



Engineering Antioxidant Surfaces for Titanium-Based Metallic Biomaterials

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Abstract: Prolonged inflammation induced by orthopedic metallic implants can critically affect the success rates, which can even lead to aseptic loosening and consequent implant failure. In the case of adverse clinical conditions involving osteoporosis, orthopedic trauma and implant corrosion-wear in peri-implant region, the reactive oxygen species (ROS) activity is enhanced which leads to increased oxidative stress. Metallic implant materials (such as titanium and its alloys) can induce increased amount of ROS, thereby critically influencing the healing process. This will consequently affect the bone remodeling process and increase healing time. The current review explores the ROS generation aspects associated with Ti-based metallic biomaterials and the various surface modification strategies developed specifically to improve antioxidant aspects of Ti surfaces. The initial part of this review explores the ROS generation associated with Ti implant materials and the associated ROS metabolism resulting in the formation of superoxide anion, hydroxyl radical and hydrogen peroxide radicals. This is followed by a comprehensive overview of various organic and inorganic coatings/materials for effective antioxidant surfaces and outlook in this research direction. Overall, this review highlights the critical need to consider the aspects of ROS generation as well as oxidative stress while designing an implant material and its effective surface engineering.

Keywords: antioxidant; surfaces; titanium; reactive oxygen species (ROS); biomaterials

1. Introduction

One of the key factors associated with inflammatory response is the oxidative stress, which is characterized by the imbalance/disparity between the generation of reactive oxygen species (ROS) and antioxidant defense system [1]. Osteoporosis, the most common bone disorder globally, is a systemic skeletal disorder associated with diminishing bone mass and micro-architectural bone tissue degradation with concomitant bone fragility and osteoporotic fracture [2–4]. Considered as one of the major global pandemics of the 21st century, osteoporosis induces more than 8.9 million bone fractures per annum, affecting about 200 million people, and in addition, poses a high risk specifically to post-menopausal women, with 40–50% prevalence in women older than 60 years [5–7]. Other leading causes for bone fracture includes road accidents, falls and sports injuries. Following the bone fracture, secondary healing ensues involving various stages such as hematoma formation, acute inflammation, callus formation and bone remodeling [8]. The fracture trauma results in blood vessel rupture in the region of fracture leading to hematoma [9]. The hematoma



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Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). micro-environment in the fracture site is structurally unstable, hypoxic and acidic, which requires a cross-talk between inflammatory cells and cells related to bone healing in order to re-establish normal homeostatic state [9,10]. The bone remodeling involves a collective involvement of various bone cells such as osteoclasts (removal of damaged and old bones), osteoblasts (synthesis and secretion of osteoid matrix during mineralization) and osteocytes (regulating new bone formation and old bone resorption) [11–13]. Oxidative stress is a predominant factor which negatively affects the bone remodeling process resulting in a deteriorated bone mineral density, contributing in the etiology of osteoporosis [14–16].

Clinical intervention of bone fractures and defects considers the usage of orthopedic implants for the treatment of orthopedic trauma with minimal harm to the patients. Metallic, ceramic and polymeric biomaterials have been explored and researched for orthopedic implant applications, with each class of materials possessing its own advantages and disadvantages [17–19]. Thanks to their superior mechanical properties, metallic materials are the most widely used material for internal fracture fixation components. The three dominant material classes in this aspect are 316L stainless steel, Co-Cr alloys and Ti and its alloys [20–23]. However, metallic materials are prone to degradation due to corrosion-wear synergy (tribocorrosion) in complex physiological environments capable of eliciting the release of ions and debris in the peri-implant region [24,25]. Such wear-debris release from articulating components can result in the activation and senescence of resident cells including macrophages, fibroblasts, osteoclasts and osteoblasts, eventually leading to the production and release of pro-inflammatory cytokines, chemokines, ROS and reactive nitrogen species (RNS) [26,27]. This elicits chronic inflammatory cascades and oxidative stress reactions eventually resulting in bone resorption and osteolysis induced implant failure [26,28].

During normal healing process, osteoblasts express antioxidant enzymes such as superoxide dismutase (SOD) for inducing the conversion of ROS into O and H₂O to induce osteoblast differentiation [29,30]. However, during adverse conditions as mentioned above, enhanced ROS activity results in oxidative stress reducing the bone mineralization density by affecting the remodeling process [31,32]. In spite of the presence of endogenous antioxidants, excessive generation of free radicals and inflammatory processes result in oxidative stress [33]. The occurrence of oxidative stress can be ascribed to abnormal activation of enzymes which generates ROS. ROS are highly reactive, short-lived molecules formed as by-products during molecular oxygen reduction which are capable of oxidative damage to macromolecules in biological cells [34–38]. ROS include radical and non-radical oxygen species such as superoxide anion (O^{2-}), hydroxyl radical (OH^{-}) and hydrogen peroxide (H_2O_2) [39–42]. The mechanism of ROS formation via electrochemical corrosion reaction, radical transformation via Fenton and Haber–Weiss Reactions, light induction and surface catalytic reactions is elaborately reviewed by Kessier et al. [43].

Antioxidants are naturally occurring reducing agents which can hinder the generation of ROS via the phenomenon of scavenging free radicals and eradicating ROS derivatives. Hence, the origin of oxidative stress can be linked to the imbalance between ROS and antioxidants which encompass enzymatic antioxidants (e.g., polyphenols, carotenoids, glutathione, tocopherols) and antioxidant enzymes (SOD, catalase, glutathione peroxidase) [44]. An increment in antioxidant levels can be potentially harmful as it could induce molecular damages, apoptosis or necrosis, and oxidative stress is found to be associated with several diseases including cardiovascular, neurodegenerative, carcinoma, diabetes, ischemia/reperfusion injury, rheumatoid arthritis and aging [45]. Endogenous enzymatic antioxidants include SOD, catalase, glutathione peroxidase and glutathione reductase, whereas non-enzymatic endogenous antioxidants include glutathione and lipoic acid [46].

Metallic implant materials are widely used for bone-anchored therapy for orthopedic and dental treatments. Apart from the wear-induced oxidative stress as discussed above, metallic material insertion during surgical procedure induces large amount of ROS generation and is incapable of generating antioxidants, thereby critically influencing the healing process which elevates the healing time.

Implant surface plays a pivotal role in dictating the host response of the implanted material. In most cases, surface modification of implants alters the surface morphology, topography, chemistry and surface energy, particularly aimed at improving matrix protein adhesion, cellular adhesion and proliferation, to attain better osseointegration [47,48]. A variety of surface modification strategies involving surface texturing and surface coatings have been developed to improve the interfacial mechanical strength, wear resistance, tribocorrosion resistance and biocompatibility in order to enhance the longevity of orthopedic implants [49,50]. Recently, surface modification of Ti implants has been gaining research attention to repair the impaired osseointegration by developing surfaces with antioxidant activity [51–53]. In summary, it is imperative to gain more insights into the advancements in this field to further improve the antioxidant activities of Ti implant surface by proper surface modification to improve its clinical efficiency. In view of these aspects, the present review is focused towards the various surface engineering techniques to combat the undesirable ROS generation associated with Ti-based metallic implants. Several review articles have been published reporting the underlying mechanism of ROS formation and antioxidative mechanisms [30,54,55]. In addition, review articles comprehensively describing surface modification techniques for Ti surface are published [56,57]. The novelty aspect of the present review lies in collating the available reported works in improving the antioxidant properties of Ti-based metallic implant surfaces via various organic and inorganic coatings. Even though several research articles have explored the antioxidant activity of surfaces developed for antibacterial and biocompatible applications, this review exempts these articles and is focused on research associated with surfaces/materials specifically developed for antioxidant purpose. This review initially presents an outline of ROS generation associated with the insertion of Ti implants. This is followed by sections describing organicand inorganic-based coatings on Ti surfaces to ameliorate the antioxidant aspects along with prospective future perspectives. The major objective of the present review is to provide an overall idea about how surface modification can assist in improving the ROS scavenging activity and reduce oxidative damage to improve the clinical efficiency of Ti-based implants.

2. Titanium Alloys and Reactive Oxygen Species Metabolism

Titanium (Ti) and its alloys are the widely used material for a variety of load-bearing orthopedic implant applications thanks to the excellent mechanical aspects, lower modulus values, corrosion resistance and excellent biocompatibility [58]. Ti is a transition metal which exists in a hexagonal closed pack (hcp) crystal structure (α -Ti), which transforms into its allotropic form with a body-centered cubic (bcc) structure (β -Ti) above a temperature of 882 °C, which is retained up to its melting point (1688 °C). Several Ti-based alloys such as commercially pure Ti (cp-Ti, ASTM-F67), Ti-6Al-4V (ASTM-F136), Ti-6Al-7Nb (ASTM-F1472, F1295) and Ti-13Nb-13Zr (ASTM-F1713-08) have been explored for dental implants, bone fixation plates, screws and hip joint stems [59,60]. Current research focus is more shifted towards β -Ti alloys as they possess comparatively lower elastic modulus (as low as 46–55 GPa), high strength, good cold workability and, most importantly, the beneficial biocompatibility aspects due to β -phase stabilizing alloying additions (Nb, Ta, Mo, Mn, Fe etc.) [61–63]. In addition, Ti-based shape memory alloys are prospective materials for various biomedical applications owing to the shape memory and superelasticity effects [64]. Despite these beneficial aspects, wear-induced aseptic loosening is a limiting factor hampering the efficiency of Ti-based orthopedic implants [65]. Wear-particle phagocytosis by macrophages can induce cytokine and free radical release, resulting in an aseptic inflammatory response, capable of promoting osteoclast resorption [66]. The role of high oxidative stress as one of the main causative factors in various inflammatory and degenerative disorders points towards the contribution of ROS towards aseptic loosening. As a response to the released metallic particles in a physiological condition, the immune system elicits an inflammation process, which involves generation of ROS through a series of enzyme-assisted biochemical reactions (schematic figure as shown in Figure 1) [67].



Figure 1. Schematic depicting the ROS generation associated with Ti implants with associated biochemical reactions.

Superoxide radical generation is catalyzed by NADPH (nicotinamide adenine dinucleotide phosphate) oxidase (Equation (1)). Electrons from NADPH is accepted by the cytosolic domain of gp91^{phox} (electron transferase of NADPH oxidase) and is transferred across membrane to O₂ to generate superoxide radical (O₂⁻) as the primary product [68]. Gp91^{phox} contains all the required co-factors to effectuate electron transfer reaction, in which electrons transfer from NADPH onto flavin adenine dinucleotide (FAD) and to the haem group in the following step, inducing reduction of O₂ to O₂⁻ [69].

$$NADPH + O_2 \leftrightarrow NADP^+ + O_2^- + H^+$$
(1)

In response to this, antioxidant scavenging enzymes such as SOD promote dismutation to convert superoxide to hydrogen peroxide and an oxygen molecule (Equation (2)), which occurs spontaneously (rate constant = $5 \times 10^5 \text{ M}^{-1} \text{s}^{-1}$ at neutral pH) [70]. This reaction is greatly accelerated by SOD, and the corresponding catalytic activity is attributed partly to the electrostatic interactions in active center of SOD protein [71].

$$2O_2^- + 2H^+ \to O_2 + H_2O_2$$
 (2)

Stimulation of neutrophils results in oxygen consumption in a respiratory burst that produces O_2^- and H_2O_2 . Simultaneous discharge of abundant myeloperoxidase enzyme occurs, which utilizes H_2O_2 to oxidize halides (chlorides, bromides) and thiocyanates to corresponding hypohalous acids and hypothiocyanite [72]. Myeloperoxidase, also called verdoperoxidase, is a heme-containing peroxidase generated mostly from polymorphonuclear neutrophils and found in primary granules of granulocytic cells [73]. The reaction between hydrogen peroxide with halides (such as Cl^- in physiological environment) is catalyzed by granule-localized myeloperoxidase to form hypochlorous acid (bleach) (Equation (3)).

$$H_2O_2 + Cl^- \to HOCl + OH^-$$
(3)

In addition, hydrogen peroxide can generate hydroxide and hydroperoxyl radicals by reacting with ferrous and ferric cations (Fenton reactions). Fenton chemistry can significantly enhance the degradation of many transition metals (including Ti alloys, Co-Cr alloys) [74]. Fenton reaction involves an initial electron transfer with neither bond formation nor breaking and the generation of hydroxyl radicals [75]. Haber Weiss reaction which makes use of Fenton chemistry involves vital mechanism in which highly reactive hydroxyl radical generation occurs [76]. Another possible cathodic reaction taking place at implant/bone interface is oxygen reduction to generate hydrogen peroxide (Equation (4)). The cathodic oxygen reduction can be sub-divided into several reactions, resulting in the generation of hydroxyl radicals and hydrogen peroxide.

$$O_2 + 2H_2O + 2e^- \to H_2O_2 + 2OH^-$$
 (4)

Hence, ROS are additional products of overall electrochemical reactions occurring in the implant interface other than the metallic ions and/or particles. The presence of ROS (hydroxyl radicals and hydrogen peroxide) can further promote degradation of Ti implants [77]. Among the various ROS molecules, hydrogen peroxide can mix with water and diffuse through membranes of peri-implant tissues, critically affecting intracellular redox status, thereby increasing the chances of implant failure [78].

3. Surface Modification for Antioxidant Ti Surfaces

Surface modification of Ti alloys offers an effective strategy to combat the limitations associated with ROS activity. To develop such surfaces/coatings, several surface modification techniques such as layer-by-layer technique, immersion/dip coating, spin coating, plasma immersion ion implantation and radiofrequency plasma-enhanced chemical vapor deposition (enlisted in Table 1) are being researched. A limited number of coating surfaces/materials have been explored to improve the antioxidant activity of Ti surfaces which can be conveniently categorized as organic and inorganic materials for surface modification.

Technique	Description	Ref.
Layer-by-layer technique	Bottom-up adsorption technique which involves the development of multi-layered (layers of oppositely charged species) thin films bound together through electrostatic interactions.	[79,80]
Immersion/dip coating	Solution-based deposition method which involves the immersion of substrate into a solution of material to be coated which depends on parameters such as dwelling time, substrate-withdrawal speed, number of dip-coating cycles and coating evaporation factor.	[81,82]
Spin coating	A technique which uses centrifugal force for deposition, in which a suspension is dropped from top into the rotating substrate, and the resulting centrifugal force will assist in spreading out the coating on the substrate, thereby coating it. The process is dependent on parameters such as dispense volume, spin speed, solution viscosity, solution concentration, spin time, etc.	[83–85]
Plasma immersion ion implantation (PIII)	Method to improve biocompatibility aspects of material surfaces by immersing in a plasma environment and applying negative-high-voltage pulsed bias. Compared to traditional plasma techniques, PIII can extend to some tens of nanometers beneath sample surface and can treat complex geometries.	[86–88]
Plasma enhanced chemical vapor deposition	Low temperature chemical vapor deposition in which plasma is used to drive chemical reactions between plasma-generated-reactive species and substrate instead of high temperatures.	[89–91]

Table 1. Various reported techniques used for developing antioxidant surfaces on Ti/Ti alloy surfaces.

3.1. Organic Materials for Surface Modification

3.1.1. Tannic Acid

Tannic acid is a water-soluble natural polyphenol compound, which is often present in tea, wine and fruits and possesses excellent antioxidant and antibacterial activity owing to the presence of pyrogallol and catechol groups [92]. The antioxidant activity of tannic acid is dependent on its capability to chelate metal ions such as Fe(II) and interfering one of the reaction steps in Fenton reaction, thereby retarding oxidation [93]. There are several been used to deposition [90,97], electroducposition, of vassisted deposition [90] and minicipation [90] have been used to deposit tannic acid coatings. The presence of catechol groups renders tannic acid substrate-independent adhesive properties. Polyphenol group interactions can occur via several catechol–surface interactions ranging from noncovalent interactions (hydrogen bonding, pi–pi interactions) to chemical bonding (coordination, covalent) [99]. In addition, polyphenol tannic acid is capable of forming functional coatings on various materials by means of an intrinsic auto-oxidative surface polymerization. Sebastian et al. investigated the deposition kinetics of tannic acid on Ti surfaces which revealed a multiphase layer generation [100]. An initial growth phase revealed build-up of layer which is compact as well as rigid (approx. 2 h), followed by adsorption of an increasingly dissipative layer (approx. 5 h). Following this, a coating discontinuation was observed which was corroborated with large particle precipitation in coating solutions.

In order to develop multifunctional coatings on Ti surface, tannic acid is often codeposited along with other functional biomaterial coatings for prospective implant applications. Hydroxyapatite $(Ca_{10}(PO_4)_6(OH)_2)$ is a bioactive material, the main inorganic bone component which possesses excellent osteoinduction and osteoconduction properties. In view of rendering Ti surfaces (which are bioinert) with bioactive and antioxidant properties, hydroxyapatite and tannic acid based composite coatings have been explored. A consistent and strong antioxidant activity was displayed by hydroxyapatite/tannic acid coatings deposited on Ti substrates modified by titania (TiO₂) nanotubes (Figure 2a-c) [101]. Gelatin added to hydroxyapatite can improve the osteogenic aspects to enhance bone formation. However, gelatin-hydroxyapatite coatings failed in bone conduction function due to weak bonding between them. Tannic acid has been found to strongly adsorb to hydroxyapatite surface and firmly glued gelatin and hydroxyapatite [96]. The resultant tannic acid-hydroxyapatite-gelatin complex coating demonstrated significantly higher antioxidant activity and reduced cell damage/changes in the presence of H_2O_2 . There are limitations reported with the adherence of tannic acid onto hydroxyapatite and salivary acquired pellicle peptide modified tannic acid exhibited better adsorption performance on hydroxyapatite surface [102]. Tightly adsorbed coating exhibited smooth, superhydrophilic surface with excellent antibacterial and antibiofouling performance.

In order to develop multifunctional antioxidant and antibacterial coatings, tannic acid is coated along with antibacterial elements which can be contact killing, release killing or anti-adhesive. Despite being widely explored for a wide spectrum of antibacterial applications, silver (Ag) usage for bio-surfaces is limited by dose dependent cytotoxicity. Hydroxyapatite-tannic acid coating developed by immersion technique on a Ag-loaded TiO₂ nanotubular Ti surface demonstrated high antibacterial activity, improved cytocompatibility and revealed slow release of tannic acid from surface, which contributed towards persistent antioxidant activity as shown in Figure 2d–f [53]. Polyethylene glycol is a promising antifouling polymeric interface, an appropriate proton acceptor and can generate hydrogen bonds with tannic acid [103]. Simultaneous deposition of polyethylene glycol resulted in increased coating thickness and improved surface coverage [104]. A novel pH-bacteria triggered antibiotic release mechanism has been developed by layer-by layer deposition of tannic acid with cationic antibiotics such as tobramycin, gentamicin and polymyxin B [105]. Unlike linear polymer molecules which are incapable of retaining antibiotics, tannic acid through its hydrogen bonding and electrostatic interactions interacted well with the antibiotics. The interesting aspect is the non-eluting characteristic of the tannic acid/antibiotic coating which is capable of triggering antibiotic release created by pH reduction induced by bacterial pathogens. Hizal et al. reported an ultrathin tannic acid/gentamicin layer-by-layer film on 3D nano-pillared structures, which exhibited a 10-fold decrease in bacterial attachment due to larger surface area of nanostructured surface and lower bacterial adhesion forces on nanopillar tips [106]. Apart from these, strontium



 (Sr^{2+}) incorporated tannic acid functionalized on Ti surface revealed enhanced osteoblast differentiation and reduced osteoclast activity [107].

Figure 2. Antioxidant activity of HAP/tannic acid composite coating. (a) Mechanism illustrating antioxidant activity; (**b**,**c**) antioxidant activity as a function of incubation time of HAP/tannic acid coating compared to Trolox; (**c**) antioxidant activity of (I) negative control (deionized water), (II–IV) 10, 30 and 50 g/L of tannic acid, (V) positive control (Trolox). Data reported as means \pm standard deviations, n = 3 (* *p* < 0.05, ** *p* < 0.01, *** *p* < 0.001). Reprinted from [101] with permission from Elsevier. (**d**) Formation mechanism depiction of Ag nanoparticles; (**e**) antioxidant activity with respect to incubation time of Ag/HAP/tannic acid coating compared to Trolox; and (f) antioxidant activity of (I) negative control (deionized water), (II–IV) 0.05, 0.1 and 0.2 M AgNO₃, (V) positive control (Trolox). Reprinted from [53] with permission from Elsevier.

3.1.2. Chitosan

Chitosan is a polycationic natural macromolecule (with a molecular structure of (1,4)linked 2-amino-2-deoxy- β -d-glucan), which is capable of reacting with many physiologically relevant ROS [108–111]. Owing to its various beneficial aspects such as improvement of osseointegration and cellular interactions, minimal foreign body response, favorable degradation rate and, most importantly, due to antioxidant and free radical scavenging activities, this partly deacetylated form of chitin is a prospective material for surface modification [112,113]. Chitosan can form functional coatings on Ti surface owing to the existence of amine groups in chitosan polymer chains, which are capable of developing covalent bonds with Ti via silanization [114]. Reasonable mechanisms for antioxidative action of chitosan include presence of intra-molecular hydrogen bonding [115], residual-free amino groups in water-soluble chitosan which may induce metal chelation [116] and the ability of NH₂ amino groups to react with hydroxyl groups (OH⁻) to generate stable macromolecule radicals [117].

Lieder et al. studied the effect of degree of deacetylation of chitosan membranes coated on Ti surfaces which resulted in an improved fibronectin adsorption, cellular attachment and proliferation, but with no instigation of spontaneous osteogenic differentiation [114]. Chitosan coating (85–90% deacetylated) on porous Ti surface evidenced excellent antioxidant effect and favored osteoblast activity under diabetic conditions through reactivation of P13K/AKT pathway [118]. The study elucidated that chitosan can play a role in the reactivation of P13K/AKT pathway which mediates diabetes-induced ROS overproduction at bone-implant interface (Figure 3c,d). A multi-step layer-by-layer self-assembly was employed to deposit biofunctional composite coatings of chitosan and alginate enriched with caffeic acid on Ti-6Al-7Nb surface [119].



Figure 3. Antioxidant activity of chitosan based surfaces on Ti. (**a**) Antioxidant activity measured over a period of 8 weeks; (**b**) osteoblast interaction with and without treatment of hydrogen peroxide. Reprinted from [51] with permission from Elsevier. (**c**) intracellular ROS activity measured by DCF fluorescence intensity and (**d**) intracellular GSH-Px activity to quantify the rate of oxidation of the reduced glutathione to the oxidized glutathione by H₂O₂ (* *p* < 0.05 vs. TI + NS; # *p* < 0.05 vs. TI + DS; ** *p* < 0.01). Reprinted from [118] with permission from Elsevier.

Multiple steps consisted of piranha solution treatment of Ti alloy surface, plasma chemical activation and dip coating. Antioxidant activity measured in terms of DPPH-scavenging activity was higher for chitosan coating due to its potent reducing activity by hydrogen-donating ability. Conjugation of caffeic acid on chitosan resulted in the generation of amide linkages, increasing the amount of electron-donating groups. Another chitosan-based composite coating consisting of chitosan-catechol, gelatin and hydroxyapatite nanofibres deposited on Ti substrates exhibited high level of ROS scavenging activity and decreased oxidative damage on cellular level as displayed in Figure 3a,b [51]. This coating was able to retain increased level of p-FAK (assists in cell spreading and migration) and p-Akt (control cell survival and apoptosis) compared to pure Ti. The developed multilayer coating improved cell matrix adhesion and intercellular adhesion, while attenuating ROS-induced damage by interfering expressions of integrin αv and $\beta 3$, cadherin genes, anti-apoptotic and pro-apoptotic gene amounts. Electrophoretic deposition technique is also explored recently to coat chitosan-based composite coatings with hydroxyapatite, graphene and gentamycin [120].

3.1.3. Proanthocyanidin

Proanthocyanidin is condensed tannins (comprising of flavan-3-ol monomeric units), which belongs to the class of naturally occurring polyphenol flavonoid (non-thiol natural antioxidant), is found abundantly in berries and fruits [121,122]. Proanthocyanidin possesses excellent ROS scavenging activity, can regulate macrophage behavior and is capable of stimulating bone formation under oxidative stress conditions [123–126]. Tang et al. reported layer-by-layer self-assembly method to deposit hyaluronic acid/chitosan multilayers with proanthocyanidins [127]. The three-dimensional multilayered network of hyaluronic acid/chitosan on Ti surface facilitated proanthocyanidin incorporation into the micro interspaces between hyaluronic acid and chitosan, eventually leading to its controlled release. Proanthocyanidin incorporation is based on the electrostatic interaction between reactive OH⁻ radical in proanthocyanidin and positive amine groups in chitosan. Layer-by-layer assembly is a self-assembly technique based on the electrostatic attractions (polyanions and polycations) between the assembled components to generate polyelectrolyte multilayers. Layer-by-layer technique involves charging Ti substrates by

conjugating polyethylenimine for the purpose of obtaining higher binding forces. A sustained release of proanthocyanidin for a prolonged period of 14 days and mitigation of ROS-mediated inflammatory response were inferred. In other work, layer-by-layer technique was employed to integrate collagen type-II with proanthocyanidin which assisted in accelerating proliferation and osteogenic differentiation via Wnt/b-catenin signaling pathway and improved bone generation in vivo [128]. A novel covalent-conjugation strategy is reported to immobilize chitosan-encapsulated proanthocyanidin on Ti surface based on coupling agents (3-aminopropyl) triethoxysilane and glutaraldehyde [129]. Effective attenuation of the inhibitory effect of oxidative stress was induced by proanthocyanidin by the decrease of p53 gene expression. This study also indicated the improved stability of covalently immobilized coatings with improved wear and compression resistances attributed to strong chemical bonding and possessed the advantage of using nanoparticles as roller bearings.

4. Inorganic Materials for Surface Engineering Antioxidative Properties

4.1. Ceria Based Coatings

Cerium is a rare earth metallic element in lanthanide series and can exist in either free metal or metallic oxide form. Cerium possesses dual oxidation states: trivalent cerium sesquioxide (cerous Ce^{3+}) and tetravalent cerium dioxide (ceric Ce^{4+}) forms. Cerium oxide nanoparticles have received widespread attention for biocompatibility improvement, oph-thalmic applications [130], cardiovascular pathology, treating neurodegenerative disorders and spinal cord injury owing to its ROS-scavenging ability [131,132]. The role of cerium oxide nanoparticles to effectuate ROS-scavenging activity and antioxidant mimicking role has been extensively reviewed by Nelson et al. [133]. Cerium oxide nanoparticles exhibit rapid and expedient switches in oxidation state between Ce^{3+} and Ce^{4+} during redox reactions. Owing to its lower reduction potential, cerium oxide exhibits redox-cycling property.

Cerium oxide is one of the most interesting oxides due to the presence of oxygen vacancy defects (which can be quickly generated and eliminated), and it can act as an oxygen buffer. The presence of oxygen vacancy sites on nanoceria lattice is responsible for the unusual catalytic activity of this class of material which is dependent on the efficient supply of lattice oxygen at reaction sites governed by the formation of oxygen vacancy sites [134]. Catalytic reaction of cerium oxide nanoparticles with super oxide anion (O^{2-}) and hydrogen peroxide (H_2O_2) mimics biological action of SOD-mimetic and catalase thereby protecting cells against ROS induced damage [135]. Multi-enzymatic antioxidant activity is based on the ability of cerium oxide to rapidly switch between the multiple valence states. SOD mimic activity is elicited by a shift from Ce³⁺ to Ce³⁺ (deactivating hydrogen peroxide) [135–137]. SOD and catalase mimic activity of cerium oxide nanoparticles is particularly relevant under physiological pH condition (pH-7.5), rendering ROS-scavenging properties and inhibiting inflammatory mediator production.

Plasma-sprayed cerium oxide coating with a hierarchical topography was developed for antioxidant surfaces to preserve the intracellular antioxidant defense system [138]. Ceria oxide coating was found to be successful in decreasing SOD activity, reducing ROS generation and suppressing malondialdehyde development in hydrogen peroxide-treated osteoblasts. Li et al. reported magnetron sputtering (2, 3 and 5 min, $\approx 10^{-4}$ Pa) and vacuum annealing (450 °C) to deposit tiny homogenously distributed cerium oxide nanoparticle coatings with varying surface Ce⁴⁺/Ce³⁺ ratios by tuning of deposition time [131]. Quite interestingly, the Ce⁴⁺/Ce³⁺ ratio in this work reported the opposite trend for SOD and catalase mimetic activity. This work also highlighted the effective antioxidative mechanism of cerium oxide only when SOD and catalase mimetic activities are coordinated (H₂O₂ decomposition rate \geq generation rate). The observed Ce⁴⁺/Ce³⁺ ratio resulted in improved cytocompatibility, new bone formation, bone integration and upregulation of osteogenic genes and protein expressions (Figure 4a). Apart from the surface chemistry, the shape of ceria-based nanoparticles has also been reported to influence its ROS scavenging activity. Nanowire-shaped ceria is reported to occupy the extracellular space as its cellular internalization rate is slow [139]. Hence, nanowire-shaped ceria present on the cell surface can level down the hydrogen peroxide molecules and induce ROS consumption as schematically shown in Figure 4b. Spin coating represents a quick and facile surface modification technique to obtain coatings of thickness ranging from a few nanometers to few micrometers. Spin coating technique was used to deposit hydrothermally synthesized nano-CeO₂ with varying morphologies (nanorod, nano-cube and nano-octahedra) which yielded uniform coatings in Ti surfaces [140]. Total antioxidant capacity was in the order of nano-octahedron > nano-cube > nanorod. The anti-inflammation ability was correlated to the relative Ce³⁺ or Ce⁴⁺ content (XPS results displayed in Figure 4c) which, in turn, was influenced by the particle size and exposed crystalline lattice planes. With decreased particles size, Ce³⁺ content increased and rendered nano-octahedron improved SOD mimetic activity.



Figure 4. Influence of ceria surface chemistry and shape on the ROS scavenging activity. (**a**) Influence of Ce^{4+}/Ce^{3+} ratio on MSC and macrophage. Reprinted from [131] with permission from John Wiley & Sons. (**b**) X-ray-photoelectron spectroscopy analysis of nanorod, nano-cube and nano-octahedron-shaped ceria with varying Ce^{4+}/Ce^{3+} ratio. Reprinted from [139] with permission from Elsevier. (**c**) Schematic depicting the scavenging of extracellular matrix ROS by nanowire-shaped ceria present at the cell surface. Reprinted from [140] with permission from Elsevier.

High energy Ce plasma was used to develop cerium-modified Ti surface by using plasma immersion ion implantation technology [141]. A shift in the appearance of surface from nano-grains to nano-pits was noted, with treatment time increased from 30 to 60 and 90 min. Cerium implantation on Ti surface resulted in reducing the hydroxyl radical generation on Ti surface with increase in plasma immersion time. Reduction in fluorescence intensities and enhanced protection of E.coli model from oxidative stress are attributed to the improved corrosion resistance of the modified surface and the capability of the CeO_x on Ti surface to consume hydroxyl radical and hydrogen peroxide. In a similar work, atmospheric plasma was used to deposit cerium oxide-incorporated calcium silicate coating on Ti-6Al-4V substrates [142]. The developed surfaces demonstrated good biochemical stability, cellular viability and antibacterial activity against *E. faecalis*. Recently metal organic framework (MOF) coating was developed in situ on Ti surface to develop bioresponsive Ce/Sr incorporated bio-MOF coating based on hydrothermal technique [143]. Hydrothermally treated Ce-MOF and Ce/Sr-MOF revealed a Ce⁴⁺/Ce³⁺ ratio of 1.186 and 2.76, respectively. Both of these Ce-containing surfaces revealed excellent H₂O₂ decomposition, superoxide anion disintegrating capacity, 80% of radical clearance during DPPH assay and persistence of antioxidant activity with TMB assay.

4.2. Silica

Various silicon based coatings have been explored for biocompatibility applications such as amorphous silicon oxygen thin films (a-SiOx) [144,145], calcium silicate [146], solgel-based silica bioglasses [147], silicon nitride [148], etc. Silicon is an important element which possesses an influential role in the activity of SOD to improve the ROS scavenging ability. Ilias et al. studied the plasma-enhanced chemical vapor deposition of amorphous silicon oxynitride in view of attaining rapid bone regeneration and fracture healing [149]. This study is the first of its kind to reveal the dependence of Si^{4+} on SOD1 to improve osteogenesis. For an effective bone healing, a sustained released of Si⁴⁺ should be ensured from the implant surface. Nitrogen incorporation into amorphous silica effectuated a continual Si⁴⁺ release which can be fine-tuned based on the surface chemistry (O/N ratio), and thickness of deposited film dictates the total release period. Plasma-enhanced chemical vapor deposition was similarly utilized by the same research group to develop coatings in the form of silicon oxynitrophosphide [150]. Up-regulation of SOD1 and cat-1 was observed in cells exposed to silicon oxynitrophosphide with varying oxygen and nitrogen contents. In other work, a radio frequency plasma-enhanced chemical vapor deposition (RF-PECVD) method which makes use of silane as Si source was used to deposit hydrogenated amorphous silicon coatings on Ti-6Al-4V substrate [151]. Hydrogen incorporation into the coating resulted in lower surface oxidation and amorphous silicon coating influenced fibroblasts with no significant effect on keratinocytes. A table enlisting the summary of advantages and limitations of different organic and inorganic coatings/materials described is provided in Table 2.

Table 2. Summary of	t various coatings	s used for antioxid	dant properties.
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Type of Coating/Surface	Antioxidant Mechanism	Advantages	Limitations	Ref.
Tannic acid	Ability to chelate metal ions such as Fe(II), thereby interfering with one of the reaction steps in the Fenton reaction and thereby slowing oxidation	Antibacterial, antioxidant, high hemostatic efficiency, anticancer property, regenerative potential	Weak lipid solubility, low bioavailability, and short half-life, release rate should be controlled to exclude potential cytotoxicity, unstable adhesion	[152–155]
Chitosan	Reasonable mechanisms include presence of intra-molecular hydrogen bonding, metal chelation, ability of NH ₂ amino groups to react with hydroxyl groups	Biological activity, antimicrobial activity, hydrophilicity, and biodegradability	Delamination, unstable adhesion	[156–158]
Proanthocyanidin	By scavenging free radicals and by modifying signaling pathways, including those involving nuclear factor erythroid 2-related factor 2 (Nrf2), mitogen-activated protein kinase (MAPK), nuclear factor-kappaB (NF-κB), and phosphoinositide 3-kinase (PI3K)/Akt	Antioxidant, anticancer, antidiabetic, neuroprotective, and antimicrobial	High cost, low chemical stability and limited binding sites, difficulties in resolving the chemical labeling pattern of PAs with their proposed biosynthetic pathway, and defining the subcellular sites of biosynthesis	[121,124,159]

Type of Coating/Surface	Antioxidant Mechanism	Advantages	Limitations	Ref.
Ceria	Ability to rapidly switch between multiple valence states. SOD mimic activity is elicited by a shift from Ce^{3+} to Ce^{4+} (scavenging of O^{2-}) and catalase mimic activity is induced by a shift from Ce^{4+} to Ce^{3+} (deactivating hydrogen peroxide)	Antioxidant, anticancer and anti-inflammatory properties, biosensors	Toxicity associated with small-sized nano-ceria	[160,161]
Silica	Hydroxylation degree, By regulation of antioxidants enzyme activity	Accelerated bone fracture healing, biomineral synthesis	Lipid peroxidation induced toxicity	[149,162]

Table 2. Cont.

5. Conclusions and Future Perspectives

There are several titanium-surface modification techniques being used which can be classified as mechanical (polishing, blasting, peening), chemical (chemical treatment, solgel, anodic oxidation, chemical vapor deposition) and physical (thermal spraying, plasma spraying, physical vapor deposition, evaporation, ion plating, sputtering, glow discharge plasma, ion implantation and deposition) techniques [57,163]. In spite of the fact that several surface modification strategies are being researched with focus on antibacterial and cytocompatible surfaces, Ti surfaces with improved antioxidant properties require further research focus. The most common techniques explored on depositing functional molecules on the Ti surfaces are based on physical adsorption, based on weak hydrogen bonding and van der Waals forces. This is a limiting factor as it restricts the bond strength and coating life, which will potentially affect the efficacy of the implant. This can be tackled based on chemical immobilization via covalent bonding, in which case a more sustained release of functional molecules can be achieved as compared to physical adsorption techniques.

A critical limitation hampering the potentiality of Ti and its alloys is the inferior wear resistance to be used in articulating surfaces. In spite of the fact that various organic coatings on Ti are bioactive and can develop antioxidant activity, these coatings are mechanically instable, which is a particularly relevant aspect to be considered in terms of wear resistance. During surgical procedures, these implants often encounter mechanical forces of up to 15 N, which will critically affect the life of the coatings and sometimes can lead to coating spalling [129]. One way to tackle this will be the immobilization of such molecules on an already-modified surface layer [163]. Hence, a prospective idea is the development of bi-layer coating consisting of (a) an inner wear/corrosion resistant layer and (b) an outer bioactive layer with antioxidant activity. More research should be focused towards the extraction of exogenous antioxidants (mainly derived from food and medicinal plants, such as fruits, vegetables, cereals, mushrooms, beverages, flowers, spices and traditional medicinal herbs [164]) and its immobilization on Ti surface. Increasing the complexity of a surface modification process will render the process difficult and expensive for rapid commercialization.

One of the critical factors to be assessed while developing such surface is the effect of surface oxide layer on Ti surface. Ti and its alloys, when exposed to air, form a spontaneous native titanium dioxide (TiO_2) layer with thickness in the range of 2–20 nm. This possesses a profound influence on the binding of molecules on the Ti surface and coating adhesion. Antioxidant release kinetics should also be given prior focus, as burst release in physiological environment can induce harmful enzymatic imbalances. More computational studies focused towards the stimulatory effect of various prospective coating materials on inducing oxidative stress and ROS need emphasis. Apart from these aspects, various factors to stimulate physiological conditions to assess the antioxidant activity of the developed

surfaces shall be incorporated in studies, as synergetic influence of factors can alter the ROS scavenging activity.

Despite the beneficial aspects possessed by Ti and its alloy for load-bearing implant applications, there is a plenty of room for investigating the complex biological phenomena associated with ROS activity in the physiological environment. Most of the published works intended to improve the antioxidant properties of Ti surface are based on organic materials such as tannic acid, chitosan and proanthocyanidin and inorganic coatings based on ceria and silica. Future relevant research trends can be foreseen in improving the mechanical stability and controlled drug elution associated with organic coatings. It is also highly desirable that multifactorial aspects in a real physiological environment shall be considered while assessing the ROS scavenging activity of the developed surfaces. Overall, it is suggested to consider the ROS generation and antioxidant aspects with more research in this direction to develop an efficient implant surface of metallic biomaterials for improving the clinical efficiency. Most importantly, complex challenges associated with translation of lab research to clinical practice demands effective collaboration between material scientists, engineers, biologists and clinicians.

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References

- Mohamad, N.-V.; Ima-Nirwana, S.; Chin, K.-Y. Are Oxidative Stress and Inflammation Mediators of Bone Loss due to Estrogen Deficiency? A Review of Current Evidence. *Endocr. Metab. Immune Disord.-Drug Targets* 2020, 20, 1478–1487. [CrossRef] [PubMed]
- Consensus development conference: Diagnosis, prophylaxis, and treatment of osteoporosis. Am. J. Med. 1993, 94, 646–650. [CrossRef] [PubMed]
- 3. Harvey, N.; Dennison, E.; Cooper, C. Osteoporosis: Impact on health and economics. *Nat. Rev. Rheumatol.* **2010**, *6*, 99–105. [CrossRef] [PubMed]
- Salari, N.; Ghasemi, H.; Mohammadi, L.; Behzadi, M.H.; Rabieenia, E.; Shohaimi, S.; Mohammadi, M. The global prevalence of osteoporosis in the world: A comprehensive systematic review and meta-analysis. J. Orthop. Surg. Res. 2021, 16, 609. [CrossRef]
- 5. Al Anouti, F.; Taha, Z.; Shamim, S.; Khalaf, K.; Al Kaabi, L.; Alsafar, H. An insight into the paradigms of osteoporosis: From genetics to biomechanics. *Bone Rep.* 2019, *11*, 100216. [CrossRef]
- Johnell, O.; Kanis, J.A. An estimate of the worldwide prevalence and disability associated with osteoporotic fractures. *Osteoporos. Int.* 2006, 17, 1726–1733. [CrossRef]
- 7. Ji, M.X.; Yu, Q. Primary osteoporosis in postmenopausal women. Chronic Dis. Transl. Med. 2015, 1, 9–13. [CrossRef]
- Loi, F.; Córdova, L.A.; Pajarinen, J.; Lin, T.-H.; Yao, Z.; Goodman, S.B. Inflammation, fracture and bone repair. *Bone* 2016, *86*, 119–130. [CrossRef]
- Schell, H.; Duda, G.N.; Peters, A.; Tsitsilonis, S.; Johnson, K.A.; Schmidt-Bleek, K. The haematoma and its role in bone healing. J. Exp. Orthop. 2017, 4, 5. [CrossRef]
- 10. Walters, G.; Pountos, I.; Giannoudis, P.V. The cytokines and micro-environment of fracture haematoma: Current evidence. J. Tissue Eng. Regen. Med. 2018, 12, e1662–e1677. [CrossRef]
- Raggatt, L.J.; Partridge, N.C. Cellular and Molecular Mechanisms of Bone Remodeling. J. Biol. Chem. 2010, 285, 25103–25108.
 [CrossRef]
- 12. Bellido, T. Osteocyte-Driven Bone Remodeling. Calcif. Tissue Int. 2014, 94, 25–34. [CrossRef] [PubMed]
- Xiong, J.; O'Brien, C.A. Osteocyte RANKL: New insights into the control of bone remodeling. J. Bone Miner. Res. 2012, 27, 499–505. [CrossRef] [PubMed]
- Marcucci, G.; Domazetovic, V.; Nediani, C.; Ruzzolini, J.; Favre, C.; Brandi, M.L. Oxidative Stress and Natural Antioxidants in Osteoporosis: Novel Preventive and Therapeutic Approaches. *Antioxidants* 2023, 12, 373. [CrossRef] [PubMed]

- 15. Bădilă, A.E.; Rădulescu, D.M.; Ilie, A.; Niculescu, A.-G.; Grumezescu, A.M.; Rădulescu, A.R. Bone Regeneration and Oxidative Stress: An Updated Overview. *Antioxidants* **2022**, *11*, 318. [CrossRef] [PubMed]
- León-Reyes, G.; Argoty-Pantoja, A.D.; Becerra-Cervera, A.; López-Montoya, P.; Rivera-Paredez, B.; Velázquez-Cruz, R. Oxidative-Stress-Related Genes in Osteoporosis: A Systematic Review. *Antioxidants* 2023, 12, 915. [CrossRef]
- 17. Hallab, N.J.; Jacobs, J.J. 2.5.4—Orthopedic Applications. In *Biomaterials Science*, 4th ed.; Wagner, W.R., Sakiyama-Elbert, S.E., Zhang, G., Yaszemski, M.J., Eds.; Academic Press: Cambridge, MA, USA, 2020; pp. 1079–1118. [CrossRef]
- 18. Ratner, B.D.; Zhang, G. 1.1.2—A History of Biomaterials. In *Biomaterials Science*, 4th ed.; Wagner, W.R., Sakiyama-Elbert, S.E., Zhang, G., Yaszemski, M.J., Eds.; Academic Press: Cambridge, MA, USA, 2020; pp. 21–34. [CrossRef]
- 19. Wagner, W.R. 1.3.1—The Materials Side of the Biomaterials Relationship. In *Biomaterials Science*, 4th ed.; Wagner, W.R., Sakiyama-Elbert, S.E., Zhang, G., Yaszemski, M.J., Eds.; Academic Press: Cambridge, MA, USA, 2020; pp. 83–84. [CrossRef]
- Bai, L.; Gong, C.; Chen, X.; Sun, Y.; Zhang, J.; Cai, L.; Zhu, S.; Xie, S.Q. Additive Manufacturing of Customized Metallic Orthopedic Implants: Materials, Structures, and Surface Modifications. *Metals* 2019, 9, 1004. [CrossRef]
- Pilliar, R.M. Metallic Biomaterials. In *Biomedical Materials*; Narayan, R., Ed.; Springer International Publishing: Cham, Germany, 2021; pp. 1–47. [CrossRef]
- Prasad, K.; Bazaka, O.; Chua, M.; Rochford, M.; Fedrick, L.; Spoor, J.; Symes, R.; Tieppo, M.; Collins, C.; Cao, A.; et al. Metallic Biomaterials: Current Challenges and Opportunities. *Materials* 2017, 10, 884. [CrossRef]
- 23. Chen, Q.; Thouas, G.A. Metallic implant biomaterials. Mater. Sci. Eng. R Rep. 2015, 87, 1–57. [CrossRef]
- 24. Villanueva, J.; Trino, L.; Thomas, J.; Bijukumar, D.; Royhman, D.; Stack, M.M.; Mathew, M.T. Corrosion, Tribology, and Tribocorrosion Research in Biomedical Implants: Progressive Trend in the Published Literature. *J. Bio-Tribo-Corros.* **2016**, *3*, 1. [CrossRef]
- 25. Eliaz, N. Corrosion of Metallic Biomaterials: A Review. Materials 2019, 12, 407. [CrossRef]
- Steinbeck, M.J.; Jablonowski, L.J.; Parvizi, J.; Freeman, T.A. The Role of Oxidative Stress in Aseptic Loosening of Total Hip Arthroplasties. J. Arthroplast. 2014, 29, 843–849. [CrossRef]
- Primožič, J.; Poljšak, B.; Jamnik, P.; Kovač, V.; Čanadi Jurešić, G.; Spalj, S. Risk Assessment of Oxidative Stress Induced by Metal Ions Released from Fixed Orthodontic Appliances during Treatment and Indications for Supportive Antioxidant Therapy: A Narrative Review. *Antioxidants* 2021, 10, 1359. [CrossRef]
- 28. Goodman, S.B.; Gallo, J. Periprosthetic Osteolysis: Mechanisms, Prevention and Treatment. J. Clin.Med. 2019, 8, 2091. [CrossRef]
- 29. Wang, Y.; Branicky, R.; Noë, A.; Hekimi, S. Superoxide dismutases: Dual roles in controlling ROS damage and regulating ROS signaling. *J. Cell Biol.* **2018**, *217*, 1915–1928. [CrossRef]
- Atashi, F.; Modarressi, A.; Pepper, M.S. The role of reactive oxygen species in mesenchymal stem cell adipogenic and osteogenic differentiation: A review. *Stem Cells Dev.* 2015, 24, 1150–1163. [CrossRef]
- Cerqueni, G.; Scalzone, A.; Licini, C.; Gentile, P.; Mattioli-Belmonte, M. Insights into oxidative stress in bone tissue and novel challenges for biomaterials. *Mater. Sci. Eng. C* 2021, 130, 112433. [CrossRef] [PubMed]
- Domazetovic, V.; Marcucci, G.; Iantomasi, T.; Brandi, M.L.; Vincenzini, M.T. Oxidative stress in bone remodeling: Role of antioxidants. *Clin. Cases Miner. Bone Metab. Off. J. Ital. Soc. Osteoporos. Miner. Metab. Skelet. Dis.* 2017, 14, 209–216. [CrossRef] [PubMed]
- 33. Pizzino, G.; Irrera, N.; Cucinotta, M.; Pallio, G.; Mannino, F.; Arcoraci, V.; Squadrito, F.; Altavilla, D.; Bitto, A. Oxidative Stress: Harms and Benefits for Human Health. *Oxidative Med. Cell. Longev.* **2017**, 2017, 8416763. [CrossRef]
- Sies, H.; Belousov, V.V.; Chandel, N.S.; Davies, M.J.; Jones, D.P.; Mann, G.E.; Murphy, M.P.; Yamamoto, M.; Winterbourn, C. Defining roles of specific reactive oxygen species (ROS) in cell biology and physiology. *Nat. Rev. Mol. Cell Biol.* 2022, 23, 499–515. [CrossRef] [PubMed]
- 35. Thannickal, V.J.; Fanburg, B.L. Reactive oxygen species in cell signaling. *Am. J. Physiol.-Lung Cell. Mol. Physiol.* 2000, 279, L1005–L1028. [CrossRef] [PubMed]
- 36. Brieger, K.; Schiavone, S.; Miller, F.J., Jr.; Krause, K.-H. Reactive oxygen species: From health to disease. *Swiss Med. Wkly.* 2012, 142, w13659. [CrossRef] [PubMed]
- 37. Nosaka, Y.; Nosaka, A.Y. Generation and Detection of Reactive Oxygen Species in Photocatalysis. *Chem. Rev.* 2017, 117, 11302–11336. [CrossRef] [PubMed]
- Parham, S.; Kharazi, A.Z.; Bakhsheshi-Rad, H.R.; Nur, H.; Ismail, A.F.; Sharif, S.; RamaKrishna, S.; Berto, F. Antioxidant, Antimicrobial and Antiviral Properties of Herbal Materials. *Antioxidants* 2020, *9*, 1309. [CrossRef] [PubMed]
- Collin, F. Chemical Basis of Reactive Oxygen Species Reactivity and Involvement in Neurodegenerative Diseases. *Int. J. Mol.Sci.* 2019, 20, 2407. [CrossRef]
- Murphy, M.P.; Bayir, H.; Belousov, V.; Chang, C.J.; Davies, K.J.A.; Davies, M.J.; Dick, T.P.; Finkel, T.; Forman, H.J.; Janssen-Heininger, Y.; et al. Guidelines for measuring reactive oxygen species and oxidative damage in cells and in vivo. *Nat. Metab.* 2022, 4, 651–662. [CrossRef]
- Andrés, C.M.; Pérez de la Lastra, J.M.; Andrés Juan, C.; Plou, F.J.; Pérez-Lebeña, E. Superoxide Anion Chemistry— Its Role at the Core of the Innate Immunity. *Int. J. Mol.Sci.* 2023, 24, 1841.
- 42. Pervaiz, S.; Clement, M.-V. Superoxide anion: Oncogenic reactive oxygen species? *Int. J. Biochem. Cell Biol.* **2007**, *39*, 1297–1304. [CrossRef]

- Kessler, A.; Hedberg, J.; Blomberg, E.; Odnevall, I. Reactive Oxygen Species Formed by Metal and Metal Oxide Nanoparticles in Physiological Media&Mdash; A Review of Reactions of Importance to Nanotoxicity and Proposal for Categorization. *Nanomaterials* 2022, 12, 1922.
- Bešlo, D.; Golubić, N.; Rastija, V.; Agić, D.; Karnaš, M.; Šubarić, D.; Lučić, B. Antioxidant Activity, Metabolism, and Bioavailability of Polyphenols in the Diet of Animals. *Antioxidants* 2023, 12, 1141. [CrossRef]
- 45. Valko, M.; Leibfritz, D.; Moncol, J.; Cronin, M.T.D.; Mazur, M.; Telser, J. Free radicals and antioxidants in normal physiological functions and human disease. *Int. J. Biochem. Cell Biol.* **2007**, *39*, 44–84. [CrossRef] [PubMed]
- Kotha, R.R.; Tareq, F.S.; Yildiz, E.; Luthria, D.L. Oxidative Stress and Antioxidants&Mdash; A Critical Review on In Vitro Antioxidant Assays. *Antioxidants* 2022, 11, 2388.
- Smeets, R.; Stadlinger, B.; Schwarz, F.; Beck-Broichsitter, B.; Jung, O.; Precht, C.; Kloss, F.; Gröbe, A.; Heiland, M.; Ebker, T. Impact of Dental Implant Surface Modifications on Osseointegration. *BioMed Res. Int.* 2016, 2016, 6285620. [CrossRef]
- Zhu, G.; Wang, G.; Li, J.J. Advances in implant surface modifications to improve osseointegration. *Mater. Adv.* 2021, 2, 6901–6927. [CrossRef]
- 49. Ghosh, S.; Abanteriba, S. Status of surface modification techniques for artificial hip implants. *Sci. Technol. Adv. Mater.* **2016**, *17*, 715–735. [CrossRef] [PubMed]
- 50. Liu, Y.; Rath, B.; Tingart, M.; Eschweiler, J. Role of implants surface modification in osseointegration: A systematic review. *J. Biomed. Mater. Res. Part A* **2020**, *108*, 470–484. [CrossRef]
- Chen, W.; Shen, X.; Hu, Y.; Xu, K.; Ran, Q.; Yu, Y.; Dai, L.; Yuan, Z.; Huang, L.; Shen, T.; et al. Surface functionalization of titanium implants with chitosan-catechol conjugate for suppression of ROS-induced cells damage and improvement of osteogenesis. *Biomaterials* 2017, 114, 82–96. [CrossRef]
- 52. Dumitriu, C.; Ungureanu, C.; Popescu, S.; Tofan, V.; Popescu, M.; Pirvu, C. Ti surface modification with a natural antioxidant and antimicrobial agent. *Surf. Coat. Technol.* **2015**, *276*, 175–185. [CrossRef]
- 53. Di, H.; Qiaoxia, L.; Yujie, Z.; Jingxuan, L.; Yan, W.; Yinchun, H.; Xiaojie, L.; Song, C.; Weiyi, C. Ag nanoparticles incorporated tannic acid/nanoapatite composite coating on Ti implant surfaces for enhancement of antibacterial and antioxidant properties. *Surf. Coat. Technol.* **2020**, *399*, 126169. [CrossRef]
- Clanton, T.L. Hypoxia-induced reactive oxygen species formation in skeletal muscle. J. Appl. Physiol. 2007, 102, 2379–2388. [CrossRef]
- 55. Chapple, I.L.C. Reactive oxygen species and antioxidants in inflammatory diseases. J. Clin. Periodontol. **1997**, 24, 287–296. [CrossRef]
- 56. Han, X.; Ma, J.; Tian, A.; Wang, Y.; Li, Y.; Dong, B.; Tong, X.; Ma, X. Surface modification techniques of titanium and titanium alloys for biomedical orthopaedics applications: A review. *Colloids Surf. B Biointerfaces* **2023**, 227, 113339. [CrossRef]
- 57. Liu, X.; Chu, P.K.; Ding, C. Surface modification of titanium, titanium alloys, and related materials for biomedical applications. *Mater. Sci. Eng. R Rep.* **2004**, 47, 49–121. [CrossRef]
- 58. Geetha, M.; Singh, A.K.; Asokamani, R.; Gogia, A.K. Ti based biomaterials, the ultimate choice for orthopaedic implants—A review. *Prog. Mater. Sci.* 2009, *54*, 397–425. [CrossRef]
- Kaur, M.; Singh, K. Review on titanium and titanium based alloys as biomaterials for orthopaedic applications. *Mater. Sci. Eng. C* 2019, 102, 844–862. [CrossRef] [PubMed]
- 60. Quinn, J.; McFadden, R.; Chan, C.-W.; Carson, L. Titanium for Orthopedic Applications: An Overview of Surface Modification to Improve Biocompatibility and Prevent Bacterial Biofilm Formation. *iScience* **2020**, *23*, 101745. [CrossRef] [PubMed]
- Chen, L.-Y.; Cui, Y.-W.; Zhang, L.-C. Recent Development in Beta Titanium Alloys for Biomedical Applications. *Metals* 2020, 10, 1139. [CrossRef]
- 62. Kolli, R.P.; Devaraj, A. A Review of Metastable Beta Titanium Alloys. *Metals* 2018, *8*, 506. [CrossRef]
- Abdel-Hady Gepreel, M.; Niinomi, M. Biocompatibility of Ti-alloys for long-term implantation. J. Mech. Behav. Biomed. Mater. 2013, 20, 407–415. [CrossRef]
- 64. Biesiekierski, A.; Wang, J.; Abdel-Hady Gepreel, M.; Wen, C. A new look at biomedical Ti-based shape memory alloys. *Acta Biomater.* **2012**, *8*, 1661–1669. [CrossRef]
- 65. Vishnu, J.; Manivasagam, G. Surface Modification and Biological Approaches for Tackling Titanium Wear-Induced Aseptic Loosening. J. Bio-Tribo-Corros. 2021, 7, 32. [CrossRef]
- 66. Peng, K.T.; Hsu, W.H.; Shih, H.N.; Hsieh, C.W.; Huang, T.W.; Hsu, R.W.W.; Chang, P.J. The role of reactive oxygen species scavenging enzymes in the development of septic loosening after total hip replacement. *J. Bone Jt. Surg. Br. Vol.* 2011, *93-B*, 1201–1209. [CrossRef] [PubMed]
- 67. Prestat, M.; Thierry, D. Corrosion of titanium under simulated inflammation conditions: Clinical context and in vitro investigations. *Acta Biomater.* **2021**, *136*, 72–87. [CrossRef] [PubMed]
- Nguyen, G.T.; Green, E.R.; Mecsas, J. Neutrophils to the ROScue: Mechanisms of NADPH Oxidase Activation and Bacterial Resistance. *Front. Cell. Infect. Microbiol.* 2017, 7, 373. [CrossRef] [PubMed]
- 69. Groemping, Y.; Rittinger, K. Activation and assembly of the NADPH oxidase: A structural perspective. *Biochem. J.* 2005, 386, 401–416. [CrossRef]
- 70. Fujii, J.; Homma, T.; Osaki, T. Superoxide Radicals in the Execution of Cell Death. Antioxidants 2022, 11, 501. [CrossRef]

- 71. Getzoff, E.D.; Tainer, J.A.; Weiner, P.K.; Kollman, P.A.; Richardson, J.S.; Richardson, D.C. Electrostatic recognition between superoxide and copper, zinc superoxide dismutase. *Nature* **1983**, *306*, 287–290. [CrossRef]
- 72. Paumann-Page, M.; Furtmüller, P.G.; Hofbauer, S.; Paton, L.N.; Obinger, C.; Kettle, A.J. Inactivation of human myeloperoxidase by hydrogen peroxide. *Arch. Biochem. Biophys.* **2013**, 539, 51–62. [CrossRef]
- 73. Khan, A.A.; Alsahli, M.A.; Rahmani, A.H. Myeloperoxidase as an Active Disease Biomarker: Recent Biochemical and Pathological Perspectives. *Med. Sci.* **2018**, *6*, 33. [CrossRef]
- 74. Prousek, J. Fenton chemistry in biology and medicine. Pure Appl. Chem. 2007, 79, 2325–2338. [CrossRef]
- 75. Winterbourn, C.C. Toxicity of iron and hydrogen peroxide: The Fenton reaction. Toxicol. Lett. 1995, 82–83, 969–974. [CrossRef]
- 76. Kehrer, J.P. The Haber—Weiss reaction and mechanisms of toxicity. Toxicology 2000, 149, 43–50. [CrossRef] [PubMed]
- Peñarrieta-Juanito, G.; Sordi, M.B.; Henriques, B.; Dotto, M.E.R.; Teughels, W.; Silva, F.S.; Magini, R.S.; Souza, J.C.M. Surface damage of dental implant systems and ions release after exposure to fluoride and hydrogen peroxide. *J. Periodontal Res.* 2019, 54, 46–52. [CrossRef] [PubMed]
- Kalbacova, M.; Roessler, S.; Hempel, U.; Tsaryk, R.; Peters, K.; Scharnweber, D.; Kirkpatrick, J.C.; Dieter, P. The effect of electrochemically simulated titanium cathodic corrosion products on ROS production and metabolic activity of osteoblasts and monocytes/macrophages. *Biomaterials* 2007, 28, 3263–3272. [CrossRef] [PubMed]
- 79. Brown, P.S.; Bhushan, B. Mechanically durable, superomniphobic coatings prepared by layer-by-layer technique for self-cleaning and anti-smudge. *J. Colloid Interface Sci.* 2015, 456, 210–218. [CrossRef]
- 80. Ariga, K.; Hill, J.P.; Ji, Q. Layer-by-layer assembly as a versatile bottom-up nanofabrication technique for exploratory research and realistic application. *Phys. Chem. Chem. Phys.* 2007, *9*, 2319–2340. [CrossRef]
- Neacşu, I.A.; Nicoară, A.I.; Vasile, O.R.; Vasile, B.Ş. Chapter 9—Inorganic micro- and nanostructured implants for tissue engineering. In *Nanobiomaterials in Hard Tissue Engineering*; Grumezescu, A.M., Ed.; William Andrew Publishing: Norwich, NY, USA, 2016; pp. 271–295. [CrossRef]
- Alfieri, M.L.; Riccucci, G.; Ferraris, S.; Cochis, A.; Scalia, A.C.; Rimondini, L.; Panzella, L.; Spriano, S.; Napolitano, A. Deposition of Antioxidant and Cytocompatible Caffeic Acid-Based Thin Films onto Ti6Al4V Alloys through Hexamethylenediamine-Mediated Crosslinking. ACS Appl. Mater. Interfaces 2023, 15, 29618–29635. [CrossRef]
- Rased, N.H.; Vengadaesvaran, B.; Raihan, S.R.S.; Rahim, N.A. Chapter 6—Introduction to solar energy and its conversion into electrical energy by using dye-sensitized solar cells. In *Energy Materials*; Dhoble, S.J., Kalyani, N.T., Vengadaesvaran, B., Kariem Arof, A., Eds.; Elsevier: Amsterdam, The Netherlands, 2021; pp. 139–178. [CrossRef]
- Sahu, N.; Parija, B.; Panigrahi, S. Fundamental understanding and modeling of spin coating process: A review. *Indian J. Phys.* 2009, 83, 493–502. [CrossRef]
- 85. Hashizume, M.; Kunitake, T. Preparation of Self-Supporting Ultrathin Films of Titania by Spin Coating. *Langmuir* 2003, 19, 10172–10178. [CrossRef]
- 86. Chen, Y.; Xu, C.; Wang, C.-H.; Bilek, M.M.M.; Cheng, X. An effective method to optimise plasma immersion ion implantation: Sensitivity analysis and design based on low-density polyethylene. *Plasma Process. Polym.* **2022**, *19*, 2100199. [CrossRef]
- Sotoudeh Bagha, P.; Paternoster, C.; Khakbiz, M.; Sheibani, S.; Gholami, N.; Mantovani, D. Surface Modification of an Absorbable Bimodal Fe-Mn-Ag Alloy by Nitrogen Plasma Immersion Ion Implantation. *Materials* 2023, 16, 1048. [CrossRef]
- Walschus, U.; Hoene, A.; Patrzyk, M.; Lucke, S.; Finke, B.; Polak, M.; Lukowski, G.; Bader, R.; Zietz, C.; Podbielski, A.; et al. A Cell-Adhesive Plasma Polymerized Allylamine Coating Reduces the In Vivo Inflammatory Response Induced by Ti6Al4V Modified with Plasma Immersion Ion Implantation of Copper. J. Funct. Biomater. 2017, 8, 30. [CrossRef]
- Terasawa, T.-O.; Saiki, K. Growth of graphene on Cu by plasma enhanced chemical vapor deposition. *Carbon* 2012, 50, 869–874.
 [CrossRef]
- 90. Gan, Z.; Wang, C.; Chen, Z. Material Structure and Mechanical Properties of Silicon Nitride and Silicon Oxynitride Thin Films Deposited by Plasma Enhanced Chemical Vapor Deposition. *Surfaces* **2018**, *1*, 59–72. [CrossRef]
- Vasudev, M.C.; Anderson, K.D.; Bunning, T.J.; Tsukruk, V.V.; Naik, R.R. Exploration of Plasma-Enhanced Chemical Vapor Deposition as a Method for Thin-Film Fabrication with Biological Applications. ACS Appl. Mater. Interfaces 2013, 5, 3983–3994. [CrossRef]
- 92. Kaczmarek, B. Tannic Acid with Antiviral and Antibacterial Activity as A Promising Component of Biomaterials—A Minireview. *Materials* 2020, 13, 3224. [CrossRef]
- Lopes, G.K.B.; Schulman, H.M.; Hermes-Lima, M. Polyphenol tannic acid inhibits hydroxyl radical formation from Fenton reaction by complexing ferrous ions1This study is dedicated to the memory of Botany Professor Luiz F.G. Labouriau (1921–1996).1. *Biochim. Et Biophys. Acta BBA-Gen. Subj.* 1999, 1472, 142–152. [CrossRef]
- 94. Sathishkumar, G.; Gopinath, K.; Zhang, K.; Kang, E.-T.; Xu, L.; Yu, Y. Recent progress in tannic acid-driven antibacterial/antifouling surface coating strategies. *J. Mater. Chem. B* 2022, *10*, 2296–2315. [CrossRef]
- 95. Wang, Z.; Gao, J.; Zhu, L.; Meng, J.; He, F. Tannic acid-based functional coating: Surface engineering of membranes for oil-in-water emulsion separation. *Chem. Commun.* 2022, *58*, 12629–12641. [CrossRef]
- 96. Zhu, Y.; Zhou, D.; Zan, X.; Ye, Q.; Sheng, S. Engineering the surfaces of orthopedic implants with osteogenesis and antioxidants to enhance bone formation in vitro and in vivo. *Colloids Surf. B Biointerfaces* **2022**, 212, 112319. [CrossRef]

- 97. Iqbal, M.H.; Schroder, A.; Kerdjoudj, H.; Njel, C.; Senger, B.; Ball, V.; Meyer, F.; Boulmedais, F. Effect of the Buffer on the Buildup and Stability of Tannic Acid/Collagen Multilayer Films Applied as Antibacterial Coatings. *ACS Appl. Mater. Interfaces* **2020**, *12*, 22601–22612. [CrossRef]
- 98. He, X.; Gopinath, K.; Sathishkumar, G.; Guo, L.; Zhang, K.; Lu, Z.; Li, C.; Kang, E.-T.; Xu, L. UV-Assisted Deposition of Antibacterial Ag–Tannic Acid Nanocomposite Coating. ACS Appl. Mater. Interfaces 2021, 13, 20708–20717. [CrossRef]
- 99. Saiz-Poseu, J.; Mancebo-Aracil, J.; Nador, F.; Busqué, F.; Ruiz-Molina, D. The Chemistry behind Catechol-Based Adhesion. *Angew. Chem. Int. Ed.* **2019**, *58*, 696–714. [CrossRef] [PubMed]
- Geißler, S.; Barrantes, A.; Tengvall, P.; Messersmith, P.B.; Tiainen, H. Deposition Kinetics of Bioinspired Phenolic Coatings on Titanium Surfaces. *Langmuir* 2016, 32, 8050–8060. [CrossRef] [PubMed]
- 101. Qiaoxia, L.; Yujie, Z.; Meng, Y.; Yizhu, C.; Yan, W.; Yinchun, H.; Xiaojie, L.; Weiyi, C.; Di, H. Hydroxyapatite/tannic acid composite coating formation based on Ti modified by TiO₂ nanotubes. *Colloids Surf. B Biointerfaces* **2020**, *196*, 111304. [CrossRef] [PubMed]
- 102. Yang, X.; Huang, P.; Wang, H.; Cai, S.; Liao, Y.; Mo, Z.; Xu, X.; Ding, C.; Zhao, C.; Li, J. Antibacterial and anti-biofouling coating on hydroxyapatite surface based on peptide-modified tannic acid. *Colloids Surf. B Biointerfaces* 2017, 160, 136–143. [CrossRef] [PubMed]
- 103. Camós Noguer, A.; Olsen, S.M.; Hvilsted, S.; Kiil, S. Long-term stability of PEG-based antifouling surfaces in seawater. *J. Coat. Technol. Res.* **2016**, *13*, 567–575. [CrossRef]
- 104. Guo, L.L.; Cheng, Y.F.; Ren, X.; Gopinath, K.; Lu, Z.S.; Li, C.M.; Xu, L.Q. Simultaneous deposition of tannic acid and poly(ethylene glycol) to construct the antifouling polymeric coating on Titanium surface. *Colloids Surf. B Biointerfaces* 2021, 200, 111592. [CrossRef]
- Zhuk, I.; Jariwala, F.; Attygalle, A.B.; Wu, Y.; Libera, M.R.; Sukhishvili, S.A. Self-Defensive Layer-by-Layer Films with Bacteria-Triggered Antibiotic Release. ACS Nano 2014, 8, 7733–7745. [CrossRef]
- 106. Hizal, F.; Zhuk, I.; Sukhishvili, S.; Busscher, H.J.; van der Mei, H.C.; Choi, C.-H. Impact of 3D Hierarchical Nanostructures on the Antibacterial Efficacy of a Bacteria-Triggered Self-Defensive Antibiotic Coating. ACS Appl. Mater. Interfaces 2015, 7, 20304–20313. [CrossRef] [PubMed]
- 107. Steffi, C.; Shi, Z.; Kong, C.H.; Chong, S.W.; Wang, D.; Wang, W. Use of Polyphenol Tannic Acid to Functionalize Titanium with Strontium for Enhancement of Osteoblast Differentiation and Reduction of Osteoclast Activity. *Polymers* 2019, *11*, 1256. [CrossRef] [PubMed]
- 108. Lin, W.; Qi, X.; Guo, W.; Liang, D.; Chen, H.; Lin, B.; Deng, X. A barrier against reactive oxygen species: Chitosan/acellular dermal matrix scaffold enhances stem cell retention and improves cutaneous wound healing. *Stem Cell Res. Ther.* 2020, 11, 383. [CrossRef]
- Khokon, M.A.R.; Uraji, M.; Munemasa, S.; Okuma, E.; Nakamura, Y.; Mori, I.C.; Murata, Y. Chitosan-Induced Stomatal Closure Accompanied by Peroxidase-Mediated Reactive Oxygen Species Production in Arabidopsis. *Biosci. Biotechnol. Biochem.* 2010, 74, 2313–2315. [CrossRef]
- Banerjee, M.; Mallick, S.; Paul, A.; Chattopadhyay, A.; Ghosh, S.S. Heightened Reactive Oxygen Species Generation in the Antimicrobial Activity of a Three Component Iodinated Chitosan–Silver Nanoparticle Composite. *Langmuir* 2010, 26, 5901–5908. [CrossRef] [PubMed]
- 111. Parham, S.; Kharazi, A.Z.; Bakhsheshi-Rad, H.R.; Kharaziha, M.; Ismail, A.F.; Sharif, S.; Razzaghi, M.; RamaKrishna, S.; Berto, F. Antimicrobial Synthetic and Natural Polymeric Nanofibers as Wound Dressing: A Review. Adv. Eng. Mater. 2022, 24, 2101460. [CrossRef]
- 112. Kumari, S.; Tiyyagura, H.R.; Pottathara, Y.B.; Sadasivuni, K.K.; Ponnamma, D.; Douglas, T.E.L.; Skirtach, A.G.; Mohan, M.K. Surface functionalization of chitosan as a coating material for orthopaedic applications: A comprehensive review. *Carbohydr. Polym.* 2021, 255, 117487. [CrossRef]
- 113. Abinaya, B.; Prasith, T.P.; Ashwin, B.; Viji Chandran, S.; Selvamurugan, N. Chitosan in Surface Modification for Bone Tissue Engineering Applications. *Biotechnol. J.* **2019**, *14*, 1900171. [CrossRef]
- 114. Lieder, R.; Darai, M.; Thor, M.B.; Ng, C.H.; Einarsson, J.M.; Gudmundsson, S.; Helgason, B.; Gaware, V.S.; Másson, M.; Gíslason, J.; et al. In vitro bioactivity of different degree of deacetylation chitosan, a potential coating material for titanium implants. *J. Biomed. Mater. Res. Part A* 2012, 100A, 3392–3399. [CrossRef]
- 115. Tomida, H.; Fujii, T.; Furutani, N.; Michihara, A.; Yasufuku, T.; Akasaki, K.; Maruyama, T.; Otagiri, M.; Gebicki, J.M.; Anraku, M. Antioxidant properties of some different molecular weight chitosans. *Carbohydr. Res.* **2009**, 344, 1690–1696. [CrossRef]
- 116. Xue, C.; Yu, G.; Hirata, T.; Terao, J.; Lin, H. Antioxidative Activities of Several Marine Polysaccharides Evaluated in a Phosphatidylcholine-liposomal Suspension and Organic Solvents. *Biosci. Biotechnol. Biochem.* **1998**, *62*, 206–209. [CrossRef]
- 117. Xie, W.; Xu, P.; Liu, Q. Antioxidant activity of water-soluble chitosan derivatives. *Bioorganic Med. Chem. Lett.* **2001**, *11*, 1699–1701. [CrossRef] [PubMed]
- 118. Li, X.; Ma, X.-Y.; Feng, Y.-F.; Ma, Z.-S.; Wang, J.; Ma, T.-C.; Qi, W.; Lei, W.; Wang, L. Osseointegration of chitosan coated porous titanium alloy implant by reactive oxygen species-mediated activation of the PI3K/AKT pathway under diabetic conditions. *Biomaterials* 2015, *36*, 44–54. [CrossRef] [PubMed]
- Jabłoński, P.; Kyzioł, A.; Pawcenis, D.; Pucelik, B.; Hebda, M.; Migdalska, M.; Krawiec, H.; Arruebo, M.; Kyzioł, K. Electrostatic self-assembly approach in the deposition of bio-functional chitosan-based layers enriched with caffeic acid on Ti-6Al-7Nb alloys by alternate immersion. *Biomater. Adv.* 2022, 136, 212791. [CrossRef] [PubMed]

- 120. Stevanović, M.; Djošić, M.; Janković, A.; Kojić, V.; Stojanović, J.; Grujić, S.; Bujagić, I.M.; Rhee, K.Y.; Mišković-Stanković, V. The chitosan-based bioactive composite coating on titanium. *J. Mater. Res. Technol.* **2021**, *15*, 4461–4474. [CrossRef]
- 121. Rauf, A.; Imran, M.; Abu-Izneid, T.; Iahtisham Ul, H.; Patel, S.; Pan, X.; Naz, S.; Sanches Silva, A.; Saeed, F.; Rasul Suleria, H.A. Proanthocyanidins: A comprehensive review. *Biomed. Pharmacother.* **2019**, *116*, 108999. [CrossRef]
- 122. de la Iglesia, R.; Milagro, F.I.; Campión, J.; Boqué, N.; Martínez, J.A. Healthy properties of proanthocyanidins. *BioFactors* **2010**, *36*, 159–168. [CrossRef] [PubMed]
- 123. Park, Y.S.; Jeon, M.H.; Hwang, H.J.; Park, M.R.; Lee, S.H.; Kim, S.G.; Kim, M. Antioxidant activity and analysis of proanthocyanidins from pine (*Pinus densiflora*) needles. *Nutr. Res. Pract.* 2011, *5*, 281–287. [CrossRef]
- Yang, L.; Xian, D.; Xiong, X.; Lai, R.; Song, J.; Zhong, J. Proanthocyanidins against Oxidative Stress: From Molecular Mechanisms to Clinical Applications. *BioMed Res. Int.* 2018, 2018, 8584136. [CrossRef]
- 125. Andersen-Civil, A.I.S.; Leppä, M.M.; Thamsborg, S.M.; Salminen, J.-P.; Williams, A.R. Structure-function analysis of purified proanthocyanidins reveals a role for polymer size in suppressing inflammatory responses. *Commun. Biol.* **2021**, *4*, 896. [CrossRef]
- 126. Tenkumo, T.; Aobulikasimu, A.; Asou, Y.; Shirato, M.; Shishido, S.; Kanno, T.; Niwano, Y.; Sasaki, K.; Nakamura, K. Proanthocyanidin-rich grape seed extract improves bone loss, bone healing, and implant osseointegration in ovariectomized animals. *Sci. Rep.* **2020**, *10*, 8812. [CrossRef]
- 127. Tang, J.; Chen, L.; Yan, D.; Shen, Z.; Wang, B.; Weng, S.; Wu, Z.; Xie, Z.; Shao, J.; Yang, L.; et al. Surface Functionalization with Proanthocyanidins Provides an Anti-Oxidant Defense Mechanism That Improves the Long-Term Stability and Osteogenesis of Titanium Implants. Int. J. Nanomed. 2020, 15, 1643–1659. [CrossRef]
- Bai, Z.; Hu, K.; Shou, Z.; Yu, J.; Meng, H.; Zhou, H.; Chen, L.; Yu, T.; Lu, R.; Li, N.; et al. Layer-by-layer assembly of procyanidin and collagen promotes mesenchymal stem cell proliferation and osteogenic differentiation in vitro and in vivo. *Regen. Biomater.* 2023, 10, rbac107. [CrossRef] [PubMed]
- 129. Zhou, Q.; Wu, T.; Bai, Z.; Hong, G.; Bian, J.; Xie, H.; Chen, C. A silane-based coupling strategy for enhancing the mechanical properties of proanthocyanidin nanocoatings on Ti dental implants. *Appl. Surf. Sci.* **2022**, *602*, 154400. [CrossRef]
- 130. Maccarone, R.; Tisi, A.; Passacantando, M.; Ciancaglini, M. Ophthalmic Applications of Cerium Oxide Nanoparticles. *J. Ocul. Pharmacol. Ther.* **2019**, *36*, 376–383. [CrossRef]
- 131. Li, J.; Wen, J.; Li, B.; Li, W.; Qiao, W.; Shen, J.; Jin, W.; Jiang, X.; Yeung, K.W.K.; Chu, P.K. Valence State Manipulation of Cerium Oxide Nanoparticles on a Titanium Surface for Modulating Cell Fate and Bone Formation. *Adv. Sci.* 2018, *5*, 1700678. [CrossRef] [PubMed]
- 132. Dhall, A.; Self, W. Cerium Oxide Nanoparticles: A Brief Review of Their Synthesis Methods and Biomedical Applications. *Antioxidants* **2018**, *7*, 97. [CrossRef]
- Nelson, B.C.; Johnson, M.E.; Walker, M.L.; Riley, K.R.; Sims, C.M. Antioxidant Cerium Oxide Nanoparticles in Biology and Medicine. *Antioxidants* 2016, 5, 15. [CrossRef]
- 134. Esch, F.; Fabris, S.; Zhou, L.; Montini, T.; Africh, C.; Fornasiero, P.; Comelli, G.; Rosei, R. Electron Localization Determines Defect Formation on Ceria Substrates. *Science* 2005, *309*, 752–755. [CrossRef]
- 135. Heckert, E.G.; Karakoti, A.S.; Seal, S.; Self, W.T. The role of cerium redox state in the SOD mimetic activity of nanoceria. *Biomaterials* 2008, *29*, 2705–2709. [CrossRef]
- 136. Korsvik, C.; Patil, S.; Seal, S.; Self, W.T. Superoxide dismutase mimetic properties exhibited by vacancy engineered ceria nanoparticles. *Chem. Commun.* 2007, 1056–1058. [CrossRef]
- 137. Pirmohamed, T.; Dowding, J.M.; Singh, S.; Wasserman, B.; Heckert, E.; Karakoti, A.S.; King, J.E.S.; Seal, S.; Self, W.T. Nanoceria exhibit redox state-dependent catalase mimetic activity. *Chem. Commun.* **2010**, *46*, 2736–2738. [CrossRef] [PubMed]
- 138. Li, K.; Xie, Y.; You, M.; Huang, L.; Zheng, X. Plasma sprayed cerium oxide coating inhibits H2O2-induced oxidative stress and supports cell viability. *J. Mater. Sci. Mater. Med.* **2016**, *27*, 100. [CrossRef] [PubMed]
- Mahapatra, C.; Singh, R.K.; Lee, J.-H.; Jung, J.; Hyun, J.K.; Kim, H.-W. Nano-shape varied cerium oxide nanomaterials rescue human dental stem cells from oxidative insult through intracellular or extracellular actions. *Acta Biomater.* 2017, 50, 142–153. [CrossRef] [PubMed]
- Li, X.; Qi, M.; Sun, X.; Weir, M.D.; Tay, F.R.; Oates, T.W.; Dong, B.; Zhou, Y.; Wang, L.; Xu, H.H.K. Surface treatments on titanium implants via nanostructured ceria for antibacterial and anti-inflammatory capabilities. *Acta Biomater.* 2019, 94, 627–643. [CrossRef] [PubMed]
- 141. Zhang, H.; Qiu, J.; Liu, X. Enhanced antioxidant capability and osteogenic property of medical titanium by cerium plasma immersion ion implantation. *Surf. Interfaces* **2021**, *26*, 101402. [CrossRef]
- 142. Qi, S.; Wu, J.; Xu, Y.; Zhang, Y.; Wang, R.; Li, K.; Xu, Y. Chemical Stability and Antimicrobial Activity of Plasma-Sprayed Cerium Oxide–Incorporated Calcium Silicate Coating in Dental Implants. *Implant Dent.* **2019**, *28*, 564–570. [CrossRef] [PubMed]
- 143. Chen, M.; Wang, D.; Li, M.; He, Y.; He, T.; Chen, M.; Hu, Y.; Luo, Z.; Cai, K. Nanocatalytic Biofunctional MOF Coating on Titanium Implants Promotes Osteoporotic Bone Regeneration through Cooperative Pro-osteoblastogenesis MSC Reprogramming. ACS Nano 2022, 16, 15397–15412. [CrossRef]
- 144. Mandracci, P.; Mussano, F.; Ceruti, P.; Pirri, C.F.; Carossa, S. Reduction of bacterial adhesion on dental composite resins by silicon–oxygen thin film coatings. *Biomed. Mater.* **2015**, *10*, 015017. [CrossRef]

- 145. Mandracci, P.; Ceruti, P.; Ricciardi, C.; Mussano, F.; Carossa, S. a-SiOx Coatings Grown on Dental Materials by PECVD: Compositional Analysis and Preliminary Investigation of Biocompatibility Improvements. *Chem. Vap. Depos.* 2010, 16, 29–34. [CrossRef]
- 146. Alves Silva, E.C.; Tanomaru-Filho, M.; da Silva, G.F.; Delfino, M.M.; Cerri, P.S.; Guerreiro-Tanomaru, J.M. Biocompatibility and Bioactive Potential of New Calcium Silicate–based Endodontic Sealers: Bio-C Sealer and Sealer Plus BC. *J. Endod.* 2020, *46*, 1470–1477. [CrossRef]
- 147. Arcos, D.; Vallet-Regí, M. Sol-gel silica-based biomaterials and bone tissue regeneration. *Acta Biomater.* **2010**, *6*, 2874–2888. [CrossRef]
- 148. Heimann, R.B. Silicon Nitride, a Close to Ideal Ceramic Material for Medical Application. Ceramics 2021, 4, 208–223. [CrossRef]
- 149. Ilyas, A.; Odatsu, T.; Shah, A.; Monte, F.; Kim, H.K.W.; Kramer, P.; Aswath, P.B.; Varanasi, V.G. Amorphous Silica: A New Antioxidant Role for Rapid Critical-Sized Bone Defect Healing. *Adv. Healthc. Mater.* **2016**, *5*, 2199–2213. [CrossRef]
- 150. Monte, F.A.D.; Awad, K.R.; Ahuja, N.; Kim, H.K.W.; Aswath, P.; Brotto, M.; Varanasi, V.G. Amorphous Silicon Oxynitrophosphide-Coated Implants Boost Angiogenic Activity of Endothelial Cells. *Tissue Eng. Part A* **2019**, *26*, 15–27. [CrossRef] [PubMed]
- 151. Mussano, F.; Genova, T.; Laurenti, M.; Munaron, L.; Pirri, C.F.; Rivolo, P.; Carossa, S.; Mandracci, P. Hydrogenated amorphous silicon coatings may modulate gingival cell response. *Appl. Surf. Sci.* **2018**, *436*, 603–612. [CrossRef]
- 152. Bigham, A.; Rahimkhoei, V.; Abasian, P.; Delfi, M.; Naderi, J.; Ghomi, M.; Dabbagh Moghaddam, F.; Waqar, T.; Nuri Ertas, Y.; Sharifi, S.; et al. Advances in tannic acid-incorporated biomaterials: Infection treatment, regenerative medicine, cancer therapy, and biosensing. *Chem. Eng. J.* **2022**, 432, 134146. [CrossRef]
- Kaczmarek-Szczepańska, B.; Połkowska, I.; Paździor-Czapula, K.; Nowicka, B.; Gierszewska, M.; Michalska-Sionkowska, M.; Otrocka-Domagała, I. Chitosan/Phenolic Compounds Scaffolds for Connective Tissue Regeneration. *J. Funct. Biomater.* 2023, 14, 69. [CrossRef] [PubMed]
- 154. Widsten, P.; Salo, S.; Niemelä, K.; Helin, H.; Salonen, M.; Alakomi, H.-L. Tannin-Based Microbicidal Coatings for Hospital Privacy Curtains. J. Funct. Biomater. 2023, 14, 187. [CrossRef]
- 155. Amarowicz, R. Tannins: The new natural antioxidants? Eur. J. Lipid Sci. Technol. 2007, 109, 549–551. [CrossRef]
- 156. Sanguedolce, M.; Saffioti, M.R.; Rotella, G.; Curcio, F.; Cassano, R.; Umbrello, D.; Filice, L. The Effects of Substrate Material on Chitosan Coating Performance for Biomedical Application. *Procedia CIRP* **2022**, *108*, 817–820. [CrossRef]
- 157. Ganesh, S.S.; Anushikaa, R.; Swetha Victoria, V.S.; Lavanya, K.; Shanmugavadivu, A.; Selvamurugan, N. Recent Advancements in Electrospun Chitin and Chitosan Nanofibers for Bone Tissue Engineering Applications. *J. Funct. Biomater.* 2023, 14, 288. [CrossRef]
- 158. Oe, T.; Dechojarassri, D.; Kakinoki, S.; Kawasaki, H.; Furuike, T.; Tamura, H. Microwave-Assisted Incorporation of AgNP into Chitosan–Alginate Hydrogels for Antimicrobial Applications. *J. Funct. Biomater.* **2023**, *14*, 199. [PubMed]
- Yu, K.; Song, Y.; Lin, J.; Dixon, R.A. The complexities of proanthocyanidin biosynthesis and its regulation in plants. *Plant Commun.* 2023, 4, 100498. [CrossRef] [PubMed]
- 160. Banavar, S.; Deshpande, A.; Sur, S.; Andreescu, S. Ceria nanoparticle theranostics: Harnessing antioxidant properties in biomedicine and beyond. *J. Phys. Mater.* **2021**, *4*, 042003. [CrossRef]
- Yokel, R.A.; Hussain, S.; Garantziotis, S.; Demokritou, P.; Castranova, V.; Cassee, F.R. The yin: An adverse health perspective of nanoceria: Uptake, distribution, accumulation, and mechanisms of its toxicity. *Environ. Sci. Nano* 2014, 1, 406–428. [CrossRef] [PubMed]
- Huang, Y.; Li, P.; Zhao, R.; Zhao, L.; Liu, J.; Peng, S.; Fu, X.; Wang, X.; Luo, R.; Wang, R.; et al. Silica nanoparticles: Biomedical applications and toxicity. *Biomed. Pharmacother.* 2022, 151, 113053. [CrossRef] [PubMed]
- 163. Zhang, L.-C.; Chen, L.-Y.; Wang, L. Surface Modification of Titanium and Titanium Alloys: Technologies, Developments, and Future Interests. *Adv. Eng. Mater.* **2020**, *22*, 1901258. [CrossRef]
- 164. Xu, D.-P.; Li, Y.; Meng, X.; Zhou, T.; Zhou, Y.; Zheng, J.; Zhang, J.-J.; Li, H.-B. Natural Antioxidants in Foods and Medicinal Plants: Extraction, Assessment and Resources. *Int. J. Mol. Sci.* 2017, *18*, 96. [CrossRef]

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