Table S1: Ag NPs and antibacterial activity evaluation of selected publications from 2015-2018 period.

Stabilizer	NPs size (nm)	Bacteria	Bacteria origin	Protocol	Culture media	CFU/mL	MIC (μg/mL)	MBC (μg/mL)	Disk loading	ZOI (mm)	Reference
"Naked"	19.5 ± 7.7	*E. faecalis* *S. aureus* *S. epidermidis* *B. subtilis* *E. coli* *S. typhimurium* *S. enterica*	KCCM 13807 KCTC 1916 KCTC 1971 KCTC 1021 KCTC 1682 KCCM 40253 KACC 10763	Kirby-Bauer method Microdilution method MIC90	Mueller Hinton Agar Luria Bertani	10^7	256* 32–64* 32–64* 128* 128–256* 32–64* 256*	N/A	30 μg	20* 18* 21* 14* 16* 17* 16*	[3]
"Naked"	≈10	*A. baumannii* ($n = 17$) *A. baumannii* *A. nosocomialis* ($n = 10$)	Clinical isolates ATCC 1906 Clinical isolates	Microdilution method	N/A	N/A	0.39–0.78 0.78 0.39–0.78	/	/	/	[4]
"Naked"	10 20 40	*E. coli*	N/A	Kirby-Bauer method	Mueller Hinton 2 Agar	N/A	/	/	N/A	1–2 1–2 1–2	[5]
"Naked"	55.6 ± 2.9 (DLS)	*P. aeruginosa* ($n = 3$) *S. maltophilia* ($n = 3$) *B. cepacian* ($n = 3$) *S. aureus* ($n = 3$)	Clinical isolates	Kirby-Bauer method Microdilution method TKA	Cation Adjusted Mueller Hinton	1–5 × 10^9	1.06 1.06 4.25 2.125 2.125 1.06 4.25 4.25 4.25 8.5 8.5 8.5	2.125 2.125 4.25 4.25 4.25 2.125 4.25 4.25 4.25 17.0 8.5 8.5	N/A	N/A	[6]
"Naked"	2–5 (70–75% TEM)	*L. monocytogenes*	PCM 2191	Microdilution method	Tryptone Soy yeast extract broth	2 × 10^6	8	/	/	/	[7]
"Naked"	23.6 (TEM) 57.8 (DLS)	*E. coli* (K-12)	KCTC 1116	Growth Curves	Luria Bertani	1 × 10^8	/	/	/	/	[8]
"Naked"	10	*S. aureus* *S. aureus* ($n = 30$)	ATCC 25923 Clinical Isolates	Agar Well diffusion method Microdilution method	Mueller Hinton Agar Luria-Bertani	2 × 10^8	12.5 12.5 (16) 25 (14)	25 12.5 (17) 25 (13)	N/A 12.5 μg/mL (18) 25 μg/mL (12)	25–30	[9]

N/A: not available; KCCM: Korean Culture Center of Microorganisms; KCTC: Korean Collection of Type Cultures; KACC: Korean Agriculture Culture Collection; MIC: minimal inhibitory concentration; ATCC: American Type Culture Collection; DLS: dynamic light scattering; TKA: Time Kill Assay; TEM: transmission electron microscope; PCM: Polish Collection of Microorganisms; *estimated.

Stabilizer	NPs size (nm)	Bacteria	Bacteria origin	Protocol	Culture media	CFU/mL	MIC (μg/mL)	MBC (μg/mL)	Disk loading	ZOI (mm)	Reference
"Naked"	40 (TEM)	*S. aureus*	N/A	Agar Well diffusion method	Mueller Hinton Agar	N/A	/	/	N/A	27.7 ± 0.62	[10]
		B. cereus								24.1 ± 0.80	
		P. aeruginosa								15.8 ± 0.41	
		K. pneumoniae								28.2 ± 1.07	
		E. coli								29.4 ± 0.56	
Unknown	35	*E. coli* (K-12)	JM 109	Agar Well diffusion method	Mueller Hinton	1×10^4			N/A	5	[11]
"Naked"	27.2								20 μg/mL	7	
Citrate	23 ± 2 (TEM)	*L. monocytogenes* (*n* = 20)	Clinical isolates	Colony Forming Units	Mueller Hinton	1×10^8	/	/	/	/	[12]
		L. monocytogenes	ATCC 13932								
Citrate	6.0–28.2 (XRD)	*S. aureus*	ATCC 25923	Kirby-Bauer method	Nutrient Agar	1×10^4	/	/	0.25–3 mmol/mL	10 to 25	[13]
		E. coli	ATCC 35218							3 to 11	
Citrate	2.3 ± 0.5	*S. aureus*	ATCC 35696	Microdilution method Kirby-Bauer method Growth Curves	Broth medium	10^8–10^9	7.8	15.6	62.5 μg/mL	18.8	[14]
		E. coli	ATCC 23282				7.8	15.6		19.5	
	12.5 ± 2.2	*S. aureus*	ATCC 35696				15.6	31.3		14.8	
		E. coli	ATCC 23282				15.6	31.3		16.2	
	32.4 ± 6.5 (TEM)	*S. aureus*	ATCC 35696				62.5	125		9.3	
		E. coli	ATCC 23282				62.5	125		11.0	
Citrate	40–50	*S. aureus*	ATCC 25923	Microdilution method Kirby-Bauer method TKA	Nutrient Agar	10^8–10^9	190	N/A	242 μg	14*	[15]
		B. subtilis	AST5-2				195			15*	
		P. aeruginosa	AL2-14B				188			11*	
		K. pneumoniae	AWD5				184			17*	
		E. coli	ATCC 25922				190			5*	
	20 × 20–90	*S. aureus*	ATCC 25923				358			10*	
		B. subtilis	AST5-2				350			13*	
		P. aeruginosa	AL2-14B				348			9*	
		K. pneumoniae	AWD5				320			14*	
		E. coli	ATCC 25922				340			6*	
Citrate	20.1 ± 4.4 (TEM)	*E. coli*	ATCC 25922	TKA	Luria Bertani Tryptone Soy MRS	1×10^6	/	17.5	/	/	[16]
		S. aureus	ATCC 25923					14			
		L. bulgaricus	CGMCC 1.6970					9			
		L. casei	CGMCC 1.2435					8			
Citrate	10–40 (TEM)	*E. coli*	ATCC 25922	Growth Curves	Luria Bertani	/	/	/	/	/	[17]

N/A: not available; ATCC: American Type Culture Collection; DLS: dynamic light scattering; TKA: Time Kill Assay; TEM: transmission electron microscope; XRD: X-ray diffraction; CGMCC: China General Microbiological Culture Collection Center; MRS: deMan, Rogosa and Sharpe medium; *estimated from publication.

Stabilizer	NPs size (nm)	Bacteria	Bacteria origin	Protocol	Culture media	CFU/mL	MIC (µg/mL)	MBC (µg/mL)	Disk loading	ZOI (mm)	Reference
Citrate	20 ± 9 25 ± 3 11 ± 6 (TEM)	*S. aureus*	ATCC 25923	Agar Well diffusion method	Nutrient Agar	10^8–10^9	/	/	0.1 M ($AgNO_3$)	N/A	[18]
Citrate	42–58 (TEM)	*S. aureus* *S. pyogenes* *S. typhi* *P. aeruginosa*	N/A	Kirby-Bauer method	Tryptone Soy	10^7	/	/	5µg/disc	14.8 13.6 12.5 19.1	[19]
Citrate	15	*E. coli* *B. subtilis*	N/A	Kirby-Bauer method	Nutrient Agar	1.5 × 10^8	/	/	N/A	N/A N/A	[20]
	18	*E. coli* *B. subtilis*								7.0 6.5	
	30	*E. coli* *B. subtilis*								7.5 7.5	
	30 (DLS)	*E. coli* *B. subtilis*								N/A N/A	
GSH	10–50	*C. jejuni* (*n* = 22) *C. coli* (*n* = 18) *C. jejuni*	Animal or Human clinical isolates NCTC 11168	Microdilution method	Brucella Mueller Hinton	1 × 10^8	9.85–39.4 4.92–39.4 4.92	9.85–39.4 9.85–39.4 9.85	/	/	[21]
D-xylose L-arabinose D-ribose D-glucose D-galactose D-mannose D-lactose D-xylose	33 30 39 25 28 25 15 18	*E. coli* *Klebsiella* spp.	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	[22]
Citrate L-fucose	10.15 ± 3.37 (TEM)	*P. aeruginosa* (PAO1)	N/A	Microdilution method	Luria Bertani without Chloride ions	1 × 10^8	17.5 7.5	75 40	/	/	[23]
PEG	15.8 ± 2.2 (TEM)	*S. aureus* *P. aeruginosa* *S. enterica* *E. coli*	ATCC 6538 ATCC 15442 ATCC 10708 ATCC 11229	Microdilution method	Mueller Hinton	N/A	4.7 2.3 2.3 1.2	/	/	/	[24]

N/A: not available; ATCC: American Type Culture Collection; DLS: dynamic light scattering; TEM: transmission electron microscope; GSH: glutathione; NCTC: National Collection of Type Culture; PEG: polyethylene glycol.

Stabilizer	NPs size (nm)	Bacteria	Bacteria origin	Protocol	Culture media	CFU/mL	MIC (μg/mL)	MBC (μg/mL)	Disk loading	ZOI (mm)	Reference
PC	3.3 ± 0.9 4.9 ± 2.9 (TEM)	*E. coli*	OW6	Growth curves	Todd Hewitt broth	/	/	/	/	/	[25]
		S. aureus	Mu50								
PVA	31	*E. coli*	N/A	Kirby-Bauer method	Nutrient Agar	N/A	/	/	N/A	3.4	[26]
		Pseudomonas sp								4.1	
	24	*E. coli*								4.9	
		Pseudomonas sp								6.2	
	19	*E. coli*								7.0	
		Pseudomonas sp								8.1	
	14 (SEM)	*E. coli*								8.8	
		Pseudomonas sp								10.2	
PVP/citrate	50–60 70–80	*E. coli*	N/A	Colony Forming Units	Nutrient Agar	/	/	/	/	/	[27]
PVP	14.0 ± 0.3 (TEM)	*E. coli* (K-12)	MG1655	Growth curves	Tryptone Soy	/	/	/	/	/	[28]
		B. subtilis	ATCC 6051								
	5	*E. coli*	ATCC 8739	Poisoned Food Technique	Mueller Hinton	1×10^8	1	/	/	/	[29]
		P. aeruginosa	ATCC 9027				2				
		S. aureus	ATCC 6538				2				
		S. epidermidis	ATCC 12228				4				
	20	*E. coli*	ATCC 8739				2				
		P. aeruginosa	ATCC 9027				2				
		S. aureus	ATCC 6538				8				
		S. epidermidis	ATCC 12228				2				
	8	*A. hydrophila*	4AK4	Kirby-Bauer method Microdilution method	N/A Mueller Hinton	3×10^8	No activity	/	10 μg/mL	0	[30]
		P. putida	KT2442				No activity			0	
		E. coli	Trans 1-T1				No activity			0	
		B. subtilis	ATCC 28357				No activity			0	
		S. aureus	N/A				10 μg/mL			15	
	29 (TEM)	*A. hydrophila*	4AK4				No activity			0	
		P. putida	KT2442				No activity			0	
		E. coli	Trans 1-T1				No activity			0	
		B. subtilis	ATCC 28357				No activity			0	
		S. aureus	N/A				10 μg/mL			6	

N/A: not available; ATCC: American Type Culture Collection; DLS: dynamic light scattering; TKA: Time Kill Assay; TEM: transmission electron microscope; PCM: Polish Collection of Microorganisms; XRD: X-ray diffraction; CGMCC: China General Microbiological Culture Collection Center; GSH: glutathione; NCTC: National Collection of Type Culture; PEG: polyethylene glycol; CTAB: cetyl-trimethyl ammonium bromide; NTA: nanoparticle tracking analysis; PC: phosphorylcholine; PVA: polyvinyl alcohol; SEM: scanning electron microscope; PVP: polyvinylpyrrolidone.

Stabilizer	NPs size (nm)	Bacteria	Bacteria origin	Protocol	Culture media	CFU/mL	MIC (μg/mL)	MBC (μg/mL)	Disk loading	ZOI (mm)	Reference
PVP	15.6 (TEM)	*Citrobacter sp* *Enterococcus sp*	N/A	Colony Forming Units	N/A	/	/	/	/	/	[31]
	3–34 (TEM)	*S. aureus* PTCC No. 1112 *E. coli* PTCC No. 1330	ATCC 6537 ATCC 8739	N/A	Mueller Hinton	1×10^7	N/A	N/A	N/A	N/A	[32]
	10–15 (TEM)	*E. coli* *S. aureus*	ATCC 25922 ATCC 25923	Kirby-Bauer method	Nutrient Agar	1×10^7–10^8	/	/	100 μL	N/A	[33]
	60 ± 15	*E. coli*	ATCC 25922	Growth curves Microdilution method	Luria Bertani	10^3 10^4 10^5 10^6	50.0 ± 5.2 75.0 ± 2.6 100.0 ± 7.0 100.0 ± 6.0	/	/	/	[34]
	55 ± 10					10^3 10^4 10^5 10^6	25 ± 4.6 37.5 ± 5.3 50.0 ± 3.5 75.0 ± 4.6				
	60 × 2000–4000					10^3 10^4 10^5 10^6	50.0 ± 4.6 100.0 ± 5.3 > 100.0 > 100.0				
	20.6 ± 3.1	*C. jejuni* (*n* = 4) *C. jejuni* *Salmonella* spp. (*n* = 5)	Chicken isolates NCTC 11168 Chicken isolates	Microdilution method	Mueller Hinton Luria Bertani	10^5	3.125–6.25 6.25 12.5–25	/	/	/	[35]
PVP/Glycerol	31.2 (TEM) 46.5 (DLS)	*C. sakazakii*	ATCC 29544 ATCC BAA894 ATCC 29004 ATCC 12868	Microdilution method Oxford cup method	Luria Bertani	1×10^8	125 125 62.5 62.5	/	200 mg/L	14.3 13.8 16.3 16.0	[36]
Oleylamine	10 (TEM)	*B. subtilis*	ATCC 6633	Growth curves	Luria Bertani	/	/	/	/	/	[37]
Casein	12.5 ± 4 (TEM) 50.0 ± 0.7 (DLS)	*E. coli* *P. aeruginosa*	MC 1061 DS 10-129	Bioluminescence inhibition assay	Luria Bertani	1×10^9	/	/	/	/	[38]

N/A: not available; ATCC: American Type Culture Collection; TEM: transmission electron microscope; NCTC: National Collection of Type Culture; PVP: polyvinylpyrrolidone.

Stabilizer	NPs size (nm)	Bacteria	Bacteria origin	Protocol	Culture media	CFU/mL	MIC (μg/mL)	MBC (μg/mL)	Disk loading	ZOI (mm)	Reference
Sericin	3.78 ± 1.14 (TEM)	*S. aureus*	ATCC 25923	Cell counting (FCM)	Nutrient medium	N/A	/	/	/	/	[39]
		E. coli	ATCC 25922								
Thioacetic acid	20–25	*S. aureus*	ATCC 25923	Microdilution method	Mueller Hinton	N/A	1	1	/	/	[40]
		S. epidermidis	ATCC 35984				1	10			
		A. baumannii	ATCC 19606				10	10			
		P. aeruginosa	ATCC 27853				10	10			
Propionic acid	30–35	*S. aureus*	ATCC 25923				1	1			
		S. epidermidis	ATCC 35984				1	1			
		A. baumannii	ATCC 19606				1	10			
		P. aeruginosa	ATCC 27853				10	10			
Lipoid acid	2.0 ± 0.5 (TEM)	*S. aureus*	N/A	Growth curves	Luria Bertani	1×10^{6}	/	/	/	/	[41]
		E. coli	DH5α								
		E. coli	DSM4230								
PEG	44	*E. coli*	N/A	Kirby-Bauer method	Nutrient Agar	N/A	/	/	6 μL	3	[42]
		Pseudomonas spp.								3	
EDTA	39	*E. coli*								4	
		Pseudomonas spp.								5	
PVP	35	*E. coli*								6	
		Pseudomonas spp.								6	
PVA	31 (SEM)	*E. coli*								7	
		Pseudomonas spp.								8	
"Naked" Unknown	7.5	*S. aureus*	ATCC 29737	Agar Well diffusion method	Brain Heart Infusion	1.5×10^{8}	9.7×10^{-8}	/		5–40*	[43]
		S. mutans	ATCC 35668				ND			5–22*	
		S. pyogenes	ATCC 8668				ND			8–20*	
		E. coli	ATCC 15224				ND			4–31*	
		P. vulgaris	ATCC 7829				ND			5–10*	
		S. aureus	ATCC 29737	Microdilution method	Mueller Hinton		4×10^{-9}			5–40*	
		S. mutans	ATCC 35668				4×10^{-8}			5–22*	
	10.1	*S. pyogenes*	ATCC 8668				4×10^{-8}			2–29*	
		E. coli	ATCC 15224				4×10^{-10}			6–39*	
	(TEM)	*P. vulgaris*	ATCC 7829				ND			3–5*	

N/A: not available; ATCC: American Type Culture Collection; DLS: dynamic light scattering; TEM: transmission electron microscope; PEG: polyethylene glycol; PVA: polyvinyl alcohol; SEM: scanning electron microscope; PVP: polyvinylpyrrolidone; EDTA: ethylenediaminetetraacetid acid; C12mim: 3-methylimidazolium chloride; FCM: Flow Cytometry; * estimated from publication.

Stabilizer	NPs size (nm)	Bacteria	Bacteria origin	Protocol	Culture media	CFU/mL	MIC (μg/mL)	MBC (μg/mL)	Disk loading	ZOI (mm)	Reference
							MIC_{50}/MIC_{90}				
PVA	10	*A. baumannii* (*n* = 17)	Clinical isolates	Agar Well diffusion method; Microdilution method; Time kill assay	Mueller Hinton; Mueller Hinton Cation Adjusted; Tryptone Soy; Mueller Hinton Cation Adjusted	N/A	13.5; ≥ 54		50μL	0*	[44]
		P. aeruginosa (*n* = 12)	Clinical isolates				13.5; ≥ 54	≥ 54			
		P. aeruginosa	ATCC 27853 S				13.5				
		P. aeruginosa	INCQS 230								
		Enterobacteriaceae (*n* = 21)	Clinical isolates				≥ 54; ≥ 54				
		S. maltophilia (*n* = 2)	Clinical isolates				≥ 54; ≥ 54				
		S. aureus (*n* = 13)	Clinical isolates				≥ 54; ≥ 54				
		S. aureus	ATCC 29213				≥ 54	≥ 54			
		S. epidermidis	INCQS 198				27	27			
		Enterococcus sp (*n* = 14)	Clinical isolates				≥ 54; ≥ 54				
Citrate	40	*A. baumannii* (*n* = 17)	Clinical isolates				3.4; 3.4			12*	
		P. aeruginosa (*n* = 12)	Clinical isolates				3.4; 3.4				
		P. aeruginosa	ATCC 27853 S				3.4	6.7			
		P. aeruginosa	INCQS 230								
		Enterobacteriaceae (*n* = 21)	Clinical isolates				6.7; 6.7				
		S. maltophilia (*n* = 2)	Clinical isolates				1.6; 1.6				
		S. aureus (*n* = 13)	Clinical isolates				6.7; 13.5				
		S. aureus	ATCC 29213				13.5				
		S. epidermidis	INCQS 198				6.7	27			
		Enterococcus sp (*n* = 14)	Clinical isolates				6.7; 13.5	6.7			
Citrate	60	*A. baumannii* (*n* = 17)	Clinical isolates				≥ 10; ≥ 10			N/A	
		P. aeruginosa (*n* = 12)	Clinical isolates				≥ 10; ≥ 10				
		P. aeruginosa	ATCC 27853 S				≥ 10	≥ 10			
		P. aeruginosa	INCQS 230								
		Enterobacteriaceae (*n* = 21)	Clinical isolates				≥ 10; ≥ 10				
		S. maltophilia (*n* = 2)	Clinical isolates				≥ 10; ≥ 10				
		S. aureus (*n* = 13)	Clinical isolates				≥ 10; ≥ 10				
		S. aureus	ATCC 29213				≥ 10				
		S. epidermidis	INCQS 198				≥ 10	≥ 10			
		Enterococcus sp (*n* = 14)	Clinical isolates				≥ 10; ≥ 10	≥ 10			

N/A: not available; ATCC: American Type Culture Collection; DLS: dynamic light scattering; TEM: transmission electron microscope; PEG: polyethylene glycol; PVA: polyvinyl alcohol; SEM: scanning electron microscope; * estimated from publication.

Stabilizer	NPs size (nm)	Bacteria	Bacteria origin	Protocol	Culture media	CFU/mL	MIC (µg/mL)	MBC (µg/mL)	Disk loading	ZOI (mm)	Reference
Cysteine	7.6 ± 1.5	*S. aureus*	ATCC 29213	Microdilution method	Mueller Hinton Cation Adjusted	N/A	580	/	/	/	[45]
		E. coli	ATCC 23716				145				
		P. aeruginosa	ATCC 25619				35				
PVP	7.7 ± 1.6 (TEM)	*S. aureus*	ATCC 29213				69				
		E. coli	ATCC 23716				69				
		P. aeruginosa	ATCC 25619				73				
Citrate	10.2 ± 2.3	*E. coli*	MG 1655	Growth curves	Luria Bertani	1 × 10^7	/	/	/	/	[46]
MPA	10.2 ± 2.5										
MHA	10.2 ± 2.2										
MPS	9.9 ± 2.0										
Citrate	40	*E. coli*	DH5*α*	Agar Well diffusion method Growth curves	Luria Bertani	N/A	/	/	4 x 10^{-9} mg	20.2 ± 5.5	[47]
PVP										0.8 ± 1.4	
PEG										24.5 ± 2.4	
Lipoid acid	9.5 ± 1.9 (TEM)	*Actinomyces* (*n* = 1)	Clinical isolates	Plate dilution method / Microdilution method	Brucella agar supplemented / Mueller Hinton	5 × 10^5	≤ 5	/	/	/	[48]
		Bacteroides (*n* = 4)	Clinical isolates				80 - ≥ 100				
		Bacteroides fragilis	ATCC 25285				≥ 100				
		Bifidobacterium (*n* = 1)	Clinical isolates				≤ 5				
		Bifidobacterium breve	ATCC 15700				10				
		Finegoldia (*n* = 2)	Clinical isolates				5-10				
		Fusobacterium (*n* = 4)	Clinical isolates				40 - ≥ 100				
		Fusobacterium nucleatum	ATCC 25585				40				
		Parabacteroides (*n* = 1)	Clinical isolates				80				
		Parvimonas (*n* = 2)	Clinical isolates				≤ 5				
		Peptostreptococcus (*n* = 1)	Clinical isolates				≤ 5				
		Peptostreptococcus anaerobius	ATCC 25286				10				
		Porphyromonas (*n* = 3)	Clinical isolates				≤ 5–20				
		Porphyromonas levii	ATCC 29147				≤ 5				
		Prevotella (*n* = 5)	Clinical isolates				10 - ≥ 100				
		Prevotella loescheii	ATCC 15930				≥ 100				
		Propionibacterium (*n* = 2)	Clinical isolates				10–40				
		Tannerella (*n* = 1)	Clinical isolates				20				
		S. aureus	ATCC 25923				5.0				
		S. aureus	ATCC 6538				2.5				
		S. aureus	ATTC 6538P				5.0				
		S. epidermidis	ATCC 14990				5.0				
		S. mutans	ATCC 29175				5.0				

N/A: not available; ATCC: American Type Culture Collection; TEM: transmission electron microscope; PEG: polyethylene glycol; PVP: polyvinylpyrrolidone; MPA: mercaptopropionic acid; MHA: mercaptohexanoic acid; MPS: mercaptopropionic sulfonic acid.

Stabilizer	NPs size (nm)	Bacteria	Bacteria origin	Protocol	Culture media	CFU/mL	MIC (µg/mL)	MBC (µg/mL)	Disk loading	ZOI (mm)	Reference
PEG	9.8 ± 2.0 (TEM)	*Actinomyces* (*n* = 1)	Clinical isolates	Plate dilution method Microdilution method	Brucella agar supplemented Mueller Hinton	5×10^5	≤ 5	/	/	/	[48]
		Bacteroides (*n* = 4)	Clinical isolates				40 - ≥ 100				
		Bacteroides fragilis	ATCC 25285				≤ 5				
		Bifidobacterium (*n* = 1)	Clinical isolates				≤ 5				
		Bifidobacterium breve	ATCC 15700				10				
		Finegoldia (*n* = 2)	Clinical isolates				5–20				
		Fusobacterium (*n* = 4)	Clinical isolates				40				
		Fusobacterium nucleatum	ATCC 25585				≤ 5				
		Parabacteroides (*n* = 1)	Clinical isolates				40				
		Parvimonas (*n* = 2)	Clinical isolates				≤ 5				
		Peptostreptococcus (*n* = 1)	Clinical isolates				≤ 5				
		Peptostreptococcus anaerobius	ATCC 25286				10				
		Porphyromonas (*n* = 3)	Clinical isolates				≤ 5–20				
		Porphyromonas levii	ATCC 29147				≤ 5				
		Prevotella (*n* = 5)	Clinical isolates				20 - ≥ 100				
		Prevotella loescheii	ATCC 15930				20				
		Propionibacterium (*n* = 2)	Clinical isolates				10–80				
		Tannerella (*n* = 1)	Clinical isolates				40				
		S. aureus	ATCC 25923				2.5				
		S. aureus	ATCC 6538				5.0				
		S. aureus	ATTC 6538P				10.0				
		S. epidermidis	ATCC 14990				0.625				
		S. mutans	ATCC 29175				10				

N/A: not available; ATCC: American Type Culture Collection; TEM: transmission electron microscope; PEG: polyethylene glycol.

Stabilizer	NPs size (nm)	Bacteria	Bacteria origin	Protocol	Culture media	CFU/mL	MIC (μg/mL)	MBC (μg/mL)	Disk loading	ZOI (mm)	Reference
"Naked"	11.2 ± 2.1 (TEM)	*Actinomyces* (*n* = 1)	Clinical isolates	Plate dilution method Microdilution method	Brucella agar supplemented Mueller Hinton	5×10^5	≤ 5	/	/	/	[48]
		Bacteroides (*n* = 4)	Clinical isolates				40 - ≥ 100				
		Bacteroides fragilis	ATCC 25285				≥ 100				
		Bifidobacterium (*n* = 1)	Clinical isolates				≤ 5				
		Bifidobacterium breve	ATCC 15700				≤ 5				
		Finegoldia (*n* = 2)	Clinical isolates				5–10				
		Fusobacterium (*n* = 4)	Clinical isolates				≤ 5–20				
		Fusobacterium nucleatum	ATCC 25585				≥ 100				
		Parabacteroides (*n* = 1)	Clinical isolates				≤ 5				
		Parvimonas (*n* = 2)	Clinical isolates				≤ 5				
		Peptostreptococcus (*n* = 1)	Clinical isolates				≤ 5				
		Peptostreptococcus anaerobius	ATCC 25286				≤ 5				
		Porphyromonas (*n* = 3)	Clinical isolates				≤ 5–10				
		Porphyromonas levii	ATCC 29147				≥ 100				
		Prevotella (*n* = 5)	Clinical isolates				≤ 5–40				
		Prevotella loescheii	ATCC 15930				≤ 5				
		Propionibacterium (*n* = 2)	Clinical isolates				≤ 5–10				
		Tannerella (*n* = 1)	Clinical isolates				10				
		S. aureus	ATCC 25923				2.5				
		S. aureus	ATCC 6538				2.5				
		S. aureus	ATTC 6538P				10.0				
		S. epidermidis	ATCC 14990				0.3125				
		S. mutans	ATCC 29175				10				
Citrate	15 ± 4	*E. coli* (K-12)	ATCC 10798	Microdilution method	Mueller Hinton	1×10^9	/	1.7×10^3	/	/	[49]
		E. coli	ER2566					165.0×10^3			
HH	13 ± 2	*E. coli* (K-12)	ATCC 10798					18.1×10^3			
		E. coli	ER2566					166.0×10^3			
SHSH	13 ± 4	*E. coli* (K-12)	ATCC 10798					5.2×10^3			
		E. coli	ER2566					10.3×10^3			
SHST	10 ± 6	*E. coli* (K-12)	ATCC 10798					2.6×10^3			
	(TEM)	*E. coli*	ER2566					55.0×10^3			
Starch	8 ± 4 (TEM)	*S. aureus*	N/A	Kirby-Bauer method	Nutrient agar	10^5	/	/	1×10^{-7} mol	11	[50]
		E. coli								9	
AOT	20	*E. coli*	N/A	Microdilution method	Luria Bertani	10^8	512	1024	/	/	[51]
		S. aureus					256	512			
	50	*E. coli*					1024	1024			
		S. aureus					512	2048			

N/A: not available; ATCC: American Type Culture Collection; TEM: transmission electron microscope; HH: Hydroxylamine hypochlorite; SHSH: Sodium hypophosphite and sodium hexametaphosphate; SHST: Sodium hypophosphite, sodium hexametaphosphate and sodium tripolyphosphate; AOT: Bis(2-ethylhexyl) sulfosuccinate.

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