



Article Structural and Optical Sensing Properties of Nonthermal Atmospheric Plasma-Synthesized Polyethylene Glycol-Functionalized Gold Nanoparticles

Linh Nhat Nguyen ^{1,2,3}, Pradeep Lamichhane ^{1,2}, Eun Ha Choi ^{1,2}, and Geon Joon Lee ^{1,2,*}

- ¹ Department of Electrical and Biological Physics, Kwangwoon University, Seoul 01897, Korea
- Plasma Bioscience Research Center, Kwangwoon University, Seoul 01897, Korea
- ³ Laboratory of Plasma Technology, Institute of Materials Science, Vietnam Academy of Science and Technology, 18 Hoang Quoc Viet, Hanoi 100000, Vietnam
- * Correspondence: gjlee@kw.ac.kr; Tel.: +82-2-940-8619

Abstract: Polyethylene glycol-functionalized gold nanoparticles (Au@PEG NPs) were prepared by a simple plasma-assisted method without additional reducing chemicals. After irradiating tetrachloroauric acid (HAuCl₄) and polyethylene glycol (PEG) in aqueous medium with an argon plasma jet, the gold precursor transformed into an Au@PEG NP colloid that exhibited surface plasma resonance at 530 nm. When the plasma jet entered the water, additional reactive species were induced through interactions between plasma-generated reactive species and aqueous media. Interaction of the gold precursor with the plasma-activated medium allowed the synthesis of gold nanoparticles (AuNPs) without reductants. The plasma-synthesized Au@PEG NPs had a quasi-spherical shape with an average particle diameter of 32.5 nm. The addition of PEG not only helped to stabilize the AuNPs but also increased the number of AuNPs. Au@PEG NP-loaded paper (AuNP-paper) was able to detect the degradation of rhodamine B, therefore, indicating that AuNP-paper can act as a surface-enhanced Raman scattering platform. Dye degradation by plasma treatment was investigated by optical absorption and Raman spectroscopy. The method proposed for the fabrication of Au@PEG NPs is rapid, low-cost, and environment-friendly and will facilitate the application of plasma-synthesized nanomaterials in sensors.

Keywords: gold nanoparticles; plasma synthesis; polyethylene glycol; reactive oxygen species; surface plasmon resonance; surface-enhanced Raman scattering; optical sensing

1. Introduction

Metal nanoparticles are used in various applications, such as optical sensing, bioimaging, drug delivery, and photothermal therapy. Metal nanoparticle-based optical sensing is a noninvasive method that allows for the rapid diagnosis of biomaterials and cancers [1]. Owing to their outstanding physical and optical properties, gold nanoparticles (AuNPs) have become one of the most prominent nanomaterials in the nanotechnology field. AuNPs are used widely as surface-enhanced-Raman spectroscopy (SERS) platforms due to optical confinement effects stemming from their surface plasmon resonance [2,3]. Generally, AuNPs can be synthesized by chemical reduction of HAuCl₄ precursors. However, harsh reducing agents are used during chemical synthesis, and AuNP toxicity has become a serious concern [4]. To overcome this issue, the development of green synthesis processes that eliminate the use of toxic reagents during the preparation of AuNPs has become an important research direction [5,6]. Using green synthesis, environmentally and biologically friendly AuNPs suitable for a wide range of applications, including optical sensing, biomedical imaging, and physicochemical analysis, can be prepared. In this context, many studies have reported green preparations of gold nanoparticles using plant extracts, microorganisms, enzymes, polysaccharides, and other non-toxic molecules [7–11]. These



Citation: Nguyen, L.N.; Lamichhane, P.; Choi, E.H.; Lee, G.J. Structural and Optical Sensing Properties of Nonthermal Atmospheric Plasma-Synthesized Polyethylene Glycol-Functionalized Gold Nanoparticles. *Nanomaterials* **2021**, *11*, 1678. https://doi.org/10.3390/ nano11071678

Academic Editor: Paolo Pengo

Received: 12 May 2021 Accepted: 22 June 2021 Published: 25 June 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). substances contain large amounts of hydroxyl and phenol groups that reduce Au ions [12]. Generally, however, these non-hazardous substances are weak reducing agents, thus, resulting in longer processing times compared with that of chemical synthesis [13]. In this sense, non-thermal plasma is a beneficial alternative to synthesize AuNPs, because toxic agents are not used, and the processing time is shorter than that required for chemical synthesis.

The non-thermal plasma synthesis of metal nanostructures is an alternative green synthesis approach that has substantial advantages compared to chemical synthesis [14–17]. This method uses non-thermal atmospheric pressure plasmas under ambient conditions and produces large amounts of reactive oxygen and nitrogen species, excited metastable species, free electrons, and charged particles [18]. These species can be transferred into the liquid phase when the plasma is in contact with liquid medium [19,20]. Dissolved reactive species are key contributors to the chemical reactions that occur during the plasma synthesis of metallic nanoparticles in the plasma-activated water [21,22]. In the plasma synthesis of AuNPs, the reduction of Au³⁺ ions can be triggered both by short-lived solvated electrons and long-lived reactive species, such as H_2O_2 , and reduction of the gold precursor results in surfactant-free AuNPs [23]. Plasma synthesis of hybrid nanomaterials composed of AuNPs and a polymer has been investigated to improve the stability and biocompatibility of AuNPs [24–26]. Additional polymers play the role of stabilization agents that can assist the formation of AuNPs in the plasma-treated medium. The amount of plasma-synthesized AuNPs has been reported to be higher in HAuCl₄–polymer than in HAuCl₄ [25]. To improve the polyethylene glycol (PEG) bonding strength to the AuNP surface, it is necessary to modify PEG with thiol or amine groups [27]. For instance, Furusho et al. functionalized PEG with pentaethylenehexamine (PEHA) and used it as a polymer matrix for plasma synthesis of AuNPs [28]. PEG is a biocompatible polymer that has been extensively used for nanoparticle stabilization and surface modification [29]. There are numerous reports describing the preparation and applications of PEG-coated plasmonic nanomaterials, including optical sensing and biomedical applications [30,31].

In this study, we describe the synthesis of gold nanoparticles by plasma reduction of a HAuCl₄ precursor. The morphological and optical properties of AuNPs were examined by scanning electron microscopy (SEM) images and optical absorption spectra, respectively. PEG was used to improve the morphology and formation efficiency of nanoparticles during plasma synthesis of AuNPs. Next, we investigated the optical sensing properties of AuNPs using plasma-treated and untreated rhodamine B (RhB) dye. The degradation of RhB by plasma treatment was studied by optical absorption and Raman spectroscopy. For Raman spectroscopic measurements of plasma-treated RhB, AuNP-loaded paper was used as a SERS platform (Figure 1).

2. Experimental Details

2.1. Plasma Jet System

Figure 2 illustrates the experimental layout of the plasma jet used to synthesize the Au@PEG NPs in this work. The plasma jet was constructed by inserting a 12-cm-long stainless-steel syringe inside a 20-cm-long quartz tube. A copper piece was wrapped around the outside of the quartz tube and connected to the ground. A high-frequency AC sinusoidal voltage of 5 kV was applied between the steel syringe and the copper piece, and 99.999% argon at a flow rate of 700 sccm was used as a feeding gas to generate a plasma jet. Current and voltage waveforms were measured using a current probe (Tektronix, P6022, Beaverton, OR, USA) and a voltage probe (Tektronix, P6015A, Beaverton, OR, USA), respectively, both connected to a digital oscilloscope (Lecroy, WaveSurfer 434, New York, NY, USA). Optical emission spectra (OES) of the plasma plume were recorded by a high-resolution fiber optic spectrometer (Ocean Optics, HR4000, Orlando, FL, USA).

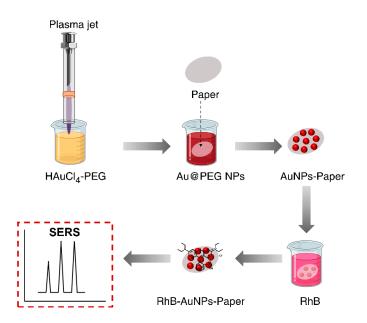


Figure 1. Schematic representation of the plasma-synthesis of polyethylene glycol-functionalized gold nanoparticles (Au@PEG NPs) and fabrication of a surface-enhanced Raman scattering platform from Au@PEG NPs.

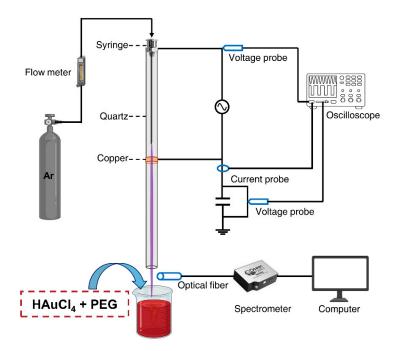


Figure 2. Schematic of the experimental setup for Au@PEG NP synthesis using a non-thermal atmospheric plasma jet.

2.2. Synthesis of Gold Nanoparticles by Plasma Jet

Tetrachloroauric acid trihydrate (HAuCl₄·3H₂O, \geq 99.9% trace metals basis) and polyethylene glycol (PEG, $M_n = 4000$) were purchased from Sigma Aldrich (Yongin-city, Kyunggi-do, Korea) and used without purification. All glassware was cleaned with aqua regia solution (a 3:1 mixture of hydrochloric acid and nitric acid) overnight and washed thoroughly with de-ionized (DI) water before use. Stock solutions of 10 mM HAuCl₄ and 50 mM PEG were prepared. A mixture of HAuCl₄ and PEG was prepared by adding 300 µL of 10 mM HAuCl₄ and 50 µL of 50 mM PEG to 4 mL of DI water. The HAuCl₄–PEG solution was treated with the non-thermal plasma jet for 10 min with vigorous magnetic stirring, which resulted in a dark purple Au@PEG solution. The obtained product was purified by centrifugation at 8000 rpm for 10 min, followed by removal of the supernatant.

2.3. Morphological and Optical Characterizations of Plasma-Synthesized Gold Nanoparticles

The SEM analysis of plasma-synthesized Au@PEG NPs was performed by a fieldemission SEM (JEOL, JSM–7001F, Tokyo, Japan) under 15 kV acceleration voltage. For better SEM observation, we functionalized Au@PEG NPs with bovine serum albumin (BSA) to prevent the aggregation of gold nanoparticles [32]. Briefly, 100 mL of Au@PEG NP solution was mixed with 100 mL of 300 μ M BSA. Ten microliters of the Au@PEG NP–BSA mixture was drop-cast onto a silicon wafer and dried under vacuum conditions. The absorption spectra of plasma-synthesized Au@PEG NP solutions were measured using an optical absorption spectrometer (Jasco, J–815, Tokyo, Japan).

2.4. Coating of Gold Nanoparticles on Filter Paper

Filter paper (Hyundai Micro, HM5301200, Seoul, Korea) was cut into 1.0-cm-diameter disk-shaped pieces and dipped in Au@PEG NP solution for 12 h at 4 °C. After dip-coating, the Au@PEG NP-loaded paper (AuNP-paper) was dried under ambient conditions.

2.5. Optical Properties of Rhodamine B before and after Plasma Treatment

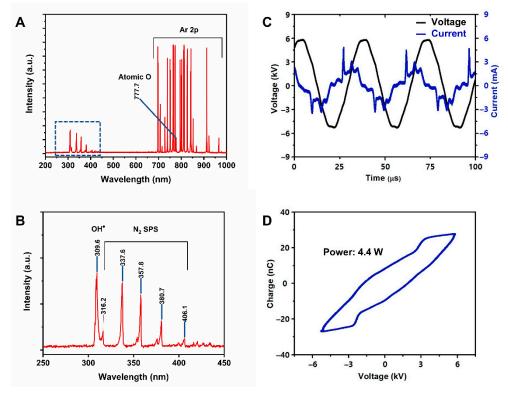
Rhodamine B (RhB, analytical standard) was purchased from Sigma Aldrich. A 10^{-4} M RhB stock solution was prepared by dissolving 4.8 mg of RhB powder in 100 mL DI water, and this solution was filtered to eliminate insoluble components. A 3-mL aliquot of 10^{-4} M RhB solution was treated with an Ar plasma jet for different amounts of times (0, 90, 180, and 600 s). The optical absorption spectra of plasma-treated and untreated RhB solutions were obtained by an optical absorption spectrometer (Jasco, J–815, Tokyo, Japan). A paper disk plate was dip-coated in the plasma-treated RhB solution for 12 h and dried at room temperature. Raman spectra of plasma-treated and untreated RhB on the AuNP-paper were measured at room temperature using a confocal Raman microscope (WITec, Alpha 300R, Ulm, Germany) with a 633-nm excitation light emitted from a He-Ne laser. In order to obtain the Raman spectra, the laser beam was focused on the sample using a microscope objective ($20 \times$).

3. Results and Discussion

3.1. Plasma Discharge Characterization

OES can provide important information about reactive species contained in a plasma jet. Figure 3A demonstrates the OES of the plasma jet over the wavelength range of 200–1000 nm; strong emission lines indicate the presence of plasma-generated reactive species. The 309-nm emission of the hydroxyl radical (OH) originated from dissociation of water vapor in ambient air [33]. Weak emission lines of the nitrogen second positive system (N₂–SPS; $C^3\Pi_u \rightarrow B^3\Pi_g$ transition of nitrogen molecules) were detected in the range of 330–380 nm [34] (Figure 3B), which is attributed to the fact that the plasma was operated under atmospheric conditions. Strong emission lines in the wavelength range of 700–1000 nm were attributed to the Ar $2p \rightarrow 1s$ transition, because argon was used as the working gas. These generated species can penetrate further into the liquid medium under the effect of gas flow and contribute to plasma–liquid interactions. Figure 3C shows the current-voltage profile of the AC-driven plasma discharge. The discharge current peaks appeared in the positive half-cycle of the applied voltage are due to the accumulation of positive surface charges while the polarity of these charges is reversed in the negative half-cycle of the applied voltage.

The measured peak voltage and peak current were about 6 kV and 5 mA, respectively. The period of voltage waveform was 35 μ s, which corresponds to a frequency of 28.5 kHz. We connected a 33-nF ceramic capacitor in series with the ground electrode and measured the voltage across this capacitor to calculate the accumulated charge. A charge-voltage Lissajous loop was constructed by plotting the accumulated charge versus the applied



voltage (Figure 3D). The dissipated power of the plasma jet was calculated to be 4.4 W based on the area of the Lissajous loop.

Figure 3. Characteristics of the plasma jet. (**A**,**B**) Optical emission spectrum, (**C**) current-voltage waveform, and (**D**) Lissajous curve of the Ar plasma jet.

3.2. Characteristics of Plasma-Treated Water for Plasma Synthesis of Gold Nanoparticles

When a plasma jet enters water, reactive species in the plasma phase are transported to the liquid phase. These solvated species significantly change the physicochemical properties of plasma-treated water and stimulate a series of complex chemical reactions. Previous studies have confirmed that hydrogen peroxide (H_2O_2) is a key factor in the plasma reduction process of Au^{3+} [23,25]. H_2O_2 is a long-lived species that is generated through several pathways [35-37]. In the plasma jet, gaseous H₂O₂ formed inside the quartz tube is delivered through the nozzle, and aqueous H_2O_2 is generated continuously at the gas-liquid interface and accumulated inside the bulk liquid, resulting in a high concentration of H_2O_2 . In the gas phase, plasma-generated OH radicals can recombine to form gaseous H_2O_2 , which can be transported to the gas-liquid interface (Equation (1)) [35]. H₂O₂ was observed at all positions along the plasma jet, as shown in Figure 4B. Most of the H_2O_2 formed in the plasma phase is immediately dissolved in water. At the gas-liquid interface, plasma-generated OH radicals can be converted to aqueous H_2O_2 , which can be transported into the liquid medium (Equation (2)) [35]. In the bulk liquid, OH radicals can be generated through plasma-induced ultraviolet (UV) photolysis of water, and solvated OH radicals can recombine to form H_2O_2 (Equation (3)) [35]. Under acidic conditions, plasma-generated superoxide can be converted to H_2O_2 (Equation (4)) [36].

$$OH_{(g)} + OH_{(g)} \rightarrow H_2O_{2(g)} \tag{1}$$

$$OH_{(g)} + OH_{(g)} \rightarrow OH_{(aq)} + OH_{(aq)} \rightarrow H_2O_{2(aq)}$$
⁽²⁾

$$H_2O_{(aq)} \xrightarrow{UV} H_{(aq)} + OH_{(aq)}; OH_{(aq)} + OH_{(aq)} \rightarrow H_2O_{2(aq)}$$
(3)

$$O_{2(aq)}^{-} + HO_{2(aq)} + H^{+}_{(aq)} \to H_2O_{2(aq)} + O_{2(aq)}$$
(4)

6 of 13

Consequently, each of these four equations represents possible pathways to generate H_2O_2 by an atmospheric plasma jet; (1) H_2O_2 in gas phase, (2) H_2O_2 at gas–liquid interface, (3) H_2O_2 via UV photolysis of water, and (4) H_2O_2 via plasma-generated superoxide.

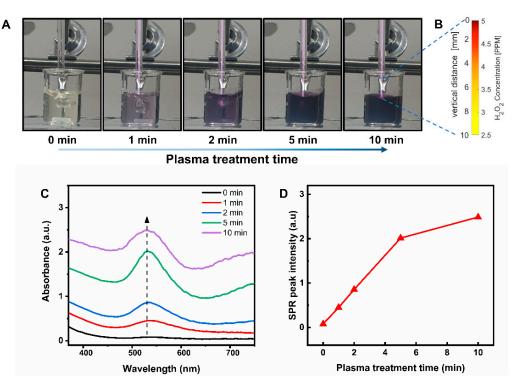


Figure 4. Effects of the plasma treatment time on the optical properties of the Au@PEG NP solution produced by plasma treatment of a HAuCl₄–PEG solution. (**A**) The color change of the Au@PEG NP solution over a plasma treatment time of 10 min. (**B**) Spatial distribution of the H₂O₂ concentration along the plasma jet. (**C**) Optical absorption spectra of the AuNP solutions with different plasma treatment times. (**D**) SPR peak intensity of the AuNP solution as a function of the plasma treatment time.

In the plasma-activated water, aqueous H_2O_2 can be produced by Equations (2)–(4) [35–37]. To verify the generation of H_2O_2 by the plasma jet, we treated 4 mL of DI water with a plasma jet for 10 min and quantified the H_2O_2 concentration of the plasma-treated water. By assessing the absorption spectrum of the plasma-treated water using a standard H_2O_2 detection colormetric assay, we determined the concentration of H_2O_2 to be about 3.3 mM. H_2O_2 concentration in the plasma-activated water is relatively high due to the accumulation of aqueous H_2O_2 generated at the gas–liquid interface and inside the bulk liquid [35–37].

3.3. Optical and Morphological Properties of Plasma-Synthesized Gold Nanoparticles

The prepared HAuCl₄–PEG solution was stable at room temperature for several hours with no significant change in color, suggesting no reduction of Au³⁺ ions. After the start of plasma treatment, the color of the HAuCl₄–PEG solution rapidly changed from light yellow to dark purple, indicating the formation of AuNPs (Figure 4A). The concentration of gold nanoparticles in solution was proportional to the absorption coefficient of the surface plasmon resonance (SPR). Therefore, the absorption spectra of HAuCl₄–PEG solutions with different plasma treatment times were measured to evaluate the nanoparticle formation (Figure 4C). The observed increase in the SPR peak intensity with the plasma treatment time is summarized in Figure 4D. These results demonstrate that plasma treatment was an

effective method to synthesize Au@PEG NPs. The plasma reduction mechanism of Au³⁺ to metallic Au⁰ has been well-established and is based on the following two reactions:

$$Au^{3+} + 3e^- \to Au^0 \tag{5}$$

$$2Au^{3+} + 3H_2O_2 \to 2Au^0 + 3O_2 + 6H^+$$
(6)

Equation (5) is a direct reduction of Au^{3+} to Au^0 by plasma electrons solvated in the liquid medium [38]. Equation (6) shows an indirect reduction of Au^{3+} through H_2O_2 that is formed in the medium due to plasma treatment [23]. As H_2O_2 is a stable species with a long lifetime, Equation (6) can proceed after the plasma is turned off. We also treated a HAuCl₄ solution with plasma without adding PEG. The color of the plasma-treated HAuCl₄ solution changed slightly to light orange, thereby, suggesting that the reaction occurred slower.

Figure 5A shows the optical absorption spectra of the AuNP solutions produced by plasma treatment of the HAuCl₄–PEG and HAuCl₄ solutions. The plasma-treated HAuCl₄-PEG solution exhibited an absorption peak at 530 nm, while the plasma-treated $HAuCl_4$ solution had an absorption peak at 567 nm. These results suggest that the average nanoparticle size of the plasma-treated HAuCl₄ solution was larger than when PEG was present. Small-sized Au⁰ seeds formed rapidly under plasma treatment and grew continuously, resulting in larger AuNPs. When there was no surfactant in the solution, AuNPs were free to grow in size during plasma treatment, resulting in larger AuNPs. When PEG was added, it functioned as a surfactant to stabilize AuNPs, thus, limiting their growth. The SPR peak intensity of plasma-treated HAuCl₄–PEG solution (optical density of 2.5) was much stronger than that of the plasma-treated HAuCl₄ solution (optical density of 0.8), indicating the formation of more AuNPs in the former solution. PEG likely functioned as a stabilizer and promoted the formation of AuNPs. This result is in agreement with a previous report of De Vos et al., who performed plasma synthesis of AuNPs with an additional polymer matrix [25]. They demonstrated that the amount of plasma-synthesized AuNPs was higher in HAuCl₄-PEG solution than in HAuCl₄ only solution. Based on the obtained data, the addition of PEG to the plasma-synthesis process of AuNPs affected not only the size of the resultant AuNPs but also the number of AuNPs.

The morphology of Au@PEG NPs was studied by SEM analysis. Figure 5B,C shows $60,000 \times$ and $160,000 \times$ magnification SEM images of Au@PEG NPs loaded on a silicon substrate, respectively. Plasma-synthesized Au@PEG NPs with a quasi-spherical shape were observed. The particle size distribution of Au@PEG NPs was obtained from SEM images using *ImageJ* software, v1.52a; National Institutes of Health, MD, USA, 2018 [39]. The average particle diameter of Au@PEG NPs was calculated as 32.5 ± 10.1 nm (Figure 5D). We noted that there was a small fraction of Au@PEG NPs with large non-spherical shapes (triangular and hexagonal), which were usually found in plasma-assisted nanoparticle synthesis. During the plasma treatment process, the ionic gold precursor concentration continuously depleted, while the number of formed metallic gold nanoparticles increased. When the precursor concentration is low, the anisotropic growth of the formed gold nanoparticles is favored over the nucleation of new nanoparticles [23,25].

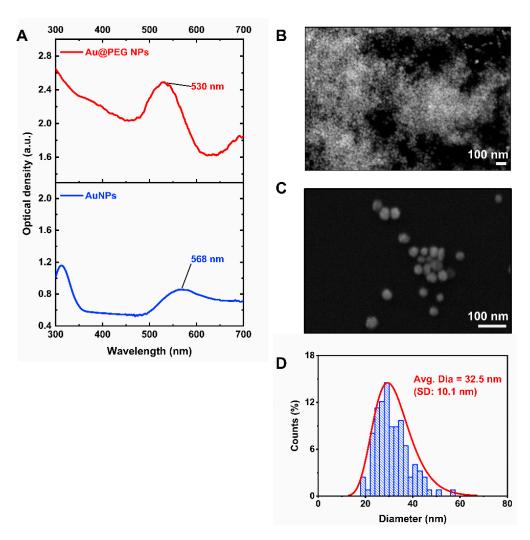


Figure 5. (A) Optical absorption spectra of the Au@PEG NP solution after plasma treatment for 10 min: HAuCl₄–PEG (red curve) and HAuCl₄ (blue curve). (B) $60,000 \times$ and (C) $160,000 \times$ SEM images of Au@PEG NPs. (D) The particle size distribution of Au@PEG NPs. The average particle size was 32.5 nm with a standard deviation of 10.1 nm.

3.4. Characteristics of Gold Nanoparticles Loaded on the Paper Disk

To investigate the optical sensing properties of AuNPs as a SERS platform, we deposited Au@PEG NPs on paper by dipping paper disks in Au@PEG NP colloids for 12 h. After dip-coating, the paper color changed from white to black, implying the successful deposition of Au@PEG NPs. A SEM image of AuNPs loaded on the paper is shown in Figure 6A. A large amount of Au@PEG NPs accumulated on the paper. We also performed energy-dispersive X-ray spectroscopy elemental mapping to confirm the existence of Au@PEG NPs on the paper (Figure 6B–D). Intense gold elemental signals (yellow dots) were detected across the surface of the AuNP-paper, indicating the homogenous deposition of Au@PEG NPs by the dip-coating method. O and C element signals from the paper disk plate were also present.

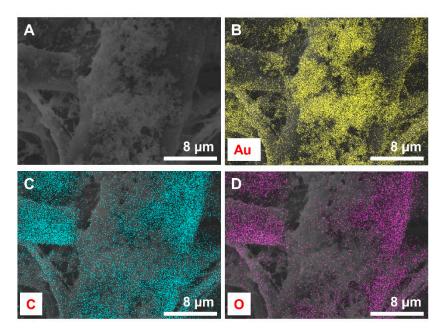


Figure 6. (**A**) 2300× SEM image of AuNP-paper and (**B**–**D**) the corresponding energy-dispersive X-ray spectroscopy elemental mapping, (**B**): Gold, (**C**): Carbon, and (**D**): Oxygen.

3.5. Optical Sensing Properties of Plasma-Treated Rhodamine B by Gold Nanoparticles

To study the optical sensing activity of plasma-synthesized gold nanoparticles, we investigated the effect of plasma treatment on rhodamine B (RhB). RhB was selected as a model analyte as it is a nonbiodegradable and commonly used organic dye. The RhB solution (10^{-4} M) was treated with a plasma jet for 0, 90, 180, and 600 s. The plasmainduced degradation of organic dyes in aqueous solution has been studied previously with optical absorption spectroscopy [40-42]. In this research, we investigated plasmainduced dye degradation by both optical absorption and Raman spectroscopy. Figure 7A shows the optical absorption spectra of plasma-treated and untreated RhB solutions. The pristine RhB solution exhibited a broad absorption band with a strong peak at 550 nm and a shoulder at 520 nm. The 550 nm absorption was assigned to RhB monomers, and the 520 nm absorption peak was assigned to RhB dimers [43]. The main absorption peak located at 550 nm was chosen as an indicator for the degree of dye degradation. A change in the primary absorption peak at 550 nm with increasing plasma treatment time is shown in Figure 7B. When comparing the 550 nm absorption peak of plasma-treated RhB with that of untreated RhB, the 550 nm absorption peak intensity of 90 s plasma-treated RhB was around 26% of that of pristine RhB, indicating that the RhB molecules were degraded significantly by the plasma jet. When the plasma treatment time was increased to 180 s, the absorbance of RhB was reduced to about 5% of that of pristine RhB. Plasma treatment for 600 s reduced the absorbance of RhB to almost zero, suggesting that most of the RhB molecules had been destroyed by the plasma jet.

To further investigate the degradation of RhB by plasma treatment, we measured the Raman spectra of plasma-treated and untreated RhB. For SERS detection of RhB adsorbed on the AuNP-paper, paper disks were submerged in plasma-treated and untreated RhB solutions for 12 h, and RhB coated onto the AuNP-paper was subjected to Raman measurements after washing with fresh DI water. Figure 8A shows the Raman spectra of plasma-treated and untreated RhB adsorbed on the AuNP-paper. In this experiment, the Raman spectra were obtained using 633-nm excitation light with a power of 0.5 mW. The Raman spectra were measured at five different positions on the AuNP-paper. Spatially averaged Raman spectra were plotted in Figure 8A. The Raman spectra of pristine RhB exhibited Raman bands at 620.5 cm⁻¹, 1197.7 cm⁻¹, 1278.6 cm⁻¹, 1356.6 cm⁻¹, 1502.8 cm⁻¹, and 1642.4 cm⁻¹, which we attributed to C – C stretching, C = C stretching, and C – H bending modes of the aromatic ring [44]. The effects of the plasma treatment time on the

620 cm⁻¹-Raman peak are shown in Figure 8B. Error bars indicate standard deviations of Raman scattering intensities for those five positions on the AuNP-paper. The Raman peak intensities of the major Raman bands of plasma-treated RhB decreased with increasing plasma treatment time, indicating that RhB molecules were degraded by plasma treatment.

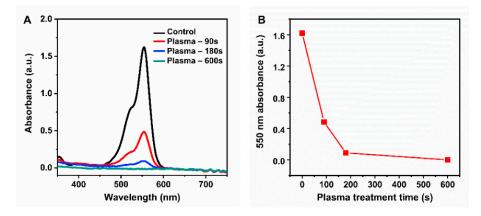


Figure 7. (**A**) Optical absorption spectra of the rhodamine B solutions with different plasma treatment times. The control sample corresponds to a 10^{-4} M Rh B solution not exposed to plasma. (**B**) 550 nm-absorption peak intensity of the RhB solution as a function of the plasma treatment time.

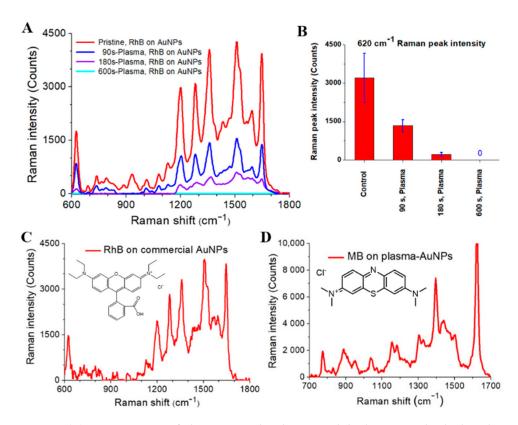


Figure 8. (**A**) Raman spectra of plasma-treated and untreated rhodamine B adsorbed on the AuNPpaper. The control sample was prepared using 10^{-4} M Rh B solution not exposed to plasma. The Raman spectra were averaged after measurements at five different positions on the AuNP-paper. (**B**) Effects of the plasma treatment time on the 620 cm⁻¹–Raman peak. Error bars indicate the standard deviation of the Raman scattering intensities for five different positions on the AuNPpaper. (**C**) Raman spectra of the rhodamine B adsorbed on the AuNP-paper with commercial gold nanoparticles. (**D**) Raman spectra of the methylene blue adsorbed on the AuNP-paper with plasma-synthesized gold nanoparticles.

To compare the optical sensing activity of plasma-synthesized AuNPs with that of commercial gold nanoparticles, we measured the Raman spectra of RhB coated on the AuNP-paper with commercial gold nanoparticles (Sigma-Aldrich, 741973, Yongin-city, Kyunggi-do, Korea; 30 nm diameter, OD 1, stabilized suspension in citrate buffer). Commercial AuNP-loaded papers and plasma-synthesized Au@PEG NP-loaded papers were prepared using gold nanoparticle solutions with the same SPR absorption peak. As shown in Figure 8C, the Raman spectra of RhB on commercial gold nanoparticles exhibited characteristic Raman peaks at 620.5 cm⁻¹, 1197.7 cm⁻¹, 1278.6 cm⁻¹, 1356.6 cm⁻¹, 1502.8 cm⁻¹, and 1642.4 cm^{-1} , similar to the Raman frequencies of RhB on plasma-synthesized AuNPs. In addition, major Raman peak intensities of RhB on commercial gold nanoparticles were slightly different from those of plasma-synthesized gold nanoparticles. These differences can be ascribed to the different fabrication method of gold nanoparticles and the different shell surrounding the AuNP core. A PEG shell surrounding the AuNP core can affect the binding between AuNPs and RhB molecules. Next, plasma-synthesized AuNPs were tested for the optical sensing of another molecule, methylene blue (MB). For this purpose, we coated MB on the AuNP-paper and measured the Raman spectra. Figure 8D shows the Raman spectra of MB adsorbed on the AuNP-paper. The Raman spectra of MB on AuNPs exhibited characteristic Raman peaks at 448.3 cm⁻¹ and 1621.4 cm⁻¹, which we attributed to C - N - C skeletal bending and C - C stretching modes [45,46]. This result indicates that plasma-synthesized AuNPs can be used to produce a Raman-sensitive AuNP-paper platform. To summarize, such an AuNP-paper-based Raman sensing platform has several merits: (1) it requires a small volume sample, and is cheap, flexible, and biocompatible; (2) it can discriminate between normal and damaged areas; and, (3) it is easy to load gold nanoparticles and sensing molecules on the paper surface. The spatial resolution and homogeneity of the sensing materials on the AuNP-paper are limited due to the use of filter paper and dip-coating method [47].

4. Conclusions

Herein, we described the synthesis of PEG-functionalized gold nanoparticles from tetrachloroauric acid and PEG using an argon plasma jet without additional reducing chemicals. Au@PEG NPs with a mean particle diameter of 32.5 nm were obtained after plasma treatment for 10 min. The presence of PEG in the solution assisted the formation of Au nanoparticle cores. Au@PEG NPs were coated on filter paper by a simple soaking process to produce a Raman-sensitive AuNP-paper platform. The AuNP-paper acted as a surface-enhanced Raman scattering platform when used to analyze the degradation of RhB dye. In summary, this study presents a quick and cost-effective process for the Raman sensing of plasma-synthesized Au@PEG NPs.

Author Contributions: G.J.L. conceived and performed the experiments, analyzed the data, and wrote the manuscript. L.N.N. performed the experiments and research and wrote the manuscript. P.L. performed the experiments. E.H.C. provided assistance in the plasma setup and the analysis of plasma-generated reactive oxygen and nitrogen species. All authors have read and agreed to the published version of the manuscript.

Funding: This research was supported by the Basic Science Research Program through the National Research Foundation of Korea funded by the Korea government (Grant number NRF-2018R1D1A1B0 7040386), and by Leading Foreign Research Institute Recruitment Program through the National Research Foundation of Korea (NRF) funded by the Korea government (NRF-2016K1A4A3914113), and by the Excellent researcher support project of Kwangwoon University in 2021.

Conflicts of Interest: The authors declare no conflict of interest.

References

- 1. Stiles, P.L.; Dieringer, J.A.; Shah, N.C.; Van Duyne, R.R. Surface-enhanced Raman spectroscopy. *Annu. Rev. Anal. Chem.* 2008, 1, 601–626. [CrossRef] [PubMed]
- Dasary, S.S.R.; Singh, A.K.; Senapati, D.; Yu, H.; Ray, P.C. Gold nanoparticle-based label-free SERS probe for ultrasensitive and selective detection of trinitrotoluene. *J. Am. Chem. Soc.* 2009, *131*, 13806–13812. [CrossRef]

- 3. Lee, G.J.; Kang, M.; Kim, Y.; Choi, E.H.; Cho, M.J.; Choi, D.H. Optical assessment of chiral–achiral polymer blends based on surface plasmon resonance effects of gold nanoparticles. *J. Phys. D Appl. Phys.* **2019**, *53*, 95102. [CrossRef]
- 4. Boisselier, E.; Astruc, D. Gold nanoparticles in nanomedicine: Preparations, imaging, diagnostics, therapies and toxicity. *Chem. Soc. Rev.* **2009**, *38*, 1759–1782. [CrossRef] [PubMed]
- 5. Anastas, P.; Eghbali, N. Green chemistry: Principles and practice. Chem. Soc. Rev. 2010, 39, 301–312. [CrossRef] [PubMed]
- Anastas, P.T.; Kirchhoff, M.M. Origins, current status, and future challenges of green chemistry. Acc. Chem. Res. 2002, 35, 686–694.
 [CrossRef]
- 7. Iravani, S. Green synthesis of metal nanoparticles using plants. Green Chem. 2011, 13, 2638–2650. [CrossRef]
- 8. Mandal, D.; Bolander, M.E.; Mukhopadhyay, D.; Sarkar, G.; Mukherjee, P. The use of microorganisms for the formation of metal nanoparticles and their application. *Appl. Microbiol. Biotechnol.* **2006**, *69*, 485–492. [CrossRef]
- Singh, P.; Kim, Y.J.; Zhang, D.; Yang, D.C. Biological synthesis of nanoparticles from plants and microorganisms. *Trends Biotechnol.* 2016, 34, 588–599. [CrossRef] [PubMed]
- 10. Huang, H.Z.; Yang, X.R. Synthesis of polysaccharide-stabilized gold and silver nanoparticles: A green method. *Carbohydr. Res.* **2004**, *339*, 2627–2631. [CrossRef]
- 11. Willner, I.; Baron, R.; Willner, B. Growing metal nanoparticles by enzymes. Adv. Mater. 2006, 18, 1109–1120. [CrossRef]
- 12. Zhu, H.; Du, M.; Zou, M.; Xu, C.; Li, N.; Fu, Y. Facile and green synthesis of well-dispersed Au nanoparticles in PAN nanofibers by tea polyphenols. *J. Mater. Chem.* **2012**, *22*, 9301–9307. [CrossRef]
- 13. Chien, Y.H.; Huang, C.C.; Wang, S.W.; Yeh, C.S. Synthesis of nanoparticles: Sunlight formation of gold nanodecahedra for ultra-sensitive lead-ion detection. *Green Chem.* **2011**, *13*, 1162–1166. [CrossRef]
- 14. Kaushik, N.K.; Kaushik, N.; Linh, N.N.; Ghimire, B.; Pengkit, A.; Sornsakdanuphap, J.; Lee, S.J.; Choi, E.H. Plasma and nanomaterials: Fabrication and biomedical applications. *Nanomaterials* **2019**, *9*, 98. [CrossRef]
- 15. Chiang, W.H.; Mariotti, D.; Sankaran, R.M.; Eden, J.G.; Ostrikov, K. Microplasmas for advanced materials and devices. *Adv. Mater.* **2020**, *32*, 1905508. [CrossRef]
- 16. Li, Z.; Zhang, J.; Wang, H.; Li, Z.; Zhang, X.; Di, L. Preparation of Pd/C by atmospheric-pressure ethanol cold plasma and its preparation mechanism. *Nanomaterials* **2019**, *9*, 1437. [CrossRef] [PubMed]
- 17. Tasche, D.; Weber, M.; Mrotzek, J.; Gerhard, C.; Wieneke, S.; Möbius, W.; Höfft, O.; Viöl, W. In situ investigation of the formation kinematics of plasma-generated silver nanoparticles. *Nanomaterials* **2020**, *10*, 555. [CrossRef] [PubMed]
- 18. Bruggeman, P.J.; Kushner, M.J.; Locke, B.R.; Gardeniers, J.G.E.; Graham, W.G.; Graves, D.B.; Hofman-Caris, R.; Maric, D.; Reid, J.P.; Ceriani, E.; et al. Plasma–liquid interactions: A review and roadmap. *Plasma Sources Sci. Technol.* **2016**, *25*, 053002. [CrossRef]
- 19. Rumbach, P.; Bartels, D.M.; Sankaran, R.M.; Go, D.B. The solvation of electrons by an atmospheric-pressure plasma. *Nat. Commun.* **2015**, *6*, 7248. [CrossRef]
- 20. Rumbach, P.; Witzke, M.; Sankaran, R.M.; Go, D.B. Decoupling interfacial reactions between plasmas and liquids: Charge transfer vs. plasma neutral reactions. *J. Am. Chem. Soc.* 2013, 135, 16264–16267. [CrossRef] [PubMed]
- 21. Rumbach, P.; Go, D.B. Perspectives on plasmas in contact with liquids for chemical processing and materials synthesis. *Top. Catal.* **2017**, *60*, 799–811. [CrossRef]
- 22. Chen, Q.; Li, J.; Li, Y. A review of plasma–liquid interactions for nanomaterial synthesis. J. Phys. D Appl. Phys. 2015, 48, 424005. [CrossRef]
- 23. Patel, J.; Němcová, L.; Maguire, P.; Graham, W.G.; Mariotti, D. Synthesis of surfactant-free electrostatically stabilized gold nanoparticles by plasma-induced liquid chemistry. *Nanotechnology* **2013**, *24*, 245604. [CrossRef]
- Zhang, R.C.; Sun, D.; Zhang, R.; Lin, W.F.; Macias-Montero, M.; Patel, J.; Askari, S.; McDonald, C.; Mariotti, D.; Maguire, P. Gold nanoparticle-polymer nanocomposites synthesized by room temperature atmospheric pressure plasma and their potential for fuel cell electrocatalytic application. *Sci. Rep.* 2017, 7, 46682. [CrossRef] [PubMed]
- De Vos, C.; Baneton, J.; Witzke, M.; Dille, J.; Godet, S.; Gordon, M.J.; Sankaran, R.M.; Reniers, F. A comparative study of the reduction of silver and gold salts in water by a cathodic microplasma electrode. J. Phys. D Appl. Phys. 2017, 50, 105206. [CrossRef]
- Nguyen, L.N.; Kaushik, N.; Lamichhane, P.; Mumtaz, S.; Paneru, R.; Bhartiya, P.; Kwon, J.S.; Mishra, Y.K.; Nguyen, L.Q.; Kaushik, N.K.; et al. In situ plasma-assisted synthesis of polydopamine-functionalized gold nanoparticles for biomedical applications. *Green Chem.* 2020, 22, 6588–6599. [CrossRef]
- 27. Jokerst, J.V.; Lobovkina, T.; Zare, R.N.; Gambhir, S.S. Nanoparticle PEGylation for imaging and therapy. *Nanomedicine* **2011**, *6*, 715–728. [CrossRef] [PubMed]
- 28. Furusho, H.; Kitano, K.; Hamaguchi, S.; Nagasaki, Y. Preparation of stable water-dispersible PEGylated gold nanoparticles assisted by nonequilibrium atmospheric-pressure plasma jets. *Chem. Mater.* **2009**, *21*, 3526–3535. [CrossRef]
- Otsuka, H.; Nagasaki, Y.; Kataoka, K. PEGylated nanoparticles for biological and pharmaceutical applications. *Adv. Drug Deliv. Rev.* 2003, 55, 403–419. [CrossRef]
- Shkilnyy, A.; Soucé, M.; Dubois, P.; Warmont, F.; Saboungi, M.L.; Chourpa, I. Poly (ethylene glycol)-stabilized silver nanoparticles for bioanalytical applications of SERS spectroscopy. *Analyst* 2009, *134*, 1868–1872. [CrossRef]
- Fernández-López, C.; Mateo-Mateo, C.; Álvarez-Puebla, R.A.; Pérez-Juste, J.; Pastoriza-Santos, I.; Liz-Marzán, L.M. Highly controlled silica coating of PEG-capped metal nanoparticles and preparation of SERS-encoded particles. *Langmuir* 2009, 25, 13894–13899. [CrossRef] [PubMed]

- Michen, B.; Geers, C.; Vanhecke, D.; Endes, C.; Rothen-Rutishauser, B.; Balog, S.; Petri-Fink, A. Avoiding drying-artifacts in transmission electron microscopy: Characterizing the size and colloidal state of nanoparticles. *Sci. Rep.* 2015, *5*, 9793. [CrossRef] [PubMed]
- Ershov, A.; Borysow, J. Dynamics of OH (X² Π v = 0) in high-energy atmospheric pressure electrical pulsed discharge. *J. Phys. D Appl. Phys.* 1995, 28, 68–74. [CrossRef]
- 34. Rahman, A.; Yalin, A.P.; Surla, V.; Stan, O.; Hoshimiya, K.; Yu, Z.; Littlefield, E.; Collins, G.J. Absolute UV and VUV emission in the 110–400 nm region from 13.56 MHz driven hollow slot microplasmas operating in open air. *Plasma Sources Sci. Technol.* 2004, *13*, 537–547. [CrossRef]
- Zhou, R.; Zhang, T.; Zhou, R.; Wang, S.; Mei, D.; Mai-Prochnow, A.; Weerasinghe, J.; Fang, Z.; Ostrikov, K.; Cullen, P.J. Sustainable plasma-catalytic bubbles for hydrogen peroxide synthesis. *Green Chem.* 2021, 23, 2977–2985. [CrossRef]
- 36. Giorgio, M.; Trinei, M.; Migliaccio, E.; Pelicci, P.G. Hydrogen peroxide: A metabolic by-product or a common mediator of ageing signals. *Nat. Rev. Mol. Cell Bio.* 2007, *8*, 722–728. [CrossRef]
- 37. Takeuchi, N.; Ishibashi, N. Generation mechanism of hydrogen peroxide in dc plasma with a liquid electrode. *Plasma Sources Sci. Technol.* **2018**, *27*, 045010. [CrossRef]
- Richmonds, C.; Sankaran, R.M. Plasma-liquid electrochemistry: Rapid synthesis of colloidal metal nanoparticles by microplasma reduction of aqueous cations. *Appl. Phys. Lett.* 2008, 93, 131501. [CrossRef]
- Urban, M.J.; Holder, I.T.; Schmid, M.; Fernandez Espin, V.; Garcia de la Torre, J.; Hartig, J.S.; Cölfen, H. Shape Analysis of DNA–Au Hybrid Particles by Analytical Ultracentrifugation. ACS Nano 2016, 10, 7418–7427. [CrossRef]
- 40. Gao, J.; Wang, X.; Hu, Z.; Deng, H.; Hou, J.; Lu, X.; Kang, J. Plasma degradation of dyes in water with contact glow discharge electrolysis. *Water Res.* 2003, *37*, 267–272. [CrossRef]
- 41. Jiang, B.; Zheng, J.; Liu, Q.; Wu, M. Degradation of azo dye using non-thermal plasma advanced oxidation process in a circulatory airtight reactor system. *Chem. Eng. J.* 2012, 204, 32–39. [CrossRef]
- Attri, P.; Yusupov, M.; Park, J.H.; Lingamdinne, L.P.; Koduru, J.R.; Shiratani, M.; Choi, E.H.; Bogaerts, A. Mechanism and comparison of needle-type non-thermal direct and indirect atmospheric pressure plasma jets on the degradation of dyes. *Sci. Rep.* 2016, *6*, 34419. [CrossRef] [PubMed]
- 43. Nishikiori, H.; Fujii, T. Molecular forms of rhodamine B in dip-coated thin films. J. Phys. Chem. B 1997, 101, 3680–3687. [CrossRef]
- Zhang, J.; Li, X.; Sun, X.; Li, Y. Surface-enhanced Raman scattering effects of silver colloids with different shapes. J. Phys. Chem. B 2005, 109, 12544–12548. [CrossRef] [PubMed]
- 45. Awada, C.; Dab, C.; Grimaldi, M.G.; Alshoaibi, A.; Ruffino, F. High optical enhancement in Au/Ag alloys and porous Au using surface-enhanced Raman spectroscopy technique. *Sci. Rep.* **2021**, *11*, 4714. [CrossRef]
- Sarkar, S.; Pande, S.; Jana, S.; Sinha, A.K.; Pradhan, M.; Basu, M.; Chowdhury, J.; Pal, T. Exploration of electrostatic field force in surface-enhanced Raman scattering: An experimental investigation aided by density functional calculations. *J. Phys. Chem. C* 2008, 112, 17862–17876. [CrossRef]
- Nery, E.W.; Kubota, L.T. Sensing approaches on paper-based devices: A review. Anal. Bioanal. Chem. 2013, 405, 7573–7595. [CrossRef] [PubMed]