



Supplementary Materials

Multifunctional Hydroxyapatite/Silver Nanoparticles/Cotton Gauze for Antimicrobial and Biomedical Applications

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Figure S1 shows the IR spectrum of chitosan. The strong band in the region 3291–3361 cm^{-1} corresponds to N–H and O–H stretching, as well as the intramolecular hydrogen bonds. The absorption bands at around 2921 and 2877 cm^{-1} are attributed to C–H symmetric and asymmetric stretching. The presence of residual N-acetyl groups was confirmed by the bands at around 1645 cm^{-1} (C=O stretching of amide I) and 1325 cm^{-1} (C–N stretching of amide III), respectively. The small band at 1550 cm^{-1} corresponds to the N–H bending of amide II. A peak at 1589 cm^{-1} corresponds to the N–H bending of the primary amine. The $-\text{CH}_2$ bending and $-\text{CH}_3$ symmetrical deformations were confirmed by the presence of bands at around 1423 and 1375 cm^{-1} , respectively. The absorption band at 1153 cm^{-1} can be attributed to asymmetric stretching of the C–O–C bridge. The bands at 1066 and 1028 cm^{-1} correspond to C–O stretching.

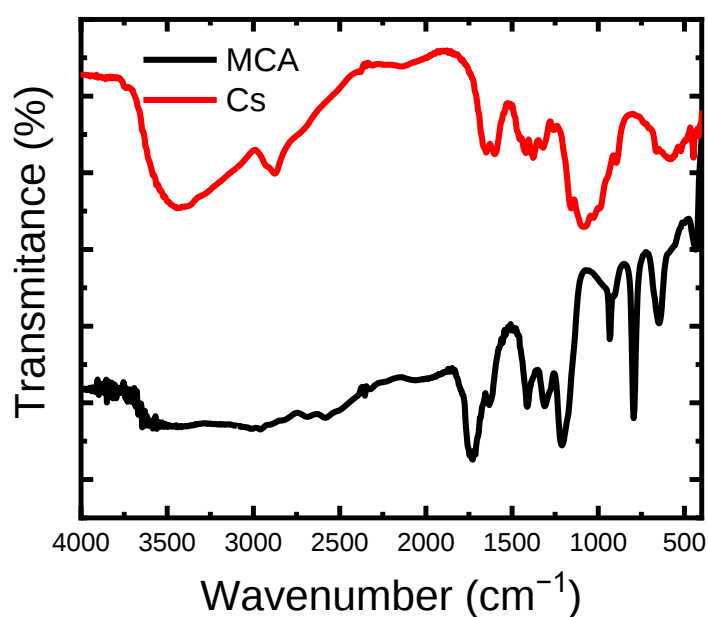


Figure S1. FTIR spectra of pure monochloroacetic acid (MCA) and cationic chitosan (Cs).

Figure S2 shows the Raman spectra of the cotton-Ag and cotton-H5-Ag. The obtained peaks are related to the cellulose chain of the cotton gauze. Cotton gauze samples (cotton-Ag and cotton-H5-Ag) showed strong, well-resolved peaks corresponding to ν of the C–C ring asymmetric stretching and C–O–C for glycoside link asymmetric and symmetric stretching at 1116, 1331 and 1088 cm^{-1} , respectively. The other peaks obtained at 1478, 379 and 1292 cm^{-1} are attributed to different types of $-\text{CH}_2$ group vibrations, while peaks at 1337 and 995 cm^{-1} are assigned to C–OH groups (Figure S2a). The peak at 2897 cm^{-1} is assigned to CH and $-\text{CH}_2$ stretching in cellulose. While peak obtained at 2736 cm^{-1} is attributed to the methine group in cotton (Figure S2b). The spectrum of cotton-Ag and cotton-H5-Ag were similar. A new peak that appeared at 540 cm^{-1} is assigned to HAp (Figure S2a).

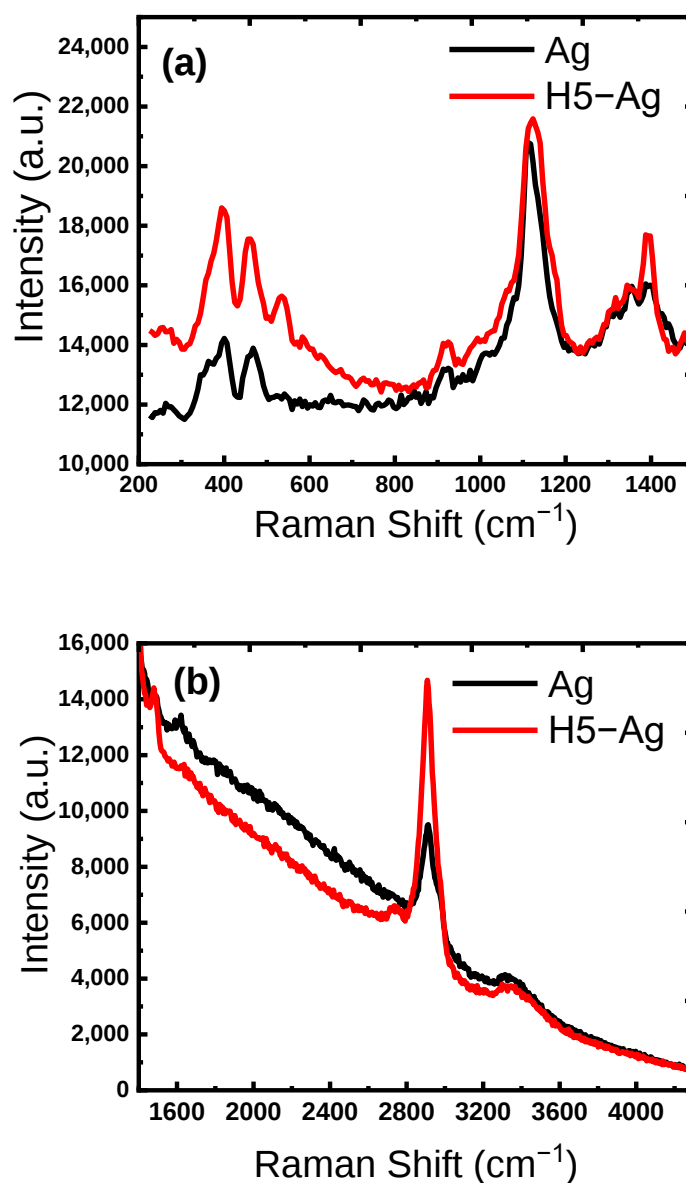


Figure S2. Raman Spectra of the cotton gauze fabrics coated with Ag NPs and HAp-Ag NPs after washing and drying: (a) spectrum from 200 to 1500 cm^{-1} ; and (b) spectrum from 1500 to 4200 cm^{-1} .

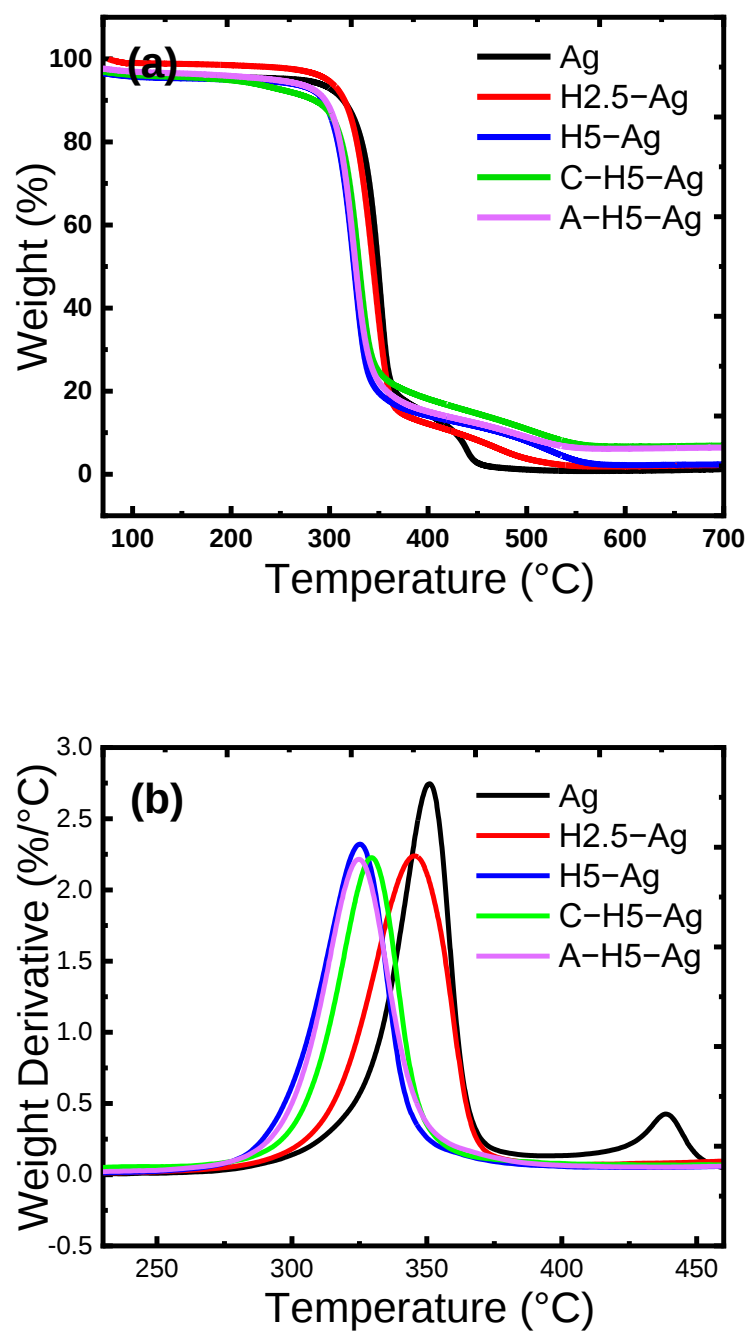


Figure S3. TGA thermal analysis of cotton fiber samples coated with Ag and HAp-Ag NPs: (a) TGA weight loss; and (b) Weight derivatives. Spectroscopic characteristics of the unmodified (blank) and modified cotton gauze.

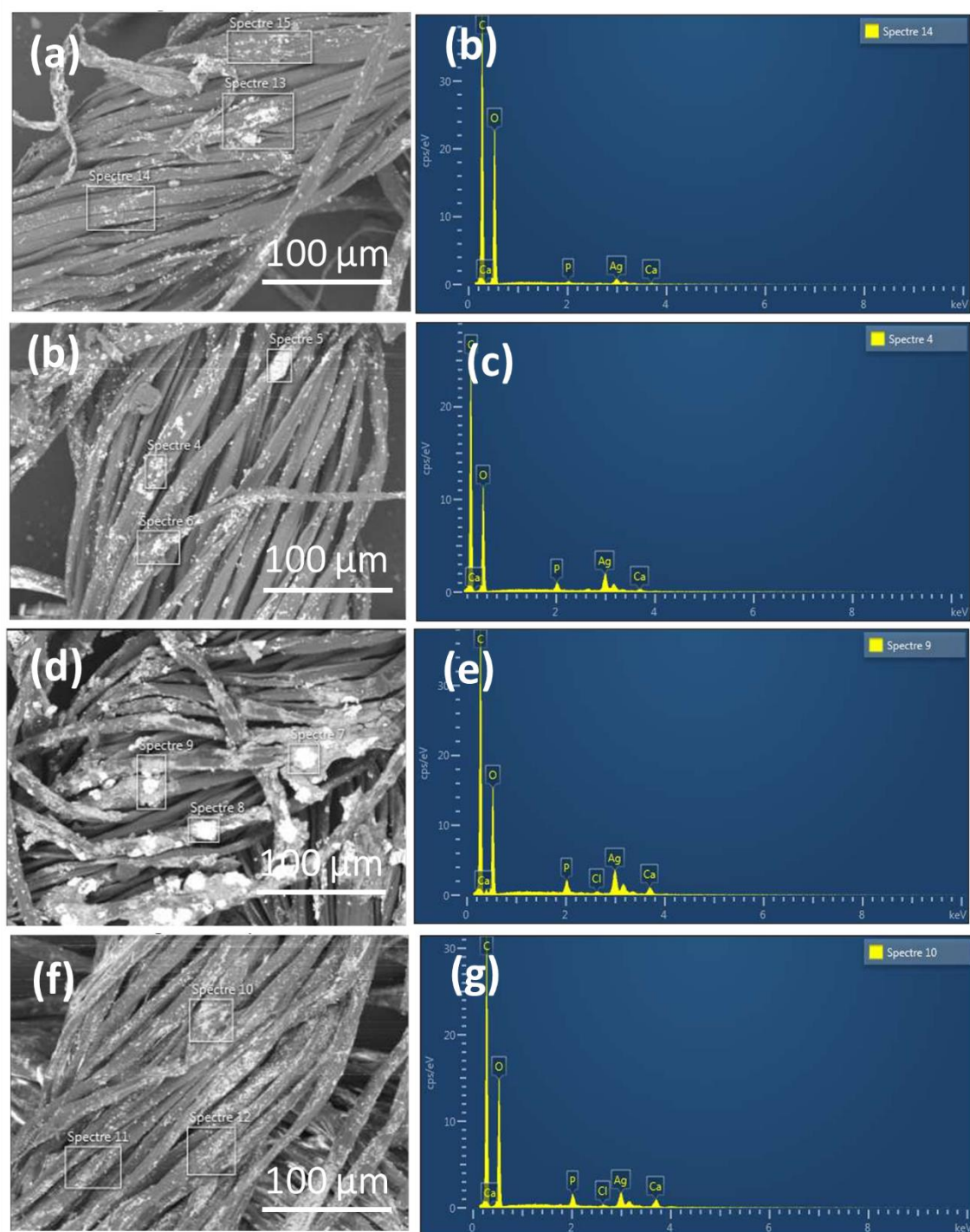


Figure S4. SEM-EDX analysis and their corresponding SEM image of the cotton gauze coated with HAp-Ag NPs: (a,b) cotton gauze coated with 2.5% HAp and 500 ppm Ag NPs; (c,d) cotton gauze coated with 5% HAp and 500 ppm Ag NPs; (e,f) cationic modified cotton gauze coated with 5% HAp and 500 ppm Ag NPs; (g,h) anionic modified cotton gauze coated with 5% HAp and 500 ppm Ag NPs.

Table S1. Antimicrobial activity, air permeability, water absorption, and tensile strength of the prepared cotton gauze samples.

Sample	Antimicrobial Activity (Inhibition Zone, nm)				Air Permeability cm ³ /cm ² /sec	Water Absorption % (30 min)	Tensile Strength (N/cm ²)
	Gram (+) Bacteria	Gram (-) Bacteria	Fungi	Fungi			
	<i>S. aureus</i>	<i>E. coli</i>	<i>C. albicans</i>	<i>A. niger</i>			
Blank	0	0	0	0	255.3 ± 1.4	98 ± 1.4	510 ± 2
H2.5	0	0	0	0	252.8 ± 1.8	111 ± 1.3	535 ± 2
C-H2.5	0	0	0	0	249.1 ± 1.0	117 ± 1.2	537 ± 1
A-H2.5	0	0	0	0	250.5 ± 1.6	120 ± 1.4	536 ± 3
H5	0	0	0	0	249.7 ± 1.0	115 ± 1.1	539 ± 6
C-H5	0	0	0	0	248.8 ± 1.3	118 ± 1.4	538 ± 3
A-H5	0	0	0	0	249.6 ± 1.4	122 ± 1.5	537 ± 5
Ag	13	14	13	0	253.3 ± 1.4	105 ± 1.2	520 ± 3
H2.5-Ag	16	17	16	0	252.4 ± 1.2	114 ± 1.1	536 ± 4
C-H2.5-Ag	15	16	14	0	248.9 ± 1.1	119 ± 1.4	538 ± 5
A-H2.5-Ag	18	20	18	0	250.3 ± 1.9	122 ± 1.1	537 ± 4
H5-Ag	15	15	14	0	249.3 ± 1.1	117 ± 1.3	540 ± 3
C-H5-Ag	14	16	13	0	248.5 ± 1.5	120 ± 1.5	539 ± 2
A-H5-Ag	18	19	18	0	249.2 ± 1.1	124 ± 1.4	539 ± 5