

Supporting Information

Dendritic Mesoporous Silica Hollow Spheres for Nano-Bioreactor Application

Qian Zhang ¹, Minying Wu ¹, Yuanyuan Fang ¹, Chao Deng ², Hsin-Hui Shen ³, Yi Tang ^{1,*} and Yajun Wang ^{2,*}

¹ Department of Chemistry, Fudan University, Shanghai 200433, China; 13110220006@fudan.edu.cn (Q.Z.); 18110220035@fudan.edu.cn (M.W.); 19110220078@fudan.edu.cn (Y.F.)

² College of Chemistry & Materials Engineering, Wenzhou University, Wenzhou 325027, China; dengchao@wzu.edu.cn

³ Department of Materials Science and Engineering, Monash University, Clayton, VIC 3800, Australia; hsin-hui.shen@monash.edu

* Correspondence: yitang@fudan.edu.cn (Y.T.); yajunwang@wzu.edu.cn (Y.W.)

Synthesis of BaSO₄ NP-S and BaSO₄ NP-L

Based on the synthesis of BaSO₄ NP, when using a Ba-EDTA solution of pH 6.0, the sample denoted as **BaSO₄ NP-S** was obtained. And when using a Ba-EDTA solution of pH 9.0, 50 mL of Na₂SO₄ and 200 mL of Ba-EDTA, the **BaSO₄ NP-L** was obtained. As shown in Figure S1a, most of the BaSO₄ NP-S have the shape of an ellipse other than spherical morphology of BaSO₄ NP used above, while a few particles of the BaSO₄ NP-L are elliptic in Figure S1c. The conditional stability constant of the complex of Ba²⁺ and EDTA ions at pH 9.0 is higher than at pH 6.0, resulting in a slower rate of the precipitation of BaSO₄ and smaller size. When amplifying the reaction system at pH 6.0, same with BaSO₄ NP, once the large volumes of reaction solution was added, a faster rate and local high concentration led to a larger particle and wider size distribution.

Specific Enzyme Activity Measurement

First, immobilized or free catalase was dispersed into a 2 mL of phosphate buffered solution (PBS, 50 mmol L⁻¹, pH 7.0, 25 °C) containing H₂O₂ (20 mmol L⁻¹), and the concentration of catalase was 2.0 μg mL⁻¹. The absorbance at 240 nm was monitored to track changes in H₂O₂ concentration. The standard curve is shown in Figure S7a.

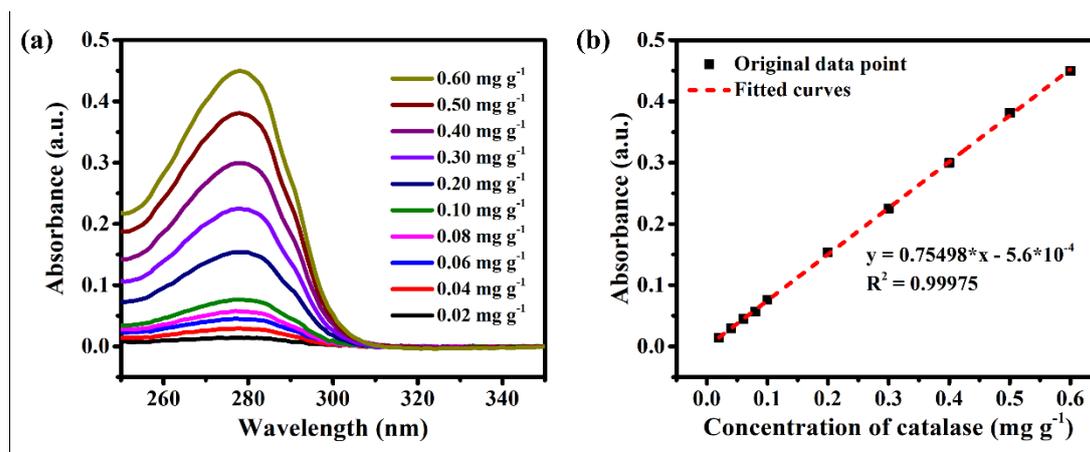


Figure S1. (a) UV-vis spectra of standard catalase solution and (b) the corresponding standard curve.

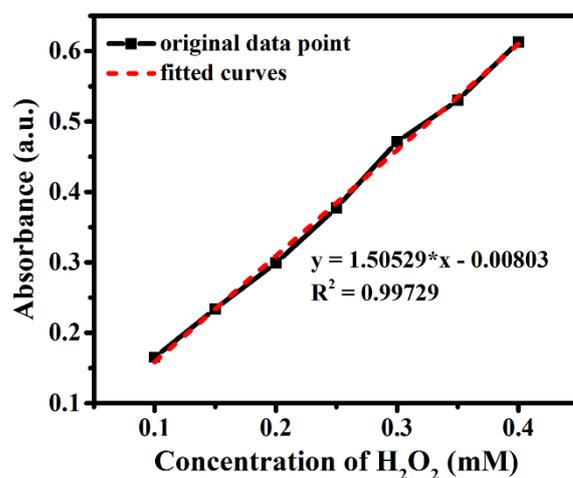


Figure S2. The standard curve for the H₂O₂ detection using HRP-TMB method.

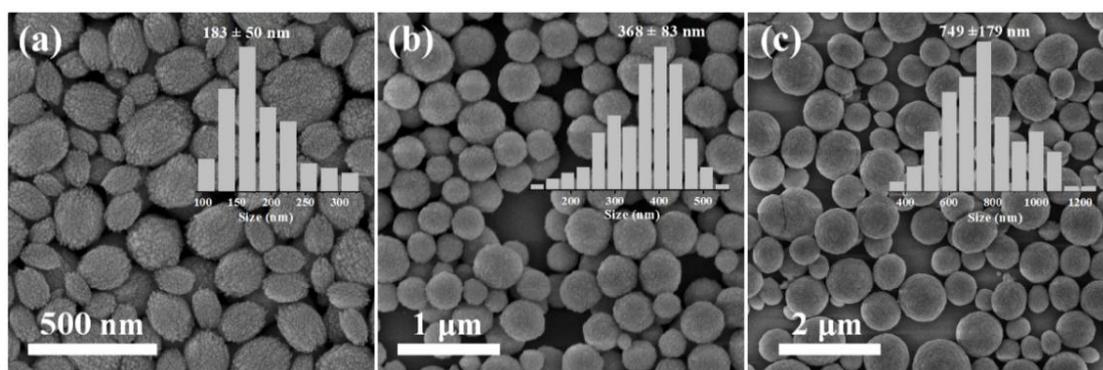


Figure S3. SEM images of different BaSO₄ NPs: (a) BaSO₄ NP-S, (b) BaSO₄ NP, and (c) BaSO₄ NP-L, and their corresponding particle size distributions (insets) obtained by the SEM images statistics. The numbers of particles counted for BaSO₄ NP-S, BaSO₄ NP, and BaSO₄ NP-L were 109, 127, and 139, respectively.

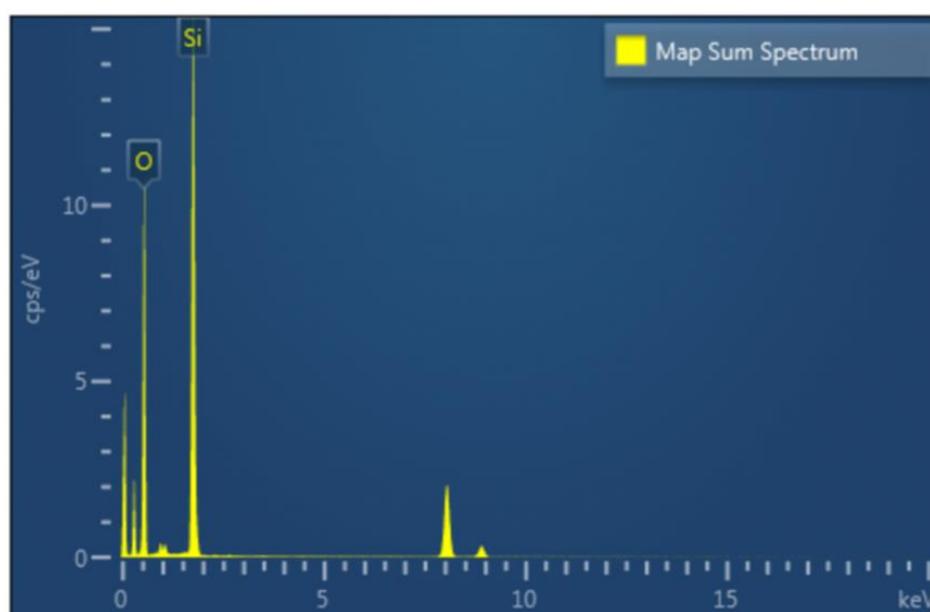


Figure S4. EDX spectrum of HMSN.

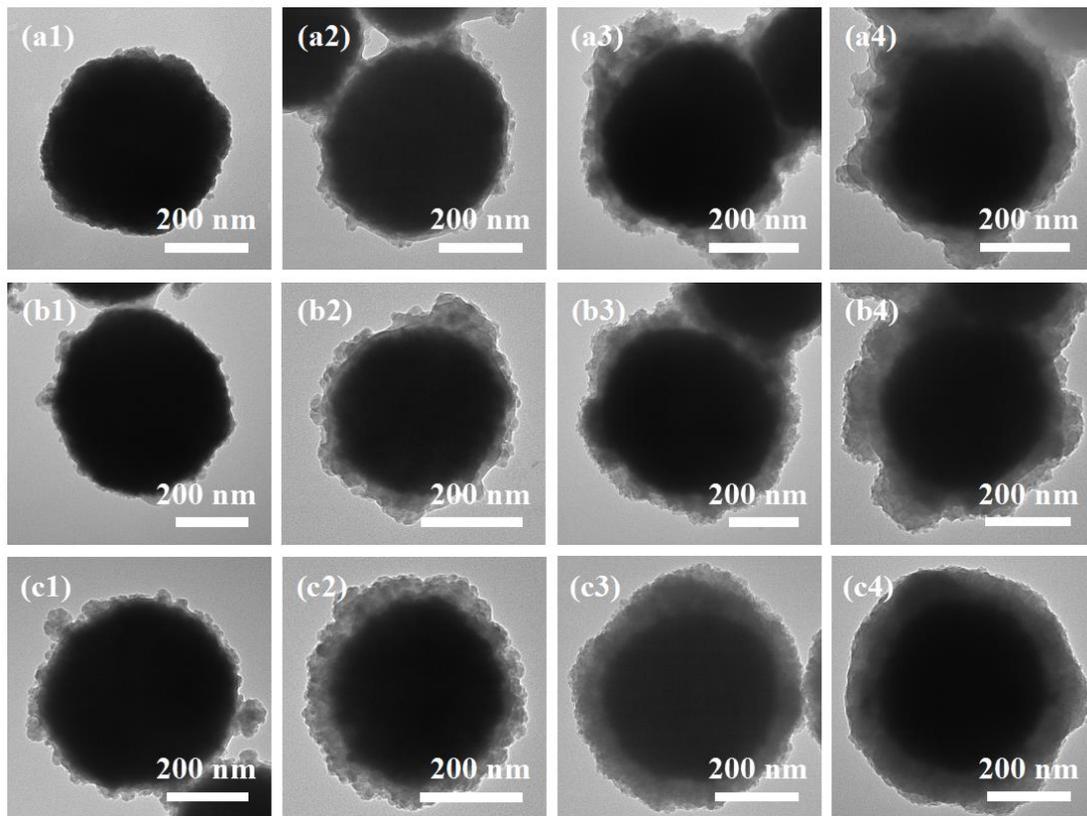


Figure S5. TEM images of BaSO₄@APF-SiO₂ with different reaction time: 15 min (**a1**, **b1**, and **c1**), 30 min (**a2**, **b2**, and **c2**), 60 min (**a3**, **b3**, and **c3**), and 120 min (**a4**, **b4**, and **c4**). (**a**), (**b**), and (**c**) correspond to different TEOS addition of 0.545, 1.095, and 2.190 mL.

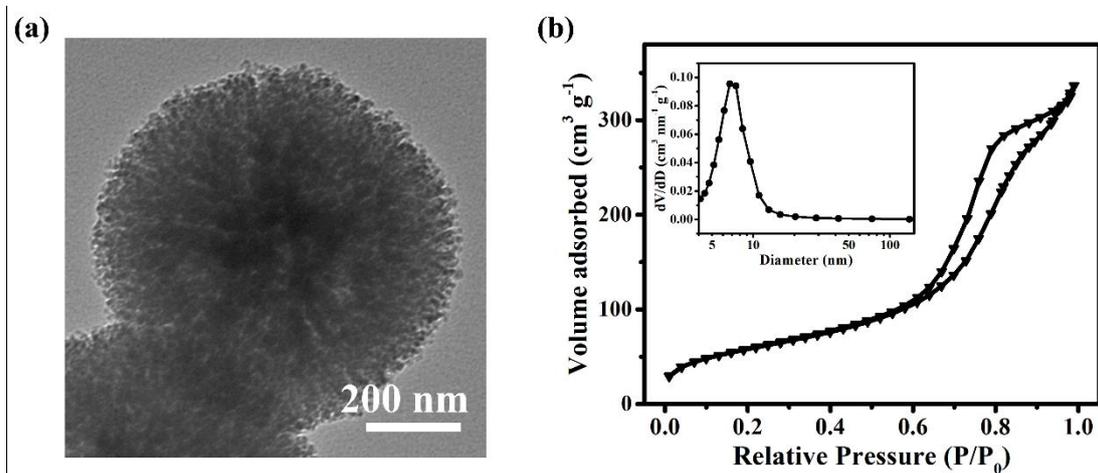


Figure S6. (a) TEM image of SMSN; (b) N₂ sorption isotherm and pore size distribution (inset) of SMSN calculated from desorption branch with BJH model.

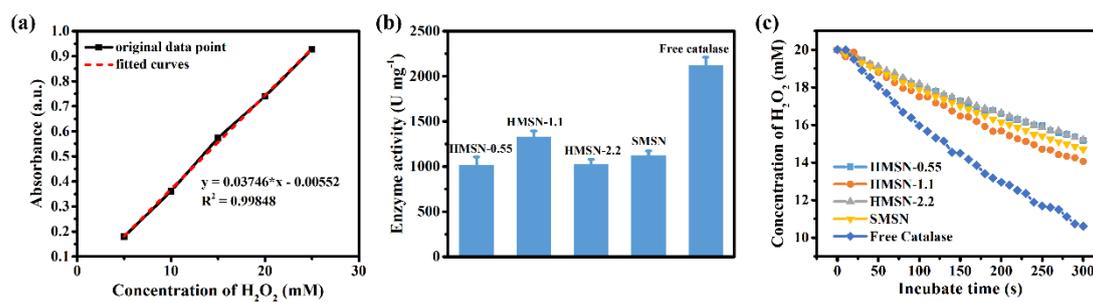


Figure S7. (a) Standard curve for H₂O₂ detection by UV-vis at 240 nm; (b) Enzyme activity of the catalase loaded in HMSN-0.55, HMSN-1.1, HMSN-2.2, and SMSN, and free catalase; (c) the dynamic experiments for the calculation of enzyme activities in (b).