AIEgens)-based photosensitizers (PSs) showed enhanced ROS generation upon aggregation, which showed great potential for hypoxic tumor treatment with enhanced PDT effect. In this review, we summarized the design strategies and applications of AIEgen-based PSs with improved PDT efficacy since 2019. Firstly, we introduce the research background and some basic knowledge in the related field. Secondly, the recent approaches of AIEgen-based PSs for enhanced PDT are summarized in two categories: (1) organelle-targeting PSs that could cause direct damage to organelles to enhance PDT effects, and (2) PSs with tumor-targeting abilities to selectively suppress tumor growth and reduce side effects. Finally, current challenges and future opportunities are discussed. We hope this review can offer new insights and inspirations for the development of AIEgen-based PSs for better PDT effect.

Keywords: organelle targeting; photodynamic therapy; aggregation-induced emission; tumor targeting



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1. Introduction

Cancer, as one of the most fatal diseases, caused nearly 10 million deaths in 2020, as reported by the World Health Organization International Agency for Research on Cancer (IARC) [1]. According to the Chinese National Cancer Center, the survival rate has increased by 10% to 40.5% compared to 10 years ago. This surprising result could be attributed to the emergence of various therapies to overcome cancer [2]. At present, cancer treatment methods mainly include surgery, radiotherapy, chemotherapy, immunotherapy, etc. [3–5]. However, these therapies still come with limitations, such as nausea and vomiting, which are common side effects of chemotherapy and radiation [6], and the effectiveness of immunotherapy for specific tumors [7]. In recent years, people have gradually turned their attention to the field of phototherapy [4,8–14], which shows various attractive features such as non-invasiveness, low toxicity and good biocompatibility [11,15].

From the perspective of photodynamic therapy (PDT) [16], photosensitizers (PSs) produce ROS to kill tumor cells under light irradiation. PSs generate ROS through two processes [8,11,17–20]: (a) Type I: through proton or electron transfer [21], direct reaction with substrate or solvent molecules to form O₂^{•−} and •OH [22]; (b) Type II: the triplet PSs transfer their energy to triplet oxygen molecules to form ¹O₂ [10]. Nevertheless, PDT suffers from problems such as aggregation in aqueous solutions, which leads to both fluorescence quenching (ACQ effect) [23,24] and ROS quenching. In addition, the short lifetime of ¹O₂

shortens its effective radius, which might limit the Type II PDT efficacy [25]. The PSs with Type I ROS production capacity will be described in this paper, and the remaining examples without additional description are those of Type II ROS or ROS species not described in the original literature. Aggregation-induced emission (AIE) was proposed to inhibit PSs aggregation and promote ROS production. On the other hand, organelle-targeting strategies could generate ROS in situ and cause severe damage to organelles, which could help to solve the problem of short $^1\text{O}_2$ lifetime [26–28]. In addition, tumor-targeting strategies can effectively improve the specificity of PDT and reduce side effects, which can help improve PDT performance [28–31].

In this review, we summarized current approaches to enhance photodynamic therapy based on AIE from two different perspectives (Figure 1). First, the design strategies and antitumor applications of AIEgen-based PSs that are able to target cell plasma membranes, mitochondria, lysosomes, lipid droplets, nuclei, the endoplasmic reticulum, and the Golgi apparatus were introduced. Second, different approaches for tumor-targeting AIEgen-based PSs were covered. This review does not aim to be comprehensive; readers are also encouraged to refer to other excellent reviews for more information [32–37]. Due to the fast advances in this field, we only summarized some representative works during the past three years. We hope that this review can provide some ideas for related research and promote the progress of PDT therapy.

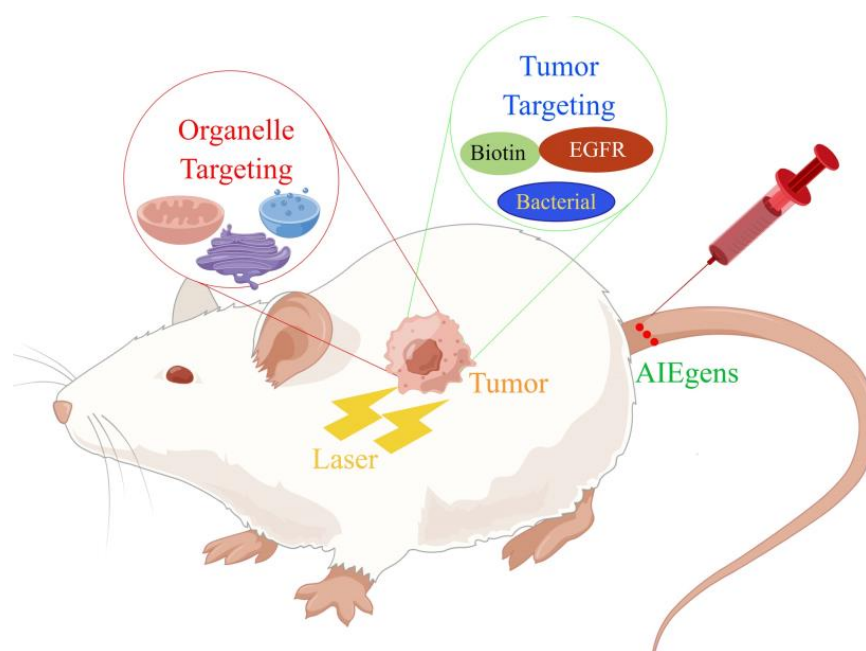


Figure 1. Different methods of treating tumor hypoxia.

2. Organelle-Targeting AIE-PDT

PSs should be designed to be intracellularly targetable due to high ROS reactivity and short diffusion distance, which may enhance PDT efficacy significantly. Currently, many AIEgen-based PSs that showed subcellular organelle-targeting abilities have been reported. Most of them were constructed to target mitochondria, lysosomes, lipid droplets, and plasma membranes because the targeting strategies were well-established [28]. AIEgen-based PSs targeting other organelles, such as the endoplasmic reticulum (ER) and Golgi apparatus (GA), were less reported.

2.1. Membrane Targeting

In addition to secreting and transporting proteins, the cell membrane also absorbs and excretes substances inside and outside through endocytosis and exocytosis. If the cell membrane is damaged, this results in the increased permeability of tumor cells. It

may promote the uptake of deleterious material [38–41]. There are two main strategies for targeting cell membranes [28]. One is to build a positive charge structure based on a similar method of targeting mitochondria, and the introduction of amphipathic structures is able to obtain stronger cell-membrane-targeting capabilities [42]. The other strategy is using membrane-specific ligand modification [43]. The antigen-modified PS, for example, can specifically recognize receptors abundantly expressed on tumor membranes.

Recently, Tang et al. designed and synthesized AIE-PSs with NIR-emission (735 nm) that could target cell membranes [38]. They infused the cationation structure into TBMPEI, triggering the cell-membrane-targeting characterization (Figure 2). The fluorescence signals of DCFH incubated with TBMPEI significantly increased nearly 900 times in 80 s under white light, indicating remarkable ROS generation ability. The species of ROS produced by TBMPEI were shown to be a mixture of Type I and Type II. After being stained with TBMPEI, multiple cells showed high Pearson's correlation coefficients and high signal-to-noise ratio. They also confirmed that TBMPEI had good cytotoxicity through the IC₅₀ values of 4T1, A549 and Hela cells under light excitation. The researchers found that the integrity of the cell membrane was destroyed, and DNA degradation even occurred. TBMPEI was shown to induce necroptosis of tumor cells by targeting cell membranes with the Annexin V-FITC test and tumor section. The tumor in vivo had been inhibited significantly in the experiment group treated with TBMPEI.

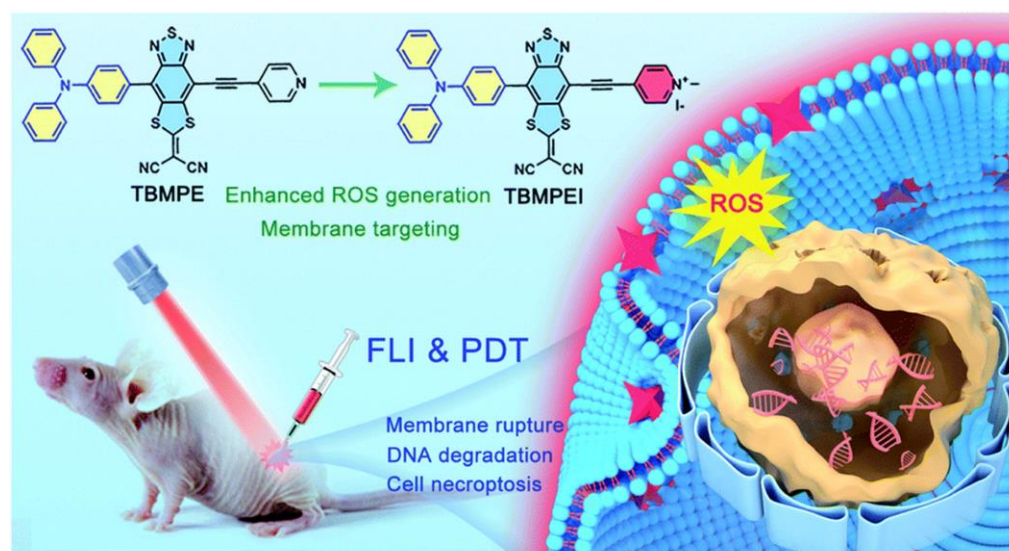


Figure 2. Schematic illustration of TBMPEI with cell-membrane-targeting function. Reprinted with permission from Ref. [41]. Copyright 2022 Royal Society of Chemistry.

Recently, Liu et al. developed a cancer immunotherapy that causes pyroptosis through a membrane-targeted photosensitizer TBD-3C with AIE characteristics (Figure 3) [40]. In addition to stimulating macrophage M1 polarization, it can cause maturation of dendritic cells (DCs) and activate CD8 + cytotoxic T lymphocytes (CTLs) (Figure 3). The result of flow cytometry showed that TBD-3C is capable of successful membrane anchoring. Flow cytometry and DCFH-DA upon light irradiation (40 mW cm^{-2} for 10 min) were further used to confirm the ROS generation ability of TBD-3C in pancreatic cancer. After irradiation, TBD-3C induced swelling on KPC and panc02 cells, showing the typical morphological features of pyroptosis. KPC and Panc02 cells treated with TBD-3C release lactate dehydrogenase (LDH) as the typical signal of pyroptosis. This study developed a pyroptosis-based photodynamic anti-tumor immunotherapy approach.

Yang et al. reported an amphiphilic perylene derivative AIE-PSs named AP ($\phi_{\Delta} = 0.22$, methylene blue as reference) which is capable of cell-membrane-targeting [44]. AP could form nanoparticles by self-assembly in an aqueous solution and decompose into free monomeric molecules after membrane anchoring (Figure 4). CLSM images of MCF-7 cells

stained with AP were clearly eliminated. H&E staining and TUNEL images indicates that apoptosis occurred in many tumor cells. In addition, in vivo biocompatibility tests revealed that AP was almost nontoxic.

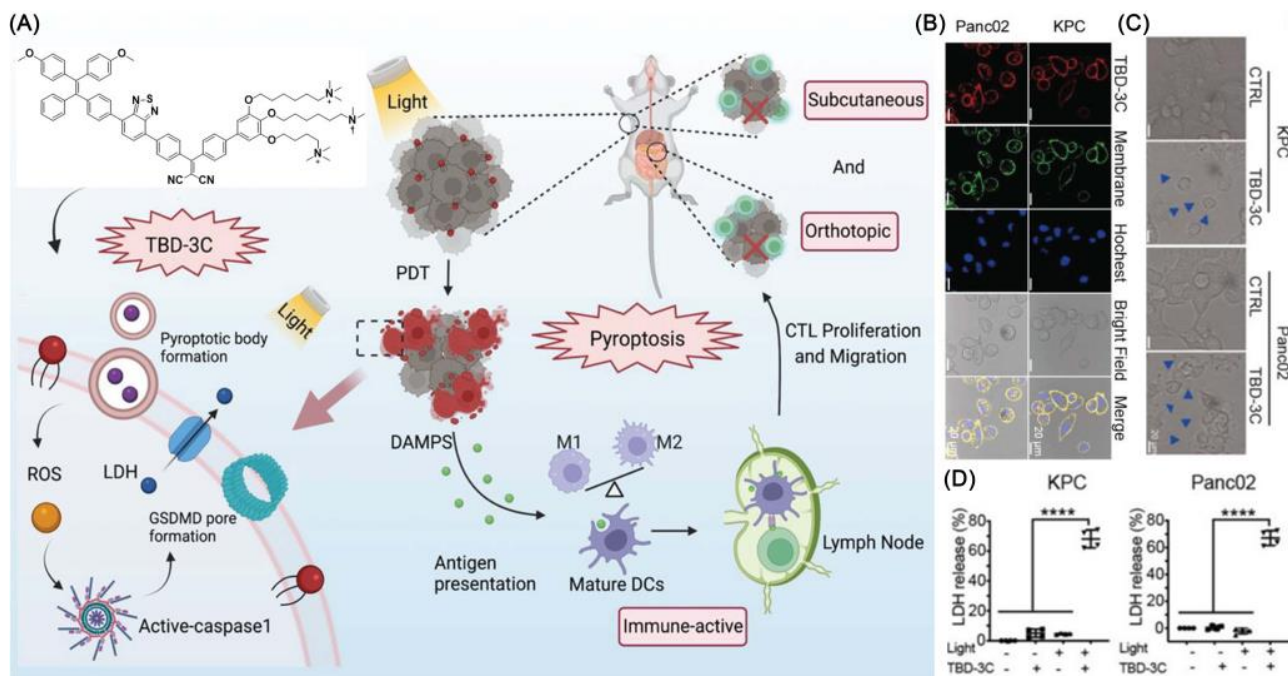


Figure 3. (A) Photodynamic pyroptosis for antitumor immunotherapy; (B) images of cells incubated with TBD-3C captured with CLSM; (C) KPC and Panc02 cells were analyzed by confocal microscopy after being exposed to 40 mW cm^{-2} for 10 min; bright blue arrows indicate membrane expansion; (D) Generated LDH from KPC and Panc02 cells irradiated with light at 40 W cm^{-2} for 10 min. (**** $p < 0.0001$). Reprinted with permission from Ref. [40]. Copyright 2022 WILEY-VCH.

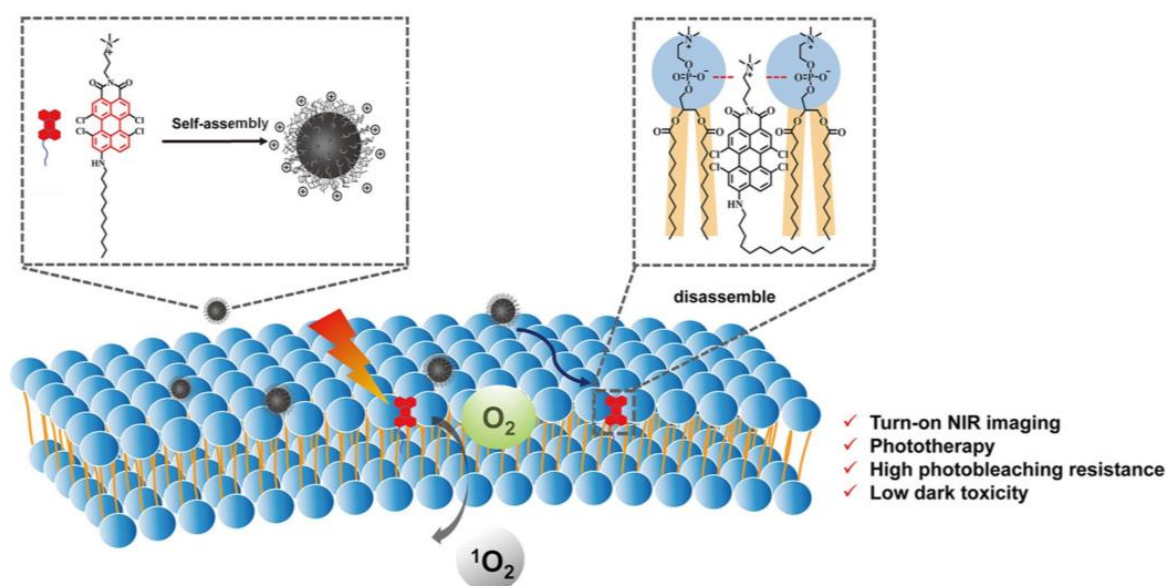


Figure 4. Schematic illustration of AP-based turn-on NIR fluorescence imaging and PDT. Reprinted with permission from Ref. [44]. Copyright 2021 American Chemical Society.

2.2. Lysosomal Targeting

As the main digestive site in cells, lysosomes degrade intruding toxic substances using more than 60 hydrolytic enzymes, making them crucial in autophagy and secre-

tion [28,34,45]. Lysosome-targeted strategies have also become instructive and meaningful for PDT therapy. There are two main approaches: (1) Most examples target lysosomes through the modification of lipophilic amines with the addition of morpholines and other amine groups [46–60]. (2) A promotion of endocytosis can also transport PSs from the endosome into the cell and capture them in the lysosome [61–70].

According to Niu et al., a BTZPP molecule with A-D-A structure was synthesized and the PDT effect was investigated (Figure 5) [47]. It had been demonstrated in experiments that BTZPP NPs are capable of high $^1\text{O}_2$ quantum yield (72.3%, rose bengal as a reference), NIR emission (635 nm) and good photostability under harsh conditions such as acidity. Moreover, BTZPP with LysoTracker Green had a good Pearson coefficient (0.91). Apoptosis was observed in Hela cells treated with BTZPP under light conditions. Based on MTT assay, BTZPP NPs showed IC_{50} value. In addition, BTZPP NPs exhibit the characteristic of long-term *in vivo* imaging, which is helpful for diagnosis. Meanwhile, the *in vivo* experiment indicated BTZPP had no obvious systemic toxicity.

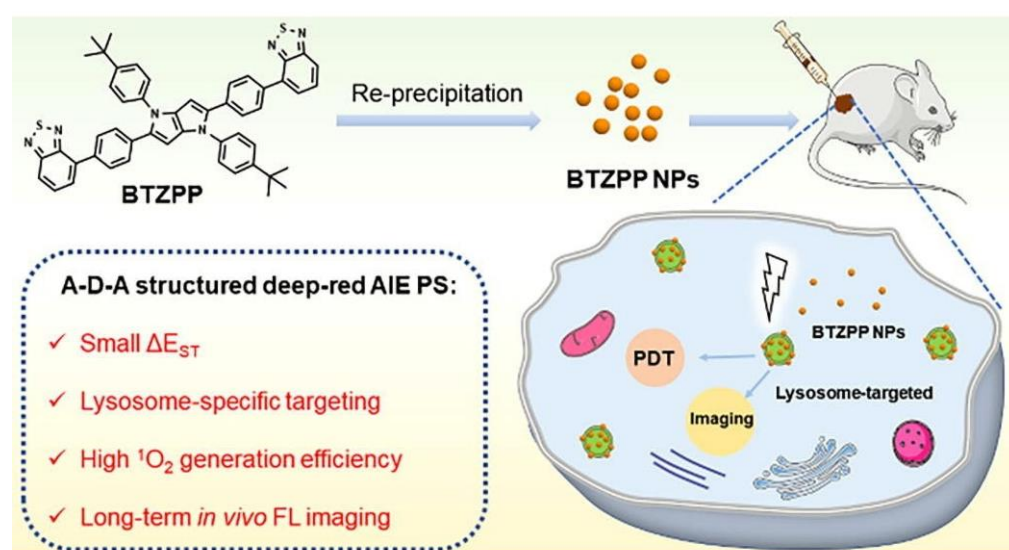


Figure 5. Schematic illustration of AIE-PSs (BTZPP) designed for long-term imaging. Reprinted with permission from Ref. [47]. Copyright 2022 Elsevier Ltd.

Tang et al. synthesized a series of molecules with varying amounts of thiophene spacers [70]. As the first single molecules to deliver all phototheranostics, TSSIs can effectively deliver fluorescence and photoacoustic imaging, such as modalities including PDT, photothermal imaging (PTI), photothermal therapy (PTT) and so on (Figure 6). The TSSI NPs simultaneously exhibit NIR-II emission (1000 nm), high ROS generation (Type I), and good photothermal conversion efficiency (46%). TSSIs have excellent ROS generation ability, with the emission intensity of DCFH increased over 250-fold after irradiation. The generation of Type I ROS made the PDT effect more effective under hypoxic conditions. TSSI showed good lysosomal targeting ability, and the Pearson coefficient was up to 0.964. Additionally, the authors demonstrated that TSSI NPs are ingested by cells using energy-dependent endocytosis. H&E staining of tumor slices revealed that tumor cell apoptosis occurred under NIR irradiation. Moreover, compared with the control group, tumor growth was significantly inhibited until extinct with TSSI treatment. Meanwhile, no significant systemic toxicity was discovered.

Similarly, the authors reported another example, TTT-4, with better photoacoustic-guided imaging ability in 2021 (Figure 7) [59] and a better therapeutic effect on tumor tissue with a powerful lysosomal targeting ability. TTT-4 also generates Type I ROS with high generation ability. The fluorescence of DCFH increased 160-fold under white light ($22.1 \text{ mW}/\text{cm}^2$). MTT assay indicated that the IC_{50} value of 4T1 cells incubated with TTT-4 was less than $1 \mu\text{mol}$, which should be attributed to the excellent effects of PDT and PTT.

Furthermore, H&E staining of tumor slices revealed the tumor tissue exhibited lots of apoptotic cells.

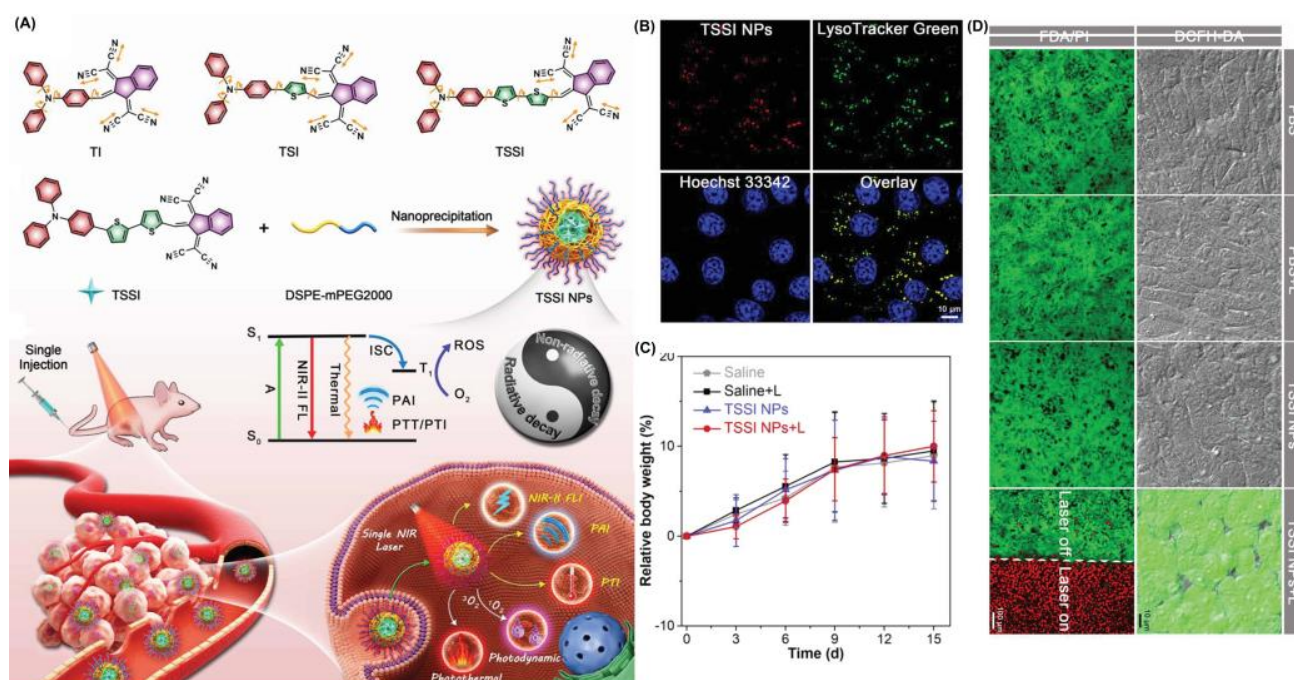


Figure 6. (A) Schematic illustration of mechanism and phototheranostic function of TSSI; (B) co-localization of 4T1 cells after incubation with TSSI NPs; (C) body weight changes of mice in vivo biosafety evaluation; (D) live/dead cell staining and intracellular ROS of 4T1 cells. Reprinted with permission from Ref. [70]. Copyright 2020 WILEY-VCH.

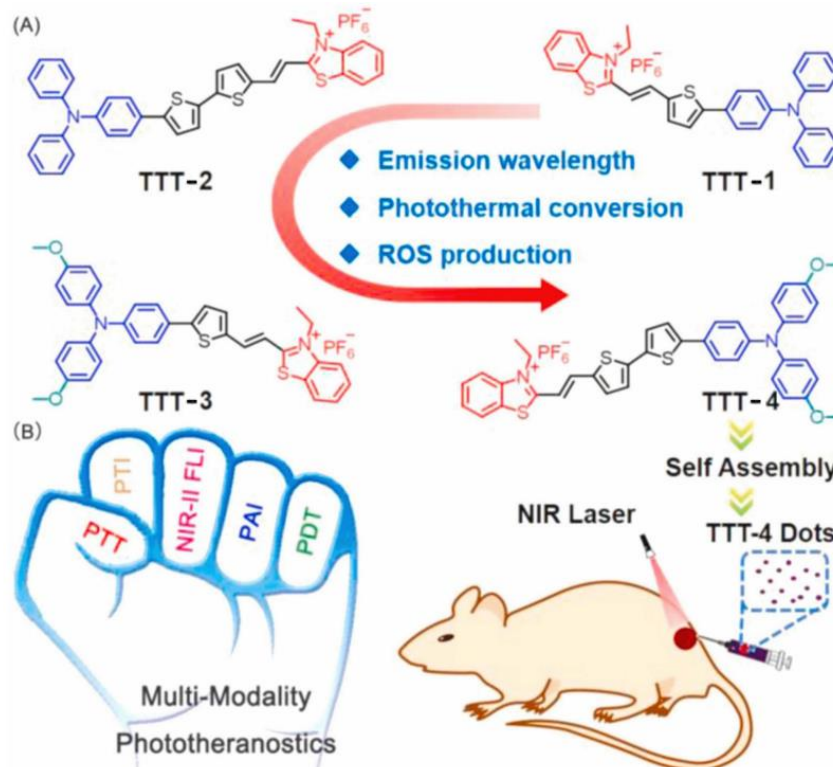


Figure 7. (A) Molecular structures and (B) multifunctional phototheranostic application of AIEgens. Reprinted with permission from Ref. [59]. Copyright 2021 Elsevier Ltd.

2.3. Mitochondrion Targeting

The mitochondria play vital roles in energy production and intracellular signal transmission in cells [71]. The overexpression of ROS in mitochondria may cause oxidative stress, which may disrupt the mitochondrial microenvironment and lead to apoptosis, autophagy and necroptosis [72–74]. Consequently, the mitochondrion is a suitable organelle for PDT. In order to realize mitochondrial localization, several strategies have been explored, including: (1) introducing lipophilic cations structures that are intrinsically or modified [75–109]; and (2) using mitochondria-specific peptides [110–113].

Zheng et al. developed a series of positively charged AIEgens (Figure 8) [71]. DCQu is capable of specific mitochondrial targeting with high $^1\text{O}_2$ generation efficiency and NIR emission. DCPy and DCQu had superior AIE properties compared to other compounds. Following co-incubation of DCQu and H₂DCF-DA with HeLa cells, fluorescence imaging experiments revealed that DCQu generates $^1\text{O}_2$ efficiently (2.1-fold higher than DCPy) during irradiation. DCQu exhibits high Pearson's correlation coefficients of 0.95, indicating superior specificity for mitochondrial staining. Furthermore, MTT assay showed that DCQu had a good therapeutic effect on tumor tissues. Using hematoxylin and eosin (H&E) analysis, it was evident that many cells with highly condensed nuclei were apoptotic. Compared with the control group, the survival rate of mice treated with DCQu increased significantly.

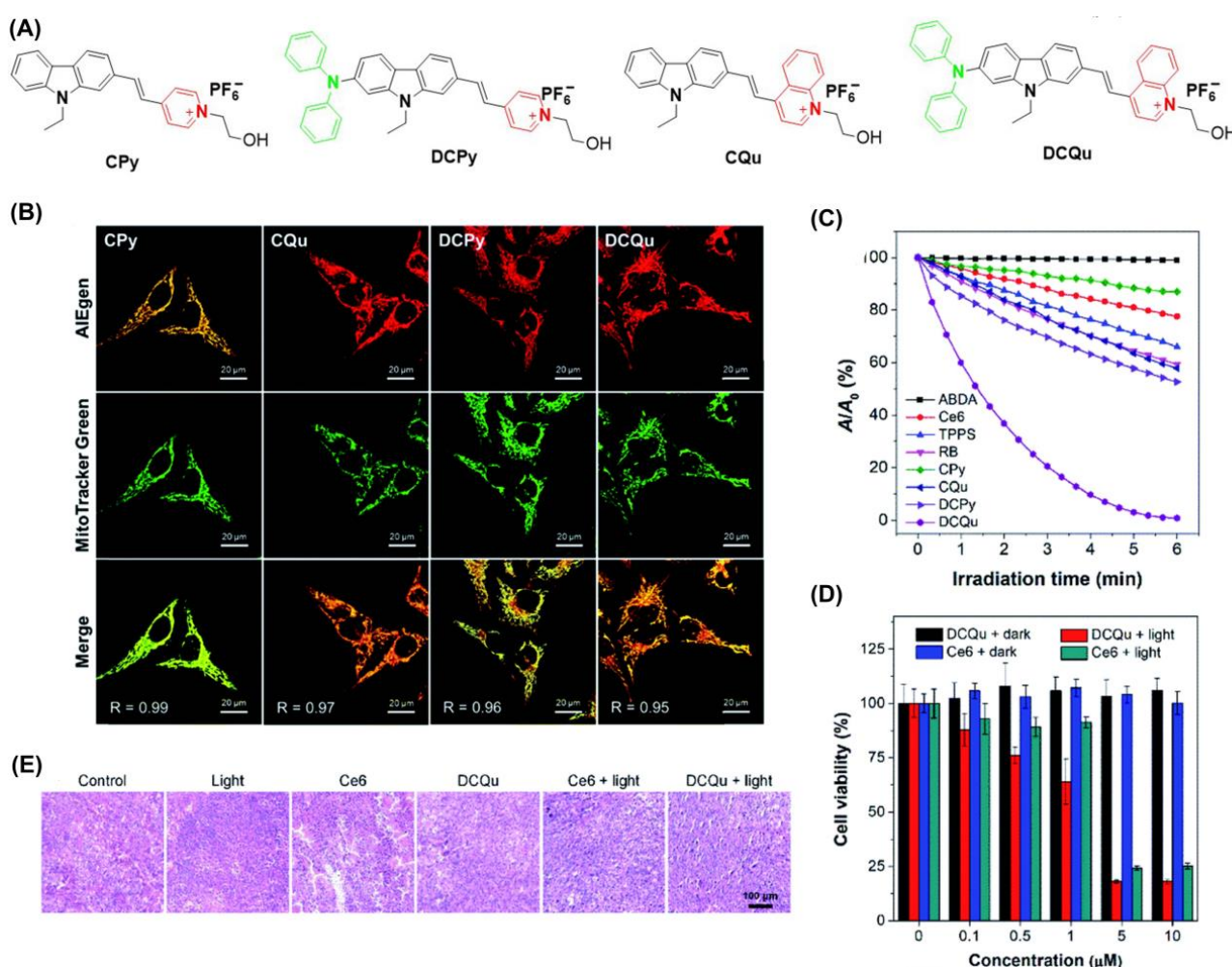


Figure 8. (A) Structure of CPy, CQu, DCPy and DCQu; (B) CLSM images of HeLa cells stained with CPy, CQu, DCPy and DCQu and various probes; (C) detection of $^1\text{O}_2$ through ABDA; (D) cell viability of HeLa cancer cells stained with DCQu; (E) H&E staining analysis of tumor tissues treated with DCQu. Reprinted with permission from Ref. [75]. Copyright 2020 Royal Society of Chemistry.

Tang et al. drew on a cationization strategy to synthesize DTPAN, DTPAPy, DTPANPF₆ and DTPAPyPF₆ (Figure 9) [78]. Using injected cationization with a strong ICT effect, DTPANPF₆ and DTPAPyPF₆ are endowed with high Type I radical production capacity and the ability to target mitochondria. The HPF intensity enhancement of DTPANPF₆ and DTPAPyPF₆ was 37.4- and 30.0-fold under irradiation (20 mW cm⁻²). Both DTPANPF₆ and DTPAPyPF₆ exhibited high Pearson coefficients for both HeLa and MCF-7 cells. The viability of HeLa cells suggests they both have therapeutic potential under hypoxic conditions. H&E staining suggested prominent cell necroptosis occurred in tumor tissues. In *in vivo* PDT experiments, tumor growth was significantly inhibited when treated with DTPANPF₆ and DTPAPyPF₆.

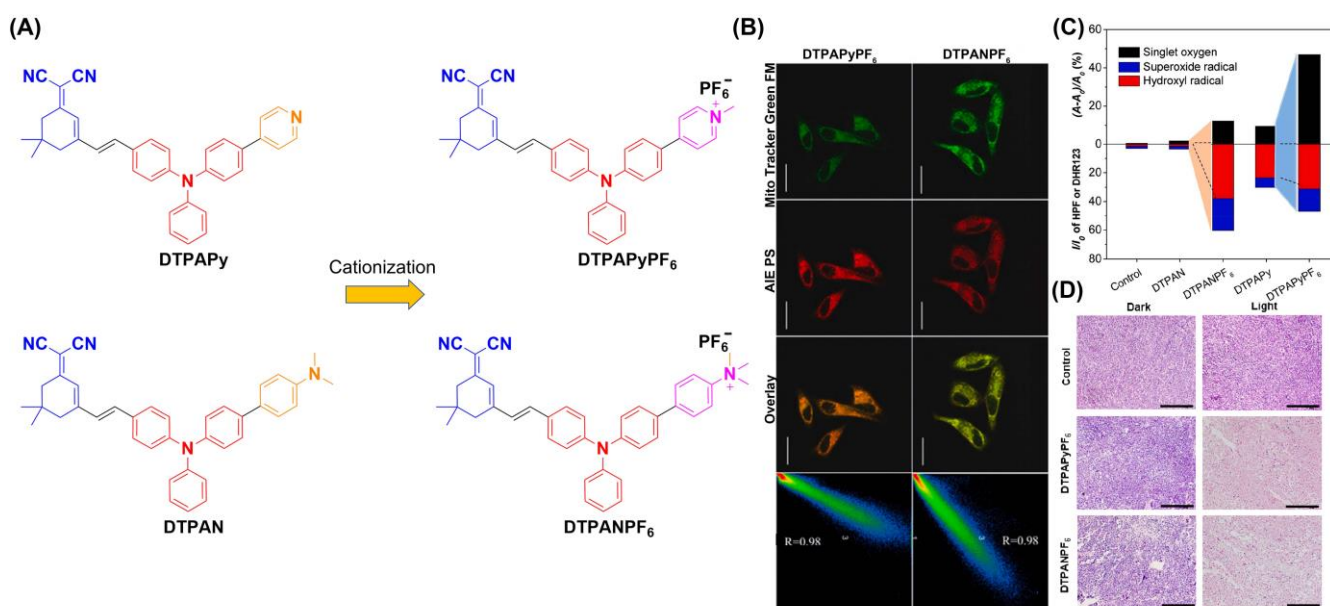


Figure 9. (A) Structure of DTPAN, DTPAPy, DTPANPF₆ and DTPAPyPF₆; (B) co-localization of HeLa cells incubated with DTPANPF₆ or DTPAPyPF₆; (C) summary of different ROS generation of DTPAN, DTPAPy, DTPANPF₆ and DTPAPyPF₆; (D) H&E staining of tumor tissues. Reprinted with permission from Ref [78]. Copyright 2022 Elsevier Ltd.

Tang et al. reported another study on PDT treatment of mitochondrial targeting to initiate autophagy (Figure 10) [90]. TACQ exhibits near-infrared emission (635 nm), high photothermal conversion efficiency (55%), and high ¹O₂ generation capacity. The reduction of absorbance for ABDA treated with TACP at 378 nm after irradiation for 60 s reached 90.3%. The quinoline cation of TACQ selectively accumulated in the mitochondria. The Pearson coefficient of TACQ with Mito-Tracker Green (MTG) was as high as 0.95. In comparison with MTG, TACQ provided better 3D mitochondrial images with higher lateral resolution. TEM characterization of HeLa cells indicates that autophagosomes are formed inducing mitophagy after TACQ treatment. The authors suggest that TACQ accelerates mitochondrial instability and leads to apoptosis in cancer cells.

2.4. Lipid Droplet Targeting

Lipid droplets are lipid-rich organelles found mainly in adipose tissue. They are highly dynamic organelles involved in intracellular lipid storage, metabolism, and membrane transfer. Additionally, LDs are being pursued as a target of PDT [114–120]. The LD-targeted photosensitizer should have a highly hydrophobic structure and be less polar than the other fractions in the cell [121–123].

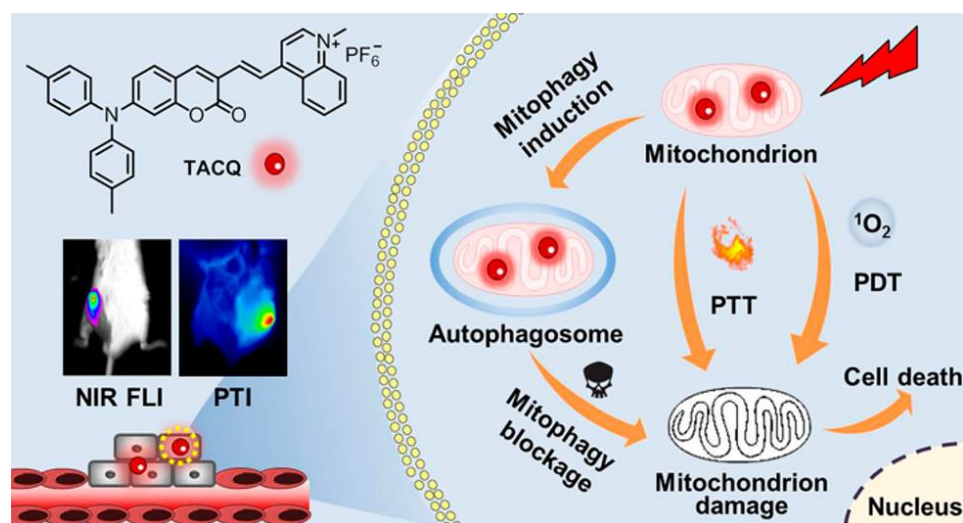


Figure 10. Schematic illustration of chemical structures and multifunctional phototheranostic of TACQ. Reprinted with permission from Ref. [90]. Copyright 2021 American Chemical Society.

Dai et al. designed and synthesized an AIE-PSS (TTI) with strong lipophilic and near-infrared emission (NIR) (Figure 11) [119]. The $^1\text{O}_2$ quantum yield of TTI, was determined to be 85.16% using an equation. In addition, the calculated $\text{Clog } p$ values of TTI and its derivatives ranged from 8.3 to 9.4, all within the range of 4.5 to 9.5 which could target LDs. The Pearson coefficient of TTI with BODIPY 493/503 was calculated to be 0.9491. Cell apoptosis was detected by Annexin V-FITC/PI co-staining. The results showed that TTI could effectively induce apoptosis of HepG2 cells under white light irradiation.

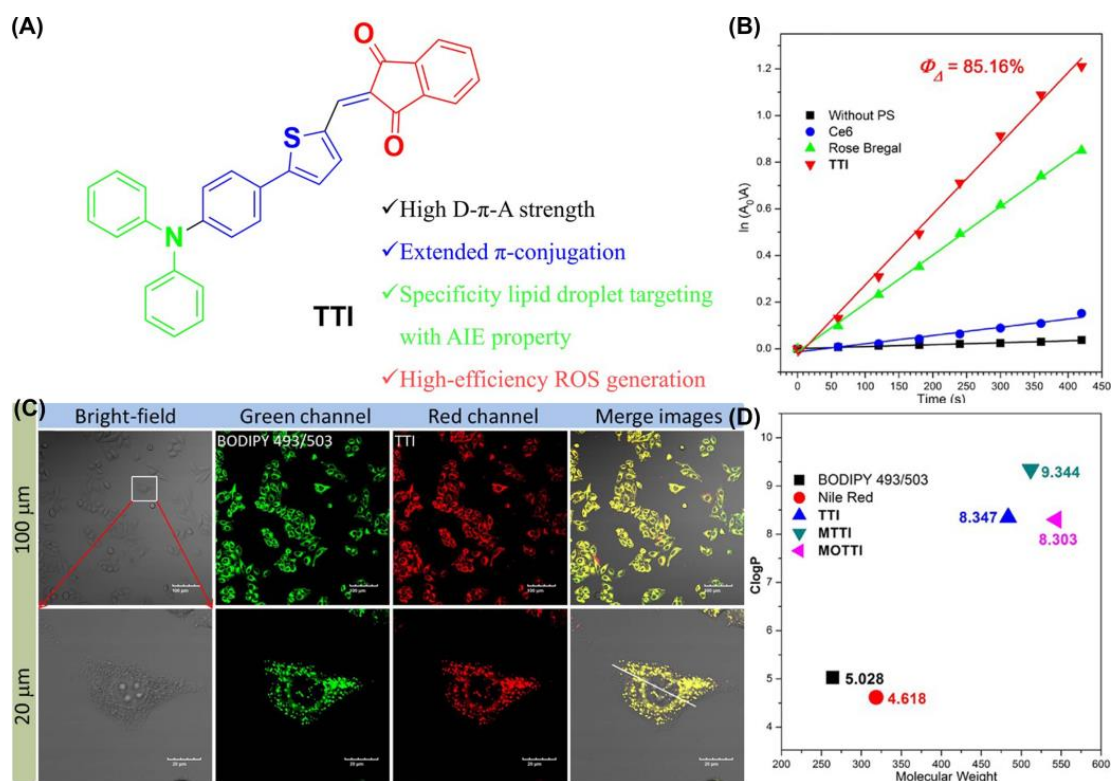


Figure 11. (A) Chemical structure and functions of TTI; (B) $^1\text{O}_2$ quantum yield with TTI and $\text{Clog } p$ values of TTI; (C) CLSM images of HepG2 cells stained with TTI and BODIPY 493/503; (D) $\text{Clog } p$ values of TTI and its derivatives. Reprinted with permission from Ref. [119]. Copyright 2021 Elsevier Ltd.

Tang et al. synthesized two AIE-PSs (PI and PTI) with near-infrared (NIR) emission properties and the ability to specifically target lipid droplets (Figure 12) [124]. PTI was obtained by introducing a thiophene ring into the PI skeleton to enhance the ISC process. The authors confirmed that targeting lipid droplets using PI and PTI caused ferroptosis by monitoring intracellular glutathione (GSH) and glutathione peroxidase 4 (GPX4) levels. Furthermore, it is important to note that the authors used homologous MCF-7 cell membranes to wrap the PLGA core, composed of PTI and PLGA, to achieve a homologous targeting ability. Besides, PTI has a distinguished ROS generation ability with the intensity of H2DCFH-DA in PBS, increasing 120-fold upon white light irradiation (50 mW cm^{-2}). The synthesized MCFCNPs have a good inhibitory effect on tumors in vivo without obvious toxic side effects.

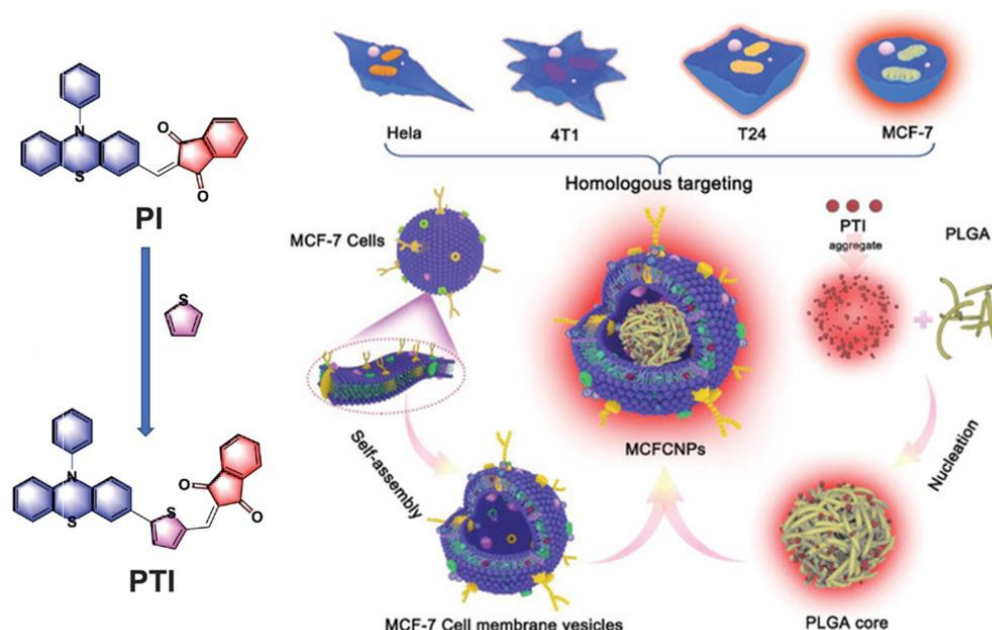


Figure 12. Schematic illustration of chemical structures and mechanisms of TACQ. Reprinted with permission from Ref. [124]. Copyright 2021 WILEY-VCH.

Similarly, Tang et al. reported another case in 2021 (Figure 13) [114]. They combined the targeted LDs AIE-PSs (MeTIND-4) with DC cell membranes to achieve antigen functioning as a biomimetic nano-photosensitizer (DC@AIEdots). While the exogenous cell membrane stimulates the proliferation and activation of T cells, internal AIE photosensitizers target tumor cells for PDT. The fluorescence intensity of DCFH treated with MeTIND-4 increased striking by 600-fold after irradiation (60 mW cm^{-2}). Therefore, PSs not only produces sufficient ROS to eliminate tumor cells, but also promotes immunogenic cell death. In addition, the efficiency of the tumor delivery of photosensitizers had been effectively improved (1.6 times). DC@AIEdots can not only kill in situ tumors, but also suppress distant tumors by activating the immune system against tumor growth. This work provides a significant guide for the development of related fields.

In another example reported by Liu et al., they constructed NIR-emitting PSs (TPET-IS, TPET-FU and TPEF-IS) with the function of targeting LDs [125]. Based on the theoretical calculations, the Log *p* values for TPET-IS, TPET-FU and TPEF-IS were 9.39, 7.89 and 8.03, respectively, which were higher than BODIPY 493/503 (a commercial LD marker), indicating a good LD-specific targeting. The Pearson's correlation coefficients were 0.94, 0.96, and 0.97. It is worth noting that the survival rate of HeLa cells at $50 \mu\text{M}$ concentration of the three compounds was more than 90% under dark conditions.

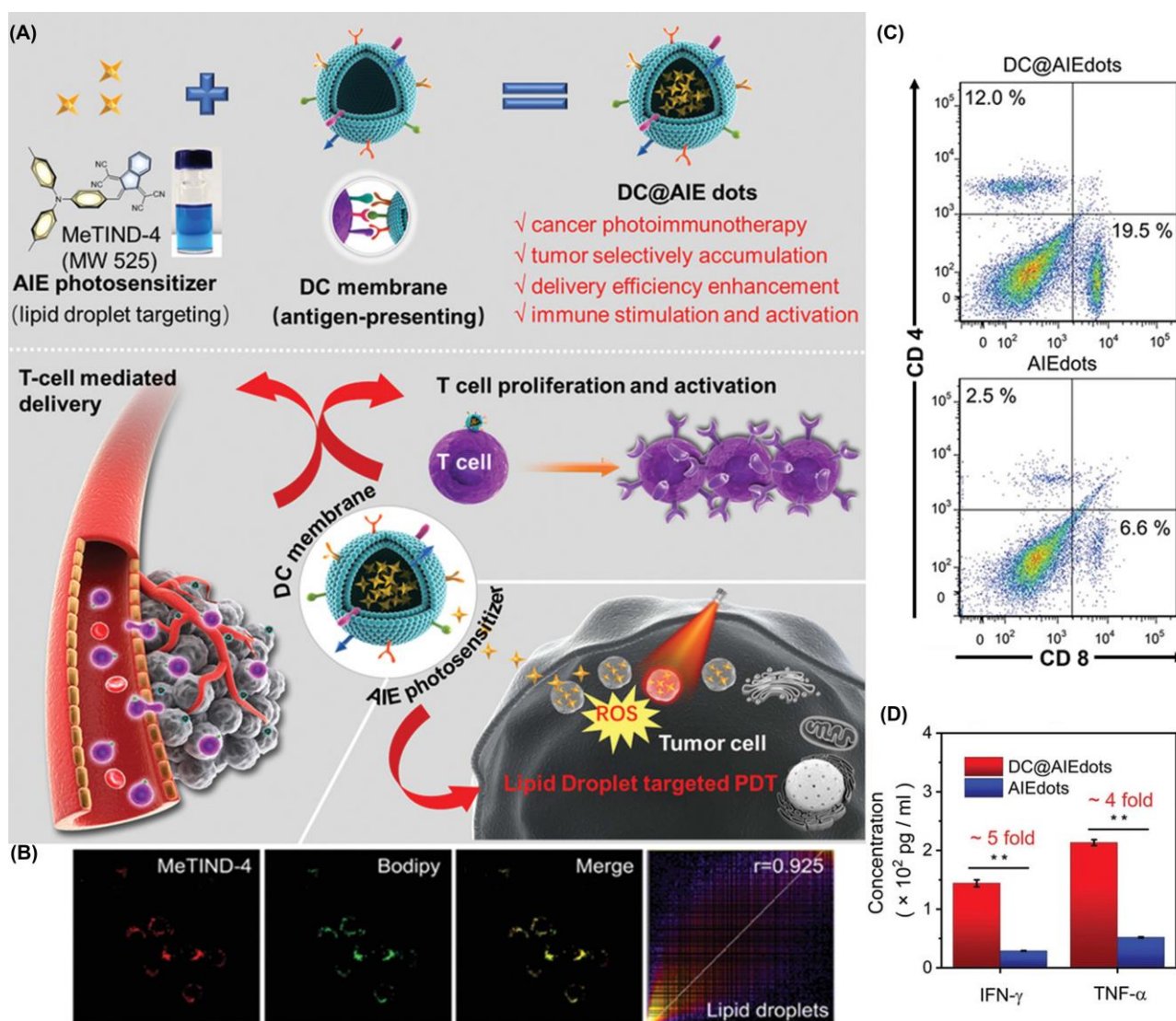


Figure 13. (A) Schematic illustration of chemical structures and in vivo photodynamic immunotherapy of DC@AIEdots; (B) colocalization experiments of 4T1 cells stained with MeTIND-4; (C) flow cytometry analysis of tumor infiltrating CD8⁺ and CD4⁺ T cells; (D) concentration of proinflammatory cytokines TNF- α and IFN- γ . * $p < 0.05$, ** $p < 0.01$. Reprinted with permission from Ref. [114]. Copyright 2021 WILEY-VCH.

2.5. Endoplasmic Reticulum Targeting

ROS-induced stress in the endoplasmic reticulum (ER) may lead to the activation of downstream immune pathways, resulting in immunogenic death of cells [28]. PSs modified by specific peptides or methyl sulfonamide usually have ER-targeting abilities, and some ring metal complexes can also target the ER [126].

Based on the reported AIE material TBP, Su et al. grew, in sulfonic acid, functional groups through a cation strategy to prepare TBP-SO₃ to obtain the ability to target the ER (Figure 14) [127]. TBP-SO₃ exhibited high Type I ROS generation capability, while the fluorescence spectra of DHR123 treated with TBP-SO₃ increased nearly 800-fold after irradiation (23.4 mW/cm²). In CLSM co-localization assay, the Pearson coefficient of HeLa cells incubated with ER-Tracker Red (targeting ER) and TBP-SO₃ was 0.93. It was found that the cell survival rate was not significantly decreased at 30 μ mol concentration without light. The IC₅₀ value of TBP-SO₃ under white light irradiation was less than 5 μ mol. The above experiments showed that TBP-SO₃ had a good application prospect in PDT.

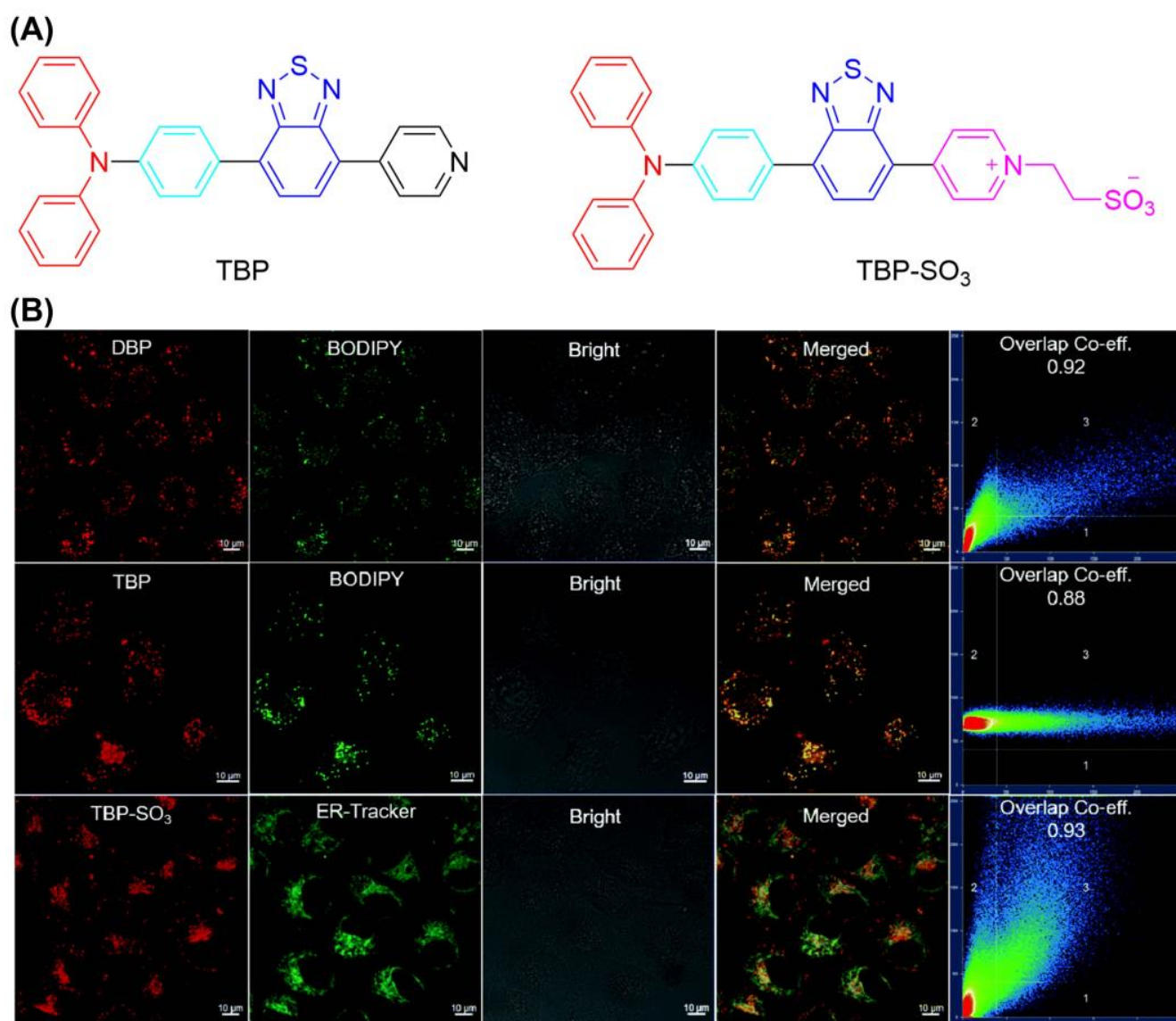


Figure 14. (A) Chemical structure of TBP and TBP-SO₃; (B) CLSM images of HeLa cells stained with TBP-SO₃. Reprinted with permission from Ref. [127]. Copyright 2022 WILEY-VCH.

Additionally, Tang et al. reported two ER-targeting Type I AIE-PSs (α -TPA-PIO and β -TPA-PIO) in 2020 (Figure 15) [128]. The results showed that β -TPA-PIO inhibited tumor cell growth under hypoxic conditions. Images from colocalization experiments showed good overlap between β -TPA-PIO and ER. The fluorescence signal of HPF containing α -TPA-PIO or β -TPA-PIO increased 6- and 11-fold after white light irradiation (20 mW cm⁻²). An in vitro study including co-localization, Western blot, and immunohistochemistry analyses found that PSC could lead to autophagy and apoptosis by inducing ER stress. Additionally, in vivo experiments indicated that β -TPA-PIO was effective in eliminate solid tumors. Researchers suggest that PIO induces immunogenic cell death, facilitating the combined effects of PDT and immunotherapy.

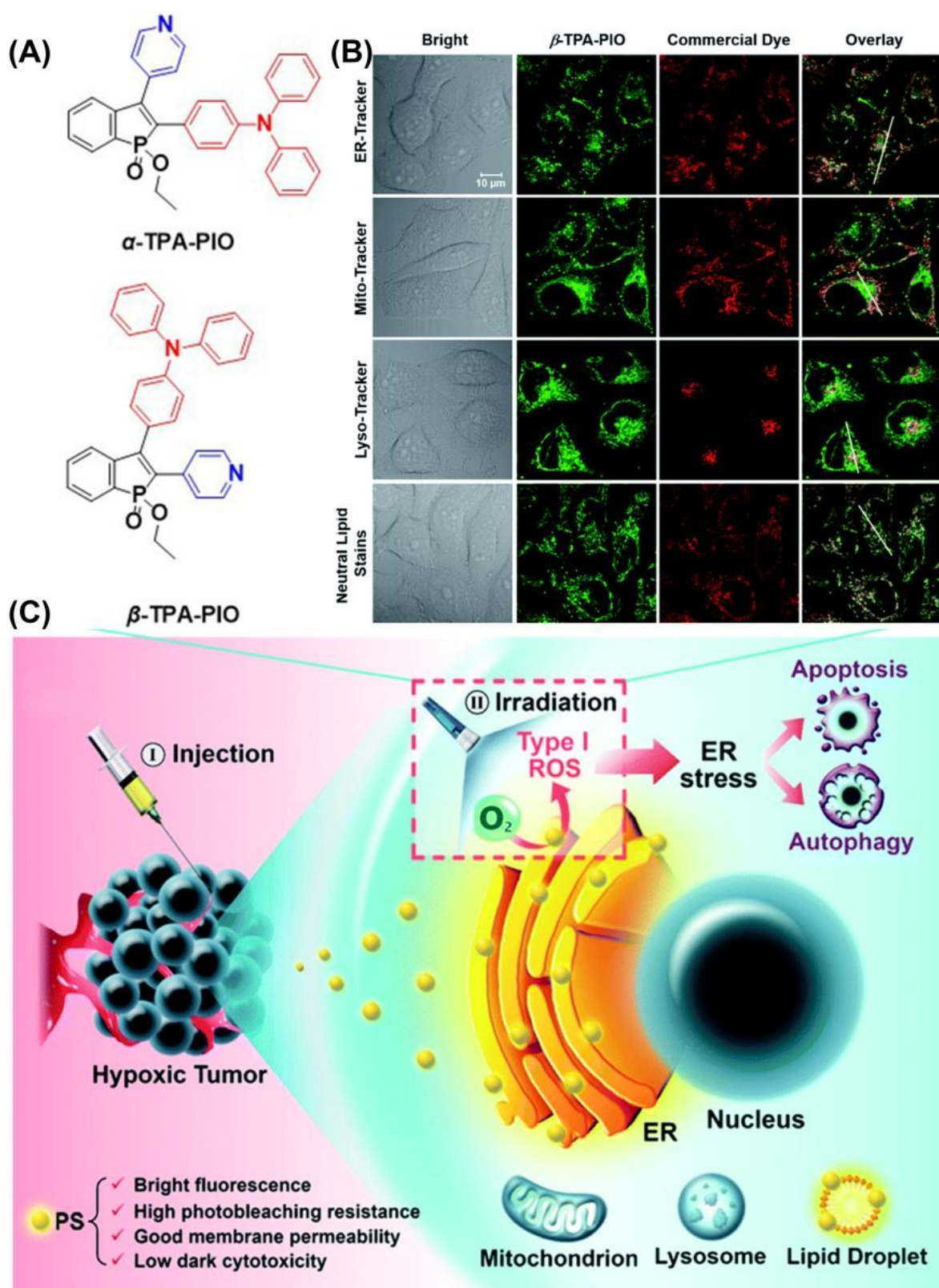


Figure 15. (A) Chemical structure of α -TPA-PIO and β -TPA-PIO; (B) CLSM images of HeLa cells co-stained with β -TPA-PIO; (C) schematic illustration of PDT treatment treated with PIO-based PSs. Reprinted with permission from Ref. [128]. Copyright 2020 Royal Society of Chemistry.

2.6. Golgi Apparatus Targeting

It has been reported for the first time that photosensitizers with AIE characterizes can target the Golgi apparatus (GA), as shown by Guo et al. (Figure 16) [129]. They synthesized and found that TPE-PYT-CPS has ER-targeting capability via caveolin/raft endocytosis. By utilizing structure–activity relationships, researchers believe cyano-induced rod-like packing in molecules plays a key role in GA targeting. Pyrene units and cyano-pyridinium salt moiety have been shown to reduce the energy gap (ΔE_{ST}) between the lowest singlet state (S_1) and the lowest triplet state (T_1), so as to promote the generation of ROS. The release of ROS causes oxidative stress and damages the GA. Then, the structural protein p115 is cleaved into N-terminal and C-terminal fragments, which are then transported into the nucleus and up-regulate apoptosis proteins p53, triggering mitochondrial dysfunction and leading to apoptosis. The decomposition rate (k_d) of ABDA in an aqueous solution of TPE-PYT-CPS, which represents ROS generation ability, was 32.85 nmol per minute. Specifically, the intracellular 1O_2 was detected using the CLSM method, and the image showed obvious green fluorescence. A Pearson correlation coefficient of 0.98 indicated that TPE-Pyt-CPS had an excellent GA targeting ability. After incubation with TPE-PYT-CPS (0.2 μ m), flow cytometry showed 56.7% apoptosis in HeLa cells. The present study provides a ground-breaking report on a promising AIE-enhanced PDT strategy, whose design has important implications for related GA-targeted photosensitizers.

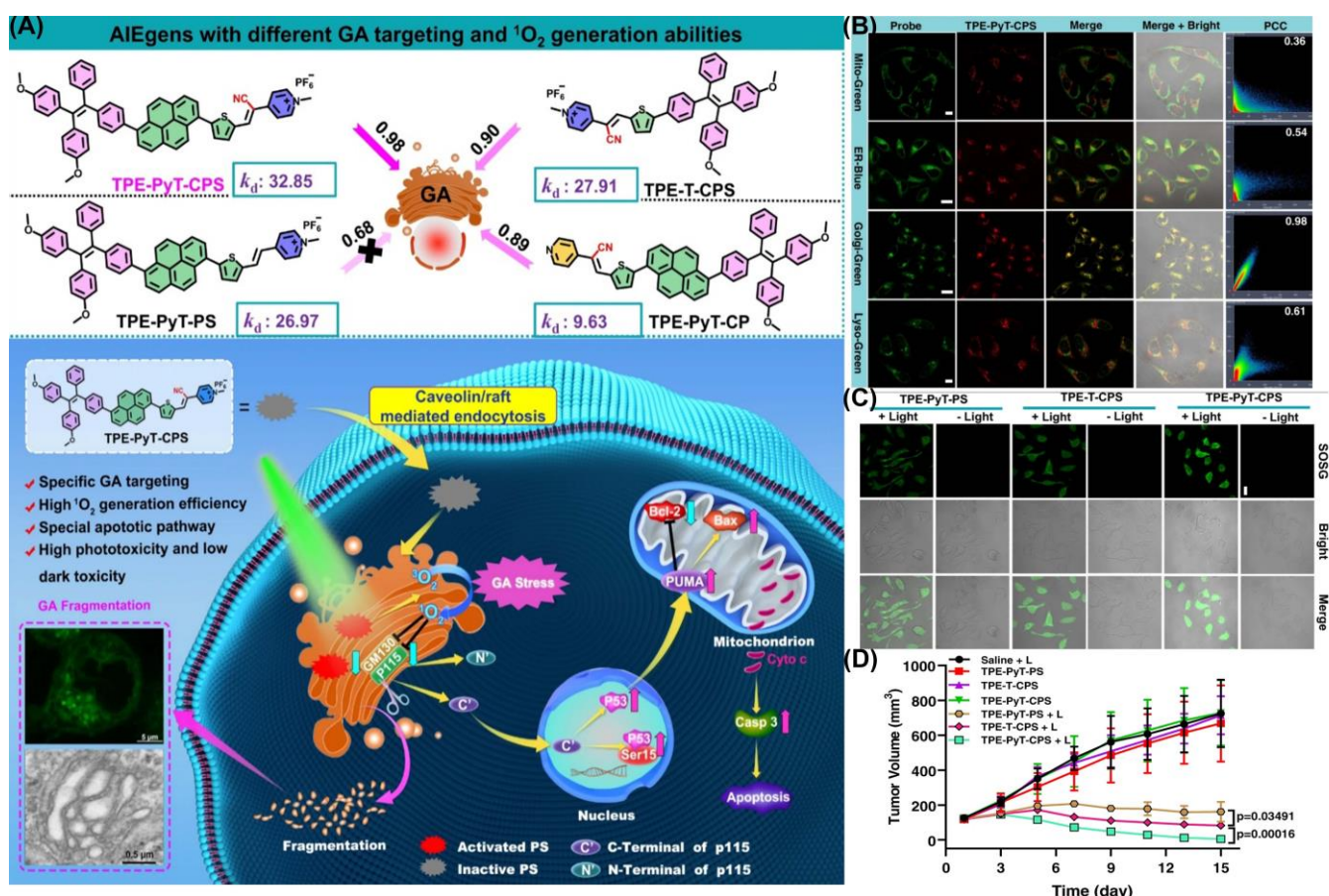


Figure 16. (A) Schematic illustration of TPE-PYT-CPS-induced GA stress and induced cell apoptosis upon PDT; (B) CLSM of HeLa cells stained with TPE-PYT-CPS and different probes; (C) intracellular 1O_2 detection by CLSM; (D) changes of tumor volume treated with different AIEgens. Reprinted with permission from Ref. [129]. Copyright 2022 Nature Communication.

2.7. Nucleus Targeting

As the “brain” in the cell, the nucleus is responsible for DNA storage, regulating cell metabolism, intracellular signaling and regulating the cell cycle [28]. The nuclear pore is located in the nuclear envelope and is approximately 40 nm in diameter, allowing some water-soluble small molecules to freely traverse the nucleus. Macromolecules such as proteins and RNA require energy and transporters to enter the nucleus. Generally, PSs target the nucleus in two ways: (1) modification of short peptide chains with nuclear targeting capability [60,61,130,131]; and (2) aptamer modification [132–135].

Recently, Mao et al. developed an AIE-PSs (MeTPAE) with nuclear targeting capability based on a triphenylamine framework (Figure 17) [130]. MeTPAE can not only combine with histone deacetylases (HDACs) to inhibit cell proliferation, but also be synergistically treated with PDT. Additionally, MeTPAE not only has high ROS generation, including Type I and Type II ROS ($\Phi_{\Delta} = 77.2\%$ in water), but its excellent two-photon absorption property also provides convenience for PDT. The fluorescence intensity of MeTPAE is further enhanced after binding to nucleic acid through electrostatic interaction and hydrogen bonding. Additionally, the Pearson coefficient of MeTPAE versus Hoechst 33342 was 0.85. Moreover, MeTPAE binding to telomeric G4 DNA caused efficient destruction of nucleic acids and inhibited telomerase activity in nucleic acid titration experiments.

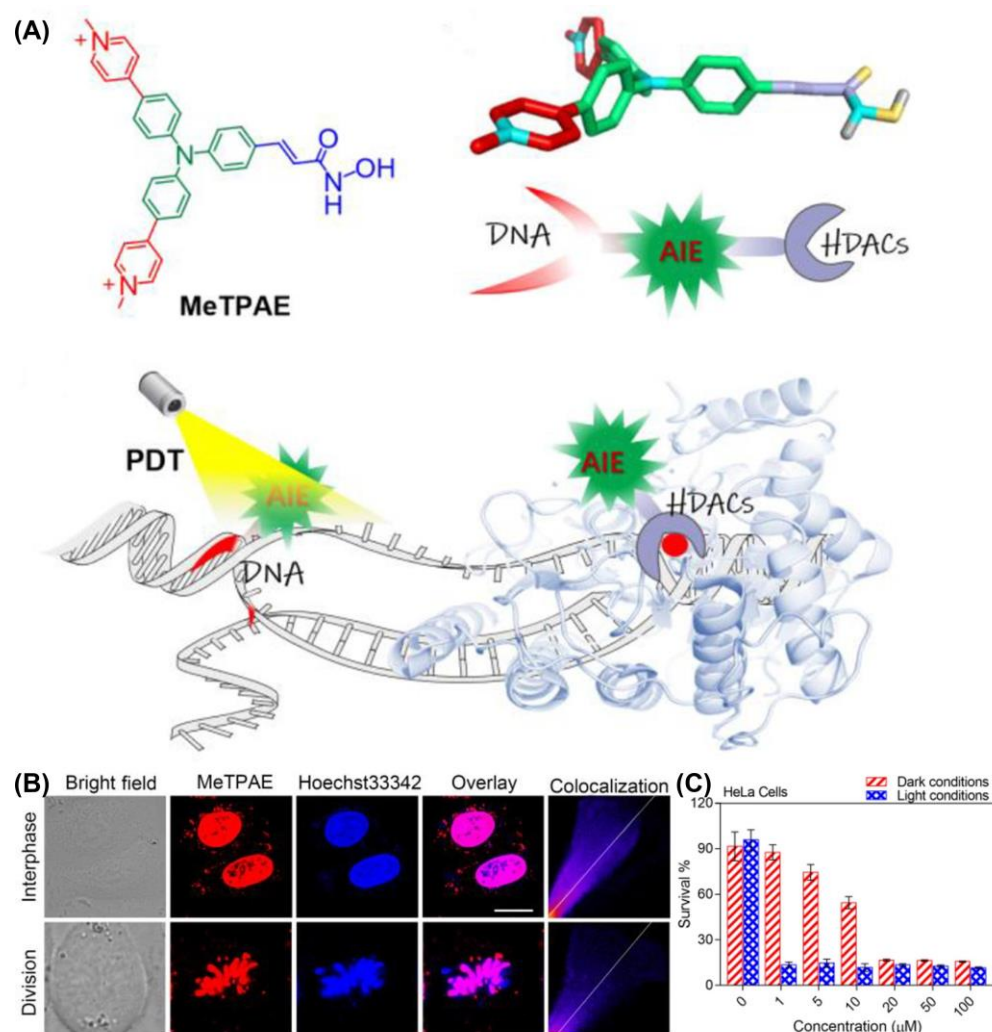


Figure 17. (A) Schematic illustration of chemical structure and mechanisms of MeTPAE with nucleic acids and histone deacetylase; (B) colocalization images of MeTPAE and Hoechst 33342 in HeLa cells during cell interphase or division; (C) cell viability of HeLa cell after being treated with MeTPAE. Reprinted with permission from Ref. [130]. Copyright 2022 WILEY-VCH.

Tang et al, developed the first AIE-PSs (TPE-4EP+) that can monitor its own photodynamic therapy response in real time in situ (Figure 18). It has an extremely high singlet oxygen production efficiency (ABDA decomposition rate of TPE-4EP+ reached $118.5 \text{ nmol min}^{-1}$ under 4.2 mW/cm^2 white light) and undergoes a process of transfer from mitochondria to the nucleus during the induction of apoptosis. The authors believe that this is because the charged TPE-4EP+ gradually dissociates from the binding to the mitochondrial membrane due to the loss of mitochondrial membrane potential during apoptosis. Moreover, due to the expansion of nuclear membrane permeability, it binds to a large number of DNA in the nucleus through electrostatic adsorption and illuminates the nucleus.

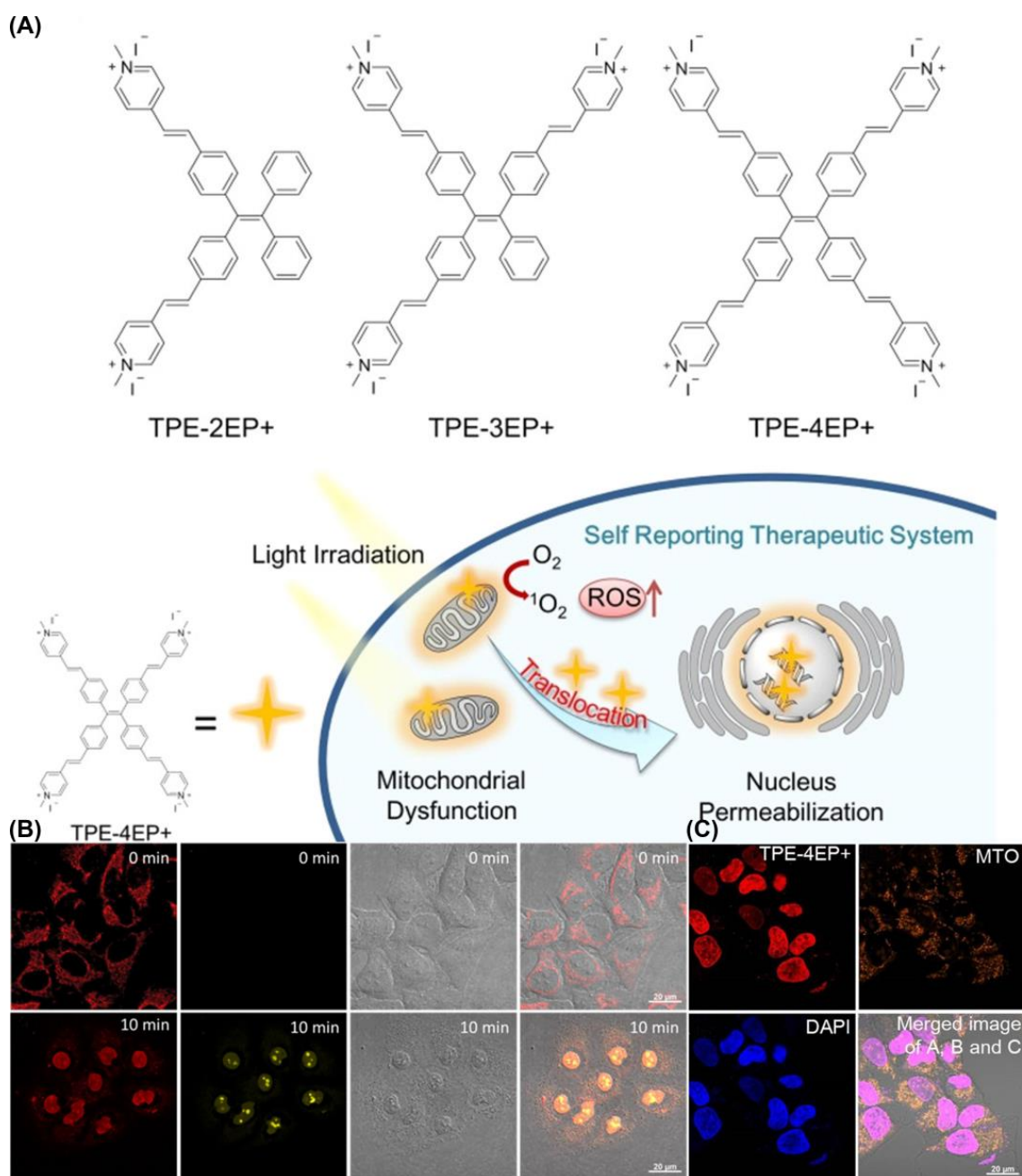


Figure 18. (A) Molecular structures and mechanism of mitochondria-to-nucleus translocation of TPE-4EP+; (B) mitochondria-to-nucleus translocation images of HeLa Cells stained with TPE-4EP+; (C) colocalization experiments of HeLa Cells co-incubation with TPE-4EP+. Reprinted with permission from Ref. [131]. Copyright 2019 American Chemical Society.

2.8. Multiple Organelle Targeting

In terms of multi-organelle targeting, a single AIE-PS has multiple-organelle-targeting capabilities by structural design. Another strategy is the use of different AIE-PSs targeting various organelles to generate ROS and damage organelles. At the same drug concentration, the therapeutic effect of drugs with the ability to target multiple organelles is better than that of drugs only target one specific organelle. This can effectively improve the therapeutic effect of PDT and reduce the use of drug concentration and the toxic side effects. Therefore, the development of PSs with multiple targeting sites has attracted much attention [41,136–141].

Tang et al. encapsulated TTFMN by introducing triphenylethylene units on the basis of TFMN [61]. To facilitate the PDT antitumor effect of TTFMN, a pH-activated TAT peptide-modified amphiphilic polymer encapsulates TTFMN to transport PSs into the tumor nucleus (Figure 19). Both TFMN and TTFMN emission wavelengths reached NIR (651 nm). Moreover, these two compounds had great application potential as they all produced ROS through Type I mechanisms (intensity of DCFH containing TTFMN was enhanced by nearly 500-fold under 22.1 mW cm^{-2} white light irradiation in 5 min). After entering the cell, TTFMN first enters the lysosome, where it is activated by acid and then transported to the nucleus. With the growth of incubation time, some TTFMN-NPs translocated to the perinuclear region. When the time reached 12 h, a large number of TTFMN-NPs crossed the perinuclear region and even partially entered the nucleus. The apoptosis of tumor cells induced by TTFMN-NPs was confirmed by terminal deoxynucleotidyl transferase-mediated nick end labeling (TUNEL) staining. In all organs of mice treated with TTFMN-NP, H&E-stained slices showed no significant organ damage.

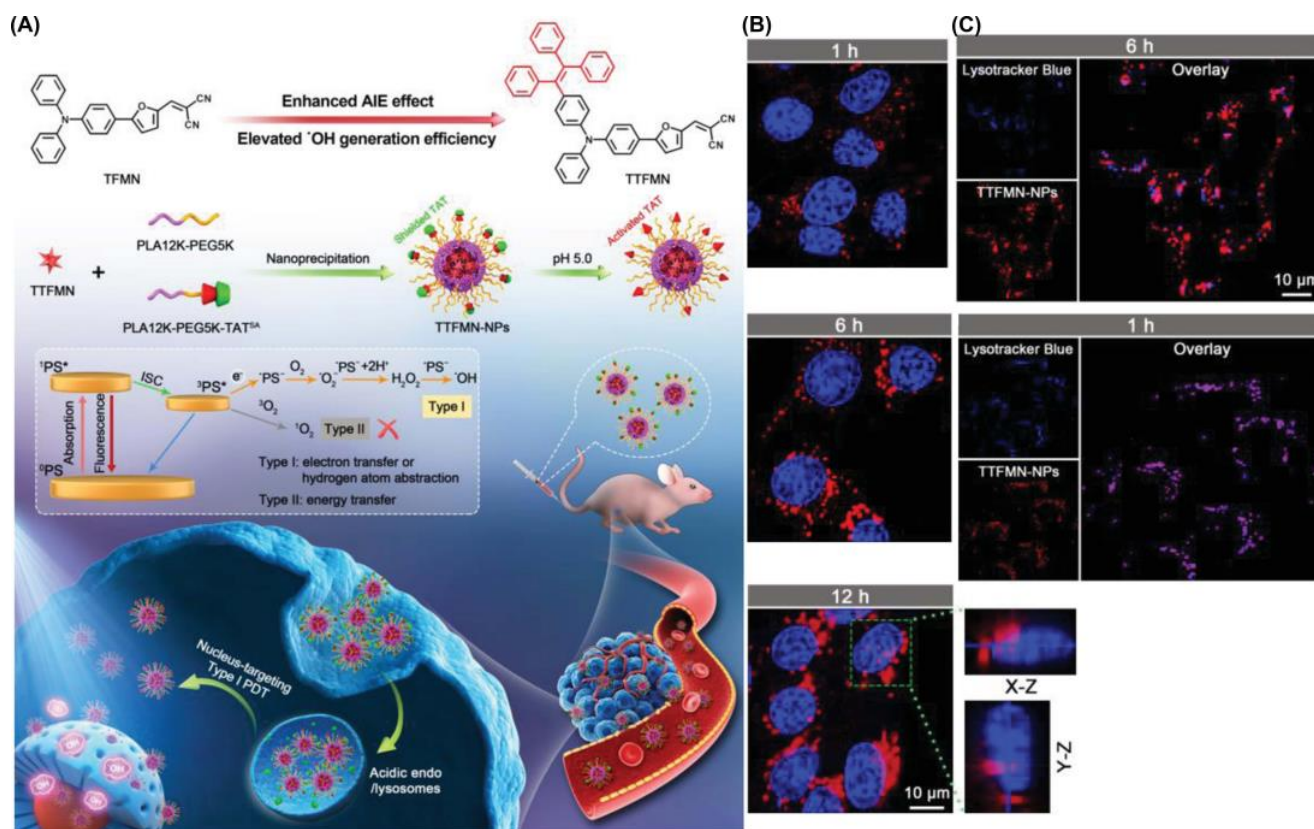


Figure 19. (A) Molecular structures and multifunctional phototheranostic application of TTFMN-NPs; (B) intracellular tracking on 4T1 cells; (C) CLSM images of nuclear targeting delivery on 4T1 cells. Reprinted with permission from Ref. [61]. Copyright 2021 WILEY-VCH.

After that, the authors reported another modified AIE-PS for nuclear targeting (Figure 20) [60]. TPE-TTMN-TPA was synthesized by infusing a diphenylamine structure on the basis of TTMN. Compared with the previous work, the TPE-TTMN-TPA nanoparticles had more red-shifted NIR emissions, higher Type I ROS generation capacity (intensity of DCFH enhanced nearly 600-fold after 10 min of 22.1 mW cm^{-2} white light irradiation) and better nuclear-targeted delivery. T4-NPs composed of TPE-TTMN-TPA and SA-TAT also needed to be activated by lysosomal acid. After incubation for 1 h, the Pearson coefficient of T4-NPs and lysosomes reached 0.94, while it decreased to 0.62 after 6 h, proving its effective escape from lysosomes. As shown in the image, more and more T4-NPs entered the nucleus as time elapsed. Flow cytometry analysis showed that a large number of tumor cells underwent apoptosis after photoexcitation, which proved the effectiveness of the PDT effect.

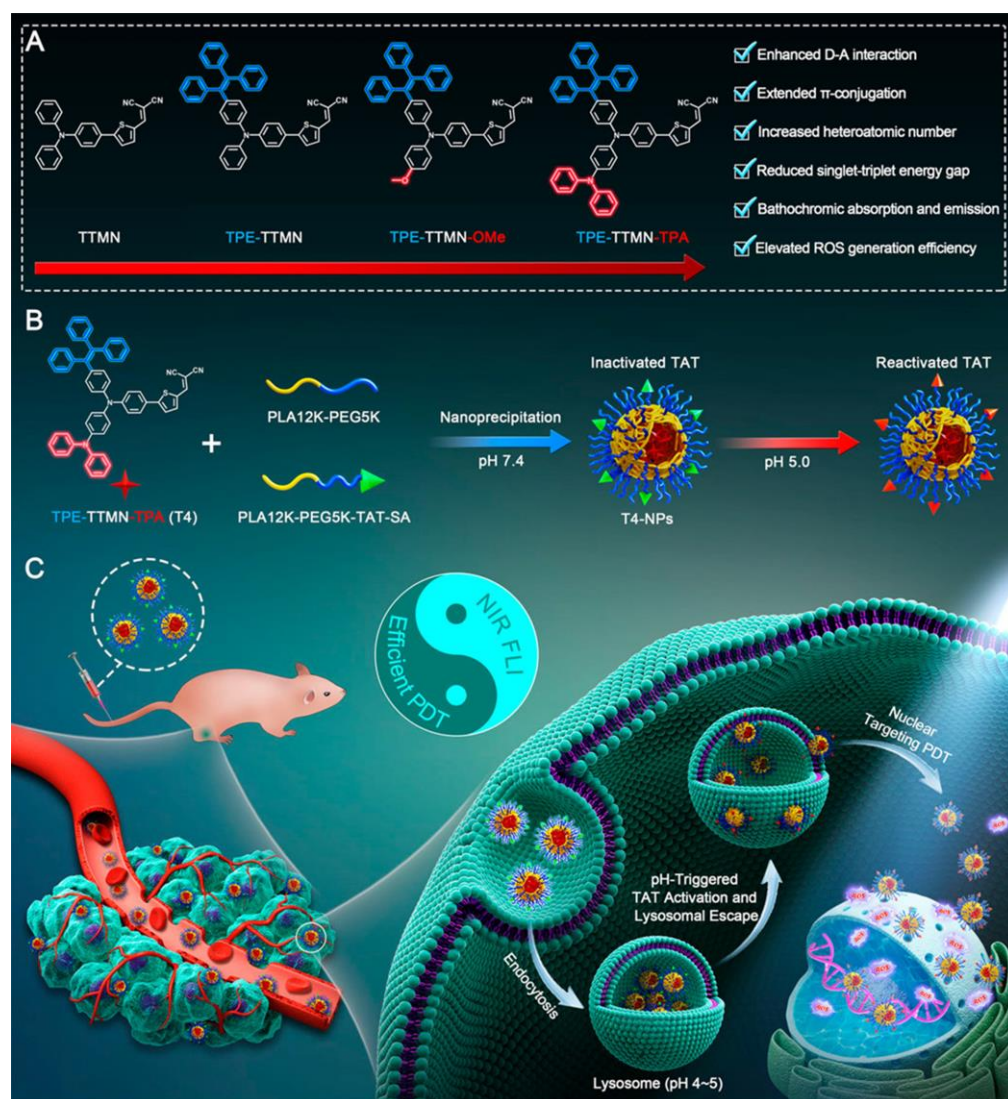


Figure 20. Schematic illustration of (A) molecular engineering of TPE-TTMN-TPA, (B) construction of nucleus-targeted T4-NPs, and (C) applications in PDT. Reprinted with permission from Ref. [60]. Copyright 2021 American Chemical Society.

Another example is the single AIE-PS (TBZPy, MTBZPy, TNZPy, MTNZPy) with lysosomal and mitochondrial targeting capabilities reported by Tang et al., (Figure 21) [138]. By constructing strong intramolecular charge transfer (ICT), the electron-rich system can facilitate the progress of Type I PDT by providing electrons. In the presence of TNZPy, they used H2DCF-DA as an indicator to define the ROS generation where the intensity increased

140-fold after white light irradiation (50 mW cm^{-2}). The Pearson coefficients of TNZPy for lysosomes and mitochondria were 0.81 and 0.88, respectively. With the development of time, the Pearson coefficient of lysosomes gradually decreased, and the corresponding mitochondrial gradually increased. This suggests that TNZPy can efficiently escape from lysosomes and accumulate in mitochondria. The authors suggest that ROS generation after photoexcitation synergistically destroys lysosomes and mitochondrial organelles to induce apoptosis. The IC_{50} value of TNZPy for HeLa cells was less than $6 \mu\text{mol}$ under both hypoxic and normoxic conditions, indicating potential for treating tumors under hypoxia conditions. In the *in vivo* experiment, the body weight of tumor-bearing mice did not change significantly, while the tumor growth was significantly inhibited.

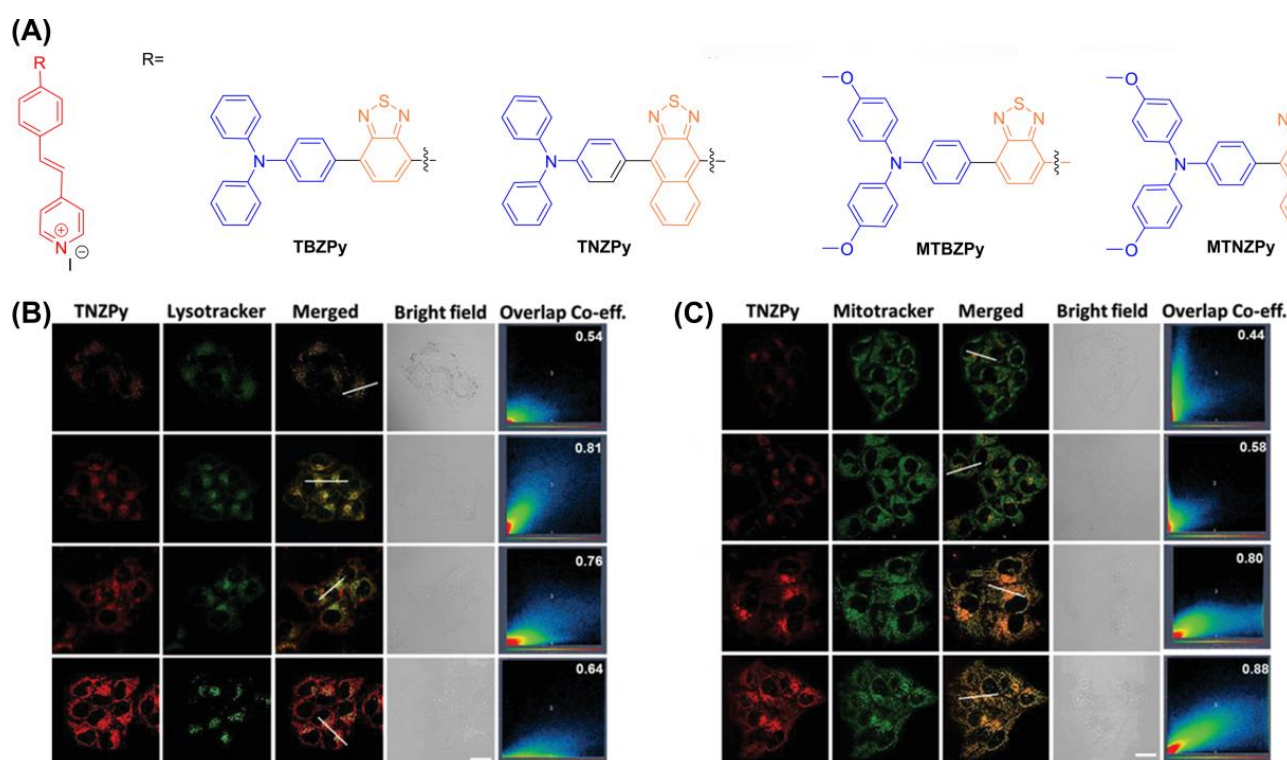


Figure 21. (A) Chemical structure of TBZPy, MTBZPy, TNZPy, MTNZPy; (B) CLSM images of HeLa cells stained with TNZPy and Lysotracker green; (C) CLSM images of HeLa cells stained with TNZPy and Mitotracker green. Reprinted with permission from Ref. [138]. Copyright 2020 WILEY-VCH.

In 2020, Tang et al. proposed a pioneering strategy of $1 + 1 + 1 > 3$ (Figure 22) [41]. They synthesized a series of different AIE-PSs (TFPy, TFVP and TPE-TFPy) but with the same skeletal structure through subtle structural adjustments, and they were able to specifically anchor to mitochondria, cell membranes, and lysosomes to damage organelles by producing ROS. The $^1\text{O}_2$ quantum yield of TFPy, TFVP and TPE-TFPy compared with rose bengal were 25.2%, 18.3% and 63.0%, respectively, suggesting good ROS production. In situ, TPE-TFPy aggregates easily form nanosized aggregates that endocytose into lysosomes and specifically illuminate them. The positively charged pyridine moiety of TFPy may bind to mitochondria, leading to mitochondrial targeting capability. In part, TFVP's lower membrane permeability coefficient may be due to its higher free-energy barrier, which confers its specific aggregation properties on the cell membrane. Fluorescence imaging results showed that TFPy, TFVP and TPE-TFPy showed strong targeting ability towards mitochondria, cell membranes and lysosomes, respectively. Compared with the single photosensitizer treatment, the combination of the three treatments significantly enhanced the anti-tumor effect of PDT. Notably, the synergistic treatment of the three did not affect their biocompatibility.

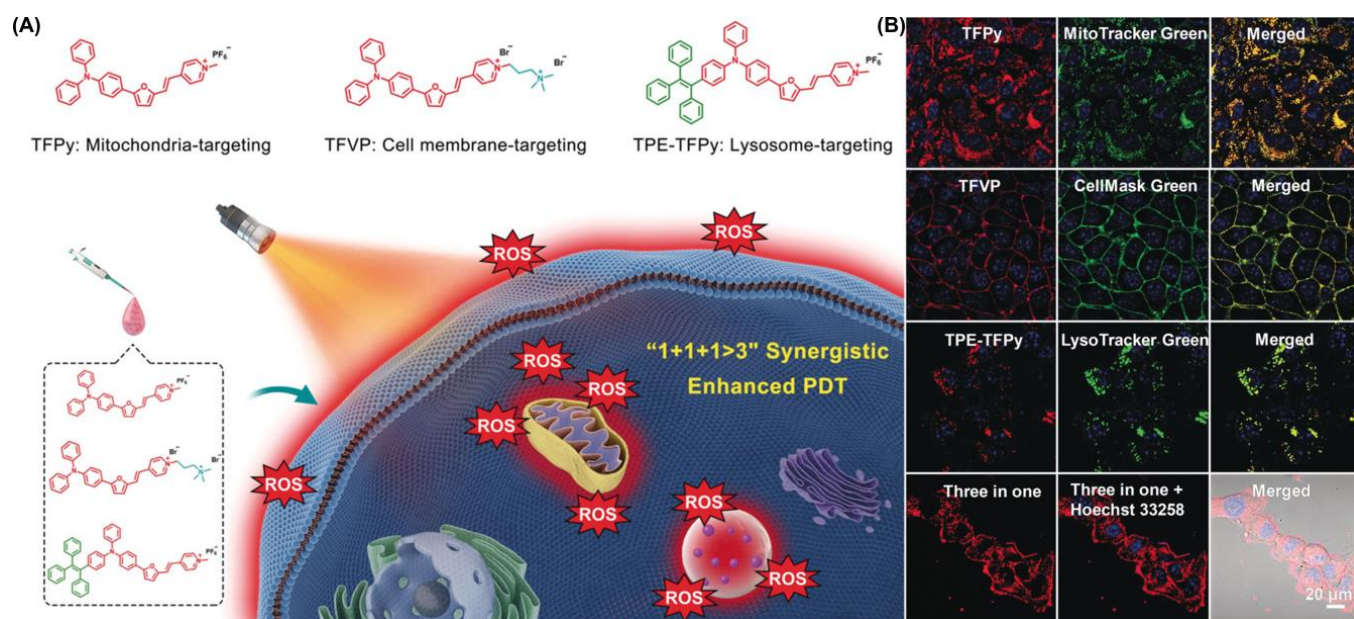


Figure 22. (A) Schematic illustration using TFPy, TFVP and TPE-TFPy for achieving $1 + 1 + 1 > 3$ synergistic enhanced PDT; (B) co-localization of these three AIEgens. Reprinted with permission from Ref. [41]. Copyright 2020 WILEY-VCH.

3. Tumor-Targeting AIE-PDT

With the continuous research of anti-tumor drugs and the rapid development of tumor biology, PSs are usually encapsulated into nanoparticles to enhance their absorption in the biological environment and obtain better targeting ability [142,143]. For nanoparticles (NPs), tumor-targeting strategies are divided into active-targeting and passive-targeting strategies [144–146]. Nanomedicine can extravasate and remained in the pathological site mainly based on the enhanced permeability and retention (EPR) effect, which can be classified as passive targeting [147,148]. However, the EPR effect can only deliver very limited amounts of PS to tumor tissues and its efficiency has been challenged in recent years. In this regard, some active targeting strategies have been proposed to help enhance tumor-targeting ability, such as the modification of some ligands that specifically bind to tumor-overexpressed receptors, and encapsulating PSs into some engineered cell membranes as camouflage for tumor targeting, etc.

3.1. Passive Targeting

Generally, passive targeting is based on the EPR effect, optimizing the size or surface properties of nanoparticles [143,149]. Nano-systems (20–200 nm) can selectively penetrate tumor stroma via newly formed leaky vessels [150–152]. Compared with free drug molecules, NPs are preferentially accumulated at tumor sites through the EPR effect. There are two main ways to enhance the EPR effect: (1) A greater penetration of NPs through the extracellular matrix (ECM) could improve the EPR effect [144,153,154]. Injecting hyaluronidase (HAase) to decompose the ECM structure would be an efficient method. However, there is no AIE-PS based on this approach that has been reported. (2) Since albumin has a long circulation half-life and continuous uptake in tumor tissues, it can help enhance the EPR effect. (3) Using a carrier such as PEG-encapsulating drugs to form nanoparticles. There are several AIE-PSs with an association with albumin that have been reported [155–158].

Recently, Tang et al. first proposed a mitochondria-targeting two-photon PSs (TPABP-Ir) with AIE properties based on an Ir(III) structure to generate Type I and Type II ROS (Figure 23) [156]. TPABP-Ir was coated with BSA to form TPABP-Ir@BSA nanoparticles (Ir-NPs). Colocalization imaging showed that the Pearson coefficient of Ir-NPs with mi-

tochondria reached 0.86, indicating specific targeting of mitochondria. They also used DCFH as an indicator to detect total ROS production, and the fluorescence of the group treated with TPABP-Ir was 10 times higher than RB, and 17 times higher than Ce6. In addition, a significant increase in ROS production was observed in MCF-7 cells treated with Ir-NPs compared with the control cells, indicating a good ROS generation capacity. For in vivo experiments, the Annexin V-FITC and MTT assays demonstrated a good inhibitory ability of Ir-NPs by inducing apoptosis. Similar to in vitro experiments, Ir-NPs significantly inhibited tumor growth in tumor-bearing mice without obvious systemic toxicity.

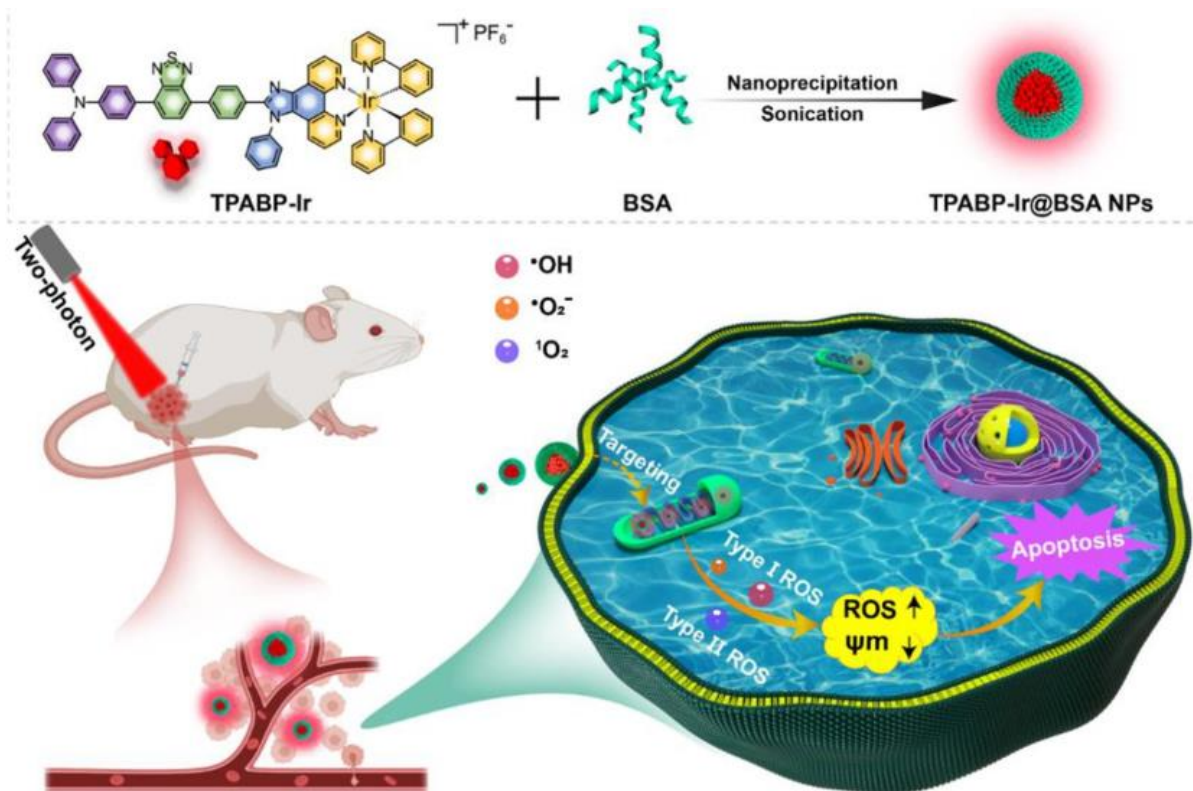


Figure 23. Schematic illustration of TPABP-Ir@BSA NPs for PDT treatment. Reprinted with permission from Ref. [156]. Copyright 2022 Elsevier Ltd.

In 2021, Liu et al. [159] reported a new coordination polymer nanoparticle (CPN) which could achieve synchronous radiotherapy (RT) and radiodynamic therapy (RDT) under X-ray irradiation (Figure 24). They synthesized Hf-AIE-PEG-DBCO nanoparticles with a significant tumor inhibition effect by encapsulating TPEDC-DAC (AIE-PSs) using PEG modified with DBCO (dibenzocyclooctyne). Bioorthogonal click chemistry was performed by adding the metabolic precursor Ac₄ManNAz (used to modify azide groups on cell membrane glycans) and DBCO-modified PEG to enhance the accumulation and prolong the retention of CPNs in tumors. Compared with the control group, Hf-AIE-PEG-DBCO exhibited a more obvious increase in fluorescence of DCFH-DA pretreated with Ac₄ManNAz under X-ray irradiation, which is consistent with better tumor inhibition in vitro. Hf-AIE-PEG-DBCO showed a strong inhibitory effect on tumor growth in vivo as well as in vitro. H&E staining of tumor tissues showed significant tumor death after co-incubation with Ac₄ManNAz and HF-AIE-PEG-DBCO under light. In this study, the combination of radiotherapy and RDT had a significant killing effect following intravenous injection of CPNs, due to the high penetration of X-rays and the DBCO-mediated bioorthogonal click chemistry.

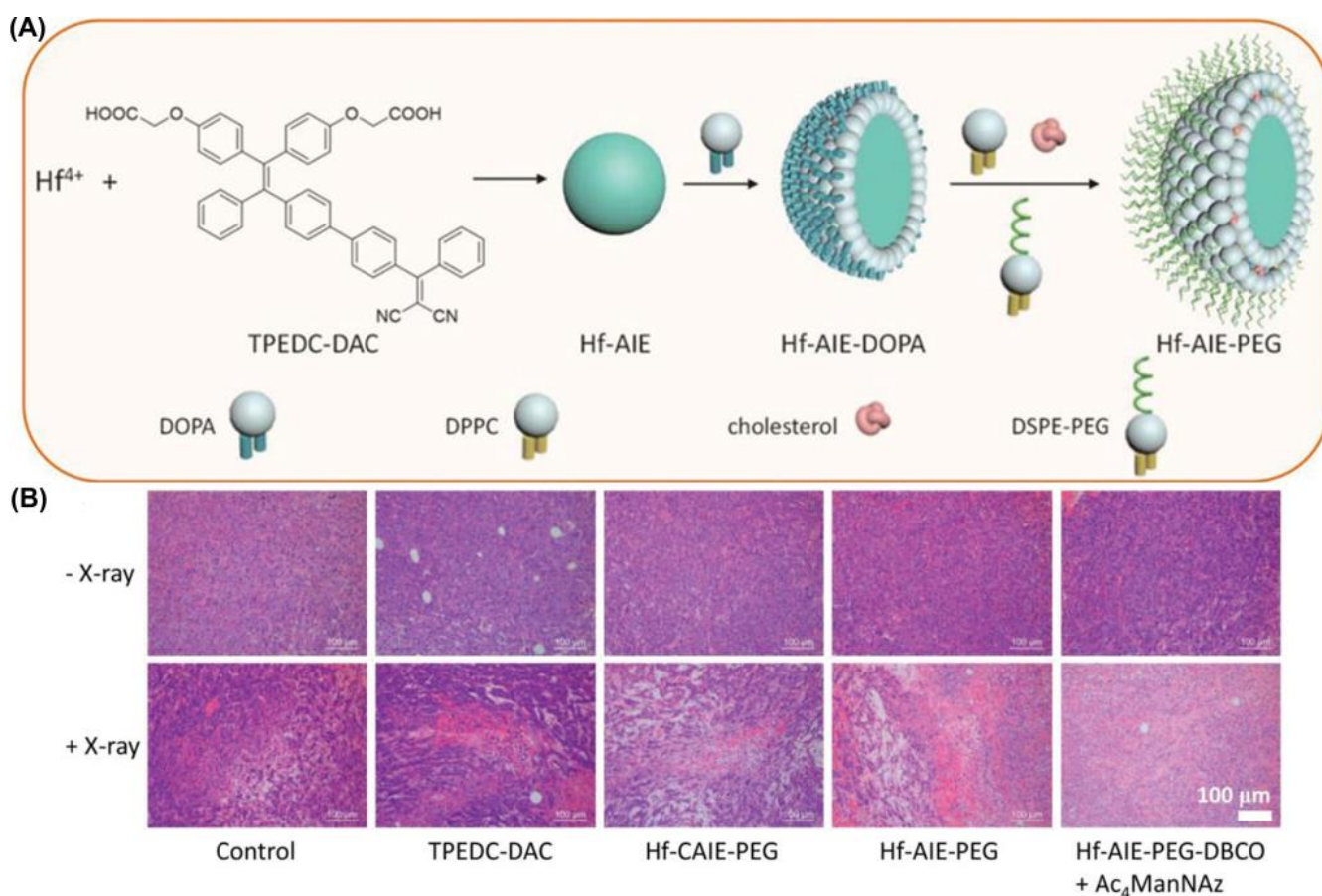


Figure 24. (A) Schematic illustration of fabrication of Hf-AIE-PEG; (B) H&E staining analysis of tumor tissues treated with various treatments. Reprinted with permission from Ref. [159]. Copyright 2021 WILEY-VCH.

Liu et al. also developed another example of PEG-modified nanoparticles of variable size [160]. NPs with a size of 100–200 nm had a better tumor enrichment effect through the EPR effect, while smaller NPs (<50 nm) had minimal adhesion to the extravasation site of tumor blood vessels and extracellular matrix, which favored intratumoral penetration. They developed Dox-PEG-PS@MIL-100 NPs for the pH-response of photosensitization and the nanoparticle size-reducing process (Figure 25). H_2O_2 could break down the tumor intake of Dox-PEG-PS@MIL-100 NPs and release TPABTDCT (AIE-PSs) for the activatable PDT process. Meanwhile, Dox-PEG can self-assemble into ultra-small nanoparticles (DOX NPs) that can penetrate deep into tumors. After that, Dox was released into the nucleus to damage DNA under a low-pH environment. It was proved that TPABTDCT has better $^1\text{O}_2$ generation efficiency than Ce6. This work achieved advanced photodynamic–chemotherapy combination therapy.

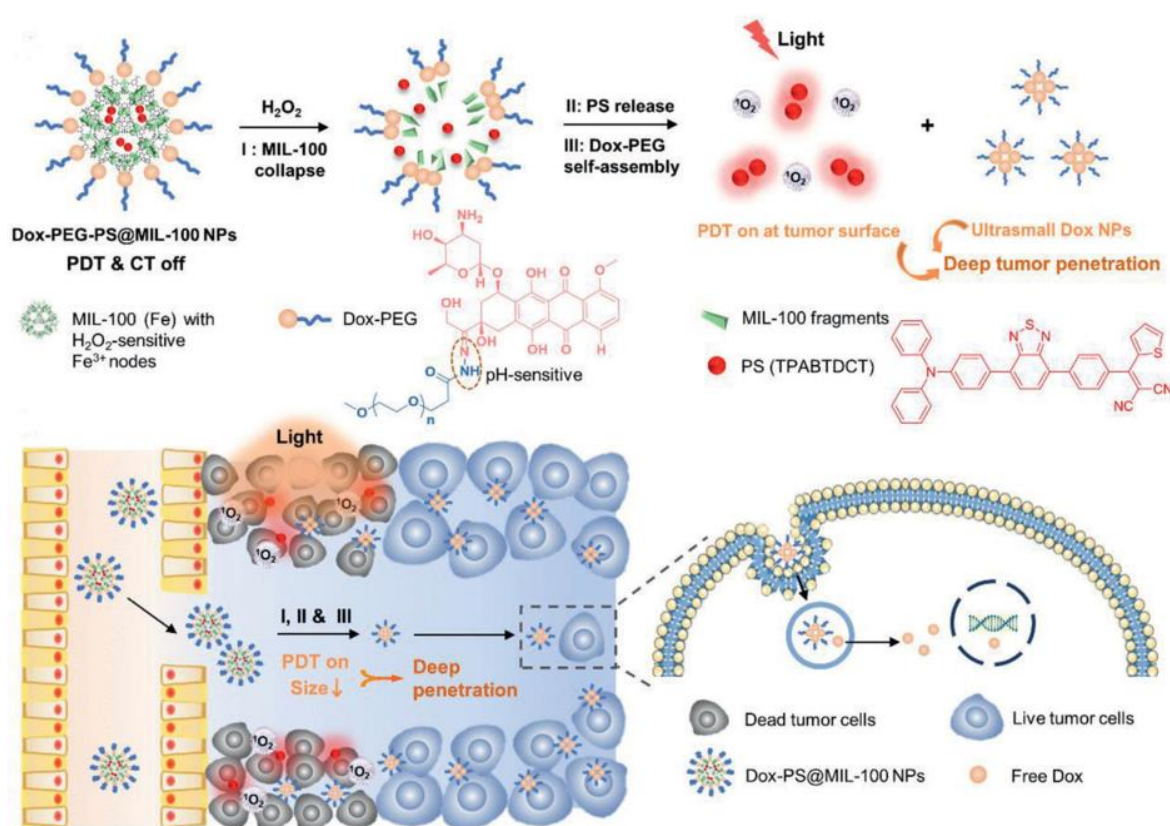


Figure 25. Schematic illustration of Dox-PEG-PS@MIL-100 NPs for advanced photodynamic-chemotherapy combination therapy. Reprinted with permission from Ref. [160]. Copyright 2021 WILEY-VCH.

3.2. Active Targeting

In recent years, scientists have become aware that EPR effects vary over time during tumor development, and that they are highly heterogeneous [145,161–164]. To enhance specificity, active targeting is increasingly preferred. Active targeting is primarily performed by modifying bioligands on the surface of NPs that have the ability to target specific receptors on tumor cells [149,165]. Current active targeting strategies mainly target growth factor receptors overexpressed in cancers of different tissue origins, such as folate (FA) [166], transferrin (Tf) receptor [167], epidermal growth factor receptor (EGFR) [168,169] and so on [170–172]. Active-targeting strategies for PDT are well-summarized in other review papers [172].

As tumors require more biotin than normal tissues, linking biotin units can be used to target tumors through overexpressed biotin receptors. In a recent work, Chen et al. introduced nitrobenzoic acid (TTVBA) and biotin units to synthesize AIE-PSs (TTVBA) that avoided fluorescence quenching caused by PET [173]. Moreover, the fluorescence enhancement was the highest that had ever been reported during the aggregation process. The colocalization experiment of HeLa cells demonstrates that Biotin-TTVBA was accumulated in the cytoplasm with a bright red color. Besides, Biotin-TTVBA also had high photobleaching resistance after 350 s irradiation. Additionally, the IC_{50} values for Biotin-TTVBA were 2.5, 2.5 and 10 μ M for HeLa, MCF-7, and L-O2 cells, which means that Biotin-TTVBA could selectively kill tumor cells with a high expression of the biotin receptor.

Another example is using EGFR to achieve the ability to target tumors. DCTBT is a newly developed photosensitizer with AIE characteristics, which enables NIR-II (1000 nm) fluorescence imaging, Type-I PDT and photothermal therapy (PTT), as reported by Tang et al. (Figure 26) [174]. Amphiphilic polymers modified with an EGFR-targeting

peptide were doped to encapsulate DCTBT. The ROS species results showed that the DCTBT produced only $\bullet\text{OH}$ and $\text{O}_2^{\bullet-}$ through the Type I pathway. Besides, the DCTBT NP has comparable ROS production efficiency with rose bengal. The photothermal efficiency experiment showed that DCTBT NPs had better photothermal performance (59.6%) than CTBT NPs. In vivo testing of DCTBT NPs on PANC-1 tumors revealed that they remained fluorescent 48 h after injection. It was noted that the Target-NPs (with EGFR-targeting peptide) had a better anti-cancer effect than the non-Target-NPs (without EGFR-targeting peptide). The combination of Type I PDT-PTT and DCTBT significantly inhibited the growth of PANC-1 tumors in vitro and in vivo. This approach shows great promise for overcoming tumors in hypoxic environments.

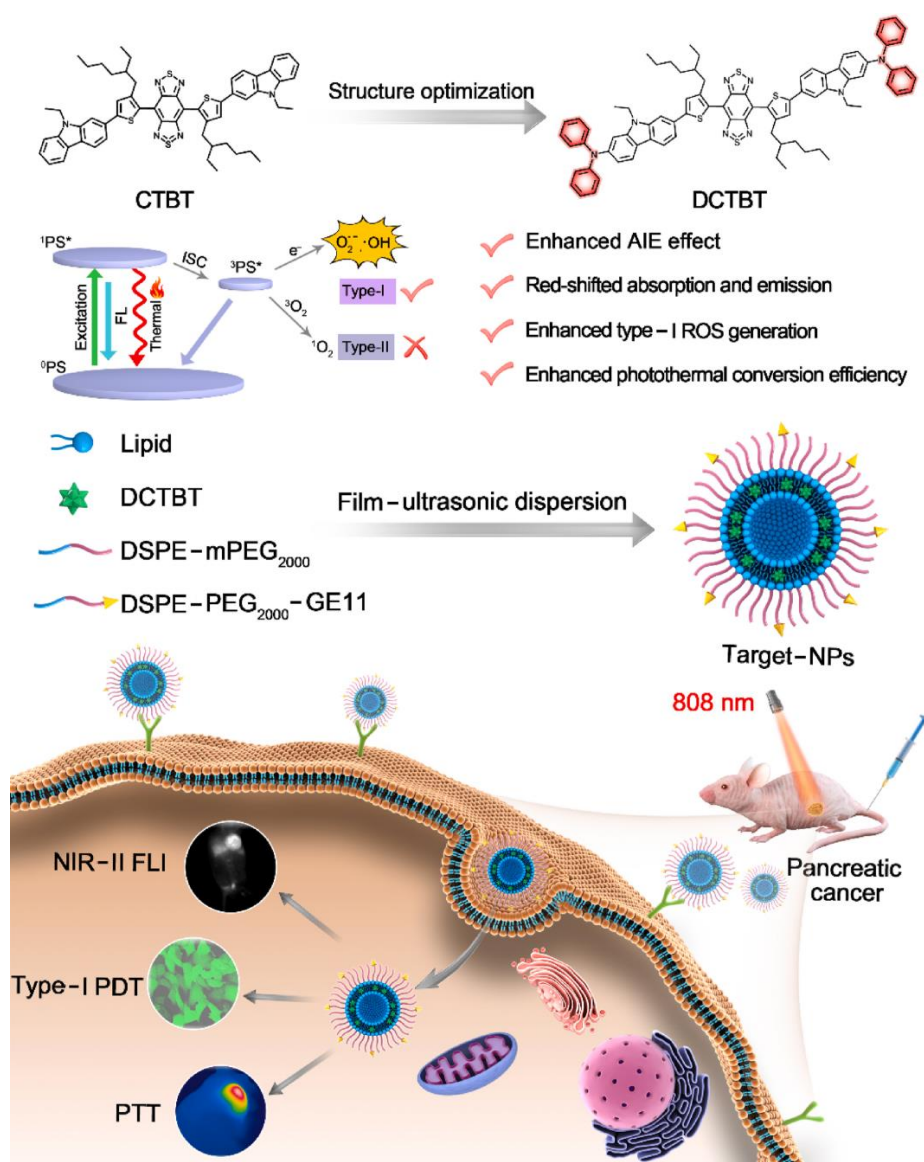


Figure 26. Schematic illustration of DCTBT, and the application on NIR-II FLI-guided Type-I PDT-PTT. Reprinted with permission from Ref. [174]. Copyright 2022 Elsevier Ltd.

In recent years, a series of new active-targeting methods have emerged, including targeting tumor tissues under hypoxic conditions by the anaerobic nature of some bacteria [175,176], such as *Escherichia coli*, or by using cell membrane camouflage or liposomes to target tumors [177–181]. In addition, using red blood cell membranes to disguise NPs can deceive the immune system and reduce the immune response to NPs. The use of stimuli-

responsive PDT therapy such as ROS-responsive, pH-responsive, hypoxia-responsive, redox-responsive, and so on, is also a major research topic [182–185]. Simply put, a tumor-targeting strategy can effectively reduce the toxic and side effects of photosensitizers and improve the killing ability of tumors, especially hypoxic tumors.

Under hypoxic conditions, Tang developed a novel approach for addressing the problem of drug resistance by combining bacteria with the Type I PDT system of the TBP-2 (Figure 27) [186]. A PDT intervention of this kind has never been reported before, which has shown to significantly impair orthotopic colon cancer growth and overcome pre-treatment toxicity. PDT-mediated cancer treatment can be delivered effectively to hypoxic tumors, because TBP-2 contains two cationic structures enabling *E. coli* to absorb it into the periplasmic space. The authors found that AE had brighter pictures and mainly concentrated near the cell membrane. The co-localization experiment involving AE and a hypoxic area proved that AE appeared in a hypoxic environment. Similarly, another work by Tang et al. using TBPP with similar structure and uropathogenic *Escherichia coli* (UPEC) to induce urinary tract infection (UTI) also had the function of targeting hypoxic tumors and had a good effect on the treatment of bladder cancer [187].

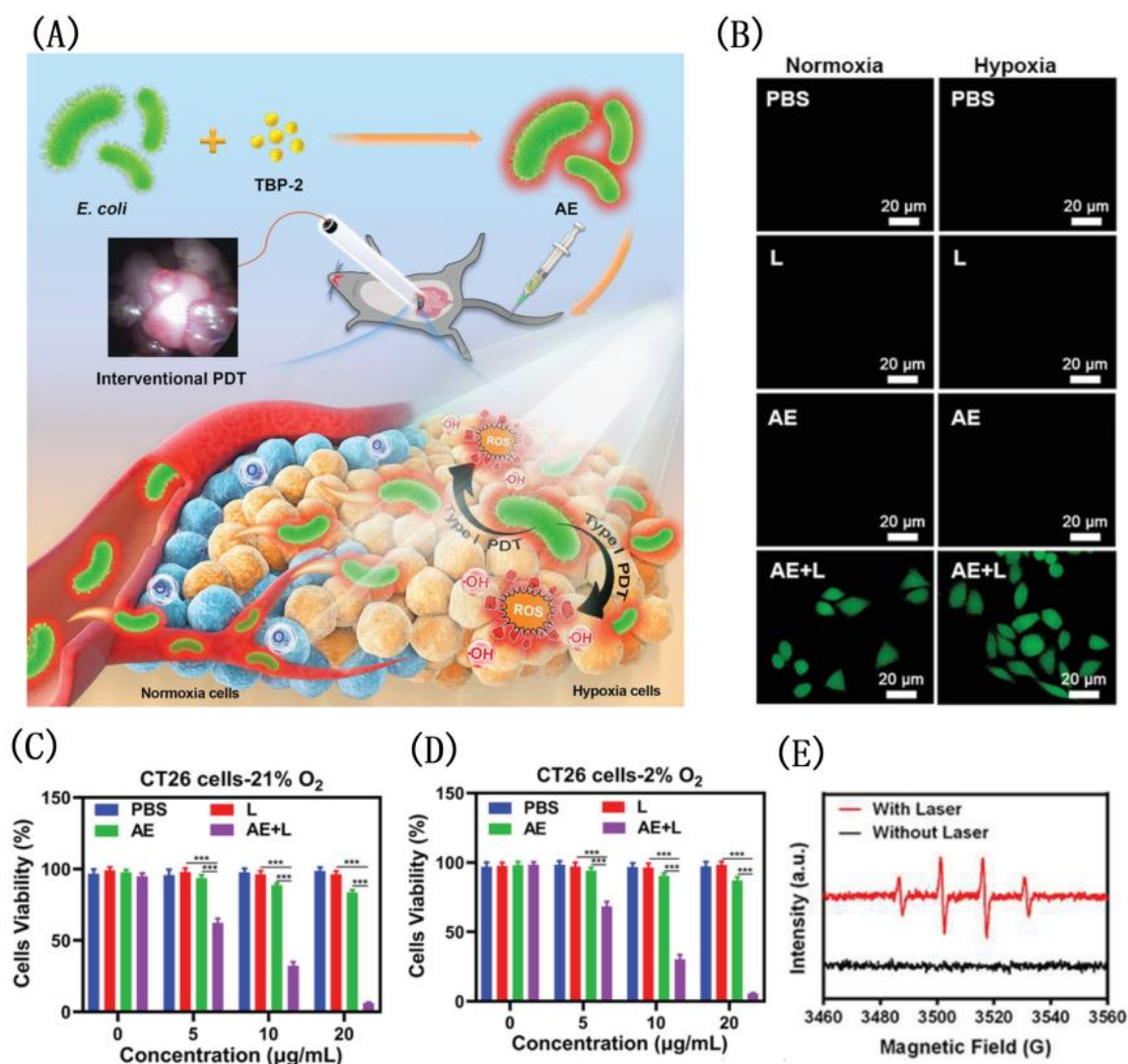


Figure 27. (A) The structure of TBP-2; viability of CT26 cells under normoxia (B) and (C) hypoxic conditions; (D) CLSM image of intracellular ROS; (E) schematic illustration of bright bacteria with TBP-2 for photodynamic therapy. Reprinted with permission from Ref. [186]. Copyright 2021 WILEY-VCH.

Using PLT-derived vesicles (PV) from mouse blood samples, Tang et al. synthesized a biomimetic nano-enzyme (PMD) by wrapping DCPy and MnO_2 nanoparticles (Figure 28) [188]. In the co-localization experiment, CT26 cell imaging showed that PMD was more closely related to cells. Subsequently, the authors found that CT26 cells treated with PMD had stronger fluorescence and higher Mn content at the same concentration compared with erythrocyte-membrane-coated MD (RMD). Compared with control, the group containing PMD had better ROS production (nearly 15-fold). Hypoxia-inducible-factor (HIF-1 α) staining treated with PMD found that there was almost no HIF-1 α . MTT assay of CT26 cells showed that PMD had a good and similar IC_{50} value regardless of hypoxia or normoxia. This indicates that it has a good inhibitory effect on tumor under hypoxic condition. A superior tumor-targeting effect of PMD particles was demonstrated by intravenous injection into CT26 tumor-bearing mice of RMD or PMD, and PMD accumulation was significantly higher than that of RMD. This work proved that molecules with PLT derived vesicles have better PDT efficacy than nanoparticles disguised by red blood cells alone, which provides guidance for the future development of related work.

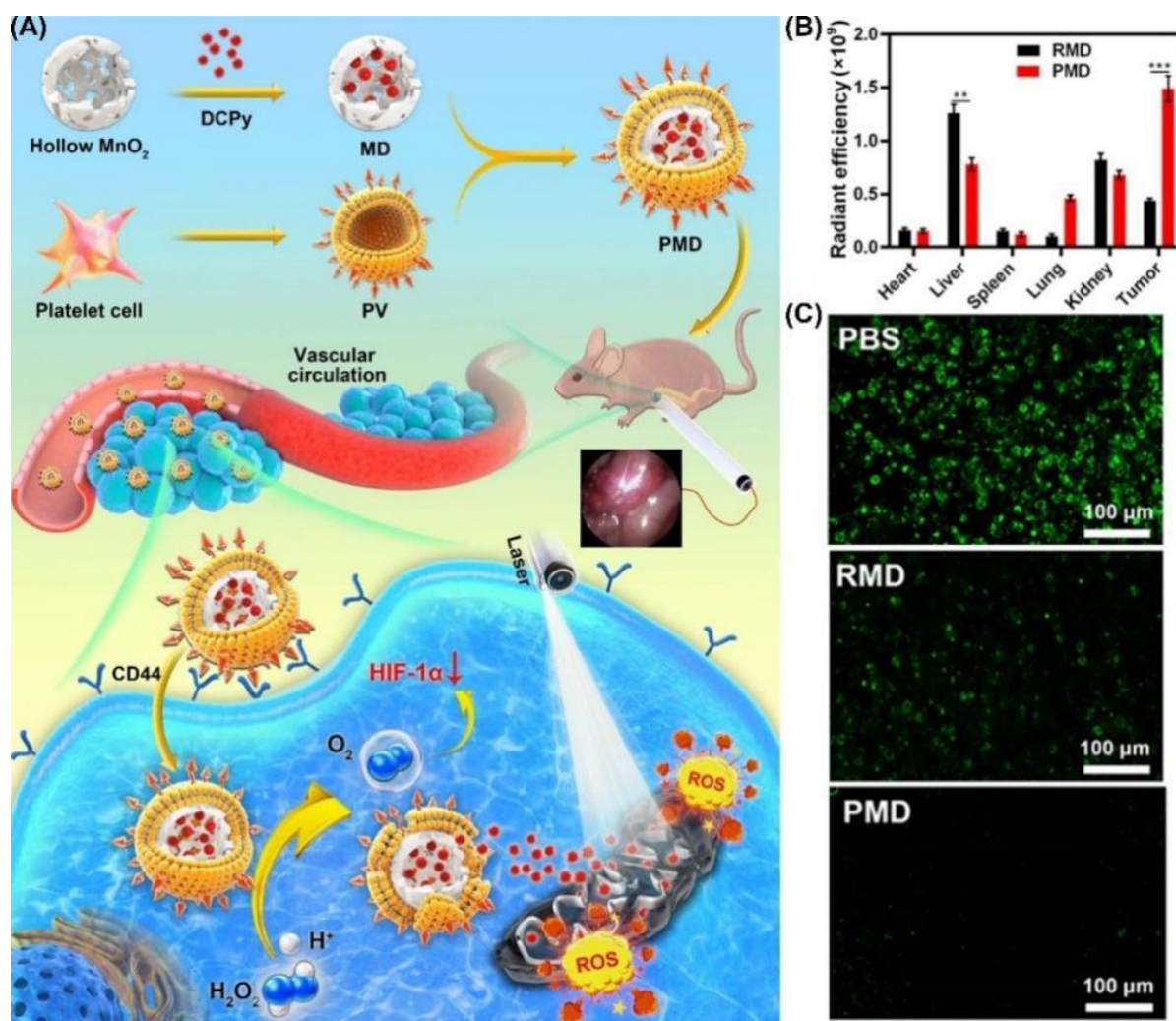


Figure 28. (A) Schematic illustration of the use of DCPy and PV as bionic nanozymes for photodynamic therapy; (B) radiant efficiency in tumors and different organs after PMD or RMD injection; (C) fluorescent staining for HIF-1 α (representing intratumoral hypoxia). ** $p < 0.01$, *** $p < 0.001$. Reprinted with permission from Ref. [188]. Copyright 2022 American Chemical Society.

4. Conclusions and Perspectives

As a new promising anti-tumor method, PDT has attracted wide attention. This review introduces organelle-targeting and tumor-targeting PSs based on AIE strategies from the past three years. We introduced the chemical structures of various PSs and their functions as well as the mechanisms of PSs to treat tumor cells. Additionally, we classified their targeting mechanisms and principles. Although the emergence of AIE strategies has solved the problem of poor efficacy of PDT in vivo to a certain extent, PDT still faces many problems at present [36,189]. However, the main challenges of PDT are not only limited to light intensity in tissues, tumor hypoxia, and low accumulation efficiency of PSs in tumors [190–192]. Future efforts should be devoted to the following aspects: (1) Developing AIE-based PSs targeting the less-reported organelles, such as the GA or nucleus, is worthy of investigation; (2) developing novel nanoparticle carriers and targeting conjugates to modify AIE-PSs and improve their water solubility, tumor targeting and delivery efficiency [193]; (3) developing AIEgen-based PSs with NIR I or NIR II absorption and Type I PDT abilities to overcome the limited penetration depth and the drug resistance of tumors under hypoxia; (4) the construction of AIE-PSs with both tumor-targeting and organelle-targeting abilities to optimize the therapeutic performance; (5) smart AIEgen-based PSs which show stimuli-responsive abilities and combined therapeutic effects such as PDT, PTT, immunotherapy and sonodynamic therapy [193] are also highly desirable; (6) some tumor cells are resistant to one certain cell death mode, and therefore developing AIE-PSs that can cause multiple cell death pathways are appealing for the effective inhibition of these tumor cells.

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References

1. Sung, H.; Ferlay, J.; Siegel, R.L.; Laversanne, M.; Soerjomataram, I.; Jemal, A.; Bray, F. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA Cancer J. Clin.* **2021**, *71*, 209–249. [[CrossRef](#)] [[PubMed](#)]
2. Jahanban-Esfahlan, R.; de la Guardia, M.; Ahmadi, D.; Yousefi, B. Modulating tumor hypoxia by nanomedicine for effective cancer therapy. *J. Cell. Physiol.* **2018**, *233*, 2019–2031. [[CrossRef](#)] [[PubMed](#)]
3. Sharma, A.; Arambula, J.F.; Koo, S.; Kumar, R.; Singh, H.; Sessler, J.L.; Kim, J.S. Hypoxia-targeted drug delivery. *Chem. Soc. Rev.* **2019**, *48*, 771–813. [[CrossRef](#)] [[PubMed](#)]
4. Shen, Y.; Shuhendler, A.J.; Ye, D.; Xu, J.-J.; Chen, H.-Y. Two-photon excitation nanoparticles for photodynamic therapy. *Chem. Soc. Rev.* **2016**, *45*, 6725–6741. [[CrossRef](#)]
5. Ding, Y.; Wang, Y.; Hu, Q. Recent advances in overcoming barriers to cell-based delivery systems for cancer immunotherapy. *Exploration* **2022**, *2*, 20210106. [[CrossRef](#)]
6. Basak, S.; Gicheru, Y.; Kapoor, A.; Mayer, M.L.; Filizola, M.; Chakrapani, S. Molecular mechanism of setron-mediated inhibition of full-length 5-HT(3A) receptor. *Nat. Commun.* **2019**, *10*, 3225. [[CrossRef](#)]
7. Grabovska, Y.; Mackay, A.; O'Hare, P.; Crosier, S.; Finetti, M.; Schwalbe, E.C.; Pickles, J.C.; Fairchild, A.R.; Avery, A.; Cockle, J.; et al. Pediatric pan-central nervous system tumor analysis of immune-cell infiltration identifies correlates of antitumor immunity. *Nat. Commun.* **2020**, *11*, 4324. [[CrossRef](#)]

8. Li, X.; Lovell, J.F.; Yoon, J.; Chen, X. Clinical development and potential of photothermal and photodynamic therapies for cancer. *Nat. Rev. Clin. Oncol.* **2020**, *17*, 657–674. [\[CrossRef\]](#)
9. Xie, Z.; Fan, T.; An, J.; Choi, W.; Duo, Y.; Ge, Y.; Zhang, B.; Nie, G.; Xie, N.; Zheng, T.; et al. Emerging combination strategies with phototherapy in cancer nanomedicine. *Chem. Soc. Rev.* **2020**, *49*, 8065–8087. [\[CrossRef\]](#)
10. Zhou, Z.; Song, J.; Nie, L.; Chen, X. Reactive oxygen species generating systems meeting challenges of photodynamic cancer therapy. *Chem. Soc. Rev.* **2016**, *45*, 6597–6626. [\[CrossRef\]](#)
11. Li, X.; Kwon, N.; Guo, T.; Liu, Z.; Yoon, J. Innovative Strategies for Hypoxic-Tumor Photodynamic Therapy. *Angew. Chem. Int. Ed.* **2018**, *57*, 11522–11531. [\[CrossRef\]](#) [\[PubMed\]](#)
12. Chen, H.; Wan, Y.; Cui, X.; Li, S.; Lee, C.S. Recent Advances in Hypoxia-Overcoming Strategy of Aggregation-Induced Emission Photosensitizers for Efficient Photodynamic Therapy. *Adv. Healthc. Mater.* **2021**, *10*, e2101607. [\[CrossRef\]](#) [\[PubMed\]](#)
13. Zhang, Y.; Xu, C.; Yang, X.; Pu, K. Photoactivatable Protherapeutic Nanomedicine for Cancer. *Adv. Mater.* **2020**, *32*, 2002661. [\[CrossRef\]](#) [\[PubMed\]](#)
14. Li, J.; Pu, K. Semiconducting Polymer Nanomaterials as Near-Infrared Photoactivatable Protherapeutics for Cancer. *Acc. Chem. Res.* **2020**, *53*, 752–762. [\[CrossRef\]](#) [\[PubMed\]](#)
15. Cui, X.; Lu, G.; Dong, S.; Li, S.; Xiao, Y.; Zhang, J.; Liu, Y.; Meng, X.; Li, F.; Lee, C.-S. Stable π -radical nanoparticles as versatile photosensitizers for effective hypoxia-overcoming photodynamic therapy. *Mater. Horiz.* **2021**, *8*, 571–576. [\[CrossRef\]](#)
16. Dolmans, D.E.J.G.J.; Fukumura, D.; Jain, R.K. Photodynamic therapy for cancer. *Nat. Rev. Cancer* **2003**, *3*, 380–387. [\[CrossRef\]](#)
17. Dichiaro, M.; Prezzavento, O.; Marrazzo, A.; Pittalà, V.; Salerno, L.; Rescifina, A.; Amata, E. Recent advances in drug discovery of phototherapeutic non-porphyrinic anticancer agents. *Eur. J. Med. Chem.* **2017**, *142*, 459–485. [\[CrossRef\]](#)
18. Dang, J.; He, H.; Chen, D.; Yin, L. Manipulating tumor hypoxia toward enhanced photodynamic therapy (PDT). *Biomater. Sci.* **2017**, *5*, 1500–1511. [\[CrossRef\]](#)
19. Ge, J.; Lan, M.; Zhou, B.; Liu, W.; Guo, L.; Wang, H.; Jia, Q.; Niu, G.; Huang, X.; Zhou, H.; et al. A graphene quantum dot photodynamic therapy agent with high singlet oxygen generation. *Nat. Commun.* **2014**, *5*, 4596. [\[CrossRef\]](#)
20. Li, J.; Pu, K. Development of organic semiconducting materials for deep-tissue optical imaging, phototherapy and photoactivation. *Chem. Soc. Rev.* **2019**, *48*, 38–71. [\[CrossRef\]](#)
21. DeRosa, M.C.; Crutchley, R.J. Photosensitized singlet oxygen and its applications. *Coord. Chem. Rev.* **2002**, *233*, 351–371. [\[CrossRef\]](#)
22. Agostinis, P.; Berg, K.; Cengel, K.A.; Foster, T.H.; Girotti, A.W.; Gollnick, S.O.; Hahn, S.M.; Hamblin, M.R.; Juzeniene, A.; Kessel, D.; et al. Photodynamic Therapy of Cancer: An Update. *CA Cancer J. Clin.* **2011**, *61*, 250–281. [\[CrossRef\]](#) [\[PubMed\]](#)
23. Zhang, L.; Li, Y.; Che, W.; Zhu, D.; Li, G.; Xie, Z.; Song, N.; Liu, S.; Tang, B.Z.; Liu, X.; et al. AIE Multinuclear Ir(III) Complexes for Biocompatible Organic Nanoparticles with Highly Enhanced Photodynamic Performance. *Adv. Sci.* **2019**, *6*, 1802050. [\[CrossRef\]](#) [\[PubMed\]](#)
24. Tang, B.Z. Aggregology: Exploration and innovation at aggregate level. *Aggregate* **2020**, *1*, 4–5. [\[CrossRef\]](#)
25. Sagadevan, A.; Hwang, K.C.; Su, M.D. Singlet oxygen-mediated selective C-H bond hydroperoxidation of ethereal hydrocarbons. *Nat. Commun.* **2017**, *8*, 1812. [\[CrossRef\]](#)
26. Fan, W.; Huang, P.; Chen, X. Overcoming the Achilles' heel of photodynamic therapy. *Chem. Soc. Rev.* **2016**, *45*, 6488–6519. [\[CrossRef\]](#)
27. Kulsi, G.; Song, J.M. Sub Cellular Organelles-targeting Photo Dynamic Therapy (PDT). *Mini-Rev. Org. Chem.* **2016**, *13*, 336–348. [\[CrossRef\]](#)
28. Wang, R.; Li, X.; Yoon, J. Organelle-Targeted Photosensitizers for Precision Photodynamic Therapy. *ACS Appl. Mater. Interfaces* **2021**, *13*, 19543–19571. [\[CrossRef\]](#)
29. Gomer, C.J.; Ferrario, A.; Luna, M.; Rucker, N.; Wong, S. Photodynamic therapy: Combined modality approaches targeting the tumor microenvironment. *Lasers Surg. Med.* **2006**, *38*, 516–521. [\[CrossRef\]](#)
30. Olivo, M.; Bhuvaneswari, R.; Lucky, S.S.; Dendukuri, N.; Soo-Ping Thong, P. Targeted Therapy of Cancer Using Photodynamic Therapy in Combination with Multi-faceted Anti-Tumor Modalities. *Pharmaceuticals* **2010**, *3*, 1507–1529. [\[CrossRef\]](#)
31. Shirasu, N.; Nam, S.O.; Kuroki, M. Tumor-targeted Photodynamic Therapy. *Anticancer Res.* **2013**, *33*, 2823–2831. [\[PubMed\]](#)
32. Sharath Kumar, K.S.; Girish, Y.R.; Ashrafizadeh, M.; Mirzaei, S.; Rakesh, K.P.; Hossein Gholami, M.; Zabolian, A.; Hushmandi, K.; Orive, G.; Kadumudi, F.B.; et al. AIE-featured tetraphenylethylene nanoarchitectures in biomedical application: Bioimaging, drug delivery and disease treatment. *Coord. Chem. Rev.* **2021**, *447*, 214135. [\[CrossRef\]](#)
33. Korneev, O.V.; Sakhno, T.V.; Korotkova, I.V. Nanoparticles-based photosensitizers with effect of aggregation-induced emission. *Biopolym. Cell* **2019**, *35*, 249–267. [\[CrossRef\]](#)
34. Hu, F.; Liu, B. Organelle-specific bioprobes based on fluorogens with aggregation-induced emission (AIE) characteristics. *Org. Biomol. Chem.* **2016**, *14*, 9931–9944. [\[CrossRef\]](#)
35. Suman, G.R.; Pandey, M.; Chakravarthy, A.S.J. Review on new horizons of aggregation induced emission: From design to development. *Mater. Chem. Front.* **2021**, *5*, 1541–1584.
36. Dai, J.; Wu, X.; Ding, S.; Lou, X.; Xia, F.; Wang, S.; Hong, Y. Aggregation-Induced Emission Photosensitizers: From Molecular Design to Photodynamic Therapy. *J. Med. Chem.* **2020**, *63*, 1996–2012. [\[CrossRef\]](#)
37. Yi, G.; Hong, S.H.; Son, J.; Yoo, J.; Park, C.; Choi, Y.; Koo, H. Recent advances in nanoparticle carriers for photodynamic therapy. *Quant. Imaging Med. Surg.* **2018**, *8*, 433–443. [\[CrossRef\]](#)

38. Niu, N.; Yu, Y.; Zhang, Z.; Kang, M.; Wang, L.; Zhao, Z.; Wang, D.; Tang, B.Z. A cell membrane-targeting AIE photosensitizer as a necroptosis inducer for boosting cancer theranostics. *Chem. Sci.* **2022**, *13*, 5929–5937. [\[CrossRef\]](#)
39. Wu, M.; Liu, X.; Chen, H.; Duan, Y.; Liu, J.; Pan, Y.; Liu, B. Activation of Pyroptosis by Membrane-Anchoring AIE Photosensitizer Design: New Prospect for Photodynamic Cancer Cell Ablation. *Angew. Chem. Int. Ed.* **2021**, *60*, 9093–9098. [\[CrossRef\]](#)
40. Wang, M.; Wu, M.; Liu, X.; Shao, S.; Huang, J.; Liu, B.; Liang, T. Pyroptosis Remodeling Tumor Microenvironment to Enhance Pancreatic Cancer Immunotherapy Driven by Membrane Anchoring Photosensitizer. *Adv. Sci.* **2022**, *9*, e2202914. [\[CrossRef\]](#)
41. Xu, W.; Lee, M.M.S.; Nie, J.J.; Zhang, Z.; Kwok, R.T.K.; Lam, J.W.Y.; Xu, F.J.; Wang, D.; Tang, B.Z. Three-Pronged Attack by Homologous Far-red/NIR AIEgens to Achieve $1 + 1 + 1 > 3$ Synergistic Enhanced Photodynamic Therapy. *Angew. Chem. Int. Ed.* **2020**, *59*, 9610–9616. [\[CrossRef\]](#) [\[PubMed\]](#)
42. Choromanska, A.; Chwilikowska, A.; Kulbacka, J.; Baczynska, D.; Rembalkowska, N.; Szewczyk, A.; Michel, O.; Gajewska-Naryniecka, A.; Przystupski, D.; Saczko, J. Modifications of Plasma Membrane Organization in Cancer Cells for Targeted Therapy. *Molecules* **2021**, *26*, 1850. [\[CrossRef\]](#) [\[PubMed\]](#)
43. Salazar, G.T.A.; Huang, Z.; Zhang, N.; Zhang, X.-G.; An, Z. Antibody Therapies Targeting Complex Membrane Proteins. *Engineering* **2021**, *7*, 1541–1551. [\[CrossRef\]](#)
44. Yang, N.; Song, S.; Ren, J.; Liu, C.; Li, Z.; Qi, H.; Yu, C. Controlled Aggregation of a Perylene-Derived Probe for Near-Infrared Fluorescence Imaging and Phototherapy. *ACS Appl. Bio Mater.* **2021**, *4*, 5008–5015. [\[CrossRef\]](#) [\[PubMed\]](#)
45. Gao, P.; Pan, W.; Li, N.; Tang, B. Boosting Cancer Therapy with Organelle-Targeted Nanomaterials. *ACS Appl. Mater. Interfaces* **2019**, *11*, 26529–26558. [\[CrossRef\]](#) [\[PubMed\]](#)
46. Chen, J.; Li, S.; Wang, Z.; Pan, Y.; Wei, J.; Lu, S.; Zhang, Q.W.; Wang, L.H.; Wang, R. Synthesis of an AIEgen functionalized cucurbit[7]uril for subcellular bioimaging and synergistic photodynamic therapy and supramolecular chemotherapy. *Chem. Sci.* **2021**, *12*, 7727–7734. [\[CrossRef\]](#) [\[PubMed\]](#)
47. Huang, L.; Qing, D.; Zhao, S.; Wu, X.; Yang, K.; Ren, X.; Zheng, X.; Lan, M.; Ye, J.; Zeng, L.; et al. Acceptor-donor-acceptor structured deep-red AIE photosensitizer: Lysosome-specific targeting, in vivo long-term imaging, and effective photodynamic therapy. *Chem. Eng. J.* **2022**, *430*, 132638. [\[CrossRef\]](#)
48. Huang, W.; Zhang, Y.; Tan, X.; Wang, N.; Wang, J.; He, M.; Peng, J.; Hu, J.; Zhao, Y.; Wang, S. An AIEgen-based photosensitizer for lysosome imaging and photodynamic therapy in tumor. *Sens. Actuators B Chem.* **2021**, *335*, 129698. [\[CrossRef\]](#)
49. Lee, M.M.S.; Yan, D.; Chau, J.H.C.; Park, H.; Ma, C.C.H.; Kwok, R.T.K.; Lam, J.W.Y.; Wang, D.; Tang, B.Z. Highly efficient phototheranostics of macrophage-engulfed Gram-positive bacteria using a NIR luminogen with aggregation-induced emission characteristics. *Biomaterials* **2020**, *261*, 120340. [\[CrossRef\]](#)
50. Liu, X.; He, X.; Zhang, X.; Wang, Y.; Liu, J.; Hao, X.; Zhang, Y.; Yuan, X.A.; Tian, L.; Liu, Z. New Organometallic TetraphenylethyleneIridium (III) Complexes with Antineoplastic Activity. *Chembiochem* **2019**, *20*, 2767–2776. [\[CrossRef\]](#)
51. Meng, F. Aggregation induced emission-active molecules bearing tunable singlet oxygen generation: The different length alkyl chain matters. *Spectrochim. Acta A Mol. Biomol. Spectrosc.* **2021**, *248*, 119233. [\[CrossRef\]](#) [\[PubMed\]](#)
52. Song, S.L.; Zhao, Y.; Kang, M.M.; Zhang, Z.J.; Wu, Q.; Fu, S.; Li, Y.M.; Wen, H.F.; Wang, D.; Tang, B.Z. Side-Chain Engineering of Aggregation-Induced Emission Molecules for Boosting Cancer Phototheranostics. *Adv. Funct. Mater.* **2021**, *31*, 2107545. [\[CrossRef\]](#)
53. Xu, Y.; Zhang, Y.; Li, J.; An, J.; Li, C.; Bai, S.; Sharma, A.; Deng, G.; Kim, J.S.; Sun, Y. NIR-II emissive multifunctional AIEgen with single laser-activated synergistic photodynamic/photothermal therapy of cancers and pathogens. *Biomaterials* **2020**, *259*, 120315. [\[CrossRef\]](#) [\[PubMed\]](#)
54. Yang, M.Q.; Deng, J.R.; Su, H.F.; Gu, S.X.; Zhang, J.; Zhong, A.G.; Wu, F.S. Small organic molecule-based nanoparticles with red/near-infrared aggregation-induced emission for bioimaging and PDT/PTT synergistic therapy. *Mater. Chem. Front.* **2021**, *5*, 406–417. [\[CrossRef\]](#)
55. Yang, Z.; Zhang, Z.; Sun, Y.; Lei, Z.; Wang, D.; Ma, H.; Tang, B.Z. Incorporating spin-orbit coupling promoted functional group into an enhanced electron D-A system: A useful designing concept for fabricating efficient photosensitizer and imaging-guided photodynamic therapy. *Biomaterials* **2021**, *275*, 120934. [\[CrossRef\]](#)
56. Yao, H.M.; Dai, J.; Zhuang, Z.Y.; Yao, J.Y.; Wu, Z.X.; Wang, S.X.; Xia, F.; Zhou, J.; Lou, X.D.; Zhao, Z.J. Red AIE conjugated polyelectrolytes for long-term tracing and image-guided photodynamic therapy of tumors. *Sci. China-Chem.* **2020**, *63*, 1815–1824. [\[CrossRef\]](#)
57. Zhuang, J.; Yang, H.; Li, Y.; Wang, B.; Li, N.; Zhao, N. Efficient photosensitizers with aggregation-induced emission characteristics for lysosome- and Gram-positive bacteria-targeted photodynamic therapy. *Chem. Commun.* **2020**, *56*, 2630–2633. [\[CrossRef\]](#)
58. Zou, H.; Zhang, J.; Wu, C.; He, B.; Hu, Y.; Sung, H.H.Y.; Kwok, R.T.K.; Lam, J.W.Y.; Zheng, L.; Tang, B.Z. Making Aggregation-Induced Emission Luminogen More Valuable by Gold: Enhancing Anticancer Efficacy by Suppressing Thioredoxin Reductase Activity. *ACS Nano* **2021**, *15*, 9176–9185. [\[CrossRef\]](#)
59. Wen, H.; Zhang, Z.; Kang, M.; Li, H.; Xu, W.; Guo, H.; Li, Y.; Tan, Y.; Wen, Z.; Wu, Q.; et al. One-for-all phototheranostics: Single component AIE dots as multi-modality theranostic agent for fluorescence-photoacoustic imaging-guided synergistic cancer therapy. *Biomaterials* **2021**, *274*, 120892. [\[CrossRef\]](#)
60. Zhang, Z.; Xu, W.; Xiao, P.; Kang, M.; Yan, D.; Wen, H.; Song, N.; Wang, D.; Tang, B.Z. Molecular Engineering of High-Performance Aggregation-Induced Emission Photosensitizers to Boost Cancer Theranostics Mediated by Acid-Triggered Nucleus-Targeted Nanovectors. *ACS Nano* **2021**, *15*, 10689–10699. [\[CrossRef\]](#)

61. Kang, M.; Zhang, Z.; Xu, W.; Wen, H.; Zhu, W.; Wu, Q.; Wu, H.; Gong, J.; Wang, Z.; Wang, D.; et al. Good Steel Used in the Blade: Well-Tailored Type-I Photosensitizers with Aggregation-Induced Emission Characteristics for Precise Nuclear Targeting Photodynamic Therapy. *Adv. Sci.* **2021**, *8*, 2100524. [[CrossRef](#)] [[PubMed](#)]
62. Dai, Y.; He, F.; Ji, H.; Zhao, X.; Misal, S.; Qi, Z. Dual-Functional NIR AIEgens for High-Fidelity Imaging of Lysosomes in Cells and Photodynamic Therapy. *ACS Sens.* **2020**, *5*, 225–233. [[CrossRef](#)] [[PubMed](#)]
63. Gao, Y.; Zheng, Q.C.; Xu, S.; Yuan, Y.; Cheng, X.; Jiang, S.; Kenry; Yu, Q.; Song, Z.; Liu, B.; et al. Theranostic Nanodots with Aggregation-Induced Emission Characteristic for Targeted and Image-Guided Photodynamic Therapy of Hepatocellular Carcinoma. *Theranostics* **2019**, *9*, 1264–1279. [[CrossRef](#)] [[PubMed](#)]
64. Liao, Y.; Wang, R.; Wang, S.; Xie, Y.; Chen, H.; Huang, R.; Shao, L.; Zhu, Q.; Liu, Y. Highly Efficient Multifunctional Organic Photosensitizer with Aggregation-Induced Emission for In Vivo Bioimaging and Photodynamic Therapy. *ACS Appl. Mater. Interfaces* **2021**, *13*, 54783–54793. [[CrossRef](#)]
65. Shao, L.; Pan, Y.; Hua, B.; Xu, S.; Yu, G.; Wang, M.; Liu, B.; Huang, F. Constructing Adaptive Photosensitizers via Supramolecular Modification Based on Pillararene Host-Guest Interactions. *Angew. Chem. Int. Ed.* **2020**, *59*, 11779–11783. [[CrossRef](#)]
66. Wang, S.W.; Chen, H.; Liu, J.; Chen, C.J.; Liu, B. NIR-II Light Activated Photosensitizer with Aggregation-Induced Emission for Precise and Efficient Two-Photon Photodynamic Cancer Cell Ablation. *Adv. Funct. Mater.* **2020**, *30*, 2002546. [[CrossRef](#)]
67. Wang, X.; Tong, J.; He, Z.; Yang, X.; Meng, F.; Liang, H.; Zhang, X.; Luo, L. Paclitaxel-Potentiated Photodynamic Theranostics for Synergistic Tumor Ablation and Precise Anticancer Efficacy Monitoring. *ACS Appl. Mater. Interfaces* **2020**, *12*, 5476–5487. [[CrossRef](#)]
68. Xu, J.; Zheng, Q.; Cheng, X.; Hu, S.; Zhang, C.; Zhou, X.; Sun, P.; Wang, W.; Su, Z.; Zou, T.; et al. Chemo-photodynamic therapy with light-triggered disassembly of theranostic nanoplatfrom in combination with checkpoint blockade for immunotherapy of hepatocellular carcinoma. *J. Nanobiotechnol.* **2021**, *19*, 355. [[CrossRef](#)]
69. Yi, X.; Hu, J.J.; Dai, J.; Lou, X.; Zhao, Z.; Xia, F.; Tang, B.Z. Self-Guiding Polymeric Prodrug Micelles with Two Aggregation-Induced Emission Photosensitizers for Enhanced Chemo-Photodynamic Therapy. *ACS Nano* **2021**, *15*, 3026–3037. [[CrossRef](#)]
70. Zhang, Z.; Xu, W.; Kang, M.; Wen, H.; Guo, H.; Zhang, P.; Xi, L.; Li, K.; Wang, L.; Wang, D.; et al. An All-Round Athlete on the Track of Phototheranostics: Subtly Regulating the Balance between Radiative and Nonradiative Decays for Multimodal Imaging-Guided Synergistic Therapy. *Adv. Mater.* **2020**, *32*, e2003210. [[CrossRef](#)]
71. Smith, R.A.J.; Hartley, R.C.; Cochemé, H.M.; Murphy, M.P. Mitochondrial pharmacology. *Trends Pharmacol. Sci.* **2012**, *33*, 341–352. [[CrossRef](#)] [[PubMed](#)]
72. Rajendran, L.; Knölker, H.-J.; Simons, K. Subcellular targeting strategies for drug design and delivery. *Nat. Rev. Drug Discov.* **2010**, *9*, 29–42. [[CrossRef](#)] [[PubMed](#)]
73. Wallace, D.C. Mitochondria and cancer. *Nat. Rev. Cancer* **2012**, *12*, 685–698. [[CrossRef](#)] [[PubMed](#)]
74. Huang, H.; Dong, C.; Chang, M.; Ding, L.; Chen, L.; Feng, W.; Chen, Y. Mitochondria-specific nanocatalysts for chemotherapy-augmented sequential chemoreactive tumor therapy. *Exploration* **2021**, *1*, 50–60. [[CrossRef](#)]
75. Zheng, Z.; Liu, H.; Zhai, S.; Zhang, H.; Shan, G.; Kwok, R.T.K.; Ma, C.; Sung, H.H.Y.; Williams, I.D.; Lam, J.W.Y.; et al. Highly efficient singlet oxygen generation, two-photon photodynamic therapy and melanoma ablation by rationally designed mitochondria-specific near-infrared AIEgens. *Chem. Sci.* **2020**, *11*, 2494–2503. [[CrossRef](#)]
76. Wei, D.; Chen, Y.; Huang, Y.; Li, P.; Zhao, Y.; Zhang, X.; Wan, J.; Yin, X.; Liu, T.; Yin, J.; et al. NIR-light triggered dual-cascade targeting core-shell nanoparticles enhanced photodynamic therapy and immunotherapy. *Nano Today* **2021**, *41*, 101288. [[CrossRef](#)]
77. Zhang, T.; Zhang, J.; Wang, F.-B.; Cao, H.; Zhu, D.; Chen, X.; Xu, C.; Yang, X.; Huang, W.; Wang, Z.; et al. Mitochondria-Targeting Phototheranostics by Aggregation-Induced NIR-II Emission Luminogens: Modulating Intramolecular Motion by Electron Acceptor Engineering for Multi-Modal Synergistic Therapy. *Adv. Funct. Mater.* **2022**, *32*, 2110526. [[CrossRef](#)]
78. Yu, Y.; Wu, S.; Zhang, L.; Xu, S.; Dai, C.; Gan, S.; Xie, G.; Feng, G.; Tang, B.Z. Cationization to boost both type I and type II ROS generation for photodynamic therapy. *Biomaterials* **2022**, *280*, 121255. [[CrossRef](#)]
79. Liu, P.; Ren, F.; Son, S.; Ji, M.S.; Li, P.; Cai, Z.; Shi, J.; Liu, Y.; Dong, Y.; Kim, J.S. Mitochondrial targeted AIEgen phototheranostics for bypassing immune barrier via encumbering mitochondria functions. *Biomaterials* **2022**, *283*, 121409. [[CrossRef](#)]
80. Yan, S.; Sun, P.; Niu, N.; Zhang, Z.; Xu, W.; Zhao, S.; Wang, L.; Wang, D.; Tang, B.Z. Surfactant-Inspired Coassembly Strategy to Integrate Aggregation-Induced Emission Photosensitizer with Organosilica Nanoparticles for Efficient Theranostics. *Adv. Funct. Mater.* **2022**, *32*, 2200503. [[CrossRef](#)]
81. He, B.; Huang, J.; Zhang, J.; Sung, H.H.Y.; Lam, J.W.Y.; Zhang, Z.; Yan, S.; Wang, D.; Zhang, J.; Tang, B.Z. Novel Quinolizine AIE System: Visualization of Molecular Motion and Elaborate Tailoring for Biological Application. *Angew. Chem. Int. Ed.* **2022**, *61*, e202117709.
82. Pandey, N.K.; Xiong, W.; Wang, L.; Chen, W.; Bui, B.; Yang, J.; Amador, E.; Chen, M.; Xing, C.; Athavale, A.A.; et al. Aggregation-induced emission luminogens for highly effective microwave dynamic therapy. *Bioact. Mater.* **2022**, *7*, 112–125. [[CrossRef](#)] [[PubMed](#)]
83. Zhang, Y.; Huang, W.; Tan, X.; Wang, J.; Zhao, Y.; Hu, J.; Wang, S. A mitochondria-targeted dual-functional aggregation-induced emission luminogen for intracellular mitochondrial imaging and photodynamic therapy. *Biomater. Sci.* **2021**, *9*, 1232–1236. [[CrossRef](#)] [[PubMed](#)]
84. Duan, C.; Hu, J.J.; Liu, R.; Dai, J.; Duan, M.; Yuan, L.; Xia, F.; Lou, X. Spatial Order of Functional Modules Enabling Diverse Intracellular Performance of Fluorescent Probes. *Angew. Chem. Int. Ed.* **2021**, *60*, 18280–18288. [[CrossRef](#)]

85. Cao, S.; Xia, Y.; Shao, J.; Guo, B.; Dong, Y.; Pijpers, I.A.B.; Zhong, Z.; Meng, F.; Abdelmohsen, L.; Williams, D.S.; et al. Biodegradable Polymersomes with Structure Inherent Fluorescence and Targeting Capacity for Enhanced Photo-Dynamic Therapy. *Angew. Chem. Int. Ed.* **2021**, *60*, 17629–17637. [\[CrossRef\]](#) [\[PubMed\]](#)
86. Wang, L.; Huang, Y.; Yu, Y.; Zhong, H.; Xiao, H.; Zhang, G.; Zhang, D. Photosensitizer with High Efficiency Generated in Cells via Light-Induced Self-Oligomerization of 4,6-Dibromothieno[3,4-b]thiophene Compound Entailing a Triphenyl Phosphonium Group. *Adv. Healthc. Mater.* **2021**, *10*, e2100896. [\[CrossRef\]](#) [\[PubMed\]](#)
87. He, Z.; Gao, Y.; Zhang, H.; Xue, Y.; Meng, F.; Luo, L. Mitochondrion-Anchored Photosensitizer with Near Infrared-I Aggregation-Induced Emission for Near Infrared-II Two-Photon Photodynamic Therapy. *Adv. Healthc. Mater.* **2021**, *10*, e2101056. [\[CrossRef\]](#)
88. Yuan, G.; Lv, C.; Liang, J.; Zhong, X.; Li, Y.; He, J.; Zhao, A.; Li, L.; Shao, Y.; Zhang, X.; et al. Molecular Engineering of Efficient Singlet Oxygen Generators with Near-Infrared AIE Features for Mitochondrial Targeted Photodynamic Therapy. *Adv. Funct. Mater.* **2021**, *31*, 2104026. [\[CrossRef\]](#)
89. Song, N.; Zhang, Z.; Liu, P.; Dai, D.; Chen, C.; Li, Y.; Wang, L.; Han, T.; Yang, Y.W.; Wang, D.; et al. Pillar[5]arene-Modified Gold Nanorods as Nanocarriers for Multi-Modal Imaging-Guided Synergistic Photodynamic-Photothermal Therapy. *Adv. Funct. Mater.* **2021**, *31*, 2009924. [\[CrossRef\]](#)
90. Li, Y.; Zhuang, J.; Lu, Y.; Li, N.; Gu, M.; Xia, J.; Zhao, N.; Tang, B.Z. High-Performance Near-Infrared Aggregation-Induced Emission Luminogen with Mitophagy Regulating Capability for Multimodal Cancer Theranostics. *ACS Nano* **2021**, *15*, 20453–20465. [\[CrossRef\]](#)
91. Zhou, C.; Peng, C.; Shi, C.; Jiang, M.; Chau, J.H.C.; Liu, Z.; Bai, H.; Kwok, R.T.K.; Lam, J.W.Y.; Shi, Y.; et al. Mitochondria-Specific Aggregation-Induced Emission Luminogens for Selective Photodynamic Killing of Fungi and Efficacious Treatment of Keratitis. *ACS Nano* **2021**, *15*, 12129–12139. [\[CrossRef\]](#) [\[PubMed\]](#)
92. Zhang, L.; Wang, J.L.; Ba, X.X.; Hua, S.Y.; Jiang, P.; Jiang, F.L.; Liu, Y. Multifunction in One Molecule: Mitochondrial Imaging and Photothermal & Photodynamic Cytotoxicity of Fast-Response Near-Infrared Fluorescent Probes with Aggregation-Induced Emission Characteristics. *ACS Appl. Mater. Interfaces* **2021**, *13*, 7945–7954. [\[PubMed\]](#)
93. Zhu, D.; Zheng, Z.; Luo, G.; Suo, M.; Li, X.; Duo, Y.; Tang, B.Z. Single injection and multiple treatments: An injectable nanozyme hydrogel as AIEgen reservoir and release controller for efficient tumor therapy. *Nano Today* **2021**, *37*, 101091. [\[CrossRef\]](#)
94. Zhao, X.; Dai, Y.; Ma, F.; Misal, S.; Hasrat, K.; Zhu, H.; Qi, Z. Molecular engineering to accelerate cancer cell discrimination and boost AIE-active type I photosensitizer for photodynamic therapy under hypoxia. *Chem. Eng. J.* **2021**, *410*, 128133. [\[CrossRef\]](#)
95. Feng, H.T.; Li, Y.; Duan, X.; Wang, X.; Qi, C.; Lam, J.W.Y.; Ding, D.; Tang, B.Z. Substitution Activated Precise Phototheranostics through Supramolecular Assembly of AIEgen and Calixarene. *J. Am. Chem. Soc.* **2020**, *142*, 15966–15974. [\[CrossRef\]](#) [\[PubMed\]](#)
96. Zhu, D.; Duo, Y.; Suo, M.; Zhao, Y.; Xia, L.; Zheng, Z.; Li, Y.; Tang, B.Z. Tumor-Exocytosed Exosome/Aggregation-Induced Emission Luminogen Hybrid Nanovesicles Facilitate Efficient Tumor Penetration and Photodynamic Therapy. *Angew. Chem. Int. Ed.* **2020**, *59*, 13836–13843. [\[CrossRef\]](#)
97. Huang, J.; He, B.; Zhang, Z.; Li, Y.; Kang, M.; Wang, Y.; Li, K.; Wang, D.; Tang, B.Z. Aggregation-Induced Emission Luminogens Married to 2D Black Phosphorus Nanosheets for Highly Efficient Multimodal Theranostics. *Adv. Mater.* **2020**, *32*, e2003382. [\[CrossRef\]](#)
98. Wang, J.; Zhu, X.; Zhang, J.; Wang, H.; Liu, G.; Bu, Y.; Yu, J.; Tian, Y.; Zhou, H. AIE-Based Theranostic Agent: In Situ Tracking Mitophagy Prior to Late Apoptosis to Guide the Photodynamic Therapy. *ACS Appl. Mater. Interfaces* **2020**, *12*, 1988–1996. [\[CrossRef\]](#)
99. Zhao, X.; Fan, Z.; Qiao, Y.; Chen, Y.; Wang, S.; Yue, X.; Shen, T.; Liu, W.; Yang, J.; Gao, H.; et al. AIEgens Conjugation Improves the Photothermal Efficacy and Near-Infrared Imaging of Heptamethine Cyanine IR-780. *ACS Appl. Mater. Interfaces* **2020**, *12*, 16114–16124. [\[CrossRef\]](#)
100. Liu, J.; Liu, X.; Wu, M.; Qi, G.; Liu, B. Engineering Living Mitochondria with AIE Photosensitizer for Synergistic Cancer Cell Ablation. *Nano Lett.* **2020**, *20*, 7438–7445. [\[CrossRef\]](#)
101. Guo, B.; Wu, M.; Shi, Q.; Dai, T.J.; Xu, S.D.; Jiang, J.W.; Liu, B. All-in-One Molecular Aggregation-Induced Emission Theranostics: Fluorescence Image Guided and Mitochondria Targeted Chemo-and Photodynamic Cancer Cell Ablation. *Chem. Mater.* **2020**, *32*, 4681–4691. [\[CrossRef\]](#)
102. He, X.; Situ, B.; Gao, M.; Guan, S.; He, B.; Ge, X.; Li, S.; Tao, M.; Zou, H.; Tang, B.Z.; et al. Stereotactic Photodynamic Therapy Using a Two-Photon AIE Photosensitizer. *Small* **2019**, *15*, e1905080. [\[CrossRef\]](#) [\[PubMed\]](#)
103. Liu, Z.; Zou, H.; Zhao, Z.; Zhang, P.; Shan, G.G.; Kwok, R.T.K.; Lam, J.W.Y.; Zheng, L.; Tang, B.Z. Tuning Organelle Specificity and Photodynamic Therapy Efficiency by Molecular Function Design. *ACS Nano* **2019**, *13*, 11283–11293. [\[CrossRef\]](#) [\[PubMed\]](#)
104. Chen, Y.; Ai, W.; Guo, X.; Li, Y.; Ma, Y.; Chen, L.; Zhang, H.; Wang, T.; Zhang, X.; Wang, Z. Mitochondria-Targeted Polydopamine Nanocomposite with AIE Photosensitizer for Image-Guided Photodynamic and Photothermal Tumor Ablation. *Small* **2019**, *15*, e1902352. [\[CrossRef\]](#)
105. Yao, D.; Yang, S.; Wang, Y.; Bian, K.; Yang, W.; Wang, D.; Zhang, B. An ALP-activatable and mitochondria-targeted probe for prostate cancer-specific bimodal imaging and aggregation-enhanced photothermal therapy. *Nanoscale* **2019**, *11*, 6307–6314. [\[CrossRef\]](#)
106. Chen, C.; Ni, X.; Jia, S.; Liang, Y.; Wu, X.; Kong, D.; Ding, D. Massively Evoking Immunogenic Cell Death by Focused Mitochondrial Oxidative Stress using an AIE Luminogen with a Twisted Molecular Structure. *Adv. Mater.* **2019**, *31*, e1904914. [\[CrossRef\]](#)

107. Zhang, J.; Wang, Q.; Guo, Z.Q.; Zhang, S.Z.; Yan, C.X.; Tian, H.; Zhu, W.H. High-Fidelity Trapping of Spatial-Temporal Mitochondria with Rational Design of Aggregation-Induced Emission Probes. *Adv. Funct. Mater.* **2019**, *29*, 1808153. [\[CrossRef\]](#)
108. Zhuang, W.; Yang, L.; Ma, B.; Kong, Q.; Li, G.; Wang, Y.; Tang, B.Z. Multifunctional Two-Photon AIE Luminogens for Highly Mitochondria-Specific Bioimaging and Efficient Photodynamic Therapy. *ACS Appl. Mater. Interfaces* **2019**, *11*, 20715–20724. [\[CrossRef\]](#)
109. Zhao, N.; Li, P.; Zhuang, J.; Liu, Y.; Xiao, Y.; Qin, R.; Li, N. Aggregation-Induced Emission Luminogens with the Capability of Wide Color Tuning, Mitochondrial and Bacterial Imaging, and Photodynamic Anticancer and Antibacterial Therapy. *ACS Appl. Mater. Interfaces* **2019**, *11*, 11227–11237. [\[CrossRef\]](#)
110. Lin, F.; Bao, Y.-W.; Wu, F.-G. Improving the Phototherapeutic Efficiencies of Molecular and Nanoscale Materials by Targeting Mitochondria. *Molecules* **2018**, *23*, 3016. [\[CrossRef\]](#)
111. Kagan, V.E.; Wipf, P.; Stoyanovsky, D.; Greenberger, J.S.; Borisenko, G.; Belikova, N.A.; Yanamala, N.; Arias, A.K.S.; Tunekar, M.A.; Jiang, J.; et al. Mitochondrial targeting of electron scavenging antioxidants: Regulation of selective oxidation vs random chain reactions. *Adv. Drug Deliv. Rev.* **2009**, *61*, 1375–1385. [\[CrossRef\]](#) [\[PubMed\]](#)
112. Heller, A.; Brockhoff, G.; Goepferich, A. Targeting drugs to mitochondria. *Eur. J. Pharm. Biopharm.* **2012**, *82*, 1–18. [\[CrossRef\]](#) [\[PubMed\]](#)
113. Allemailem, K.S.; Almatroudi, A.; Alsahli, M.A.; Aljaghawani, A.; El-Kady, A.M.; Rahmani, A.H.; Khan, A.A. Novel Strategies for Disrupting Cancer-Cell Functions with Mitochondria-Targeted Antitumor Drug-Loaded Nanoformulations. *Int. J. Nanomed.* **2021**, *16*, 3907–3936. [\[CrossRef\]](#) [\[PubMed\]](#)
114. Xu, X.; Deng, G.; Sun, Z.; Luo, Y.; Liu, J.; Yu, X.; Zhao, Y.; Gong, P.; Liu, G.; Zhang, P.; et al. A Biomimetic Aggregation-Induced Emission Photosensitizer with Antigen-Presenting and Hitchhiking Function for Lipid Droplet Targeted Photodynamic Immunotherapy. *Adv. Mater.* **2021**, *33*, e2102322. [\[CrossRef\]](#)
115. Jiang, G.Y.; Li, C.B.; Liu, X.; Chen, Q.Q.; Li, X.K.; Gu, X.G.; Zhang, P.F.; Lai, Q.F.; Wang, J.G. Lipid Droplet-Targetable Fluorescence Guided Photodynamic Therapy of Cancer Cells with an Activatable AIE-Active Fluorescent Probe for Hydrogen Peroxide. *Adv. Opt. Mater.* **2020**, *8*, 2001119. [\[CrossRef\]](#)
116. Zhang, F.; Liu, Y.; Yang, B.; Guan, P.; Chai, J.; Wen, G.; Liu, B. Tunable NIR AIE-active optical materials for lipid droplet imaging in typical model organisms and photodynamic therapy. *J. Mater. Chem. B* **2021**, *9*, 2417–2427. [\[CrossRef\]](#)
117. Sun, Z.; Shi, S.; Guan, P.; Liu, B. Construction of heteroaryl-bridged NIR AIEgens for specific imaging of lipid droplets and its application in photodynamic therapy. *Spectrochim. Acta A Mol. Biomol. Spectrosc.* **2022**, *272*, 120946. [\[CrossRef\]](#)
118. Zhang, F.; Liu, Y.M.; Yang, B.S.; Wen, G.M.; Liu, B. Near-infrared AIEgens for lipid droplets imaging in corpus adiposum or trachea of *Locusta migratoria* and its application in photodynamic therapy. *Sens. Actuators B Chem.* **2020**, *322*, 128589. [\[CrossRef\]](#)
119. Dai, Y.P.; Zhao, X.X.; Ji, H.F.; Zhang, D.D.; Zhang, P.; Xue, K.; Misal, S.; Zhu, H.Y.; Qi, Z.J. Multifunctional aggregation-induced emission nanoparticle for high-fidelity imaging of lipid droplets in living cells and its application in photodynamic therapy. *Chem. Eng. J.* **2021**, *410*, 128186. [\[CrossRef\]](#)
120. Li, Y.; Peng, Q.; Li, S.; Cai, Y.; Zhang, B.; Sun, K.; Ma, J.; Yang, C.; Hou, H.; Su, H.; et al. A multifunctional quinoxalin-based AIEgen used for fluorescent thermo-sensing and image-guided photodynamic therapy. *Sens. Actuators B Chem.* **2019**, *301*, 127139. [\[CrossRef\]](#)
121. Zhang, C.; Liu, P. The New Face of the Lipid Droplet: Lipid Droplet Proteins. *Proteomics* **2019**, *19*, 1700223. [\[CrossRef\]](#) [\[PubMed\]](#)
122. Hashemi, H.F.; Goodman, J.M. The life cycle of lipid droplets. *Curr. Opin. Cell Biol.* **2015**, *33*, 119–124. [\[CrossRef\]](#) [\[PubMed\]](#)
123. Bersuker, K.; Olzmann, J.A. Establishing the lipid droplet proteome: Mechanisms of lipid droplet protein targeting and degradation. *Biochim. Biophys. Acta-Mol. Cell Biol. Lipids* **2017**, *1862*, 1166–1177. [\[CrossRef\]](#) [\[PubMed\]](#)
124. Li, Y.; Zhang, R.; Wan, Q.; Hu, R.; Ma, Y.; Wang, Z.; Hou, J.; Zhang, W.; Tang, B.Z. Trojan Horse-Like Nano-AIE Aggregates Based on Homologous Targeting Strategy and Their Photodynamic Therapy in Anticancer Application. *Adv. Sci.* **2021**, *8*, 2102561. [\[CrossRef\]](#) [\[PubMed\]](#)
125. Sun, Z.; Liu, Y.; Guan, P.; Yang, B.; Liu, B. Near-infrared dual-functional AIEgens for lipid droplets imaging in multispecies and photodynamic therapy. *Dyes Pigm.* **2021**, *185*, 108884. [\[CrossRef\]](#)
126. Pool, M.R. Targeting of Proteins for Translocation at the Endoplasmic Reticulum. *Int. J. Mol. Sci.* **2022**, *23*, 3773. [\[CrossRef\]](#)
127. Ding, G.; Tong, J.; Gong, J.; Wang, Z.; Su, Z.; Liu, L.; Han, X.; Wang, J.; Zhang, L.; Wang, X.; et al. Molecular engineering to achieve AIE-active photosensitizers with NIR emission and rapid ROS generation efficiency. *J. Mater. Chem. B* **2022**, *10*, 5272–5278. [\[CrossRef\]](#)
128. Zhuang, Z.; Dai, J.; Yu, M.; Li, J.; Shen, P.; Hu, R.; Lou, X.; Zhao, Z.; Tang, B.Z. Type I photosensitizers based on phosphindole oxide for photodynamic therapy: Apoptosis and autophagy induced by endoplasmic reticulum stress. *Chem. Sci.* **2020**, *11*, 3405–3417. [\[CrossRef\]](#)
129. Liu, M.; Chen, Y.; Guo, Y.; Yuan, H.; Cui, T.; Yao, S.; Jin, S.; Fan, H.; Wang, C.; Xie, R.; et al. Golgi apparatus-targeted aggregation-induced emission luminogens for effective cancer photodynamic therapy. *Nat. Commun.* **2022**, *13*, 2179. [\[CrossRef\]](#)
130. Wang, K.N.; Liu, L.Y.; Mao, D.; Hou, M.X.; Tan, C.P.; Mao, Z.W.; Liu, B. A Nuclear-Targeted AIE Photosensitizer for Enzyme Inhibition and Photosensitization in Cancer Cell Ablation. *Angew. Chem. Int. Ed.* **2022**, *61*, e202114600.
131. Zhang, T.; Li, Y.; Zheng, Z.; Ye, R.; Zhang, Y.; Kwok, R.T.K.; Lam, J.W.Y.; Tang, B.Z. In Situ Monitoring Apoptosis Process by a Self-Reporting Photosensitizer. *J. Am. Chem. Soc.* **2019**, *141*, 5612–5616. [\[CrossRef\]](#) [\[PubMed\]](#)

132. Wan, L.-Y.; Yuan, W.-F.; Ai, W.-B.; Ai, Y.-W.; Wang, J.-J.; Chu, L.-Y.; Zhang, Y.-Q.; Wu, J.-F. An exploration of aptamer internalization mechanisms and their applications in drug delivery. *Expert Opin. Drug Deliv.* **2019**, *16*, 207–218. [[CrossRef](#)] [[PubMed](#)]
133. Shrivastava, G.; Bakshi, H.A.; Aljabali, A.A.; Mishra, V.; Hakkim, F.L.; Charbe, N.B.; Kesharwani, P.; Chellappan, D.K.; Dua, K.; Tambuwala, M.M. Nucleic Acid Aptamers as a Potential Nucleus Targeted Drug Delivery System. *Curr. Drug Deliv.* **2020**, *17*, 101–111. [[CrossRef](#)] [[PubMed](#)]
134. Shigdar, S.; Schrand, B.; Giangrande, P.H.; de Franciscis, V. Aptamers: Cutting edge of cancer therapies. *Mol. Ther.* **2021**, *29*, 2396–2411. [[CrossRef](#)]
135. Kau, T.R.; Silver, P.A. Nuclear transport as a target for cell growth. *Drug Discov. Today* **2003**, *8*, 78–85. [[CrossRef](#)]
136. Huang, Y.; Liu, D.E.; An, J.; Liu, B.; Sun, L.; Fu, H.; Yan, S.; Sun, W.; Gao, H. Reactive Oxygen Species Self-Sufficient Multifunctional Nanoplatfor for Synergistic Chemo-Photodynamic Therapy with Red/Near-Infrared Dual-Imaging. *ACS Appl. Bio Mater.* **2020**, *3*, 9135–9144. [[CrossRef](#)]
137. Chai, C.; Zhou, T.; Zhu, J.; Tang, Y.; Xiong, J.; Min, X.; Qin, Q.; Li, M.; Zhao, N.; Wan, C. Multiple Light-Activated Photodynamic Therapy of Tetraphenylethylene Derivative with AIE Characteristics for Hepatocellular Carcinoma via Dual-Organelles Targeting. *Pharmaceutics* **2022**, *14*, 459. [[CrossRef](#)]
138. Wan, Q.; Zhang, R.; Zhuang, Z.; Li, Y.; Huang, Y.; Wang, Z.; Zhang, W.; Hou, J.; Tang, B.Z. Molecular Engineering to Boost AIE-Active Free Radical Photogenerators and Enable High-Performance Photodynamic Therapy under Hypoxia. *Adv. Funct. Mater.* **2020**, *30*, 2002057. [[CrossRef](#)]
139. Feng, G.X.; Wang, C.; Chen, C.J.; Pan, Y.T.; Wu, M.; Wang, Y.B.; Liu, J.; Liu, B. Modulating Cell Specificity and Subcellular Localization by Molecular Charges and Lipophilicity. *Chem. Mater.* **2020**, *32*, 10383–10393. [[CrossRef](#)]
140. Ni, J.S.; Lee, M.M.S.; Zhang, P.; Gui, C.; Chen, Y.; Wang, D.; Yu, Z.Q.; Kwok, R.T.K.; Lam, J.W.Y.; Tang, B.Z. SwissKnife-Inspired Multifunctional Fluorescence Probes for Cellular Organelle Targeting Based on Simple AIEgens. *Anal. Chem.* **2019**, *91*, 2169–2176. [[CrossRef](#)]
141. Chen, K.Q.; Zhang, R.Y.; Wang, Z.M.; Zhang, W.J.; Tang, B.Z. Structural Modification Orientated Multifunctional AIE Fluorescence Probes: Organelles Imaging and Effective Photosensitizer for Photodynamic Therapy. *Adv. Opt. Mater.* **2020**, *8*, 1901433. [[CrossRef](#)]
142. Madathiparambil Visalakshan, R.; González García, L.E.; Benzigar, M.R.; Ghazaryan, A.; Simon, J.; Mierczynska-Vasilev, A.; Michl, T.D.; Vinu, A.; Mailänder, V.; Morsbach, S.; et al. The Influence of Nanoparticle Shape on Protein Corona Formation. *Small* **2020**, *16*, 2000285. [[CrossRef](#)] [[PubMed](#)]
143. Koo, H.; Huh, M.S.; Sun, I.-C.; Yuk, S.H.; Choi, K.; Kim, K.; Kwon, I.C. In Vivo Targeted Delivery of Nanoparticles for Theranosis. *Acc. Chem. Res.* **2011**, *44*, 1018–1028. [[CrossRef](#)] [[PubMed](#)]
144. Lee, D.; Kwon, S.; Jang, S.-Y.; Park, E.; Lee, Y.; Koo, H. Overcoming the obstacles of current photodynamic therapy in tumors using nanoparticles. *Bioact. Mater.* **2022**, *8*, 20–34. [[CrossRef](#)] [[PubMed](#)]
145. Danhier, F. To exploit the tumor microenvironment: Since the EPR effect fails in the clinic, what is the future of nanomedicine? *J. Control. Release* **2016**, *244*, 108–121. [[CrossRef](#)]
146. Dutta, B.; Barick, K.C.; Hassan, P.A. Recent advances in active targeting of nanomaterials for anticancer drug delivery. *Adv. Colloid Interface Sci.* **2021**, *296*, 102509. [[CrossRef](#)]
147. Jain, R.K.; Stylianopoulos, T. Delivering nanomedicine to solid tumors. *Nat. Rev. Clin. Oncol.* **2010**, *7*, 653–664. [[CrossRef](#)]
148. Son, J.; Yi, G.; Yoo, J.; Park, C.; Koo, H.; Choi, H.S. Light-responsive nanomedicine for biophotonic imaging and targeted therapy. *Adv. Drug Deliv. Rev.* **2019**, *138*, 133–147. [[CrossRef](#)]
149. Byrne, J.D.; Betancourt, T.; Brannon-Peppas, L. Active targeting schemes for nanoparticle systems in cancer therapeutics. *Adv. Drug Deliv. Rev.* **2008**, *60*, 1615–1626. [[CrossRef](#)]
150. Chen, F.; Hableel, G.; Zhao, E.R.; Jokerst, J.V. Multifunctional nanomedicine with silica: Role of silica in nanoparticles for theranostic, imaging, and drug monitoring. *J. Colloid Interface Sci.* **2018**, *521*, 261–279. [[CrossRef](#)]
151. Oh, J.Y.; Kim, H.S.; Palanikumar, L.; Go, E.M.; Jana, B.; Park, S.A.; Kim, H.Y.; Kim, K.; Seo, J.K.; Kwak, S.K.; et al. Cloaking nanoparticles with protein corona shield for targeted drug delivery. *Nat. Commun.* **2018**, *9*, 4548. [[CrossRef](#)] [[PubMed](#)]
152. Ma, Y.; Mou, Q.; Wang, D.; Zhu, X.; Yan, D. Dendritic Polymers for Theranostics. *Theranostics* **2016**, *6*, 930–947. [[CrossRef](#)] [[PubMed](#)]
153. Gong, H.; Chao, Y.; Xiang, J.; Han, X.; Song, G.; Feng, L.; Liu, J.; Yang, G.; Chen, Q.; Liu, Z. Hyaluronidase to Enhance Nanoparticle-Based Photodynamic Tumor Therapy. *Nano Lett.* **2016**, *16*, 2512–2521. [[CrossRef](#)] [[PubMed](#)]
154. Golombek, S.K.; May, J.N.; Theek, B.; Appold, L.; Drude, N.; Kiessling, F.; Lammers, T. Tumor targeting via EPR: Strategies to enhance patient responses. *Adv. Drug. Deliv. Rev.* **2018**, *130*, 17–38. [[CrossRef](#)]
155. Liu, W.; Li, Z.; Qiu, Y.; Li, J.; Yang, J.; Li, J. Biomineralization of Aggregation-Induced Emission-Active Photosensitizers for pH-Mediated Tumor Imaging and Photodynamic Therapy. *ACS Appl. Bio Mater.* **2021**, *4*, 5566–5574. [[CrossRef](#)]
156. Jiang, Y.; Zhu, W.; Xu, Z.; Zhang, Z.; Tang, S.; Fan, M.; Li, Z.; Zhang, J.; Yang, C.; Law, W.-C.; et al. A mitochondrion-targeting two-photon photosensitizer with aggregation-induced emission characteristics for hypoxia-tolerant photodynamic therapy. *Chem. Eng. J.* **2022**, *448*, 137604. [[CrossRef](#)]
157. Gao, F.; Wu, J.; Gao, H.; Hu, X.; Liu, L.; Midgley, A.C.; Liu, Q.; Sun, Z.; Liu, Y.; Ding, D.; et al. Hypoxia-tropic nanozymes as oxygen generators for tumor-favoring theranostics. *Biomaterials* **2020**, *230*, 119635. [[CrossRef](#)]

158. Ding, K.; Wang, L.; Zhu, J.; He, D.; Huang, Y.; Zhang, W.; Wang, Z.; Qin, A.; Hou, J.; Tang, B.Z. Photo-Enhanced Chemotherapy Performance in Bladder Cancer Treatment via Albumin Coated AIE Aggregates. *ACS Nano* **2022**, *16*, 7535–7546. [\[CrossRef\]](#)
159. Liu, J.; Hu, F.; Wu, M.; Tian, L.; Gong, F.; Zhong, X.; Chen, M.; Liu, Z.; Liu, B. Bioorthogonal Coordination Polymer Nanoparticles with Aggregation-Induced Emission for Deep Tumor-Penetrating Radio- and Radiodynamic Therapy. *Adv. Mater.* **2021**, *33*, 2007888. [\[CrossRef\]](#)
160. Wang, Y.; Shi, L.; Wu, W.; Qi, G.; Zhu, X.; Liu, B. Tumor-Activated Photosensitization and Size Transformation of Nanodrugs. *Adv. Funct. Mater.* **2021**, *31*, 2010241. [\[CrossRef\]](#)
161. Harrington, K.J.; Mohammadtaghi, S.; Uster, P.S.; Glass, D.; Peters, A.M.; Vile, R.G.; Stewart, J.S.W. Effective Targeting of Solid Tumors in Patients with Locally Advanced Cancers by Radiolabeled Pegylated Liposomes. *Clin. Cancer Res.* **2001**, *7*, 243–254. [\[PubMed\]](#)
162. Tanaka, N.; Kanatani, S.; Tomer, R.; Sahlgren, C.; Kronqvist, P.; Kaczynska, D.; Louhivuori, L.; Kis, L.; Lindh, C.; Mitura, P.; et al. Whole-tissue biopsy phenotyping of three-dimensional tumours reveals patterns of cancer heterogeneity. *Nat. Biomed. Eng.* **2017**, *1*, 796–806. [\[CrossRef\]](#) [\[PubMed\]](#)
163. Maeda, H.; Khatami, M. Analyses of repeated failures in cancer therapy for solid tumors: Poor tumor-selective drug delivery, low therapeutic efficacy and unsustainable costs. *Clin. Transl. Med.* **2018**, *7*, 11. [\[CrossRef\]](#)
164. Natfji, A.A.; Ravishankar, D.; Osborn, H.M.I.; Greco, F. Parameters Affecting the Enhanced Permeability and Retention Effect: The Need for Patient Selection. *J. Pharm. Sci.* **2017**, *106*, 3179–3187. [\[CrossRef\]](#) [\[PubMed\]](#)
165. Yoo, J.; Park, C.; Yi, G.; Lee, D.; Koo, H. Active targeting strategies using biological ligands for nanoparticle drug delivery systems. *Cancers* **2019**, *11*, 640. [\[CrossRef\]](#)
166. Cheung, A.; Bax, H.J.; Josephs, D.H.; Ilieva, K.M.; Pellizzari, G.; Opzoomer, J.; Bloomfield, J.; Fittall, M.; Grigoriadis, A.; Figini, M.; et al. Targeting folate receptor alpha for cancer treatment. *Oncotarget* **2016**, *7*, 52553–52574. [\[CrossRef\]](#)
167. Qian, Z.M.; Li, H.; Sun, H.; Ho, K. Targeted drug delivery via the transferrin receptor-mediated endocytosis pathway. *Pharmacol. Rev.* **2002**, *54*, 561–587. [\[CrossRef\]](#) [\[PubMed\]](#)
168. Bazak, R.; Hourri, M.; El Achy, S.; Kamel, S.; Refaat, T. Cancer active targeting by nanoparticles: A comprehensive review of literature. *J. Cancer Res. Clin. Oncol.* **2015**, *141*, 769–784. [\[CrossRef\]](#)
169. Danhier, F.; Feron, O.; Préat, V. To exploit the tumor microenvironment: Passive and active tumor targeting of nanocarriers for anti-cancer drug delivery. *J. Control. Release* **2010**, *148*, 135–146. [\[CrossRef\]](#)
170. Wang, X.; Luo, D.; Basilion, J.P. Photodynamic Therapy: Targeting Cancer Biomarkers for the Treatment of Cancers. *Cancers* **2021**, *13*, 2992. [\[CrossRef\]](#)
171. Chitgupi, U.; Qin, Y.; Lovell, J.F. Targeted Nanomaterials for Phototherapy. *Nanotheranostics* **2017**, *1*, 38–58. [\[CrossRef\]](#) [\[PubMed\]](#)
172. Gierlich, P.; Mata, A.I.; Donohoe, C.; Brito, R.M.M.; Senge, M.O.; Gomes-da-Silva, L.C. Ligand-Targeted Delivery of Photosensitizers for Cancer Treatment. *Molecules* **2020**, *25*, 5317. [\[CrossRef\]](#)
173. Wang, Y.; Pan, X.; Dai, T.; Wang, L.; Shi, H.; Wang, H.; Chen, Z. An AIE photosensitizer with unquenched fluorescence based on nitrobenzoic acid for tumor-targeting and image-guided photodynamic therapy. *Biomater. Sci.* **2022**, *10*, 4866–4875. [\[CrossRef\]](#)
174. Li, D.; Chen, X.; Wang, D.; Wu, H.; Wen, H.; Wang, L.; Jin, Q.; Wang, D.; Ji, J.; Tang, B.Z. Synchronously boosting type-I photodynamic and photothermal efficacies via molecular manipulation for pancreatic cancer theranostics in the NIR-II window. *Biomaterials* **2022**, *283*, 121476. [\[CrossRef\]](#)
175. Li, S.; Konstantinov, S.R.; Smits, R.; Peppelenbosch, M.P. Bacterial Biofilms in Colorectal Cancer Initiation and Progression. *Trends Mol. Med.* **2017**, *23*, 18–30. [\[CrossRef\]](#) [\[PubMed\]](#)
176. Dickson, I. Bacterial biofilms and toxins prompt a perfect storm for colon cancer. *Nat. Rev. Gastroenterol. Hepatol.* **2018**, *15*, 129. [\[CrossRef\]](#) [\[PubMed\]](#)
177. Jin, C.S.; Zheng, G. Liposomal Nanostructures for Photosensitizer Delivery. *Lasers Surg. Med.* **2011**, *43*, 734–748. [\[CrossRef\]](#) [\[PubMed\]](#)
178. Derycke, A.S.L.; de Witte, P.A.M. Liposomes for photodynamic therapy. *Adv. Drug Deliv. Rev.* **2004**, *56*, 17–30. [\[CrossRef\]](#) [\[PubMed\]](#)
179. Zhang, W.; Huang, X. Stem cell membrane-camouflaged targeted delivery system in tumor. *Mater. Today Bio* **2022**, *16*, 100377. [\[CrossRef\]](#)
180. Wang, S.; Duan, Y.; Zhang, Q.; Komarla, A.; Gong, H.; Gao, W.; Zhang, L. Drug Targeting via Platelet Membrane-Coated Nanoparticles. *Small Struct.* **2020**, *1*, 2000018. [\[CrossRef\]](#)
181. Guo, Y.; Wang, Z.; Shi, X.; Shen, M. Engineered cancer cell membranes: An emerging agent for efficient cancer theranostics. *Exploration* **2022**, *2*, 20210171. [\[CrossRef\]](#)
182. Yu, W.; Liu, R.; Zhou, Y.; Gao, H. Size-Tunable Strategies for a Tumor Targeted Drug Delivery System. *ACS Cent. Sci.* **2020**, *6*, 100–116. [\[CrossRef\]](#) [\[PubMed\]](#)
183. Yao, X.; Mu, J.; Zeng, L.; Lin, J.; Nie, Z.; Jiang, X.; Huang, P. Stimuli-responsive cyclodextrin-based nanoplatfoms for cancer treatment and theranostics. *Mater. Horiz.* **2019**, *6*, 846–870. [\[CrossRef\]](#)
184. Yang, N.; Xiao, W.; Song, X.; Wang, W.; Dong, X. Recent Advances in Tumor Microenvironment Hydrogen Peroxide-Responsive Materials for Cancer Photodynamic Therapy. *Nano-Micro Lett.* **2020**, *12*, 15. [\[CrossRef\]](#)
185. Hu, J.-J.; Jiang, W.; Yuan, L.; Duan, C.; Yuan, Q.; Long, Z.; Lou, X.; Xia, F. Recent advances in stimuli-responsive theranostic systems with aggregation-induced emission characteristics. *Aggregate* **2021**, *2*, 48–65. [\[CrossRef\]](#)

186. Zhu, D.; Zhang, J.; Luo, G.; Duo, Y.; Tang, B.Z. Bright Bacterium for Hypoxia-Tolerant Photodynamic Therapy Against Orthotopic Colon Tumors by an Interventional Method. *Adv. Sci.* **2021**, *8*, 2004769. [[CrossRef](#)]
187. Zhang, T.; Deng, Y.; Liu, Y.S.; Chua, S.L.; Tang, B.Z.; Khoo, B.L. Bacterial targeted AIE photosensitizers synergistically promote chemotherapy for the treatment of inflammatory cancer. *Chem. Eng. J.* **2022**, *447*, 137579. [[CrossRef](#)]
188. Duo, Y.; Suo, M.; Zhu, D.; Li, Z.; Zheng, Z.; Tang, B.Z. AIEgen-Based Bionic Nanozymes for the Interventional Photodynamic Therapy-Based Treatment of Orthotopic Colon Cancer. *ACS Appl. Mater. Interfaces* **2022**, *14*, 26394–26403. [[CrossRef](#)]
189. Liu, S.; Feng, G.; Tang, B.Z.; Liu, B. Recent advances of AIE light-up probes for photodynamic therapy. *Chem. Sci.* **2021**, *12*, 6488–6506. [[CrossRef](#)]
190. Hong, L.; Li, J.; Luo, Y.; Guo, T.; Zhang, C.; Ou, S.; Long, Y.; Hu, Z. Recent Advances in Strategies for Addressing Hypoxia in Tumor Photodynamic Therapy. *Biomolecules* **2022**, *12*, 81. [[CrossRef](#)]
191. Zhao, X.; Liu, J.; Fan, J.; Chao, H.; Peng, X. Recent progress in photosensitizers for overcoming the challenges of photodynamic therapy: From molecular design to application. *Chem. Soc. Rev.* **2021**, *50*, 4185–4219. [[CrossRef](#)] [[PubMed](#)]
192. Hu, T.; Wang, Z.; Shen, W.; Liang, R.; Yan, D.; Wei, M. Recent advances in innovative strategies for enhanced cancer photodynamic therapy. *Theranostics* **2021**, *11*, 3278–3300. [[CrossRef](#)] [[PubMed](#)]
193. Liu, X.; Duan, Y.; Liu, B. Nanoparticles as contrast agents for photoacoustic brain imaging. *Aggregate* **2021**, *2*, 4–19. [[CrossRef](#)]