



Article Efficient Detection of Nerve Agents through Carbon Nitride Quantum Dots: A DFT Approach

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Abstract: V-series nerve agents are very lethal to health and cause the inactivation of acetylcholinesterase which leads to neuromuscular paralysis and, finally, death. Therefore, rapid detection and elimination of V-series nerve agents are very important. Herein, we have carried out a theoretical investigation of carbon nitride quantum dots (C_2N) as an electrochemical sensor for the detection of V-series nerve agents, including VX, VS, VE, VG, and VM. Adsorption of V-series nerve agents on C_2N quantum dots is explored at M05-2X/6-31++G(d,p) level of theory. The level of theory chosen is quite adequate in systems describing non-bonding interactions. The adsorption behavior of nerve agents is characterized by interaction energy, non-covalent interaction (NCI), Bader's quantum theory of atoms in molecules (QTAIM), frontier molecular orbital (FMO), electron density difference (EDD), and charge transfer analysis. The computed adsorption energies of the studied complexes are in the range of -12.93 to -17.81 kcal/mol, which indicates the nerve agents are physiosorbed onto C₂N surface through non-covalent interactions. The non-covalent interactions between V-series and C2N are confirmed through NCI and QTAIM analysis. EDD analysis is carried out to understand electron density shifting, which is further validated by natural bond orbital (NBO) analysis. FMO analysis is used to estimate the changes in energy gap of C_2N on complexation through HOMO-LUMO energies. These findings suggest that C_2N surface is highly selective toward VX, and it might be a promising candidate for the detection of V-series nerve agents.

Keywords: organophosphorus compounds; V-series nerve agents; frontier molecular orbital (FMO); electron density differences (EDD); sensing

1. Introduction

V-series nerve agents are lethal organophosphorus compounds that can inhibit acetylcholinesterase present in the central nervous system. The inactivation of acetylcholinesterase results in the accumulation of acetylcholine in the synapse, which can lead to neuromuscular paralysis and, finally, death [1–3]. These nerve agents display a potential threat to the community due to their physical properties (odorless, colorless, high volatility), higher toxicity, facile synthesis, and rapid action [4]. V-series nerve agents are devastating weapons used by terrorists due to their dispersive and highly lethal nature [5,6]. For the last two decades, V-type nerve agents have been used in various terrorist attacks [7–11].

V-series are considered more potent than G-series due to their greater resistance to detoxification and higher stability [12]. Therefore, quick, effective analytical tools are required to detect V-type nerve agents that will help to prevent terrorist attacks utilizing



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Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). these agents. A variety of alternative techniques have been developed and used, such as gas chromatography along with mass spectrometry [13,14], fluorescence spectroscopy [15], ion mobility spectrometry (IMS), desorption electrospray ionization (DESI) [16], paper-based sensors [17], and membrane technology [18]. Although the mentioned techniques are desirable to capture and destroy harmful CWAs, these techniques are accompanied by some drawbacks such as complex procedures, user-unfriendly, and being expensive. In contrast, electrochemical sensors are most commonly employed for the detection of various analytes due to their rapid response, low cost, user-friendly nature, and simplicity [19].

Literature reveals that several types of materials have been investigated as electrochemical sensors for rapid and selective detection of nerve agents and other lethal CWA. For example, magnesium oxide nano-sheets, 3D graphene, boron nitrite nano-sheets, metal organic frameworks (MOFs) [20–22], covalent organic frameworks (COFs) [23,24], graphitic carbon nitride [25], robust hydrophobic MOFs [26], chemo sensors [27], and graphenebased composite sensors [28,29] have been used in this regard. These materials show many valuable properties in the detection and destruction of toxic pollutants and harmful CWAs. Changes in electronic properties upon adsorption of aromatic analytes (coronene, benzene, circumcirumcoronene, and circumcoronene) over Cu(111) and Au(111) are described well through interface dipole interactions [30,31]. However, these materials possess a small surface area, low porosity, rapid deactivation, less active sites for adsorption, and low reproducibility. Thus, a space is available to explore new sensing material, which must exhibit rapid response, large surface area, high sensitivity, and selectivity.

Herein, we designed and studied nitrogen-containing carbon nitride quantum dots (C₂N) as a sensor for the detection of V-series nerve agents [32]. The C₂N quantum dots have shown some remarkable properties, such as greater thermal stability, high electron-rich nitrogen content, and high surface area [33]. C₂N shows higher specific surface area, good thermal stability, and conductivity than other CN-based compounds. According to the study of Tian et al., the limit of thermal stability of different material composites with C₂N in different stoichiometry is \approx 700–750 °C [34]. We previously showed through MD simulations that the structure of C₂N is quite stable beyond 400 °C [35]. Many promising applications of C₂N have been reported in the literature, such as fluorescence spectroscopy [36] and photocatalysis [37–40]. Similarly, C₂N has been extensively explored theoretically for hydrogen storage [41], selective detection of picric acid [42], detection of ammonia [43], and batteries [44,45]. Recently, 2D C₂N surface has been investigated for the selective detection of NH₃ as an electrochemical sensor [43,46]. The presence of an electron-rich nitrogenated cavity makes the adsorption process facile; therefore, C₂N surface is a promising candidate for adsorption and sensing applications [47].

In our previously reported studies on C_2N surface, C_2N showed high selectivity towards NH₃ (among studied toxic analytes; HF, PH₃, HCN, NH₃, and H₂S), NM (Nmustard) (among studied analytes; GA, GB, GF, GD, NM, and SM) and NI₃ (among studied nitrogen halides; NBr₃, NCl₃, and NI₃) on conductivity basis. In summary, the C_2N surface has remained selective towards analytes that exhibit S--H, N--H, or C--H bonds of a methyl group attached to a phosphorus or nitrogen group [40,43,47,48]. Additionally, C_2N surface bears a well-defined activity that is better suited for the adsorption of V-type nerve agents. An extensive literature survey reveals that literature is silent upon the evaluation of interactions of V-type nerve agents (VX, VE, VM, VR, VG) with C_2N surface [49]. In the current study, we have carried out DFT calculations to study the mode of adsorption of five different V-series nerve agents on C_2N surface. The perturbation in band gap upon complexation is evaluated through frontier molecular orbital (FMO) analysis. The nature of the interactions is examined by the quantum theory of atoms in molecules (QTAIM), non-covalent interaction (NCI), symmetry adapted perturbation theory (SAPT0) analysis, and electron density differences (EDD).

2. Computational Methodology

All optimizations were performed using the M05-2X method with a 6-31++G(d,p)level of theory, and calculations were computed via Gaussian 09 software. M05-2X, a calibrated hybrid method with two non-local exchange functions, is best for capturing non-covalent interactions [50]. We used the M05-2X functional for our DFT study based on already reported benchmark studies in the literature. This functional is best specifically when dispersion interactions dominate among non-covalent interactions, as in the case of our study. A benchmark study was performed by Burns et al. on several hybrid functionals, and they reported that dispersion dominated non-covalent interactions could be better studied through the M05-2x hybrid functional [51]. Quite rich literature is available on the efficiency of the M05-2X hybrid functional for non-bonding interactions [25,52–57]. While frontier molecular orbitals (FMO) analysis was performed using the B3LYP method with 6-31G(d) level of theory because the accuracy of this method is well reported in the literature [58–60]. Visualization of the optimized geometries was performed through GaussView 5.0 and Chemcraft software [61,62]. Several possible orientations for each analyte were studied to obtain the most stable configuration with the lowest energy of each complex. The interaction energies of interacting fragments were evaluated by the following equation:

$$\Delta E = E_{\text{complex}} - (E_{\text{V-series}} + E_{\text{C2N}})$$
(1)

In Equation (1): $E_{complex}$, E_{C2N} , and $E_{V-series}$ represent the electronic energies of V-series nerve agents C_2N complexes, isolated C_2N surface, and V-series nerve agents, respectively. Frontier molecular orbital (FMO) analysis is carried out to analyze the change in the energy band gap [63]. To study the charge transfer between V-series nerve agents and C_2N surface, natural bond orbital (NBO) and electron density differences (EDD) analysis was performed [64]. EDD analysis provides a visual portrayal of the charge transfer among interacting fragments through colored isosurfaces and further validates NBO results [65].

NCI analysis is computed to visualize and differentiate the interactive forces existing among the analyte and C₂N surface. Multiwfn 3.7 software is employed to construct 2D RDG graphs and 3D isosurfaces of V-series@C₂N complexes. The non-covalent interaction index is generally dependent on two variables, RDG (reduced density gradient) and ρ (electron density), and they are interrelated by the equation given below:

$$RDG = \frac{1}{2(3\pi)^{1/3}} \frac{\nabla \rho}{\rho^{3/4}}$$
(2)

According to the divergence theorem, the density of net gradient flux is indicated by the Laplacian ($\nabla^2 \rho$) sign. A positive sign of Laplacian ($\nabla^2 \rho > 0$) indicates that the flux is leaving, whereas the negative sign ($\nabla^2 \rho < 0$) shows that flux is entering in an infinitesimally small volume in the vicinity of the reference point [47]. Therefore, the sign of Laplacian indicates the accumulation or depletion of electron density at the reference point. In NCI analysis, the nature of interactions is described via a color scheme dependent on the value of sign(λ_2) ρ . The appearance of blue spikes and isosurfaces show strong electrostatic interactions with a corresponding negative value of sign(λ_2) ρ in 2D-RDG graphs and 3D plots, respectively. While sign(λ_2) ρ with small negative values indicates weak van der Waals forces, appearing as green spikes and isosurfaces in 2D-RDG graphs and 3D plots, respectively. Red isosurfaces appear when sign(λ_2) ρ has large positive values and indicates repulsive interactions [42,56,65].

Symmetry-adapted perturbation theory (SAPT) analysis is also carried out to understand the mode of interaction present between the V-series and C₂N surface. SAPT0 calculations were computed with PSI4 software. SAPT0 analysis consists of four components of interaction energies, i.e., induction (ΔE_{ind}), dispersion (ΔE_{disp}), electrostatic (ΔE_{elstat}), and exchange (ΔE_{exch}). The equation for SAPT0 ΔE_{int} can be written as:

$$\Delta E_{int} = \Delta E_{ind} + \Delta E_{disp} + \Delta E_{elstat} + \Delta E_{exch}$$
(3)

Hence, SAPT0 total consists of the sum of these four contributing factors, i.e., ΔE_{ind} , ΔE_{disp} , ΔE_{elstat} , and ΔE_{exch} , which stabilizes V-series nerve agents with C₂N upon complexation [66]. Among these components, ΔE_{disp} , ΔE_{elstat} , and ΔE_{ind} are attractive forces, whereas the ΔE_{exch} factor is a repulsive force.

Another useful tool for quantifying non-covalent interactions is QTAIM analysis. Important topological parameters, including electronic density (ρ), Laplacian ($\nabla^2 \rho$), and total energy density (H(r)), are employed to confirm the types of interactions via bond critical points (BCPs). Similarly, other parameters, such as the local potential energy V(r), local Lagrangian kinetic energy G(r), and total energy density H(r), are being used to understand the types of interactions existing between the V-series and C₂N surface. Whereas, in topological analysis, Laplacian ($\nabla^2 \rho$) and electronic density (ρ) are the two main parameters used to examine the strength of a bond for particular BCPs [42,67–69]. Overall, we used Multiwfn, VMD, Gnuplot, and GIMP for NCI, QTAIM, and EDD analyses, while the SAPT0 graph was generated through MS excel. The features of PCs used are Workstation Intel Core i7 6700 K processor, 8 MB Cache 6th Generation Gigabit H 110 main board, with DDR IV, RAM 1900 MHZ, 16 GB, tower casing 400-Watt supply, DVD RW, SATA 10,000 GB hard disk. The average time taken for optimization in this study is two days, 19 h 7 min, while for properties average run time is 5 h 11 min.

3. Results and Discussion

3.1. Geometric Optimization and Adsorption Energies

In the current study, the adsorption behavior of five different V-series nerve agents was examined against C₂N surface. The chemical structures of these V-series nerve agents are presented in Figure 1. V-series nerve agents selected as analytes are VX; O-Ethyl-S-[2(diisopropylamino)ethyl] methylphosphonothioate, VS; O-Ethyl-S-[2(diisopropylamino)ethyl] ethylphosphonothioate VE; O-Ethyl-S-[2-(diethylamino)ethyl] ethylphosphonothioate VE; O-Ethyl-S-[2-(diethylamino)ethyl] ethylphosphonothioate, VG; O,O-Diethyl-S-[2-(diethylamino)ethyl] phosphorothioate and VM; O-Ethyl-S-[2-(diethylamino)ethyl] methylphosphonothioate (Figure 1). In V-type nerve agents, a thiophosphate (S-P=O) group makes up the central unit. On one side of the thiophosphate group, an -OR group is attached, while on the other side, R group is present (the R group could be methyl or ethyl) (see Figure 1) [70].



Figure 1. Chemical structures of studies V-type nerve agents.

The optimized C_2N quantum dot is shown in Figure 2. C_2N consists of alternate pyrazine and benzene rings. In C_2N material, the presence of an electron-rich nitrogenated cavity might help in the adsorption of analytes and thus make them suitable candidates for sensing studies against CWAs. The C_2N surface consists of four appropriate binding sites; **A**; the center of C_2N surface, **B**; triangular region made by C-N atoms, **C**; at the top of benzene rings and **D**; is over pyrazine rings (see Figure 2) [71,72].



Figure 2. Optimized geometry of C_2N surface with four binding sites (A, B, C, and D) at the M05-2X/6-31++G(d,p) level of theory (yellow shows nitrogen, red represents hydrogen and light purple represents carbon).

All possible orientations were explored over four favorable binding sites to get the most stable and lowest energy configuration for all V-series@C₂N complexes. The most stable optimized geometries of V-series@C₂N complexes are presented in Figure 3.

Interaction distance and energy (E_{int}) are two important parameters for real estimation of interaction behavior [73]. The bond interaction distance decreases when the analyte approaches the surface (C_2N) for complexation. Interactions existing between V-series nerve agents and C_2N surface are attributed to the minimum interaction distance, which can be understood via interaction energy. We have presented interaction distances for all complexes (V-series@ C_2N) along with corresponding interaction energy in Table 1.



Figure 3. The optimized geometries of V-series@C₂N complexes at M05-2X/6-31++G(d,p) level of theory.

Nitro@C ₂ N						
Complex	Intermolecular Bond	Bond Length (Å)	E _{int} (kcal/mol)			
	H10N1	2.80				
	H9N2	2.75				
VX@C2N	H9N3	2.38	-17.64			
	H9N4	2.85				
	H8N5	2.40				
	H7H6	2.78				
	H7N1	2.87				
VEOC N	H6N3	2.96	10.02			
v S@C2IN	H6N4	2.84	-12.93			
	H5N2	2.99				
	H9N1	2.87				
	H8N2	2.55				
VEQC N	H8N3	2.72	12.07			
VE@C2IN	H10N5	2.72	-13.97			
	H10N6	2.65				
	H7N4	2.72				
	H7N1	2.84				
	H7N2	2.40				
VC@C-N	H10N3	2.42	17 01			
VG@C2IV	H10N4	2.72	-17.61			
	H8H6	2.90				
	O9C5	2.84				
	H7N1	2.86				
VM@C-N	H6N2	2.79	17.60			
VIVI@C2IN	H6N3	2.91	-17.09			
	H5H4	2.66				

Table 1. Bond lengths (Å) and interaction energies (kcal/mol) of the V-series@C₂N complexes at the M05-2X/ 6-31++G(d,p) level of theory.

The interaction energies of VG@C₂N, VM@C₂N, and VX@C₂N complexes are quite comparable, and interaction energy values are -17.81 kcal/mol, -17.69 kcal/mol, and -17.64 kcal/mol, respectively (see Table 1). Whereas the other two complexes, i.e., VE@C₂N and VS@C₂N, have interaction energy values of -13.97 kcal/mol and -12.93 kcal/mol, respectively. The optimized stable geometry of **VX@C2N** showed strong non-covalent interactions between the H-atoms of **VX** and the N-atoms of **C₂N**. The strongest H---N interactions are observed between H9---N3 and H8---N5 atoms (see Figure 3) of the **VX@C2N** complex with interaction distances of 2.38 Å and 2.40 Å, respectively. Whereas relatively weak H---N interactions are observed between H10---N1, H9---N2, and H9---N4 atoms of **VX@C2N** complex with interaction distances of 2.75 Å, 2.80 Å, and 2.85 Å, respectively. Moreover, van der Waals interactions are also observed between the H7 atom of VX and the H6 atom of C₂N with an interaction distance of 2.78 Å (see Figure 3).

The most stable configuration of $VS@C_2N$ shows that H-atoms of the ethyl group of VS are projecting towards the C_2N surface and interacting with N-atoms of the C_2N cavity. The bond distances of H---N interactions range from 2.84 Å to 2.99 Å in the $VS@C_2N$ complex. On the other hand, the stable geometry of the $VE@C_2N$ complex shows the least interaction distance of 2.55 Å for H8---N2 atoms. In this case, the H-atoms of VE are inclined towards C_2N surface, thus interacting with the N-atoms of C_2N .

Similarly, the most stable configuration of the VG@C₂N complex was obtained after optimizing several interactive modes on C₂N surface. The stable geometry of the VG@C₂N complex reveals that the H-atoms of VG interact with the N-atoms of the surface. H10 and H11 atoms of VG both show strong interactions with N-atoms of C₂N, i.e., H10---N3, H10---N4, H7---N1, and H7---N2 with interaction distances of 2.42 Å, 2.72 Å, 2.84 Å, and 2.40 Å, respectively. VG@C₂N complex also shows interaction distances of 2.84 Å (O9---C5) and 2.90 Å (H8---H6).

In the case of the VM@C₂N complex, the H-atoms of the ethyl group of VM were heading towards the N-atom's surface, whereas the remaining part of the VM analyte flipped away from the surface (C₂N), giving an umbrella-like appearance. Strong H----N interactions are observed at interaction distances of 2.86, 2.79, and 2.913 Å for H7---N1, H6---N2, and H6---N3, respectively, between the H-atoms of VR and the six nitrogen atoms of the C₂N cavity. One weak van der Waals interaction is observed at a bond distance of 2.66 Å (H5---H4).

For all V-series@C₂N, the interaction energy values reveal that all analytes are physiosorbed on C₂N surface. The strongest interaction from studied V-series nerve agents is observed for VG nerve agents with an interaction energy of -17.81 kcal/mol. The interaction energy trend for studied V-series nerve agents is VG@C2N > VM@C2N > VX@C2N > VE@C2N > VS@C2N.

In the case of $VS@C_2N$ and $VE@C_2N$ complexes, the longer bond distance between analytes and C_2N atoms resulted in a lowering of interaction energy after complexation as compared to the rest of the studied V-series nerve agents.

3.2. Non-Covalent Interactions (NCI) Analysis

NCI analysis is a useful tool for characterizing the nature of interactions, and it helps in differentiating between different nonbonding interactions, such as weak van der Waals interactions, H-bonding, and repulsive interactions. The NCI analysis gives a clear explanation of non-covalent interactions based on the second eigen value with the first differential of electron density (ρ), known as RDG (reduced density gradient) [74]. In NCI analysis, the 2D-RDG graphs are observed by plotting the reduced density gradient (RDG) at the y-axis versus sign $\lambda 2(\rho)$ at the x-axis. In sign $\lambda 2(\rho)$, ρ indicates the bonding strength, whereas sign λ 2 gives the information regarding the type of bonding. For repulsion, the positive value of sign $\lambda 2(\rho)$ indicates the presence of weak van der Waals interactions. The color map in the 3D isosurface is also dependent on the value of sign $\lambda 2(\rho)$ of NCI analysis. If the values are positive, i.e., sign $\lambda 2(\rho) > 0$, red isosurfaces result, indicating steric repulsive interaction. Negative values, i.e., sign $\lambda 2(\rho) < 0$, resulted in the appearance of green isosurfaces, which represent van der Waals interactions. The appearance of blue isosurfaces in 3D images among interacting fragments of complex resulted from a large negative value of sign $\lambda 2(\rho)$, i.e., sign $\lambda 2(\rho) > -0.02$, which corresponds to strong electrostatic interactions [69].

The 2D-RDG graphs and colored 3D isosurfaces are presented in Figures 4 and 5, respectively. In NCI analysis, the colored map reveals the appearance of a green isosurface between C2N surface and V-series nerve agents, which indicates the existence of weak van der Waals interactions. The 3D green isosurfaces in the case of VG@C₂N, VM@C₂N, and VX@C₂N complexes are more prominent as compared to the other two complexes, which shows the stability of these complexes, which is in accordance with the results of interactions energy. Furthermore, 2D-RDG plots depict that the projection of scattered green spikes in all V-series@ C₂N complexes appears in the range of 0.00 a.u. to -0.015 a.u., which confirms weak van der Waals interactions in all complexes. The existence of steric repulsion in all complexes is also confirmed through red-colored 3D isosurfaces, which are observed in the center of pyrazine and benzene rings of the C₂N unit (Figure 4).

Similarly, more negative values of λ_2 and deep RDG confirms the presence of strong electrostatic interactions, specifically hydrogen bonding. 2D NCI graphs depict that the spikes appear at sign $\lambda_2(\rho) > -0.01$ (a.u.). which presents strong electrostatic interactions, whereas, below this negative value, London dispersion force exists.



VM@C₂N

Figure 4. NCI 3D isosurfaces of optimized geometries of stable V-series@C₂N complexes computed at M05-2X method (iso value 0.05 a.u.) here red color is for repulsive interaction, green color indicates weak van der Waal's forces interactions, and blue color is for strong electrostatic interactions.

0.020

0.015

0.010

0.005

0.000

-0.005

-0.010

-0.015

-0.020

-0.025

-0.030

-0.035





VS@C₂N









3.3. Quantum Theory of Atoms in Molecules (QTAIM) Analysis

The non-covalent interactions among V-series analytes and C₂N surface are further explored via Bader's quantum theory of atoms in molecules (QTAIM) analysis. The topological parameters employed to study the nature of non-covalent interactions at BCPs are electronic density $\rho(\mathbf{r})$, Laplacian of electronic density $\nabla^2 \rho(\mathbf{r})$, local Potential energy V(r), local Lagrangian kinetic energy G(r), and total energy density H(r). For covalent bonds, values of ρ must be positive and greater than 0.1 a.u., while Laplacian ($\nabla^2 \rho$) is always a large value with a negative sign. On the contrary, for non-covalent interactions, the values of ρ are always less than 0.1 a.u. ($\rho < 0.1$ a.u.) and $\nabla^2 \rho$ is positive with small values [75,76]. Similarly, for covalent interactions, the ratio of -V/G > 2, whereas for non-covalent interactions, the ratio of -V/G must be less than 1 (-V/G < 1) [77].

Geometries of all complexes are optimized at M05-2X/6-31++G(d,p) level of theory to characterize QTAIM results. The results of topological parameters calculated via QTAIM analysis for V-series@C₂N complexes are reported in Table 2. Topological 3D isosurfaces of all studied V-series@C₂N complexes obtained through QTAIM analysis are shown in Figure 6. The values of electron density (ρ) and Laplacian ($\nabla^2 \rho$) reported in Table 2 justify the existence of non-covalent interactions in all studied V-series@C₂N complexes. A total of 11 BCPs are observed in the case of the VX@C₂N complex with six H---N, four H---C, and one S---C bond interactions. The values of electron density (ρ) and Laplacian ($\nabla^2 \rho$) are in the range of 0.004 to 0.012 a.u. and 0.011 to 0.037 a.u., respectively, which clearly indicates the existence of non-covalent weak interactions. The values of total energy density H(r), local Potential energy V(r), and local Lagrangian kinetic energy G(r) are also observed in the range of non-covalent weak interactions. The maximum number of BCPs are observed for the VS@C₂N complex, which is 14 (see Table 2). The values of electron density ρ for observed BCPs is in the range of 0.003 to 0.009 a.u., and the range of values for Laplacian $\nabla^2 \rho$ is 0.012 to 0.026 a.u.

Table 2. Topological parameters obtained through QTAIM analysis for studied V-series@C₂N complexes.

AnalyteC ₂ N	ρ (a.u)	∇ ² ρ (a.u)	G (r) (a.u)	V(r) (a.u)	H(r) (a.u)	-V/G	E _{int} (kcal/mol)
			VX@C	C ₂ N			
H19C11	0.007	0.022	0.004	-0.0035	0.0010	0.87	-1.10
H17N1	0.012	0.037	0.008	-0.0077	0.0007	0.96	-2.41
H20N10	0.007	0.022	0.005	-0.0037	0.0010	0.74	-1.16
H16N2	0.006	0.020	0.004	-0.0031	0.0009	0.77	-0.97
H15N6	0.006	0.022	0.004	-0.0035	0.0010	0.87	-1.10
H15N3	0.011	0.036	0.008	-0.0073	0.0009	0.91	-2.29
H17N8	0.006	0.021	0.004	-0.0032	0.0010	0.80	-1.00
H14C4	0.004	0.011	0.002	-0.0017	0.0006	0.85	-0.53
H13C5	0.005	0.017	0.004	-0.0021	0.0008	0.52	-0.66
S21C22	0.008	0.023	0.005	-0.0037	0.0010	0.74	-1.16
H18C12	0.005	0.015	0.003	-0.0023	0.0008	0.77	-0.72
			VS@C	C_2N			
H20C10	0.003	0.012	0.002	-0.0018	0.0006	0.90	-0.56
H22N1	0.004	0.014	0.003	-0.0019	0.0007	0.63	-0.59
H22N3	0.005	0.019	0.004	-0.0029	0.0009	0.72	-0.91
H20N12	0.005	0.018	0.004	-0.0029	0.0008	0.72	-0.91
H22N4	0.005	0.019	0.004	-0.0028	0.0009	0.70	-0.88
H22N8	0.006	0.021	0.004	-0.0032	0.0010	0.80	-1.00
H22N11	0.005	0.019	0.004	-0.0029	0.0009	0.72	-0.91
H19C10	0.004	0.014	0.003	-0.0020	0.0008	0.67	-0.62
H18C9	0.004	0.013	0.003	-0.0018	0.0007	0.60	-0.56
O16N8	0.008	0.026	0.006	-0.0049	0.0009	0.82	-1.54
H14C5	0.005	0.017	0.003	-0.0027	0.0008	0.90	-0.85
H14C6	0.005	0.017	0.003	-0.0027	0.0008	0.90	-0.85
H21C13	0.003	0.012	0.002	-0.0018	0.0006	0.90	-0.56
O16N4	0.009	0.032	0.007	-0.0067	0.0007	0.96	-2.10
O16C7	0.008	0.029	0.006	-0.0055	0.0009	0.92	-1.72

AnalyteC ₂ N	ρ (a.u)	∇²ρ (a.u)	G (r) (a.u)	V(r) (a.u)	H(r) (a.u)	-V/G	E _{int} (kcal/mol)
			VE@C	C_2N			
H9N3	0.002	0.008	0.002	-0.0011	0.0005	0.55	-0.34
H9N2	0.002	0.008	0.002	-0.0010	0.0005	0.50	-0.31
S12C6	0.003	0.009	0.002	-0.0011	0.0005	0.55	-0.34
H10N4	0.002	0.008	0.002	-0.0011	0.0005	0.55	-0.34
H11N1	0.003	0.010	0.002	-0.0014	0.0006	0.70	-0.44
H11N5	0.003	0.010	0.002	-0.0014	0.0006	0.70	-0.44
			VG@0	C_2N			
H12N10	0.007	0.025	0.005	-0.0040	0.0010	0.80	-1.25
O14C6	0.004	0.014	0.003	-0.0023	0.0007	0.77	-0.72
O16C4	0.007	0.025	0.005	-0.0040	0.0011	0.80	-1.25
H13N1	0.011	0.033	0.008	-0.0070	0.0007	0.87	-2.19
H15N3	0.011	0.036	0.008	-0.0073	0.0008	0.91	-2.29
H15N5	0.005	0.018	0.004	-0.0030	0.0008	0.75	-0.94
H13N2	0.006	0.020	0.004	-0.0034	0.0008	0.85	-1.07
H11C7	0.006	0.019	0.004	-0.0029	0.0010	0.72	-0.91
H18C9	0.004	0.014	0.003	-0.0021	0.0007	0.70	-0.66
H17N8	0.006	0.020	0.004	-0.0035	0.0008	0.87	-1.10
H19N8	0.004	0.014	0.003	-0.0023	0.0006	0.77	-0.72
			VM@	C ₂ N			
O21C6	0.005	0.018	0.004	-0.0031	0.0008	0.77	-0.97
H16N12	0.005	0.017	0.003	-0.0025	0.0008	0.83	-0.78
O21C4	0.005	0.017	0.004	-0.0029	0.0007	0.72	-0.91
O21C5	0.005	0.018	0.004	-0.0031	0.0007	0.77	-0.97
O20N10	0.009	0.029	0.007	-0.0060	0.0006	0.86	-1.88
H17C13	0.007	0.023	0.005	-0.0037	0.0010	0.74	-1.16
H19C11	0.004	0.014	0.003	-0.0020	0.0008	0.67	-0.62
H16N2	0.006	0.019	0.004	-0.0029	0.0010	0.72	-0.91
H16N3	0.006	0.020	0.004	-0.0031	0.0010	0.77	-0.97
H16N7	0.005	0.016	0.003	-0.0024	0.0008	0.80	-0.75
H16N10	0.004	0.014	0.003	-0.0020	0.0007	0.67	-0.63
H15N1	0.005	0.018	0.004	-0.0029	0.0008	0.72	-0.91
H23N9	0.004	0.015	0.003	-0.0024	0.0007	0.80	-0.75

Table 2. Cont.

Similarly, six BCPs are present for the VE@C₂N complex with five H---N and one S---C bond interactions. The strongest interactions are obtained for H11---N1 and H11---N5 bonds with electron density ρ and Laplacian $\nabla^2 \rho$ values of 0.003 a.u. 0.010 a.u., respectively. In the case of the VG@C₂N complex, 11 BCPs are observed with seven H---N, two H---C, and two O---C bond interactions. The values of electron density (ρ) and Laplacian ($\nabla^2 \rho$) are in the range of 0.004 to 0.011 a.u. and 0.014 to 0.036 a.u., respectively. For the VM@C₂N complex, the total BCPs observed are 13, with seven H---N, three O---C, two H---C, and one O---N bond interaction (see Figure 6 & Table 2). The values of electron density (ρ) and Laplacian ($\nabla^2 \rho$) are observed in the range of 0.004 to 0.009 a.u. and 0.014 to 0.029 a.u., respectively, which indicate that only non-covalent interactions exist between VM analyte and C₂N surface.



VM@C₂N

Figure 6. QTAIM analysis of studied V-series@C₂N complexes and yellow lines between nerve agent molecules and C₂N sheet indicates bond paths, whereas orange-colored dots present on bond paths show the bond critical points (BCPs).

The ratio of -V/G is also calculated for each BCPs of all studied V-series@C₂N complexes. The highest individual -V/G values for studied complexes i.e., **VX@C2N**, **VS@C2N**, **VE@C2N**, **VG@C2N** and **VM@C2N** are 0.96, 0.96, 0.70. 0.91 and 0.86, respectively (Table 2). Moreover, the ratio of -V/G also indicates that the local potential energy V(r) parameter is dominant in all V-series@C₂N complexes. Potential energy mainly rises due to the rise in values of Laplacian and electron density. The values of Laplacian, electron density, and -V/G also indicate that non-covalent interactions exist in all studied complexes.

Apparently, the observed values of all topological parameters, i.e., electron density $\rho(r)$, Laplacian $\nabla^2 \rho(r)$ total energy density H(r), Potential energy V(r), and Lagrangian kinetic energy G(r) for all studied V-series@C₂N complexes indicate that only non-covalent interactions exist among nerve agents and 2D surface. Therefore, all the V-series nerve agents are physiosorbed on C₂N surface. Among studied V-series nerve agents, the lowest BCPs are examined in the case of the VE@C₂N complex, i.e., six. Furthermore, the topological parameters such as ρ , $\nabla^2 \rho$, G, V, and H have the lowest values in cases of VE@C₂N complex as compared to the rest of V-series@C₂N complexes.

3.4. SAPTO Analysis

Symmetry adapted perturbation theory (SAPT0) analysis is used to characterize the interactions quantitatively between V-series nerve agents and C₂N surface. SAPT0 analysis consists of four contributing factors (interaction energies), i.e., electrostatic (ΔE_{elst}), exchange (ΔE_{exch}), induction (ΔE_{ind}), and dispersion (ΔE_{disp}) [78]. The sum of these four components, E_{elst} , E_{exch} , E_{ind} , and E_{dis} , gives the total SAPT0 energy (E_{SAPT0}) [79]. The contribution of each interaction energy component is obtained through SAPT0 analysis, and values are reported in Table 3, whereas a graphical representation of V-series@C₂N complexes is shown in Figure 7.

Table 3. SAPT0 analysis of V-series@C₂N complexes (kcal/mol).

Complexes	ΔE_{elst}	%	ΔE_{exch}	ΔE_{ind}	%	ΔE_{dis}	%	ΔE_{SAPT0}
VX@C ₂ N	-13.18	26.60	23.28	-4.49	9.06	-31.88	64.34	-26.27
VS@C ₂ N	-10.28	21.24	22.20	-5.12	10.59	-32.99	68.17	-26.20
VE@C2N	-10.66	26.18	18.23	-3.47	8.52	-26.58	65.29	-22.49
VG@C ₂ N	-15.39	29.20	25.14	-5.83	11.06	-31.48	59.73	-27.56
VM@C ₂ N	-9.37	21.39	19.75	-4.62	10.55	-29.80	68.05	-24.04



Figure 7. SAPT0 analysis of V-series@C₂N complexes graphical representation.

The highest contribution of the E_{exch} component is observed in the VG@C₂N complex (25.14 kcal/mol), followed by VX@C₂N, VS@C₂N, VM@C₂N, and VE@C₂N complexes with E_{exch} values of 23.28, 22.20, 19.75, and 18.23 kcal/mol, respectively. The contribution of remaining energy components for the VX@C₂N complex are -13.18 kcal/mol (E_{elst}), -4.49 kcal/mol (E_{ind}), and -31.88 kcal/mol (E_{disp}). SAPT0 results clearly indicate that the dispersion component is the major stabilizing factor (64.34%), followed by the E_{elst} component, which contributes 26.60%. Similarly, for the VS@C₂N complex, the values of E_{elst} , E_{disp} , and E_{ind} are -10.28, -32.99, and -5.12 kcal/mol, respectively. In the case of the VE@C₂N complex, again, E_{disp} is a dominant contributing factor with 65.29% (-26.58 kcal/mol) contribution. Whereas E_{elst} and E_{ind} are less dominant towards total SAPT0 with 26.18% (-10.66 kcal/mol) and 8.52% (-3.47 kcal/mol) contribution, respectively. VM@C₂N complex again follows the same trend of SAPT energy components, where the E_{disp} factor shows the highest contribution (68.05%), while E_{elest} and E_{ind} indicate less contribution of 21.39% and 10.55% towards total SAPT0, respectively.

SAPT0 analysis indicates that the majority of SAPT0 energy components are negative, which reveals that attractive forces are more dominating between analytes and C₂N surface (see Table 3). Whereas the exchange component (ΔE_{exch}) shows positive values, which reveals the presence of repulsive force between two interacting components. The highest stabilization energy is observed in the case of the VG@C₂N complex among E_{SAPT0} energy values, which is in accordance with interaction energy (E_{int}) analysis (see Table 1). The overall order of the SAPT0 component's contribution towards total SAPT0 energy (E_{SAPT0}) is $E_{disp} > E_{elest} > E_{ind}$. This trend indicates that the major stabilizing factor among SAPT0 components is E_{disp} .

4. Electronic Properties

4.1. Natural Bond Orbital (NBO) Analysis

The NBO analysis is carried out to analyze the quantity of charge transfer after the complexation of V-series nerve agents with C_2N surface. Electronic properties play an important role in understanding the nature of interactions existing among nerve agents and C_2N surface. The values of NBO charges upon the interaction of analytes and surface are given in Table 3. The negative value of NBO charges shows that the charge is transferred from surface to analyte and vice versa [80].

NBO analysis reveals that the values of NBO charges appeared in the range of -0.023 e^- to -0.002 e^- . Here the negative sign indicates that, for all V-series@C₂N complexes, the charge is transferred from C₂N surface to V-series agents. The NBO charge values of studied nerve agents are -0.023 e^- , -0.020 e^- , -0.013 e^- , -0.012 e^- , and -0.002 e^- for VX, VS, VE, VG, and VM, respectively (Table 4). This trend is observed due to charge transfer upon the interaction of the positively charged H-atoms of nerve agents (VX, VS, VE, VG, and VM) with the electron-rich C₂N surface. The highest amount of charge transfer (-0.023 e^- and -0.020 e^-) was examined for VX@C₂N and VS@C₂N complexes, respectively, which might be due to the strong electrostatic interaction (hydrogen bonding) of H-atoms with N-atoms of C₂N cavity. Furthermore, small NBO charge transfer in the London dispersion force case of VM@C₂N complex reveals the existence of weaker non-covalent interactions.

Table 4. FMO and NBO results of most stable V-series@C₂N complexes.

Complexes	HOMO eV	LUMO eV	E _{H-L}	NBO (e ⁻)
VX@C ₂ N	-5.02	-2.89	2.13	-0.023
VS@C ₂ N	-5.37	-2.74	2.63	-0.020
VE@C ₂ N	-5.35	-2.88	2.47	-0.012
VG@C ₂ N	-5.56	-2.75	2.81	-0.013
VM@C ₂ N	-5.57	-2.78	2.80	-0.002
C_2N	-6.40	-2.69	3.71	-

4.2. Electron Density Differences (EDD)

The type of interactions upon adsorption of nerve agents on C_2N surface was further characterized through Electron Density Difference (EDD) analysis. EDD plots of V-series@ C_2N complexes (Isovalue = 0.004 a.u.) are given in Figure 7. The isosurfaces were obtained using multiwfn 3.7 software(Tian Lu, Beijing Kein Research Center for Natural Sciences). Electron density difference was calculated through the variance of electron density among V-series@ C_2N complexes and the aggregate electron density of bare C_2N surface and isolated V-series nerve agents.

The appearance of blue and purple isosurfaces in EDD analysis indicates orbital interaction between C_2N surface and nerve agents (Figure 8). Blue isosurfaces depict an accumulation of electronic density, whereas purple isosurfaces show a depletion of electron density. Blue isosurfaces appeared due to the electrostatic interaction of H-atoms of considered V-type nerve agents and N-atoms of C_2N surface, which resulted in a higher accumulation of electron density among nerve agents and C_2N surface. Moreover, blue surfaces also reveal sigma (σ) donation of charge from N-atoms of C_2N surface to V-type nerve agents, thereby confirming charge transfer from surface towards analytes. The appearance of purple isosurfaces has also been noticed in Figure 8, which indicates the reduction of electron density at the interacting H-atoms of V-series agents. EDD plots revealed electronic density shifting among analytes and surface rings C_2N , i.e., benzene and pyrazine rings. Electron Density Difference (EDD) results are also validated through charge transfer (NBO) analysis.

4.3. Frontier Molecular Orbital (FMO) Analysis

Upon complexation with V-series nerve agents, the variation in electronic properties of C_2N sheet is evaluated via frontier molecular orbitals analysis. FMO analysis was employed to study the conductivity of the considered material through a change in band gap. Generally, conductivity rises with the decrease in the energy gap (HOMO-LUMO gap) and vice versa [81].

The values of HOMO-LUMO energies in a.u. and eV and their differences (band gap) in eV are reported in Table 4. Isosurfaces of HOMO and LUMO for studied complexes are presented in Figure 9. For bare C₂N surface, the HOMO energy is -6.40 eV, the LUMO energy is -2.69 eV, and the energy gap (E_{H-L}) for bare C₂N surface is **3.71** eV. The HOMO-LUMO energy gaps (E_{H-L}) of V-series@C₂N complexes are **2.13** eV (VX@C2N), **2.63** eV (VS@C2N), **2.47** eV (VE@C2N), **2.81** eV (VG@C2N), and **2.80** eV (VM@C2N). For all studied V-series nerve agents, an appreciable reduction in the HOMO-LUMO energy gap (E_{H-L}) is observed upon the adsorption of V-series analytes on C₂N surface.

As a result of the complexation of C₂N surface with analyte VX, the HOMO energy increases from -6.40 eV to -5.02 eV, and LUMO energy reduces from -2.69 eV to -2.89. Moreover, the HOMO-LUMO energy gap (E_{H-L}) is changed from 3.71 eV to 2.13 eV. Similar behavior with slight differences is observed for the rest of the V-series nerve agents in terms of variation in energies of HOMO and LUMO and energy gap (E_{H-L}). Upon adsorption of VS over C₂N, the energy of HOMO increases (-5.37 eV), which results in a reduced E_{H-L} gap of 2.63 eV, compared to 3.71 eV for isolated C₂N. Similarly, the interaction of VE with C₂N surface increases the HOMO energy value and decreases LUMO energy; E_{H-L} is reduced to 2.47 eV (see Table 4). Moreover, VG and VM nerve agents also show the same trend upon complexation; HOMO energy values increase to -5.56 eV and -5.57, and LUMO energy values reduce to -2.75 eV and -2.78 eV, respectively. The E_{H-L} are also reduced to 2.81 eV and 2.80 eV for VG and VM complexes, respectively. Among studied V-series@C₂N, the most prominent HOMO-LUMO energy gap (E_{H-L}) reduction (2.13 eV) is observed for the VX@C₂N complex.



VX@C₂N

VS@C₂N





VM@C₂N

Figure 8. EDD plots of V-series@ C_2N with Iso value = 0.004 a.u. (Blue isosurfaces show accumulation of electron density, while purple isosurfaces represent depletion of electron density).









VE@C₂N

Figure 9. Cont.



VM@C₂N

Figure 9. HOMO-LUMO densities of the studied V-series@C₂N surface.

In all studied V-series@C₂N complexes, HOMO orbital density is located over C₂N surface, whereas, in the case of LUMO, the orbital density is mostly distributed on V-series nerve agents (analytes). Overall, FMO data clearly displays that all studied V-series nerve agents show an appreciable reduction in the E_{H-L} gap upon complexation, which revealed that C₂N surface shows higher sensitivity towards V-series nerve agents.

5. Recovery Time

The stability of an ideal sensor can be measured through its ability to reprocess. Suitable recovery time is essential for the adsorption of nerve agents because the very high recovery response of a sensor leads to poisoning of the surface, whereas a very short recovery time does not provide appreciable time for the analyte to stay on the surface. The recovery response time of a sensor is theoretically calculated via transition theory which is given by the equation:

$$\tau = \upsilon^{-1} exp\left(\frac{-E_{ads}}{KT}\right) \tag{4}$$

where τ , υ , K, E_{ads}, and T represent recovery time, attempt frequency, Boltzmann constant interaction energy, and temperature of recovery, respectively. To evaluate the recovery time of C₂N, an attempt frequency of 10^{12} s⁻¹ is applied [82–84]. The recovery times (τ) of V-series@C₂N complexes are calculated by using Equation (4) at three different temperatures, i.e., 298, 350, and 400 K. Recovery times of 8.53 s, 11.40 s, and 9.29 s are observed in the case of VX@C₂N, VG@C₂N, and VM@C₂N complexes, respectively. Recovery time improves by rising temperature, i.e., the obtained values of recovery time at 400 K are 4.30×10^{-3} s,

 5.33×10^{-3} s, and 4.58×10^{-3} for VX@C₂N, VG@C₂N, and VM@C₂N complexes, respectively. However, the recovery time results seem much better compared to reported values for other surfaces. A short recovery time of 0.63×10^{-6} s at room temperature for the desorption of G-series nerve agent (GF) from graphdiyne (GDY) surface has been reported in the literature [85]. Moreover, for the desorption of NO₂ from C₃N surface, 6.8 s of recovery time is required. Similarly, a recovery time of 102 s is needed for the desorption of CO from Au-MoS₂ surface at room temperature [86,87]. The C₂N surface has been investigated previously by our research group, and very short recovery times of 0.027 s, 0.012 s, and 0.073 s at 298 K for VR, NM, and GF analytes, respectively, had been calculated [25,47]. Desorption results of V-type nerve agents indicates that C₂N surface can act as a potential candidate as a sensor with suitable recovery time.

6. Conclusions

In the present study, the sensing ability of carbon nitride quantum dots (C_2N) is carried out against V-type nerve agents through DFT at M05-2X/6-31++G(d,p) level of theory. Interaction energy (E_{int}) values of optimized geometries predict that all studied V-series@C₂N complexes are stable (thermodynamically), and adsorption is exothermic. The results indicate that the VE@C₂N complex is the most stable complex with the highest interaction energy of -17.81 kcal/mol. The stability of the VE@C₂N complex is attributed due to the presence of strong electrostatic forces, as revealed by QTAIM analysis. Laplacian and electron density values and -V/G ratio of all complexes characterized via QTAIM analysis indicate that only non-covalent interactions exist between V-series and C₂N units. Non-covalent interaction (NCI) studies depicted by the presence of green spikes revealed weak van der Waals interactions among interacting fragments. FMO analysis indicated that an appreciable decrease in the HOMO-LUMO energy gap (E_{H-L}) occurred for all studied complexes. Reduction in band gap also reveals that C₂N surface is highly sensitive and selective towards V-type nerve agents. A short recovery response time of 3.01 ms at 298 K is predicted for the desorption of VS from C₂N. The key findings distinctly indicate a better performance of C_2N surface as an electrochemical sensor towards the VX analyte. We believe that these results will play a crucial role for an experimentalist to tailor a highly selective electrochemical sensor using C₂N surface.

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